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### **Factors affecting the efficiency of aerosolized salbutamol delivery via a metered dose inhaler and equine spacer device**

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1 **Title:** Factors affecting the efficiency of aerosolised salbutamol delivery via a metered dose  
2 inhaler and equine spacer device

3 **Short running title:** Efficiency of aerosolised salbutamol delivery

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13

14 **Abstract**

15 Despite frequent use of metered dose inhalers (MDIs) and spacers in equine practice, limited  
16 information exists on the efficiency of aerosol delivery using such devices. We determined the  
17 particle size distribution within an MDI-generated salbutamol aerosol delivered via an equine  
18 spacer using “best practice” delivery technique and assessed the effect of variations in MDI  
19 use technique (shaking prior to each actuation, rapid repetitive actuations and MDI angulation)  
20 on aerosol delivery efficiency.

21 Under optimal conditions, only 53( $\pm$ 18) microgrammes ( $\mu$ g) salbutamol per 100 $\mu$ g actuation  
22 was delivered beyond the spacer. Although this aerosol had a high (89.6% [ $\pm$ 2.4]) fine particle  
23 (<5 micron [ $\mu$ m]) fraction, and a low mass median aerodynamic diameter (2.52 [ $\pm$ 0.29] $\mu$ m)  
24 and particle size variability (geometric SD - 1.66 [ $\pm$ 0.16] $\mu$ m), within all particle size fractions  
25 there was a high coefficient of variance (31-79%) of the percentage salbutamol delivered  
26 between experimental runs, thus impeding any effort to predict drug delivery to the patient  
27 during equine inhalation therapy. Despite observable trends and with the exception of minor  
28 statistically significant changes in the least abundant particle sizes, none of the deviations from  
29 a “best practice” delivery technique significantly altered the relative salbutamol delivery  
30 beyond the spacer, a finding which has potential relevance with regard to maintaining user  
31 compliance.

32

33 **Keywords:** horse, MDI, inhalation, aerosol, nebuliser

## 34 **Introduction**

35 The use of inhalation therapy in equine practice has recently increased in popularity,  
36 particularly in relation to corticosteroid and bronchodilator treatment of equine asthma  
37 (Robinson et al., 1993; Tesarowski et al., 1994; Derksen et al., 1996; Derksen et al., 1999;  
38 Durham, 2001) but also for the delivery of other therapeutic agents including antibiotics (Art  
39 et al., 2010; Burton et al., 2013; Ferrucci et al., 2013; Fultz et al., 2015). The proposed  
40 advantages over systemic drug delivery include a relatively lower cost, drug delivery directly  
41 to the site of action and, particularly in the case of corticosteroids, a reduced risk of systemic  
42 adverse effects (Hoffman, 1997, Duvivier et al., 1997; Duvivier et al., 1999; Lavoie, 2001).  
43 Various means of aerosol generation exist, including ultrasonic, jet and mesh nebulisation and  
44 metered dose inhalers (MDIs), each differing with respect to the variability in aerosol particle  
45 size distribution (Duvivier et al., 1997; Votion et al., 1997; Duvivier et al., 1999). Furthermore,  
46 a variety of delivery devices are available, including equine-specific and customised spacers  
47 and airtight facemasks, the use of which is indicated largely due to the inability to accurately  
48 synchronise aerosol generation with inspiration in the horse (Lavoie, 2001).

49 Although successful drug delivery to the peripheral airways is partly dependent the patient's  
50 breathing pattern and the viscosity, density, surface tension and hygroscopic growth potential  
51 of the drug solution (Silverman, 1990; Morrow, 1996), ultimately the aerodynamic diameter  
52 of the aerosolised particles is the major determinant of peripheral airway deposition  
53 (Stahlhofen, 1980). Despite the small size and low variability of the aerosolised particles  
54 generated by MDIs (Kim et al., 1985), there are a variety of factors which can significantly  
55 influence MDI-generated aerosol delivery to peripheral airways. In human respiratory  
56 medicine, this has led to established protocols for MDI use (Everard et al., 1995); protocols  
57 which have subsequently been applied to the field of equine inhalation therapy. However,  
58 despite many of the recommendations deriving from *in vitro* studies, there remains a lack of

59 concordance between the standard protocols used in human respiratory medicine and those  
60 employed in the laboratory setting (Everard et al., 1995). Such inconsistency has the potential  
61 to result in unnecessary recommendations being made to MDI users which may have a negative  
62 impact on patient compliance with appropriate self-medication. Such negative impact is likely  
63 to be amplified when unnecessary recommendations result in an extended duration of  
64 treatment, a significant consideration with equine inhalation therapy when multiple actuations  
65 are generally required. Recommendations which may significantly extend the duration of  
66 treatment include shaking the MDI prior to each actuation when multiple actuations are  
67 required and the avoidance of rapidly performed consecutive actuations (Everard et al., 1995;  
68 Wildhaber, 1996).

69 The limited data relating to drug delivery via an equine-specific spacer are largely derived from  
70 *in vivo* scintigraphic studies which revealed relatively poor and markedly varied aerosol  
71 delivery to the peripheral airways (Votion et al., 1997; Rush et al., 1999; Votion et al., 1999).  
72 Due to the lack of published *in vitro* studies on MDI-generated aerosol characteristics using  
73 equine spacers, this study was designed to measure the efficiency of delivery and particle size  
74 distribution of an MDI-generated salbutamol aerosol delivered via an equine spacer device.  
75 Specific deviations, regarded as having potential influence on owner compliance with respect  
76 to MDI use, from a “best practice” protocol, were evaluated in relation to their effect on the  
77 efficiency of aerosolised drug delivery.

78

## 79 **Materials and methods**

80 Three sets of comparative experiments were conducted within the study, each with a measured  
81 output of aerosolised salbutamol delivery to the various stages of a next generation impactor  
82 (NGI)<sup>a</sup>, as follows: *Experiment 1* - Effect of shaking the MDI prior to each sequential actuation;

83 *Experiment 2* - Effect of angulation of the MDI device within the spacer; *Experiment 3*; Effect  
84 of multiple actuations in rapid succession. Additionally, in light of the variability in data  
85 derived from experiment 1, selected data from experiments 2 and 3 were also used to measure  
86 the efficiency of salbutamol delivery with the MDI device secured in an optimal position  
87 relative to the spacer device (*Optimal delivery measurement*), whereby salbutamol retention  
88 within the spacer was also measured.

89

### 90 *Salbutamol aerosol generation and delivery to the NGI*

91 For comparative purposes, the quantity of aerosolised salbutamol delivered was determined by  
92 the number of 100µg actuations of the MDI<sup>b</sup> directly into an equine spacer<sup>c</sup> (**Figure 1a**). The  
93 spacer was connected to the throat of the NGI, a high-performance, precision, particle  
94 classifying cascade impactor designed for testing MDIs, dry powder inhalers, nebulizers and  
95 nasal sprays, separating aerosolised particles based on particle size and aerodynamic properties.  
96 The NGI is comprised of a throat (designed to mimic the calibre and airflow directional changes  
97 within the human upper airway) and a series of eight stages, characterised by different pore  
98 sizes of sequentially decreasing diameter, thus simulating the sequential decrease in airway  
99 diameter from the trachea to the terminal bronchioles (**Figure 1b**). Consequently, aerosolised  
100 particles delivered into the NGI are fractionated and collected onto each of these stages. The  
101 distal portal of the NGI was connected to a vacuum pump<sup>d</sup>, calibrated to generate a constant  
102 flow rate of 60L/min through the entire system (spacer, throat, NGI and all connecting tubing).  
103 Leaks within the system were prevented by sealing all connections with parafilm<sup>e</sup> and the  
104 absence of leaks was confirmed by comparing airflow rate before and after each experimental  
105 run. Airflow was maintained for 30s after each experimental run. With a constant airflow of  
106 60L/min, the stage effective cut off particle diameters (at 50% efficiency) were as follows:

107 stage 1 - 8.06 $\mu$ m; stage 2 – 4.46 $\mu$ m; stage 3 – 2.82 $\mu$ m; stage 4 – 1.66 $\mu$ m; stage 5 – 0.94 $\mu$ m;  
108 stage 6 – 0.55 $\mu$ m; stage 7 – 0.34 $\mu$ m; stage 8 - 0 $\mu$ m.

109

110 Following aerosol delivery, samples were retrieved from the spacer (*Experiments 2 and 3* only),  
111 throat and each collection stage by instilling 10ml distilled water, re-suspending any deposited  
112 salbutamol with a cell scraper, pipetting into a labelled container and storing at 4°C until further  
113 analysis. The decision to measure salbutamol deposition within the spacer was made following  
114 completion of experiment 1 which revealed a relatively low drug delivery to the NGI. This  
115 additional sample collection was conducted to determine whether, and to what extent, this low  
116 output reflected retention within the spacer. A separate pipette was used for each sample to  
117 reduce the risk of cross contamination. Following sample collection, the spacer, NGI stages  
118 and throat were washed in dilute detergent and rinsed with distilled water before being air dried,  
119 to avoid accumulation of static electricity. Prior to each experiment, the NGI, NGI stages, and  
120 throat were refrigerated at 4°C for 1h to minimise subsequent evaporative losses. Between  
121 experiments, the MDI was stored up-right at room temperature.

122

### 123 *Salbutamol assay*

124 Standard salbutamol concentrations (0 to 100 $\mu$ g/ml) were prepared from the stock solution  
125 (10mg/ml salbutamol hemisulphate salt<sup>e</sup> in distilled water) and 100 $\mu$ l of standard and sample  
126 (spacer, throat and NGI stages) was pipetted in duplicate into wells of a UV-clear flat bottom  
127 microwell plate<sup>f</sup>. Absorbance was read at 224nm and standard and sample concentrations  
128 calculated using multi-detection microplate data collection and analysis software<sup>g</sup>. A mean  
129 value of duplicate results showing acceptable agreement was used for subsequent statistical  
130 analyses.

131

132 *Experimental designs*

133 Within each set of comparative experiments, the order of runs was randomised. For each  
134 experiment, the MDI device was shaken for 30s prior to each run and two ‘waste’ actuations  
135 were performed prior to connecting the MDI to the spacer. Constant airflow was established  
136 prior to aerosol generation. All comparative experiments involved 8 repetitions of the delivery  
137 of 10 x 100µg actuations (total 1mg salbutamol) with the exception of *Experiment 3* (effect of  
138 rapid actuations), whereby 8 repetitions of 8 x 100µg actuations (total 0.8mg salbutamol) were  
139 delivered .The experimental designs are summarised in Table 1. Briefly, *Experiment 1*  
140 compared 8 repeats of 10 actuations delivered at 5 s intervals without removing the MDI from  
141 the spacer with 8 repeats of 10 actuations, each preceded by a 30s period of MDI shaking;  
142 *Experiment 2* compared 3 sets of 8 repeats of 10 actuations delivered at 5s intervals; each set  
143 differing with respect to the direction of actuation within the spacer (with the output nozzle  
144 horizontal or at 10° or 20° above the horizontal) (Figure 1c); *Experiment 3* compared 3 sets of  
145 8 repeats of 8 actuations (MDI actuated in a horizontal direction), either delivered individually  
146 at 5s intervals, as 4 x double actuations in rapid succession (approximately 2 actuations per  
147 second) or as 2 x quadruple actuations in rapid succession (approximately 2 actuations per  
148 second); the *Optimal Delivery Experiment* measured the efficiency of salbutamol delivery  
149 under presumed optimal delivery conditions using selected data derived from *Experiments 2*  
150 and 3 (horizontal actuation of the MDI and 5s interval between individual actuations).

151

152 **Statistical analyses**



153 For the comparative delivery experiments, values are presented as, and analyses applied to,  
154 measured salbutamol expressed as a percentage of the anticipated total aerosolised salbutamol  
155 actuated (median and range) (*Experiments 1 and 2* – 1mg; *Experiment 3* – 800µg). When only  
156 2 experimental conditions were compared, a Mann Whitney test for non-parametric data was  
157 applied directly. When more than 2 experimental conditions were compared, a Mann Whitney  
158 test for non-parametric data was applied only if differences were revealed by a Kruskal-Wallis  
159 analysis. Significance was assumed at  $P < 0.05$ . For the *Optimal Delivery Experiment*,  
160 salbutamol delivery is expressed as both percentage (median and range) anticipated total  
161 aerosolised salbutamol per series of actuations and micrograms salbutamol per actuation (mean  
162 and SD). The fine ( $< 5\mu\text{m}$ ) particle fraction is expressed as a percentage (mean and SD) per  
163 actuation and the mean aerodynamic particle size is expressed in  $\mu\text{m}$  (mean and SD) per  
164 actuation.

165

## 166 **Results**

### 167 *Optimal delivery measurement*

168 Data relating to percentage of the anticipated total aerosolised salbutamol (assuming 100µg per  
169 actuation) deposited within the spacer and the different stages of the NGI are summarised in  
170 **Figure 2**. The greatest deposition of aerosolised salbutamol was within stage 4 (23% [8-33]) of  
171 the NGI, the spacer (21% [8-32]) and stage 3 (17% [6-31]) of the NGI, followed by stages 5  
172 (6% [3-11]) and 2 (6% [2-10]).

173 The mean ( $\pm$  SD) measured output (per single 100µg actuation) from the MDI was  $75 \pm 16\mu\text{g}$ ,  
174 with a mean calculated aerosol delivery to the NGI of  $53 \pm 18\mu\text{g}$ , of which,  $48 \pm 16\mu\text{g}$  was within  
175 the “fine particle” ( $< 5\mu\text{m}$ ) range, equating to a fine particle fraction of  $89.6 \pm 2.4\%$ . The mass  
176 median aerodynamic diameter (MMAD) of the aerosol, calculated over 16 repetitions (8 from

177 *Experiments 2 and 3, respectively*), was  $2.52 \pm 0.29 \mu\text{m}$  (namely 50% of the total sample mass  
178 was present in particles with aerodynamic diameters  $< 2.5 \mu\text{m}$ , and 50% was present in particles  
179 having an aerodynamic diameter  $> 2.52 \mu\text{m}$ , with a geometric standard deviation of  
180  $1.66 \pm 0.16 \mu\text{m}$ .

181

182 *Experiment 1: Effect of shaking the MDI prior to each sequential actuation*

183 There was no significant difference between shaking the MDI at the beginning of 10 actuations  
184 and shaking the MDI prior to each of the 10 actuations with regard to percentage of total  
185 salbutamol delivered to the NGI (43% [20-66] versus 41% [17-60], respectively) or percentage  
186 of total salbutamol delivered to each stage of the NGI (Figure 3).

187

188 *Experiment 2: Effect of angulation of the MDI within the spacer*

189 Compared with a horizontal orientation of MDI output nozzle, there was no significant effect  
190 of the other MDI angulations ( $10^\circ$  and  $20^\circ$  upward deviation) on percent salbutamol delivered  
191 to the spacer, the NGI or the NGI and spacer combined. There was a statistically significant,  
192 yet small effect of MDI angulation on the percent salbutamol delivered to stage 8 ( $P=0.035$ ) of  
193 the NGI, whereby the  $10^\circ$  angulation resulted in significantly ( $P=0.005$ ) less salbutamol  
194 delivery than the horizontal orientation (0.4% [0-0.7] vs 0.8% [0.4-1.4];  $P=0.005$ ); otherwise  
195 there was no significant effect of MDI angulation on salbutamol delivery to any of the NGI  
196 stages (Figure 4).

197 When considering only the median drug delivery calculated from the 3 experimental  
198 conditions, increasing the MDI angle from a horizontal orientation to a  $20^\circ$  upward deviation  
199 resulted in a 15% reduction in total output to the NGI and a 24% increase in retention within  
200 the spacer.

201 *Experiment 3: Effect of multiple rapid MDI actuations on salbutamol delivery*

202 Compared with 8 single actuations, there was no significant effect of multiple rapid actuations  
203 (4x2 or 2x4) on percent salbutamol delivered to the spacer, the NGI or the NGI and spacer  
204 combined. There was a statistically significant, yet small effect of multiple rapid actuations on  
205 the percent salbutamol delivered to stages 1 (P=0.007), 2 (P=0.032) and 8 (P=0.032) of the  
206 NGI (Figure 5). Four x 2 rapid actuations resulted in significantly less salbutamol delivery to  
207 stages 1 (0.7% [0-1.1] vs 1.1% [0.8-1.5]; P=0.021) and 2 (4.9% [2.4-5.7] vs 6.8% [4.4-8.3];  
208 P=0.01) than 8 x single actuations. Two x 4 rapid actuations resulted in significantly less  
209 salbutamol delivery to stages 1 (0.1% [0-1.1] vs 1.1% [0.8-1.5]; P=0.007) and 8 (0% [0-1.0] vs  
210 0.5% [0-1.0]; P=0.025) than 8 x single actuations and to stage 8 than 4 x 2 rapid actuations (0%  
211 [0-1.0] vs 0.3% [0-1.0]; P=0.021).

212 When considering only the median drug delivery calculated from the 3 experimental  
213 conditions, 4 sets of double rapid actuations resulted in a 15% reduction in total MDI output,  
214 16% reduction in drug delivery to the NGI and 14% reduction to stages 3 and 4 of the NGI. In  
215 comparison, 2 sets of quadruple rapid actuations resulted in a 21% reduction in total MDI  
216 output, 24% reduction in drug delivery to the NGI and 21% reduction to stages 3 and 4 of the  
217 NGI.

218

219 **Discussion**

220 Despite the increasing popularity of inhalation therapy in the horse, the results of this study  
221 highlighted a variety of important considerations with this mode of drug delivery. Importantly,  
222 only half of the anticipated MDI output was detected within the NGI. Although a significant  
223 proportion of the deficit could be explained by drug retention within the spacer, there remained  
224 a proportion which could not be accounted for following sampling from all NGI stages

225 (including the throat). Therefore, either the MDI did not always achieve a 100µg output during  
226 each actuation or drug was deposited within other components of the system which were not  
227 subsequently sampled or there was a failure to optimally solubilise all precipitated drug within  
228 each NGI component. It is unlikely that significant losses occurred within the tubing between  
229 the spacer and NGI. In contrast, significant drug losses may have occurred around the exit  
230 nozzle of the MDI because a white residue was often visible at this site during cleaning of the  
231 MDI prior to each experiment. Importantly, losses could not be attributed to drug depletion  
232 within the MDI as the number of actuations per MDI device were recorded and the MDI  
233 replaced well in advance of the calculated drug depletion threshold. This is an important  
234 consideration during therapeutic use of such devices as the drug will often become depleted  
235 prior to depletion of the propellant (Rubin & Durotoye, 2004).

236 Despite significant losses within the spacer, the drug delivered to the NGI had a consistently  
237 high small particle fraction, with almost 90% of particles being less than 5µm. Furthermore,  
238 the calculated MMAD of the aerosol consistently approximated 2.5µm, indicating that 50% of  
239 the total sample mass was present in particles with aerodynamic diameters less than 2.5µm and  
240 50% was present in particles having an aerodynamic diameter greater than 2.5µm, with a  
241 geometric standard deviation (GSD) of  $1.66 \pm 0.16 \mu\text{m}$ . This narrow range of particle size  
242 distribution is predicted with an MDI device and contrasts with the more heterodispersed  
243 distribution associated with other methods of drug aerosolisation (e.g. ultrasonic, and mesh  
244 nebulisation). For example, using the same experimental set up, the authors have demonstrated  
245 the generation of an aerosol with a MMAD of 1.4µm and a GSD of 3.2µm using an active mesh  
246 nebuliser device<sup>i</sup> commonly used in equine practice (*unpublished observations*).

247

248 Although the MDI-generated particle size distribution was considered to be optimal for drug  
249 delivery to the smaller airways, it should be emphasised that such assumptions, as they relate  
250 to equine inhalation therapy, are largely based on human patient derived data. With regard to  
251 the prediction of the likelihood of an aerosol penetrating each region of the human respiratory  
252 tract, The American Conference of Governmental Industrial Hygienists (ACGIH) describes  
253 three fractions (inhalable, thoracic, respirable) generally defined by the aerodynamic diameter  
254 at which 50% penetration of that fraction occurs (50% cut-point), with the 50% cut-point for  
255 the respirable fraction generally assumed to be 4 $\mu$ m. The likelihood of significant differences  
256 between the size-dependent penetration of particles into the equine lung and the human lung  
257 has been proposed (Ivester et al., 2014). Although the obligate nasal breathing strategy of  
258 horses may predominantly influence the deposition of larger particles, other differences may  
259 bring into question the appropriateness of applying human derived data to the horse in relation  
260 to the deposition of smaller particles, such as those generated by a MDI. These include the  
261 considerably greater (10 to 12-fold) resting tidal volume in the horse and its role in determining  
262 linear flow rates within the respiratory tract, with a resultant effect on particle impaction  
263 (Ivester et al., 2014). However, in the absence of experimental data to define equine-specific  
264 particle fractions or detailed anatomic descriptions of airway dimensions which would permit  
265 the construction of predictive models of particle penetration, it is generally assumed that  
266 particles less than 4-5 $\mu$ m are likely to reach the lower airways in the horse (Hoffman, 1997;  
267 Lavoie, 2001).

268

269 An airflow of 60L/min was used as it more closely approximated the minute volume of an adult  
270 horse. The calculation of the particle characteristics (e.g. MMAD and fine particle fraction)  
271 was reliant on a constant flow rate through the system and the flow rate applied determines the  
272 region of particle deposition within the NGI. However, this differs markedly from the

273 fluctuating inspiratory flow rates associated with tidal breathing at rest which can typically  
274 reach peaks of 120-240L/min. It is likely that the application of a variable flow rate would have  
275 had some influence on the degree of drug delivery to the NGI, although the nature of this  
276 influence is difficult to predict. Peaks in fluctuating airflow may promote particle impaction at  
277 the NGI throat, thus reducing delivery; alternatively, periods of zero flow may facilitate aerosol  
278 suspension within the spacer, thus increasing delivery (Duvivier et al., 1997). In human  
279 respiratory medicine, the generation of a slow inspiratory flow rate immediately following  
280 actuation is recommended to maximise particle delivery to the peripheral airways (Everard et  
281 al., 1995; Wildhaber et al., 1996).

282

283 Even under optimal delivery conditions, this study revealed a significant degree of variation  
284 both in drug retention in the spacer and drug delivery to all stages of the NGI. This variation  
285 could not be attributed to repeated use of the MDI as no association was detected between drug  
286 output during each series of actuations and the total number of previous actuations of the device  
287 (*data not shown*). Although prior knowledge of the predicted losses prior to aerosol delivery to  
288 the patient (e.g. within the spacer) will permit some degree of compensation (i.e. delivery of a  
289 larger dose), it is not possible to compensate for the unknown delivery achieved with each  
290 actuation or series of actuations. The clinical significance of this variation is greatest in relation  
291 to the sites of greatest particle deposition; namely within the spacer (CoV - 32%) and stages 3  
292 and 4 (CoV – 49 and 41%, respectively). This equated to a 5-fold difference between the lowest  
293 and highest deposition in stages 3 and 4 out of the 16 repetitions performed in the *Optimal*  
294 *Delivery Experiment*. Such variation in delivery has previously been reported by Votion et al.,  
295 (1997) in relation to both ultrasonic nebulisation and jet aerosol delivery and by Janssens et al.,  
296 (1999) in relation to MDI delivery via a spacer device in asthmatic children, whereby  
297 coefficient of variance values ranging from 23 to 37% were reported, depending on the spacers

298 used. Such variation will inevitably render any efforts to make accurate dosing  
299 recommendations problematic; consequently, recommended doses should be used only as  
300 guidelines and the drug should ultimately be administered “to effect” (Lavoie, 2001).

301 All the data used for the *optimal delivery experiment* were derived from experiments 2 and 3,  
302 whereby a constant horizontal orientation of the MDI nozzle was maintained within the spacer.  
303 Therefore the variation in both aerosol delivery to the NGI and retention within the spacer  
304 could not be attributed to the occasional actuation in a suboptimal direction resulting in the  
305 high velocity propulsion of drug directly onto the inner surface of the spacer. Furthermore, the  
306 coefficient of variance of drug delivered to the NGI in experiment 1, whereby the MDI device  
307 was not secured in position, was no greater than that derived from the data included in the  
308 *Optimal Delivery Experiment*. Indeed, the results of experiment 2 confirmed that a 20°  
309 deviation from the optimal direction of actuation failed to significantly alter the percent  
310 salbutamol delivered to the spacer, the NGI or the NGI and spacer combined. Despite this lack  
311 of statistical significance, which may partly be attributable to the wide variation in drug  
312 delivery between each series of actuations, there was an obvious trend towards a lower drug  
313 delivery to the NGI and a greater drug retention within the spacer with increasing angulation  
314 of the MDI device.

315 Owner compliance with respect to the correct use of the MDI device constitutes a major factor  
316 in the likelihood of success of inhalation therapy in the horse. Consequently, instructions are  
317 regularly provided by the attending clinician, highlighting the “dos” and “don’ts” of MDI and  
318 spacer use which are largely based on recommendations applied within the medical profession.  
319 These generally involve factors such as shaking the MDI prior to each actuation, exhaling fully  
320 prior to actuation, holding the MDI vertically, coinciding actuation with inspiration, adopting  
321 a slow inspiratory effort, initiating only a single actuation per breath, and subsequent breath  
322 holding for a minimum of 5s (Resnick et al., 1996). However, despite the relative simplicity of

323 these steps, several studies have revealed poor knowledge of correct MDI use protocol,  
324 particularly amongst medical professionals (Jones et al., 1995; Resnick et al., 1996; Stelmach  
325 et al., 2007). Furthermore, certain studies have identified particular recommendations to be  
326 inappropriate and potentially detrimental with regard to their potential influence on user  
327 compliance (Everard et al., 1995). Such recommendations are likely to have greater influences  
328 on compliance when they significantly increase the time required for drug administration;  
329 particularly in equine inhalation therapy when multiple actuations are required.

330 Everard et al., (1995) clearly demonstrated the importance of MDI shaking prior to drug  
331 administration, likely reflecting the importance of mixing the active drug and the propellant  
332 within the MDI device. However, the current study failed to demonstrate any significant benefit  
333 of shaking the MDI device before each actuation in a series of sequential actuations, in relation  
334 to both total and respirable particle delivery to the NGI. This finding likely reflects an  
335 insufficient time period (5s) between each actuation to permit separation of the salbutamol and  
336 the propellant. Everard et al., (1995) also demonstrated a reduction in both total and respirable  
337 particle generation with multiple actuations in rapid succession. Although the current study  
338 failed to identify a statistically significant effect of rapid double and quadruple actuations on  
339 drug delivery to the spacer, NGI or spacer and NGI combined, there was a trend for increased  
340 rapid sequential actuations to reduce drug delivery. However, in light of the small magnitude  
341 of the reduction, this could readily be compensated for by a small increase the number of  
342 actuations; for example, an extra actuation for every 4 rapid consecutive actuations.

343 In conclusion, this study demonstrates the difficulties in predicting the magnitude of drug  
344 delivery to the peripheral airways using an MDI and equine spacer device. Therefore, when  
345 selecting the most appropriate route of drug administration, this shortcoming must be  
346 considered and weighed up against the advantages of this therapeutic approach, including the  
347 reduced risk of systemic adverse drug effects and relatively lower drug costs. Furthermore, the



348 apparent lack of requirement to shake the MDI prior to each actuation and the limited effect of  
349 multiple rapid actuations can significantly reduce the time required to administer the treatment  
350 and therefore have the potential to improve owner compliance with regard to MDI use in the  
351 horse. Finally, it is important to appreciate that any conclusions derived from this study can  
352 only be applied clinically to the use of salbutamol. It remains unknown whether similar results  
353 would be obtained with other drug aerosols generated by an MDI device (e.g. corticosteroids)  
354 as differences in viscosity, density and surface tension have the potential to influence both the  
355 particle size distribution of the aerosol generated as well as the rapidity with which the drug  
356 and propellant separate between actuations.

357

## 358 **Manufacturers**

359 <sup>a</sup> Next Generation Impactor, Copley Scientific, Colwick Quays Business Park, Private Road No. 2,  
360 Colwick, Nottingham, NG4 2JY, United Kingdom

361 <sup>b</sup> Ventolin® Evohaler® 100 micrograms, Glaxo Wellcome UK Limited, Stockley Park, West Uxbridge Middlesex  
362 UB11 1BT

363 <sup>c</sup> Equine Haler®, Jørgen Kruuse A/S, Havretoften 4 DK-5550 Langeskov Denmark

364 <sup>d</sup> Copley High Capacity Pump, Copley Scientific, Colwick Quays Business Park, Private Road No. 2,  
365 Colwick, Nottingham, NG4 2JY, United Kingdom

366 <sup>e</sup> S5013 Sigma-Aldrich Company Ltd. Dorset, England

367 <sup>f</sup> Corning® 3675 96 well plates, UV-transparent , Sigma-Aldrich Company Ltd. Dorset, England

368 <sup>g</sup> Synergy HT Biotek, BioTek Instruments Inc 2005, Papermakers House, Rivenhall Road, Swindon SN5 7BD,  
369 United Kingdom

370 <sup>h</sup> Copley Inhaler Testing Data Analysis software (CITDAS), Copley Scientific, Colwick Quays Business Park,  
371 Private Road No. 2, Colwick, Nottingham, NG4 2JY, United Kingdom

372 <sup>i</sup> Flexineb, Nortev, Unit 18, Claregalway Corporate Park, Galway H91 KFX3, Ireland

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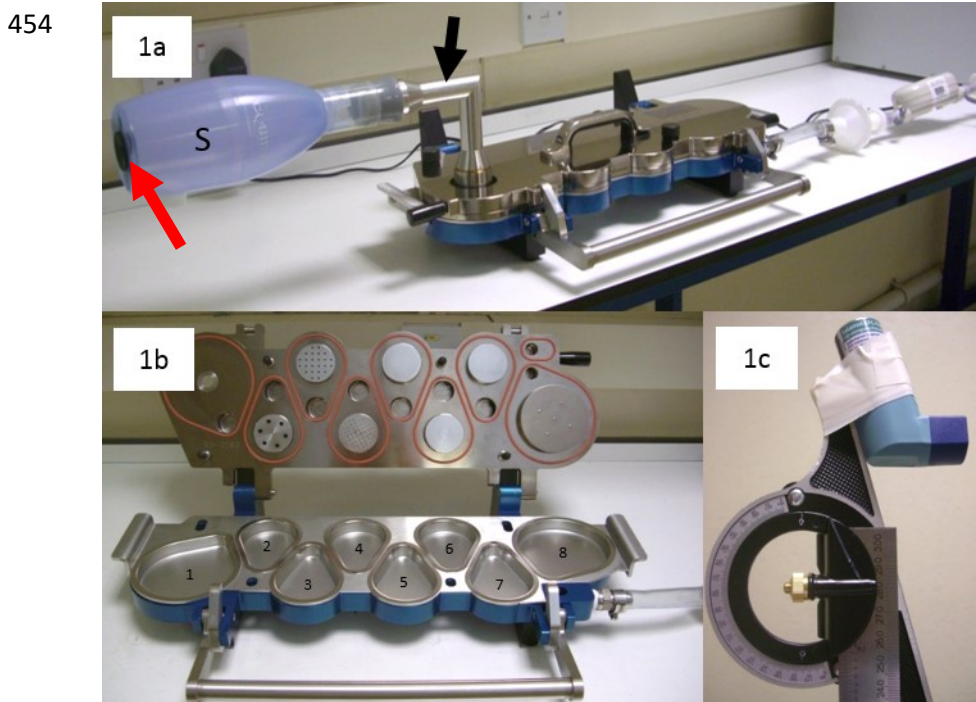
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444 **Figures**

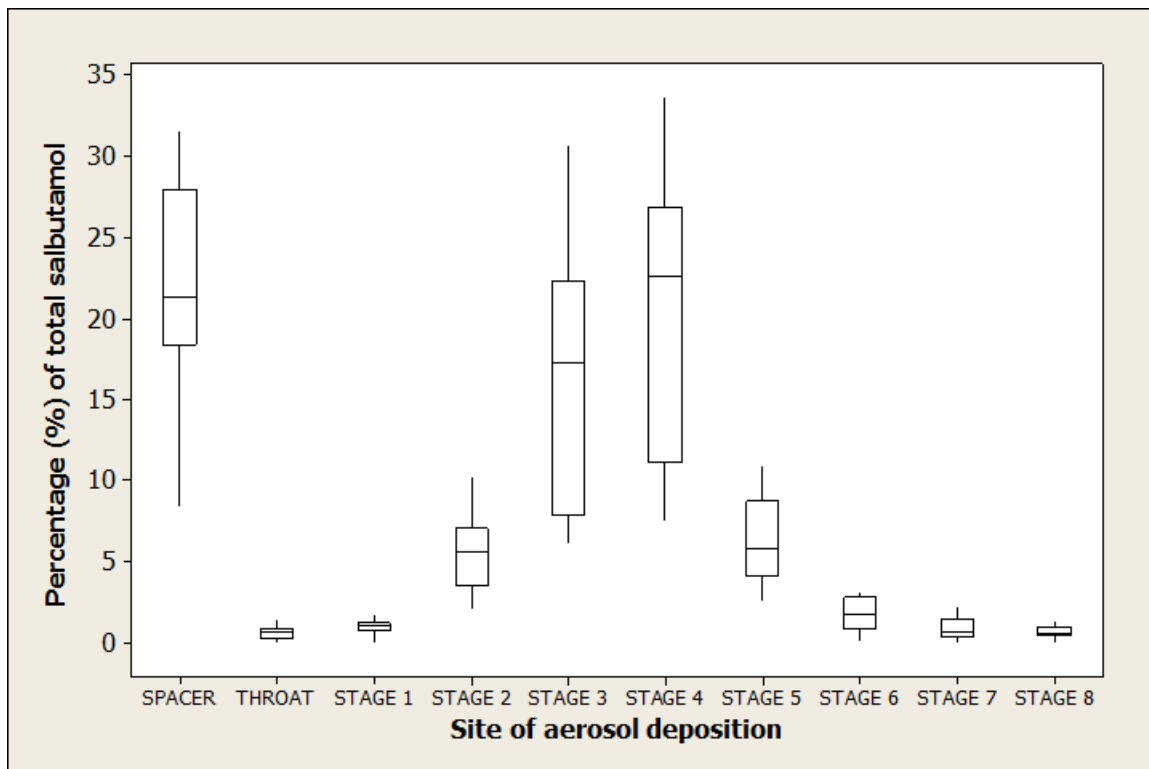
445 Figure 1 (a-c): Experimental setup. (a) The spacer (S) was fixed to the throat (black arrow) of  
446 the NGI in a horizontal position and the MDI was inserted into the spacer adjacent to the  
447 inspiratory valves (red arrow). (b) The NGI was comprised of a throat (not shown) and a series  
448 of eight stages through which air (containing the generated aerosol) flowed at a constant flow  
449 rate via pore sizes of sequentially decreasing diameter and consisting of eight particle collection  
450 plates (labelled 1-8). (c) For *Experiment 2*, the orientation of the MDI nozzle relative to the  
451 horizontal position was determined by its attachment to a combination square angle finder  
452 which was fixed to the bench.

453



455 Figure 2: Box and whiskers plot depicting the percentage of anticipated total aerosolised  
456 salbutamol (assuming 100µg per actuation) deposited within the spacer and the different stages  
457 of the NGI (throat and stages 1 – 8) under perceived optimal conditions. Data were derived  
458 from Experiments 2 and 3 when the MDI nozzle was orientated in a horizontal position with a  
459 5 s delay between actuations. Median (horizontal line), interquartile range (box limits) and  
460 range (whisker limits) derived from 2 x 8 repeats of either 10 (*Experiment 2*) or 8 (*Experiment*  
461 *3*) actuations.

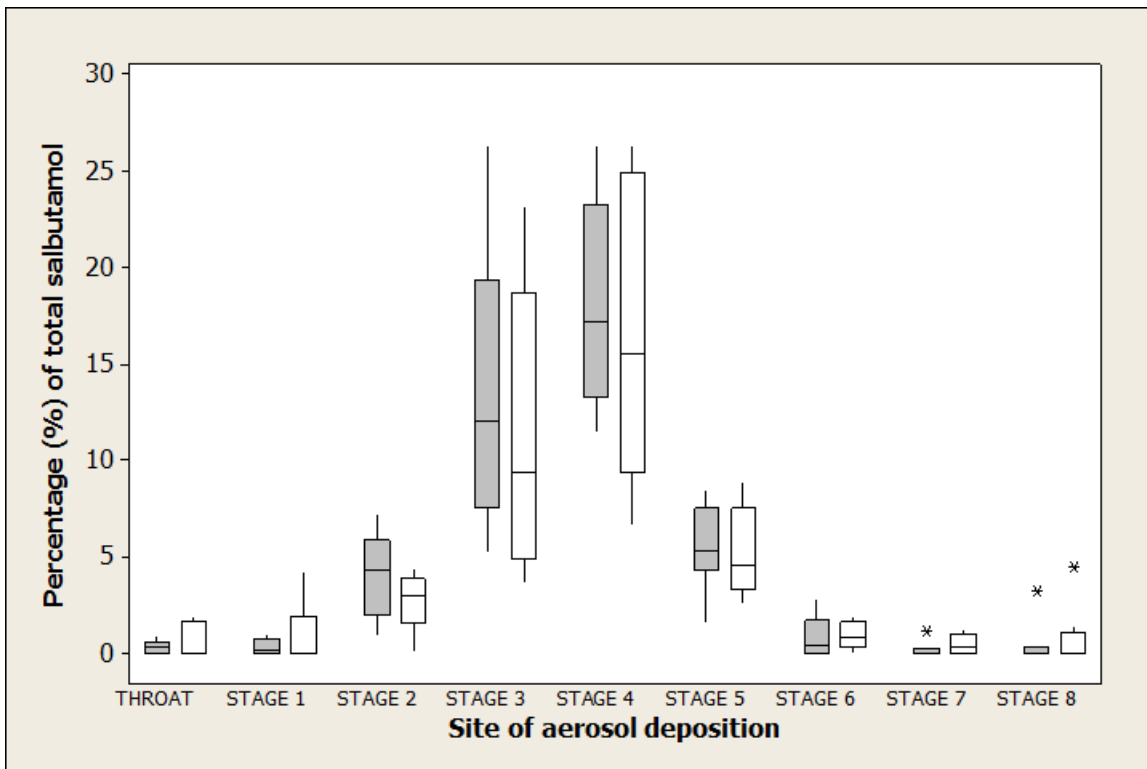
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464 Figure 3: Box and whiskers plot depicting the percentage of anticipated total aerosolised  
465 salbutamol (assuming 100µg per actuation) deposited within the different stages of the NGI  
466 (throat and stages 1 – 8) when the MDI was either shaken for 30s prior to the first actuation  
467 and then actuated at 5 s intervals without further shaking (solid boxes) or shaken for 30s prior  
468 to each actuation (open boxes). Median (horizontal line), interquartile range (box limits) and  
469 range (whisker limits) derived from 2 x 8 repeats of 10 actuations. Asterisk = outlier.

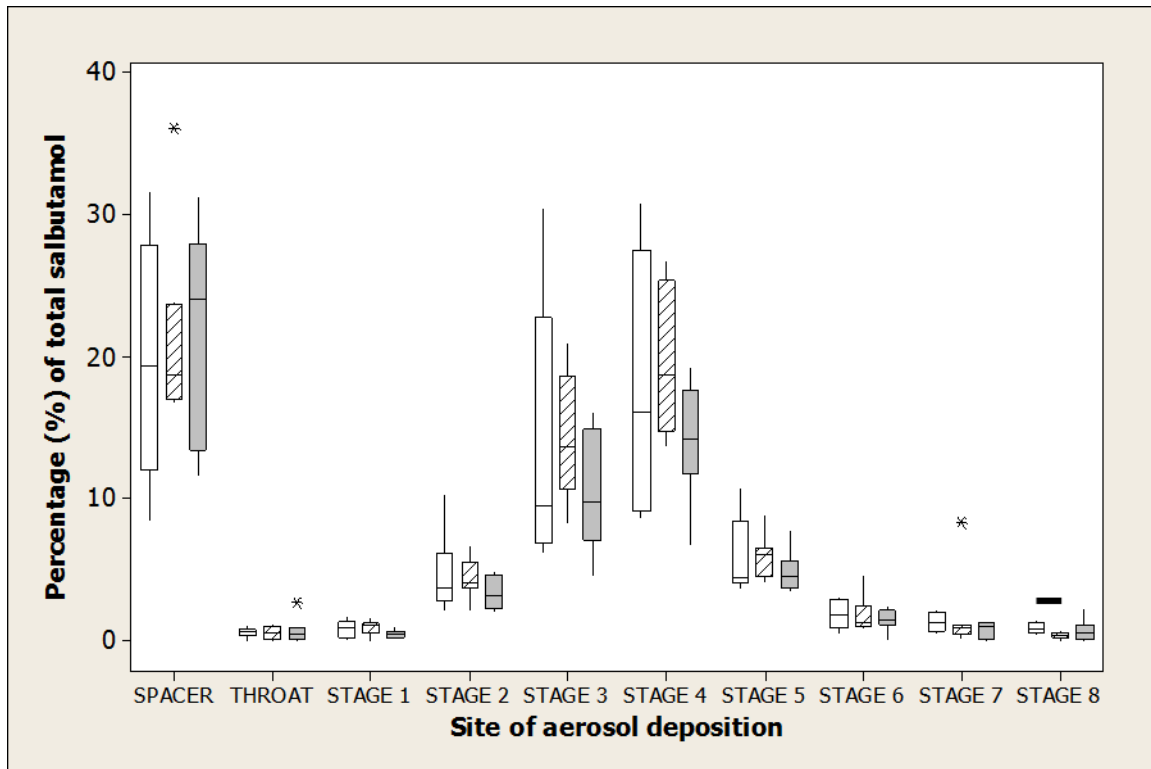
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472 Figure 4: Box and whiskers plot depicting the percentage of anticipated total aerosolised  
 473 salbutamol (assuming 100µg per actuation) deposited within the spacer and the different stages  
 474 of the NGI (throat and stages 1 – 8) following actuation of the MDI device when the output  
 475 nozzle was horizontal (open boxes) or deviated 10° (hatched boxes) or 20° (solid boxes) above  
 476 the horizontal. Median (horizontal line), interquartile range (box limits) and range (whisker  
 477 limits) derived from 3 x 8 repeats of 10 actuations. Asterisk = outlier; horizontal bar – limits  
 478 depict significantly different data sets (P<0.05).

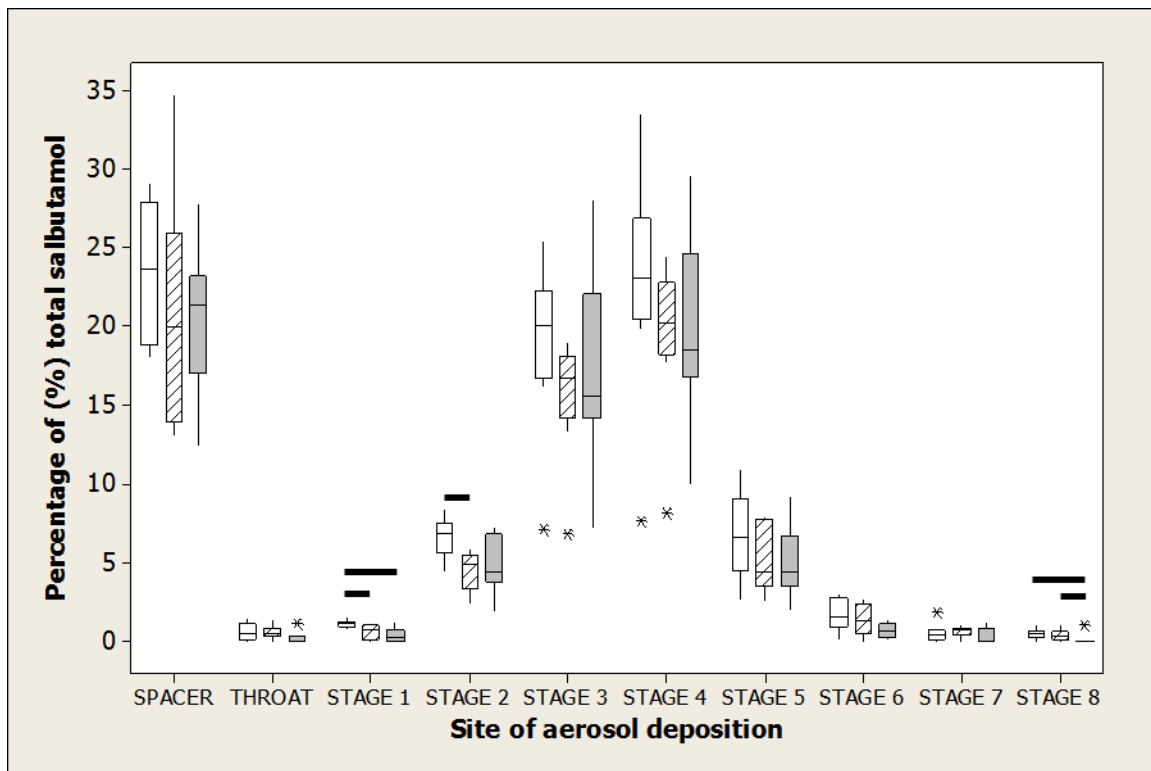
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481 Figure 5: Box and whiskers plot depicting the percentage of anticipated total aerosolised  
 482 salbutamol (assuming 100µg per actuation) deposited within the spacer and the different stages  
 483 of the NGI (throat and stages 1 – 8) following 8 actuations, either delivered individually at 5s  
 484 intervals (open boxes), as 4 x double actuations in rapid succession (hatched boxes) or as 2 x  
 485 quadruple actuations in rapid succession (solid boxes). Median (horizontal line), interquartile  
 486 range (box limits) and range (whisker limits) derived from 3 x 8 repeats of 8 actuations.  
 487 Asterisk = outlier; horizontal bar – limits depict significantly different data sets (P<0.05).

488



489



**Table 1:** Summary of experimental designs. Shaded cell indicates the comparisons made for each experiment. Bold text indicates the data used to measure aerosol characteristics in the Optimal Delivery Experiment.

	<b>MDI shake</b>	<b>MDI angulation</b>	<b>Rapidity of successive actuations</b>
<b>Experiment 1</b>	single shake prior to series of 10 actuations <i>versus</i> shake prior to each actuation	angle of actuation not standardised	10 x single actuations
<b>Experiment 2</b>	single shake prior to series of 10 actuations	<b>horizontal</b> <i>versus</i> 10° upward angulation <i>versus</i> 20° upward angulation	10 x single actuations
<b>Experiment 3</b>	single shake prior to series of 8 actuations	horizontal	<b>8 x single actuations</b> <i>versus</i> 4 x double rapid actuations <i>versus</i> 2 x quadruple rapid actuations
<b>Optimal Delivery Experiment</b>	single shake prior to series of 8 or 10 actuations	horizontal	8 or 10 x single actuations

