

Arsenic-phosphorus interactions in the soil-plant-microbe system: dynamics of uptake, suppression and toxicity to plants

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1	Arsenic-phosphorus interactions in the soil-plant-microbe
2	system: dynamics of uptake, suppression and toxicity to plants
3	
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16	ABSTRACT
17	High arsenic (As) concentrations in the soil, water and plant systems can pose a direct health
18	risk to humans and ecosystems. Phosphate (Pi) ions strongly influence As availability in soil,
19	its uptake and toxicity to plants. Better understanding of As(V)-Pi interactions in soils and
20	plants will facilitate a potential remediation strategy for As contaminated soils, reducing As
21	uptake by crop plants and toxicity to human populations via manipulation of soil Pi content.
22	However, the As(V)-Pi interactions in soil-plant systems are complex, leading to
23	contradictory findings among different studies. Therefore, this review investigates the role of
24	soil type, soil properties, minerals, Pi levels in soil and plant, Pi transporters, mycorrhizal

association and microbial activities on As-Pi interactions in soils and hydroponics, and uptake 25 by plants, elucidate the key mechanisms, identify key knowledge gaps and recommend new 26 research directions. Although Pi suppresses As uptake by plants in hydroponic systems, in 27 soils it could either increase or decrease As availability and toxicity to plants depending on 28 the soil types, properties and charge characteristics. In soil, As(V) availability is typically 29 increased by the addition of Pi. At the root surface, the Pi transport system has high affinity 30 for Pi over As(V). However, Pi concentration in plant influences the As transport from roots 31 to shoots. Mycorrhizal association may reduce As uptake via a physiological shift to the 32 mycorrhizal uptake pathway, which has a greater affinity for Pi over As(V) than the root 33 34 epidermal uptake pathway.

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Capsule: Understanding As-Pi interactions in the soil-plant systems can help in reducing the
As uptake by crop plants and protecting the food chain.

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Keywords: Arsenic toxicity. As-Pi interactions. As-Pi uptake by plants. Soil mineralogy. Soil
types. Mycorrhizal association

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63 **1. Introduction**

64 *1.1. Arsenic in the environment*

Arsenic is generally considered one of the top priority elements for carcinogenicity and 65 toxicity to humans, animals and plants (ATSDR, 2007). Several severe incidents of As 66 67 poisoning have occurred in regions such as Bangladesh, India (West Bengal) (Anawar et al., 2002), Vietnam, Taiwan and northwest China (Wang et al., 2002). Sources of high contents of 68 As in the environment are either geogenic in groundwater aquifers (Salmon et al., 2014) or 69 anthropogenic, typically associated with mining, industrial waste and agricultural chemicals 70 (Anawar et al., 2011). Average concentration of As in the Earth's crust is 1.5 µg/g with the 71 median value of 6.0 µg As /g for uncontaminated soils (Bowen, 1979). The average As 72 contents in agricultural fields that received As-containing pesticides and defoliants range from 73 5-2553 mg/kg (Walsh and Kenny, 1975). Early mining and smelting operations led to 74 contamination of the agricultural land, with soil As levels of 539-9380 mg/kg in Korea (Jung 75

et al., 2002), 11.1-651.1 mg/kg in Portugal (Pratas et al., 2005), etc. The As concentration in human hair exhibits significant relationship with that in rice (*Oryza sativa L.*), wheat (*Triticum aestivum* L.) and agricultural soils (Rahman et al., 2007, 2008) in As-affected areas reflecting human exposure to As via the soil–plant transfer pathways (Geng et al., 2006; Huang et al., 2006). These studies indicate widespread occurrence of As, and its contamination effects on agricultural soils.

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83 *1.2. As-Pi chemistry and their competition*

Under aerobic conditions, As and phosphorus (Pi) form stable tetrahedral oxyanions 84 (As(V) and orthophosphate, Pi) of +5 oxidation state due to similar chemical characteristics 85 (Manning and Goldberg, 1996; Lambkin and Alloway, 2003). In soil, As predominantly exists 86 in the inorganic forms (arsenate, As(V) and arsenite, As(III)) with minor concentration of 87 88 dimethylarsinic acid (DMAA) and monomethylarsenic acid (MMAA) (Pantsar-Kallio and Manninen, 1997; Quaghebeur et al., 2003; Quaghebeur and Rengel, 2005). Because of their 89 90 physico-chemical similarity, As(V) and Pi ions interact strongly competing for the same charged surfaces in soil. Arsenate uptake by plants occurs via the similar carrier process as Pi 91 (Pickering et al., 2000; Zhao et al., 2009) with Pi having more affinity for transport sites than 92 As(V) (Ullrich-Eberius et al., 1989; Meharg et al., 1994; Dunlop et al., 1997). For plant 93 growth, Pi is an essential macronutrient. The nucleic acids, phospholipids and ATP contain 94 Pi, that is involved in regulating key enzyme reactions and metabolic pathways (Schachtman 95 et al., 1998; Rose et al., 2013). 96

97

98 1.3. As-Pi interactions and significance of this review

A lot of research works have studied the As-Pi interactions in the environment with
conflicting results (Vetterlein et al., 2007; Zhao et al., 2009; Szegedi et al., 2010). On one

hand, Pi can decrease plant As uptake, and on the other hand, it may promote the availability 101 102 of As in soil and increase plant uptake. Therefore, it is necessary to explore the mechanism of action between As and Pi under different environmental conditions. Most studies, to-date, 103 104 have failed to account for the complexity of interactions between Pi and As in soil-plant system, and the variability in competition between As and Pi depending on soil types and 105 substrate conditions. Furthermore, the mechanisms of how Pi, when used as fertilizer, 106 ameliorate As toxicity to crop plants (Peryea, 1998) are not yet clear (Geng et al., 2005). 107 108 Although the major factors have been identified, the net effect of their interactions is poorly understood. Therefore, further research is essential to assess the available knowledge, identify 109 110 the research gaps for the future work, understand the As-Pi interactions in soil-plant systems, and their effects on exacerbation and/or amelioration of As toxicity, because these results can 111 help to decrease the risk of As toxicity to plants, animals and humans. Therefore, this study 112 113 reviewed (1) As-Pi interactions in soil-plant continuum, (2) the effect of soil type, soil properties and minerals on As-Pi interaction in soils, (3) the role of different Pi transporters in 114 115 Pi and As uptake by plants, (4) the mechanisms of As and Pi uptake in As-tolerant and Asintolerant plants, (5) how mycorrhizal symbioses and bacteria influence As and Pi uptake by 116 plants and As toxicity, and (6) case studies of As and Pi uptake by As-tolerant and As-117 intolerant genotypes, and P-deficient and P-sufficient plants. This review investigates the role 118 of Pi on As uptake and toxicity to plants, elucidate the key mechanisms and factors affecting 119 the As : Pi interactions in soils and plants, identify key knowledge gaps and recommend new 120 research directions. 121

122

123 2. Influence of soil property, mineralogy and soil types on As and Pi sorption to soils

124 Due to a smaller size and higher charge, Pi anions bind to soil more strongly and higher

than As(V) (Lambkin and Alloway, 2003) at similar Pi and As concentrations (Woolson et al.,

1973) making more As available by plant uptake (Zou et al., 2009). Due to slow sorption to 126 soil, Pi become more competitive than As(V) over time (Lambkin and Alloway, 2003). 127 According to the Steindorf-Rebhun-Sheintuch equation, ligand exchange theory and a share 128 charge hypothesis, Pi has more probability to replace As(V) from soils (Roy et al., 1986; 129 McBride, 1994). However, Pi could also be desorbed by As due to a mass action effect of 130 high As:Pi concentrations in soil solution (Lambkin and Alloway, 2003). The fast adsorption 131 of As and Pi occurred by exchange adsorption, while slow adsorption by specific adsorption 132 in the acid, neutral and calcareous purple soils (Zou et al., 2009). 133

The soil mineralogy, soil texture, soil properties and environmental factors that may 134 greatly affect the As-Pi interaction, mobility and availability of As in soils (Bissen and 135 Frimmel, 2003) and uptake by plants include mineral components, presence of anions (e.g., 136 citrate, phosphate, phytates/phytic acid (Dalal, 1977; Marschner, 1995), residence time, pH, 137 138 redox potential, organic matter (OM), microbial activity and especially, Fe- and Al-oxide/ hydroxides (Melamed et al., 1995; Turpeinen et al., 1999; Fitz and Wenzel, 2002; Kubicki, 139 140 2005). Among these factors, goethite and pH strongly control the behaviour of As and Pi. Knowledge of As(V) adsorption on individual soil minerals may predict As(V) adsorption in 141 whole soils and its response to Pi addition (Manning and Goldberg, 1996). Fordham and 142 Norrish (1979) and Violante and Pigna (2002) found that Fe and Mn-rich minerals such as 143 goethite, nontronite, ferruginous smectites, birnessite and pyrolusite absorbed more As(V) 144 than Pi when supplied in equal molar ratio, but more Pi than As(V) was sorbed on 145 noncrystalline Al containing minerals e.g., gibbsite, boehmite, allophane and clay fractions 146 (e.g., kaolinite, illite and vermiculite) resulting in higher As(V) desorption and uptake by 147 Indian mustard (Brassica juncea L.) (Bolan et al., 2013). A high rate of goethite addition to 148 soils can reduce Pi and As uptake by plants (Vetterlein et al., 2007). The Pi and Fe addition to 149

high As soils using a sequential incorporation method can significantly decrease As toxicityto lettuce root growth by supplying a nutrient source (Koo et al., 2013).

In case of sandy soils/sandy loam, Pi addition displaced As(V) ions from the surface of 152 sand grains due to a low number of sorption sites and increased As uptake by plants, thus 153 decreasing plant growth (O'Neill, 1992; Woolson et al., 1973), but increased vegetative yields 154 of wheat, barley, sudangrass and lucerne (Creger and Peryea, 1994) decreasing As uptake and 155 toxicity on a silty clay loam (Woolson et al., 1973), a silty loam (Jacobs and Keeney, 1970) 156 157 and clay loam and sandy clay loam soils (Pigna et al., 2010). Arsenic mobility and solubility in anion-fixing soils or alkaline soils can be strongly affected in case of high P supply (Smith 158 159 et al., 1998; Violante and Pigna, 2002). As(V) and Pi adsorption-desorption had the similar characteristics and followed the Langmuir and Freundlich equations in the low pH (acid), 160 neutral and high pH soils (calcareous purple soils) (Zou et al., 2009), but they are not always 161 162 dependent on each other due to dissolution and mobility of un-reactive P at low pH (Mwamila and Gustafsson, 2011). 163

164

165 **3. As-Pi interaction, uptake and arsenic toxicity in plants**

166 *3.1. Mechanisms of As toxicity to plants*

167 The predominant effect of As toxicity in the cell is oxidative stress (Finnegan and Chen, 2012) that reduces seed germination, root and shoot growth and root surface area, etc. (Smith 168 et al., 2010), restrict nutrients and water uptake, and cause nutrient deficiency in plants 169 (Paivoke and Simola, 2001). After reduction arsenic (As(III) forms chemical bond with 170 sulphydryl groups and deactivates some enzymes (Delnomdedieu et al., 1993), and increases 171 reactive oxygen species and lipid peroxidation, but decreases the superoxide dismutase 172 activity and antioxidants levels (Raab et al., 2004) resulting in cellular membrane damage 173 (Mascher et al., 2002; Tuan et al., 2008). In contrast, Pi supply to As-treated plants decreased 174

activities of CAT, APX and lipid peroxidation, and thus decreased membrane damage (Gunes 175 et al., 2009). Arsenate in the cytoplasm competes with Pi forming unstable ADP-As(V), and 176 disrupt the energy flows in cells (Hartley-Whitaker et al., 2001, 2002). High S concentrations 177 in soil (e.g., via micronutrient fertilizers) alleviated the toxicity effect of As on plant growth 178 although Pi addition to soil enhanced As uptake by plants (Grifoni et al., 2015). This 179 phenomenon occurred due to formation of glutathione and phytochelatins, their chelation with 180 181 As and subsequent sequestration of these chelated complexes in the vacuoles (Cobbett, 2000; Hartley-Whitaker et al., 2001). Although Pi supply suppressed the As uptake by plants in the 182 hydroponics, high S addition increased As uptake and decreased Pi content influencing the 183 184 As-Pi interaction in plants. Application of As and Zn to soils shows an antagonistic relationship such as Zn in soils (at application rate of 3 mg/L) reduced As availability in soils 185 and uptake in plants (Sanchary and Huq, 2017). Arsenic accumulation by plants is controlled 186 187 by the oxidation state, As and P forms and plant species.

Arsenic hyperaccumulator plant species (e,g., Pteris vittata) detoxify As toxicity in a 188 number of ways such as transformation of As(V) to As(III) by enzyme and 189 compartmentalization in subcell; capturing reactive oxygen species by phytochelatin and 190 thiols, preventing lipid peroxidation (Shoji et al., 2008; Bona et al., 2011) and maintaining a 191 greater ratio of P/As in the roots, at least 1.2 in soil solution or 1.0 in fronds (Tu and Ma, 192 193 2003a; Singh and Ma, 2006). Therefore, Pi application may be an important strategy for As detoxification, enhanced growth of Pteris vittata and efficient As uptake to phytoremediate 194 As-contaminated soils (Kertulis et al., 2005; Ye et al., 2011; Yan et al., 2012). The in-situ 195 196 measurement of As/Pi molar concentration ratio can predict As phytotoxicity, where As/Pi ratio of 0.168–0.360 resulted in a 50% reduction of wheat (Triticum aestivum) growth 197 198 (Mojsilovic et al., 2011).

199

200 3.2. As-Pi interactions and uptake by plants grown in soils

The competition between As and Pi, and the effect on their uptake by plants grown in soils 201 are described briefly in Table 1. Arsenate uptake from soil occurs via the same transport 202 203 system as Pi (Pickering et al., 2000), however, the effects of As on plant metabolism are complex and only partially understood. An application of Pi had variable effects on the 204 sorption and mobility of As in soils, uptake by plants and toxicity effects with both increases 205 (Lambkin and Alloway, 2003; for 10 mg P/ kg in Geng et al., 2005; 238.4-423.2 mg As/kg 206 207 soil and 10.0 mM phosphate addition in Tao et al., 2006) and decreases (Hanada et al., 1975; for 40 mg P/ kg in Geng et al., 2005; Pigna et al., 2012) reported. Nevertheless, increasing Pi 208 209 concentration could not resist a fraction of 'steady state' As(V) uptake, while As(V), in contrast, can inhibit 'steady state' Pi uptake mildly (Geng et al., 2005). Therefore, the 210 211 interaction of As and Pi, and suppressive effect of one on another is not straight forward. It is 212 not yet studied how As interacts with organic P in soil. Irrigating As-rich water may change the As-Pi balance in soil solution, causing mobilisation of Pi and subsequent losses by 213 214 leaching or higher Pi availability for plant nutrition. But higher As toxicity can affect crop yields as well (Talukder et al., 2011, 2012). The mobility of As and Pi, and As toxicity 215 decreases due to formation of recalcitrant forms in aged soil (Onken and Adriano, 1997; 216 Lombi et al., 1999). 217

The soil Pi status was related to As uptake by *Urtica dioica*, but not by *Phragmites australis* indicating a differential response of plants to Pi (Ahmed et al., 2011). The uptake kinetics of As and Pi by different plant species over short or long time vary depending on the genus and species of plants. Therefore, further research is necessary to understand a timecourse of Pi and As(V) uptake by different plant species. Compared to non-tolerant *Holcus lanatus*, the tolerant clones generally contained a higher proportion of As(V) (Quaghebeur and Rengel, 2003). Arsenic uptake by and growth of rice cutgrass (*Leersia oryzoides* Sw.) and tall fescue (*Festuca arundinacea* Schreb.) was not enhanced by P fertilization above a minimal amount (Klaber and Barker, 2014). The application of Fe^{2+} to the rice fields under flooded and reduced condition decreased the As uptake in rice grain and increased their yield, while Pi application demonstrated the opposite results (Hossain et al., 2009).

229

230 3.3. Impact of Pi availability on As uptake by plants in hydroponics

Phosphate and As(V) interactions in hydroponics and plant accumulation are largely 231 investigated (Khattak et al., 1991; Clark et al., 2000; Pickering et al., 2000; Esteban et al., 232 2003; Panuccio et al., 2012; Shaibur et al., 2013) and briefly summarized in Table 2 and Fig. 233 234 1. Most studies found that Pi deficiency in nutrient solutions increased the capacity of rice and Lemma gibba plants to take up more Pi and As(V), but at low As concentration, high Pi 235 treatment decreased As(V) uptake and toxicity (Macnair and Cumbes, 1987; Mkandawire et 236 237 al., 2004; Lihong and Guilan, 2009; Panuccio et al., 2012) via suppression of the high-affinity Pi/As(V) transport system. The supply of Pi also influenced the As speciation and transport 238 239 (Lei et al., 2012). The Isatis cappadocica, a robust perennial rosette plant, can grow in highly impacted As-contaminated areas and hyperaccumulates As in its areal parts in hydroponic 240 media (Karimi and Souri, 2015). 241

242 Phosphate can suppress As(V) uptake in phytoplankton (Planas and Healey, 1978), Oryza sativa (Abedin et al., 2002), Lupinus albus (Esteban et al., 2003), barley (Shaibur et al., 2013) 243 and As-tolerant species such as Holcus lanatus, Cytisus striatus (Meharg and MacNair, 1992; 244 Bleeker et al., 2003) and Pteris vittata (Wang et al., 2002; Tu and Ma, 2003b) grown in 245 hydroponic systems. The time-dependent split-Pi application (P₁₃₄₊₆₆ and P₆₆₊₁₃₄) with low 246 initial Pi application increased the growth of Pteris vittata L. and the efficiency of As 247 removal, that was higher than high initial Pi-supply (P₂₀₀₊₀) (Santos et al., 2008). The radial 248 loss of O₂ from aerenchyma structures converts the rice root surface into much more highly 249

oxidized condition compared to the surrounding environment (Colmer, 2003) and forms Feoxide plaques on the root surface (Taylor et al., 1984).

The As-Pi interactions and their uptake by plants produced different and often 252 contradictory results depending on soil substrates and hydroponic conditions. The kinetics of 253 Pi and As accumulation in plants in hydroponics is overestimated compared to soil 254 experiments, because some processes such as water flow, redox potential, diffusion, 255 adsorption/desorption and ion exchange by minerals (Fe, Mn and Al oxyhydroxides, clay 256 257 minerals, sulfide minerals, etc.) and organic components are common in soil systems that control As and Pi solubility and mobility in soils (Bissen and Frimmel, 2003; Anawar et al., 258 259 2008), while these processes do not occur in hydroponics (Fitz and Wenzel, 2002). Hence, the results from the hydroponic experiments can not be extrapolated to and may have limited 260 validity for soil-grown plants (Fitz and Wenzel, 2002). However, the hydroponics studies can 261 262 provide new knowledge on the membrane transport.

263

264 *3.4. Relation of plant P status with As uptake*

Quaghebeur and Rengel (2004) showed that P level in plant and As-Pi interactions in soil 265 are pivotal factors controlling As and P uptake by plants. The Pi deficiency can enhance As 266 uptake by plants e.g., Pteris vittata (Lei et al., 2012), P-deficient canola (Quaghebeur and 267 Rengel, 2004) and white lupin (Lupinus albus) (Esteban et al., 2003), causing a decline in 268 plant growth (Wang et al., 2002; Geng et al., 2006) except Pteris vittata. The low As uptake 269 and amelioration of As(V) toxicity in most of the +P plants occurs by competition with Pi 270 except lupin (Esteban et al., 2003; Reina et al., 2005). The P-deficient white lupin plants 271 develop proteoid roots and take up more Pi and As(V) (Esteban et al., 2003; Reina et al., 272 273 2005), but lower translocation of As to shoot compared to corn plants (Vetterlein et al., 2009). The presence of As(V) stimulated P uptake by P-deficient plants and reduced it for the P 274

sufficient ones. However, the concentration of As in rice and barley shoots grown in hydroponics was very lower in P-deficient plants than in P-sufficient ones due to reddish iron plaque formed on the root surface sequestering As and decreasing its uptake by roots (Chen et al., 1980; Liu et al., 2004; Shaibur et al., 2013). Furthermore, it is assumed that high P level in As-tolerant plants could alleviate As toxicity more efficiently than non-tolerant ones, and despite the decreased As(V) uptake, As-tolerant plants might have higher total concentration of As in their biomass over long time (Campos et al., 2014).

Higher molar ratio of Pi/As in rice (*Oryza sativa*) shoots of Indica cultivar than for the hybrid Indica variety (Lu et al., 2010) suggested the genotypic difference in response to As(V) toxicity and potential capacity to breed rice cultivars for As affected paddy soils (Geng et al., 2006).

286

287 3.5. High-affinity and low-affinity Pi transporter for Pi and As uptake

The different families of transporter proteins are involved in Pi transport through plant 288 289 plasma membranes by a highly complex network of regulation, e.g., PHT1 transporters for Pi accumulation from soil (Nussaume et al., 2011). Plants have developed different controlled 290 adaptative mechanisms to acquire necessary amount of external Pi and maintain Pi 291 homeostasis as well as overcome low Pi availability (reviewed in Schachtman et al., 1998; 292 293 Rouached et al., 2010). In the low Pi environment, Pi and As(V) accumulation by plants are controlled by the Pht1;9 and Pht1;8 membrane transporters through activation of high-affinity 294 Pi transport system (Rausch and Bucher, 2002; Remy et al., 2012). Arsenate inhibited the root 295 296 growth of non-tolerant Holcus lanatus plants more strongly than the tolerant genotypes. The tolerant plants (e.g., *Holcus lanatus*, *Silene vulgaris* and Agrostis species) took up less As(V) 297 298 than non-tolerant ones over short time (Porter and Peterson, 1977; Zhao et al., 2009) supporting the hypothesis that a suppressed high-affinity Pi transport system provides As 299

resistance together with increased Pi/As ratio and strong selectivity against As(V) (Macnair
and Cumbes, 1987). However, total As accumulation is higher in tolerant than non-tolerant
plants over long time, as the latter become unhealthy and die relatively quickly (Puckett et al.,
2012).

As(V) and Pi compete for transporters across root plasma membrane (Smith et al., 2010). Recently, Panuccio et al. (2012) indicated that As(V) resistance of *Pennisetum clandestinum* Hochst (kikuyu perennial grass) occurs due to enhanced Pi nutrition, and not due to downregulation of the Pi uptake system. Aquaglyceroporins (AQP3, AQP7, AQP9, and AQP10) mediate the bidirectional movement of arsenite across cell membranes (Jung et al., 2012). The As(III) transport was not affected by Pi in rice.

310

311 *3.6. Effect of root exudates on As and Pi uptake*

312 The plant root exudates play critical roles in As and Pi release from soil matrix. The main factors controlling the mobility of As in surrounding environment of roots and uptake by 313 314 plants include solubilisation of As from the soil matrix/minerals, very large surface areas of roots (Pollard et al., 2002), rhizosphere pH, and chelating agent (Quaghebeur et al., 2005; 315 Marschner, 1995). The root exudates including organic acids and phenolics released by the P-316 deficient plants can mobilize Pi and As from soil matrix including Fe-oxides/hydroxides, and 317 enhance availability to plants by changing soil pH and through forming soluble metal-chelate 318 complexes (Hoffland, 1992; Strom et al., 1994; Kirk et al., 1999). 319

320

321 *3.7. Does As resistance occur via decreased As uptake or protective effect of high Pi uptake?*

Arsenic tolerance in higher plants hinges on decreased As accumulation by suppression of the high-affinity Pi/As(V) uptake system in roots (Meharg and Macnair, 1992) and decreased As transport to shoots (Pigna et al., 2009), both of which are dependent on high shoot Pi

status that outcompetes As in metabolic reactions as reported for the arsl Arabidopsis 325 326 mutants (Lee et al., 2003) and vacuolar sequestration of As in shoots. However, our data analysis from Pigna et al. (2009, 2010) and Lewinska and Karczewska (2013) demonstrate the 327 328 new results as follows: Pi application to soils not only augmented plant growth, but also increased As and Pi concentrations in shoot of wheat (Triticum durum L.) irrigated with As-329 330 rich water (Fig. 2). Therefore, it is clear that Pi addition to soils alleviated As toxicity not by 331 decreasing As uptake by plants, but increasing the plant biomass, Pi nutrition and metabolic reactions. Joardar and Kawai (2014) and Christophersen et al. (2009a) also reported that the 332 decreased As toxicity occurred not due to lower As uptake by plants, but increased Pi uptake 333 334 by plant root surface and transport to shoot (Campos et al., 2014). A more detailed investigation of uptake of As(V) and Pi is necessary to clarify this hypothesis. Our recent 335 336 study indicated that P addition to topsoil (upper 5 cm) of three-layered soil system with As in 337 the subsoil (> 10 cm) was a more effective strategy to impove the plant growth by alleviating As toxicity, increasing P concentration, but reducing As level in plant biomass than P 338 339 application in subsoil (Anawar et al., 2016).

340

4. Role of mycorrhiza on As-P interaction in soil system and uptake by plant

342 The symbiotic association of arbuscular mycorrhizal (AM) fungi with plant roots growing in As-contaminated soil has been demonstrated to improve P nutrition, reduce As(V) uptake, 343 increase P/As ratios in the shoots, and alleviate As toxicity for several plant species (Fig. 3) 344 (Meharg et al., 1994; Chen et al., 2007; Ultra et al., 2007; Xia et al., 2007; Xu et al., 2008; 345 346 Christophersen et al., 2009b, 2012; Ahmed et al., 2006, 2011). Albeit some studies did not demonstrate good responses to AM fungi for tolerance to soil As (Knudson et al., 2003; Zhu 347 et al., 2003; Li et al., 2006; Grace et al., 2009), indicating that the benefits of the AM 348 association may not be universal for all scenarios (e.g., basin wildrye). 349

Based on demonstrated studies it is reported that AM plant roots may have physiologically 350 351 altered Pi uptake systems, whereby there is a switch from the epidermal uptake pathway to an AM uptake pathway following colonisation. Considering the physiological and molecular 352 353 evidence, Smith et al. (2003) showed that AM plants have two pathways for Pi and As(V) uptake: direct pathway via high-affinity Pi transporters Pht1;1 and Pht1;2 in the epidermis and 354 root hairs in AM and NM (non-mycorrhizal) plants (Schunmann et al., 2004a,b; Shin et al., 355 2004) regardless of their responsiveness (Smith et al., 2010). The down-regulation of 356 357 expression of high-affinity Pi-uptake pathway lowered Pi and As(V) uptake in AM barley and genotypes of As-tolerant Holcus lanatus, but other studies indicated no such effect (Grace et 358 359 al., 2009). However, it suggested that the AM and direct Pi uptake pathways are integrated, and Pi and As(V) uptake may be inter-dependent (Smith et al., 2003, 2004; Smith and Read, 360 361 2008), although details of the mechanisms are still unclear.

362 The transporters in external hyphae of the AM fungi in symbiosis condition uptake Pi via the mycorrhizal pathway (Maldonado-Mendoza et al., 2002). The hyphae transports Pi along 363 a long way (Ezawa et al., 2002) and transfer to plant across a symbiotic interface to cortical 364 cells by induction of Pht1;8 in roots (Paszkowski et al., 2002; Glassop et al., 2005). Thus, a 365 higher amount of Pi and exclusion of As are promoted by the AM pathway that protect plants 366 against As uptake and toxicity (Christophersen et al., 2009b). It suggested that AMF 367 decreased As toxicity in soil by converting As speciation from inorganic to organic forms by 368 hyphae-produced glomalin (Ultra Jr et al., 2007; Chern et al., 2007; Smith and Read, 2008) 369 and acting as a barrier for As uptake (Ultra Jr et al., 2007; Ahmed et al., 2011) through several 370 371 physical mechanisms: adsorption onto plant or fungal cell walls in plant tissues or extraradical mycelium in soil (Hildebrandt et al., 1999; Kaldorf et al., 1999; Joner et al., 2000); 372 373 exclusion by precipitation onto polyphosphate granules (Turnau et al., 1993). Some studies reported that lower As uptake in shoot occurred mainly due to a "dilution effect" from 374

increased growth of AM plants and decreased As partitioning to shoots, rather than decreased 375 As uptake per plant (Chen et al., 2007; Zhao et al., 2009; Ahmed et al., 2011). Therefore, 376 further research is needed to determine the effects of AM fungi-induced various metabolic 377 378 strategies for As resistance mechanism and As uptake by plants. A recent study suggests that mycorrhizal colonization converts As into non-toxic compexes by enhancing the production 379 of cysteine, glutathione, non-protein thiols, and activity of glutathione-S-transferase in plants 380 indicating the multifarious role of AMF in alleviation of As toxicity (Sharma et al., 2017). 381 Furthermore, Pi from superphosphate fertilizer or other sources may suppress the AMF 382 colonization, change the structure of the AMF community (Shetty et al., 1995; Ahmed et al., 383 2011) and thus strongly influence any potential AM effect on As resistance. Combined 384 application of inoculum and Pi to soil augmented plant biomass, reduced As toxicity and 385 increased plant Pi nutrition; and these effects were higher than inoculation only with native 386 387 AMF (Cozzolino et al., 2010; Cattani et al., 2015).

Glomus mosseae that was more tolerant and unaffected by the high concentrations of As 388 (e.g., 200 mg/kg) than Medicago truncatula plant, increased host plant As tolerance and Pi 389 nutrition (Xu et al., 2008). Not only AMF, but also ericoid and ectomycorrhizal fungi can 390 increase their tolerance to As(V) contamination via different mechanisms (Sharples et al., 391 1999, 2000a). The ericoid mycorrhizal fungus Hymenoscyphus ericae from an As and Cu 392 393 mine spoil has adapted to As(V) contamination by decreasing As uptake via enhanced As(III) efflux (Sharples et al., 2000b), but the mechanisms behind this process are still unclear. Three 394 species of ectomycorrhizal fungi showed variable response to arsenate toxicity under the 395 influence of PO4³⁻ concentration (Chen and Tibbett, 2007). The Suillus variegatus and 396 Hebeloma crustuliniforme exhibited the high sensitivity to As toxicity at low Pi treatments, 397 398 while the higher Pi treatments reduced As toxicity. In contrast, the Cenococcum

geophilum exhibited higher tolerance to As; and Pi did not show ameliorating effect onAs toxicity in *C. geophilum*.

401

402 5. Influence of soil microbial communities on As and Pi uptake by plants

Microbial activity causes transformation of As sepcies by reduction (arsenate to arsenite), 403 404 oxidation (arsenite to arsenate), and methylation (Wang et al., 2004; Qin et al., 2006; Cai et al., 2009). Arsenic reduces the soil functional gene diversity, while Pi increases the soil 405 microbial community structure. The rhizobacteria accelerate As hyperaccumulation by P. 406 vittata (Xiong et al., 2010), even under a Pi-limiting environment (e.g., insoluble phosphate 407 408 rock in alkaline soil) in As-rich media (Lessl and Ma, 2013; Lessl et al., 2014). By excreting acids, protons, and siderophores, the rhizobacteria can cause dissolution of phosphate rock 409 and minerals that improve plant Pi nutrition and As uptake by plants (Duponnois et al., 2005; 410 411 Ghosh et al., 2015). Due to the continuous excretion of plant released organic substrates, the number of microorganisms in the rhizosphere increases compared to the non-rhizosphere soil 412 413 resulting in a more diverse bacterial community (Marschner, 1995). Microbial reduction of As(V) to As(III) occurs by dissimilatory reduction and detoxification activities of microbes 414 (Fitz and Wenzel, 2002) using As(V) as a terminal electron acceptor, and also As(V) 415 reductase and As(III) extrusion by an As(III)-efflux pump (Cervantes et al., 1994). 416

417

418 **6.** Conclusions

The soil type, structure, properties, mineral components and pH may greatly affect the As-Pi interactions, mobility and availability of As in soils and uptake by plants with increased As availability and toxicity on sandy soil, but lower on a silty and sandy clay loam soils. Although all hydroponic studies indicated that Pi additions decreased As uptake by plant resulting in mitigating As toxicity, Pi application to soil either increased or decreased As

uptake and toxicity effects by tolerant and non-tolerant plants depending on substrate 424 425 conditions. What is the main reason behind these processes is not yet explained in any study. Therefore, this review hypothesized these processes as follows. The concentration of both Pi 426 427 and As(V) in soil solution is typically orders of magnitude lower than the concentrations employed in the majority of hydroponic studies. For example, Wenzel et al. (2002) reported 428 As(V) concentrations in the soil solutions from a range of uncontaminated and moderately 429 contaminated soils to be \leq 53 nM and up to 2.3 μ M in a highly contaminated soil. Similarly, 430 Pi concentrations in soil solution are typically $< 10 \mu M$ (Bieleski, 1973), which again is orders 431 of magnitude lower than the range of Pi concentrations employed in hydroponic experiments 432 433 (e.g., Tu and Ma, 2003b, 202-1000 µM; Clark et al., 2000, 10-250 µM; Meharg et al., 1994, 25-1000 µM). Unrealistically high concentrations of Pi may elicit (low affinity) uptake 434 pathways and metabolic processes which are not reflected in soil culture. Therefore, further 435 436 hydroponic studies are recommended using the As and Pi concentrations that are commonly available in soil porewater. 437

438 In soils, Pi additions generally result in increased Pi and As uptake, and also increased As resistance indicating that increased levels of Pi in plant biomass reduced the toxicity effects of 439 As. The effects of As(V) on Pi uptake are relatively weak due to high affinity of the 440 transporters for Pi than As(V) in both hydroponics and soils. At low As(V) concentration, a 441 high-Pi treatment decreased uptake and toxicity of As(V) in non-tolerants but not in As-442 tolerant plants. The As-tolerant plants might have distinct Pi uptake system with high 443 selectivity for Pi and against As(V) resulting in higher Pi/As ratio and As resistance. New 444 data analysis indicates that Pi addition promoted plant growth and As resistance not by 445 reducing As uptake by plants, but increasing Pi concentrations in shoot and protective activity 446 by internal P-induced metabolic reactions. Sulphur in soil can enhance As uptake by plants 447 due to its ability to reduce the toxicity effects. The mycorrhizal association contributes to the 448

increased Pi nutrition, plant growth and in ameliorating As toxicity. Microorganisms in the rhizosphere can cause transformation of As sepcies by redox reactions and methylation. The rhizobacteria release different types of acids to cause the dissolution of phosphate rock and minerals resulting in higher Pi nutrition and As uptake by plants.

453

454 **7. Recommendations for future work**

455 The previous studies related to As-Pi interactions in different substrates and the role of soil types on Pi and As(V) mobility and uptake by plants produced the contradictory results. 456 Although the results from all hydroponic studies demonstrated the similar results, the studies 457 458 conducted on soils showed a lot of discrepancies. Therefore, further research works are highly recommended to study the As-Pi interactions and uptake by plants grown in different soil 459 types. After Pi addition to soil, higher Pi uptake and translocation from root to shoot can 460 461 contribute to more resistance to As toxicity to plants than decreased As uptake. However, more detailed study is required to have a clear idea about how uptake of Pi ameliorates the As 462 toxicity to plants. Several mechanisms for amelioration of As toxicity by AMF have been 463 proposed and discussed in this review, and further research is needed to characterize these 464 mechanisms, particularly a lack of As transport and increased Pi uptake via the AM pathway 465 in plants. Since high Pi supply decreases mycorrhizal colonisation, proper management of Pi 466 fertilization can increase the efficiency of mycorrhizae for enhanced Pi and a decreased As 467 uptake. Therefore, this hypothesis should be further investigated in areas where --high As 468 concentrations may exist in agricultural soils, irrigation water and/or groundwater. Using the 469 470 isotopes can help to reveal these mechanisms as well.

471

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967	
968	Figure captions
969	
970	Fig. 1. As-Pi interactions and uptake by plants in hydroponics. As and Pi supply variably
971	control As and Pi uptake by plant roots. Pi supply strongly decreases As uptake, while As
972	supply weakly decreases Pi uptake. Pi has higher affinity for plan roots than As.
973	
974	Fig. 2. Mean shoot biomass, As and P concentrations in shoots and total As uptake by wheat
975	plants grown in soil irrigated with As containing water at three concentrations amended and
976	non-amended with P (data analysis from Pigna et al., 2009).
977	
978	Fig. 3. Role of AM in modifying As-Pi interactions, and As and Pi uptake by plant root. The
979	arbuscular mycorrhizal (AM) symbioses increase Pi uptake by plant roots and Pi/As ratio in
980	shoot, while they decrease As uptake by roots.

Table 1

Soil types	Plant species	Addition	Effect	Reference
Soil	Zea mays	Fe (1-4 g/kg)	At low goethite - low As/high P uptake. At high goethite - no As/Pi	Vetterlein et al. (2007)
			uptake	
Silt loam/	Zea mays	Pi (50-300	Pi had little effect on As toxicity to plants in silt loam, but enhanced	Jacobs and Keeney
sand		mg/kg)	As uptake and toxicity in sand at 80 mgAs/kg	(1970)
Soil	Pteris vittata	As (2.7-	Low As increased Pi uptake, but high As decreased Pi uptake. Pi	Tu and Ma (2003a),
		5.3mM)	increased plant biomass and As uptake at high As supply	Kertulis et al. (2005)
Soil	Oryza sativa	Pi	Increasing shoot Pi for breeding rice with low grain As	Lu et al. (2010)
Soil	Oryza sativa	Pi (0-0.5	Suppressed As(V), but not As(III) uptake; little difference in As	Abedin et al. (2002),
		mM)	uptake between two P-deprived rice genotypes	Geng et al. (2006)
Clay loam	T. durum	Pi (75 kg/ha)	Prevented As uptake and translocation in plants	Pigna et al. (2010)
Soil	H. vulgare	Pi/ As	Pi strongly inhibited As(V) uptake. As weakly inhibited Pi uptake	Asher and Reay(1979)
Soil-sand,	H. vulgare	Pi (20-120	Increased As uptake; decreased As(V) toxicity not by lowering As	Christophersen et al.
soil	T. vulgare	mg/kg)	uptake, but by emhancing Pi nutrition. No competition between As	(2009a), Tao et al.
			and Pi.	(2006)
Soil	C. arietinum	Pi (50-400)	Pi increased As uptake, but partially protected membranes from	Gunes et al. (2009)
			damage	
Soil	L. cinereus	As/Pi	High level of As and low Pi supply decreased plant growth	Knudson et al. (2003)
Loam soil	P. armeniaca	Pi	Increased shoot and root As in soil contaminated with Pb-arsenate	Creger & Peryea
			pesticide residue.	(1994)
Soil	Arabidopsis	Pi (10-40	Low Pi supply increased As uptake and plant growth. High Pi	Geng et al. (2005)
	thaliana	mg/kg)	supplydecreased As uptake due to competition on surface of soil	
			particles and plant roots.	
Soil	Solanum	Pi (6 mM),	Pi was more strongly adsorbed to soil than As(V), Pi desorbed As	Pigna et al. (2012),
	lycopersicum	As (4 mg/L)	and increased As uptake by plants depending on soil charge	Bolan et al. (2013)
			properties	
Soil	H. lanatus	Pi (0.2 g/kg)	Increased As uptake by plant because of increased As desorption by	Lewińska and
			competition	Karczewska (2013)
Soil	Plants	Pi	Enhanced As(V) and As(III) desorption from soil and thus leaching	Fitz and Wenzel, 2002
			or uptake by plants	

Effects of competition between As and Pi on uptake by plants grown in soil.

As-Soil	Vegetable	Pi (3867)	Increased As uptake- 4.6–9.3 times forcarrot, 2.5–10for lettuce	Cao and Ma (2004)
Soil	No plants	Pi/As	At As:Pi ratio equal to 1, more Pi was sorbed than As. At As:Pi	Woolson et al. (1973)
			ratios > 1 , Pi was desorbed due to a mass action effect	
Fe-soil	No plants	Pi	Reduced As(V) sorption to low Fe oxide soils and increased As	Zupancic et al. (2012),
/mine soil			mobility by competitive adsorption	Smith et al. (2002)

Table 2Effect of competition between As and Pi on uptake by plants in hydroponics.

Plant species	Addition	Effect	References
Pteris vittata	Pi (0.1-2 mM)	High Pi decreased As(V) but not As(III) accumulation in	Lou et al. (2010), Tu et al.
		roots/shoot; it enhanced As(V) reduction; high As(V) decreased	(2004), Wang et al. (2002)
		Pi uptake	
Oryza sativa	Pi, 0.1 mM	High Pi concentration decreased As uptake; high As	Lihong and Guilan (2009)
		concentration slightly decreased Pi uptake	
Holcus lanatus	Pi	Decreased As(V) uptake in nonresistant, but less in resistant	Meharg & MacNair (1992)
		plants	
Lemna gibba	Pi, 40 mg/L	High Pi reduced As(V, III) uptake; high As reduced Pi uptake	Mkandawire et al. (2004)
M.sativa	Pi	Strongly suppressed As uptake	Khattak et al. (1991)
Silene vulgaris	Pi, 0.3-3 mg/L	As supply did not influence root growth at high Pi, but did at low	Sneller et al. (1999)
		Pi supply	
Avena sativa	Pi	Decreased As(V) uptake, but little effect on As(III) uptake.	Rumberg et al. (1960)
Glycine max	As, 32-96 μM	Decreased Pi content in soybean organs	Milivojevic et al.(2006)
T. aestivum	Pi	High-affinity uptake system switched on at 25 mM Pi.	Zhu et al. (2006)



Fig. 1. As-Pi interactions and uptake by plants in hydroponics. As and Pi supply variably control As and Pi uptake by plant roots. Pi supply strongly decreases As uptake, while As supply weakly decreases Pi uptake. Pi has higher affinity for plan roots than As.



Fig. 2. Mean shoot biomass, As and Pi concentrations in shoots and total As uptake by wheat plants grown in soil irrigated with As containing water at three concentrations amended and non-amended with Pi (data analysis from Pigna et al., 2009).



Fig. 3. Role of AM in modifying As-Pi interactions, and As and Pi uptake by plant root. The arbuscular mycorrhizal (AM) symbioses increase Pi uptake by plant roots and Pi/As ratio in shoot, while they decrease As uptake by roots.