Genetic Control of Hypothalamo-Pituitary Axis Development and Function in Mice

Eva Szarek, BSc (Hons)



A thesis submitted for the degree of

Doctor of Philosophy

March 2011

School of Molecular and Biomedical Science

Discipline of Biochemistry

The University of Adelaide

Advisors: Associate Professor Paul Thomas

Professor Jeffrey Schwartz

© 2011 Eva Szarek

Discipline of Biochemistry School of Molecular and Biomedical Science The University of Adelaide Adelaide Australia 5000

Printed on acid-free paper at Kwik Kopy, Adelaide, South Australia.

For my parents,

Jan and Dorota Szarek

"Glands rarely become ill, but when they do, they give their disease to the rest of the body"

Hippocrates, Glands, circa 500 B.C.

CONTENTS

List of Figures ix
List of Tables xi
A Note on Nomenclature xiii
Abstract xiv
Statement of Contribution by Others to this
Work xvii
Declaration of Originality xviii
Acknowledgements xix
Acronyms and Abbreviations xx
Publications xxii
Conference Preceedings xxii

1. INTRODUCTION

I. Molecular Genetics of the Hypothalamic-Pituitary Axis 23

- A. Vertebrate Hypothalamic Development 23
 - 1. Structure and function of the hypothalamus 24
 - 2. Development of the hypothalamus 25
- B. Vertebrate Pituitary Development 26
 - 1. Structure and function of the pituitary
 - 2. Development of the pituitary 30
 - 3. Early patterning of the pituitary 31 a. Signaling molecules 32
 - b. Transcription factors 34
- C. The Hypothalamo-Pituitary Axis 38
 - 1. Anatomical and functional connections 38
 - 2. Blood supply of the hypothalamopituitary axis 38
 - 3. Angiogenesis 41
 - 4. Importance of VEGF and VE-Cadherin in vascular development 43

II. Sox Family of Transcription Factors 50

- A. The SOX Family 50
- B. The SOXB1 Subgroup 50
- C. The Role of Sox3 in Hypothalamo-Pituitary Axis Development 56

- 1. Sox3 is expressed throughout early embryonic development 56
- 2. Sox3 plays an important role during brain development 56
- 3. Importance of genetic background 57

III. Consequences of Mutations in Transcription Factors: Congenital Hypopituitarism 58

IV. The Growth Hormone Axis 59

- A. Growth-Hormone and Growth Hormone-Releasing Factor 59
- B. Growth Hormone Deficiency 63

V. Generation of Novel Mice by ENU Mutagenesis 64

VI. Hypothesis, Aims and Significance 66

- A. Project 1: Identification of Sox3 Target Genes 66
- B. Project 2: Novel Dwarf Mouse Generated by ENU Mutagenesis 66

2. MATERIALS AND METHODS

I. Buffers and Solutions 67

- A. Commercially Obtained 67
 - 1. Compounds, buffers and solutions 67
 - 2. Histology 67
 - 3. Indicators, antibodies and enzymes 68
 - 4. Specialty kits 69
 - 5. Preparation of DNA oligonucleotides 69
- B. Laboratory Prepared Buffers and Solutions
 71

II. Mouse Breeding and Lines 72

- A. Maintenance and Breeding 72
 - 1. General maintenance 72

- 2. Timed matings 72
- B. Mouse Lines 72
 - 1. Sox3 transgenic lines 72
 - a. Sox3-null 73
 - b. Extra-Sox3 73
 - c. Sox3-GFP reporter (Green-Sox3) 73
 - 2. Dwarf Mouse Line Generated by ENU Mutagenesis 74

III. Embryo and Tissue Collection 76

- A. Embryo Collection 76
- B. Tissue Collection and Processing 77
 - RNA processing of mouse embryonic 10.5 dpc mouse heads used in microarray analysis 77
 - RNA processing of hypothalamic sections used in mRNA expression analysis by qPCR 78
 - 3. Isolation of protein from whole pituitaries for GH analysis 78
 - 4. Fixation and Tissue Preparation 79
 - a. Frozen Sections 79
 - b. Paraffin Sections 79
- C. Preparation of Tail Tip Genomic DNA for PCR Genotyping 79

IV. PCR Genotyping 80

- A. Sox3 Transgenic Lines 80
- B. ENU Generated Dwarf Mice 81

V. Mouse Physiological Studies 82

- A. Growth Analysis of Dwarf Mice 82
 - 1. Weight over time 82
 - a. Post-weaning 82
 - b. Pre-Weaning 82
 - 2. Body Length 83
- B. Pituitary Growth Hormone Levels 83
- C. Blood Biochemistry: Examining IGF-1 levels 83
- D. Expression of Hypothalamic GHRH and Sst by aPCR 84
- E. Statistical Analysis 84

VI. Purification of DNA for Sequencing 84

- A. Purification of DNA from Agarose Gels 84
- B. Sequencing 84

VII. Bacterial Techniques 85

- A. Media and Solutions 85
- B. Preparation of Chemically Competent E.coli
- C. Bacterial Transformation by Heat Shock 85
- D. Purification of Plasmid DNA 86

VIII.Fluorescence Immunohistochemistry 87

IX. In Situ Hybridization 87

- A. Purification of Plasmid DNA by Restriction Enzymes 87
- B. Transcription Reaction and Generation of In Situ Hybridization Probes 88
- C. In Situ Hybridization 88

X. Morphology Stain 88

- A. Hematoxylin and Eosin 88
- B. Cresyl Staining 89
- C. Masson Trichome 89

XI. Protein Immunblot 90

- A. Tissue Collection 90
- B. Whole Cell Extract 90
- C. Determining Protein Concentration Using Bradford Protein Assay 90
- D. Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis 91
- E. Protein Immunblot (Western Blot Preparation) 91

XII.Cell Dissociation 91

- A. Method 1: Cell Dissociation using Trypsin 92
- B. Method 2: Cell Dissociation using Dispase II and Collagenase B 92

XIII. Fluorescence Activated Cell Sorting 92

XIV. Microarray Using The Illumina BeadChip 93

- A. RNA Preparation 93
- B. Analysis of RNA Quality 93
- C. The Illumina® BeadChip Technology 96
- D. Microarray Processing 98
 - 1. RNA amplification 98
 - 2. Hybridization to the Illumina® BeadChip
 - 3. Array design 99
- E. Data Collection and Analysis 99
 - 1. Statistical programming environment for analyzing microarray data 99
 - 2. Normalizing data 99
 - 3. Statistical analysis to determine differentially expressed genes 100
 - 4. Data collation 101
- F. Criteria for the Identification of Differentially Expressed Genes 101

XV.cDNA Generation 102

XVI. qPCR 102

XVII.Software Programs 103

3. IDENTIFICATION OF SOX3 TARGET GENES

I. Introduction 105

II. Aims 107

III. Results 108

Aim 1: Identify Potential Sox3 target Genes by Microarray Analysis 108

- A. Cell Dissociation and FACS sorting of GFP-positive cells for use in Microarray Analysis 108
- B. Extraction of RNA from Whole Mouse Embryonic Heads for use in Microarray Analysis - RNA Quality Analysis 111
- C. Microarray Analysis 111
- D. Normalizing Microarray Data 116
- E. Identification of Differentially Expressed Genes 120
- F. Genes Chosen for qPCR Validation 122

Aim 2: Confirm Microarray Identified Potential Sox3 Target Genes by qPCR 127

G. Validation of Microarray Data by qPCR 127

Aim3: Expression of Sox3 and Sox3 Target Gene(s) 132

- H. Ngn3 is Expressed in the Developing Hypothalamus 132
- I. Ngn3 is Co-expressed with Sox3 in the Developing Hypothalamus 132

IV. Discussion 141

- A. FACS Does Not Yield Enough GFP+ Cells from Sox3-null 10.5 dpc Mouse Embryonic Heads 141
- B. Microarray Analysis Validation 143
- C. Microarray Analysis and qPCR Validation Reveal Ngn3 as a Likely Target Gene of Sox3 145
- D. Expression Studies by Immunohistochemistry Reveal that Sox3 and Ngn3 are Co-expressed in the Developing Hypothalamus 147
- E. Conclusion and Future Directions 148

4. NOVEL DWARF MOUSE GENERATED BY ENU MUTAGENESIS

I. Introduction 151

II. Aims 152

III. Results 154

Aim 1: Characterize the primary pathology of the dwarfism phenotype that links the function of the mutation to dwarfism by focusing on altered regulation Of the GH axis 154

- A. Dwarf Mice Show a Reduced and Sustained Decrease in Weight and Growth 154
- B. Dwarf Mice Show Decreased Pituitary GH and Serum IGF-1 Levels 156
- C. Dwarf Mice Show a Pronounced Hypoplasia of the Anterior Pituitary Gland 156
- D. Histopathology of the Dwarf Mouse Brain 157
- E. Hypothalamic GHRH and Somatostatin Levels Are Significantly Reduced in Dwarf Mice 160

Aim 2: Confirm the Mutation by Sequencing 164

F. The Dwarf Mutation is a Non-Conservative Substitution of Leucine to Proline in the Gene Tryptophanyl-tRNA Synthetase 164

Aim 3: Examine the expression of the mutant protein 170

- G. Wars is Expressed in Pituitary Vasculature
 - 1. Wars is co-expressed with PECAM in pituitary vasculature 170
 - 2. VE-Cadherin and Wars are expressed in pituitary vasculature 173
 - 3. Steady state level of Wars protein is not altered in the pituitary 178

Additional and Preliminary Data 179

- H. Wars^{L30P} Mutation Affects Angiogenesis
 - 1. Generation of Wars Isoforms 179
 - 2. Angiostatic activity of WARS isoforms 179
- I. Vascularity in the Mouse Brain and Pituitary 182
- J. Comparison of the Major Organs Reveals Proportionate Decrease in Size 184
- K. Dwarf Mice Show Delayed Gonadal Development and are Sub-fertile 184

IV. Discussion 190

- A. Wars^{L30P} Dwarf Mice are Proportionally Smaller with Pituitary Hypoplasia 191
- B. Pituitary GH and Serum IGF-1 are Reduced in Wars^{L30P} Dwarf Mice 192
- C. Hypothalamic Ghrh and Sst Expression Levels are Reduced in Wars^{L30P} Dwarf Mice 192
- D. Dwarf Mice Show Delayed Reproductive Development 193
- E. Wars is Expressed Within Blood Vessels of the Pituitary 194
- F. Wars^{L30P} Mutation Inhibits the Formation of New Vessels in Cell Culture 199
- G. Conclusion and Future Directions 200

APPENDICES 203

PERL script used in the collation of microarray statistical data 203

Microarray DATA showing differentially expressed genes 210

Three-dimensional cerebral vasculature of the CBA mouse brain: Circle of Willis 230

Three-dimensional analysis of vascular development in the mouse embryo 231

Microarray Validation by qPCR 232

Generatioin of Mouse Wars Isoforms 234

REFERENCES 236

INDEX 248

PUBLICATIONS 250

LIST OF FIGURES

Figure 1-1 Illustration of the organization of the hypothalamic nuclei in the mouse brain	25
Figure 1-2 Schematic representation of the structure of the pituitary	28
Figure 1-3 Schematic representation of the stages of pituitary development in rat and mice	32
Figure 1-4 Ontogeny of signaling molecules and selected transcriptional factors during mouse pituitary organogenesis	35
Figure 1-5 Anatomical location of the hypothalamus and pituitary in humans	39
Figure 1-6 The hypothalamic-pituitary axis	40
Figure 1-7 Diagrammatic representation of the blood supply and venous drainage of the hypothalamo- pituitary axis	41
Figure 1-8 Vasculogenesis and angiogenesis	42
Figure 1-9 Angiogenesis	44
Figure 1-10 The VEGF:VE-Cadherin Pathway	49
Figure 1-11 SOX family of proteins showing homology relationship	52
Figure 1-12 Expression of Sox1, Sox2, and Sox3 in the developing mouse pituitary and central nervous system	55
Figure 1-13 Abnormal morphogenesis of the hypothalamus, pituitary and midline CNS in Sox3-null mice58	
Figure 1-14 Major actions of growth hormone	61
Figure 1-15 GHRH signaling pathway	62
Figure 1-16 Spontaneous and experimental alterations in the GHRH signaling pathway that result in either somatotrope hypoplasia or hyperplasia	63
Figure 2-1 Strategy of ENU breeding for screening recessive pedigrees	75
Figure 2-2 Embryo dissection at 10.5 dpc showing live GFP in Sox3-null embryos	76
Figure 2-3 Theiler staging of mouse embryos between 9 – 15 dpc	77
Figure 2-4 Schematic representation of hypothalamic dissection in 8-week old mice	78
Figure 2-5 Electropherograms used in the analysis of RNA quality using the Agilent 2100 Bioanalyzer	95
Figure 2-6 Physical layout of twelve equally spaced strips in a Illumina® Sentrix-6 BeadChip	96
Figure 2-7 Schematic view of an Illumina® bead coupled with an oligonucleotide, consisting of the address code and a 50 base gene-specific sequence	97
Figure 3-1 Schematic representation of the wild-type, Sox3-null, Extra-Sox3 and Green-Sox3 (GFP-reporter) mice.	106
Figure 3-2 Fluorescence activated cell sorting of GFP+ mouse 10.5 dpc embryonic heads comparing two cell dissociation methods: trypsin (method 1) and a combination of dispase II and collagenase B (method 2)	112
Figure 3-3 RNA quality from whole mouse embryo heads used in microarray analysis	114
Figure 3-4 Microarray experimental outline and design	115
Figure 3-5 Box-plots showing intensity distributions before and after normalization of non-background subtracted data sets	117
Figure 3-6 M versus A plots prior and after quantile normalization of background subtracted arrays	119

Figure 3-7 q	PCR analysis showing relative quantitation of 10.5 dpc embryonic heads showing the expression profile of four microarray identified genes Sox3, Nfya, Nenf, Sfrp1 and Ngn3 in wild-type and Sox3-null mice	130
Figure 3-8 q	PCR analysis showing relative quantitation of 10.5 dpc embryonic heads showing the expression profile of Ngn3 and Sox3 in wild-type, Sox3-null and Extra-Sox3 mice	130
Figure 3-9 q	PCR analysis showing relative quantitation of 10.5 dpc embryonic heads used in microarray analysis showing the expression profile of Ngn3 and Sox3 in wild-type and Sox3-null mice	131
Figure 3-10	Expression of ngn3 in the developing hypothalamus of wild-type and Sox3-null 12.5 dpc coronal sections by in situ hybridization	135
Figure 3-11	Ngn3 is co-expressed with Sox3 in the developing hypothalamus at 10.5 dpc wild-type and Sox3-null mice – coronal orientation.	136
Figure 3-12	Expression of Ngn3 and Sox3 in the developing hypothalamus at 12.5 dpc wild-type and Sox3-null mice – coronal orientation	137
Figure 3-13	Expression of Ngn3 and Sox3 in the developing hypothalamus at 12.5 dpc in wild-type and Sox3-null mice – sagittal orientation	138
Figure 3-14	Expression of Ngn3 and Sox3 in the developing mouse hypothalamus at 14.5 dpc – coronal orientation	139
Figure 3-15	Model of Ngn3 and Sox3 cells during mouse HP axis development between 10.5 – 12.5 dpc.140	
Figure 4-1 A	Aminoacylation of tRNA	153
Figure 4-2 I	Body weight and length of dwarf and control littermates	155
Figure 4-3 (Gross brain morphology of dwarf and wild-type littermates	158
Figure 4-4 n	nRNA expression of Ghrh and Sst in wild-type and dwarf hypothalamic extracts	161
Figure 4-5 V	Nhole-pituitary GH and serum IGF-1 levels including pituitary gland weight and expression of GH in wild-type and dwarf littermates	162
Figure 4-6 T	The dwarf region showing sequence confirmation	165
Figure 4-7	Гhe WARS protein	168
Figure 4-8 I	Expression of WARS in the mouse pituitary at 8-weeks	171
Figure 4-9 I	Expression of WARS and PECAM (CD-31) in female wild-type and dwarf mouse pituitary at 8 weeks	172
Figure 4-10	Action of WARS on VE-Cadherin and proposed role of the Wars ^{L30P} mutation during angiogenesis	175
Figure 4-11	Expression of VE-Cadherin and Wars in wild-type and dwarf mouse pituitary at 8 weeks	177
Figure 4-12	Western blot analysis of Wars expression in pituitaries, brains and kidney in wild-type and dwarf mice	178
Figure 4-13	Schematic representation of the mouse tryptophanyl-tRNA (Wars) synthetase isoforms used in the tube-formation assay	180
Figure 4-14	Preliminary data of tube-formation assay showing that the Wars ^{L30P} mutation has angiostatic properties	181
Figure 4-15	Vascular pattern in the cerebral cortical surface, lateral hypothalamus and pituitary from wild-type and dwarf brains and pituitary at 8-weeks	182
Figure 4-16	Gross morphology of organs of 8-week old age matched wild-type and dwarf littermates	186
Figure 4-17	Gross morphology and histology of reproductive organs from wild-type and dwarf mice	188
Figure 4-18	Schematic representation of the human full length and truncated WARS	197

Figures in Appendices

Figure A 1 Circle of Willis on mouse brain surface and all arteries on brain surface and a slice plane	230
Figure A 2 Development of the cephalic plexus between the 5 and 20 somite in the mouse embryo	231
Figure A 3 Cloning strategy used for generating mouse Wars isoforms	235

LIST OF TABLES

Table 1-1 Hormone secretions of the anterior lobe of the pituitary gland and their control	30
Table 1-2 Signal pathways and transcription factors critical for pituitary and hypothalamic development and function	36
Table 1-3 Important role and functions of VEGF during blood vessel formation	46
Table 1-4 SOX family of proteins	51
Table 1-5 Expression and biological function of SoxB1 members	54
Table 1-6 Phenotypes of Sox3 transgenic mouse models	57
Table 1-7 Transcription factors that affect pituitary function and are associated with autosomal forms of congenital hypopituitarism	60
Table 1-8 Isolated growth hormone deficiencies associated with severe short stature	65
Table 2-1 Compounds, buffers and solutions	67
Table 2-2 Indicators and antibodies	68
Table 2-3 Antibodies used in the detection of proteins by immunofluorescence analysis	68
Table 2-4 Secondary fluorescence antibodies used in the detection of proteins by immunofluorescence	68
Table 2-5 Enzymes	69
Table 2-6 Specialty kits	69
Table 2-7 PCR primers for genotyping Sox3-null mice	70
Table 2-8 PCR primers for genotyping Sox3 transgenic and GFP reporter mice	70
Table 2-9 PCR primers for genotyping the dwarf mouse line	70
Table 2-10 qPCR primers used for the validation of microarray results (Project 1)	70
Table 2-11 qPCR primers used for analyzing Ghrh and Sst in dwarf mouse hypothalamic extracts (Project 2)	71
Table 2-12 Laboratory prepared general buffers and solutions	71
Table 2-13 PCR analysis cycling conditions for Sox3 transgenic and null mouse lines	81
Table 2-14 PCR analysis cycling conditions for dwarf mouse lines.	81
Table 2-15 Mouse numbers used in growth analysis over time	82
Table 2-16 Mouse numbers used in growth analysis at P1, P7 and P14	83
Table 2-17 Bacterial growth media composition	85
Table 3-1 Down-regulated genes identified by microarray analysis and t-test statistical analysis in Sox3-null 10.5 dpc embryonic heads	123
Table 3-2 Down-regulated genes identified by microarray analysis using three statistical analyses (LIMMA_SAM_and t-test) in Sox3-null 10.5 dnc embryonic heads	126

Table 3-3 List of four differentially expressed genes chosen for validation by qPCR showing the fold change as determined by the three statistical tests. Expression location of these genes is also shown	127
Table 4-1 Candidate genes in the dwarf critical region	166
Table 4-2 Reproductive fitness of dwarf mice	185
Table 4-3 Phenotypic characteristics comparing GH/IGF-1 long-living mutant mice with the L30P dwarf mouse	195
Table 4-4 Noncanonical activities of AARSs in vascular development	196
Tables in Appendices	
Table A 1 Genes used in the validation of microarray data	232
Table A 2 Average intensity values for Xist, Sox3, β-Actin and Gapdh	233

A NOTE ON NOMENCLATURE

Relevant nomenclature guidelines were taken into account when referring to genes and gene products throughout this thesis. To unambiguously refer to mouse (*Mus musculus*) genes and gene products, and to distinguish these from mammalian nomenclature, the following conventions were adhered to. Mouse gene names are italicized and in lower case, whereas gene products are non-italicized and the first letter is capitalized. Human gene names are italicized and all capitalized, whereas proteins are non-italicized and all capitalized. In addition, reference may be given to *Drosophila* genes and gene products. To differentiate these from mouse and/or human genes and gene products *Drosophila* genes are italicized and the protein are non-italicized and in lower case. Additionally, when referring to both the gene and protein, the protein name is given.

Species	Gene (abbreviation)	Protein (abbreviation)
Mouse	Sox3	Sox3 or mSox3
Human	SOX3	SOX3 or hSOX3
Drosophila	sox3	sox3

With reference to the Sox3 knock-out, transgenic and reporter mice, these will be referred within this thesis as follows:

Mouse Line	Nomenclature within thesis
Sox3-null	Sox3-null
Sox3-transgenic (Sox3 ^{iRES-eGFP})	Extra-Sox3
Sox3-GFP reporter	Green-Sox3

With reference to the novel dwarf mouse line described herein, we have given this mouse line the name Tukkuburko. The name is the Kaurna Aboriginal word referring to "small mouse". The Kaurna Indigenous people are the custodians of the greater Adelaide region and their cultural and heritage beliefs are still important to the living Kaurna people today.

ABSTRACT

Congenital dysfunction of the hypothalamic-pituitary (HP) axis occurs in approximately one birth per 2,200 and is associated with a broad range of common disease states including impaired growth (short stature), infertility, hypogonadism poor responses to stress and slow metabolism (Pescovitz and Eugster, 2004). Although, a number of genes have been linked to diseases of the HP axis, the genetic cause in many patients remains unknown.

This thesis examines two aspects of HP axis development and function. The first aim was to identify *Sox3* targets by examining gene expression differences between three mouse lines: *Sox3*-null (mice lacking *Sox3*; loss of function), Extra-Sox3 (mice over-expressing Sox3; gain of function) and wild-type, by genome wide profiling using the Illumina BeadChip microarray platform. The second aim was to characterize the downstream effects relative to HP development in a novel recessive dwarf mouse model with pituitary hypoplasia and growth hormone (GH) deficiency, generated by N-ethyl-N-nitrosourea (ENU) mutagenesis that produces a point mutation in the gene for the enzyme tryptophanyl-tRNA synthetase (WARS).

The first project (project 1) examined Sox3, the causative gene associated with Xlinked hypopituitarism (XH), in wild-type and transgenic mice. SOX3 is a member of the SOX (SRY-related HMG box) gene family of transcription factors that is expressed in progenitor cells of the mouse embryonic central nervous system (CNS) including the developing and postnatal hypothalamus (Rizzoti et al., 2004). It is the only member of the SOXB1 subfamily positioned on the X chromosome (Collignon et al., 1996; Stevanovic, 2003). Appropriate dose- and time-dependent expression of Sox3 in the developing hypothalamus is required for normal neuroendocrine function, particularly related to growth and growth hormone (GH). Changes associated with a loss-of-function and/or gain-of-function of Sox3 may contribute to a better understanding of other important genes, currently not known, involved in XH and/or X-linked mental retardation. At this point, however, the mechanisms linking SOX3 to its direct targets and their interplay within other downstream signaling cascades regulating HP axis development remain unknown. In order to identify Sox3-dependent genes, in mice, I performed microarray analysis of RNA extracted from embryonic mouse heads at 10.5 days post coitum (dpc) and compared RNA from wild-type, loss-of-function (Sox3-null) and gain-of-function (Extra-Sox3) mice. Several emergent candidate genes were further tested by quantitative mRNA expression analysis (qPCR). One of these was Neurogenin-3 (Ngn3), which showed a 2.5fold decrease (P<0.001) in expression by microarray in *Sox*3-null (n=6), compared with wild-type (WT; n=6) mice and 1.8-fold decrease (P<0.001) by qPCR between *Sox*3-null (n=6) and WT (n=6) mice. To evaluate the relationship between Ngn3 and Sox3 at a cellular level immunohistochemistry was performed on 10.5 dpc and 12.5 dpc brains. In WT mice at 10.5 dpc and 12.5 dpc Ngn3 and Sox3 expression overlapped in a subset of cells across the ventral-midline of the developing hypothalamus. In addition and in contrast to WT mice, in *Sox*3-null mice, there were few Ngn3 positive cells, localized to the arcuate hypothalamic nucleus. Neurogenin-3 (Ngn3) is a member of the Neurogenin gene family of proneural basic helix-loop-helix proteins. Although previous data show the importance of Ngn3 during pancreatic development, there is no information on the mechanisms and actions of Ngn3 or a relationship between NGN3 action and SOX3 during hypothalamic development. These results suggest Ngn3 is a downstream target of *Sox*3 that is contributing to appropriate development of the hypothalamic-pituitary axis.

The second study (project 2) aimed to characterize and further examine a novel recessive ENU mouse mutant, called Tukku¹, exhibiting HP axis dysfunction resulting in dwarfism, pituitary hypoplasia and GH deficiency. Adult Tukku mice are 30-40% smaller than their WT littermates. The primary focus was to characterize the dwarfism phenotype in relation to the somatotropic axis and to identify the causative gene. The mutation was identified as a leucine to proline substitution in tryptophanyl-tRNA synthetase (WARS), a member of the aminoacyl-tRNA synthetase (AARS) enzyme family that link amino acids to their specific tRNAs. For proper function of this enzyme the specific recognition of substrates is critical for the fidelity of protein synthesis. The Wars mutation is contained within the N-terminal WHEP domain, from residue 16-69, and likely causes the disruption of the alpha helical structure. The N-terminal WHEP domain has only been found in eukaryote Wars enzyme. Importantly, AARS have been linked to regulating the noncanonical activity of angiogenesis (Otani et al., 2002; Wakasugi, 2010; Wakasugi and Schimmel, 1999; Wakasugi et al., 2002b). Along with pituitary hypoplasia, Tukku mice show a significant reduction in pituitary GH and serum levels of IGF-1, suggesting the defect leading to pituitary hypoplasia involves brain regions implicated in growth of the anterior pituitary. The reduction in pituitary GH levels may also involve delivery of GHreleasing hormone (GHRH) to GH-secreting cells since preliminary data also indicate that WARS is expressed within blood vessels of the pituitary and hypothalamus. To assess this, quantitative mRNA expression analysis (qPCR) of GHRH and somatostatin (Sst) was

_

¹ Tukku, meaning 'small' in Kaurna Aboriginal language.

performed. qPCR revealed a decrease in both GHRH and Sst (fold change >2) indicating that the defect is likely to be within the hypothalamic hypophysial vasculature that extends and makes a connection with the pituitary. To evaluate the relationship between Wars and pituitary vasculature, immunohistochemistry was performed on pituitaries at 8-weeks postnatal. Pituitary sections were co-stained with antibodies against platelet endothelial cell adhesion molecule (PECAM) + Wars or vascular-endothelial cadherin (VE-Cadherin; an endothelial specific, transmembrane protein, which clusters at adheren junctions where it promotes homotypic cell-cell adhesion) + Wars. Wars immunostaining was expressed within the endothelial cells of the pituitary vasculature, both in the anterior and posterior pituitary. Both PECAM and Wars appeared co-expressed within the vascular wall. VE-Cadherin was expressed in vessels together with Wars.

Overall, the data gathered from these projects highlight important insights into the identification of *Ngn3* as a likely *Sox3* target gene (*project 1*) and have identified a novel dwarf mouse model with a genetic determinant of HP axis function (*project 2*). These results have application to the study of HP axis development, to the study of vascular development during embryology and postnatally, and to possible avenues of genetic screen testing and development of new treatments related to GH deficiencies.

STATEMENT OF CONTRIBUTION BY OTHERS TO THIS WORK

Ms Sandra Piltz contributed to routine technical assistance in the maintenance of mouse colonies as well as purification of genomic DNA and genotyping by PCR.

Mr Dale McAninch contributed to the validation of genes identified by microarray (Chapter 3. Identification of *Sox3* Target Genes, p.105). This work formed part of his honors thesis in 2008.

Dr Stuart Reed, Dr Chris Goodnow and the team from The Australian Phenomics Centre (Canberra, ACT, Australia) the dwarf mouse line generated and identified the mutated gene by sequencing used in Project 2 (Chapter 4. Novel Dwarf Mouse Generated by ENU Mutagenesis, p.151).

Ms Carlie Delaine, Ms Siti Hadzir and Dr Briony Forbes contributed to the analysis of pituitary growth hormone and serum IGF-1 levels (Figure 4-5, p.162).

Ms Nadia Gagliardi performed paraffin embedding of tissues, sectioning and histological staining of mouse ovaries and testis (Figure 4-17, p.188) and brains (using Cresyl violet stain; Figure 4-3, p.158).

Ms Chin Ng contributed to the analysis of the dwarf mouse line by generating murine Wars constructs for analysis of angiostatic activity in cell culture (Figure 4-14, p.181). Ms Chin Ng also performed western blotting of mouse brain, pituitary and kidney samples (Figure 4-12, p.178). This work formed part of her honors thesis in 2010 (Ng, 2010).

A/Prof Paul Thomas and Prof Jeffrey Schwartz provided critical reading and proofing of the thesis manuscript.

Eva Szarek was responsible for the remainder of the work.

DECLARATION OF ORIGINALITY

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary instituition to Eva Szarek and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis (as listed below*) resides with the copyright holder(s) of those works.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

* Szarek, E., Cheah, P. S., Schwartz, J., Thomas, P., 2010. Molecular genetics of the developing neuroendocrine hypothalamus. Mol Cell Endocrinol. 323, 115-23

Eva Szarek, B.Sc (Hons)

ACKNOWLEDGEMENTS

There are many people that I would like to thank for making my Ph.D possible and for offering support along the way.

My sincerest gratitude goes to my advsors A/Prof Paul Thomas and Prof Jeffrey Schwartz for their guidance, encouragement and support. It has been an honor to work under your guidance and supervision. I am grateful to A/Prof Paul Thomas for his patience, persistence and wealth of knowledge in molecular biology; I have learned an enormous amount. I am grateful to Prof Jeffrey Schwartz for introducing me to and sharing with me his wealth of knowledge in the wonderful world that is the pituitary. Working with both of you has been an amazing experience, at the bench and over a few drinks. I would like to thank you both for giving me an amazing opportunity to pursue my Ph.D in the fascinating field of brain development and neuroendocrinology. Thank you for the discussion of ideas, for giving me the freedom to follow my intuition, for offering invaluable advice.

I would also like to thank all the members of the Thomas lab (past and present), for discussions, useful tips at the lab bench, and for being a fun group to work with.

To all my friends outside the lab, thank you for being so supportive and getting my mind off the Ph.D every now and then. I really appreciated it!

A very special thank you to my loving parents and family for believing in me and taking an interest in my project. Thank you for your encouragement, and ongoing support. A special thank you to my wonderful husband for putting up with my late nights and weekends in front of the computer and my less than seldom grumpy mood. I love you and... I can finally say... "koniec!"

ACRONYMS AND ABBREVIATIONS

3'UTR	3' untranslated region	FCS	fetal calf serum
ACTH	Adrenocorticotropic hormone	FSC	forward scatter
ADH	antidiuretic hormone (same as AVP)	FSH	Follicle Stimulating Hormone
AGRF	Australian Genome Research Facility	G1	First Generation
AH	anterior hypothalamus	gDNA	genomic DNA
ARC	arcuate nucleus	GFP	green fluorescent protein
AVP	arginine vasopressin (same as ADH)	GFP+	GFP-positive
BAC	Bacterial Artificial Chromosome	GH	Growth hormone
ВСІР	5-Bromo-4-Chloro-3-Indolyl phosphate	GHRH	Growth-hormone-releasing hormone
BM	basement membrane	GHRH	R growth hormone-releasing hormone
ВМР	bone morphogenic protein	receptor	r
bр	base pair	h	hour
BSA	bovine serum albumin	H_2O	water
C-termi	inal carboxyterminal	HEPES ethansu	N-[2-hydroxyethyl]-piperazin-N'-[2- lfonic acid]
cAMP	cyclic adenosine mono phosphate	HISS	heat-inactivated horse serum
cDNA	complimentary deoxyribonucleic acid	HMG	high mobility group
СН	congenital hypopituitarism	HP	hypothalamo-pituitary
ChIP	chromatin immunoprecipitation	IGF	insulin-like growth factor
CNS	Central nervous system	IGHD	isolated growth hormone deficiency
CoIP	co-immunoprecipitation	ΙΡ	immunoprecipitation
DEPC	diethylpyrocarbonate	IPTG	isopropylthiogalactosid
DIG	digoxigenin	IRES	internal ribosome entry site
DMEN	1 Dubelcco's Modified Eagle Medium	kb	kilobase pair = 1000bp
DMN	dorsal-medial nucleus	kDa	Kilo Dalton
DMSO	dimethylsulfoxide	KO	Knockout
DNA	Deoxyribonucleic acid	LH	Luteinizing hormone
dpc	days post coitum	M	Molar
Ε	Embryonic day	m	mouse
E. coli	Escherichia coli		mitogen-activated protein kinase
ЕСМ	extracellular matrix	ME ME	median eminence
EDTA	ethylene diaminetetra acetic acid	min	minute
EGF	epidermal growth factor	ml	millilitre
eGFP	enhanced green fluorescent protein		millimolar
EGTA	ethylenglycolbis-(2-aminoethyl)-tetraacetic	mM	
acid			20 milliQ H20
ENU	N-ethyl-N-nitrosurea		messenger ribonucleic acid
<i>FACS</i>	fluorescence activated cell sorting	mRNA	messenger RNA

NBT 4-nitroblue tetrazolium chloride SSCSalt Sodium Citrate N-terminal aminoterminal Sst somatostatin ΤE ng nanograms Tris-EDTA NGN/Ngn neurogenin tg transgenic NGN3/Ngn3 neurogenin-3 TGF_B transforming growth factor-beta nМ TRHnanomolar Thyrotropin-releasing hormone ORF TRIS Open reading frame Tris-(hydroxymethyl)-aminomethan OToxytocin TrpRStryptophan-tRNA synthetase (see also WARS) Р postnatal day TSHThyroid-stimulating hormone PAGE polyacrylamide-gel electrophoresis U units PBSPhosphate buffered saline UTR untranslated region PCRPolymerase Chain Reaction VEGF vascular endothelial growth factor PDGF platelet-derived growth factor VMNVentro-medial nucleus; PFAparaformaldehyde WARS see also TrpRS PΙ propidium Iodide WTwild-type PKAprotein kinase A XН X-linked hypopituitarism PKCprotein kinase C zf zebrafish POApreoptic area; μg microgram POMC Pro-opiomelanocortin micromolar μM PVNparaventricular nucleus; aPCR quantitative real-time polymerase chain reaction qRT-PCR quantitative real-time polymerase chain reaction rat RE restriction emzyme RIN RNA integrity number RNAribonucleic acid revolutions per minute rpm rRNAribosomal RNA RTreverse transcription rt room temperature RT-PCR reverse transcriptase-polymerase chain reaction SCNsupra-chiasmatic nucleus; SDS sodium dodecyl sulfate SHHsonic hedgehog SOCM Sox consensus motif

SON

SOX

supra-optic nucleus;

Sry-related HMG box containing

PUBLICATIONS

First author publications arising from the work presented within this thesis. A copy of this publication can be found in the Publications section of this thesis.

Szarek, E., Cheah, P. S., Schwartz, J., Thomas, P., 2010. Molecular genetics of the developing neuroendocrine hypothalamus. Mol Cell Endocrinol. 323, 115-23.

CONFERENCE PRECEEDINGS

The results described in this thesis have been presented as seminar communications at the following conferences:

Szarek, E., Read, S., Forbes, B., Delaine, C., Schwartz, J., Thomas, P. A novel ENU mutation, WARS, causes dwarfism in mice. *Gold Coast Health and Medical Research Conference*, Gold Coast, Queensland, <u>Australia</u>. December 2nd-3rd 2010

Szarek, E., Read, S., Forbes, B., Delaine, C., Schwartz, J., Thomas, P. A Novel ENU mutation, WARS, causes dwarfism in mice. *Program in Developmental Endocrinology and Genetics (PDEGEN) Research Conference*, National Institutes of Health, Bethesda, MD, USA. July 9th 2010.

Szarek, E., Read, S., Forbes, B., Schwartz, J., Thomas, P. Identification of the sequence responsible for and further phenotypic characterization of a novel dwarf mouse produced by ENU-induced mutagenesis. *Gold Coast Health and Medical Research Conference*, Gold Coast, Queensland, <u>Australia</u>. December 3rd – 4th 2009.

Szarek, E., Lovell-Badge, R., Schwartz, J., Thomas, P.Q. Expression of NGN3 in the developing hypothalamus: dependence on and co-localization with SOX3 in the mouse model of altered pituitary function. *ENDO2009*, Washington DC, <u>USA</u>. June 10th – 13th 2009.