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1 **Genetic and genomic tools to improve drought tolerance in wheat**

2 Delphine Fleury* and Peter Langridge

3 Australian Centre for Plant Functional Genomics ACPFG, University of Adelaide,
4 PMB1, Glen Osmond SA 5064, Australia

5 * To whom correspondence should be addressed: E-mail:
6 delphine.fleury@acpfg.com.au

7 **Abstract**

8 Tolerance to drought is a quantitative trait, with a complex phenotype, often
9 confounded by plant phenology. Breeding for drought tolerance is further
10 complicated since several types of abiotic stress, such as high temperatures, high
11 irradiance and nutrient toxicities or deficiencies can challenge crop plants
12 simultaneously. Although marker-assisted selection is now widely deployed in
13 wheat, it has not contributed significantly to cultivar improvement for adaptation to
14 low yielding environments and breeding has relied largely on direct phenotypic
15 selection for improved performance in these difficult environments. The limited
16 success of the physiological and molecular breeding approaches now suggests that a
17 careful rethink is needed of our strategies to better understand and breed for drought
18 tolerance. A research program for increasing drought tolerance of wheat should
19 tackle the problem in a multi-disciplinary approach, considering interaction between
20 multiple stresses and plant phenology, and integrating the physiological dissection of
21 drought tolerance traits and the genetic and genomics tools, such as quantitative trait
22 loci (QTL), microarrays and transgenic crops. In this paper, recent advances in the
23 genetics and genomics of drought tolerance in wheat and barley are reviewed and
24 used as a base for revisiting approaches to analyze drought tolerance in wheat. We

25 then describe a strategy where we target a specific environment and select
26 appropriate germplasm adapted to the chosen environment based on extensive
27 definition of the morpho-physiological and molecular mechanisms of tolerance of
28 the parents. This information was used to create structured populations and develop
29 models for QTL analysis and positional cloning.

30 **The complexity of drought tolerance**

31 Drought tolerance is defined as the ability of a plant to live, grow and reproduce
32 satisfactorily with limited water supply or under periodic conditions of water deficit
33 (Turner, 1979). Crop plants should not only have the ability to survive under drought
34 but also the ability to produce a harvestable yield. Research into the molecular
35 aspects of drought tolerance has tended to focus on plant survival at the expense of
36 yield. However, severe water deficits are rare in viable agriculture, and asking how
37 crops respond to or survive extreme drought is unlikely to have much of a practical
38 impact (Passioura, 2002). The aim is not to ‘convert wheat to a cactus’ but to allow
39 wheat to continue to grow and yield grain under water limited conditions.

40 Drought tolerance is a quantitative trait, with complex phenotype and genetic control
41 (McWilliam, 1989). Understanding the genetic basis of drought tolerance in crop
42 plants is a pre-requisite for developing superior genotypes through conventional
43 breeding. Given the complexity of the genetic control of drought tolerance
44 (multigenic, low-heritability and high G x E interactions), marker assisted selection
45 has not contributed significantly to cultivar improvement for dry environments and
46 breeding has relied on direct phenotypic selection. There are additional problems in
47 investigating the genomics of drought tolerance in species such as wheat: most
48 pathways and candidates can be more effectively studied in model species with

49 smaller and sequenced genomes such as *Arabidopsis* and even amongst the cereals
50 there is more extensive data available for rice and maize when compared to wheat.
51 However, recent technological advances and the imperative to ensure sustainable
52 food production has driven research programs to genetically improve this crop
53 despite the size and complexity of the genome (bread wheat is hexaploid with 16 Gb;
54 Feuillet *et al.*, 2008).

55 Breeding for drought tolerance is further complicated by the fact that several types of
56 abiotic stress can challenge crop plants simultaneously. High temperatures, high
57 irradiance, scarcity of water and nutrient deficiencies are commonly encountered
58 under normal growing conditions but may not be amenable to management through
59 traditional farm practices. Certain soil properties such as composition and structure
60 can also affect the balance of these different stresses (examples reviewed by
61 Whitmore and Whalley, 2009). Higher plants have evolved multiple, interconnected
62 strategies that enable them to survive unpredictable environmental fluctuations.
63 However, these strategies are not always well developed in the cereal cultivars
64 grown by farmers. At the molecular scale, pathways and gene networks between
65 abiotic stresses overlap; for example, about 40% of drought or high salinity inducible
66 genes are also induced by cold stress in rice (Shinozaki and Yamaguchi-Shinozaki,
67 2007). Some biochemical mechanisms may have opposing effects under different
68 stresses; therefore tackling tolerance to one stress may lead to sensitivity to another.
69 For example, some plants avoid heat stress by increasing stomatal conductance and
70 consequently evaporative cooling. However, closing the stomata helps to decrease
71 the loss of water and maintain turgor under conditions of low soil water potential.
72 The two mechanisms will conflict when high temperature and drought occur
73 simultaneously, which is frequently the case in a Mediterranean climate. Moreover,

74 the osmo-protectant amino acid proline has a toxic effect under heat stress and its
75 accumulation may not be an appropriate tolerance mechanism in field conditions
76 when heat and drought stresses are combined (Rizhsky *et al.*, 2004; Salekdeh *et al.*,
77 2009). Although the reductionist approach of studying isolated stress has
78 considerably increased our knowledge of tolerance mechanisms, interaction between
79 multiple stresses and stress combinations should be studied so as to make progress
80 relevant to field conditions.

81 In addition to these confounding environmental factors, a drought research program
82 should also consider plant phenology. By completing its life cycle before the onset
83 of severe water deficit, plants are often able to escape drought (Chaves *et al.*, 2003).
84 This mechanism of avoidance is deployed by rapid phenological development,
85 developmental plasticity and remobilization of pre-anthesis assimilates to grain
86 (Turner, 1979). A short life cycle is particularly advantageous in environments with
87 terminal drought stress or where physical or chemical barriers inhibit root growth
88 (Bidinger and Witcombe, 1989; Blum, 1988; Turner, 1986). The plant's response to
89 drought can be confounded by the environmental covariates as a result of differing
90 phenology. Plant maturity strongly influences grain yield under dry conditions (Jiang
91 and Zeng, 1995; Ouk *et al.*, 2006). A further confounding factor is plant
92 morphology, particularly plant height and tillering. Small plants with few tillers can
93 show higher Water Use Efficiency (WUE, ratio of the volume of water consumed to
94 the total biomass produced, or ratio of biomass to total evapotranspiration) than tall
95 multi-tillered plants. Since the genotypic variation of WUE is mainly driven by
96 variations in water use rather than by variations in plant assimilation, the selection
97 for high WUE may result in smaller plants, instead of high yield under drought
98 (Blum, 2005). Some QTLs for carbon isotope discrimination (a measurement of

99 WUE) in wheat were actually associated with variation in heading date and plant
100 height (Rebetzke *et al.*, 2008). Breeding for a shortened crop lifecycle has been a
101 very successful strategy in Mediterranean conditions (Araus *et al.*, 2002). However,
102 in well-developed agricultural regions, crop flowering time has already been
103 optimised by breeders so that the plant's phenology matches its environment
104 (Passioura, 2007). Therefore research should now focus on optimizing vegetative
105 development to manage biomass and ensure effective assimilates remobilization to
106 grain when water supply becomes limiting.

107 **QTLs of drought tolerance in wheat and barley**

108 Most QTLs for drought tolerance in wheat and its close relative barley have been
109 identified through yield and yield component measurements under water limited
110 conditions (Maccaferri *et al.*, 2008; Mathews *et al.*, 2008; McIntyre *et al.*, 2009;
111 Quarrie *et al.*, 2006; von Korff *et al.*, 2008). Although yield is the most relevant trait
112 to breeders, it is very difficult to accurately describe with respect to water use and to
113 identify candidate regions for positional cloning. Few studies have identified QTLs
114 associated with specific components of drought response (Table 3). Although the
115 development of gene-based molecular markers and genome sequencing should
116 accelerate positional cloning (reviewed in (Collins *et al.*, 2008), the genomic regions
117 associated to individual QTL are still very large and are usually unsuitable for
118 screening in a breeding program.

119 Despite their importance in drought tolerance, reproductive organs and the root have
120 attracted little attention in genetic studies. The effect of drought on reproductive
121 processes has been extensively described in cereals (see review by Barnabas *et al.*,
122 2008). Passioura (2007) suggested that floral infertility resulting from water deficit

123 could be a promising target for improvement but no QTL studies for this trait have
124 been published in wheat or barley. Improving the competence of the root systems to
125 extract water from the soil also seems an obvious target for genetic analysis. A
126 simulation analysis of root trait modification suggested that an extra 10 mm of water
127 extracted during the grain filling would increase yield by 500 kg.ha⁻¹, representing a
128 25% increase of the average wheat yield in Australia (~2000 kg.ha⁻¹) (Manschadi *et*
129 *al.*, 2006). The identification of markers or genes associated with root growth and
130 architecture would be particularly useful for breeding programs to improve root traits
131 by molecular marker-assisted selection. Few papers have described work on the
132 identification of QTLs for root traits in wheat. Ma *et al.*, (2005) found a QTL for
133 root-growth rate under Al treatment. QTLs of root traits (primary/lateral root length
134 and number, root dry matter) under control conditions and during nitrogen deficiency
135 were identified in wheat (Laperche *et al.*, 2006). Relative root growth was also used
136 by Jefferies *et al.*, (1999) to map QTL for tolerance to toxic levels of soil boron.
137 However, QTLs corresponding to root architecture in dry environments are yet to be
138 discovered in wheat and barley.

139 Over the past few years there have been several mapping studies that have targeted
140 drought tolerance and other abiotic stress tolerance loci associated with performance
141 in low yielding environments (Table 4). However, despite this substantial research
142 effort the only markers that have found their way into practical plant breeding
143 programs are those for boron and aluminium tolerance (Gupta *et al.*, 2010).

144 **‘Omics of drought tolerance**

145 The tools of genomics offer the means to produce comprehensive datasets on
146 changes in gene expression, protein profiles and metabolites that result from

147 exposure to drought. Comparison of gene expression in Arabidopsis and rice showed
148 that the two species share many common stress-inducible genes (Shinozaki and
149 Yamaguchi-Shinozaki, 2007). Abiotic stress tolerance involves similar transcription
150 factors in both dicotyledonous and monocotyledonous plants and some molecular
151 mechanisms of drought tolerance have been extensively described (reviewed by
152 Yamaguchi-Shinozaki and Shinozaki, 2006). It includes signal transduction cascade
153 and activation/regulation of transcription, functional protection of proteins by late-
154 embryogenesis abundant proteins (*eg* dehydrins) and chaperon proteins (*eg* heat
155 shock proteins), accumulation of osmolytes (proline, glycine betaine, trehalose,
156 mannitol, myo-inositol), induction of chemical antioxidants (ascorbic acid and
157 glutathione) and enzymes reducing the toxicity of reactive oxygen species
158 (superoxide dismutase, glutathione S-transferase). Homologous genes of these
159 different classes were also identified in transcriptomic experiments comparing wheat
160 lines grown under well watered and water stressed conditions (Aprile *et al.*, 2009;
161 Ergen and Budak, 2009; Ergen *et al.*, 2009; Mohammadi *et al.*, 2007, 2008a; Xue *et*
162 *al.*, 2006).

163 Despite the existence of common regulatory mechanisms across species, the
164 conservation of the molecular response to dehydration across experiments (Aprile *et*
165 *al.*, 2009; Mohammadi *et al.*, 2007) is low due to variation in stress dynamics, stage
166 of development and tissue analyzed. Interestingly, microarray assays revealed
167 unexpected results such as a decrease in the expression of glutathione-related genes
168 following withholding of water in a tolerant synthetic wheat line (Mohammadi *et al.*,
169 2007), or the accumulation of proline in drought sensitive emmer wheat line (Ergen
170 and Budak, 2009), suggesting that some pathways/mechanisms are dependent upon
171 genotype and the duration, intensity and type of stress applied.

172 Comparison of transcript profiles between tolerant and susceptible lines under water
173 stress has revealed differences in regulatory pathways. A drought tolerant emmer
174 wheat genotype showed induction in *bZIP* and *HD-ZIP* gene expression
175 (transcription factors known to be related to the ABA regulatory pathway) in
176 response to shock-like drought stress, whereas the sensitive genotype induced some
177 genes encoding transcription factors that bind to ethylene responsive elements. The
178 two genotypes also showed differences in expression of the *phospholipase C* gene,
179 involved in 1,4,5-triphosphate (IP3) signaling, and MAPK cascade elements (Ergen
180 *et al.*, 2009).

181 A direct link between a gene expression profiles and specific physiological
182 mechanisms has not been demonstrated, probably because the drought stress
183 scenario and the germplasm used for microarrays and physiological experiments are
184 seldom the same. Mohammadi *et al.* (2007) identified drought responsive genes in
185 the root of a synthetic hexaploid wheat line considered as a drought avoider. This
186 line retained a higher net photosynthetic rate and relative water content compared to
187 a sensitive line, Opata. However, these researchers could not establish a causal
188 relationship between the transcripts and the maintenance of relative water content. In
189 another experiment, the synthetic line showed higher cell membrane stability and a
190 lower rate of water loss from excised leaves, longer roots, larger root:shoot ratios
191 and a larger number of seminal roots than Opata (Mohammadi *et al.*, 2008a).

192 **A broad strategy for studying the genetics of drought tolerance in low yielding** 193 **environments**

194 Three major approaches for improving drought tolerance in wheat have been
195 employed for many years:

196 1. The empirical selection for yield under water limited condition. This has been
197 widely used and the good performance of modern cultivars is testimony to
198 the success of this approach. However, there are clear signs that the rates of
199 gain are declining and are insufficient to meet demand (Tester and Langridge,
200 2010).

201 2. Define physiological ideotypes for improved yield under water limited
202 conditions, identify sources of variation for these traits and introduce these
203 traits into elite varieties (Reynolds *et al.*, 2009; Richards *et al.*, 2010).
204 Although this approach has been followed for several decades it has met with
205 only limited success. The use of carbon isotope discrimination to screen for
206 WUE is probably the only trait to lead to new cultivars.

207 3. Marker assisted selection based around screening for desirable alleles at QTL
208 for drought tolerance. Despite many publications of QTL associated with
209 drought tolerance (Table 4) a recent survey of molecular markers being
210 deployed in wheat breeding programs failed to identify a single case where a
211 drought or drought related marker was being implemented (Gupta *et al.*,
212 2010). The survey found that almost 50 loci were currently being tracked
213 with molecular markers but the only loci associated with performance in low
214 yielding environments were for tolerance to high soil boron (*Bo1*),
215 aluminium toxicity (*Almt-1*), nematode resistance (cereal cyst and root lesion
216 nematodes) and plant height. This strongly implies that the previous drought
217 QTL studies have failed to identify loci of value to wheat breeding programs.

218 The limited success of the physiological and molecular breeding approaches until
219 now suggests a careful rethink is needed of our strategies to better understand and
220 breed for drought tolerance. Some of the new plant genomic techniques and

221 platforms may allow us to overcome the previous limitations but some may simply
222 lead us further down the tried and failed paths.

223 A research program for increasing drought tolerance of wheat should tackle the
224 problem in a multi-disciplinary approach, integrating the physiological dissection of
225 drought tolerance traits and the genetic and genomics tools, such as quantitative trait
226 loci (QTL), microarrays and transgenic crops. To do so, teams should include
227 molecular biologists, physiologists and breeders. This is the approach taken in major
228 companies which have successfully demonstrated application of patented
229 technologies for drought tolerance in the field (Passioura, 2007). In this paper, we
230 describe a strategy where we target a specific environment, select appropriate
231 germplasm adapted to the given environment to create structured populations,
232 extensively describe the morpho-physiological and molecular mechanisms of
233 tolerance of the parents and combine this knowledge in models for QTL analysis and
234 positional cloning (Figure 1). These various components of the drought genetics
235 strategy are discussed below.

236 **Target a realistic and specific environment**

237 Firstly, if the strategy is to have practical relevance it should look at drought
238 tolerance under field conditions. Molecular biologists have often reported the effect
239 of genes on drought tolerance in unrealistic environments (desiccation of detached
240 leaves or of seedlings transferred from hydroponics to air, osmotic shock by
241 applying polyethylene glycol) and rarely proved their phenotype in the field and their
242 expected value in breeding (Blum, 2005; Passioura, 2007).

243 Secondly, it is clear that there are different types of drought and that plants have
244 developed different morpho-physiological mechanisms to address them. For

245 example, low water use by moderated growth is an advantage in dry land conditions
246 of stored soil moisture (eastern Australia type) but might be detrimental in other
247 environments where drought may develop at any time during the season (Western
248 Australia type) (Blum, 2005). Plants have developed different root architecture to
249 optimize the timing of water extraction from soil depending of the drought
250 conditions. A compact and deep root system with a uniform root branching pattern
251 reduces water use early in the season but increases access to water during grain
252 filling and increases yield in conditions of stored soil moisture (Manschadi *et al.*,
253 2006). In the Mediterranean environment, with seasonal rainfall and terminal
254 drought, a large and shallow root system allows water extraction early in the season
255 in the top soil layers where the water is available (Manschadi *et al.*, 2006).

256 Due to strong G x E interactions, a QTL can have positive, null or negative additive
257 effects depending on the drought conditions (Collins *et al.*, 2008). Many QTLs
258 identified for response to drought are not “stable” in different environments. Yang *et*
259 *al.*, (2007) found that 7 of 10 significantly additive QTLs for stem water-soluble
260 carbohydrates content (compound which are stored for further grain filling in dry
261 conditions) in wheat interacted with the environment. Thus, a research program on
262 drought tolerance should first define a target drought scenario. Salekdeh *et al.* (2009)
263 proposed the creation of a Minimum Information About Drought Experiment
264 (MIADE) as a standard for the drought community, similar to the MIAME standard
265 for transcriptomics data. The environment of South Australia, which is the focus of
266 the strategy presented in this paper, is a Mediterranean-type of drought (Izanloo *et*
267 *al.*, 2008). Wheat production relies on rainfall during the growing season when
268 precipitation decreases with the rise of temperature during spring. The availability of
269 water is cyclic with a succession of precipitation and drought periods from anthesis

270 to grain-filling stages. The drought is combined with a cyclic heat stress between
271 rainfall events. Additional stress factors such as high wind, high irradiance, low air
272 humidity, hostile subsoil with salinity, boron toxicity and nutrient deficiency are
273 common in Southern Australia.

274 **Population structure for genetic study of drought tolerance**

275 The size and structure of the population used for genetic analysis must be carefully
276 assessed. Previous attempts by our group to use association mapping or
277 multiparental populations to study the genetics of drought were not regarded as
278 successful. These studies simply identified loci controlling phenology, largely
279 maturity, height and tillering, as major components of the drought response. The loci
280 identified were all previously known and had little practical relevance. As described
281 below, our approach was to use several large populations based exclusively on
282 parents adapted to our targeted environment but differing in drought responses.

283 Germplasm should be selected based on the likelihood that the lines will produce
284 valuable new genetic combination of direct and immediate relevance to breeding
285 programs developing cultivars for the target environment. Selection for drought
286 tolerance should not have a significant negative effect on other selection targets in a
287 breeding program, such as maturity, height, disease resistance and grain quality. The
288 use of elite varieties in the targeted environment has some benefits: the lines can be
289 used directly in breeding program. Moreover, alleles discovered in non-elite
290 germplasm might not lead to improvement because it was already selected during the
291 development of elite wheat cultivars (Collins et al., 2008).

292 Based around comprehensive field data generated during a severe drought in the
293 2001/2002 season, a genetics-based drought strategy was devised with Australian

294 wheat breeders to select parental lines for the development of segregating
295 populations. Elite cultivars and breeding lines were screened and assessed based on
296 their grain yield under severe water limitation in South Australian environment. A
297 trait summary for the parental lines described below is presented in Table 1. The two
298 lines Excalibur and RAC875 represent major sources of drought tolerance in the
299 Southern Australian environment. RAC875 was also believed to show superior
300 tolerance to heat stress during grain filling relative to other material in the breeding
301 programs. In the severe drought over the 2006 season where average yields at our
302 field sites were only 0.8 t/ha, Excalibur and RAC875 were consistently higher
303 yielding than other varieties (116% and 122% of the site means, respectively).
304 Excalibur and RAC875 show similar behaviour under drought and out-yielded the
305 variety Kukri, which was chosen as drought sensitive parent, by 10 to 40% under
306 severe water stress (data provided by Dr. Steve Jefferies). Fig. 2 represents the
307 percentage of yield production of four of the wheat cultivars on the basis of site
308 means with an average grain yield below 3.0 t.ha⁻¹ (the data is based on the 2009
309 National Varietal trials and did not include RAC875). The parental line Drysdale
310 was selected based on the carbon isotope discrimination screen, while Gladius is a
311 new variety released in 2007 and is based on Excalibur and RAC875
312 (<http://pbr.ipaustralia.plantbreeders.gov.au/>). Gladius has been the highest yielding
313 variety in the severe droughts of 2001/2002 and 2006/2007. Thus, this population
314 will allow us to investigate the significance of the C-isotope discrimination trait
315 relative to other factors and also allow confirmation of loci identified from the other
316 two populations. Importantly, Gladius also shows the heat tolerance seen in
317 RAC875. This is likely to be significant both in determining general tolerance to

318 drought stress but may also be of importance in lifting yield and grain quality in
319 irrigated environments.

320 For two populations, Excalibur x Kukri and RAC875 x Kukri, doubled haploid (DH)
321 populations (approximately 300 for each cross) and genetic maps have been
322 generated, and seeds have been multiplied in Australia, in India (DRW, Karnal), in
323 Mexico (CIMMYT) and Syria (ICARDA). Field data was collected from trials sown
324 in Australia and Mexico in 2006, 2007 and 2008. Both crosses have also been used
325 to generate large single seed descent populations (SSD or recombinant inbred lines,
326 RILs) of 3,000 lines, which have been taken to F5. For the population of Gladius x
327 Drysdale, 5,000 RILs have been produced, and a subset of 250 lines used for
328 preparation of a preliminary map. In total, over 10,000 lines have been developed for
329 the drought work. This represents a unique resource for studying the genetic control
330 of drought tolerance in wheat. The nature of the populations means that they are of
331 immediate relevance to wheat breeding programs targeted to low yielding
332 Mediterranean type environments and they are being used directly for selecting new
333 breeding lines.

334 The basic strategy for deploying these populations is to use the small doubled
335 haploid populations and 250 randomly selected SSD lines for the Gladius x Drysdale
336 for preliminary mapping of a wide range of drought related traits under controlled
337 glasshouse and field conditions. In addition, confounding factors such as boron and
338 salinity tolerance and nematode resistance and tolerance (both cereal cyst and root
339 lesion nematodes) have been assessed. Since the major loci controlling boron
340 tolerance and nematode resistance are known (Gupta *et al.*, 2010), these can be
341 readily accounted for but salt and nematode tolerance must be measured. The field

342 data have been generated at multiple sites in Australia and at droughted and irrigated
343 trials at the CIMMYT station at Obregon in Mexico.

344 The SSD populations are sufficiently large (3,000 for each cross) to permit
345 immediate transfer to positional cloning projects and detailed genetic dissection of
346 traits. They also provide an opportunity to explore epistatic interactions. As loci
347 influencing particular components of drought tolerance are identified these will be
348 used to divide the population for a second round of analysis. With populations of
349 3,000 lines we can explore the behaviour of five or six loci alone or in combination.
350 This will allow complex genetic interactions to be explored. In particular, this
351 approach may permit analysis of genome interactions. Subsets of lines can also be
352 selected to fix genes of major effect such as the flowering time genes *Vrn* or *Ppd* to
353 overcome the confounding effect of maturity on other traits (Distelfeld *et al.*, 2009).

354 **Extensive description of the drought tolerant and sensitive parental lines**

355 During the period of population development, detailed analyses of the parents can
356 help define the physiological, biochemical and molecular components of drought
357 response and the difference in behavior of parents. This information provides an
358 opportunity for determining the relative significance and reliability of different
359 phenotyping options in the segregating populations and also provides valuable
360 resources to support gene discovery work.

361 The growth of the three key parents, Kukri, Excalibur and RAC875 has been
362 analysed in detail in controlled conditions and under a cyclic drought regime that
363 reproduces the Southern Australian environment or Mediterranean type drought
364 stress (Izanloo *et al.*, 2008). Interestingly, Excalibur and RAC875 showed different
365 strategies of tolerance to the same drought scenario. In the drought treatment,

366 RAC875 produced fewer tillers, maintained a higher number of grains per tiller and
367 showed moderate osmotic adjustment (Table 2). The mechanism of tolerance of
368 RAC875 seemed more conservative: under both well-watered and dry conditions,
369 plants stored more water soluble carbohydrates in the stem, and leaves were more
370 waxed and thicker, showing constitutive aspects of tolerance to drought. The
371 genotype Excalibur showed higher osmotic adjustment, low ABA content, high
372 stomatal conductance and rapid recovery after stress. The plant morphology and
373 yield component reflected this responsive mechanism: leaves rolled in dry conditions
374 reducing radiation interception; a high number of tillers were produced and then
375 aborted in water-limiting conditions; the plants were able to compensate for the tiller
376 abortion by producing more grains per tiller after recovery. Excalibur showed a more
377 responsive mechanism with a strong interaction with environmental conditions.

378 Another component of the parental analysis is the generation of molecular and
379 biochemical data on the drought responses. ‘Omics profiles of the wheat parents
380 Excalibur, RAC875 and Kukri have been developed during a cyclic drought under
381 the same conditions as used for the morpho-physiological study by Izanloo *et al.*
382 (2008). We have used the *Triticum aestivum* 17K oligo microarray developed by the
383 Genome Canada/Genome Prairie/Genome Quebec program Functional Genomics of
384 Abiotic Stress (FGAS), the USDA-ARS-Genomics and Gene Discovery Research
385 Unit and the Australian Centre Plant Functional Genomics. The probe sequences of
386 the microchip were from cold-stressed cv Norstar crown and leaf, cold and salt
387 stressed root, unstressed controls, NSF-mapped ESTs, wheat zygotic and early
388 embryo and ESTs of genes involved in cell-wall metabolism. Two treatments (cyclic
389 drought and well-watered) of the three wheat lines, 5 sampling time-points (during
390 development of drought, before and after rewatering) with 5 biological replicates per

391 sampling time-point and 3 tissues per replicate (leaf, stem and spike) generated more
392 than 450 samples. Preliminary results of leaf samples showed a total of 6,537
393 differentially expressed genes. A database, DroughtComparator, is being developed
394 to support analysis and interpretation of the data. This database will be publicly
395 released on completion of the experiment. A cDNA series, for use in mRNA
396 quantification by Q-PCR, is now being used to validate the gene expression profiles.

397 Metabolite profiles of leaves and grains have also been generated during the same
398 experiment. The results showed significantly different responses to drought stress for
399 RAC875, Kukri and Excalibur. Interestingly, the control and water-stress data
400 formed distinct clusters for Excalibur, the drought responsive genotype. Other cell
401 components such as fructans, betaines, lipids, waxes are being measured. Detailed
402 comparison with the transcript data should help us to understand the biochemical
403 bases and regulation of the different tolerance mechanisms of Excalibur and
404 RAC875.

405 **Use of mathematical models for QTL analysis**

406 A growing concept in biology is the use of mathematical models to understand
407 complex trait such as yield during drought. The procedure consists of dissecting the
408 phenotype and the response to environment into elementary and simpler responses
409 (Reymond *et al.*, 2003; Tardieu, 2003). Such modeling has been successfully used to
410 study leaf growth in maize (Chenu *et al.*, 2009). Manschadi *et al.*, (2006) proposed
411 integration of a physiological understanding with plant breeding to develop
412 mechanistic crop models and design ideotypes to targeted environments.

413 The problem of non-stable QTL because of the differences of environmental
414 conditions between experiments (Reymond *et al.*, 2003) may be overcome by

415 measuring accurately the environmental variables and using ecophysiological
416 models. Combining QTL and ecophysiological models can help predict a phenotype
417 from combinations of alleles by analyzing QTL for each parameter of the model. For
418 example, the response of leaf growth to temperature and water deficit in maize has
419 been broken down into traits such as intrinsic elongation rate (Reymond *et al.*, 2003;
420 Tardieu, 2003). The QTL for each trait did not coincide with the same regions of the
421 genome suggesting that the traits were regulated by different genes. In this approach
422 the QTL correspond to well-defined functions and hypotheses about the function of
423 the genes underlying the QTL are likely to be more accurate and the number of
424 candidate genes narrower than in convention QTL mapping.

425 The gene networks are regulated and coordinated so that a plant react in a predictable
426 way to a given environmental condition (Tardieu, 2003). The ‘omics datasets serve
427 to help develop models of networks and pathways that are triggered in particular
428 genotypes in response to drought. The network information can feed directly into
429 building models of the relationship of specific pathways and processes to the
430 physiological responses to drought. The comparison of gene regulatory networks and
431 QTL for each parameter of the model could also help identify candidate genes for
432 QTL cloning. By studying all possible aspects from molecular aspects to plant
433 physiology in the same germplasm (RAC875, Excalibur, Kukri) and in a specific
434 drought scenario (terminal drought of South Australia), we should be able to dissect
435 the phenotype from gene to a plant mechanism and build the appropriate models.
436 The approach is empowered by the use of the same germplasm for creating the
437 genetic populations (DH and RIL) and allowing the analysis of different allelic
438 combinations.

439 **The QTL under fine-mapping at the ACPFG**

440 Genetic maps using DArT and SSR markers have been constructed for Excalibur x
441 Kukri, RAC875 x Kukri and Gladius x Drysdale populations. For each population,
442 between 600 and 1,000 markers have been included in the maps. The two Kukri
443 populations were grown at over ten sites in Southern Australian and at the CIMMYT
444 field site in Obregon, for the past three years. There are now data available for over
445 20 sites per year. In addition to standard assessment of lines during the growing
446 season and recording of detailed climatic data at each site, the soil at the field sites
447 was sampled extensively and assessed for nutrient levels and the presence of soil
448 pathogens and pests. The sites range from severe drought stress with average yields
449 at below 0.5 t.ha⁻¹ to sites where average yields were around 7 t.ha⁻¹. The Excalibur x
450 Kukri population has also been screened for tolerance to resistance to nematodes
451 (both cereal cyst and root lesion), boron toxicity, sodium exclusion and high pH
452 using established lab based hydroponic assays.

453 The first targets for positional cloning were common loci that appeared in both Kukri
454 populations. For the target loci around 6,000 RILs are available to support fine
455 mapping. Four loci have been identified for positional cloning: a locus on
456 chromosome 1B associated with yield under drought, a locus on 3B associated with
457 yield under heat stress and also canopy temperature suppression, a 6A locus that
458 shows a correlation between flag leaf width and grain size and a 7A locus that shows
459 increased spike length, higher grain number and increased yield, particularly under
460 severe drought stress (Izanloo et al., unpublished results). Loci of potential value to
461 the breeding programs will be confirmed using the larger RILs populations and
462 through analysis of selected lines from the Gladius x Drysdale population. These will
463 be made available to breeding programs as rapidly as possible.

464 **Moving forward to positional cloning of drought tolerance QTLs**

465 Fine mapping of a drought response may define a QTL to a region containing a large
466 number of genes. Genes that show a drought responsive expression pattern or may
467 encode an enzyme or other protein involved in a metabolic pathway that responds to
468 drought stress, would become strong candidates for further analysis. In maize, 22
469 differentially expressed genes were identified in a microarray experiment on four
470 susceptible and tolerant recombinant inbred lines, co-located on the genetic map with
471 QTLs for drought tolerance (Marino *et al.*, 2009). A physical map of genes involved
472 in drought tolerance has been attempted in wheat by Ramalingam *et al.* (2006) who
473 assigned 259 EST (811 loci) to chromosome deletion bins of wheat. However, the
474 analysis of transcript profiles in wheat is impaired by the absence of a genome
475 sequence and knowledge of homeologous sequences. The comparison of QTL and
476 microarray data is also difficult due to the low number of sequence-based markers in
477 the wheat genetic map. To overcome the problem, a SNP database is under

478 construction based on deep sequencing (over 1 million reads) of normalized full-
479 length cDNA of the Australian parental lines described above. Preliminary results
480 indicated that this work will generate around 30,000 SNPs per cross, which should
481 greatly facilitate the fine-mapping of the QTL and the use of grass synteny to select
482 candidate genes for cloning. It will also facilitate the genetic mapping of drought
483 responsive genes in Excalibur and RAC875 identified by the microarray and reverse
484 genetics programs.

485 Major changes in gene expression, protein or metabolite profiles can be directly
486 mapped onto the mapping populations. Coincidence of loci controlling gene
487 expression (eQTL), protein (pQTL) or metabolite (mQTL) with physiological or
488 yield related loci, indicates possible biochemical processes underlying the
489 physiological response. The co-localization of eQTL and physiological QTL for
490 diverse traits showed that it may facilitate the identification of candidate genes and
491 accelerate positional cloning (Kliebenstein, 2009). The gene expression profile of
492 lines in a mapping population using Affymetrix microarrays successfully identified
493 eQTL associated with seed development in wheat (Jordan *et al.*, 2007) and resistance
494 to leaf rust in barley (Chen *et al.*, 2010b). This method also allows the identification
495 of *trans* eQTL and provides valuable information on the regulatory network involved
496 in different tolerance mechanisms. However, the cost of microarray analysis for a
497 segregating population is prohibitive. To reduce the cost of such experiment, Xue *et*
498 *al.* (2006) compared the transcript profiles of selected progeny lines showing
499 difference in transpiration efficiency (carbon isotope discrimination). They identified
500 93 differentially expressed genes between high and low transpiration efficiency lines
501 of a wheat progeny Quarrion/Genaro. In a similar approach, recombinant lines of the
502 Excalibur/Kukri and RAC875/Kukri population will be selected based on the

503 outcomes of field screening and will also feed into the metabolomics and
504 transcription profiling projects.

505 **Conclusions**

506 Our knowledge of the mechanism of drought tolerance has been enhanced by
507 research programs targeting specific physiological, genetics or molecular aspect of
508 the drought response. However, in wheat these approaches have not lead to an
509 increase in tolerance over that already achieved by breeders using empirical
510 selection. Although the idea of linking physiology, 'omics and quantitative genetics
511 have been already proposed, only few research programs have taken this integrative
512 approach. The great strength of genetic and genomics analysis in wheat has been the
513 ability to generate large populations and well developed field phenotyping
514 capabilities. However, genetic studies have often been too ambitious and not
515 permitted effective dissection of the drought response or focused on specific drought
516 scenarios or regimes. Analysis of response to drought has been further complicated
517 by the absence of a genome sequence and the generally poor genomics resources has
518 been limiting. New developments in sequencing, marker development and genome
519 analysis have created the opportunity to revisit the way in which we structure
520 populations for analysis and tackle specific components of drought tolerance.
521 Phenotyping has now become the major cost and rate-limiting step in the genetic
522 analysis of drought tolerance and many other traits, and the development of rapid
523 and cheap procedures to characterize components of the drought response will be
524 critical in improving genetic resolution.

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530 of NSW.

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752

753 **Table 1.** Characteristics of wheat parents used for the drought mapping populations.

754 The traits listed are expected to affect field performance of these lines under water

755 limiting conditions.

Traits	RAC875	Kukri	Excalibur	Drysdale	Gladius
Drought tolerance	high	low	high	high	high
Heat tolerance	high	low	intermediate	-	high
Boron uptake	Low	Intermediate	Very high	Intermediate-High	Intermediate-High
Sodium uptake	Low-Intermediate	Intermediate	Low	High	Low-Intermediate
Zn efficiency	Low	Low	Intermediate-High	Low	Unknown
Nematodes					
Cereal cyst	Unknown	Susceptible	Susceptible	Susceptible	Unknown
Root lesion	Susceptible	Susceptible	Moderately resistant	Susceptible	Unknown
High pH	Intermediate	Intermediate	Intolerant	Unknown	Unknown

756

757 **Table 2.** Characteristics of the “drought tolerant” wheat genotypes RAC875 and
 758 Excalibur under a cyclic terminal drought regime.

RAC875	Excalibur
Lowest tiller number <i>per se</i>	High tiller number, more tiller abortion under stress
Thick green leaves	Higher total biomass
Stronger leaf waxiness	Higher root-shoot ratio under water stress
Stay-green phenotype	Highest osmotic adjustment
Moderate osmotic adjustment	Low ABA content under stress
Low stomatal conductance	Highest stomatal conductance
Slower recovery in stomatal aperture	Rapid recovery after re-watering
High water soluble carbohydrate	

759

760

761 **Table 3.** QTLs of physiological responses to drought stress identified in wheat and
 762 barley.

Trait	Species	Drought condition	Chromosome location	Reference
Water soluble carbohydrate	wheat	Rainfed field	1A, 1D, 2D, 4A, 6B, 7B, 7D	Yang <i>et al.</i> , 2007
Carbon isotope ratio, osmotic potential, chlorophyll content, flag leaf rolling index	durum wheat	Rainfed field	2B, 4A, 5A, 7B	Peleg <i>et al.</i> , 2009
Grain carbon isotope discrimination	barley	Mediterranean rain fed field	2H, 3H, 6H, 7H	Teulat <i>et al.</i> , 2002
Relative water content	barley	Mediterranean rain fed field	6HL	Teulat <i>et al.</i> , 2003
Leaf osmotic potential, osmotic potential at full turgor, osmotic adjustment, carbon isotope discrimination	barley	Water-deficit in growth chamber	6HL	Teulat <i>et al.</i> , 2001; Diab <i>et al.</i> , 2004
Water soluble carbohydrate	barley	Water-deficit in growth chamber	4H	Diab <i>et al.</i> , 2004
Chlorophyll and chlorophyll	barley	Post-flowering	2H, 4H, 6H, 7H	Guo <i>et al.</i> , 2008

fluorescence parameters

Relative water content	barley	Water- withholding	1H, 2H, 6H	Chen <i>et al.</i> , 2010a
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763

764 **Table 4.** QTLs identified for tolerance to several abiotic stresses in wheat and barley
 765 (update of the review by Langridge *et al.* (2006) which covered studies earlier than
 766 2006).

Stress	Chromosome location	Reference
wheat		
Drought	20 QTLs	Kirigwi <i>et al.</i> , 2007; Mathews <i>et al.</i> , 2008; Salem <i>et al.</i> , 2007
Cold	5A, 1D	Baga <i>et al.</i> , 2007
Copper toxicity	1AL, 2DS, 3DS, 4AL, 5AL, 5DL, 5BL and 7DS	Balint <i>et al.</i> , 2007; Balint <i>et al.</i> , 2009
Aluminium toxicity	4DL, 3BL, 2A, 5AS and 2DL	Cai <i>et al.</i> , 2008; Ma <i>et al.</i> , 2006
Salinity	47 QTLs	Ma <i>et al.</i> , 2007
Heat	1B, 5B and 7B	Mohammadi <i>et al.</i> , 2008b
Nitrogen deficiency	2D, 4B and 5A	Laperche <i>et al.</i> , 2008
barley		
Drought	38 QTLs	von Korff <i>et al.</i> , 2008
Salinity	30 QTLs	Witzel <i>et al.</i> , 2010; Xue <i>et al.</i> , 2009
Water-logging	20 QTLs	Li <i>et al.</i> , 2008
Aluminium toxicity	2H, 3H and 4H	Navakode <i>et al.</i> , 2009

767

768

769 **Figure legends**

770 **Fig. 1.** Schematic representation of the pathway from parental lines selection to gene
771 discovery. The diagram shows the role of physiological analysis, population
772 development and phenotyping and the various ‘omics technologies in supporting a
773 gene discovery path. Given the complexity and variability of the droughted
774 environment, the first task is defining the type of drought regime or regimes that are
775 being investigated. The second issue is selecting germplasm that will be suitable for
776 the target environments and is likely to reveal major loci associated with tolerance.
777 These lines are used to develop the segregating populations that form the base for the
778 genetic analysis. Mathematical models of physiological traits and omics of parental
779 and selected recombinant lines provide functional data to select candidate genes for
780 the QTL. A further component in the process of defining target regions and
781 candidates for drought tolerance in wheat is the use of the sequenced cereal
782 genomes. The strong conservation of gene order between the grasses means that the
783 rice and *Brachypodium* genomes provide a valuable resource in developing markers
784 for fine mapping in a target region and for identifying candidate genes for the QTL.

785

786 **Fig. 2.** Relative yield of mapping parents in low yielding environments in 2009. The
787 data are taken from the National Variety Trials
788 (<http://www.nvtonline.com.au/home.htm>). The yields of the four varieties are
789 expressed as a percentage of the site mean. RAC875 was not included in these trials.