

Hydrogeological controls on regional-scale indirect nitrous oxide (N₂O) emission factors for rivers

Richard James Cooper, Sarah Katrina Wexler, Christopher Adams, and Kevin M. Hiscock

Environ. Sci. Technol., **Just Accepted Manuscript** • DOI: 10.1021/acs.est.7b02135 • Publication Date (Web): 25 Aug 2017

Downloaded from <http://pubs.acs.org> on August 31, 2017

Just Accepted

“Just Accepted” manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides “Just Accepted” as a free service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. “Just Accepted” manuscripts appear in full in PDF format accompanied by an HTML abstract. “Just Accepted” manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are accessible to all readers and citable by the Digital Object Identifier (DOI®). “Just Accepted” is an optional service offered to authors. Therefore, the “Just Accepted” Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the “Just Accepted” Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these “Just Accepted” manuscripts.

1 Hydrogeological controls on regional-scale indirect nitrous oxide 2 (N_2O) emission factors for rivers

3 Richard J. Cooper*, Sarah K. Wexler, Christopher A. Adams, Kevin M. Hiscock

4 *School of Environmental Sciences, University of East Anglia, Norwich Research Park, Norwich, NR4 7TJ, UK*

5 *Correspondence: Richard.J.Cooper@uea.ac.uk

6

7 Abstract

8 Indirect nitrous oxide (N_2O) emissions from rivers are currently derived using poorly constrained
9 default IPCC emission factors (EF_{5r}) which yield unreliable flux estimates. Here, we demonstrate how
10 hydrogeological conditions can be used to develop more refined regional-scale EF_{5r} estimates required
11 for compiling accurate national greenhouse gas inventories. Focusing on three UK river catchments
12 with contrasting bedrock and superficial geologies, N_2O and nitrate (NO_3^-) concentrations were
13 analyzed in 651 river water samples collected from 2011 to 2013. Unconfined Cretaceous Chalk
14 bedrock regions yielded the highest median N_2O -N concentration ($3.0 \mu\text{g L}^{-1}$), EF_{5r} (0.00036) and
15 N_2O -N flux ($10.8 \text{ kg ha}^{-1} \text{ a}^{-1}$). Conversely, regions of bedrock confined by glacial deposits yielded
16 significantly lower median N_2O -N concentration ($0.8 \mu\text{g L}^{-1}$), EF_{5r} (0.00016) and N_2O -N flux (2.6 kg
17 $\text{ha}^{-1} \text{ a}^{-1}$), regardless of bedrock type. Bedrock permeability is an important control in regions where
18 groundwater is unconfined, with a high N_2O yield from high permeability Chalk contrasting with
19 significantly lower median N_2O -N concentration ($0.7 \mu\text{g L}^{-1}$), EF_{5r} (0.00020) and N_2O -N flux (2.0 kg
20 $\text{ha}^{-1} \text{ a}^{-1}$) on lower permeability unconfined Jurassic mudstone. The evidence presented here
21 demonstrates EF_{5r} can be differentiated by hydrogeological conditions and thus provide a valuable
22 proxy for generating improved regional-scale N_2O emission estimates.

23 **Keywords:** Denitrification; streams; climate change; greenhouse gas; IPCC; agriculture

24 1. Introduction

25 Nitrous oxide (N_2O) is a powerful greenhouse gas with a global warming potential 265 times greater
26 than carbon dioxide (CO_2) over a 100-year timescale.¹ At a current atmospheric concentration of 329
27 ppb,² N_2O is the third most important well-mixed greenhouse gas behind CO_2 and methane (CH_4),
28 accounting for 6% of total anthropogenic radiative forcing (0.17 W m^{-2}).^{1, 3} N_2O is also the single
29 most dominant stratospheric ozone (O_3) depleting substance emitted in the 21st century through its
30 role in the catalytic reduction of O_3 to oxygen (O_2).^{1, 4} Importantly, concentrations are estimated to

31 have increased by 22% since 1750 (270 ppb)⁵ and have been growing at an annual rate of 0.75 ppb
32 since the late 1970s.³

33 N₂O is produced as a byproduct of bacterially-driven aerobic nitrification in soils, sediments and
34 waterbodies during the oxidation of ammonium (NH₄⁺) to nitrate (NO₃⁻), by predominantly
35 autotrophic *Nitrosomonas* and *Nitrobacter* sp.⁶⁻⁸ N₂O also forms as an obligate intermediate product
36 of denitrification under low oxygen conditions through the bacterial reduction of NO₃⁻ to nitrogen gas
37 (N₂).⁷⁻⁹ Furthermore, in oxygen-deficient environments, NH₄⁺ can be oxidized to nitrite (NO₂⁻) and
38 then reduced to nitric oxide (NO), N₂O and N₂ via nitrifier denitrification.¹⁰

39 Current estimates of the total global flux of N₂O into the atmosphere as a result of nitrogen (N)
40 cycling are ~18.8 Tg N a⁻¹, of which ~10.5 Tg N a⁻¹ (55%) originate in natural sources.¹¹ The
41 remaining 45% of emissions are derived from anthropogenic sources (~8.3 Tg N a⁻¹) as a result of
42 perturbations to the N cycle.¹¹ Agriculture represents the largest anthropogenic source (5.3 – 8.0 Tg N
43 a⁻¹) and can be divided into direct emissions from soils (1.8 – 2.1 Tg N a⁻¹), animal production (2.1 –
44 2.3 Tg N a⁻¹) and indirect emissions (1.3 – 2.6 Tg N a⁻¹).¹¹⁻¹⁴ Whilst direct soil emissions have been
45 extensively studied,^{6, 15-20} indirect emissions arising from atmospheric deposition (~0.3 – 0.4 Tg N a⁻¹)
46 human sewage (~0.2 – 0.3 Tg N a⁻¹) and N leaching and runoff (~0.6 – 1.9 Tg N a⁻¹) are less well
47 constrained and remain a major source of uncertainty in the global N₂O budget.²¹⁻²⁹

48 The Intergovernmental Panel on Climate Change (IPCC) uses emission factors to estimate indirect
49 N₂O emissions from waterbodies arising from N leaching and runoff (EF₅).^{13, 30} These are based either
50 on the fraction (Frac_{LEACH}) of the original total fertilizer N input into the system that is lost to
51 waterbodies as a result of leaching and runoff from agricultural soils (eq. 1), or simply the ratio of
52 dissolved N₂O to dissolved inorganic nitrogen (DIN) within the waterbody (eq. 2):

$$53 \quad (1) \quad EF_5 = \frac{N_2O-N}{(total \ N \ input \times \ Frac_{LEACH})}$$

54 or

$$55 \quad (2) \quad EF_5 = \frac{N_2O-N}{NO_3-N}$$

56 The IPCC divides EF₅ into three components based on the site of N₂O production in either
57 groundwater (EF_{5g}), rivers (EF_{5r}) or estuaries (EF_{5e}). Since 2006, each component has been assigned a
58 default value of 0.0025 (i.e. 2.5 g of N₂O-N emitted per kg of N in leachate/runoff), thus giving a
59 combined EF₅ of 0.0075.³⁰ However, these default ‘Tier 1’ emission factors are poorly constrained
60 due to a paucity of studies, highly uncertain water-air gaseous exchange relationships, and large
61 variability in environmental conditions.²⁹ Thus, EF₅ has a wide range of uncertainty (0.0005 – 0.025)
62 and has been broadly criticized for either over^{14, 31, 32} or under^{9, 29} estimating actual N₂O fluxes.

63 In order to produce more accurate emission estimates, refined ‘Tier 3’ EF₅ emission factors need to be
64 derived which reflect regional variability in climate, soil type, geology, hydrochemistry, river
65 morphology and land management.^{14, 33} In this research, we investigated the impact of
66 hydrogeological conditions upon riverine N₂O emissions as a way of generating improved ‘regional-
67 scale’ EF_{5r} estimates. Focusing on three UK river catchments with contrasting bedrock (chalk,
68 limestone, sandstone, mudstone, volcanic) and superficial (glacial till, glacial sands/gravels, absent)
69 geologies, we explored whether hydrogeological conditions (high/low permeability,
70 confined/unconfined groundwater) exerted a sufficiently robust control over EF_{5r} that it could be used
71 as a proxy for upscaling N₂O emission estimates that are required for producing national greenhouse
72 gas inventories. We hypothesized such an association could arise due to hydrogeological conditions
73 controlling the infiltration and upwelling of water and dissolved N fertilizers in catchments, which in
74 turn impacts upon the formation and movement of dissolved N₂O gas. It is envisaged the outcomes of
75 this research will provide useful evidence for updating indirect N₂O emission factors used in future
76 IPCC assessment reports.

77 **2. Materials and Methods**

78 **2.1 Study Locations**

79 The three river catchments (Avon, Eden and Wensum) and sampling locations investigated in this
80 study were selected to align with the counterpart UK government-funded Demonstration Test
81 Catchments (DTC) program which is evaluating the extent to which on-farm mitigation measures can
82 cost-effectively reduce the impact of agricultural pollution on river ecology.³⁴

83 *2.1.1 River Wensum*

84 The River Wensum, Norfolk, is a 78 km length groundwater-dominated lowland (source = 75 m
85 above sea level (a.s.l.)) calcareous river that drains an area of 660 km² and has a mean annual
86 discharge of 4.1 m³ s⁻¹ near its outlet³⁵ (Figure 1; hydrological summaries provided in Figure S1 and
87 Table S1). The catchment is underlain by Cretaceous White Chalk bedrock which is unconfined in the
88 upper catchment and along sections of the river valley where the baseflow index (BFI) is ~0.7–0.9.
89 Over much of the rest of the catchment, the Chalk is confined by superficial deposits of Mid-
90 Pleistocene diamicton glacial tills principally comprising chalky, flint-rich boulder clays of the
91 Sheringham Cliffs (0.2–0.5 m depth) and Lowestoft (0.2–20 m depth) Formations. These are
92 interspersed with layers of glaciofluvial and glaciolacustrine sands and gravels where the BFI is ~0.5–
93 0.7. Within the river valley, Holocene-age alluvium and river terrace deposits are present.³⁶ Surface
94 soils across the catchment range from low permeability clay loams and sandy peats, to free draining
95 sandy loams. Arable agriculture (wheat, barley, sugar beet, oilseed rape) dominates land use (63%)
96 with the remainder comprising 19% improved grassland, 9% mixed woodland, 5% unimproved

97 grassland and 4% urban. The mean annual temperature is 10.1°C and the mean annual rainfall total is
98 674 mm (1981-2010).³⁷

99 2.1.2 River Eden

100 The River Eden, Cumbria, is a 145 km length surface runoff-dominated upland (source = 675 m a.s.l.)
101 river draining 2288 km² with a mean annual discharge of 53.4 m³ s⁻¹ near its outlet.³⁵ The catchment
102 bedrock comprises a mixture of Permo-Triassic sandstone, lower Palaeozoic igneous formations
103 (Borrowdale Volcanics) and steeply dipping fractured Carboniferous limestone interbedded with
104 sandstone and mudstone units (Figure 1). Quaternary glacial till confines the majority of the bedrock,
105 varying in thickness from 0–30 m across the catchment, whilst alluvium is present in the river valley.
106 The BFI in these confined areas is 0.3–0.5. Soils are mainly sandy clay loam and clay loam; locally
107 deep and well-drained in the headwaters, seasonally wet in the central elevations, moving through to
108 slowly permeable and seasonally waterlogged in lower parts of the catchment. Livestock farming
109 (sheep and dairy) dominates land use, with approximately 50% of land under improved pasture, 20%
110 rough grazing, 16% arable and 8% mixed woodland. The mean annual temperature is 9.4°C and the
111 mean annual precipitation total is 1197 mm (1981-2010).³⁷

112 2.1.3 River Avon

113 The River Avon, Hampshire, is a 96 km length groundwater-dominated, lowland (source = 120 m
114 a.s.l.) river draining 1717 km² with a mean annual discharge of 20.3 m³ s⁻¹ near its outlet.³⁵
115 Approximately 85% of the main river flow is supplied by the underlying Cretaceous White Chalk and
116 Upper Greensand bedrock aquifers (BFI = 0.75–0.95). The catchment also contains extensive
117 expanses of low permeability Jurassic Kimmeridge Clay mudstone (BFI = 0.2–0.5). The Chalk and
118 mudstone are largely unconfined across the catchment, but are locally covered by pockets of
119 Quaternary alluvium, head and river terrace deposits. Free-draining, shallow, lime-rich soils overlay
120 the Chalk across much of the catchment, alongside smaller areas of low permeability base-rich clay
121 loam. The catchment has a mixed farming system, with 48% of land under arable cultivation and 32%
122 in grassland for lowland grazing and intensive dairy production. The mean annual temperature is
123 10.1°C and the mean annual rainfall total is 857 mm (1981-2010).³⁷

124 2.2 Sample Collection

125 For the River Wensum, samples were collected from 20 sites across the catchment at approximately
126 monthly intervals between February 2011 and May 2013, such that 26 samples were collected from
127 each site and 520 samples were collected in total (Table 1). Of the 20 sites, 16 were tributary streams
128 <10 m width, of which 12 were from sites where the Chalk is confined by glacial deposits ($n = 312$)
129 and four sites where the Chalk is largely unconfined ($n = 104$). A further four sites were located on the
130 main channel of the River Wensum (>10 m width) and drained an upstream area encompassing both

131 confined and unconfined Chalk; these samples are henceforth referred to as ‘semi-confined’ Chalk (n
132 = 104). The higher sampling resolution for the River Wensum enabled temporal variability in N_2O
133 dynamics to be assessed for this catchment.

134 For the River Avon, samples were collected from six headwater tributaries on four separate occasions
135 (February, June and October 2012, March 2013). 2-3 replicates were collected from each tributary on
136 each sampling occasion, giving 56 samples in total. Of these, 29 samples came from sites on
137 unconfined Chalk and 27 were on unconfined mudstone.

138 For the River Eden, samples were collected from nine headwater tributaries on four occasions (March,
139 June and October 2012, March 2013). 2–3 replicates were collected from each tributary on each
140 sampling occasion, giving 75 samples in total. Of these, 21 samples were from sites on limestone, 27
141 on sandstone and 27 on volcanics, all confined by glacial deposits.

142 Water samples for dissolved N_2O analysis were collected in 20 mL glass syringes that were flushed
143 three times with river water and any trapped air expelled before the final sample was taken. Samples
144 were returned to cold storage (4°C) within 3 h and analyzed for N_2O within 72 h of collection. Water
145 samples for NO_3^- , NO_2^- , NH_4^+ and dissolved organic carbon (DOC) analysis were grab sampled in 1 L
146 polypropylene bottles and also analyzed within 72 h of collection after filtering. For 240 of the River
147 Wensum grab samples collected between May 2012 and May 2013, an aliquot was filtered through a
148 0.22 μm cellulose acetate filter and frozen at -20°C in preparation for nitrogen and oxygen stable
149 isotope analysis. River water temperatures were measured in-situ with a handheld alcohol
150 thermometer.

151 **2.3 Sample Analysis**

152 Dissolved N_2O concentrations were determined by purge-and-trap gas chromatography with an
153 electron capture detector (Shimadzu GC-ECD) which had a measurement accuracy within $\pm 3\%$ and a
154 detection limit of 0.0008 $\mu g N L^{-1}$. Dissolved NO_3^- concentrations were determined by ion
155 chromatography (Dionex ICS-2000) with a precision of $\pm 0.2 mg N L^{-1}$, whilst dissolved NO_2^- and
156 NH_4^+ were determined by a continuous flow analyzer (Skalar SAN++) with precisions of ± 1.5 and 5
157 $\mu g L^{-1}$, respectively. DOC was determined by a Shimadzu TOC/TN analyzer with a precision of ± 0.5
158 $mg L^{-1}$. Samples for isotopic analysis were prepared using the denitrifier method and analyzed on a
159 GEO 20:20 GC-IRMS with a TG II prep system with a precision of $\pm 0.4\%$ for $\delta^{15}N_{NO_3}$ and $\pm 0.6\%$ for
160 $\delta^{18}O_{NO_3}$ (Table S2).

161 **2.4 Fluxes and Emission Factors**

162 N₂O emission factors for each river water sample were calculated by the mass ratio approach (eq. 2)
 163 derived from the measured concentrations of N₂O (mg N L⁻¹) and NO₃⁻ (mg N L⁻¹). Fluxes of N₂O
 164 from the river to the atmosphere were calculated using the water-air gas exchange eq. 3:³⁸

$$165 \quad (3) \quad F = kC_w - \frac{C_a}{k'_h}$$

166 where F is the flux of N₂O (mol cm⁻² h⁻¹), subsequently converted into kg N ha⁻¹ a⁻¹; k is the gas
 167 transfer velocity of N₂O across the water-air interface (cm h⁻¹); C_w is the concentration of N₂O in river
 168 water (mol cm⁻³); C_a is the concentration of N₂O in the atmosphere (mol cm⁻³); and k'_h is the
 169 dimensionless Henry's law constant for N₂O. Estimation of the gas transfer velocity represents a
 170 major source of uncertainty in water-atmosphere gas flux calculations and currently no definitive
 171 method exists to define k values.^{39,40} The use of benthic turbulence models which incorporate aspects
 172 of stream velocity, stream depth, bed roughness and bed slope, likely produce more accurate gas
 173 exchange rates for small-to-medium sized rivers such as those studied here.⁴¹ However, with only
 174 three of our 35 study sites being gauged (Figure 1), a lack of velocity and depth data meant we instead
 175 adopted a wind-based turbulence model where k was calculated as eq. 4:⁴²

$$176 \quad (4) \quad k = 1.91e^{0.35U} \left(\frac{Sc}{600} \right)^{-0.5}$$

177 where U is the wind speed (m s⁻¹) and Sc is the Schmidt number for N₂O in freshwater adjusted for
 178 temperature.⁴³ Although likely less accurate than a benthic turbulence model, the approach adopted is
 179 consistent across all sites and it yields mean k values of 4.0, 4.5 and 4.6 for the Avon, Eden and
 180 Wensum catchments, respectively, which are within the range of 3–7 previously recommended.⁴²
 181 Mean wind speed data (15-min resolution) for the time of sample collection were obtained from local
 182 weather stations within each catchment.

183

184 The N₂O saturation level (%) was calculated as eq. 5:²⁸

$$185 \quad (5) \quad N_2O_{(sat)} = \frac{N_2O_{(water)}}{N_2O_{(eq)}} \times 100$$

186 where N₂O_(water) is the measured N₂O concentration in river water and N₂O_(eq) is the concentration
 187 when water is in equilibrium with the atmosphere.⁴⁴

188

189 **3. Results and Discussion**

190 **3.1 Spatial variability**

191 In 99.9% of river water samples, N₂O saturation levels exceeded the atmospheric equilibrium
 192 implying almost all sites were acting as a net source of N₂O to the atmosphere. Saturation levels
 193 ranged from 90–1305% (median = 283%) for the River Wensum, 116–455% (median = 158%) for the
 194 River Eden and 136–17070% (median = 1178%) for the River Avon. Whilst the Wensum and Eden

195 values are comparable with other studies, saturation levels in the River Avon are towards the upper
196 end of the range previously reported.^{14, 26, 45, 46}

197 Unconfined Chalk regions of the rivers Avon (16.83 $\mu\text{g N L}^{-1}$) and Wensum (2.53 $\mu\text{g N L}^{-1}$) had the
198 highest median N_2O concentrations, contributing to a median concentration for all unconfined Chalk
199 sites of 3.03 $\mu\text{g N L}^{-1}$ (Table 1; Figure 2). Comparatively high median NO_3^- concentrations were also
200 observed for the unconfined Chalk of the Avon (7.01 mg N L^{-1}) and Wensum (9.21 mg N L^{-1}), and
201 together these unconfined Chalk regions yielded the highest median EF_{5r} (0.00036) and N_2O flux
202 (10.8 $\text{kg N ha}^{-1} \text{a}^{-1}$). These indirect fluxes are towards the upper end of the range previously reported
203 for rivers draining arable and grassland sites in Europe.⁴⁷ However, despite unconfined Chalk regions
204 having the highest EF_{5r} of the different hydrogeological types, the overall median EF_{5r} was 7 times
205 lower than the IPCC default value of 0.0025. Only unconfined Chalk regions of the River Avon had a
206 median EF_{5r} value (0.00235) comparable to the IPCC default.

207 Regions with confined hydrogeological conditions under glacial deposits yielded the lowest riverine
208 N_2O concentrations, regardless of bedrock type or geographical location. Median N_2O concentrations
209 in river water samples from confined limestone (0.52 $\mu\text{g N L}^{-1}$), volcanic (0.57 $\mu\text{g N L}^{-1}$), sandstone
210 (0.61 $\mu\text{g N L}^{-1}$) and Chalk (0.79 $\mu\text{g N L}^{-1}$) bedrock areas were ~ 4 times lower (t -test $p < 0.05$) than
211 recorded in rivers draining areas of unconfined Chalk. The confined Wensum Chalk (5.20 mg N L^{-1})
212 and confined Eden sandstone (4.52 mg N L^{-1}) also had lower NO_3^- concentrations than the unconfined
213 Wensum Chalk sites, although higher riverine NO_3^- concentrations were recorded on confined
214 limestone (8.61 mg N L^{-1}) in the River Eden. Together, the median EF_{5r} (0.00016) and N_2O flux (2.6
215 $\text{kg ha}^{-1} \text{a}^{-1}$) for all sites confined by glacial deposits were the lowest and second lowest recorded,
216 respectively, with an emission factor 16 times lower than the IPCC default.

217 The semi-confined Chalk hydrogeological grouping, composed of main River Wensum sites which
218 receive a mix of N_2O and NO_3^- enriched water from unconfined Chalk tributaries and N_2O and NO_3^-
219 depleted water from confined Chalk tributaries, had median N_2O (1.31 $\mu\text{g N L}^{-1}$) and NO_3^- (5.99 mg N
220 L^{-1}) concentrations, EF_{5r} (0.00022) and flux rate (5.1 $\text{kg N ha}^{-1} \text{a}^{-1}$) between that of the confined and
221 unconfined sites. However, in pronounced contrast to the unconfined Chalk sites, streams on
222 unconfined mudstone in the River Avon yielded low median N_2O (0.69 $\mu\text{g N L}^{-1}$) and NO_3^- (4.46 mg
223 N L^{-1}) concentrations. This indicates that bedrock permeability may exert an important control on
224 N_2O production where it is unconfined, with mudstone permeability being substantially lower than
225 that of Chalk. Emission factors for the unconfined mudstone were highly variable, but median values
226 were ~ 12 times lower than unconfined Chalk in the same catchment (0.00020), with an emission rate
227 of 2.0 $\text{kg ha}^{-1} \text{a}^{-1}$.

228 Importantly, despite river discharge varying substantially between sampling locations (Figure S1)
229 there is no evidence of a dilution effect in N_2O concentrations at the larger main river sites, nor is

230 there evidence of a strong N₂O degassing signal as water moves further down the catchment. The
231 hydrogeological conditions at the sampling sites remain the dominant classifier of N₂O concentration
232 and EF_{5r} regardless of discharge or stream order. Evidence for this can be seen in Table 1, where the
233 semi-confined Chalk grouping of the River Wensum is composed solely of the four main river sample
234 locations which have the greatest discharges and highest stream orders. If downstream degassing and
235 dilution were major controls on N₂O, we would expect these semi-confined sites to have lower N₂O
236 concentrations than the other 16 Wensum tributary locations, but this is not the case. Instead, median
237 N₂O concentrations and EF_{5r} values were significantly ($p < 0.05$) higher at the semi-confined main
238 river sites (N₂O = 1.31 μg N L⁻¹; EF_{5r} = 0.00022) than recorded in the confined upstream tributary
239 locations (N₂O = 0.79 μg N L⁻¹; EF_{5r} = 0.00016). Similarly, we also observe that despite having
240 substantially higher discharge (Figure S1), confined tributary sites in the River Eden yield comparable
241 median N₂O concentrations (0.52–0.61 μg N L⁻¹) and EF_{5r} values (0.00007–0.00019) to the confined
242 tributary locations in the River Wensum.

243 3.2 Temporal variability

244 Seasonally, riverine N₂O and NO₃⁻ concentrations were lowest during spring (MAM) and summer
245 (JJA), respectively, regardless of hydrogeological conditions (Table 2). Likewise, the highest NO₃⁻
246 concentrations typically occurred across all locations during the winter (DJF), consistent with higher
247 N leaching rates under wetter antecedent conditions. Highest N₂O concentrations did, however, differ
248 by hydrogeological type, being greatest during summer and autumn (SON) in unconfined and semi-
249 confined regions, and during winter in areas of confined Chalk. Emission factors were highest during
250 summer/autumn and lowest during spring, irrespective of hydrogeological type, with these seasonal
251 contrasts being statistically significant (t -test $p < 0.05$). Such patterns are broadly consistent with the
252 temporal variability in N₂O concentrations reported previously,^{26, 48, 49} and demonstrate that
253 application of a single default EF_{5r} value fails to capture the significant temporal variability in N₂O
254 dynamics and could lead to a misrepresentation of the true N₂O flux. Here, median N₂O flux rates
255 were greatest during winter irrespective of hydrogeological conditions due to significantly higher
256 wind speeds at this time of year yielding a higher gas transfer velocity for the wind disturbance-based
257 Equation 4.

258 Unconfined Chalk sites consistently had the highest dissolved N₂O concentrations throughout the 28-
259 month period, with concentrations ranging from 0.81–4.70 μg L⁻¹ (Figure 3). Conversely, at Chalk
260 sites confined by glacial deposits N₂O concentrations were consistently the lowest and least variable,
261 ranging from 0.32–2.35 μg L⁻¹. The semi-confined Chalk sites were intermediate to the confined and
262 unconfined locations. Peaks in N₂O concentration (e.g. December 2011, June 2012) were associated
263 with rainfall events <24 h before sample collection, which also yielded peaks in the concentration of
264 NO₃⁻ (Figure S2), NH₄ (Figure S3), NO₂⁻ (Figure S4) and in EF_{5r} values (Figure S6). Note that the

265 IPCC typically derives EF_{5r} values based on annual N loads and annual N_2O emissions and thus the
266 instantaneous EF_{5r} values presented in Figure S6 will inherently exhibit greater temporal variability.

267 3.3 Controls on N_2O dynamics

268 It is clear from these data that N_2O concentrations, fluxes and emission factors vary between regions
269 of contrasting hydrogeological conditions and it is important to understand why this differentiation
270 occurs in order to confidently upscale EF_{5r} estimates nationally based on this characteristic. We
271 hypothesize that in all three catchments, fertilizer inputs are hydrolyzed to NH_4^+ and readily nitrified
272 to NO_3^- in the soil with further nitrification occurring in stream. In unconfined Chalk regions, NO_3^-
273 and N_2O from the soil are rapidly transported in infiltrating water down to the well-mixed
274 groundwater zone in the high permeability Chalk. As N_2O and NO_3^- enriched groundwater comprises
275 the major proportion of river flow in these regions (BFI = 0.70–0.95), high N_2O and NO_3^-
276 concentrations are subsequently observed instream, alongside elevated EF_{5r} values. Conversely, in
277 regions of confined groundwater and smaller BFI (0.40–0.70), lower permeability glacial deposits
278 reduce infiltration rates and allow low oxygen conditions to develop where denitrification and/or
279 nitrifier denitrification can occur before the infiltrating water recharges groundwater. This process
280 partially protects groundwater from NO_3^- leaching and results in lower NO_3^- and N_2O concentrations
281 at these sites.⁵⁰ Although regions of unconfined mudstone are not protected by overlying glacial
282 deposits, the lower permeability of the mudstone relative to the Chalk results in similar opportunities
283 for denitrification and/or nitrifier denitrification, thus resulting in lower NO_3^- concentrations, reduced
284 rates of soil-to-river N_2O transfer and lower EF_{5r} values. This hypothesis is supported by the stable
285 isotope data (Figure 4b).

286 The nitrogen ($\delta^{15}N$) and oxygen ($\delta^{18}O$) isotopic composition of NO_3^- can be used to infer mixing of
287 sources of NO_3^- with differing isotopic composition and to indicate the dominance of nitrification and
288 denitrification (see Supporting Information). The fractionation ratio of $\delta^{15}N_{NO_3}$ to $\delta^{18}O_{NO_3}$ for the
289 Wensum samples is 0.41 (Figure 4b), providing some evidence of denitrification across the catchment
290 drainage network. Mixing of atmospheric and fertilizer direct NO_3^- sources with partially denitrified
291 NO_3^- raises the bulk $\delta^{18}O_{NO_3}$ and produces scatter above the denitrification slope (Figure 4b).
292 Variation in pre-nitrification $\delta^{15}N_{NH_4}$ is reflected in post-nitrification $\delta^{15}N_{NO_3}$, producing scatter on the
293 $\delta^{15}N_{NO_3}$ axis. Together these mixing effects result in a weak relationship between $\delta^{18}O_{NO_3}$ and $\delta^{15}N_{NO_3}$
294 ($R^2 = 0.375$), reflecting the combined effects of mixing and denitrification.

295 The heaviest expected $\delta^{18}O_{NO_3}$ produced from nitrification of NH_4^+ in the Wensum catchment is
296 3.8%.⁵¹ This value is derived from the incorporation of oxygen from ambient water and air during
297 nitrification at an initial ratio of 2:1,^{52, 53} as well as from measurements of $\delta^{18}O_{H_2O}$ in rivers,
298 tributaries, field drains, streambed piezometers and boreholes in the Wensum catchment ($\delta^{18}O_{H_2O} =$
299 6.0% to -7.5%).⁵⁰ This upper limit would not be affected by any abiotic oxygen exchange between

300 NO_2^- and H_2O ,⁵⁴ because this would result in isotopically lighter $\delta^{18}\text{O}_{\text{NO}_3}$. Fertilizer and precipitation
301 direct sources of NO_3^- ($\delta^{18}\text{O}_{\text{NO}_3} > 20\text{‰}$) have $\delta^{18}\text{O}_{\text{NO}_3}$ values clearly differentiated from that of NO_3^-
302 produced from nitrification in the Wensum catchment, thus $\delta^{18}\text{O}_{\text{NO}_3} \leq 3.8\text{‰}$ can be used as an
303 indicator of NO_3^- from nitrification (Figure 4b).

304 There is a positive relationship between the relative dominance of nitrification NO_3^- and median N_2O
305 concentrations with hydrogeological setting. The highest proportion of samples containing
306 nitrification NO_3^- ($\delta^{18}\text{O}_{\text{NO}_3} \leq 3.8\text{‰}$) were from unconfined Chalk sites (42% of 48 samples). These
307 sites also produced the highest median N_2O concentrations ($2.53 \mu\text{g N}_2\text{O L}^{-1}$). In comparison, the
308 confined Chalk produced the lowest proportion of samples containing nitrification NO_3^- (7% of 132
309 samples), with the lowest median N_2O concentrations ($0.79 \mu\text{g N}_2\text{O L}^{-1}$). The proportion of
310 nitrification NO_3^- from semi-confined sites was between that of unconfined and confined Chalk sites
311 (22% of 60 samples) with a median N_2O concentration of $1.31 \mu\text{g N}_2\text{O L}^{-1}$. This relationship indicates
312 that nitrification, rather than denitrification, is the dominant N_2O production process in the Wensum
313 catchment. We suggest that in unconfined Chalk sites, infiltration of recharge water occurs rapidly to
314 well-mixed shallow groundwater. Baseflow transports dissolved nitrification NO_3^- , by-product N_2O
315 and denitrification-inhibiting dissolved oxygen (DO saturation = 89–97%⁵⁰) into the river.
316 Denitrification may also be inhibited in the unconfined sites by a relatively low availability of labile
317 carbon, with a mean DOC: NO_3^- ratio < 1 at unconfined Chalk sites and > 1 at confined sites.

318 Across all hydrogeological types in the Wensum catchment, N_2O concentrations and saturation levels
319 were only weakly correlated with pH ($R^2 = < 0.08$) and water temperature ($R^2 = < 0.07$), indicating
320 these variables were not directly acting as abiotic controls on N_2O production (Figure S9). Stronger
321 negative correlations were, however, established between EF_{5r} and total N ($R^2 = 0.35\text{--}0.59$) providing
322 evidence of decreasing (de)nitrification efficiency with increasing N inputs due to progressive
323 biological saturation.³¹ Low NH_4^+ concentrations at unconfined Chalk sites indicate nitrification is
324 acting as a sink for NH_4^+ (Figure S9).

325 **3.4 Implications and recommendations**

326 The evidence presented here clearly demonstrates that N_2O emission factors vary significantly
327 between regions of contrasting hydrogeological type. Given the inherent regional nature of
328 hydrogeological variability as determined by the distribution of bedrock and superficial geologies, this
329 robust association with EF_{5r} values indicates that hydrogeological conditions could be used as a
330 defining environmental characteristic for upscaling N_2O emission estimates. Undoubtedly there is a
331 need to further explore whether this association is maintained across a wider range of hydrogeological
332 settings than those investigated here. Nevertheless, regional variability in hydrogeological conditions
333 could be used to generate improved regional-scale EF_{5r} estimates that are essential for developing
334 more accurate national greenhouse gas inventories. Such an approach would address the pressing need

335 to produce more refined EF₅ values than the current broad-brush method adopted by the IPCC; a call
336 which has been repeatedly emphasized in numerous studies over the past decade.^{14, 26, 32, 38} The
337 unsuitable nature of the default IPCC EF_{5r} value is again highlighted here with 98% of samples having
338 an EF_{5r} lower than the default 0.0025 value, this despite the downward revision of EF_{5r} during the
339 2006 IPCC Fourth Assessment Report.³⁰ We therefore encourage researchers to investigate if the
340 association between hydrogeology and EF_{5r} is maintained across contrasting river basins. If it is, it
341 should be possible to calibrate hydrogeological-specific EF_{5r} emission factors which can be overlain
342 onto existing global spatial lithological^{55, 56} and hydrological⁵⁷ databases as a means of effectively
343 upscaling indirect N₂O emissions from rivers draining defined hydrogeological regions.

344 **Supporting Information**

345 The Supporting Information contains hydrological summaries for the study locations (Figure S1;
346 Table S1); description of the laboratory procedures; water quality time-series for the River Wensum
347 (Figures S2–S7); extended discussion of the stable isotope data (Figure S8; Table S2); and regression
348 plots for nitrogen species (Figure S9). The project data are provided in Excel spreadsheet format.

349 **Author Information**

350 *Corresponding Author: Richard J. Cooper; Richard.J.Cooper@uea.ac.uk; +44(0)1603592922;
351 School of Environmental Sciences, University of East Anglia, Norwich Research Park, Norwich NR4
352 7TJ, UK

353 **Acknowledgements**

354 This research was funded by Defra under the UK Inventory of Agricultural Greenhouse Gas
355 Emissions Platform (AC0116). We are grateful to the Avon and Eden Demonstration Test Catchments
356 for providing logistical support and to Lee Gumm for assistance with data analysis.

357 **References**

- 358 1. Myhre, G.; Shindell, D.; Bréon, F.-M.; Collins, W.; Fuglestedt, J.; J., H.; Koch, D.;
359 Lamarque, J.-F.; Lee, D.; Mendoza, B.; Nakajima, T.; Robock, A.; Stephens, G.; Takemura, T.;
360 Zhang, H., Anthropogenic and Natural Radiative Forcing. In *Climate Change 2013: The Physical
361 Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the
362 Intergovernmental Panel on Climate Change.*, Stocker, T. F.; Qin, D.; Plattner, G.-K.; Tignor, M.;
363 Allen, S. K.; Boschung, J.; Nauels, A.; Xia, Y.; V., B.; Midgley, P. M., Eds. Cambridge University
364 Press, Cambridge, United Kingdom: 2013.
- 365 2. NOAA Earth System Research Laboratory.
366 https://www.esrl.noaa.gov/gmd/dv/data/?parameter_name=Nitrous%2BOxide (03/03/2017),
- 367 3. Hartmann, D. L.; Klein Tank, A. M. G.; Rusticucci, M.; Alexander, L. V.; Brönnimann, S.;
368 Charabi, Y.; Dentener, F. J.; Dlugokencky, E. J.; Easterling, D. R.; Kaplan, A.; Soden, B. J.; Thorne,
369 P. W.; Wild, M.; Zhai, P. M., Observations: Atmosphere and Surface. In *Climate Change 2013: The
370 Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the
371 Intergovernmental Panel on Climate Change*, Stocker, T. F.; Qin, D.; Plattner, G.-K.; Tignor, M.;

- 372 Allen, S. K.; Boschung, J.; Nauels, A.; Xia, Y.; Bex, V.; Midgley, P. M., Eds. Cambridge University
373 Press, Cambridge, United Kingdom: 2013.
- 374 4. Ravishankara, A. R.; Daniel, J. S.; Portmann, R. W., Nitrous oxide (N₂O): the dominant
375 ozone-depleting substance emitted in the 21st century. *Science* **2009**, *326*, 123-125.
- 376 5. Davidson, E. A., The contribution of manure and fertilizer nitrogen to atmospheric nitrous
377 oxide since 1860. *Nature Geoscience* **2009**, *2*, (9), 659-662.
- 378 6. Bremner, J. M.; Blackmer, A. M., Nitrous oxide: emission from soils during nitrification of
379 fertilizer nitrogen. *Science* **1978**, *199*, (4326), 295-296.
- 380 7. Strauss, E. A.; Richardson, W. B.; Bartsch, L. A.; Cavanaugh, J. C.; Bruesewitz, D. A.;
381 Imker, H.; Heinz, J. A.; Soballe, D. M., Nitrification in the Upper Mississippi River: patterns,
382 controls, and contribution to the NO₃ budget. *Journal of the North American Benthological Society*
383 **2004**, *23*, (1), 1-14.
- 384 8. Mulholland, P. J.; Helton, A. M.; Poole, G. C.; Hall, R. O.; Hamilton, S. K.; Peterson, B. J.;
385 Tank, J. L.; Ashkenas, L. R.; Cooper, L. W.; Dahm, C. N.; Dodds, W. K.; Findlay, S. E.; Gregory, S.
386 V.; Grimm, N. B.; Johnson, S. L.; McDowell, W. H.; Meyer, J. L.; Valett, H. M.; Webster, J. R.;
387 Arango, C. P.; Beaulieu, J. J.; Bernot, M. J.; Burgin, A. J.; Crenshaw, C. L.; Johnson, L. T.;
388 Niederlehner, B. R.; O'Brien, J. M.; Potter, J. D.; Sheibley, R. W.; Sobota, D. J.; Thomas, S. M.,
389 Stream denitrification across biomes and its response to anthropogenic nitrate loading. *Nature* **2008**,
390 *452*, (7184), 202-205.
- 391 9. Beaulieu, J. J.; Tank, J. L.; Hamilton, S. K.; Wollheim, W. M.; Hall, R. O.; Mulholland, P. J.;
392 Peterson, B. J.; Ashkenas, L. R.; Cooper, L. W.; Dahm, C. N.; Dodds, W. K.; Grimm, N. B.; Johnson,
393 S. L.; McDowell, W. H.; Poole, G. C.; Maurice Valett, H.; Arango, C. P.; Bernot, M. J.; Burgin, A. J.;
394 Crenshaw, C. L.; Helton, A. M.; Johnson, L. T.; O'Brien, J. M.; Potter, J. D.; Sheibley, R. W.; Sobota,
395 D. J.; Thomas, S. M., Nitrous oxide emission from denitrification in stream and river networks. *PNAS*
396 **2011**, *108*, (1), 214-219.
- 397 10. Kool, D. M.; Dolfing, J.; Wrage, N.; Van Groenigen, J. W., Nitrifier denitrification as a
398 distinct and significant source of nitrous oxide from soil. *Soil Biology and Biochemistry* **2011**, *43*, (1),
399 174-178.
- 400 11. Syakila, A.; Kroeze, C., The global nitrous oxide budget revisited. *Greenhouse Gas*
401 *Measurement and Management* **2011**, *1*, (1), 17-26.
- 402 12. Mosier, A.; Kroeze, C.; Nevison, C.; Oenema, O.; Seitzinger, S.; van Cleemput, O., Closing
403 the global N₂O budget: nitrous oxide emissions through the agricultural nitrogen cycle. *Nutrient*
404 *Cycling in Agroecosystems* **1998**, *52*, 225-248.
- 405 13. Nevison, C., Review of the IPCC methodology for estimating nitrous oxide emissions
406 associated with agricultural leaching and runoff. *Chemosphere* **2000**, *2*, 493-500.
- 407 14. Clough, T. J.; Buckthought, L. E.; Kelliher, F. M.; Sherlock, R. R., Diurnal fluctuations of
408 dissolved nitrous oxide (N₂O) concentrations and estimates of N₂O emissions from a spring-fed river:
409 implications for IPCC methodology. *Global Change Biology* **2007**, *13*, (5), 1016-1027.
- 410 15. Bouwman, A. F., Direct emission of nitrous oxide from agricultural soils. *Nutrient Cycling in*
411 *Agroecosystems* **1996**, *46*, 53-70.
- 412 16. Davidson, E. A.; Kinglerlee, W., A global inventory of nitric oxide emissions from soils.
413 *Nutrient Cycling in Agroecosystems* **1997**, *48*, 14.
- 414 17. Lesschen, J. P.; Velthof, G. L.; de Vries, W.; Kros, J., Differentiation of nitrous oxide
415 emission factors for agricultural soils. *Environ Pollut* **2011**, *159*, (11), 3215-22.
- 416 18. Smith, K. A.; Dobbie, K. E.; Thorman, R.; Watson, C. J.; Chadwick, D. R.; Yamulki, S.; Ball,
417 B. C., The effect of N fertilizer forms on nitrous oxide emissions from UK arable land and grassland.
418 *Nutrient Cycling in Agroecosystems* **2012**, *93*, 127-149.
- 419 19. Butterbach-Bahl, K.; Baggs, E. M.; Dannenmann, M.; Kiese, R.; Zechmeister-Boltenstern, S.,
420 Nitrous oxide emissions from soils: how well do we understand the processes and their controls?
421 *Philos Trans R Soc Lond B Biol Sci* **2013**, *368*, (1621), 20130122.
- 422 20. Griffis, T. J.; Lee, X.; Baker, J. M.; Russelle, M. P.; Zhang, X.; Venterea, R.; Millet, D. B.,
423 Reconciling the differences between top-down and bottom-up estimates of nitrous oxide emissions for
424 the U.S. Corn Belt. *Global Biogeochemical Cycles* **2013**, *27*, (3), 746-754.
- 425 21. Reay, D. S.; Davidson, E. A.; Smith, K. A.; Smith, P.; Melillo, J. M.; Dentener, F.; Crutzen,
426 P. J., Global agriculture and nitrous oxide emissions. *Nature Climate Change* **2012**, *2*, (6), 410-416.

- 427 22. Reay, D. S.; Smith, K. A.; Edwards, A. C., Nitrous oxide emission from agricultural drainage
428 waters. *Global Change Biology* **2003**, *9*, 195-203.
- 429 23. Reay, D. S.; Smith, K. A.; Edwards, A. C.; Hiscock, K. M.; Dong, L. F.; Nedwell, D. B.,
430 Indirect nitrous oxide emissions: Revised emission factors. *Environmental Sciences* **2005**, *2*, (2-3),
431 153-158.
- 432 24. Well, R.; Weymann, D.; Flessa, H., Recent research progress on the significance of aquatic
433 systems for indirect agricultural N₂O emissions. *Environmental Sciences* **2005**, *2*, (2-3), 143-151.
- 434 25. Beaulieu, J. J.; Arango, C. P.; Hamilton, S. K.; Tank, J. L., The production and emission of
435 nitrous oxide from headwater streams in the Midwestern United States. *Global Change Biology* **2008**,
436 *14*, (4), 878-894.
- 437 26. Hinshaw, S. E.; Dahlgren, R. A., Dissolved nitrous oxide concentrations and fluxes from the
438 eutrophic San Joaquin River, California. *Environ Sci Technol* **2013**, *47*, (3), 1313-1322.
- 439 27. Yu, Z.; Deng, H.; Wang, D.; Ye, M.; Tan, Y.; Li, Y.; Chen, Z.; Xu, S., Nitrous oxide
440 emissions in the Shanghai river network: implications for the effects of urban sewage and IPCC
441 methodology. *Glob Chang Biol* **2013**, *19*, (10), 2999-3010.
- 442 28. Chen, N.; Wu, J.; Zhou, X.; Chen, Z.; Lu, T., Riverine N₂O production, emissions and export
443 from a region dominated by agriculture in Southeast Asia (Jiulong River). *Agriculture, Ecosystems &*
444 *Environment* **2015**, *208*, 37-47.
- 445 29. Turner, P. A.; Griffis, T. J.; Lee, X.; Baker, J. M.; Venterea, R. T.; Wood, J. D., Indirect
446 nitrous oxide emissions from streams within the US Corn Belt scale with stream order. *Proc Natl*
447 *Acad Sci U S A* **2015**, *112*, (32), 9839-9843.
- 448 30. De Klein, C.; Novoa, R. S. A.; Ogle, S.; Smith, K. A.; Rochette, P.; Wirth, T. C., N₂O
449 emissions from managed soils, and CO₂ emissions from lime and urea application. In *2006 IPCC*
450 *guidelines for national greenhouse gas inventories*, Cambridge University Press, Cambridge, United
451 Kingdom: 2006; p 54.
- 452 31. Hu, M.; Chen, D.; Dahlgren, R. A., Modeling nitrous oxide emission from rivers: a global
453 assessment. *Glob Chang Biol* **2016**, *22*, (11), 3566-3582.
- 454 32. Hama-Aziz, Z. Q.; Hiscock, K. M.; Cooper, R. J., Indirect Nitrous Oxide Emission Factors
455 for Agricultural Field Drains and Headwater Streams. *Environ Sci Technol* **2017**, *51*, (1), 7.
- 456 33. Skiba, U.; Jones, S. K.; Dragosits, U.; Drewer, J.; Fowler, D.; Rees, R. M.; Pappa, V. A.;
457 Cardenas, L.; Chadwick, D.; Yamulki, S.; Manning, A. J., UK emissions of the greenhouse gas
458 nitrous oxide. *Philos Trans R Soc Lond B Biol Sci* **2012**, *367*, (1593), 1175-1185.
- 459 34. McGonigle, D. F.; Burke, S. P.; Collins, A. L.; Gartner, R.; Haft, M. R.; Harris, R. C.;
460 Haygarth, P. M.; Hedges, M. C.; Hiscock, K. M.; Lovett, A. A., Developing Demonstration Test
461 Catchments as a platform for transdisciplinary land management research in England and Wales.
462 *Environ Sci Process Impacts* **2014**, *16*, (7), 1618-1628.
- 463 35. CEH National River Flow Archive. <https://nrfa.ceh.ac.uk/data/search> (17/07/2017),
- 464 36. Hiscock, K. M.; Dennis, P. F.; Saynor, P. R.; Thomas, M. O., Hydrochemical and stable
465 isotope evidence for the extent and nature of the effective Chalk aquifer of north Norfolk, UK.
466 *Journal of Hydrology* **1996**, *180*, 29.
- 467 37. Meteorological-Office UK Climate Averages.
468 <http://www.metoffice.gov.uk/public/weather/climate/?tab=climateStations> (17/07/2017),
- 469 38. Outram, F. N.; Hiscock, K. M., Indirect nitrous oxide emissions from surface water bodies in
470 a lowland arable catchment: a significant contribution to agricultural greenhouse gas budgets?
471 *Environ Sci Technol* **2012**, *46*, (15), 8156-8163.
- 472 39. Beaulieu, J. J.; Shuster, W. D.; Rebholz, J. A., Controls on gas transfer velocities in a large
473 river. *Journal of Geophysical Research: Biogeosciences* **2012**, *117*, (G2).
- 474 40. Raymond, P. A.; Zappa, C. J.; Butman, D.; Bott, T. L.; Potter, J.; Mulholland, P.; Laursen, A.
475 E.; McDowell, W. H.; Newbold, D., Scaling the gas transfer velocity and hydraulic geometry in
476 streams and small rivers. *Limnology and Oceanography: Fluids and Environments* **2012**, *2*, (1), 41-
477 53.
- 478 41. Baulch, H. M.; Venkiteswaran, J. J.; Dillon, P. J.; Maranger, R., Revisiting the application of
479 open-channel estimates of denitrification. *Limnology and Oceanography: Methods* **2010**, *8*, (5), 202-
480 215.

- 481 42. Raymond, P. A.; Cole, J., Gas exchange in rivers and estuaries: choosing a gas transfer
482 velocity. *Estuaries* **2001**, *24*, (2), 312-317.
- 483 43. Wanninkhof, R., Relationship between wind speed and gas exchange over the ocean. *Journal*
484 *of Geophysical Research* **1992**, *97*, (C5), 7373-7382.
- 485 44. Weiss, R. F.; Price, B. A., Nitrous oxide solubility in water and seawater. *Marine Chemistry*
486 **1980**, *8*, 13.
- 487 45. Cole, J.; Caraco, N., Emissions of nitrous oxide (N₂O) from a tidal, freshwater river, the
488 Hudson River, New York. *Environmental Science & Technology* **2001**, *35*, (6), 6.
- 489 46. Xia, Y.; Li, Y.; Li, X.; Guo, M.; She, D.; Yan, X., Diurnal pattern in nitrous oxide emissions
490 from a sewage-enriched river. *Chemosphere* **2013**, *92*, (4), 421-428.
- 491 47. Garnier, J.; Billen, G.; Vilain, G.; Martinez, A.; Silvestre, M.; Mounier, E.; Toche, F., Nitrous
492 oxide (N₂O) in the Seine river and basin: Observations and budgets. *Agriculture, Ecosystems &*
493 *Environment* **2009**, *133*, (3-4), 223-233.
- 494 48. Wilcock, R. J.; Sorrell, B. K., Emissions of Greenhouse Gases CH₄ and N₂O from Low-
495 gradient Streams in Agriculturally Developed Catchments. *Water, Air, and Soil Pollution* **2008**, *188*,
496 (1-4), 155-170.
- 497 49. Hama-Aziz, Z. Q.; Hiscock, K. M.; Cooper, R. J., Dissolved nitrous oxide (N₂O) dynamics in
498 agricultural field drains and headwater streams in an intensive arable catchment. *Hydrological*
499 *Processes* **2017**, *31*, (6), 1371-1381.
- 500 50. Wexler, S. K. An investigation into the sources, cycling and attenuation of nitrate in an
501 agricultural lowland catchment using stable isotopes of nitrogen and oxygen in nitrate. University of
502 East Anglia, 2011.
- 503 51. Wexler, S. K.; Hiscock, K. M.; Dennis, P. F., Catchment-scale quantification of hyporheic
504 denitrification using an isotopic and solute flux approach. *Environ Sci Technol* **2011**, *45*, (9), 3967-
505 3973.
- 506 52. Andersson, K. K.; Hooper, A. B., O₂ and H₂O are each the source of one O in NO⁻² produced
507 from NH₃ by Nitrosomonas: ¹⁵N-NMR evidence. *FEBS Letters* **1983**, *164*, (2), 236-240.
- 508 53. Kumar, S.; Nicholas, D. J. D.; Williams, E. H., Definitive ¹⁵N NMR evidence that water
509 serves as a source of 'O' during nitrite oxidation by Nitrobacter agilis. *FEBS Letters* **1983**, *152*, (1),
510 71-74.
- 511 54. Snider, D. M.; Spoelstra, J.; Schiff, S. L.; Venkiteswaran, J. J., Stable oxygen isotope ratios of
512 nitrate produced from nitrification: ¹⁸O-labeled water incubations of agricultural and temperate forest
513 soils. *Environmental Science & Technology* **2010**, *44*, 5358-5364.
- 514 55. Gleeson, T.; Smith, L.; Moosdorf, N.; Hartmann, J.; Dürr, H. H.; Manning, A. H.; van Beek,
515 L. P. H.; Jellinek, A. M., Mapping permeability over the surface of the Earth. *Geophysical Research*
516 *Letters* **2011**, *38*, (2), L02401.
- 517 56. Dürr, H. H.; Meybeck, M.; Dürr, S. H., Lithologic composition of the Earth's continental
518 surfaces derived from a new digital map emphasizing riverine material transfer. *Global*
519 *Biogeochemical Cycles* **2005**, *19*, (4), GB4S10.
- 520 57. Doll, P.; Fiedler, K., Global-scale modeling of groundwater recharge. *Hydrology and Earth*
521 *System Sciences* **2008**, *12*, 863-885.

522

523

524

525

526

527

528

529

530 **Figure Captions**

531 **Figure 1:** Location of the three UK study catchments with associated bedrock and superficial
532 geologies. Based upon DiGMapGB-625, with the permission of the British Geological Survey.

533 **Figure 2:** Violin plot summaries of (a) dissolved nitrous oxide concentration, (b) nitrate
534 concentration, (c) EF_{5r} emission factor and (d) nitrous oxide flux for all river water samples by
535 hydrogeological type. Various includes limestone, chalk, sandstone and volcanic bedrock.

536 **Figure 3:** Time-series of dissolved nitrous oxide concentrations in the River Wensum catchment
537 between February 2011 and May 2013.

538 **Figure 4:** (a) Dissolved nitrous oxide and nitrate concentrations for all catchments grouped by
539 hydrogeological type. Various includes limestone, chalk, sandstone and volcanic bedrock. Lines are
540 linear regressions; (b) Stable isotope composition of nitrogen and oxygen in nitrate for River Wensum
541 samples collected between May 2012 and May 2013. Diagonal line represents the denitrification
542 isotope effect (regression line for all data; $R^2=0.375$). Dark grey area delineates expected range of
543 isotopic composition from 'nitrification' nitrate in the Wensum catchment. Vertical and horizontal
544 arrows denote expected ranges of labelled processes.

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564 **Tables**

565

566 **Table 1:** Summary river water data for different catchments and contrasting hydrogeological types. Values
 567 presented as medians with one standard deviation in parentheses. Different superscript letters denote significant
 568 differences (*t*-test $p < 0.05$) between hydrogeological types within each catchment.

Catchment	Bedrock	Hydrogeological type	N samples	N ₂ O (µg N L ⁻¹)	NO ₃ ⁻ (mg N L ⁻¹)	Emission Factor (EF _{5r})	Indirect N ₂ O flux (kg N ha ⁻¹ a ⁻¹)
Wensum	Chalk	Unconfined	104	2.53 (0.92) ^a	9.21 (2.02) ^a	0.00030 (0.00015) ^a	9.75 (4.57) ^a
	Chalk	Semi-confined	104	1.31 (0.40) ^b	5.99 (1.81) ^b	0.00022 (0.00008) ^b	5.09 (2.03) ^b
	Chalk	Confined; glacial deposits	312	0.79 (0.26) ^c	5.20 (2.24) ^c	0.00016 (0.00014) ^c	2.72 (1.62) ^c
Eden	Limestone	Confined; glacial deposits	21	0.52 (0.09) ^a	8.61 (1.33) ^a	0.00007 (0.00001) ^a	1.50 (0.64) ^a
	Volcanics	Confined; glacial deposits	27	0.57 (0.34) ^a	-	-	1.72 (1.10) ^b
	Sandstone	Confined; glacial deposits	27	0.61 (0.44) ^b	4.52 (5.21) ^a	0.00019 (0.00010) ^b	2.19 (1.84) ^b
Avon	Chalk	Unconfined	29	16.83 (12.93) ^a	7.01 (0.41) ^a	0.00235 (0.00186) ^a	60.14 (51.98) ^a
	Mudstone	Unconfined	27	0.69 (0.95) ^b	4.46 (2.49) ^b	0.00020 (0.00057) ^b	1.95 (8.98) ^b
All	Chalk	Unconfined	133	3.03 (9.14) ^a	8.51 (2.02) ^a	0.00036 (0.00134) ^a	10.83 (33.70) ^a
	Chalk	Semi-confined	104	1.31 (0.40) ^b	5.99 (1.81) ^c	0.00022 (0.00008) ^b	5.09 (2.03) ^b
	Mudstone	Unconfined	27	0.69 (0.95) ^{bc}	4.46 (2.49) ^b	0.00020 (0.00056) ^a	1.95 (8.98) ^{bc}
	Various*	Confined; glacial deposits	387	0.76 (0.28) ^c	5.26 (2.65) ^d	0.00016 (0.00014) ^c	2.58 (4.62) ^c

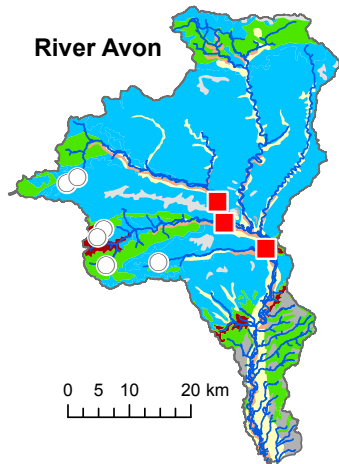
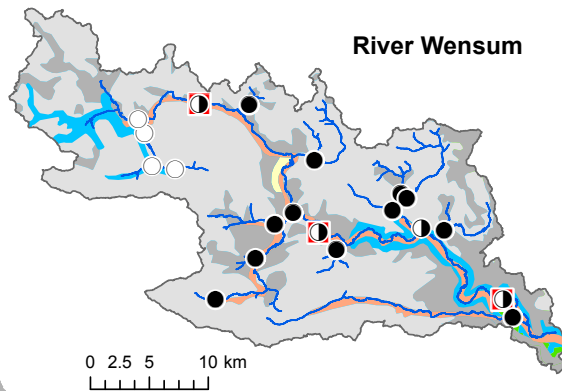
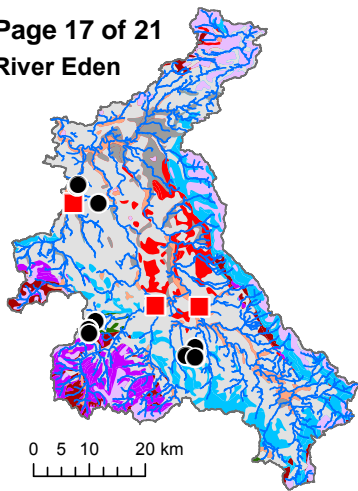
* includes limestone, chalk, sandstone and volcanic bedrock

569

570 **Table 2:** Seasonal variability in nitrogen dynamics for sites with contrasting hydrogeological types in the River
 571 Wensum catchment. Values presented as medians with one standard deviation in parentheses. Different
 572 superscript letters denote significant differences (*t*-test $p < 0.05$) between seasons with the same hydrogeological
 573 type.

Hydrogeological type	N samples	Parameter	Spring (MAM)	Summer (JJA)	Autumn (SON)	Winter (DJF)
Unconfined Chalk	104	N ₂ O (µg N L ⁻¹)	2.07 (0.90) ^a	3.06 (0.83) ^b	3.21 (0.93) ^b	2.47 (0.81) ^{ab}
		NO ₃ ⁻ (mg N L ⁻¹)	9.70 (1.44) ^a	8.22 (1.10) ^b	8.72 (2.64) ^{ab}	10.00 (2.12) ^a
		EF _{5r}	0.00021 (0.00010) ^a	0.00039 (0.00011) ^b	0.00039 (0.00020) ^b	0.00027 (0.00009) ^a
		Flux (kg N ha ⁻¹ a ⁻¹)	8.14(5.11) ^{ab}	9.06 (2.90) ^b	10.40 (3.84) ^{ab}	11.82 (5.15) ^a
Semi-confined Chalk	104	N ₂ O (µg N L ⁻¹)	1.10 (0.36) ^a	1.42 (0.37) ^b	1.39 (0.45) ^b	1.26 (0.38) ^{ab}
		NO ₃ ⁻ (mg N L ⁻¹)	6.68 (1.90) ^a	4.60 (1.62) ^b	5.90 (1.41) ^a	6.71 (1.84) ^a
		EF _{5r}	0.00019 (0.00005) ^a	0.00030 (0.00009) ^b	0.00025 (0.00005) ^c	0.00020 (0.00006) ^a
		Flux (kg N ha ⁻¹ a ⁻¹)	5.21 (2.07) ^a	4.19 (1.61) ^b	4.31 (1.99) ^{ab}	5.89 (2.00) ^c
Confined Chalk	312	N ₂ O (µg N L ⁻¹)	0.75 (0.24) ^a	0.74 (0.26) ^a	0.79 (0.24) ^a	0.87 (0.29) ^b
		NO ₃ ⁻ (mg N L ⁻¹)	5.81 (1.90) ^a	4.31 (2.27) ^b	4.72 (2.39) ^b	5.59 (2.21) ^a
		EF _{5r}	0.00015 (0.00006) ^a	0.00019 (0.00024) ^b	0.00016 (0.00013) ^{bc}	0.00016 (0.00008) ^c
		Flux (kg N ha ⁻¹ a ⁻¹)	2.81 (1.46) ^a	2.01 (1.02) ^b	2.48 (0.89) ^c	3.94 (1.93) ^d

574



Sampling Location Hydrogeology

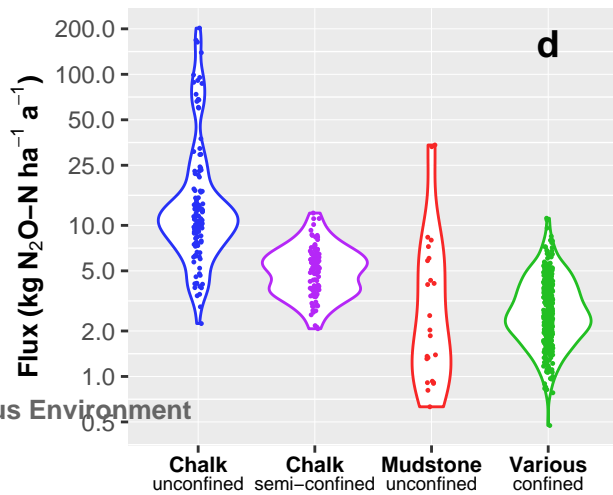
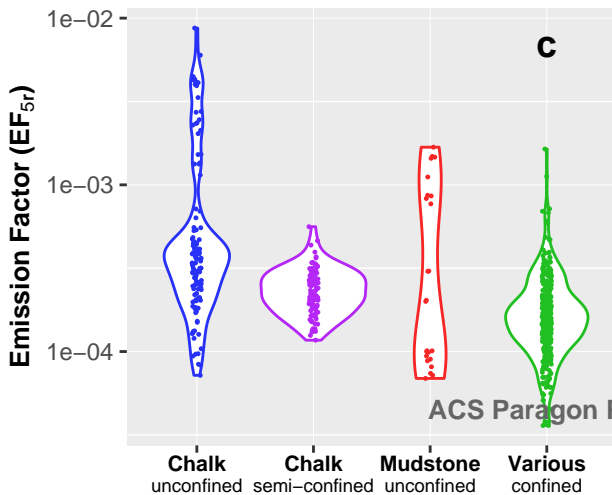
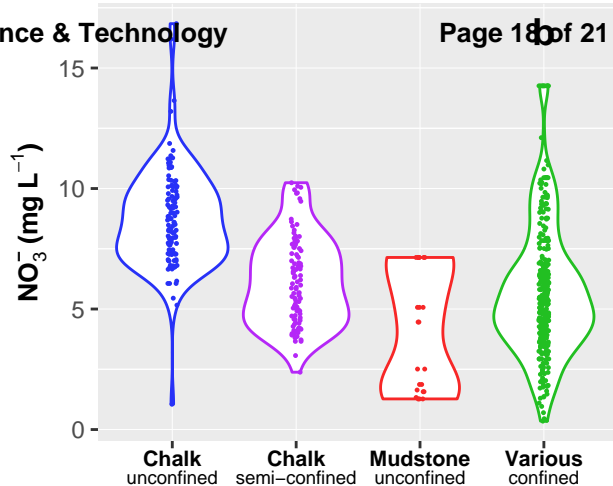
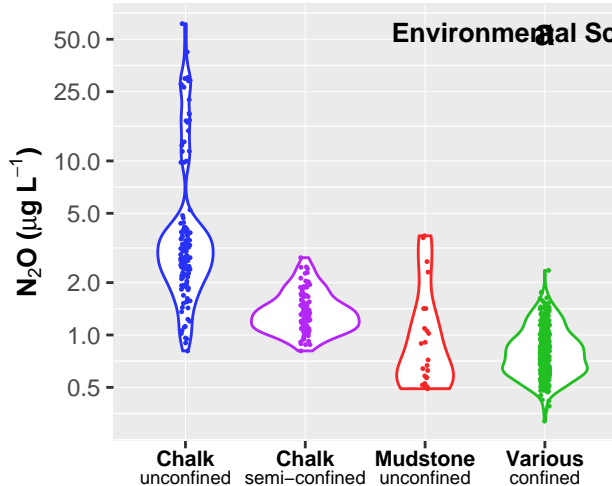
- Unconfined
- ◐ Semi-confined
- Confined
- Gauging station

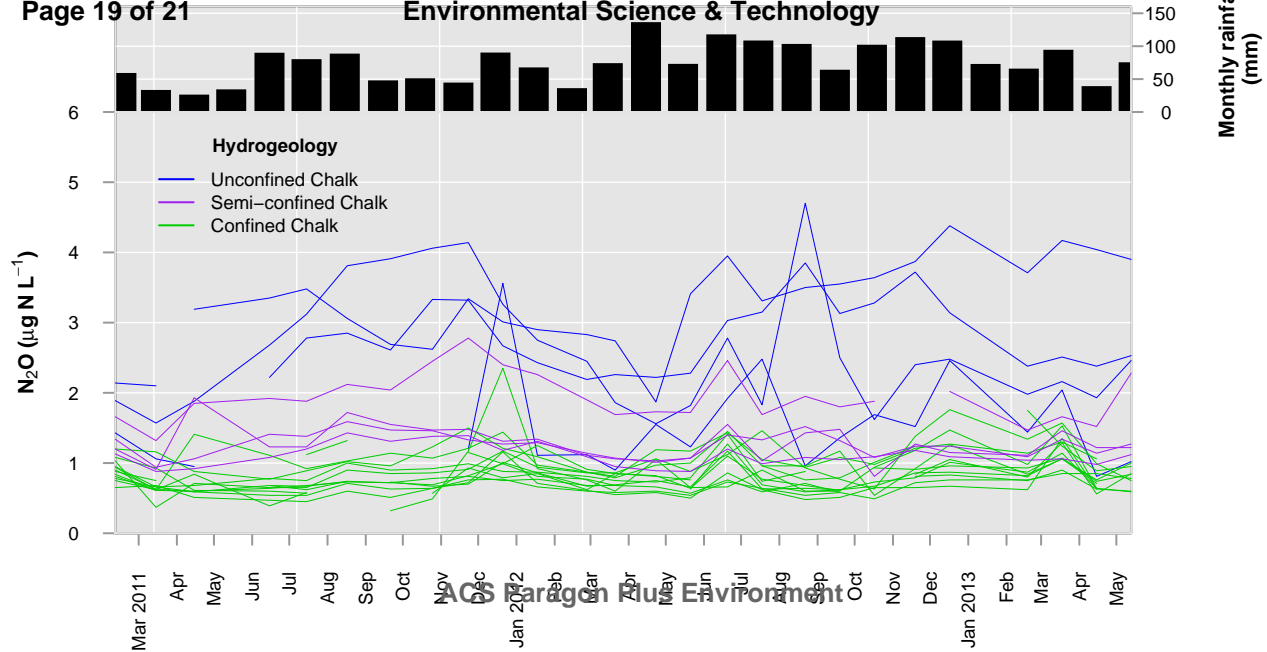
Superficial Geology

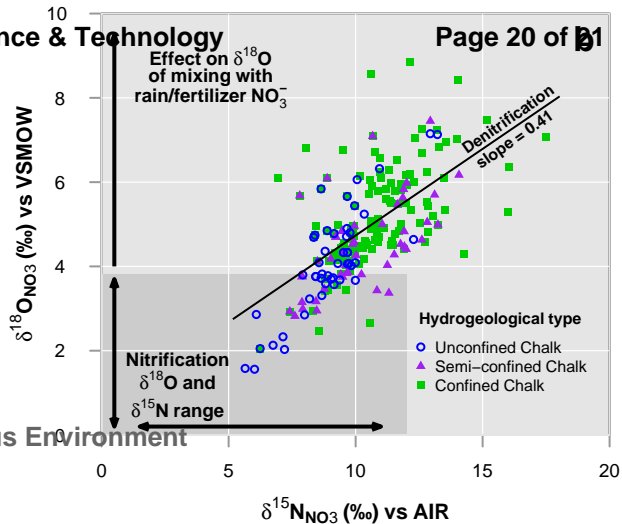
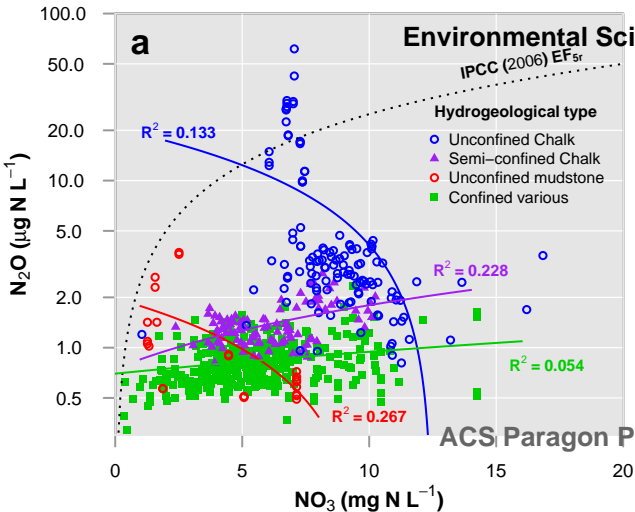
- Sand and gravel
- Diamicton till
- River terrace deposits
- Alluvium
- Peat

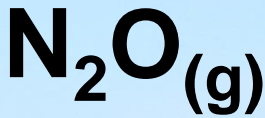
Bedrock Geology

- Limestone/Chalk
- Mixed sedimentary
- Sandstone
- Mudstone
- Volcanics
- Conglomerate









$$EF_{5r} = \text{N}_2\text{O-N} / \text{NO}_3\text{-N}_{(\text{L})}$$

ACS Paragon Plus Environment

River Wensum, UK