The Role of Aristaless Related Homeobox (ARX) Gene Mutations in Intellectual Disability

A thesis submitted for the degree of **Doctor of Philosophy** to the **University of Adelaide** by

Tod Fullston B.Sc. (Hons) Student No 1067053

Neurogenetics Laboratory

Discipline of Paediatrics School of Paediatrics and Reproductive Health Women's and Children's Hospital campus

<u>Primary supervisor:</u> Professor Jozef Gécz <u>Co-supervisor:</u> Professor John Mulley

November 2011

TABLE OF CONTENTS

ABSTRACT	9
STATEMENT AND DECLARATION	11
ACKOWLEDGEMENTS	12
TABLE OF ABBREVIATIONS	
1 INTRODUCTION	
1.1 WHAT IS INTELLECTUAL DISABILITY?	
1.2 GENETIC CAUSES OF INTELLECTUAL DISABILITY	
1.2.1 Down syndrome	
1.2.2 Autosomal recessive intellectual disability (ARID)	
1.3 X-LINKED INTELLECTUAL DISABILITY (XLID)	
1.3.1 Syndromic and non-syndromic XLID	
1.3.2 Fragile-X syndrome	
1.4 THE ARISTALESS RELATED HOMEOBOX (ARX) GENE	
1.4.1 Mutations in the ARX gene	
1.4.1.1 Mutations of the polyalanine tract 1	
1.4.1.2 Mutations of the polyalanine tract 2	
1.4.1.3 Mutations that alter homedomain residues	35
1.4.1.4 Mutations in the octapeptide domain	
1.4.1.5 Mutations in the aristaless domain	38
1.4.1.6 Mutations outside of known domains	
1.4.1.6 Copy number variations	40
1.4.2 Phenotypes of affected males with ARX mutations	42
1.4.2.1 Variable clinical expressivity of <i>ARX</i> mutations	45
1.4.2.2 A genotype/phenotype correlation	
1.4.2.3 Phenotypes of female <i>ARX</i> mutation carriers	
1.4.3 Function of ARX	49
1.4.3.1 ARX expression	
1.4.3.2 Mouse models of ARX mutations	51
1.4.3.3 Role of ARX in pancreas?	
1.4.4 Ultraconserved elements around the ARX locus	56
1.5 RESEARCH AIMS	57
2 SUBJECTS, MATERIALS AND METHODS	59
2.1 Patient cohort	59
2.2 Tables of reagents	61
2.3 Tables of primer pairs	64
2.4 PCR formulae and thermocycle profiles	
2.4.1 Standard PCR	69
2.4.2 GC rich PCR (<i>ARX</i>)	69

2.4.3 qPCR	70
2.4.4 Sanger sequencing reaction	
2.4.5 In vitro mutagenesis PCR	71 72
2.5 Gel electrophoresis	
2.6 SSCP/dHPLC screening for ARX mutations	
2.6.1 SSCP mutation screening.	
2.6.2 dHPLC screening	
2.7 In vitro mutagenesis of the ARX ORF	75
2.8 Cloning strategy of polyAlanine tract mutations into ARX ORF	75
2.9 Cell culture	77
2.10 Immunohistochemistry	78
2.11 Fluorescence microscopy	79
2.12 Western immunoblotting	80
2.13 List of URLs	81
3 SCREENING FOR MUTATIONS IN THE ARX GENE AND PUTATIVE ENHA	
3.1 Introduction	
3.2 Strategy for ARX variation screening	86
3.3 Mutations discovered in the ARX coding sequence.	88
3.4 Alterations discovered in ultraconserved (uc) element sequences	
3.5 Non pathogenic variants identified	92
3.6 Discussion	93
4. AN ARX NONSENSE MUTATION c.81 C>G (p.Y27X) LEADS TO BOTH OHTAHARA AND WEST SYNDROME, BUT NOT XLAG	101
4.1 Introduction	101
4.2 Patient and family investigations	
4.3 Cell based studies	109
4.4 Discussion	
5 EFFECT OF VARIATIONS IN THE LENGTH OF POLYALANINE TRACT 1 ARX FUNCTION	O N
5.1 Introduction	118
5.2 Small in frame deletions in pA1 are rare variants	121
5.3 Expansion of pA1 tract	124
5.4 The length of pA1 tract aligns with phenotypic severity and degree of muta ARX protein mislocalisation	
5.5 Discussion	129
6 VARIATIONS IN THE LENGTH OF ARX POLYALANINE TRACT 2	 132

6.1 Introduction	132
6.2 Three new families with the 24 bp duplication mutation	134
6.3 A 27 bp duplication mutation causes infantile spasms and early death	138
6.4 A 33 bp duplication mutation causes Partington syndrome	145
6.5 Mutant ARX protein mislocalises as a function of polyalanine tract 2 length	150
6.6 DNA Sequence analysis of the three duplication mutations	154
6.7 Is the 24 bp duplication mutation recurrent or identical by descent?	160
6.8 Discussion	161
7 MUTATIONS IN THE HOMEODOMAIN OF ARX	165
7.1 Introduction	165
7.2 A c.1074 G>T mutation alters a residue of the homoedomain (p.R358S)	167
7.3 A c.1136 G>T mutation alters a key residue of NLS3 (p.R379L)	170
7.4 Homeodomain mutations cause ARX protein mislocalisation	174
7.5 Discussion	178
8 CHARACTERISATION OF THE DUPLICATION	181
8.1 Introduction	181
8.2 Family information and ARX gene locus duplication discovery	184
8.3 qPCR confirmation and segregation testing	187
8.4 Orientations and fine mapping of the duplication	189
8.5 Investigation of ARX/POLA mRNA in patient's LCLs	191
8.6 Discussion	194
9 FINAL DISCUSSION AND CONCLUSION	197
9.1 ARX screening strategies and recommendations	197
9.2 A complex genotype-phenotype relationship exists	199
9.3 ARX related female phenotypes are complex	200
9.4 Abnormal protein localisation results from mutations	202
9.5 The DNA sequence of ARX is inherently prone to mutation	203
9.6 Is ARX subject to regulation by environmental cues?	204
9.7 Aberrant ARX over/under-expression may confer pathology	206
9.8 Concluding remarks	207
APPENDIX A. PUBLICATIONS ARISING FROM THIS PROJECT	208
APPENDIX B	321
RIRI IOCDA PHV	325

LIST OF FIGURES

Figure 1.1a.	The theoretical normal distribution of IQ scores across the population	<i>17</i>
Figure 1.4a.	The genomic context of ARX at Xp22	<u>29</u>
Figure 1.4b.	Schematic of the full length ARX protein	30
Figure 1.4.1a	a. Schematic summary of known mutations in ARX	31
_	Schematic of PCR, digest and sub-cloning strategy used to generate polyAlanine as from the <i>pCMV-myc-ARX</i> wt vector	<u>76</u>
Figure 3.1a.	Schematic of the 562 amino acid full length ARX protein	84
_	Representative images of the <i>ARX</i> c.429_452dup(24 bp) mutation from agarose gel phoresis or dHPLC analysis	<u>87</u>
Figure 3.3a.	Schematic summary of the six ARX mutations discovered in eight families	89
Figure 3.4a.	Pedigree of AGRE family AU0432 with the uc466 54 G>A variation	90
Figure 3.4b.	Pedigree of AGRE family AU0598 with the uc466 -30 G>T variation	91
_	Schematic of the 3 uc element variants identified in the context of ARX general	omic 92
Figure 3.6a.	Conservation and enhancers 3' of the ARX locus	97
_	The pedigree is compatible with an X-linked mode of inheritance for the c.81C>G on	<u>104</u>
Figure 4.2b.	EEG and MRI results for individuals IV-1 and IV-2 with the c.81C>G mutation	108
_	The relative proportion of co-transfection between the <i>ARX</i> expression construct wit GFP vector	th an
_	The expression of wildtype and p.Y27X mutant myc tagged ARX proteins by wester	rn 112
_	The pedigree of the family with the 9 bp in frame deletion (c.305_313del) in the DN des for pA1 of <i>ARX</i>	IA 122
0	The pedigree of the family with a 3 bp in frame deletion (c.305_307del) in the DNA des for pA1 of <i>ARX</i>	123
Figure 5.4a.	Representative images for the sub-classification of ARX protein localisation	126
_	Proportion of ARX positive cells displaying abnormal localisation for four pA1 trac s compared to wildtype ARX	t <u>127</u>
Figure 6.2a.	Pedigree of family 6.2a with the c.429_452dup(24bp) duplication mutation	134
Figure 6.2b.	Pedigree of family 6.2b with the c.429_452dup(24bp) duplication mutation	136
Figure 6.2c.	Pedigree of family 6.2c with the c.429_452dup(24bp) duplication mutation	137
_	A pedigree of family 6.3 with the c.430_456dup(27bp) mutation within the great graal haplotype	and _ 139

polyalanine tract 2 in exon 2 of the <i>ARX</i> gene (wt and c.430_456dup(27bp))	
Figure 6.3c. Fluorescent fragment analysis of individuals from the c.430_456dup(27bp) famil compared to a wildtype allele	y 143
Figure 6.4a. A pedigree of family 6.4 with the c.423_455dup(33bp) duplication mutation	145
Figure 6.4b. DNA sequence chromatograms of partial sequence of exon 2 of the <i>ARX</i> gene from unaffected individual and the c.423_455dup(33bp) proband	
Figure 6.5a. Proportion of ARX positive cells displaying abnormal for three mutant pA2 tract lengths compared to wildtype ARX	151
Figure 6.5b. Proportion of ARX positive cells displaying abnormal localisation for four pA1 tlengths and three pA2 tract lengths compared to wildtype ARX	
Figure 6.6a. The precise position where mutant sequence commences for each pA2 duplication mutation is ambiguous	on 155
Figure 6.6b. The predicted conformation with the lowest kinetic energy by the mfold algorithm the sequence surrounding the pA2 duplication mutations	
Figure 6.6c. Codons used within pA tracts with similar DNA sequences to that of <i>ARX</i> pA2 (<i>HOXA13</i> , <i>HOXD13</i> and <i>FOXL2</i>)	158
Figure 7.2a. Pedigree of family 7.2a with the c.1074 G>T (p.R358S) mutation	168
Figure 7.2b. DNA sequence chromatograms of wildtype and c.1074 G>T <i>ARX</i>	169
Figure 7.3a. Pedigree of family 7.3a with the c.1136 G>T (p.R379L) mutation	170
Figure 7.3b. Brain MRI findings from an individual with XLAG and the c.1136 G>T mutatio	n <i>171</i>
Figure 7.3c. DNA sequence chromatograms of wildtype and c.1136 G>T <i>ARX</i>	173
Figure 7.4a. Proportion of ARX positive cells displaying abnormal localisation for three muta the homeodomain compared to wildtype ARX	ntions ir 175
Figure 7.4b . Representative images of the localisation of the c.1074 G>T (p.R358S) mutant construct	176
Figure 8.2a. Pedigree of the large XLID family investigated for the ARX gene locus duplication	on_ <i>184</i>
Figure 8.2b. Relative Xp22 probe intensities from the comparative genome hybridisation usin DNA from the proband with an <i>ARX</i> gene locus duplication	_
Figure 8.3a. Summary of qPCR assays to refine the size of the <i>ARX</i> gene duplication	188
Figure 8.4a. Schematic of 3 theoretically possible orientations of the <i>ARX</i> gene duplication at and PCR strategies used to resolve them	•
Figure 8.4b. Products from orientation specific PCRs_	191
Figure 8.5a. Products from a RT-PCR specific to either scrambled or wt <i>ARX</i> mRNA	193
Figure 9.1a. UCSC genome browser image of human <i>ARX</i> locus with a custom exome sequer coverage track enabled	ncing 198
Figure 9.6a. UCSC genome browser image of human <i>ARX</i> locus showing methylation	205

LIST OF TABLES

Table of Abbreviations	12
Table 1.3a. Recommendations for exclusion criteria for investigation and diagnosis of a male with XLID	child 23
Table 1.3.1a. Genes that cause both nsXLID and sXLID when mutated	24
Table 1.4.1.3a. Truncation or missense mutations in the homeodomain of ARX	36
Table 1.4.1.5a. Mutations in DNA that codes for the aristaless domain of ARX	39
Table 1.4.1.6a. Mutations outside of known domains within ARX	40
Table 1.4.2a. Ten clinically distinct phenotypes are observed for mutations in the ARX gene_	43
Table 1.4.4a. The ultraconserved elements flanking the human ARX gene	56
Table 2.1a. Subsets of patients within the heterogeneous cohort screened for mutations in the ORF and ultraconserved elements	ARX 60
Table 2.2a. Details of reagents used in PCR and DNA electrophoresis protocols	61
Table 2.2b. Details of reagents used for plasmid preparations, cloning and bacterial culture	62
Table 2.2c. Details of reagents used for mammalian cell based investigations and western immunoblotting	63
Table 2.3a. PCR/Agarose gel electrophoresis screening and mutant polyalanine tract sub-clon primers	ing 64
Table 2.3b. Hexachlorofluorescein labelled SSCP primers	64
Table 2.3c. dHPLC primers	65
Table 2.3d. Primers used for haplotype analysis of the region flanking the <i>ARX</i> locus	66
Table 2.3e. Primers used for copy number variation qPCR	67
Table 2.3f. Mutagenic primers used for site directed mutagenesis	68
Table 2.3g. Primers used for sequence confirmation of mutant pCMV-myc-ARX ORFs	68
Table 2.6a. dHPLC denaturation temperatures and time-shifts used	74
Table 3.1a. Ten clinically distinct phenotypes are observed for mutations in the ARX gene	84
Table 3.1b. The ultraconserved elements flanking human the ARX gene	86
Table 3.3a. Details of the six mutations identified	89
Table 3.4a. DNA sequence variation identified in the uc elements	92
Table 3.5a. Details of presumed non-pathogenic variants identified	93
Table 3.6a. Subsets of patients within the heterogeneous cohort and mutations/sequence varia discovered within them	tions 94
Table 4.1a. Summary of mutations in ARX that cause Ohtahara syndrome	101
Table 5.4a. Amount of ARX protein mislocalisation per pA1 <i>pCMVmycARX</i> expression co 24h post transfection	onstruct 128

6.3a. STS marker positions by UCSC genome browser for <i>DXS8099-DXS8027-ARX-DXS1202-DXS8047</i> at Xp22.11-Xp21.3, relative to the <i>ARX</i> locus	141
6.3b. Amplification of wildtype and c.429_452dup(24bp) and c.430_456dup(27bp) mu <i>ARX</i> alleles from female carrier's DNA sourced from various tissues	ıtant 144
6.5a. Amount of ARX protein mislocalisation per pA2 <i>pCMVmycARX</i> mutant expression construct 24h post transfection	153
6.7a. STS marker haplotype for <i>DXS8099-DXS8027-ARX-DXS12027-DXS8047</i> at Xp21.3-Xp22.11 for probands with an <i>ARX</i> duplication mutation from 12 separate families	
7.4a. Amount of ARX protein mislocalisation per homeodomain <i>pCMVmycARX</i> me expression construct 24h post transfection	ıtant <i>177</i>
of published <i>ARX</i> mutations, ordered by the position at which they occur in the ORF (<i>APPENDIX B</i>)	209

ABSTRACT

Intellectual disability (ID) affects $\sim 1-3\%$ of the population, profoundly impacting the lives of affected individuals and their families. An approximate 30% excess of males with ID implicates X-chromosome genes. The most common inherited form of ID is fragile-X syndrome, affecting $\sim 1/5,000$ live male births. Another X-linked gene, the aristaless related homeobox (ARX) gene, is also frequently mutated causing X-linked ID (XLID).

At least 50 pathogenic mutations spanning the *ARX* open reading frame (ORF) have been reported in 110 families. These mutations cause at least 10 clinically distinct pathologies, all of which include ID. These clinical entities range in severity from X-linked lissencephaly with ambiguous genitalia (XLAG) to mild ID with no other consistent clinical features.

Of the known *ARX* mutations 60% occur in the section of the ORF that encodes for the first two tracts of uninterrupted alanine, *ie* polyalanine (pA) tracts. This is likely due to the extraordinarily high GC content of these regions of the gene (>97%). Two recurrent mutations (c.304ins(GCG)₇ – pA1 and c.429_452dup – pA2) arise from expansion of their respective pA tracts. The c.429_452dup mutation alone accounts for ~40% of all reported *ARX* mutations.

To assess the frequency of *ARX* mutations among the intellectually disabled, genomic DNA from 613 individuals were screened for the most frequent *ARX* mutations. Of these, 500/613 samples were screened for mutations in the entire *ARX* ORF by either SSCP, dHPLC or direct Sanger sequencing. A subset of 94/500 patients were also screened for sequence variations in ultraconserved (uc) elements flanking the *ARX* gene, which likely act as *ARX* enhancers. Subsequently, using transient transfection studies we assessed the subcellular localisation of selected mutations and wildtype ARX proteins.

Six different *ARX* mutations were detected in eight individuals (8/613; 1.3%) and potentially pathogenic sequence variations were found in uc elements in three more individuals. A total of five duplication mutations were discovered in pA2, two larger than the recurrent c.429_452dup, confirming exon 2 of *ARX* as a mutation 'hot spot'. Increased aggregation was observed as a function of pA1 and pA2 length, aligning with the patient's phenotypic severity.

Three missense mutations were detected. A familial c.81G>C mutation caused a premature termination codon in exon 1, leading to Ohtahara syndrome (OS) and West syndrome (WS) in two male cousins. Although the c.81G>C mutation should truncate the ARX protein, reinitiation of translation at a down-stream methionine codon (c.121_123) likely occurs, 'rescuing' these patients from the otherwise severe XLAG phenotype.

Two point mutations (c.1074G>T/p.R358S; c.1136G>T/ p.R379L) that alter key residues within the homeodomain were found in two individuals with brain/genital malformations and led to increased ARX protein mislocalisation. These mutations impair vital properties of ARX's transcription factor function by perturbing its localisation into the nucleus (p.R379L) or DNA binding (p.R358S).

This study confirms that *ARX* mutations contribute significantly to XLID and that the majority of mutations occur within exon 2, specifically within the region of pA2. Moreover, there is a correlation between the subcellular localization of the mutant protein and the clinical severity in the patients.

STATEMENT AND DECLARATION

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution to **Tod Fullston** and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue, the Australasian Digital Theses Program (ADTP) and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Signed:	Date:
ε	

ACKOWLEDGEMENTS

I sincerely thank members of the Gécz Neurogenetics Program and the Mulley epilepsy research team for their hospitality, assistance and mentoring. Specifically, I thank Bree Hodgson (SSCP), Xenia Iona and Merran Finniss (dHPLC) for their collaboration which I found invaluable for mutations screening. The people involved in the process can make the journey worthwhile and I want to thank the following people individually:

<u>Dr Mark Corbett</u>: For his companionship and always humouring me by not looking too bored during repeated conversations about aligning DNA sequences from mutant polyalanine tracts.

<u>Marie Shaw</u>: Thanks for getting me the job that led to my PhD studies and your friendship throughout my time in the Gécz Neurogenetics laboratory.

<u>Dr Cheryl Shoubridge</u>: Who acted as my surrogate supervisor and who kindly collaborated with me in developing *ARX* expression constructs.

The ghosts of ARX past: Marie Mengelsdorf, Olivia McKenzie and Desiree Cloostermann whose prior work laid the foundation for my project.

<u>Prof. Jozef Gécz</u>: I sincerely thank you for opportunity to undertake research in your lab and provide me with space/resources to do so, and the many valuable lessons you have taught me.

<u>Prof. John Mulley</u>: I may have managed to get lost along the way without your sage guidance and I have greatly appreciated your down to earth mentorship.

Both of my supervisors have managed to inspire me to work towards a career in research through their critical thinking, leadership, mentoring, guidance and expertise.

I would also like to express my gratitude toward my colleagues in my new adoptive lab, who have been very understanding and generously supportive of my situation.

Finally I must thank my out of lab support crew in the form of my long suffering wife and my two most precious daughters who have provided unconditional love and support for 'Team Tod'. My daughters have introduced a beautiful chaos into my life, which makes life so much more rich and rewarding.

'Constant dripping hollows out a stone.' Lucretius.

^{&#}x27;Long is the way, and hard, that out of hell leads up to light.' John Milton.

TABLE OF ABBREVIATIONS

Abbreviation	Full description	
A, C, G, T	adenosine, cytosine, guanine, thymine – nucleotides	
aa	amino acid	
ACC/AG	absence of the corpus callosum with abnormal genitalia	
ADI-R	autism diagnostic interview – revised	
AG-K	abnormal genitalia	
AGRE	autism genetic research exchange	
ANOVA	analysis of variance	
ARID	autosomal recessive intellectual disability	
aut	autism	
BERA	brainstem evoked response audiometry	
BLAST	basic local alignnment tool	
BLAT	BLAST-like alignment tool	
bp	base pairs	
c.	coding sequence	
CA	cytoplasmic positive with or without aggregates in either the nucleus or cy	vtonlasm
cDNA	complimentary DNA	topiusiii
CGH	comparative genome hybridisation	
CNS	central nervous system	
CNV	copy number variation – deletion or duplication (> 1 kb)	
CSF	cerebrospinal fluid	
CT	computerised tomography (scan)	
DAPI	4',6-diamidino-2-phenylindole	
del	deletion	
dHPLC	denaturing high pressure liquid chromatography	
DMSO	dimethyl sulphoxide	
DNA	deoxyribonucleic acid	
dNTP	deoxynucleoside triphosphate	
dup	duplication	
EEG	electroencephalogram	
EFMR	epilepsy and mental retardation limited to females	
EIEE	early infantile epileptic encephalopathy	
EMG	electromyogram	
epi	epilepsy	
exp	expansion	
FosTeS	fork stalling and template switching	
FXS	fragile-X syndrome	
FXTAS	fragile-X associated tremor/ataxia syndrome	
GABA	gamma-aminobutyric acid	
HEK293T	human embryonic kidney; cell line 293T hexachlorofluorescein	
hex		
HYD/AG	hydranencephaly with abnormal genitalia	
ID/TS/Dye	intellectual disability intellectual disability with tonic seizures with dystonia	
ID/TS/Dys IEDE	infantile epileptic-dyskinetic encephalopathy	
iGOLD	international genetics of learning disability	
indels	insertions and deletions and (<1 kb)	
ins	insertions and deletions and (<1 kb)	
IIS IQ	intelligence quotient	
ISSX	infantile spasms syndrome, X linked	
kb		Cont. next page)
AU	Kito ouse pair	com. nen page)

Abbreviation	Full description	Cont.
LCL	lymphoblastoid cell line	_
LGS	Lennox-Gastaut syndrome	
LOD	logarithm of the odds	
Mb	mega base pair	
MGB	minor groove binder	
milliQ	ddH ₂ O water from a Millipore milliQ system	
MIM	Mendelian inheritance in man – online reference	
miRNA	micro ribonucleic acid	
mis	missense	
MRI	magnetic resonance imaging	
mRNA	messenger ribonucleic acid	
MRS	magnetic resonance spectroscopy	
NGS	next generation sequencing	
NI	nuclear inclusions only	
NLS	nuclear localisation sequence	
NMD	nonsense mediated decay	
non	nonsense	
nsXLID	non syndromic X-linked intellectual disability	
OCF	occipital-frontal circumference	
ORF	open reading frame	
OS	Ohtahara syndrome	
	protein residue	
p.	polyalanine tract	
pA PAGE	* *	
PBS	polyacrylamide gel electrophoresis phosphate buffered saline	
PCR	polymerase chain reaction	
PRTS	Partington syndrome	
PS	• •	
PTC	Proud syndrome premature termination codon	
qPCR	•	
RNA	quantitative real-time polymerase chain reaction ribose nucleic acid	
RNA BT DCD	ribonucleic acid	
RT-PCR	reverse transcription polymerase chain reaction	
SDS	sodium dodecyl sulfate	
sil	silent	
SNP	single nucleotide polymorphism	
SSCP	single stranded conformational polymorphism	
STS	sequence-tagged site	
sXLID	syndromic X-linked intellectual disability	
Taq	Thermus aquaticus	
TBS	tris-buffered saline	
uc	ultraconserved elements	
UCSC	University of California, Southern California	
UTR	untranslated region	
UV	ultraviolet	
WS	West syndrome	
wt	wildtype	
XLAG	X-linked lissencephaly and ambiguous genitalia	
XLID	X-linked intellectual disability	
XMESID	X-linked myoclonic epilepsy with spasticity and intellectual disability	