Construction of a microsatellite based genetic linkage map of almond

By

Iraj Tavassolian

Thesis submitted in fulfilment of the requirements for the degree of

Doctor of Philosophy (PhD)

Discipline of Wine and Horticulture School of Agriculture, Food and Wine



Table of Contents

Table of Contents	i
List of Figures	iv
List of Tables	vi
Abstract	vii
Declaration	xi
Abbreviations	xii
Acknowledgments	
Chapter 1	
1.1 General introduction	
1.2 Botanical classification of almond	2
1.3 Economic importance and uses	
1.4 Genetic diversity and origin of almond	
1.5 Almond breeding	
1.6 Genetic markers	
1.6.1 Morphological markers	9
1.6.2 Biochemical markers	9
1.6.3 DNA markers	10
1.6.3.1 Hybridisation-based polymorphism	10
1.6.3.2 PCR-based polymorphisms	11
1.7 Application of the saturated linkage map in plant breeding	14
1.8 Morphological traits in Almond	16
1.9 Development of an Australian almond linkage map, prior to the	18
1.9.1 Introduction	
1.9.2 Mapping population	
1.9.2.1 Introduction	
1.9.2.2 Australian almond mapping population	
1.9.3 Molecular and morphological markers used to develop the first generation of	
1.9.4 Aims of study and expected outcome	
Chapter 2	
2.1 Introduction	
2.2 Material and Methods	
2.2.1 Survey of published and unpublished microsatellites	
2.2.2 Genomic DNA extractions	
2.2.2.1 Introduction	
2.2.2.2 Extraction method	
2.2.2.3 Modified Lamboy and Alpha (1998) protocol macrotechnique	
2.2.3 DNA quantification and PCR amplification	
2.2.4 Segregation analysis and polymorphism detection	
2.2.4.1 Fluorescent labelled microsatellite detection	
2.2.4.2 Agarose gel electrophoresis	
2.2.4.3 PAGE gel electrophoresis	45
2.3 Results	46

2.3.1 Microsatellites isolated from almond	
2.3.2 Microsatellites isolated from peach	
2.3.3 Microsatellites isolated from other <i>Prunus</i> species	48
2.4 Discussion	50
Chapter 3	55
3.1 Introduction	
3.1.1 Shell hardness	
3.1.2 Kernel size	
3.1.4 Kernel taste	
3.1.5 Percentage of doubles	
3.2 Materials and Methods	63
3.2.1 Plant material and pomological traits measurement	
3.2.1.1 Shell hardness and kernel weight assessment	
3.2.1.2 Kernel size and shape	
3.2.1.3 Testa characteristics	
3.2.1.4 Kernel taste and amygdalin quantification	
3.2.1.5 Percentage of doubles	66
3.2.2 Statistical analysis	66
3.3 Results	
3.3.1 Shell hardness	
3.3.2 In-shell weight	
3.3.3 Kernel weight	
3.3.4 Kernel length	
3.3.5 Kernel thickness	
3.3.6 Kernel width	
3.3.7 Kernel shape	
3.3.8 Testa colour and pubescence	
3.3.10 Double kernel	
3.4 Discussion.	76
3.4.1 Shell hardness	
3.4.2 In-shell weight.	
3.4.3 Kernel weight	
3.4.4 Kernel size and shape	
3.4.5 Testa colour and pubescence	
3.4.6 Kernel taste and amygdalin quantification	
3.4.7 Percentage of double kernels	
3.5 Conclusion	95
Chapter 4	98
4.1 Introduction	
4.1.1 Marker assisted selection	
4.1.2 Map-based cloning and physical mapping	
4.1.3 Comparative mapping	
4.1.4 Construction of genetic linkage maps	
4.1.4.1 Mapping population	
4.1.4.2 Population genotyping	
4.1.4.3 Linkage analysis	
4.1.5 The <i>Prunus</i> linkage maps	
4.1.5.1 The almond maps	
4.2 Materials and Methods	108
4.2.1 Mapping population and DNA extraction	108
4.2.2 Microsatellites marker analysis	
4.2.3 Verification of microsatellites as MAS for self-compatibility	109

4.2.4 Linkage analysis and map construction	109
4.3 Results	111
4.3.1 Construction of the second Australian linkage maps for almond	
4.3.1.1 NP linkage map	
4.3.1.2 LA linkage map	111
4.3.1.3 The integrated linkage map	112
4.3.2 Comparison among the new maps and previous Australian almond maps	112
4.3.3 Marker assisted selection test for <i>S</i> -alleles	
4.3.4 Comparative mapping with other <i>Prunus</i> maps	113
4.4 Discussion	114
Chapter 5	134
5.1 Polymorphism detected by microsatellites	136
5.1.1 Comparison of microsatellite detection systems	
5.1.2 Genotyping of mapping population by microsatellites	137
5.2 Pomological analysis of the mapping population	138
5.2.1 Phenotypic and inheritance studies of the shell hardness, kernel size and weight	
5. 2.2 Analysis of kernel flavour inheritance and amygdalin contents	140
5.2.3 Phenotypic variation of double kernels	
5.2.3.1 Benefit of double kernels in genetic studies	142
5.3 Almond saturated map	143
5.3.1 Comparative mapping of the integrated map with <i>Prunus</i> reference map	144
5.3.2 Marker assisted selection for <i>S</i> -alleles	
5.4 Future Prospect of the almond map	146
References	148
APPENDIX	179

List of Figures

Fig. 1.1 Almond world production during 2000-20064
Fig. 1.2 The world top producers of almond
Fig. 1.3 main almond production area in Australia
Fig. 1.4 Map of the world showing the origin for almond [<i>Prunus</i> dulcis (1)] and different relative Prunus species.
Fig. 1.5 The controlled cross hybridisation between NP and LA in Waite Campus within the bird cage and branches sealed in insect proof nets
Fig. 1.6 A linkage map of the diploid almond, developed from the markers present in the male parent 'Lauranne'
Fig. 1.7 A linkage map of the diploid almond, developed from the markers present in the female parent 'Nonpareil'
Fig. 1.8 Integrated linkage map of the diploid almond cross of LA × NP31
Fig. 2.1 Microsatellite polymorphism in some progeny amplified by marker EMPA004
Fig. 2.2 Electrophorogramme of fluorescent labelled alleles detected by loci BPPCT013
Fig. 3.1 Birds are very problematic, mostly in paper / soft-shell cultivars58
Fig. 3.2 Variation of testa colour in almond60
Fig. 3.3 Average of traits over three year's measurements (2005-2007). A. Percentage of double kernels, B. Shell hardness; C. In-shell weight and D. Kernel weight74
Fig. 3.4 The graphs depict the correlation between in-shell and kernel weight among the mapping population of cross 'Nonpareil' × 'Lauranne' in the average value of three years.
Fig. 3.5 The graph shows the high correlation between kernel width and length and low correlation between kernel thickness and the other kernel size traits in a segregating population.
Fig. 3.6 A. B. C Percentage of progeny traits over three years for A. Shell Hardness, B. Kernel Thickness and C. Kernel Shape

List of Tables

Table 1.1 Objectives of Almond Breeding Programs 7
Table 1.2 Polymorphic SSR markers used by Gregory, (2004)
Table 1.3 Polymorphic SSR markers screened by Mnejja (2004). 27
Table 2.1 Microsatellites: designation, number of screened markers and origin39
Table 2.2 Microsatellite name and fluorescine label.
Table 2.3 Summary of microsatellite marker analysis results 47
Table 3.1 Descriptor list for ten traits scored over the three years on the F1 progeny. 68
Table 3.2 Morphological characters measured on F ₁ hybrid population of cross between 'Nonpareil' × 'Lauranne', mean value for data collected for three years (2005-2007)
Table 3.3 Chi-square test (χ^2) for expected frequency 3:1 or 1:1 of kernel taste, colour, smoothness and double kernels in the population NP × LA79
Table 3.4 Broad sense heritability of traits in the population NP × LA for three years (2005-2007)
Table 4.1 Distribution of mapped markers and their coverage across linkage groups of 'Nonpareil' (NP), 'Lauranne' (LA) and Integrated (IN) maps116

Abstract

Almond (*Prunus dulcis*) is the most important nut crop in terms of world production. Due to its health benefit and high nutritional value the consumption and world supply of almond is increasing. To remain competitive in the world market, the Australian almond breeding program was established to produce cultivars with better adaptation to Australian conditions. As part of this program an almond mapping population consisting of 93 F_1 progeny derived from a cross between the American cultivar 'Nonpareil' (NP) and the European self-compatible cultivar 'Lauranne' (LA) was produced to construct the genetic linkage maps. The first almond linkage map developed prior to the commencement of this project failed to produce the eight linkage groups similar to the basic chromosome number of almond (x = 8) and many large gaps were also observed on the linkage groups. Therefore, more markers were needed to saturate the maps.

Microsatellite markers are considered one of the best choices for mapping studies. 195 microsatellite markers isolated from *Prunus* species were obtained from published papers or by personal communication. Polymorphism was revealed by three different methods, and in general, polyacrylamide gel electrophoresis (PAGE) compared to the fluorescent labelled marker detection using an automated DNA sequencer or agarose gel electrophoresis, showed the most efficient and cost effective method of genotyping. A subset of 54 markers which produced reliable and easily interpretable polymorphic bands was selected to screen the whole mapping population. Microsatellites originally isolated from almond species showed the highest rate of amplification and polymorphism followed by peach microsatellites and the least informative markers were isolated from cherry. It seems that the level of

transportability and usefulness of microsatellite markers is related to the genetic distance of the closely related species. Almond and peach belong to the same subgenus (Amygdalus) and other *Prunus* species are classified in Prunophora subgenus.

The nut, or kernel, is the commercial part of the almond tree, thus to improve the quality of fruit an understanding of environmental influence, heritability and correlation of traits is required. Pomological and quality characters such as: shell hardness, kernel size, shape, taste, pubescence, colour, and percentage of doubles were measured during three consecutive years (2005-2007) on the total mapping population, but data analysis (ANOVA) was performed only on trees that survived for all three years. Most of the traits showed high broad-sense heritability and kernel shape showed the highest heritability of $H^2 = 0.92$ suggesting high genetic control of this trait. Occasionally larger kernels than either parent were found in the progeny indicating potential for improvement of this trait even with smaller kernel size parent that encompass many desirable characters. High correlation was also found between the in-shell and kernel weight (r = 0.74), kernel length / kernel width (r = 0.67), kernel weight to kernel length (r = 0.78) and kernel width (r = 0.80). This correlation estimation pointed out in this study indicates that the improvement of one character may result the progress in another trait. Neither of the parents in the mapping population had bitter or obvious slightly bitter taste but slightly bitter kernels were observed among the progeny. Amygdalin was assumed to be responsible for bitter taste in almond; therefore we measured the amount of amygdalin in sweet and slightly bitter kernel progeny by HPLC. However, the results showed that amygdalin exists in sweet kernels as well. Although the average amount of amygdalin in slightly bitter kernels (20.34 mg kg⁻¹ FW) was higher than sweet kernels (3.67 mg kg⁻¹ FW), some

sweet kernels had higher amounts of amygdalin suggesting the impact of other components on slightly bitter kernel. The highest variability within the traits was observed in the percentage of double kernel, which showed the highest standard error. Strong environmental effects, particularly low temperature at pre-blossom time is speculated to produce much higher double kernels.

Three genetic linkage maps, one for each parent and an integrated map were constructed by the addition of 54 new microsatellite markers to the previous dataset. All the data was scored and coded according to the coding system necessary by JoinMap3 which was used for map construction. 131 markers including microsatellite, ISSR, RAPD, SCAR and S-allele markers were placed on the integrated map covering 590.7 cM with the average density of 4.5 cM/marker. The minimum number of six microsatellite markers was placed on linkage group 8 and the linkage group 1 which is the longest linkage group has 14 microsatellite markers. Comparative mapping study with other *Prunus* maps, especially with the highly saturated reference map showed complete synteny and minor changes in the order of four markers on linkage groups compared with Prunus reference map. The conservation of molecular marker order observed in this study supports the idea of looking at *Prunus* genome as a single genetic system and practical application of this similarity would be in crosstransportability of microsatellite markers from well developed linkage maps to the less studied species in *Prunus*. Ten microsatellite loci placed on our map have not been reported before and could be used to improve the density of other *Prunus* maps, especially the reference map.

This study contributed to the better understanding of the mode of inheritance and environmental effect on morphological traits and the effect of amygdalin on kernel taste. The most saturated microsatellite based almond linkage map developed

in this study can serve as a framework for future almond breeding program in Australia and benefit *Prunus* improvement programs internationally.

Declaration

This work contains no material which has been accepted for the award of any other degree or diploma in any university or tertiary institution and, to the best of my knowledge contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent for my thesis to be made available for photocopying and loan when deposited in the University library.

Iraj Tavassolian May, 2008

Abbreviations

A Amygdalin

AFLP Amplified fragment length polymorphism

ANOVA Analysis of variance

BAC Bacterial artificial chromosome

BC Before Christ

bp Base pair

BSA Bulked segregant analysis

 χ^2 Chi-squared

°C Degrees celsius

cM centi-Morgan

CsCl Cesium choloride

CTAB Cetyltrimethylammonium bromide

cv. Cultivar

CAPs Cleaved amplified polymorphic sequence

DArT Diversity arrays technology

DETC Diethylidithiocarbamic acid

DNA Deoxyribonucleic acid

dNTP Deoxynucleotide triphosphate

DPSTC Double pseudo-testcross

EDTA Ethylene diamine tetraacetic acid

EST Expressed sequence tag

FAM 6-carboxy-fluorescine

F₁ First familial generation

F₂ Second familial generation

FAM 6-carboxyfluorescein

FAO Food and agriculture organisation

FISH Fluorescence in-situ hybridisation

FLB formamide / bromophenol blue

FW Fresh weight

g gram

G Linkage group

GDR Genome database for Rosaceae

H Broad sense inheritability

HCl Hydrochloric acid

HEX Hexacholoro-6-flourescine

HEX Hexachloro-6-carboxy-fluorescein

HPLC High performance liquid chromatography

ISSR Inter simple sequence repeat

Kb Kilobase

LA Lauranne

LD Linkage disequilibrium

LOD Logarithm of odds ratio

LSD Least significant difference

M Molar

MAS Marker assisted selection

min minute

mg milligram

μL Microlitres

mL Millilitres

mM Millimolar

mm Millimetre

NH4Ac Ammonium acetate

NP Nonpareil

NSW New South Wales

PAGE Polyacrylamide gel electrophoresis

PCR Polymerase chain reaction

pH Power of hydrogen

% Percent

QTL Quantitative trait loci

Registered trademark

r coefficient of correlation

RAPD Randomly amplified polymorphic DNA

RFLP Restriction fragment length polymorphism

RHS Royal Horticultural Society

SCAR Sequence characterised amplified region

SD Standard deviation

SDS Sodium dodecylsulphate

SE Standard error

sec second

SNP Single nucleotide polymorphism

sp. Species

SSR Simple sequence repeats

TBE Tris borat- EDTA buffer

TE Tris-EDTA buffer

TEMED N,N,N',N'- tetramethylethylenediamone

 $T \times E$ 'Texas' × 'Earlygold'

U Enzyme unit

USDA United state department of agriculture

V Volts

var. Variety

v/v Volume per volume

w/v Weight per volume

YAC Yeast artificial chromosome

Acknowledgments

This thesis would not been written without the invaluable assistance provided by many people listed below. If I missed anyone I sincerely apologise. I am deeply grateful to Dr Chris Ford for his peer supervision, constructive comments and endless support and encouragement. I always remember him as a role model in my next career. I express my appreciation to Dr Michelle Wirthensohn for all her patience, expert advice and enthusiasm. She has really brilliant mind and was so helpful to me and my wife. I also extend my gratitude to Prof. Margaret Sedgley who was very inspiring and her constant interest gave me the determination about the project from the start.

I wish to specially thank Dr Shubiao Wu for his important assistance on scoring of the microsatellite markers and comments on a draft of the thesis manuscript. Many thanks also to Prof. Raffaele Testolin, Dr Pere Arús and Dr Werner Howad for providing me the sequence of some unpublished microsatellite markers. My thank goes to Dr Ian Riley for statistical assistance and helpful advice during study in Australia.

I am also grateful to many other friends: Esmail, Abbas and Nasser for joyful conversations and sharing their time with me. Many thanks to the colleagues Dr Tricia Franks, Dr Matthew Hayes, Vanessa Melino, Venetia Joscelyne and Crista Burbidge for lively and lovely atmosphere they provide during my lab work.

I gratefully acknowledge Ministry of Science, Research and Technology of Iran for financial support of this study. Finally I want to express my greatest thanks to my wife, Samaneh Poormohammad, and my parent for their patience, constant encouragement, support and trust that they placed in me throughout this study. Without their help I would never been able to do this research.