

Santa Clara University
Scholar Commons

Biology

College of Arts & Sciences

11-2014

Draft Genome Sequences of Antibiotic-Resistant Commensal *Escherichia coli*

Meghan Garrett

Jennifer Parker

Craig M. Stephens

Santa Clara University, cstephens@scu.edu

Follow this and additional works at: <http://scholarcommons.scu.edu/bio>



Part of the [Genomics Commons](#), and the [Microbiology Commons](#)

Recommended Citation

Garrett M, Parker J, Stephens CM. 2014. Draft genome sequences of antibiotic-resistant commensal *Escherichia coli*. *Genome Announc.* 2(6):e00873-14. doi:10.1128/genomeA.00873-14.

Copyright © 2014 Garrett et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](#).

This Article is brought to you for free and open access by the College of Arts & Sciences at Scholar Commons. It has been accepted for inclusion in Biology by an authorized administrator of Scholar Commons. For more information, please contact rscrogin@scu.edu.

Draft Genome Sequences of Antibiotic-Resistant Commensal *Escherichia coli*

Meghan Garrett, Jennifer Parker, Craig M. Stephens

Biology Department, Santa Clara University, Santa Clara, California, USA

Antimicrobial resistance is a significant public health issue. We report here the draft genome sequences of three drug-resistant strains of commensal *Escherichia coli* isolated from a single healthy college student. Each strain has a distinct genome, but two of the three contain an identical large plasmid with multiple resistance genes.

Received 19 August 2014 Accepted 23 October 2014 Published 4 December 2014

Citation Garrett M, Parker J, Stephens CM. 2014. Draft genome sequences of antibiotic-resistant commensal *Escherichia coli*. *Genome Announc.* 2(6):e00873-14. doi:10.1128/genomeA.00873-14.

Copyright © 2014 Garrett et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Craig M. Stephens, cstephens@scu.edu.

The antibiotic “resistome” within the normal human gut microbiome is thought to contribute to the spread of resistance genes to pathogens (1, 2). *Escherichia coli* readily moves between humans, animals, and the environment (3, 4), and its ability to propagate mobile genetic elements may make it a significant vector for the spread of resistance genes (5, 6). Here, we report the draft genome sequences for three *E. coli* strains isolated from an undergraduate student, as part of a microbiology course project. The strains were isolated from rectal swabs plated on MacConkey agar (Remel) with no antibiotic selection and then were screened for resistance to representatives of several classes of antibiotics using a disk diffusion assay (Hardy Diagnostics). *E. coli* strains CS02 and CS05, which were isolated months apart, are resistant to ampicillin, erythromycin, gentamicin, streptomycin, sulfamethoxazole, tetracycline, and trimethoprim. CS05 is additionally resistant to nalidixic acid. *E. coli* strain CS03, which was isolated from the same swab as CS02, is resistant to the β -lactams ampicillin, cephalothin, and the combination of amoxicillin and clavulanic acid.

Genomic DNA was isolated using the NucleoSpin tissue kit (Qiagen). Sequencing was performed on an Ion Torrent platform (Life Technologies) at the High-Throughput Genomics Center at the University of Washington. The average read length was ~140 bases. The read ends were trimmed in Geneious version 7.0.6 (Biomatters Ltd.) before *de novo* assembly by MIRA (version 4.0) (7), initially using 45% of the reads to facilitate rapid assembly.

Contigs <500 bp were not included in the submission. Functional annotation was performed by the NCBI Prokaryotic Genomes Automatic Annotation Pipeline. The assembly metrics are provided in Table 1.

Resistance genes were identified in each strain by ResFinder version 2.1 (8). Two of the strains (CS02 and CS05) contain contigs nearly identical to most of *Klebsiella pneumoniae* plasmid pKF3-140 (9). Sequences in common with pKF3-140 include a 23-kb gene cluster encoding resistance to streptomycin-spectinomycin (*aadA5*), gentamicin [*aac(3)-IInd*], tetracycline (*tetA*), erythromycin (*mphA*), sulfonamides (*folP* alleles *sul1* and *sul2*), and trimethoprim (*dhfrA17*). β -Lactam resistance attributable to *bla*_{TEM-1B} is chromosomally encoded. Non-plasmid-derived contigs from strains CS02 and CS05 share only 96 to 98% identity, and only strain CS05 is resistant to nalidixic acid, due to characteristic mutations in *gyrA*. Strains CS02 and CS05 were isolated at different times, and the genome sequences suggest that the plasmid responsible for multidrug resistance moved horizontally between distinct *E. coli* lineages. Further analysis of these strains will be presented in a future publication.

Nucleotide sequence accession numbers. This whole-genome shotgun (WGS) project has been deposited at DDBJ/EMBL/GenBank under the WGS accession numbers [JNOF000000000](https://www.ncbi.nlm.nih.gov/nuccore/JNOF000000000) (CS02), [JNOG000000000](https://www.ncbi.nlm.nih.gov/nuccore/JNOG000000000) (CS03), and [JNOI000000000](https://www.ncbi.nlm.nih.gov/nuccore/JNOI000000000) (CS05). The versions described here are the first versions.

TABLE 1 Accession numbers and assembly metrics for annotated *E. coli* draft whole-genome sequences

Strain	NCBI accession no.	No. of contigs >500 bp	N_{50}	No. of annotated genes	No. of predicted coding sequences
CS02	JNOF000000000	278	69,714	5,020	4,392
CS03	JNOG000000000	228	63,887	4,829	4,228
CS05	JNOI000000000	245	58,975	4,872	4,397

ACKNOWLEDGMENT

We acknowledge financial support for this project from Santa Clara University.

REFERENCES

1. Sommer MO, Dantas G, Church GM. 2009. Functional characterization of the antibiotic resistance reservoir in the human microflora. *Science* 325:1128–1131. <http://dx.doi.org/10.1126/science.1176950>.
2. Schjørring S, Krogfelt KA. 2011. Assessment of bacterial antibiotic resistance transfer in the gut. *Int. J. Microbiol.* 2011:312956. <http://dx.doi.org/10.1155/2011/312956>.
3. Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, Gill SR, Nelson KE, Relman DA. 2005. Diversity of the human intestinal microbial flora. *Science* 308:1635–1638. <http://dx.doi.org/10.1126/science.1110591>.
4. Radhouani H, Silva N, Poeta P, Torres C, Correia S, Igrejas G. 2014. Potential impact of antimicrobial resistance in wildlife, environment and human health. *Front Microbiol.* 5:23. <http://dx.doi.org/10.3389/fmicb.2014.00023>.
5. Wright GD. 2007. The antibiotic resistome: the nexus of chemical and genetic diversity. *Nat. Rev. Microbiol.* 5:175–186. <http://dx.doi.org/10.1038/nrmicro1614>.
6. Bailey JK, Pinyon JL, Anantham S, Hall RM. 2010. Commensal *Escherichia coli* of healthy humans: a reservoir for antibiotic-resistance determinants. *J. Med. Microbiol.* 59:1331–1339. <http://dx.doi.org/10.1099/jmm.0.022475-0>.
7. Chevreux B. 2005. MIRA: an automated genome and EST assembler. Ph.D. thesis. German Cancer Research Center, Heidelberg, Heidelberg, Germany.
8. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.* 67:2640–2644. <http://dx.doi.org/10.1093/jac/dks261>.
9. Bai J, Liu Q, Yang Y, Wang J, Yang Y, Li J, Li P, Li X, Xi Y, Ying J, Ren P, Yang L, Ni L, Wu J, Bao Q, Zhou T. 2013. Insights into the evolution of gene organization and multidrug resistance from *Klebsiella pneumoniae* plasmid pKF3-140. *Gene* 519:60–66. <http://dx.doi.org/10.1016/j.gene.2013.01.050>.