

LONDON
SCHOOL of
HYGIENE
& TROPICAL
MEDICINE



Nikolay, B; Diallo, M; Faye, O; Boye, CS; Sall, AA (2012) Vector competence of *Culex neavei* (Diptera: Culicidae) for Usutu virus. *The American journal of tropical medicine and hygiene*, 86 (6). pp. 993-6. ISSN 0002-9637 DOI: 10.4269/ajtmh.2012.11-0509

Downloaded from: <http://researchonline.lshtm.ac.uk/623479/>

DOI: [10.4269/ajtmh.2012.11-0509](https://doi.org/10.4269/ajtmh.2012.11-0509)

Usage Guidelines

Please refer to usage guidelines at <http://researchonline.lshtm.ac.uk/policies.html> or alternatively contact researchonline@lshtm.ac.uk.

Available under license: <http://creativecommons.org/licenses/by-nc-nd/2.5/>

Vector Competence of *Culex neavei* (Diptera: Culicidae) for Usutu Virus

Birgit Nikolay, Mawlouth Diallo, Ousmane Faye, Cheikh S. Boye, and Amadou A. Sall*

Unité des Arbovirus et Virus de Fièvres Hémorragiques, et Unité d'Entomologie Médicale, Institut Pasteur de Dakar, Dakar, Senegal;
Faculté de Médecine, Pharmacie, Odonto-Stomatologie, Université Cheikh Anta Diop, Dakar, Senegal;
Faculty of Life Sciences, University of Vienna, Vienna, Austria

Abstract. Usutu virus (USUV), a flavivirus belonging to the Japanese encephalitis serocomplex, was isolated for the first time from a *Culex neavei* mosquito in 1959 in South Africa. Despite multiple isolations of USUV from *Cx. neavei* in Africa, its vector competence remains unproven. Therefore, we infected *Cx. neavei* orally with the USUV reference strain and used reverse transcription–polymerase chain reaction and an indirect immunofluorescence assay to detect virus in bodies, legs, wings, and saliva of mosquitoes. We demonstrated the susceptibility of *Cx. neavei* mosquitoes for the USUV reference strain, its potential to be transmitted, and infection, dissemination, and transmission rates of 90.9%, 40.0%, and 81.3%, respectively. Also, we showed that infection rates are dependent on the virus titer of the blood meal. Given the bionomics of *Cx. neavei*, its role as enzootic vector for USUV in Africa in a mosquito–bird transmission cycle or as bridge vector for USUV transmission to humans is discussed.

INTRODUCTION

Usutu virus (USUV), a member of the Japanese encephalitis serocomplex of the genus *Flavivirus* (family *Flaviviridae*), was isolated for the first time from *Culex neavei* mosquitoes (Theobald, 1906) in the Natal region of South Africa.^{1,2} Since then, USUV has been isolated from mosquitoes or detected by serologic analysis in birds in Africa (Senegal, Central African Republic, Burkina Faso, Côte d'Ivoire, Morocco, Nigeria, and Uganda) and Europe (Austria, Hungary, Switzerland, Italy, Spain, Germany, Poland, Czech Republic, and the United Kingdom).^{3–12}

In 2001, USUV emerged for the first time outside Africa, in Austria, and caused severe disease and high mortality rates among blackbirds and great gray owls.¹³ Human infections with USUV reported in Central African Republic, Burkina Faso, and Italy in 1981, 2004, and 2009, respectively, have been associated with fever, rash, and jaundice among immunocompetent patients or a severe neurologic syndrome among immunodeficient patients, which emphasizes its potential threat to human health.^{3,14–16} In addition, a serosurvey of neutralizing antibodies against USUV among patients with fever and rash in Austria showed a prevalence of 25%.¹⁷ These findings suggest that USUV is transmitted by mosquitoes and its transmission cycle involves birds and humans, although their specific roles remain to be investigated.

Usutu virus has been isolated from numerous mosquito species. However, the vector competence has not been determined for any of them. In Europe, USUV isolations have been reported from *Cx. pipiens*, an urban mosquito known as a bridge vector between birds and humans for West Nile virus, but also from *Aedes albopictus* mosquitoes.^{18,19} In Africa, USUV has mostly been isolated from *Cx. neavei* and to a lesser extent from *Cx. perfuscus*, *Cx. quinquefasciatus*, *Aedes minutus*, *Mansonia africana*, and *Coquillettidia aurites*.³ Given the numerous isolations of USUV from *Cx. neavei*, we re-investigated its vector competence for USUV because a previous limited study showed only its susceptibility to USUV but no transmission.¹ We report the susceptibility and ability

to transmit USUV of *Cx. neavei* by determining infection and dissemination rates and demonstrating the presence of USUV in the mosquito saliva as an indicator of virus transmission.

MATERIALS AND METHODS

Virus stocks used for infections. Mosquitoes were analyzed for their vector competence by using USUV reference strain SAAR1776.¹ Virus stocks were prepared by infecting AP61 (*Aedes pseudoscutellaris*) cells, intracerebral inoculation of suckling mice, and obtaining virus titers as described, using PS cells (porcine stable kidney cell line; American Type Culture Collection, Manassas, VA).²⁰ The PS cells were used because plaque formation was necessary for quantification of the virus and cannot be observed in AP61 cells. The viral stock titer was 6×10^7 plaque-forming units (PFU)/mL when prepared on AP61 cells. A higher virus titer of 5.5×10^8 PFU/mL could only be obtained by intracerebral inoculation of suckling mice. Although data on USUV viremia in birds are not available, studies on closely related West Nile virus showed that several bird species develop titers of 5–10 logs PFU/mL, which justifies the choice of 7–8 logs PFU/mL as an initial infective titer for our experiments.²¹

Mosquitoes. *Culex neavei* larvae were collected from a ground pool in Barkedji (15°17'N, 14°53'W), a village in the northern Sahelian region of Senegal. For the infection experiments, F₁ generation adult mosquitoes were reared in the laboratory by using standard methods with a temperature of $27 \pm 1^\circ\text{C}$, a relative humidity of 70–75%, and a 12-hour photoperiod.²²

Oral infection of mosquitoes. Oral infections were performed as described with minor changes.²³ Five to 6 day-old F₁ female mosquitoes were sucrose-starved for 24–48 hours before being exposed for 1 hour to an infectious blood meal. The infectious meal consisted of 33% rabbit erythrocytes washed with $1 \times$ phosphate-buffered saline, 33% virus stock, 20% volume of fetal bovine serum (FBS), 1% (w/v) sucrose, and 5 mM ATP. The blood meals were administered in glass membrane feeders by using chicken skin as membranes. After each feeding, the remainder was titrated. Engorged mosquitoes were incubated at 27°C , a relative humidity of 70–80% and fed with 10% sucrose solution. After 14 days of incubation, each mosquito was processed separately. First, their legs

*Address correspondence to Amadou A. Sall, Unité des Arbovirus et Virus de Fièvres Hémorragiques, Institut Pasteur de Dakar, BP 220 Dakar, Senegal. E-mail: asall@pasteur.sn

and wings were removed and combined in a tube. Mosquitoes were allowed to salivate for 20 minutes into capillary tubes containing pure FBS and bodies were collected in a tube. All samples were stored at -80°C before testing.

Analysis of mosquito samples. Each mosquito has been tested for the presence of USUV by reverse transcription–polymerase chain reaction (RT-PCR) and indirect immunofluorescence assay (IFA) by using a staged procedure starting with screening of all bodies, screening of legs and wings of mosquitoes with virus-positive bodies, and screening of saliva from mosquitoes with virus-positive legs and wings. Bodies of mosquitoes and their legs and wings were homogenized separately in $450\ \mu\text{L}$ of cell culture medium containing 10% FBS. Samples were centrifuged for 5 minutes at $7,500\ \text{rpm}$ at 4°C . Fetal bovine serum containing saliva was removed from capillaries and diluted with $450\ \mu\text{L}$ of cell culture medium containing 10% FBS. Supernatants and medium containing saliva were filtered by using a 1-mL syringe (Artsana, Como, Italy) and sterilized $0.20\text{-}\mu\text{m}$ filters (Sartorius, Göttingen, Germany).

For each specimen, RNA was extracted by using the QiaAmp Viral RNA Extraction Kit (Qiagen, Heiden, Germany), and reverse transcribed with avian myeloblastosis virus RTase (Promega, Madison, WI) and 382_NS3R1 primer (5'-TATCCTCCTGGTCTTTCCCG-3') (TIB Molbiol, Berlin, Germany) according to the manufacturer's instructions. The resulting complementary DNA was amplified by using the Go-Taq PCR Kit (Promega) and 13_NS3F1 (5'-GATGGTGACTTCACCTTAT-3) and 382_NS3R1 primers (TIB Molbiol). Cycling conditions were 5 minutes at 95°C ; 45 cycles of 1 minute at 95°C , 1 minute at 52°C , and 1 minute at 72°C ; and 10 minutes at 72°C . The RT-PCR was performed directly for homogenized samples and for cell culture supernatants to confirm a positive IFA result. Before the IFA, 2 mL of delta cell culture tubes (NUNC, Langensfeld, Germany) containing 80% confluent AP61 cells were incubated for 1 hour at 27°C with $100\ \mu\text{L}$ of filtered supernatant. Two milliliters of cell culture medium containing 5% FBS were added and cells were incubated for 14 days. The medium was changed after 10 days. The IFA was performed by using USUV-specific hyperimmune ascitic fluid as described.²⁴ Samples positive by IFA were confirmed by RT-PCR and negative samples were passaged up to 4 times to confirm negativity.

Interpretation of results. Samples were considered positive once found positive either by RT-PCR, IFA, or both. Mosquitoes with only a virus-positive body indicated infection limited to the midgut, whereas virus in the mosquito body, legs, and wings confirmed disseminated infection. Virus in saliva indicated possible transmission of virus. Infection (no. infected mosquito bodies/no. mosquitoes tested), dissemina-

tion (no. mosquitoes with infected wings and legs/no. infected mosquitoes), and transmission (no. mosquitoes with virus in saliva/no. disseminated mosquitoes) rates were calculated and compared by using the two-tailed Fisher's exact test or chi-square test (Epi-Info Software; Centers for Disease Control and Prevention, Atlanta, GA). P values < 0.05 were considered significant.

RESULTS

Culex neavei mosquitoes were orally infected with USUV reference strain SAAR1776 in blood with initial titers of 2×10^7 PFU/mL or 1.8×10^8 PFU/mL. Infection, dissemination, and transmission rates are summarized in Table 1. Virus titers of blood meals after the infection experiment (post-infection) ranged from 2×10^4 to 4.5×10^6 PFU/mL, respectively. The decrease in virus titer during feeding experiments might indicate a higher sensitivity of USUV to temperature or other environmental factors.

The average infection rate of mosquitoes was 23.1% without observable virus dissemination and viral titers were 2×10^4 – 9×10^4 PFU/mL post-infection, whereas USUV infected 90.9% of mosquitoes and showed dissemination and transmission rates of 40.0% and 81.3%, respectively, when the viral titer was 4.5×10^6 PFU/mL post-infection. The infection rates with blood meals containing viral titers of 2×10^4 – 9×10^4 PFU/mL post-infection were not significantly different among replicates A, B, and C (Table 1) ($P = 1$, by Fisher's exact test). However, the mean infection rate of the three replicates (23.1%) was significantly different from that of the replicate with a viral titer of 4.5×10^6 PFU/mL post-infection (90.9%) ($P < 0.0001$, by Fisher's exact test). Dissemination rates of 0% and 40.0% for oral infections with viral titers of 2×10^4 – 9×10^4 PFU/mL post-infection and 4.5×10^6 PFU/mL post-infection, respectively, were not statistically different ($P = 0.2824$, by Fisher's exact test).

The combination of RT-PCR and IFA enabled detection of USUV in 43 mosquito bodies, 16 legs and wings, and 13 saliva samples. The RT-PCR performed directly on homogenized samples enabled detection of virus in 39 (90.7%) of 43 mosquito bodies, 12 (75%) of 16 legs and wings, and 2 (15.4%) of 13 saliva samples, which were virus positive after being passaged in AP61 cells. However, it seems that the quantity of virus in several mosquito samples was too low to be detected only by RT-PCR and led to the use of serial passage on AP61 cells for amplification of virus and more efficient detection. Among samples found negative by RT-PCR, USUV could be detected after first, second, or third

TABLE 1

Oral infection experiments with *Culex neavei* mosquitoes fed with blood meals initially containing 2×10^7 PFU/mL and 1.8×10^8 PFU/mL of Usutu virus reference strain SAAR1776*

Blood meal titer before infection (PFU/mL)	Replicate	Blood meal titer post-infection (PFU/mL)	Infection rate†	Dissemination rate‡	Transmission rate§
2×10^7	A	2×10^4	1/3 (33.3%)	0/1 (0%)	–
	B	4×10^4	2/9 (22.2%)	0/2 (0%)	–
	C	9×10^4	0/1 (0%)	–	–
1.8×10^8	A'	4.5×10^6	40/44 (90.9%)	16/40 (40.0%)	13/16 (81.3%)

*PFU = plaque-forming unit. Virus titer of the blood meal was additionally quantified after the feeding procedure (post-infection). After 14 days incubation, fed mosquitoes were analyzed for infection of their bodies (infection), of their legs and wings (dissemination), and the presence of virus in the saliva (transmission).

†No. infected mosquito bodies/no. mosquitoes tested.

‡No. mosquitoes with infected wings and legs/no. infected mosquitoes.

§No. mosquitoes with virus in saliva/no. disseminated mosquitoes.

passage in AP61 cells for 68.4%, 21.1%, and 10.5% of samples, respectively. The fourth passage did not show additional positive samples.

Furthermore, our experiments showed significant differences in the percentage of feeding success (no. engorged mosquitoes/no. mosquitoes exposed to blood meal \times 100) and depended on the time of day when experiments were performed. The feeding success in experiments obtained between 3:00 PM and 4:00 PM ranged from 1% to 9%, and the feeding success obtained between 9:30 PM and 10:30 PM was 45% ($P < 0.0001$, by chi-square test).

DISCUSSION

Repeated USUV isolations from *Cx. neavei* suggested its role as a potential vector species. Although, susceptibility of *Cx. neavei* to USUV could be demonstrated earlier with an infection rate of 20%, USUV transmission to hamsters failed.¹ In our experimental infections, USUV could infect *Cx. neavei* and disseminate in the mosquito bodies. Furthermore, the presence of virus in the mosquito saliva (81.3% for the blood meal with 4.5×10^6 PFU/mL post-infection) is an indicator for virus transmission, and thereby strongly suggests the potential of *Cx. neavei* mosquitoes to transmit the USUV reference strain.

The wide distribution of *Cx. neavei* in Africa indicates a possible key role in USUV dispersal and endemic transmission.²⁵ *Culex neavei* was also found associated with several other flaviviruses (Bagaza, Yaounde, West Nile, and Koutango), alphaviruses (Sindbis and Babanki), and orbivirus (ArD66707, a tentatively new orbivirus regularly isolated in Senegal and provisionally named Sanar virus).^{15,26} The prevalence of USUV among birds and the ornithophilic behavior of *Cx. neavei* suggest that these mosquitoes may play a major role as endemic vectors in a mosquito-bird transmission cycle in Africa.^{27,28} This hypothesis is further supported by the observed peak biting activity of *Cx. neavei* at night, and its high abundance in the tree canopy, which is consistent with roost and nest behavior of many bird species.^{29,30}

Oral infection rates for *Cx. neavei* with USUV were 23.1% and 90.9%, respectively, for infective blood meals with virus titers of 2×10^4 – 9×10^4 PFU/mL and 4.5×10^6 PFU/mL post-infection. The dose dependence of vector susceptibility on infecting titer has been demonstrated for other arboviruses, such as West Nile virus and St. Louis encephalitis virus.^{31,32} Although in experimental infections a viral titer of 4.5×10^6 PFU/mL post-infection was necessary for high infection rates and detectable virus dissemination, low-titer viremia might still contribute to USUV transmission in nature.

In Senegal, 141 USUV isolations from *Cx. neavei* mosquitoes have been reported.³ A recent study of the temporal distribution of *Cx. neavei* mosquitoes in Barkedji showed highest mosquito abundances during September–November, and showed a positive correlation with rainfall after a lag time of one month.³³ It is noteworthy that this period of high vector abundance, and therefore potentially high USUV transmission activity, coincides with the time when resident and migratory birds meet in Senegal.³⁴ Therefore, infection of migratory bird species with USUV might lead to its further dispersal in Africa, or even provide an explanation for USUV introduction into Europe.

Furthermore, in Senegal, *Cx. neavei* is present in all biogeographic zones, ranging from the Sahelian to the Sudan-Guinean zones.^{3,23,33} Analysis of the spatial distribution of *Cx. neavei* in Barkedji showed its high abundance in sylvatic environment but low abundance in villages.³³ Therefore, mosquitoes are likely to encounter birds and other mammals gathering around water places, facilitating an enzootic transmission cycle of USUV. The low abundance of this vector species in inhabited areas, in combination with the low anthropophily, may explain the rarity of reported human USUV infections in Africa.^{27,35} This observation has been further strengthened by a serosurvey among the human populations in an area of reported USUV activity in Senegal that did not show any evidence for USUV infections (Nikolay B and others, unpublished data). These findings suggest that in Africa, USUV transmission by *Cx. neavei* occurs mainly in a sylvatic cycle with minor impact on the human population.

An often-encountered problem during experimental oral infections is to obtain a sufficient number of fed mosquitoes necessary to draw conclusions about infection, dissemination, and transmission rates. Interestingly, we have observed that the shift of feeding time from 3:00 PM to 9:30 PM has led to a significant higher feeding success for *Cx. neavei*, which is consistent with the observed peak biting activity in nature.²⁹ This observation should be taken into account in the design of experimental infection studies on *Cx. neavei* and other closely related species.

Regarding the methods used for virus detection, we observed that analysis of homogenized samples by RT-PCR is not sufficient to detect all positive samples. Therefore, we recommend at least three serial passages in AP61 cells, followed by IFA and RT-PCR analysis to increase the sensitivity of detection.

We have demonstrated the potential of *Cx. neavei* to transmit USUV and suggest involvement of this mosquito species in sylvatic transmission in Africa. However, this study was based on one population of *Cx. neavei* mosquitoes, and the susceptibility and ability to transmit virus may vary significantly among different populations of the same species. Therefore, further studies on *Cx. neavei* populations from different countries or bioclimatic zones might provide better understanding of its vector competence.

We are still far from understanding completely the USUV transmission cycle. Besides the documented infections of birds and humans, the range of vertebrate species serving as USUV reservoir hosts is largely unknown. Furthermore, identification of other wild or domestic mosquito vector species is necessary for a better understanding of the modes of USUV transmission. In Europe, USUV has been isolated from *Cx. pipiens*, suggesting its role as a vector.¹⁷ A member of the *Cx. pipiens* complex, *Cx. quinquefasciatus*, is abundant in the domestic environment in Africa. Therefore, investigation of its vector competence might also bring insights to understand the USUV transmission cycle in the domestic environment in Africa.

Received August 5, 2011. Accepted for publication February 4, 2012.

Financial support: This study was supported by Institut Pasteur de Dakar and National Institutes of Health grant 5R01A 1069145. Birgit Nikolay is awardee of a scholarship by the Austrian Federal Ministry for Science and Research.

Authors' addresses: Birgit Nikolay, Unité des Arbovirus et Virus de Fièvres Hémorragiques, Institut Pasteur de Dakar, Dakar, Senegal and Faculty of Life Sciences, University of Vienna, Vienna, Austria,

E-mails: bnikolay@pasteur.sn or birgitnikolay@gmail.com. Mawlouth Diallo, Unité d'Entomologie Médicale, Institut Pasteur de Dakar, Dakar, Senegal, E-mail: diallo@pasteur.sn. Ousmane Faye and Amadou A. Sall, Unité des Arbovirus et Virus de Fièvres Hémorragiques, Institut Pasteur de Dakar, Dakar, Senegal, E-mails: ofaye@pasteur.sn and asall@pasteur.sn. Cheikh S. Boye, Faculté de Médecine, Pharmacie, Odonto-Stomatologie, Université Cheikh Anta Diop, Dakar, Senegal, E-mail: csboye@email.com.

REFERENCES

- McIntosh BM, 1985. Usutu (SA Ar 1776), nouvel arbovirus du groupe B. *International Catalogue of Arboviruses* 3: 1059–1060.
- Woodall JP, 1964. The viruses isolated from arthropods at the East African Virus Research Institute in the 26 years ending December 1963. *Proc E Afr Acad II*: 141–146.
- Nikolay B, Diallo M, Boye CS, Sall AA, 2011. Usutu virus in Africa. *Vector Borne Zoonotic Dis* 11: 1417–1423.
- Figuerola J, Baouab RE, Soriguer R, Fassi-Fihri O, Llorente F, Jimenez-Clavero MA, 2009. West Nile virus antibodies in wild birds, Morocco, 2008. *Emerg Infect Dis* 15: 1651–1653.
- Bakonyi T, Erdelyi K, Ursu K, Ferenczi E, Csorgo T, Lussy H, Chvala S, Bukovsky C, Meister T, Weissenbock H, Nowotny N, 2007. Emergence of Usutu virus in Hungary. *J Clin Microbiol* 45: 3870–3874.
- Busquets N, Alba A, Allepuz A, Aranda C, Ignacio Nunez J, 2008. Usutu virus sequences in *Culex pipiens* (Diptera: Culicidae), Spain. *Emerg Infect Dis* 14: 861–863.
- Linke S, Niedrig M, Kaiser A, Ellerbrok H, Muller K, Muller T, Conraths FJ, Muhle RU, Schmidt D, Koppen U, Bairlein F, Berthold P, Pauli G, 2007. Serologic evidence of West Nile virus infections in wild birds captured in Germany. *Am J Trop Med Hyg* 77: 358–364.
- Buckley A, Dawson A, Moss SR, Hinsley SA, Bellamy PE, Gould EA, 2003. Serological evidence of West Nile virus, Usutu virus and Sindbis virus infection of birds in the UK. *J Gen Virol* 84: 2807–2817.
- Hubalek Z, Halouzka J, Juricova Z, Sikutova S, Rudolf I, Honza M, Jankova J, Chytil J, Marec F, Sitko J, 2008. Serologic survey of birds for West Nile flavivirus in southern Moravia (Czech Republic). *Vector Borne Zoonotic Dis* 8: 659–666.
- Hubalek Z, Wegner E, Halouzka J, Tryjanowski P, Jerzak L, Sikutova S, Rudolf I, Kruszewicz AG, Jaworski Z, Wlodarczyk R, 2008. Serologic survey of potential vertebrate hosts for West Nile virus in Poland. *Viral Immunol* 21: 247–253.
- Manarolla G, Bakonyi T, Gallazzi D, Crosta L, Weissenbock H, Dorrestein GM, Nowotny N, 2010. Usutu virus in wild birds in northern Italy. *Vet Microbiol* 141: 159–163.
- Steinmetz HW, Bakonyi T, Weissenbock H, Hatt JM, Eulenberger U, Robert N, Hoop R, Nowotny N, 2011. Emergence and establishment of Usutu virus infection in wild and captive avian species in and around Zurich, Switzerland: genomic and pathologic comparison to other central European outbreaks. *Vet Microbiol* 148: 207–212.
- Chvala S, Bakonyi T, Bukovsky C, Meister T, Brugger K, Rubel F, Nowotny N, Weissenbock H, 2007. Monitoring of Usutu virus activity and spread by using dead bird surveillance in Austria, 2003–2005. *Vet Microbiol* 122: 237–245.
- Cavrini F, Gaibani P, Longo G, Pierro AM, Rossini G, Bonilauri P, Gerundi GE, Di Benedetto F, Pasetto A, Girardis M, Dottori M, Landini MP, Sambri V, 2009. Usutu virus infection in a patient who underwent orthotopic liver transplantation, Italy, August–September 2009. *Euro Surveill* 14: pii: 19448.
- CRORA, 2005. *African Arbovirus Database*. Available at: <http://www.pasteur.fr/recherche/banques/CRORA/fixes/data.htm>.
- Pecorari M, Longo G, Gennari W, Grottola A, Sabbatini A, Tagliazucchi S, Savini G, Monaco F, Simone M, Lelli R, Rumpianesi F, 2009. First human case of Usutu virus neuroinvasive infection, Italy, August–September 2009. *Euro Surveill* 14: pii: 19446.
- Chvala-Mannsberger S, Bakonyi T, Brugger K, Nowotny N, Weissenbock H, 2007. Epizootiology of Usutu virus associated bird mortality in Austria. *Austrian Contributions to Veterinary Epidemiology*. Volume 4. Vienna: Institute for Veterinary Public Health.
- Hamer GL, Kitron UD, Brawn JD, Loss SR, Ruiz MO, Goldberg TL, Walker ED, 2008. *Culex pipiens* (Diptera: Culicidae): a bridge vector of West Nile virus to humans. *J Med Entomol* 45: 125–128.
- Tamba M, Bonilauri P, Bellini R, Calzolari M, Albieri A, Sambri V, Dottori M, Angelini P, 2010. Detection of Usutu virus within a West Nile virus surveillance program in northern Italy. *Vector Borne Zoonotic Dis* 11: 551–557.
- De Madrid AT, Porterfield JS, 1969. A simple micro-culture method for the study of group B arboviruses. *Bull World Health Organ* 40: 113–121.
- Reisen WK, Fang Y, Martinez VM, 2005. Avian host and mosquito (Diptera: Culicidae) vector competence determine the efficiency of West Nile and St. Louis encephalitis virus transmission. *J Med Entomol* 42: 367–375.
- Gerberg EJ, Barnard DR, Ward RA, 1994. Manual for mosquito rearing and experimental techniques. *Am Mosq Control Assoc Bull*: 61–62.
- Diallo M, Sall AA, Moncayo AC, Ba Y, Fernandez Z, Ortiz D, Coffey LL, Mathiot C, Tesh RB, Weaver SC, 2005. Potential role of sylvatic and domestic African mosquito species in dengue emergence. *Am J Trop Med Hyg* 73: 445–449.
- Digoutte JP, Calvo-Wilson MA, Mondo M, Traore-Lamizana M, Adam F, 1992. Continuous cell lines and immune ascitic fluid pools in arbovirus detection. *Res Virol* 143: 417–422.
- Fontenille D, Jupp PG, 1989. The presence of the *Culex (Culex) neavei* mosquito in Madagascar: its relevance in the transmission of arboviruses [in French]. *Arch Inst Pasteur Madagascar* 56: 287–295.
- Jupp PG, McIntosh BM, Blackburn NK, 1986. Experimental assessment of the vector competence of *Culex (Culex) neavei* Theobald with West Nile and Sindbis viruses in South Africa. *Trans R Soc Trop Med Hyg* 80: 226–230.
- Snow WF, 1973. The feeding habits of some west African *Culex* (Diptera: Culicidae) mosquitoes. *Bull Entomol Res* 62: 517–526.
- Snow WF, 1978. The feeding habits of some culicine mosquitoes (Diptera: Culicidae) in The Gambia. *Bull Entomol Res*: 68: 695–706.
- Lumsden WH, Someren EC, 1953. Records of *Culex* species (Diptera: Culicidae) from West Nile district, Uganda, with notes on their behavior. *Proc R Entomol Soc Lond*: 22: 19–22.
- Chandler JA, Highton RB, 1976. Studies on some ornithophilic mosquitoes (Diptera, Culicidae) of the Kano Plain, Kenya. *Bull Entomol Res* 66: 133–143.
- Richards SL, Mores CN, Lord CC, Tabachnick WJ, 2007. Impact of extrinsic incubation temperature and virus exposure on vector competence of *Culex pipiens quinquefasciatus* Say (Diptera: Culicidae) for West Nile virus. *Vector Borne Zoonotic Dis* 7: 629–636.
- Richards SL, Lord CC, Pesko K, Tabachnick WJ, 2009. Environmental and biological factors influencing *Culex pipiens quinquefasciatus* Say (Diptera: Culicidae) vector competence for Saint Louis encephalitis virus. *Am J Trop Med Hyg* 81: 264–272.
- Diallo D, Talla C, Ba Y, Dia I, Sall AA, Diallo M, 2011. Temporal distribution and spatial pattern of abundance of the Rift valley fever and West Nile fever vectors in Barkedji, Senegal. *J Vector Ecol* 36: 426–436.
- Chevalier V, Reynaud P, Lefrancois T, Durand B, Baillon F, Balanca G, Gaidet N, Mondet B, Lancelot R, 2009. Predicting West Nile virus seroprevalence in wild birds in Senegal. *Vector Borne Zoonotic Dis* 9: 589–596.
- Ba Y, Diallo D, Dia I, Diallo M, 2006. Feeding pattern of Rift Valley Fever virus vectors in Senegal. Implications in the disease epidemiology [in French]. *Bull Soc Pathol Exot* 99: 283–289.