Experimental evidence that parasites drive eco-evolutionary feedbacks

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Host resistance to parasites is a rapidly evolving trait that can influence how hosts modify ecosystems. Eco-evolutionary feedbacks may develop if the ecosystem effects of host resistance influence selection on subsequent host generations. In a mesocosm experiment, using a recently diverged (<100 generations) pair of lake and stream three-spined sticklebacks, we tested how experimental exposure to a common fish parasite (*Gyrodactylus* spp.) affects interactions between hosts and their ecosystems in two environmental conditions (low and high nutrients). In both environments, we found that stream sticklebacks were more resistant to *Gyrodactylus* and had different gene expression profiles than lake sticklebacks. This differential infection led to contrasting effects of sticklebacks on a broad range of ecosystem properties, including zooplankton community structure and nutrient cycling. These ecosystem modifications affected the survival, body condition, and gene expression profiles of a subsequent fish generation. In particular, lake juvenile fish suffered increased mortality in ecosystems previously modified by lake adults, while stream fish showed decreased body condition in stream-fish-modified-ecosystems. Parasites reinforced selection against lake juveniles in lake fish modified ecosystems, but only under oligotrophic conditions. Overall, our results highlight the overlapping timescales and the interplay of host-parasite and host-ecosystem interactions. We provide experimental evidence that parasites influence host-mediated effects on ecosystems, and thereby change the likelihood and strength of eco-evolutionary feedbacks.

eco-evolutionary dynamics | three-spined stickleback | host-parasite interaction | Gyrodactylus | eutrophication

Integrating ecosystem changes with rapid species adaptation is at the heart of modern evolutionary theory and an emerging eco-evolutionary synthesis (1–3). This crucially depends on understanding how phenotypic evolution can affect community structure and ecosystem functions (4). When the phenotypic effects of organisms on ecosystems are sufficiently large and persistent, an eco-evolutionary feedback may emerge if the organism-mediated environmental modifications become an important agent of selection that affects evolution of subsequent generations (1). While this perspective has recently received much attention (e.g. 5–8), very little is known about how interactions between organismal traits and biotic as well as abiotic drivers of ecosystem change govern the occurrence and strength of these feedbacks (9).

Parasites play key roles in ecosystems (10, 11) and evolutionary dynamics (12) because they are ubiquitous and can have strong effects on host fitness. Host-parasite interactions can evolve rapidly (12–15) and depend strongly on prevailing environmental conditions (16–18). As a result, host-parasite and host-ecosystem interactions may evolve in tandem, functionally linking evolutionary and ecological processes (19–21). For instance, variation in the composition of prey communities can be strongly modified by hosts, but it can also influence the exposure of hosts to trophically transmitted parasites (22). Feedbacks between host evolution and ecosystem dynamics may emerge when resistance evolves rapidly and influences the effects of hosts on ecosystems. Current eco-evolutionary theory recognizes that the presence and strength of feedbacks depend on a balance between the effects of both organisms and external environmental drivers on ecosystems (18). In freshwater ecosystems, nutrient loading by humans not only alters patterns of nutrient cycling (23, 24), but can also threaten population persistence (25) and disrupt ongoing species divergence by changing selection regimes (26). Furthermore, nutrient loading can increase parasite prevalence and change evolutionary trajectories of host-parasite interactions (27–29). Although the ecological and evolutionary effects of nutrient loading are well studied, very little is known about how it affects feedbacks between hosts, parasites and ecosystems.

To test for the combined effects of nutrient inputs and parasites on host-ecosystem feedbacks, we performed a two-phase mesocosm experiment where we manipulated the presence of parasites, the host ecotype, and the level of nutrient loading (Fig. 1). In phase 1, we tested whether wild-caught lake and stream sticklebacks differed in parasite resistance, gene expression profiles, metabolic condition, diet, and ecosystem effects. Because we used wild-caught fish, we did not distinguish between ecosystem modifications originating from either genetic effects or plasticity (6, 30). In phase 2, we removed the adult fish, and tested whether the ecosystem modifications by adult fish in phase 1 altered selection pressures (measured as differences in relative survival) on the next host generation. This next generation consisted of a juvenile population with equal proportions of lake, stream and hybrid juveniles (Fig. 1). Because these juveniles were reared in common-garden conditions, we could test for the effects of adult-

Significance

Anthropogenic effects on the environment are ubiquitous and have enormous impacts on individual and ecosystem health. It is widely accepted that environmental change affects disease distribution, but how it may affect parasite-driven evolution remains elusive. Our results provide experimental evidence that parasites play a major role in ecosystem dynamics, and, as a result, can affect selection in subsequent host generations. This role is further modified by the prevailing environmental conditions that affect disease dynamics in two ways: through altered ecological opportunities for disease and through altered evolutionary effects on the host.

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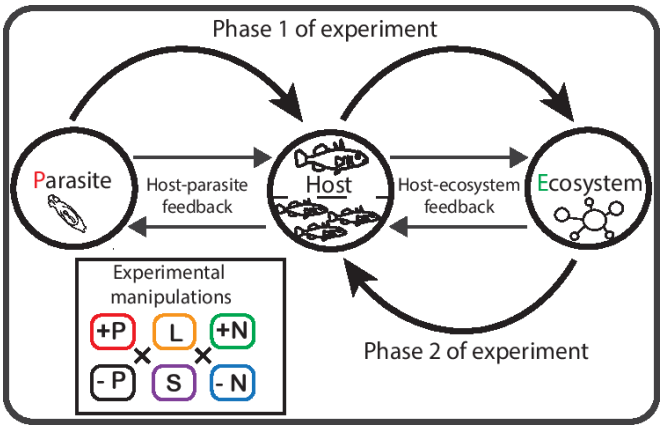

Fig. 1. Conceptual background and experimental design. During the first experimental phase, we investigated how host-parasite interactions affect surrounding ecosystems with different nutrient loadings. We characterized interactive effects of three experimental contrasts: parasite presence vs. absence (P: +P/-P), lake vs. stream host ecotype (H: L/S) and high vs. low ecosystem nutrients (E: +N/-N) throughout different biological levels. In phase 2, we tested for host-ecosystem feedbacks focusing on the next host generation and assessed selection against different host genetic backgrounds and gene expression of survivors.

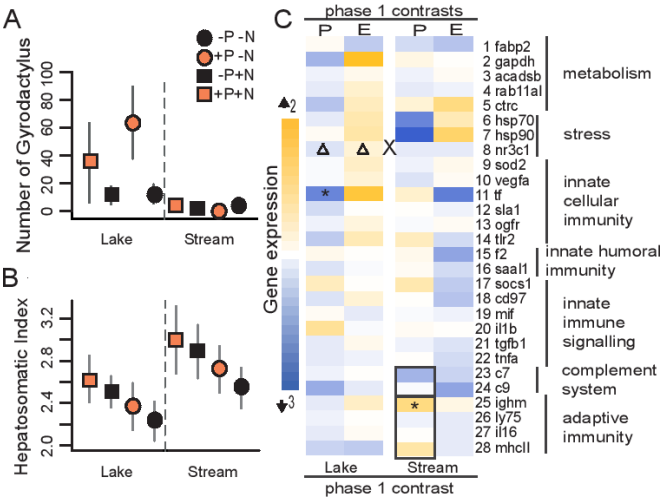

Fig. 2. Multi-level parasite and nutrient effects on sticklebacks in phase 1. Infection intensities with significant interaction of parasite exposure and host ecotype (PxH, N=159, *A*). Fish condition assessed by hepatosomatic index with effects of ecosystem nutrients (E) and infection intensity (i.i., N=159, *B*). Data is presented as means±SEM. Gene expression responses (*C*), from threefold down-regulation to twofold up-regulation in parasitized vs. control manipulations (P) and high vs. low nutrient levels (E). Significant expression changes for gene groups are highlighted by black outlines (lake: N=18, stream: N=20, test on tank averages), for single genes after Benjamini-Yekutieli correction for multiple testing (N=146, lake: N=66, stream: N=80, test on individuals) indicated by asterisks (first level effect), triangles (2way interaction) or X (3way interaction). See SI Appendix, Tables S1 & S3.

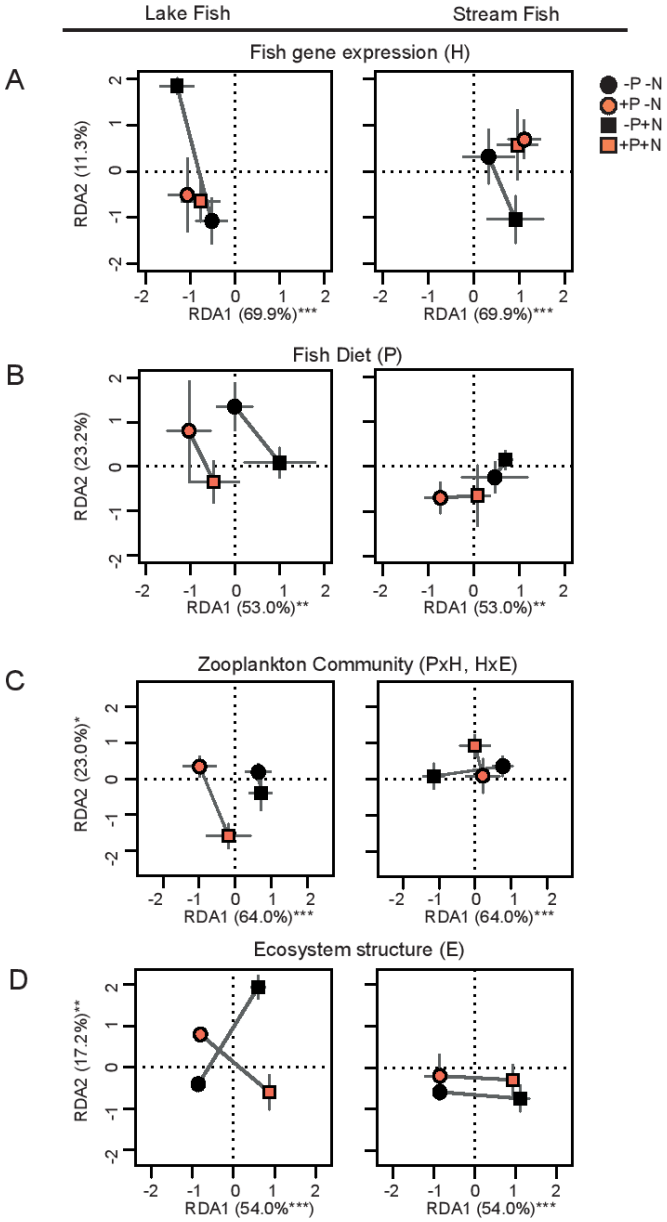

Fig. 3. Parasite effects from genes to ecosystem during phase 1 of the experiment. Gene expression (*A*),diet composition (*B*),zooplankton communities (*C*) and ecosystem parameters (*D*) are summarized by redundancy analyses (RDA, SI Appendix, Table S5a) and shown as experimental group means±SEM. Significant treatment effects for summarized data at each level are pointed out in Figure headers. Percentages are explained variance by RDA axes and asterisks indicate significance of RDA axes, * P<0.05, ** P<0.01, *** P<0.001

mediated ecosystem modifications, while controlling for rearing history and prior exposure to parasites.

Forty outdoor aquatic mesocosm ecosystems were set up with a mixture of sediments and invertebrates from multiple lakes and streams in Switzerland. We added nutrients only once before the start of the experiment to manipulate the productivity of these ecosystems (environmental contrast (E), high vs. low nutrients). We used recently diverged (<100 generations) ecotypes of lake and stream three-spined sticklebacks because these ecotypes (host contrast (H), lake vs. stream) are genetically differentiated (31, 32) and have different effects on mesocosm ecosystems (6). For phase 1 of the experiment, we manipulated parasite exposure of adults by disinfecting wild-caught fish and just prior to their introduction to the mesocosms, re-infecting half of the hosts with exactly four individuals of *Gyrodactylus* spp., a monogenean ectoparasite (parasite contrast (P), exposed vs. non-exposed). Each parasite-exposed fish received two individual parasites each from lake and stream origin to control for potential local (co)adaptation (33, 34). *Gyrodactylus* reproduces on the fish, is transmitted directly between fish hosts and can affect host condition and fitness (35). Each of the 8 factorial combinations of parasite exposure, host ecotype and nutrient level was replicated 5 times.

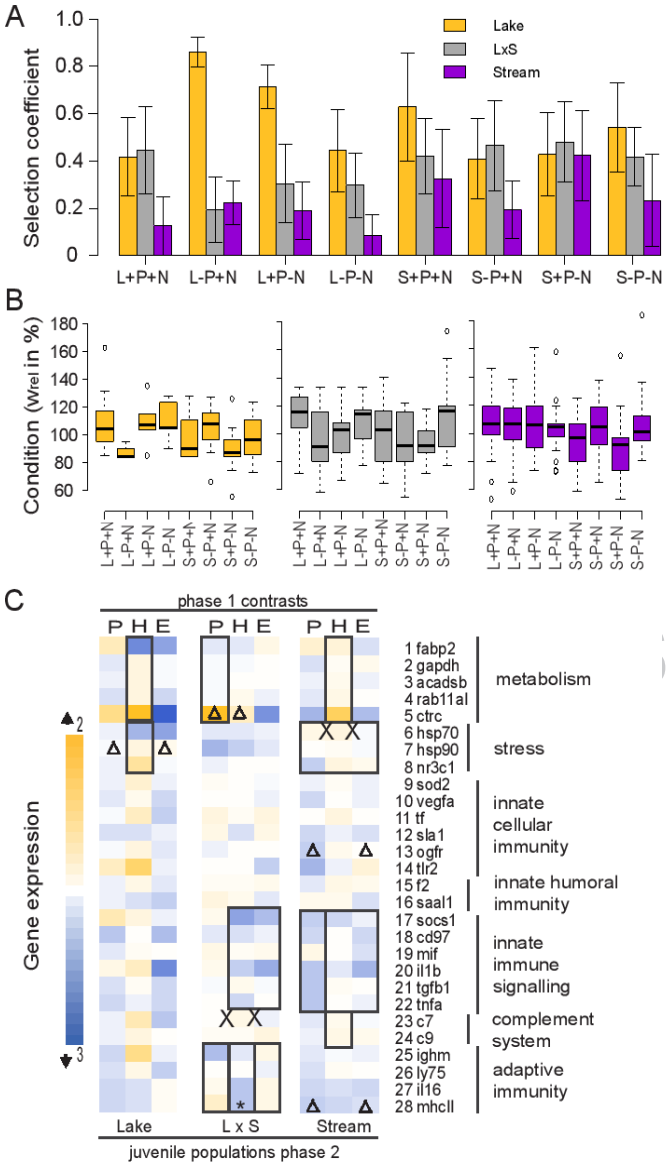

Fig. 4. Effects of ecosystem modifications on second phase fish. Selection coefficients (S = change in frequency relative to frequency of fittest genotype, subtracted from 1, within each tank (55))against different stickleback genetic backgrounds. Means±SEM across 5 replicated tanks in ecosystems modified by phase 1 manipulations are shown (*A*). Within lake fish, selection is shaped by an interaction of all previous ecosystem manipulations (PxHxE, N=39). The fittest genotype in each tank has a selection coefficient of 0 (Methods and SI Appendix, Table S7). Fish condition assessed by relative weight, showing the significant PxE effect on hybrid condition and PxH effect on stream fish condition (*B*, lake: N=73, hybrid: N=160, stream: N=184, SI Appendix, Table S7). Gene expression profiles of survivors summarized by experimental manipulation in phase 1 and for different ecotype backgrounds of the juvenile fish in phase 2 (*C*). Expression responses in parasitized vs. control tanks (P), fish introduced to the previous lake vs. stream tanks (H), and high vs. low nutrient boost tanks (E) from threefold down-regulation (blue) to twofold up-regulation (yellow). Significant regulatory changes for gene groups are highlighted by black outlines (lake: N=22, hybrid: N=32, stream: N=34, test on tank averages), for single genes after Benjamini-Yekutieli correction for multiple testing (lake: N=32, hybrid: N=79, stream: N=109, test on individuals) indicated by asterisks (first level effect), triangles (2way interaction) or X (3way interaction). See SI Appendix, Table S8.

After 7 weeks, we removed the adult fish and began phase 2 by adding juveniles to the same mesocosms that had been modified by the adults. These juvenile fish were bred by *in-vitro* fertilization using wild-caught parents and were reared on a common food source in the laboratory. Because these common-garden juveniles were not the offspring of the adults used during phase 1, we avoided possible confounding trans-generational priming effects of parasite resistance (36). Measuring variation in survival, body condition, and gene expression of these juveniles allowed us to test for an eco-evolutionary feedback by evaluating whether ecosystem modifications during phase 1 altered selection pressures during phase 2.

In order to confirm that the effects of ecotype and parasite exposure on gene expression were not solely due to plasticity (particularly in phase 1), we performed an additional common-garden experiment in the following year using lab-reared adult lake and stream fish from the same cohort as the second generation of the main experiment. To this end, we set up 12 identical outdoor tanks without sediment or zooplankton and exposed 17 lab-raised adult sticklebacks, in 6 groups of 2-3 individuals, to *Gyrodactylus* while another 17 served as control, unexposed fish (Methods and SI Appendix, Figs. S1&S2, Table S4).

Results and Discussion

At the end of phase 1 (7 weeks duration), stream fish carried fewer individual parasites than lake fish (infection intensities, defined as Σ*Gyrodactylus*/Σexposed fish (infected+non-infected): $i.i._{L+P}$= 49.8±19.1, $i.i._{S+P}$=2.67±0.85, Fig. 2A, PxH effect, SI Appendix, Table S1, infection prevalence: $prev_{L+P}$=63.0%, $prev_{S+P}$=49.5%). We observed similar infection intensity and prevalence patterns in the wild; lake fish being infected with higher numbers of *Gyrodactylus* than stream fish ($i.i._{Lwild}$= 30.4±5.23, $i.i._{Swild}$=4.68±1.75, N=40, H effect: χ^2=30.22, p<0.001, GLMM), and showing comparable infection prevalence ($prev_{Lakewild}$=57.1%, $prev_{Streamwild}$=63.2%). Even though parasites were also present at very low levels in control mesocosms, experimentally exposed fish showed significantly higher infection intensities ($i.i._{+P}$= 26.5±9.88, $i.i._{-P}$=7.2±1.9, Fig. 2A). *Gyrodactylus* numbers were highest on lake fish in ecosystems with low nutrient loading ($i.i._{Lake+P+N}$ = 35.4 ±28.4, $i.i._{Lake+P-N}$ = 64.2 ±25.7; χ^2=7.470, p=0.006, Fig. 2A), suggesting that productive environments allow the less resistant fish ecotype to compensate and reduce costs of parasitism.

To characterize the molecular phenotypes of differential parasite load between fish ecotypes, we quantified expression of 28 metabolic, immune and stress response genes. We selected i) genes from a previous transcriptomic study based on strong differential expression between fish ecotypes as well as between infection states (37) and ii) genes associated with responses to *Gyrodactylus* in other fish species (see SI Appendix, Table S2 for gene specific references). In phase 1, *Gyrodactylus* exposure of adults differently affected gene expression profiles of the two stickleback ecotypes (Fig. 2C, PxH and PxHxE effects, SI Appendix, Table S3): stream fish up-regulated genes of the adaptive immune system (P effect, p=0.004) and down-regulated genes of the complement system (P effect, p=0.024, perMANOVAs). By contrast, lake fish did not modify the expression of entire gene groups, but significantly down-regulated two genes: the antibacterial *transferrin a* and a glucocorticoid receptor involved in the general stress response (*tf*, P effect, p=0.008; *nr3c1*, PxE effect, p=0.002, LMMs). The differential gene expression profiles and infection patterns indicate that stream fish have evolved stronger immune responses against this parasite, enabling them to limit infection better than lake fish. This could potentially be achieved via mechanisms involving recognition of *Gyrodactylus* antigens by immune cell receptors (38). The observed contrasting immune gene expression responses and strong expression differences between the ecotypes (H effects throughout most genes, SI Appendix, Table S3) support the hypothesis that parasite-mediated selection between habitat types contributes to adaptive popula-

tion divergence of lake and stream ecotypes (39) and corroborate the strong immune gene expression differences between wild lake and stream sticklebacks reported in a recent study (40).

Overall, we found no persistent effects of nutrient loading on gene expression profiles of sticklebacks in phase 1 (SI Appendix, Table S3). However, we found that a stress response gene (*nr3c1*) encoding a glucocorticoid receptor, which initiates stress responses upon cortisol binding, was indirectly affected through an interaction of parasite exposure, nutrient loading and host ecotype (PxHxE effect, p=0.006, LMMs). This effect was driven by up-regulation in response to parasite pressure and down-regulation in high nutrient environments in lake fish, further highlighting how the tight interaction of biotic and abiotic selection pressures can lead to population specific patterns of gene expression.

To test whether the ecotype effects of molecular phenotypes were due to genetic differences, rather than due to differences in history of infection in the wild, we performed an additional common-garden experiment where we quantified gene expression of lab-reared adults originating from the same laboratory populations of juveniles used for phase 2. Using the same 28 genes, we found that gene expression generally differed between ecotypes (perMANOVA, H effect: $F_{1,8}=3.859$, p=0.041, SI Appendix, Table S4a). Furthermore, metabolism genes showed an ecotype-specific expression response to the parasite exposure (PxH effect: $F_{1,8}=11.20$, p=0.041, Fig. S1). These expression differences between ecotypes, as well as expression responses to *Gyrodactylus* in stream fish, were conserved between experiments (SI Appendix, Fig. S2). This demonstrates that genetic differences between the lake and stream stickleback ecotypes (32) consistently influence their molecular phenotype, and that the effects observed during phase 1 of the mesocosm experiment are likely due to both genetic differences and plasticity. The importance of metabolism genes for ecotype differences in the response to parasite exposure is also consistent with a previous study, despite the analysis of different immune organs (37).

In phase 1 of the mesocosm experiment, we found a cost of parasitism such that neither host ecotype was completely tolerant to *Gyrodactylus* (41), indicated by a decrease of the hepatosomatic index (HSI) (42) with infection intensity in both ecotypes (infection intensity effect, p=0.050, SI Appendix, Table S1, Fig. 3a in (43)). In addition, parasite exposure caused fish to feed on different prey (P effect, $R^2=0.064$, p=0.020, diet composition RDA, Fig. 3B, SI Appendix, Table S5a). Specifically, parasite-exposed individuals ate more cyclopoid copepods and fewer nymphs than control fish (SI Appendix, Table S5b, (43)). Such a diet shift could be caused either by direct parasite-mediated effects on feeding performance (44) or by changes in host feeding behavior in order to meet the nutritional requirements for coping with parasite infection (45).

Given that parasites had effects on both the condition and diet of lake and stream sticklebacks, we hypothesized that parasite exposure might further influence how sticklebacks modify other aspects of their ecosystems. We found that the composition of the zooplankton community in the mesocosms was best predicted by the interaction between the fish ecotypes and the presence of *Gyrodactylus* (PxH effect, $R^2=0.067$, p=0.028, RDA, Fig. 3C, SI Appendix, Table S5a). This effect might have been mediated by a differential top-down trophic effect of the stickleback ecotypes on the abundance of copepods in different nutrient and parasite environments (PxHxE effect, p=0.042, SI Appendix, Table S5b). Further down the food chain, the abundance of rotifers (Lepadellidae), which are a common prey of copepods, was also significantly affected by differences in how stickleback ecotypes responded to parasite exposure (i.e. a PxH effect, p=0.017, SI Appendix, Table S5b). Interestingly, interactive effects of hosts and parasites were also evident for abiotic ecosystem conditions. For example, despite the strong effects of our initial nutrient manipulation on the mesocosm ecosystems (i.e. E effects are common, SI Appendix, Table S5), the exposure of sticklebacks to parasites significantly altered the distribution of nutrients (e.g. dissolved nutrients, total nutrients, DOC) within the mesocosm ecosystems (PxE effect, $R^2=0.048$, p=0.019, nutrient concentration RDA, SI Appendix, Table S5a). A previous mesocosm experiment using these same ecotypes of sticklebacks, found that both genetic background and plasticity interactively affected prey community structure and ecosystem conditions (6). While both experiments found significant ecotype effects on a wide range of ecosystem metrics, the specific outcomes and dynamics differ between experiments. In both experiments, adult lake fish decreased copepod abundance more than stream fish in the short term (i.e. 3-7 weeks). In the previous experiment, however, this effect was reversed after 12 weeks (6). In general, mesocosms are only an approximation of natural ecosystems, and so the extent to which those effects are visible in nature remains unknown. In our experimental ecosystems, results suggest far-reaching consequences of parasitism (P effects) and host-parasite interactions (PxH effects) that extend well beyond the direct effects on host immunity, condition and diet. In phase 2 of our experiment, we tested whether such ecosystem effects alter selection regimes in the next host generation.

To initiate phase 2, we introduced lake, stream and hybrid juvenile fish (lab-bred F1) into the tanks previously modified by the adult fish. At the end of phase 2 (13 weeks duration), juvenile fish were collected and genotyped to quantify variation in survival depending on lake, hybrid, and stream fish origin (SI Appendix, Table S6). Overall, lake juveniles had a lower survival rate than either stream or hybrid juveniles ($\chi^2=67.56$, p<0.001, Pearson's χ^2-test, SI Appendix, Fig. S3). Selection against lake juveniles was linked to a three-way interaction between treatment combinations in the first phase, namely parasite exposure of adults, host ecotype, and initial nutrient additions (PxHxE effect, p=0.013, Fig. 4A, SI Appendix, Table S7). More specifically, selection against lake juveniles was higher in ecosystems previously manipulated by lake adults, particularly when these adults were either exposed to parasites in low nutrient mesocosms or unexposed to the parasite in high nutrient mesocosms. By comparison, the selection against stream and hybrid juveniles did not vary with the adult treatments in phase 1 (SI Appendix, Table S7). Among survivors however, stream juveniles had a lower body condition in ecosystems modified by parasite-exposed stream fish (PxH effect, p=0.031, Fig. 4B, SI Appendix, Table S7). Together, the observed variation in survival rate and body condition show that both lake and stream ecotypes either have a survival disadvantage (lake juveniles) or a lower condition (stream juveniles) in ecosystems manipulated by adults of the same ecotype. Such effects could be due to differential depletion of preferred prey items, in particular by adults under parasite pressure. It is also possible that parasites persisted in the mesocosms in phase 2 and had differential effects on the juvenile genotypes (see Supplementary Discussion for further extrapolations from three-way interactions).

The body condition of hybrid juveniles was unaffected by the adult ecotype, but they had a lower condition in mesocosms where adult fish had been exposed to parasites at low nutrient loading and in parasite control tanks at high nutrient loading (PxE effect, p=0.001, Fig. 4B, SI Appendix, Table S7). The dependence of hybrid juvenile condition on the interaction between parasite exposure and nutrient loading during phase 1 suggests that parasites might mediate selection against hybrids via changes in the ecosystems. Variation in the strength of selection against hybrids can influence the persistence of local adaptation, and influence the likelihood of biodiversity loss via reverse speciation (26, 46). For sticklebacks, parasite-mediated selection against hybrids has been both suggested (39) and experimentally demonstrated

(38), however independently of the ecosystem effects of sticklebacks. Our experiment suggests a previously unexplored cross-generational effect of parasites, whereby parasites influence how hosts modify their ecosystems, altering selection on a subsequent generation (Figs. 3&4). Further experiments could test whether such an effect might be even stronger in a natural environment, where multiple generations of juvenile and adult stickleback co-occur (47). Our results also illustrate a potential mechanism underlying eco-evolutionary feedbacks, namely one where host-mediated ecosystem modifications affect selection and relative fitness of a subsequent host generation.

In addition to the effects of ecosystem modifications in phase 1 on relative juvenile survival in phase 2, we found effects on the expression of metabolism genes, general stress response and innate immune signaling across juvenile ecotypes in phase 2 (Fig. 4C). In the modified ecosystems, the innate immune signaling of hybrid and stream juveniles showed an overall lower expression of genes in the high nutrient environments established in phase 1 (HxE effects, Fig. 4C, SI Appendix, Table S8a). This suggests that high nutrient environments shift either the cues or the trade-offs for investments in immune signaling by different host ecotypes. Additionally, stream juveniles exhibited differential regulation of the *mhcII* gene based on the parasite and nutrient treatments of phase 1 (PxE effect, p=0.004, Fig. 4C, SI Appendix, Table S8b). Major histocompatibility complex (MHC) class II genes are part of the adaptive immune system. They are involved in antigen recognition and specific MHC alleles are correlated with *Gyrodactylus* resistance (38). If stream fish have previously evolved under high prevalence of this parasite or in the presence of very virulent parasite strains, altering the baseline expression level of *mhcII* might be an adaptive response to reduce parasite spread and explain the selective advantage of this ecotype (Fig. 4A). The cross-generational effects of our parasite manipulation could also have been caused by the persistence of parasites in the mecocosms after the adults were removed. In this case, the regulatory response of juveniles may reflect the stronger parasite resistance of stream sticklebacks. In natural populations, the translation of parasite effects across generations, mediated by host-modified ecosystems, might be combined with transgenerational immune priming when hosts inherit epigenetic signals of their parents' previous infections (36, 48). However, we can rule out this possibility in our experiment because juveniles were not the direct offspring of phase 1 adults. Instead, the cross-generational effects we observed were solely mediated by how the presence and infection status of hosts affected the subsequent rearing environment of juveniles.

Overall, our results show that the presence of parasites and the evolution of differential parasite resistance can influence host performance (e.g. diet, and condition), and this can have cascading effects on community structure and ecosystem function. Variation in both parasite resistance and external environmental conditions can mediate the strength of eco-evolutionary feedbacks, and this can be detected at the level of molecular phenotypes and ecosystem characteristics . That host-mediated modifications of the environment caused transgenerational effects on molecular phenotypes and differential selection among ecotypes, warrants reconsidering the nature and importance of soft selection (9) and suggests that eco-evolutionary feedbacks might play an underappreciated role in adaptation. In light of our results, the effects of environmental change on infectious disease and on adaptive population divergence (26, 49) are more closely linked than previously considered.

Materials and Methods

Animal collection and treatment of phase 1 fish. We collected three-spined sticklebacks (*Gasterosteus aculeatus*) with hand nets from two stream sites in the canton of St. Gallen, Switzerland (47.321131N, 09.562395E and 47.355822N, 09.603133E) and with minnow traps at one location on the shore of Lake Constance (47.484830N, 09.542923E). Fish collection and experiments were approved by local authorities (canton of St. Gallen fishing authorities and Veterinäramt of Kanton Luzern under permit LU03/12EE). Twenty sticklebacks each of stream and lake origin were euthanized directly to assess *Gyrodactylus* spp. prevalence in the natural populations. All experimental fish were disinfected by baths in 1:4000 diluted Formalin on three consecutive days (modified from(33)). Experimental infection was achieved 7 days later by manual transfer of *Gyrodactylus* spp. individuals from non-disinfected sticklebacks collected from the same lake and stream populations. Two individual parasites from each of the lake and stream environments were transferred. Additional details are available in SI Appendix, Section SI.1.

Experimental Setup and first phase sampling. The mesocosms were plastic tanks of one cubic meter, filled with gravel, sand, sediment collected from Lake Lucerne and a nearby stream, lake water and a concentrated zooplankton inoculum from Lake Lucerne and Lake Constance. The full factorial cross design of Parasites x Host ecotype x Ecosystem Nutrients was replicated in 5 blocks for a total of 40 mesocosms. Within each block, we established contrasting nutrient environments by adding different amounts of nutrient solution containing $NaNO_3$ and HNa_2PO_4 into high and low nutrient tanks respectively (E contrast). For the first phase of the experiment, we introduced three-spined sticklebacks of either lake or stream origin to establish the host ecotype contrast (H).

We collected ecosystem data such as physico-chemical (e.g. turbidity, nutrient concentrations) as well as biological (e.g. chlorophyll levels in water and periphyton) properties of the ecosystems and sampled the zooplankton communities 6 weeks after fish introduction to the mesocosms and removed the fish one week later. Fifty-seven out of 278 sticklebacks died during the first experimental phase and were collected from the mesocosms upon detection. Mortality differed between host ecotypes, being higher among lake fish, but did not vary with other treatments (χ^2 test, H: χ^2=4.164, p=0.041, P: χ^2=0.233, p=0.629, E: χ^2=0.002, p=0.966, SI Appendix, Table S1). After euthanasia of the fish in 1M MS-222, *Gyrodactylus* specimen were counted on each fish before morphological measurements and dissection. Additional details are available in SI Appendix, Section SI.2.

Introduction and sampling of phase 2 fish. After removal of phase 1 fish, groups of juvenile lab-bred F1 sticklebacks of lake, hybrid and stream background were introduced into each tank modified throughout phase 1 of the experiment. These juvenile groups were standardized for family backgrounds within experimental blocks and ratio of stream, hybrid and lake fish across all experimental tanks (N=19-39/tank; SI Appendix, Table S6). Hybrid crosses were done in either direction, 7 with stream females, 5 with lake females. Ecosystems were all handled equally at this stage. All surviving fish were caught three months after the juvenile phase 2 fish were introduced to the mesocosms. As with phase 1 fish, after euthanasia in a 1M MS-222 solution, *Gyrodactylus* specimen were counted on each fish before length and weight measurements and removal of spleens and livers for gene expression assays. Only 10 of the 407 scanned individuals were infected with *Gyrodactylus* at the end of the experiment, with no significant effects of any previous treatment on infection levels in this second generation (binomial GLMMs, all χ^2<2.03, all P>0.15, SI Appendix, Table S7). Additional details are available in SI Appendix, Section SI.3.

Common garden experiment. To validate that part of the ecotype effect during phase 1 was based on genetic differences between lake and stream sticklebacks, we conducted a separate common garden experiment. This experiment ran for 5 weeks and consisted in 34 lab-raised adult fish kept in 12 identical outdoor tanks. Half of these fish had a genetic lake background and the other half descended from stream fish. Again, half of the experimental groups were exposed to *Gyrodactylus* on an individual basis. Gene expression data was collected from their spleens as a comparison to the wild-caught fish from the first phase of the mesocosm experiment. Additional details are available in SI appendix, Section SI.4.

Molecular analyses. We performed gene expression analyses with RNA extracted from spleens and combined spleens and livers for adults and juveniles, respectively. Because transcriptome analyses have been conducted with lake and stream three-spined sticklebacks (37), we used a target gene approach, measuring relative mRNA levels in microfluidic qPCR assays of 28 target genes. Origin of surviving juveniles was determined by parentage analysis in Colony (50), using 7 microsatellite markers (51) (Stich5196, Stich4170, Stich1125, Stich1097, Stich7033, STN18, STN75). Additional details are available in SI Appendix, Section SI.5.

Statistical analyses. All statistical analyses were performed in R version 3.1.0 (52). The following model structure was used to test for the effects of phase 1 experimental treatments: parasite exposure (P), host ecotype (H), ecosystem nutrient levels (E) and their interactions as fixed structure with block as a random factor. Univariate analyses on individual fish characteristics such as parasite burden, fish condition, gene expression and survival also included tank identity nested within block as a random effect. Fish condition for phase 1 fish was calculated as the hepatosomatic index (HSI) = 1000 x liver wet-mass (mg)/fish mass (mg) and for phase 2 fish as relative weight W_{rel} (53). HSI was tested with an LMM using infection intensity as well as the experimental treatments as fixed structure.

Diet, zooplankton communities, ecosystem parameters and gene expression were tested for experimental treatment effects in RDAs and univariate (G)LMMs. Gene expression was analyzed as ΔCt values (54) and further assessed by perMANOVAs on functional gene groups. Juvenile stocking

differences between tanks (19-39/tank) were statistically accounted for by including tank as a random factor in individual based tests and by including stocking numbers in tank based tests for phase 2 analyses. Survival differences between lake, hybrid and stream juveniles were tested with a Pearson's χ^2-test. Effects of phase 1 treatments on juvenile survival were tested in binomial GLMMs on survival rates from each tank. We calculated the selection coefficient S against each juvenile ecotype as the change in frequency of the ecotype relative to the frequency of the fittest genotype, subtracted from 1, within each tank (55). Effects of phase 1 ecosystem modifications on viability selection were tested in LMMs for each juvenile ecotype separately. Additional details are available in SI Appendix, Section SI.6.

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