

Molecular and immunological characterisation of *Acanthocheilonema viteae* chitinase

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Berlin den 23.11.2005

Babila Julius Tachu

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Dedication

This work is dedicated to:

Elizabeth Abimwoe and the one born after she transisted

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Summary

Chitinases are enzymes that break down chitin, a homopolymer of N-acetylglucosamine (GlcNAc) to its monomers. They are important molecules in the life cycle of parasitic nematodes representing important drug and vaccine targets. In this thesis, three genomic chitinase sequences (I, II and III) were obtained by characterising nine clones from a genomic library of *Acanthocheilonema viteae*. Southern blots independently confirmed the existence of three chitinase genes in *A. viteae*. The organisation of all three genomic sequences is similar, with the greatest difference occurring in exons encoding the serine-threonine rich domain of chitinases: this domain is about ten times larger in sequence III compared to I, but is absent in sequence II. Sequence I and III had features of regularly transcribed genes: start ATG, followed by an open reading frame, stop codon and polyadenylation signal. Sequence II lacked the first exon with start ATG. Screening of cDNA libraries from adult female *A. viteae* worms and L3 (third-stage larvae), respectively, as well as reverse transcriptase PCR (RT-PCR) showed transcripts in uterine microfilariae, L3 and L4 (fourth-stage larvae) for gene I only.

The N-terminal fragment of *A. viteae* chitinase was cloned into an expression vector and expressed in *Escherichia coli*. About 80% of the expressed chitinase were found in inclusion bodies and were purified under denaturing conditions. The soluble fraction was about 20% and could be purified under native conditions. Chitinase purified from inclusion bodies was 13-fold less active compared to soluble chitinase.

Synthetic peptides (P1 and P2) were designed from the active site of *A. viteae* chitinase, and used in parallel with chitinase from inclusion bodies in vaccination experiments using the *A. viteae* / *Meriones unguiculatus* filariasis model. Vaccination with recombinant protein led to a 29 % significant reduction in adult worm burden in a single experiment. Vaccination with P1 and P2 led to an overall non significant reduction in adult worm burden in two independent experiments. In the P1 group, there was a consistent reduction (39%, $p>0.05$ and 45%, $p<0.05$) in mf load that attained significance only in the second experiment. In the P2 group, there was no reduction in mf burden in the first experiment, but a significant reduction (75%, $p<0.05$) in the second.

These results suggest that filarial chitinases are potential targets for transmission blocking drugs and vaccines.

Zusammenfassung

Chitinasen sind Enzyme, die Chitin, ein Homopolymer des N-Acetylglucosamin (GlcNAc) in seine Monomere spalten. Diese sind wichtige Moleküle im Lebenszyklus der parasitischen Fadenwürmer, welche bedeutende Medikamenten- und Impfstoffziele darstellen. In der vorliegenden Arbeit wurden durch die Charakterisierung von 9 Klonen aus einer Genbank von *Acanthocheilonema viteae* drei genomische Chitinasesequenzen (I, II und III) gefunden. Diese Anzahl an Chitinasegenen wurde durch Southern-Blots bestätigt. Der Aufbau aller drei Sequenzen ist sehr ähnlich. Die größten Unterschiede sind in den Exons zu finden, welche die Serin-Threonin-reiche Domäne der Chitinasen codieren. Diese Domäne ist in Sequenz III ca. 10fach länger als in Sequenz I. In Sequenz II ist sie nicht vorhanden. Sequenz I und III hatten Eigenschaften eines regulär transkribierten Genes: eine Startcodon, gefolgt von einem offener Leserahmen, einem Stopcodon und einem Polyadenylierungssignal. Sequenz II fehlt das erste Exon mit dem Startcodon. Bei Durchmusterung einer cDNA-Bibliothek adulter *A. viteae* Würmer bzw. L3 Stadien, sowie durch Reverser Transkriptase-PCR (RT-PCR) wurden in den Mikrofilarien, L3 und L4 Transkripte für Gen I gefunden, jedoch nicht für Gen III.

Das N-terminale Fragment der *A. viteae*-Chitinase wurde in ein Expressionsplasmid ligiert und in *E. coli* exprimiert. Ungefähr 80 % der exprimierten Chitinase lagen in Einschluss-Körpern vor. Dieser Anteil konnte unter denaturierenden Bedingungen aufgereinigt werden. Der lösliche Anteil konnte unter nativen Bedingungen aufgereinigt werden. Die aus den Einschluss-Körpern aufgereinigte Chitinase zeigte im Vergleich zur löslichen Chitinase eine 13fach verminderte Aktivität.

Vom aktiven Zentrum der Chitinase wurden Peptide synthetisiert. Diese wurden parallel mit Chitinase aus den Einschluss-Körpern für Vakzinierungsexperimente im *A. viteae* / *Meriones unguiculatus*-Filarien-Modell genutzt. Die Reduzierung der Wurmlast um 29 % nach einer Immunisierung mit rekombinantem Protein zeigte eine tendenziell schützende Kapazität des verwendeten Proteins. In zwei unabhängigen Experimenten konnte nach Immunisierung mit zwei synthetischen Peptiden (P1 und P2) eine bedeutende Reduktion der Wurmlast beobachtet werden. In der P1-Gruppe gab es eine gleichmäßige Reduktion der Mf-Last, die nur im zweiten Experiment signifikant war (39 %, $p > 0,05$ und 45 %, $p < 0,05$). In der P2-Gruppe konnte in einem von zwei Experimenten eine signifikante Reduktion der Mf-Last erzielt werden (75 %, $p < 0,05$).

Diese Ergebnisse lassen vermuten, daß Filarien-Chitinasen potenzielle Ziele für Medikamente und Impfstoffe sind.

1 Introduction

Infection with filarial parasites affect about 200 million people worldwide (Saint André et al., 2002). *Onchocerca volvulus*, the cause of onchocerciasis, is a major health problem in 36 endemic countries of Africa, Latin America and the Arabian Peninsula; it is responsible for one million visually impaired subjects, 270,000 bilaterally blind cases and 6.5 million subjects suffering from severe itching or dermatitis. Globally, 18 million people are infected with the parasite, while some 120 million others are at risk of infection (<http://www.who.int/int-fs/en/fact095.html>). Lymphatic filariasis, caused by the parasites *Brugia malayi* and *Wuchereria bancrofti*, affects about 120 million people worldwide, of which 40 million are incapacitated and disfigured. It is a major health problem in India, Africa, South Asia and the Pacific (http://www.who.int/mediacentre/fact_sheets/fs102/en/).

Chemotherapeutic treatment and vector control are two strategies used to fight filariasis, but none of these strategies can have a long-lasting impact in endemic areas (Hougard et al., 1997; Cook et al., 2001, Richards et al., 2001). Vaccination is a promising additional control measure, and various strategies were used to identify and validate preventive vaccine candidates for filariasis (Abraham et al., 2002).

The search for chemotherapeutically and immunologically relevant target molecules led among others to a group of enzymatically active molecules, the chitinases.

1.1 Structure and function of chitinases

Chitinases (classified as EC 3.2.1.14 by the International Union of Biochemistry and Molecular Biology) are enzymes that break down chitin, a homopolymer of N-acetylglucosamine (GlcNAc) to its monomers. They are ubiquitous in the plant and animal kingdom (Flach et al., 1992) and play important structural, physiological, metabolic and defensive roles (Cohen-Kupiec et al., 1998; Felse et al., 1999). Chitinases are glycosyl hydrolases, a family that is subdivided according to their hydrolysis mechanisms and amino acid sequence similarities of catalytic domains (Henrissat, 1999; Henrissat and Bairoch, 1993, 1996) (<http://afmb.cnrs-mrs.fr/CAZY>). Chitinases belong to class 18 and 19 of this grouping. Family 18 chitinases are endochitinases that cleave chitin by a retaining mechanism through which the beta-linked polymer is cleaved to release beta anomer products. Family 19 chitinases are mostly found in plants, some bacteria and nematodes and cleave chitin by an inversion or retention mechanism (Robertus and Monzingo, 1999; Fukamizo, 2000;

Honda et al., 2004). Family 19 class IA/I and IB/II enzymes differ in the presence (IA/I) or absence (IB/II) of an N-terminal chitin-binding domain. Chitinases also have a carbohydrate-binding module that promotes adsorption of the enzyme to insoluble chitin. The chitin binding domain of filarial family 18 chitinases (EC 3.2.1.14) are in the carbohydrate-binding module family 14, and contain six conserved cysteins that probably form three disulfide bridges (Henrissat and Bairoch, 1993, 1996) (<http://afmb.cnrs-mrs.fr/CAZY/>).

Although the overall sequence similarity (average pair wise identity) between family 18 chitinases is only 21% (<http://www.sanger.ac.uk/Software/Pfam>), their active site regions contain many residues that are fully or highly conserved, and their structure is a TIM barrel fold as typified by chitinase-1 of *Coccidioides immitis* (Figure 1) (Hollis et al., 1998; 2000; Perakis et al., 1994; Terwischa van Scheltinga et al., 1994; van Aalten et al., 2001).

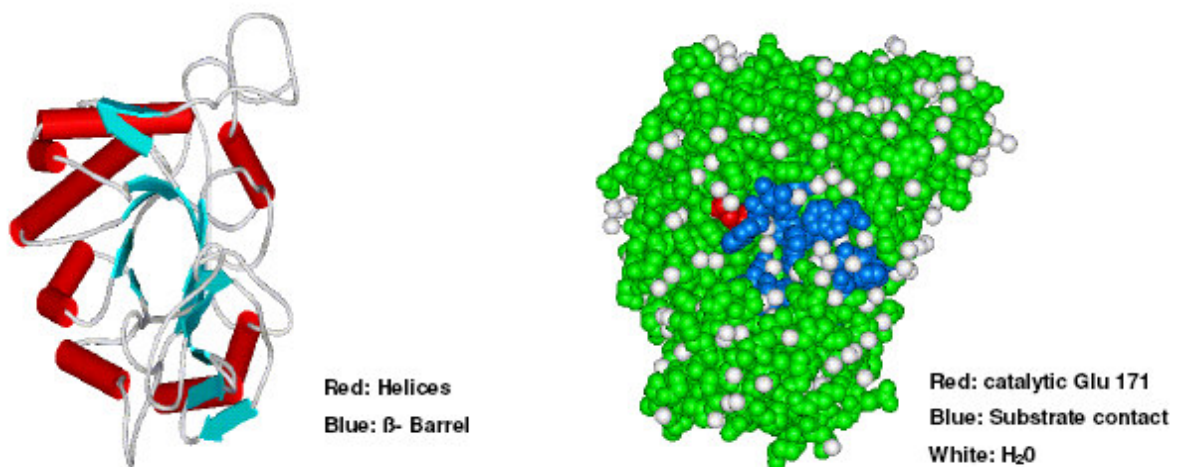


Fig. 1: Structure of chitinase-1 from *Coccidioides immitis*.

A TIM-barrel (β/α)₈ structure is shown on the left, and a model showing the catalytic Glu and positions of substrate contact is shown on the right. Diagrams were kindly provided by Prof. Dr. W. Höhne, Humboldt-University of Berlin)

In ChiB from *Serratia marcescens*, Tyr10, Ser93, Asp140, Asp142, Glu144, Tyr214, Asp215 and their corresponding subsets in other family 18 chitinases have been shown to be important for catalysis. The catalytically active residue is a glutamate in a stretch of highly conserved amino acids having the consensus DXDXE motif

(Watanabe et al., 1993; Bortone et al., 2002). The other residues important for catalysis have both mechanistic and chemical roles (Synstad et al., 2004). The carbohydrate binding clefts are present on the C-terminal side of the β strands in the (β/α)₈ barrel (Fusseti et al., 2003).

Chitinases have been described for the infective larval (L3) stage of *A. viteae* (Adam et al., 1996; Wu et al., 1996) and *O. volvulus* (Wu et al., 1996), as well as in the microfilarial stage of *B. malayi*, *B. pahangi* (Fuhrman et al., 1992) and in infective larvae (L3) and microfilariae of *W. bancrofti* (Raghavan et al., 1994). L3 chitinases of *A. viteae* and *O. volvulus* are accumulated in the glandular oesophagus of the worms in their insect vectors, and released when the larvae are transmitted to the hosts and eventually diminish following moulting from L3 to L4 (Wu et al., 1996). Microfilaria-specific chitinases (Fuhrman et al., 1992) could be recognised by a monoclonal antibody in extracts of microfilariae following several days of maturation in the vertebrate host as blood-borne microfilariae (Fuhrman et al., 1987). Interestingly, the appearance of these chitinases corresponds with the parasite's ability to infect the insect host (Fuhrman et al., 1992). To understand the possible role of these chitinases in the filariae, it is worth taking a brief look into the biology and biochemistry of the filariae relative to chitinase.

The filariae can be divided into two groups based on their mode of embryogenesis and eggshell remodelling, which arises from the two distinct patterns of intrauterine embryonic development (Rogers et al., 1976; Ellis et al., 1978). In filarial species like *D. immitis*, *O. volvulus* and *A. viteae*, the developing embryo hatches from its eggshell within the maternal uterus, and is subsequently shed as a microfilaria with no extracuticular structures or sheath. In *W. bancrofti*, *B. malayi*, and *B. pahangi*, the eggshell is remodelled as a sheath that eventually becomes part of the microfilaria. Another important difference is the route of migration of filarial microfilariae in their arthropod hosts. *B. malayi* and *B. pahangi* microfilariae penetrate the midgut of the mosquito vector and enter the haemocoel, while microfilariae of *D. immitis* enter the Malpighian tubules through the lumen at the posterior end of the gut.

The presence of chitin, the substrate of chitinases, has been biochemically documented in various nematode species and tissues. Chitin has been shown to be a component of the egg shells of many nematodes including the filariae (Brydon et

al., 1987). In addition, chitin has been demonstrated on the sheath of *B. malayi* microfilariae (Kaushal et al., 1984) and *W. bancrofti* microfilariae (Araujo et al., 1993).

In view of the biology and location of filarial chitinases, microfilaria chitinases of Brugian and Bancroftian filariae have been proposed to have a function in exsheathing the microfilaria when they are taken up in the arthropod vectors (Fuhrman et al., 1992). *A. viteae* and *O. volvulus* L3 chitinase are thought to have roles in ecdysis during post-infective development of the filariae, and in worm migration through the interstitial tissues of the host (Adam et al., 2001; Wu et al., 1996; 2001).

1.2 Filarial chitinases are potential targets for interventions

Filarial chitinases are potential targets for drug and immune interventions. Several observations implicate filarial chitinases to have roles in immune protection. In endemic areas, there are putatively immune (PIs) individuals (Elson et al., 1994; Turaga et al., 2000) with no current or past evidence of filarial infections. Interestingly, the sera of individuals putatively immune for *W. bancrofti* infection recognised a 43kD *B. malayi* L3 chitinase-like molecule, as opposed to the sera of microfilaraemic individuals (Freedman et al., 1989). A monoclonal antibody that mediated microfilaria clearance upon transfer recognised two *B. malayi* microfilaria-specific chitinases (Canlas et al., 1984a; Canlas et al., 1984b; Southworth et al., 1996). In addition, Vaccination of jirds with irradiated *A. viteae* L3 led to 90% protection (Lucius et al., 1991) and the sera of vaccinated rodents dominantly recognised *A. viteae* L3 chitinase in blots, implicating L3 chitinase as an immunodominant antigen playing an important role in the immune protection of the animals (Adam et al., 1996). These observations suggest that filarial chitinases may play a role in protective immunity.

Further more, immunisation of mice with *O. volvulus* chitinase DNA led to a significant reduction of 53% of worm burden following challenge infection (Harrison et al., 1999), experimentally confirming the relevance of filarial chitinases as vaccine molecules.

An alternative approach to immunisation with recombinant proteins would be the use of synthetic peptides designed from the active sites of filarial chitinases. Immunisation with such peptides could lead to the production of neutralising antibodies that will inhibit enzyme activity in the parasites (da Costa et al., 1999; Hirota et al., 2001; Fujii et al., 2004). A major short coming is the absence of a crystal

structure for filarial chitinase. In the absence of a crystal structure, homology modelling could be used to design peptides. The highest homology between filarial chitinases and a chitinase with known structure is between *A. viteae* chitinase and chitinase-1 of *C. immitis* (Höhne, personal communication). Using homology modelling, a structure of the active site of filarial chitinases could be obtained with *C. immitis* chitinase as template (Figure 2) (Höhne, personal communication).

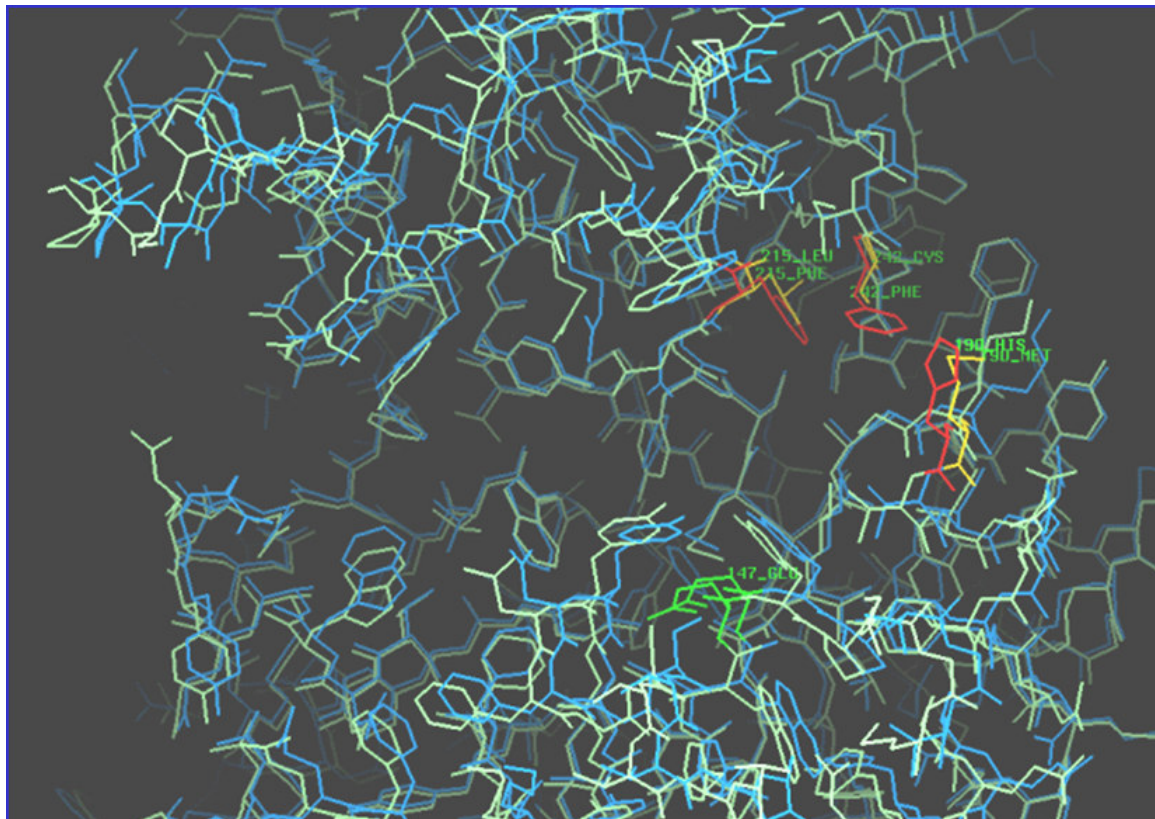


Fig. 2: Structural model of the active sites of *Acanthocheilonema viteae* (light green) and *Onchocerca volvulus* (blue) chitinases.

Non identical amino acids are shown in yellow for *A. viteae* and red for *O. volvulus* chitinase. The catalytic Glu is shown in green for both chitinases. Diagram was kindly provided by Prof. Dr. W. Höhne, Humboldt-University of Berlin.

No direct evidence exists to show that filarial chitinases are targets for drug intervention. However, chitinase molecules have been shown to be associated with parasite transmission and moulting (Wu et al., 1996; Fuhrman et al., 1987) and so will represent targets for transmission blocking drugs or vaccines. So far, the most effective inhibitor against family 18 chitinases is allosamidin (Sakuda et al., 1987). Due the cost of allosamidin, it is necessary to search for new and cheaper substitute

inhibitors. A prerequisite to the evaluation of potential inhibitors will be the production of soluble homogenous chitinase.

1.3 A. viteae as a model to investigate parasitological parameters of filarial diseases

Research in filariasis is dependent on the use of laboratory animal models owing to the inability to measure adult parasite population in humans. The filarial parasites are host-specific and so the first type of model system involves the use of parasites in surrogate models. The *Brugia spp.* / BALB/c mouse system has been used as a chemotherapeutic (Devaney et al., 1985) and immunological (Carlow and Philipp, 1987) model for the brugian filariasis. A similar approach with *Onchocerca spp.* involves the implantation of *Onchocerca spp.* in subcutaneous chambers in CBA/J or DBA/2J mice (Townson and Bianco, 1982; Abraham et al., 1992). These systems have a set back in that they rely on studies of a parasite in its non-natural host, a disadvantage that can be overcome by the use of full life-cycle models of the filaria *Brugia pahangi* in cats (Grenfell et al., 1991), bovine *Onchocerca spp.* or rodent filariae, like *Acanthocheilonema viteae* and *Litomosoides carinii*, in their natural hosts (reviewed in Abraham et al., 2002). Owing to the lack of adequate immunoreagents the underlying immunological mechanisms can however not be investigated in these model systems. The *B. pahangi* / cat model serves as a model for *Brugia* and *Wuchereria* in humans (Grenfell et al., 1991). This model system can be used to elucidate relationships between infection, immunity and disease states in lymphatic filariasis.

A. viteae in its natural host *Meriones unguiculatus* serves as a model for onchocerciasis, the disease caused by *O. volvulus*. This filarial model has some parallels to *O. volvulus* in that both reside in the subcutaneous tissue of their hosts and are therefore in the same immunological compartment. However, they do not form nodules and eye lesions like *O. volvulus*. Further more, both parasites share an array of antigenic similarities as demonstrated by anti-*O. volvulus* monoclonal antibodies (Nogami et al., 1986), and a high homology in corresponding molecules that have so far been characterised from both parasites. In addition, cross-protection between species has been shown in filariasis (Storey et al., 1982; Geiger et al., 1996) so that vaccine candidates established in one system could be tested in others. The *A. viteae* / *Meriones* system allows the study of resistance to challenge infection following immunisation (Abraham et al., 2002).

In the *A. viteae* / jird-model, it has been shown that immunisation with irradiated *A. viteae* L3 led to 90% protection against challenge infection, while immunisation with excretory-secretory products (ESP) led to 70% protection (Lucius et al., 1991). Parallel results were also obtained using irradiated L3s in other filariasis models (Lange et al., 1993; Taylor et al., 1994; Johnson et al., 1998; Trees et al., 2000) and it could be shown in this model that immunisation with irradiated L3 could also lead to resistance against homologous challenge infection (Abraham et al., 2002).

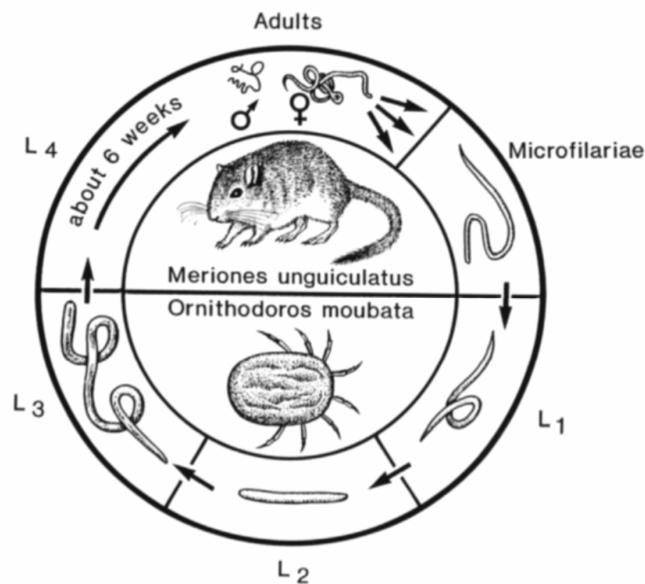


Fig. 3: Life cycle of *Acanthocheilonema viteae*

L1, L2, L3 and L4 are successive larval stages. Source: Archive of the Chair of Molecular Parasitology, Humboldt-University, Berlin.

1.4 Aims and objectives of the study

The aims of this study were three-fold. Firstly, *A. viteae* chitinase genes had to be isolated and characterised. To verify the hypothesis that there are several *A. viteae* chitinase genes, a cDNA library of female worms and L3, as well as cDNA from uterine microfilariae were screened for chitinase genes. Chitinase genes were investigated at the genomic level for copy number and gene organisation.

The second aim was the production of homogenous and enzymatically active *A. viteae* chitinase for use in inhibitor and structural studies.

The third aim was to evaluate the protective potentials of *A. viteae* chitinase and its synthetic peptides in immunisation studies. Since *A. viteae* chitinase activity has been demonstrated in two life stages of the filaria (microfilariae and infective larvae)

which could be targets for transmission blocking vaccines, another specific aim of this study was to design synthetic peptides from three-dimensional model of *A. viteae* chitinase for use in vaccination, in parallel with *A. viteae* N-terminal glycosyl hydrolase domain of chitinase.

2 Results

2.1 Characterisation of *A. viteae* chitinase genes

2.1.1 Determination of the gene structure of *A. viteae* chitinase genes

2.1.1.1 *Isolation and characterisation of A. viteae chitinase sequences from a genomic phage library*

A prerequisite to the determination of the complete genomic sequence of *A. viteae* chitinase was the isolation of clonal genomic recombinants containing the complete gene. To fulfil this requirement, an *A. viteae* genomic library in lambda dash II (Stratagene) was screened in plaque hybridisations using a DIG-labelled N-terminal cDNA sequence corresponding to nucleotides 52-1,117 of *A. viteae* L3 chitinase (Adam et al., 1996; Gene bank number U14638). The *A. viteae* genomic library (provided by Dr. Jörg Hirzmann, University of Gießen, Germany) was constructed in lambda dash II using gDNA from a mixed population of adult worms. In the first screening at high stringency, 4×10^4 plaque forming units (PFU) were investigated giving rise to 13 putatively positive recombinants. Following a second and tertiary screening round, 9 of the 13 plaques remained positive and were clonal.

2.1.1.2 *Restriction analysis of genomic inserts*

Inserts of recombinant phages were subcloned into the plasmid pBluescript for further analysis. The genomic inserts were digested out of the lambda arms, and subcloned directly into the Not I site of the plasmid, pBluescript. The advantage of such a strategy is the ease of preparing large amounts of genomic chitinase/recombinant plasmid vectors for further analysis.

Restriction digestion of the pBluescript recombinants with Not I released genomic inserts ranging in size from 10-16 kb, as well as the 3 kb pBluescript plasmid (Figure 4). For restriction analysis, recombinant DNA was digested with Not I in combination with EcoRI (Figure 5). Three main restriction patterns were observed. The first pattern was found in clones 1, 2, 11 and 13, for which digestion of full-length versions (clones 1 or 13) gave rise to 4.5, 3.5, 2, 1.8, and 1 kb bands. The second pattern was found in clones 3, 4, 9, for which digestion of the full-length versions (clones 4 and 9) gave rise to fragments corresponding to 5, 4.5, 1.2, 0.9, 0.8, 0.6, and 0.4 kb. The third pattern was found in clones 5 and 12, for which digestion of the full-length version (clone 12) gave rise to 5.5, 2.5, 1.7, 1.2, 0.8, 0.6, and 0.4 kb bands. A pair of

full-length clones from each insert group (clones 1/13, 4/9, and 5/12) was used for further sequencing and analysis.

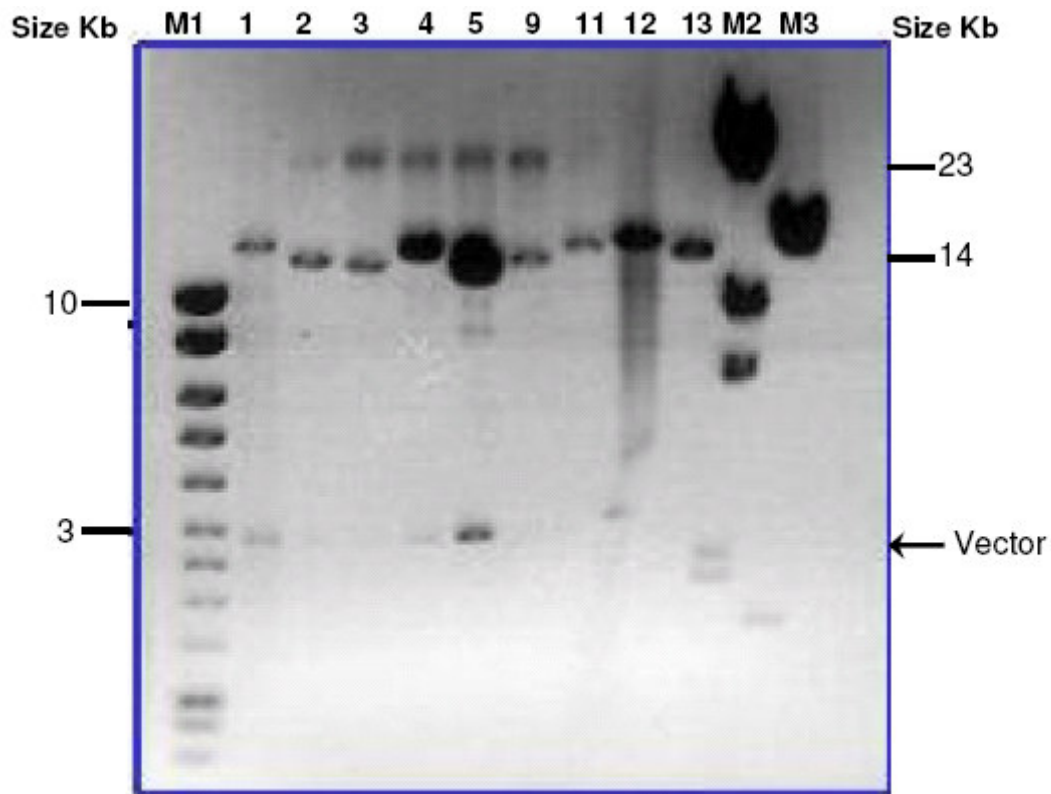


Fig. 4: Sizes of genomic inserts cloned into pBluescript

pBluescript recombinants were digested with Not I; Clone number 1-5, 9, 11-13: individual clones analysed, M1: low molecular size markers, M2: high molecular size markers, M3: 14 kb linearised plasmid.

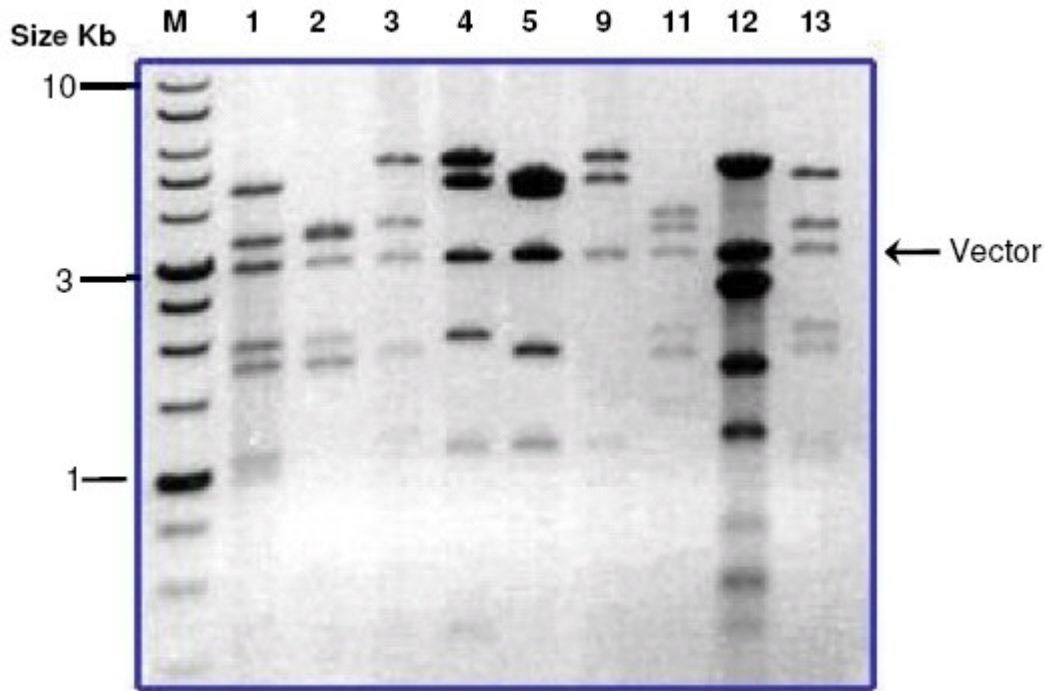


Fig. 5: Restriction analysis of *Acanthocheilonema viteae* genomic chitinase/pBluescript recombinant plasmids.

pBluescript recombinants were digested with Not I and EcoR I; Clone number 1-5, 9, 11-13: individual clones analysed, M1: low molecular size markers.

2.1.1.3 Sequencing of A. viteae genomic chitinase and determination of gene structure

Since it was technically impractical to directly sequence inserts in lambda vectors, overlapping PCR fragments were directionally amplified from recombinant lambda DNA clones using gene-specific and vector-specific primers (Figure 6) and sequenced. Alternatively, constructs of genomic inserts in pBluescript were also sequenced directly by primer walking. Three representative genomic inserts from clones 1, 9 and 12, identified following restriction mapping were used for sequencing. Independent confirmatory sequencing runs were also done with clones 4, 5, and 13 to verify data obtained for clones 9, 12 and 1, respectively. Taking clone 12 as an example, gene-specific primers were used to sequence portions of the insert corresponding to the chitinase gene, using overlapping long PCR fragments (Figure 6) or plasmids with the insert. These sequencing strategies were also applied for genomic clones 1, 4, 5, 9 and 13.

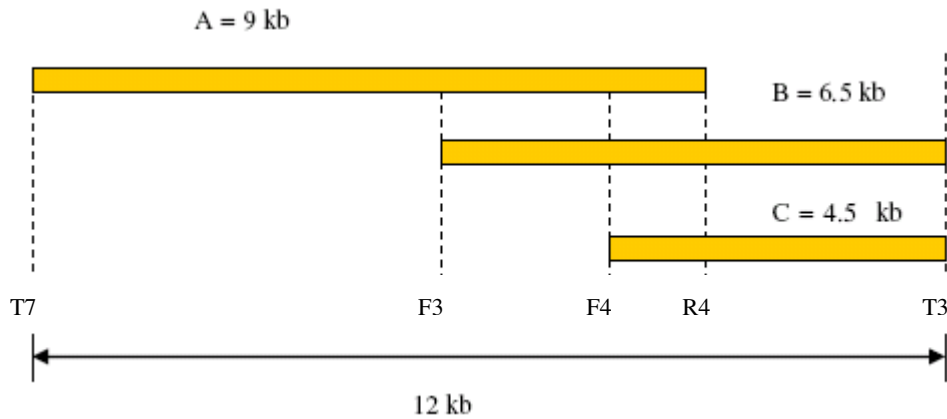


Fig. 6: Long-range PCR fragments used in sequencing of genomic insert 12.

Product A: amplified with primers T7-long range and R4 primer, product B: amplified with F3 and T3-long range primer, product C: amplified with F4 and T3-long primer.

Contiguous sequences were assembled from the nucleotide sequences obtained leading to the identification of three independent *A. viteae* chitinase genes (Figure 8). These sequences spanned about 14 kb for genomic clones 1/13, 14.5 kb for genomic clones 4/9, and 6 kb for genomic clones 5/12. The sequences were analysed and compared for the genomic organisation of the different chitinase genes using the published sequence for *A. viteae* L3 chitinase cDNA. A schematic diagram showing the extent to which the different insert groups were sequenced, and the genes predicted is shown in Figure 8. For further descriptive purposes, the chitinase genomic sequences have been named as gene sequence I, II, and III, respectively (Figure 8). Two sequences (I and III) have all features of a regularly transcribed gene: start methionine, followed by 12/13 exon sequences, and separated by 11/12 intron sequences respectively. Both sequences have a regular stop codon and a consensus polyadenylation signal. Generally, the splice donor and acceptor sites followed the common GT-AG rule (Senapathy et al., 1990), with the exception of introns e and n of gene II, and intron k of gene I that followed the GC-AG rule (Thanaraj and Clark, 2001) (Appendices 1,2 and 3) and offered for the possibility of alternative splicing.

To confirm the number of chitinase genes in the genome of *A. viteae*, Southern hybridisations were performed. A 700 bp dsDNA *A. viteae* chitinase probe was generated from *A. viteae* L3 chitinase cDNA, and hybridised under stringent

conditions to 2, 3 and 6 bands of *A. viteae* genomic DNA completely digested with Pvu II, Hind III and EcoR I, respectively (Figure 7).

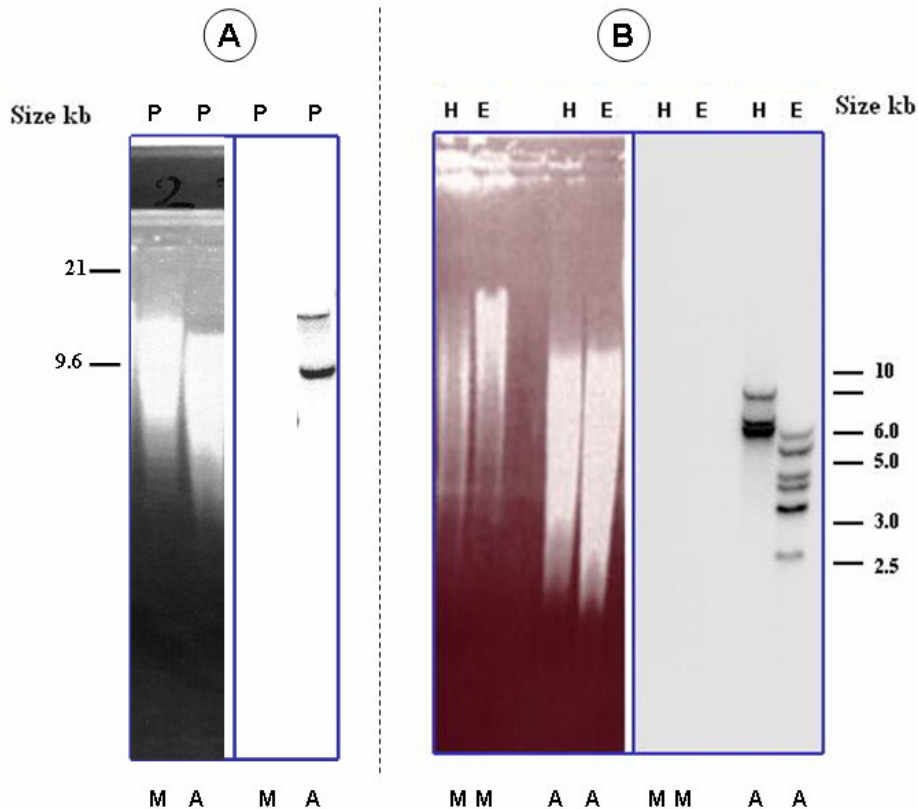


Fig. 7: Southern blot analysis of *Acanthocheilonema viteae* genomic DNA.

Meriones (M) and *A. viteae* (A) DNA (20 µg per lane) were respectively digested with Pvu II (P), EcoR I (E) and Hind III (H). Alternate results are shown for digestion of *Meriones* DNA as control, and *A. viteae* DNA on the left side of the picture, while hybridization results for both samples are on the right.

None of the restriction enzymes cut into the known cDNA sequence. In all three digested samples, the cDNA probe hybridised in different intensities to different bands. As completely digested DNA samples were used, the intensity of the bands seemed to be proportional to the number of probe molecules that bind to individual fragments. For the Pvu II digest that had two hybridisable bands, the band of approximately 15 Kb was recognised by less probe molecules than the fragment of 9.6 Kb, suggesting that there are more binding sites for the probe on the 9.6 Kb fragment. This would be consistent with a cluster of similar chitinase genes on the 9.6 restriction fragment. Further consideration of the Hind III hybridisation pattern suggests that there are at least three chitinase genes. Checking the corresponding

genomic sequences of chitinase gene I, II and III and adjacent regions up- and downstream, there are no Pvu II restriction sites, while there are two HindIII sites in each sequence that will each yield one hybridisable band within the size range observed. In addition, 6 hybridisable bands are expected for EcoRI, all falling within the range observed.

In summary, the Southern hybridisation results support the identification of three different chitinase genes in the *A. viteae* genome, for which two genes are clustered.

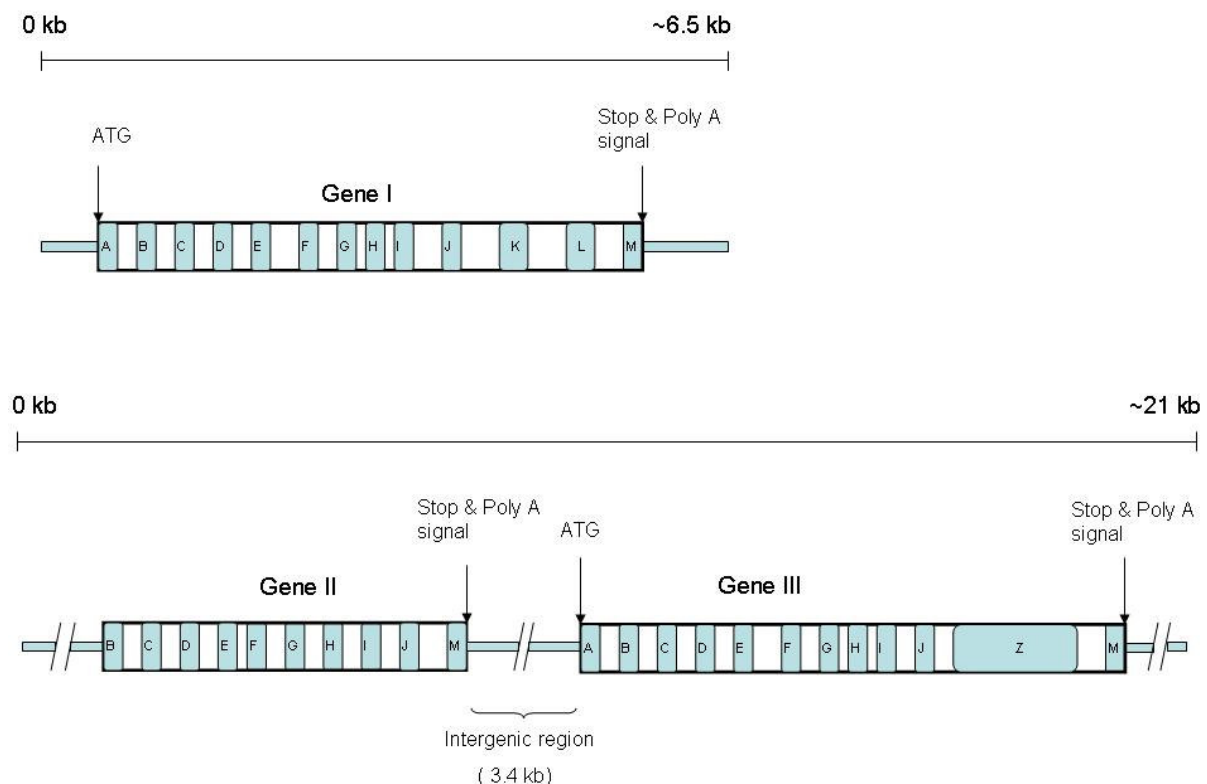


Fig. 8: Schematic diagram showing *Acanthocheilonema viteae* gene sequences.

Identified genes with lettered exons are shown in boxes, while intergenic regions are shown as grey lines. Gene sequences were obtained by sequencing different genomic clones as described in the text.

The 13 exons from gene sequence I have a 100% identity to the known cDNAs (Adam et al., 1996; Wu et al., 1996) as schematically represented in Figure 9. The signal sequence has two possible start methionines that have both been experimentally verified for gene I. The sequence can begin with the first start methionine (Wu et al., 1996) or with a second start methionine four amino acids

downstream (Adam et al., 1996). The Wu et al. sequence had additional 5' UTR of the cDNA which are found in the upstream region of the start ATG of gene I. This suggests that this sequence will be expressed in the worm. Individual exons build up different domains which represent different structural and functional parts of the molecule. Exon A contains the deduced N-terminal sequence necessary for the secretion of the protein. The sequence for a mature protein begins in exon B with the conserved YVRG sequence of filarial chitinases (Fuhrman et al., 1992). The active glutamate is found in exon E within the conserved amino acid sequence FDGFLLDWEYP (Synstad et al., 2004, Watanabe et al., 1993). The glycosyl hydrolase domain extends from exon B to exon J. This is followed by the serine-threonine region that has no direct relation to enzyme activity but has been demonstrated to facilitate enzyme secretion and to stabilize chitinase in the presence of proteolytic enzymes as shown in insect chitinases (Arakane et al., 2003). Finally exon M encodes a domain responsible for substrate binding (Fuhrman et al., 1992).

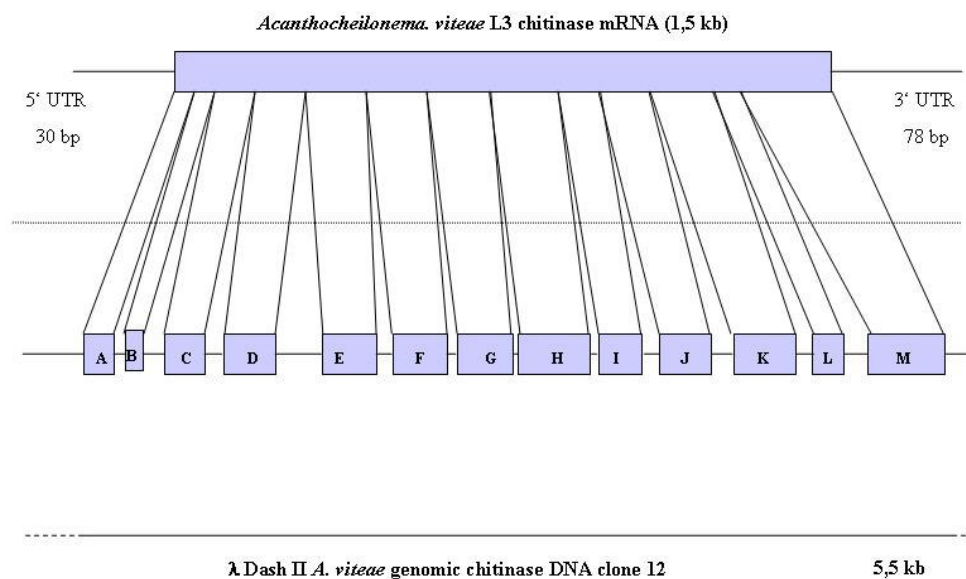


Fig. 9: Structure and organisation of *Acanthocheilonema viteae* L3 chitinase gene.

The upper panel shows the size of the cDNA, while the lower panel shows the positions and relative sizes of the 13 exons and 12 introns that make up the gene. Exons are shown as boxes, connected by introns shown as lines.

The exon / intron structure of gene sequence I and III are very similar. Therefore the exons and introns of gene I and gene III were compared as shown in Table 1. The amino acid sequences of the different exons show a high similarity of 77 to 97%.

Tab. 1: Comparison of exon/intron sizes and identities of *Acanthocheilonema viteae* genomic chitinase sequences I and III.

Exon	Size (aa)		Identities (%)		Intron	Size (bp)		Identities (%)
	Gene I	Gene III	NA	AM		Gene I	Gene III	
A	17	17	85	83	a	293	338	74
B	15	15	86	71	b	353	347	78
C	31	31	84	64	c	244	231	69
D	47	47	77	71	d	826	166	10
E	49	49	83	64	e	213	622	22
F	40	40	96	92	f	98	228	87
G	40	40	96	80	g	73	72	94
H	62	62	92	87	h	93	93	100
I	40	40	97	97	i	240	292	51
J	45	45	94	91	j	321	333	90
K	51	abs.	-	-	k	289	abs.	-
L	25	abs.	-	-	z	abs.	1276	-
Z	abs.	462	-	-	l	367	abs.	-
M	57	57	70	62				

Dashes represent identities not determined due to absence of comparable sequence in one of the genomic sequences. NA= nucleic acid; AM = amino acid; abs. = absent.

Overall, sequences I and III showed a relatively high homology. However, the greatest difference is in the region of exons K and L of gene sequence I. Exons K and L of gene I encode the serine-threonine rich region. This region is represented in gene III by one large exon named Z, which is also serine-threonine rich, but is about 6 times larger. The sequence of exon L can be aligned to the sequence of exon Z 19 times. Two different transcripts are therefore possible from both sequences: a 1.5 Kb transcript from gene I and a 2.7 Kb transcript for gene III giving a theoretical protein molecular weight of 55 kDa and 100 kDa, respectively.

Gene sequence II is apparently an incomplete gene without a regular start methionine, but having a stop codon and a consensus sequence for polyadenylation (Figure 8). An additional difference in gene II is the absence of the exons encoding for the serine-threonine rich region.

When gene II was compared with gene I and III, there was a higher amino acid identity between the exons of genes I and II (Table 2) relative to genes II and III

(Table 3). The identities of the intron sequences range from 10 to 100 % for comparable pairs.

Tab. 2: Comparison of exon/intron sizes and identities of *Acanthocheilonema viteae* genomic chitinase sequences I and II.

Exon	Size (aa)		Identities (%)		Intron	Size (bp)		Identities (%)
	Gene I	Gene II	NA	AM		Gene I	Gene II	
A	17	abs.	-	-	a	293	abs.	-
B	15	28	87	93	b	353	406	58
C	31	31	89	87	c	244	763	19
D	47	47	99	100	d	826	859	94
E	49	49	100	100	e	212	212	100
F	40	40	100	100	f	98	96	97
G	40	40	99	97.5	g	73	69	94
H	62	62	99	96	h	93	93	96
I	40	40	100	100	i	240	240	99
J	45	45	99	92	j	321	543	26
K	51	abs.	-	-	k	289	abs.	-
L	25	abs.	-	-	l	367	abs.	-
M	57	72	56	60				

Dashes represent identities not determined due to absence of comparable sequence in one of the genomic sequences. NA= nucleic acid; AM = amino acid; abs. = absent.

Tab. 3: Comparison of exon/intron sizes and identities of *Acanthocheilonema viteae* genomic chitinase sequences II and III.

Exon	Size (aa)		Identities (%)		Intron	Size (bp)		Identities (%)
	Gene II	Gene III	NA	AM		Gene II	Gene III	
A	17	abs.	-	-	a	abs.	338	-
B	28	15	70	62	b	406	347	62
C	31	31	80	76	c	763	231	18
D	47	47	78	55	d	859	166	10
E	49	49	95	74	e	212	622	22
F	40	40	90	87	f	96	228	88
G	40	40	96	90	g	69	72	98
H	62	62	96	95	h	93	93	96
I	40	40	97	97	i	240	292	51
J	45	45	98	80	z	abs.	1276	-
Z	abs.	462	-	-	j	543	333	28
M	72	57	53	45				

Dashes represent identities not determined due to absence of comparable sequence in one of the genomic sequences. NA= nucleic acid; AM = amino acid; abs. = absent.

In conclusion, the characterisation of nine genomic clones from an *A. viteae* phage library and Southern blot experiments revealed the existence of three different chitinase genes for which at least two are closely associated and might form a functional transcript. The major differences between the three genes are the serine-threonine rich region (exons K and L versus Z) almost at the 3' end of the genes.

2.2 Comparative analysis of *A. viteae* genomic chitinase sequences with genomic data bases of nematodes

At the moment, information is available from two nematode genomic data bases: the whole genomic information from the finished *C. elegans* project (*C. elegans* sequencing consortium, 1998) and the on-going sequencing project from the *Brugia malayi* genome (available at <http://www.tigr.org>). A comparison with the genomic sequence of chitinase gene I led to the identification of Tigr assembly 14274 that had the genomic sequence of *B. malayi* chitinase (Fuhrman et al., 1992). This sequence, like *A. viteae* chitinase, is organised into 13 exons and 12 introns, with similar chitinase motifs and domains being represented on the same exons.

Tab. 4: Comparison of exons/ intron sizes and identities between *Acanthocheilonema viteae* L3 (Gene I) and *Brugia malayi* MF 1 chitinases

Exon	Size (aa)		Identities (%)		Intron	Size (bp)		Identities (%)
	<i>A. viteae</i>	<i>B. malayi</i>	NA	AM		Av Chit	Bm Chit	
A	20	21	68	52	a	293	373	49
B	15	15	85	93	b	353	354	54
C	31	31	74	80	c	244	224	60
D	47	47	90	77	d	826	160	10
E	49	49	94	74	e	213	138	54
F	40	40	81	82	f	98	117	52
G	40	40	92	75	g	73	113	46
H	62	62	90	79	h	93	91	51
I	40	40	85	82	i	240	233	52
J	45	45	92	77	j	321	358	52
K	51	25	32	27	k	289	287	44
L	25	15	33	24	l	367	655	29
M	57	57	70	68				

In both genomic sequences, exon A encodes for the secretory N-terminal signal sequence, exons B-J for the glycosyl hydrolase domain, exons K and L for the serine/threonine rich linker and exon M encodes the chitin binding domain. The size of all exons, except exons K and L are essentially similar (Table 4). The distance between the ATG start codon and the stop codon is 4,566 bases as compared to 4989 bases for *A. viteae* chitinase. The introns of *B. malayi* chitinase range in size from 91 bp (intron 8) to 655 bp (intron 12) as compared to 73 bp (intron 7) to 826 bp (intron 4) for *A. viteae* L3 chitinase (Table 4). The identities between introns of both genes range from 10% to 60%. The identities of nucleotides in corresponding exons are remarkably high, except for exons K and L that have a bare 30% identity. This kind of variation was also noticed for the analysed sequences of *A. viteae* chitinase genes. This high level of homology between the chitinase genes of these related filarial species is also shown by the conservation of most of the intron phases, with the exception of intron k (Jean et al., 2001; Guiliano et al., 2002).

The situation in *A. viteae* chitinase gene sequence II in which there is no start methionine is comparable to a *Brugia malayi* genomic chitinase sequence in BAC

clone 1133599. In both sequences, there is no regular first exon A, and exon B has no regular 5' exon/intron boundary. While *A. viteae* chitinase gene II has the last exon, a regular stop codon and polyadenylation signal, the *B. malayi* sequence goes only as far as exon J. The remaining part of the sequence has homologies to nematode retroviral elements like *Ascaris lumbricoides* proretrovirus-like element (Felder et al., 1994).

In addition to these sequences, there are other large genomic assemblies with annotated genomic chitinase sequences in different genomic neighbourhoods. Tigr assembly 14932 has two chitinase genes in tandem as found on *A. viteae* genomic clones 1 and 9 (Gene II and III). The first gene (512) on this cluster has 8 exons and 7 introns and should encode only the glycosyl hydrolase domain, while the second gene (514) has 10 exons and 9 introns and should encode for all domains as found in *A. viteae* chitinase gene I.

In conclusion, *B. malayi* has two annotated chitinase genes, which have similar organisation and domain architecture like *A. viteae* chitinase gene I. This however does not exclude the existence of other chitinase genes owing to the presence of additional 14 genomic assemblies with exons having homologies to filarial chitinases.

A search in the data bank of the free-living nematode *Caenorhabditis elegans*, WormBase (<http://www.wormbase.org/>), revealed an alignment with the highest probability for the *C. elegans* sequence C04F6.3. The gene, unlike all three described *A. viteae* chitinase, has only four exons spanning 2.22 kb of genomic DNA (Figure 10). The potential cDNA encodes a protein with 617 amino acids and a molecular weight of 66.9 kDa. Like filarial chitinases, this chitinase contains one glycoside hydrolase (family 18) domain, and a chitin-binding domain. Comparison of the gene structure revealed a comparable domain for exon A responsible for the secretion of the functional protein. The glycosyl hydrolase domain containing the catalytic site of chitinase, represented in *A. viteae* by 9 different exons (B-J) is represented by exons 2 and 3 of the *C. elegans* chitinase gene. Thereby, exon 3 has an unusual size of 1313 nucleotides. Exon 3 encodes not only the glycosyl hydrolase domain, but contains also part of the serine-threonine linker region. Exon four of the *C. elegans* genomic chitinase sequence is clearly associated with the chitin binding domain as also represented by exon M of the *A. viteae* genomic sequences.

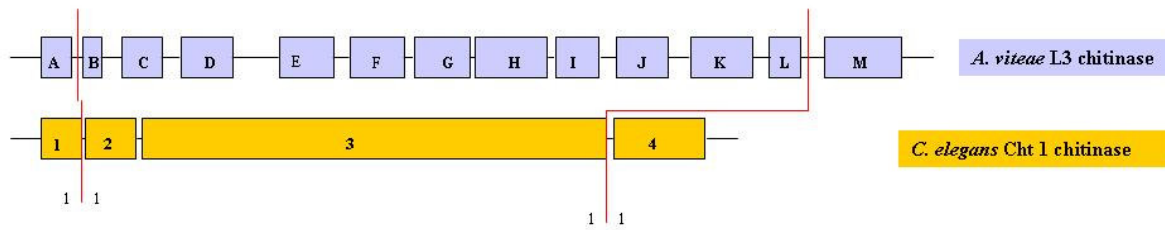


Fig. 10: Comparison of genomic organisation between *Acanthocheilonema viteae* and *Caenorhabditis elegans* chitinase genes.

Exons are shown as coloured boxes separated by lines representing introns. Red lines show two conserved intron phases with the phases indicated about the line. Intron phases are defined relative to the position of introns falling between codons (phase 0), within codon after first (phase 1) or second (phase 2) nucleotides, respectively (Jean et al., 2001a).

Despite the high degree of similarity of certain domains, there is a bigger difference in exon/ intron organisation and position of introns. But, in addition to the high homology with filarial chitinases from *B. malayi* and *A. viteae*, is also the fact that two of the three possible intron phases are conserved (Figure 10), revealing the high degree of relationship between the worm chitinase gene sequences.

2.3 *A. viteae* chitinase transcripts

The analysis of the genomic *A. viteae* chitinase genes showed the existence of at least two potential transcripts from sequences I and III. In order to determine *A. viteae* chitinase transcripts, different approaches were used like the screening of *A. viteae* cDNA libraries as well as the specific amplification of *A. viteae* chitinase nucleic acid sequences from reverse transcribed RNA material. In addition, western blot experiments were performed with material from L3 stages and compared to already existing data. Finally, using a bioinformatic approach, the resulting data were compared with information from various nematode data bases.

2.3.1 Screening of *A. viteae* L3 and adult female cDNA libraries for chitinase transcripts

In order to identify different *A. viteae* chitinase transcripts and to check the stage specificity, two cDNA libraries made from the L3s and from gravid female worms were separately screened in plaque hybridisations using an *A. viteae* cDNA specific probe. The *A. viteae* chitinase probe was a 700 nucleic acid sequence from *A. viteae* chitinase L3 cDNA, corresponding to the exons F to J of *A. viteae* chitinase gene sequence I. The cDNA probe had a 95% identity to the nucleic acids of the

corresponding exons in the *A. viteae* gene sequence III. For both cDNA libraries, 40,000 independent recombinants were screened. The screening procedure resulted in four positive plaques from the screening of the L3 cDNA library, but no positive plaques in the case of the female cDNA library. The resulting four clones of the L3 cDNA revealed a similar size of 1,500 bp for the individual inserts (data not shown). The insert of two of these four clones from the L3 library were *in vivo* excised and sequenced. Both sequences corresponded to the already published cDNA sequence for the L3 stage with minor differences. There are two published L3 sequences (Adam et al., 1996; Wu et al., 1996). These are similar along the length of the molecule with the exception of the extreme 5' end. The cDNA from Wu et al. has an additional alternative start methionine, followed by four more amino acids with an alternative start methionine. Both independently sequenced cDNA inserts from the cDNA library showed identical 5' ends as published by Wu et al. (Appendices 1, 2 and 3). In spite of the published nucleic acid sequences from Adam and Wu et al., there are some minor differences in the chitinase insert sequences identified from the L3 library (Table 5).

Tab. 5: Nucleotide differences between published *A. viteae* chitinase and the chitinase sequence used in this study

<i>A. viteae</i> chitinase		Changes		
Published	This work	Structural	Codon*	Functional
CAA	CAG	A to G transition (wobble)	29	none
GCA	ACA	G to A transition, A to T	406	none
ACC	CCC	A to C transversion, T to P	437	none
---	CAG	Insertion; Q	438/439	none
CAC	CGC	A to G transition, H to R	477	none
CAC	CAT	C to T transition (wobble)	493	none
CTT	TTT	C to T transition, L to F	501	none

The different nucleotides are indicated and the structural changes resulting in different amino acids are indicated in single letter code. Codon numbering is relative to *Acanthocheilonema viteae* cDNA chitinase (Adam et al., 1996)

Apart of the wobble in codon 29, all other differences are found in the variable serine-threonine rich region. The only difference that could have come from alternative splicing as deduced from genomic gene I was the insertion between codons 438/439. All the other differences were confirmed by sequencing independent clones and could represent allelic differences. When all the changes are considered, it may be surmised that the two sequences could code for chitinases with slightly different physical properties, but with similar enzymatic properties since the changes do not occur in sites responsible for enzyme activity.

2.3.2 Identification of *A. viteae* chitinase transcripts

A total of 21 putatively positive plaques were obtained by screening a female *A. viteae* cDNA library. However, all clones did not yield any amplicons in additional PCR screens using vector and chitinase primers, nor did they turn out positive in secondary screens.

Since the screening of a female cDNA library failed to give any chitinase transcripts, the next step to identify different chitinase transcripts from different parasite stages involved the use of RNA from uterine microfilaria and L3 stages for RT-PCR analysis. Accordingly, RT-PCR was done with total RNA of uterine *A. viteae* microfilariae and L3 using primer pairs designed to amplify segments of *A. viteae* chitinase.

RT-PCR was done with L3 and uterine microfilarial cDNA using specific primers for gene sequences I and III located on exons I and M, respectively. In the first PCR amplification experiment, two faint bands of 1.8 and 0.6 kb were amplified with primers specific for gene III, and one amplification product of 0.6 kb resulted from the PCR with the gene I specific primers (Figure 11). A nested PCR with gene III specific primers did not yield specific products, while primers specific for gene I gave rise to a specific PCR product. Identical results were obtained with RNA from both L3 and uterine microfilariae. The 0.6 kb fragment amplified with the specific primers for gene I from the microfilarial stage was subcloned and sequenced and shown to be 100 % identical to the corresponding sequence for *A. viteae* L3 chitinase (Adam et al., 1996; Wu et al., 1996).

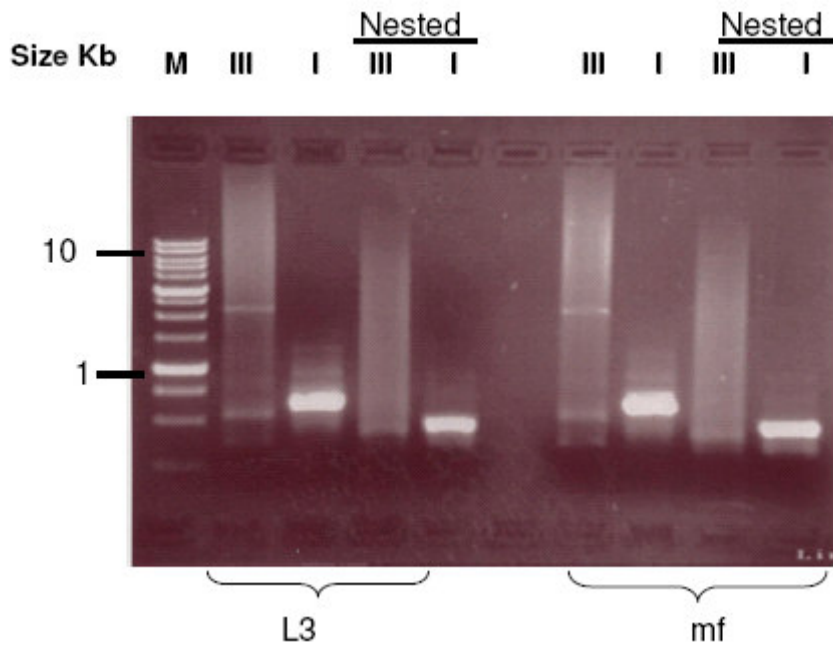


Fig. 11: Transcripts of *A. viteae* genomic sequences in L3 and microfilariae (mf)

RT-PCR results were obtained with primers specific to genomic sequences I and III. Corresponding sequence-specific primers (F8 and R3) were used in RT-PCRs, while a common forward primer (F5) was used with specific reverse primers (R3) in nested PCRs. M, Molecular size marker.

The fragment did not contain the terminal ends of the corresponding *A. viteae* L3 transcript. As the differences between sequences I and III lie particularly in the 3' end, a 3' RACE PCR was done with RNA from uterine microfilariae, L3 and L4 to determine possible variations in the 3' end. A major PCR product of 700 bp was obtained from all three stages as shown by the representative gel in Figure 12. The 700 bp product from mf cDNA was subcloned in pGEM-T vector, checked by sequencing and shown to be 100% identical to the already published 3' end of gene I. The products from L3 and L4 were confirmed by nested PCR to be transcripts of gene I.

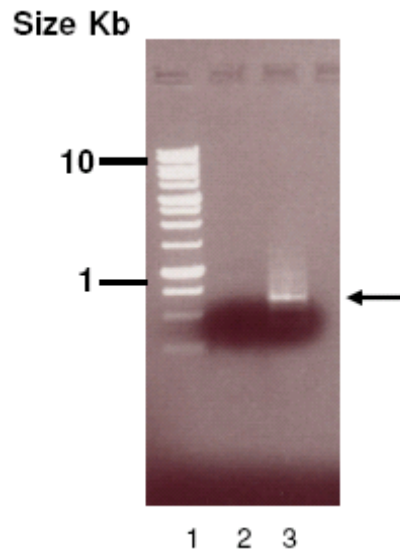


Fig. 12: Amplification of 3' end of cDNA chitinase from different stages using Rapid amplification of cDNA ends (RACE)

Lane 1, molecular size markers; lane 2, negative control; lane 3, amplified chitinase fragment (indicated by arrow).

In conclusion, a transcript for gene I could be amplified from uterine microfilariae, L3 and L4, while transcripts for gene III could not be found in these same stages. However, it can not be excluded that chitinase transcripts are found in other stages.

2.4 Identification of homologous chitinase sequences from nematode databases

A blastx analysis of public data bases with cDNA sequences corresponding to exons of genomic gene I revealed high homologies with orthologous transcripts of filarial (Table 6) and *C. elegans* nematodes (Table 8).

Tab. 6: Orthologous filarial transcripts to *A. viteae* gene sequence I

Sequence accession number	Source of chitinase sequence	Blastx e-value to gene I
L42010	<i>Acanthocheilonema viteae</i> (L3)	7.3e-267
AF250997	<i>Wuchereria bancrofti</i> (mf)	1.4e-197
M73689	<i>Brugia malayi</i> (mf)	5.0e-194
L42021	<i>Onchocerca volvulus</i> (L3)	2.1e-186
U59689	<i>Brugia pahangi</i> (mf)	4.1e-166
U59690	<i>Brugia pahangi</i> (mf)	8.4e-163
NM_076187 (C04F6.3)	<i>Caenorhabditis elegans</i> (embryos)	1.9e-93

With the exception of *Brugia pahangi*, all the other filariae have one described chitinase sequence. *B. pahangi* has two microfilarial chitinase transcripts which differ in the serine threonine rich linker region (Figure 13). Comparison of the filarial amino acid sequences of *Wuchereria bancrofti*, *Brugia malayi*, *B. pahangi*, and *Onchocerca volvulus* to the *A. viteae* L3 chitinase sequence revealed an apparently high identity in the overall amino acid composition, with the exception of a region at the 3' end that corresponds to the serine-threonine rich linker region between the glycosyl hydrolase domain and substrate binding domain (Figure 13).

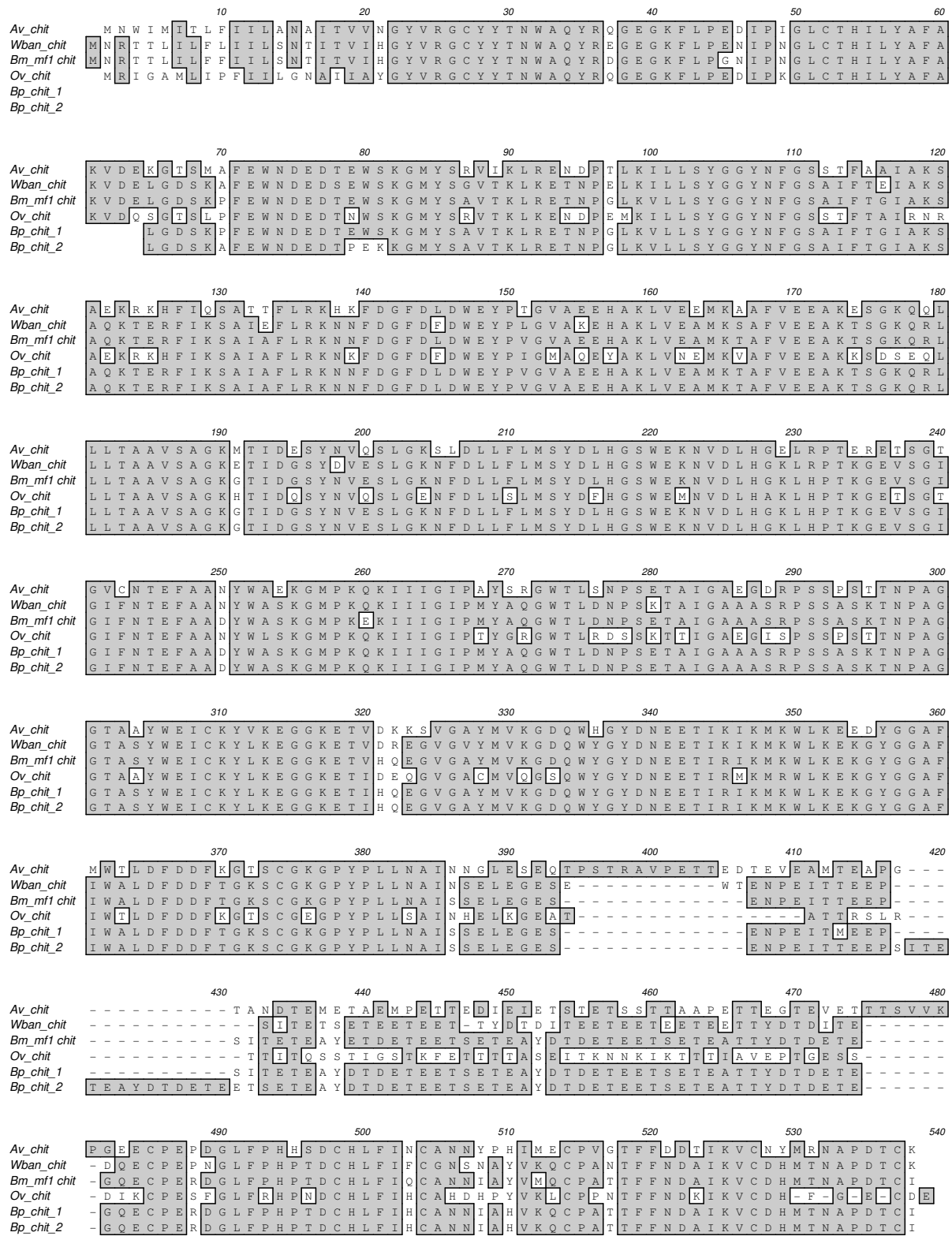


Fig. 13: Amino acid sequence comparison of different filarial chitinases.

Av_chit: *Acanthocheilonema viteae* chitinase (Wu et al., 1996); Wban_chit: *Wuchereria bancrofti* chitinase (Raghavan et al., 1994); Bm_mf1chit: *Brugia malayi* chitinase (Fuhrman et al., 1992); Ov_chit: *Onchocerca volvulus* chitinase (Wu et al., 1996); Bp_chit: *Brugia pahangi* chitinase (Arnold et al., 1996).

A detailed analysis of the EST databases of *B. malayi*, *W. bancrofti* and *O. volvulus* revealed that *B. malayi* databases contain the highest number of chitinase-like ESTs. A clustering of these ESTs resulted in three contigs. One of the three contigs in *B. malayi* is complete and corresponds to the already published sequences for *B. malayi* microfilaria chitinase (Fuhrman et al., 1992). The other contigs are not identical to the published chitinase sequence (Tab 7). In conclusion, *B. pahangi* has two published microfilarial chitinase mRNAs, *W. bancrofti* one microfilarial chitinase, *O. volvulus* and *A. viteae* each has one L3 chitinase mRNA. *B. malayi* has a total of three chitinase sequences with a possible difference in stage-specific expression.

Tab. 7: Brugia malayi ESTs

Cluster or Sequence	ESTs		Status of transcript
	Number	Stage	
BMC00298	AA022373 AA022418 AA246080 AA246111 AA598350 H48963 N44465	Microfilariae	<i>B. malayi</i> microfilaria chitinase (M 73689)
	AA257830	Female	
BMC12278	AA054927	Microfilariae	Not yet characterised: lysin (K) substitutes glutamate (E) at active site
BMC18355	CD570590	2-day irradiated L3	Not yet characterised. Identities to other chitinases: 25% to <i>A. viteae</i> chitinase; 32% to <i>B. malayi</i> microfilarial chitinase.

A blastx analysis of the *C. elegans* database led to 21 confirmed *C. elegans* sequences with some similarity to *A. viteae* chitinase (Table 8).

Tab. 8: *C. elegans* homologues of *A. viteae* chitinase gene sequence I

Wormpep number	Secretory signal sequence	Blastx e-value to sequence V
C04F6.3	yes	6.5e-102
F07G11.9	yes	1.9e-31
T13H5.3	yes	3.9e-25
Y22D7AL.14	no	2.2e-23
F15A4.8b	no	3.4e-20
F15A4.8a	no	1.5e-15
R09D1.10	no	1.5e-14
R09D1.5	no	2.9e-13
R09D1.8	no	3.2e-12
R09D1.3	no	1.1e-11
C45E5.3	no	2.7e-11
R09D1.11	no	8.4e-11
C25A8.4	yes	2.2e-10
R09D1.9	no	4.9e-10
C08H9.12	no	2.1e-09
C08H9.14	yes	3.1e-09
C08H9.13	no	4.3e-09
T19H5.1	no	9.2e-09
C08H9.11	no	1.5e-08
M176.8	no	1.7e-07
C08H9.7	no	3.9e-07

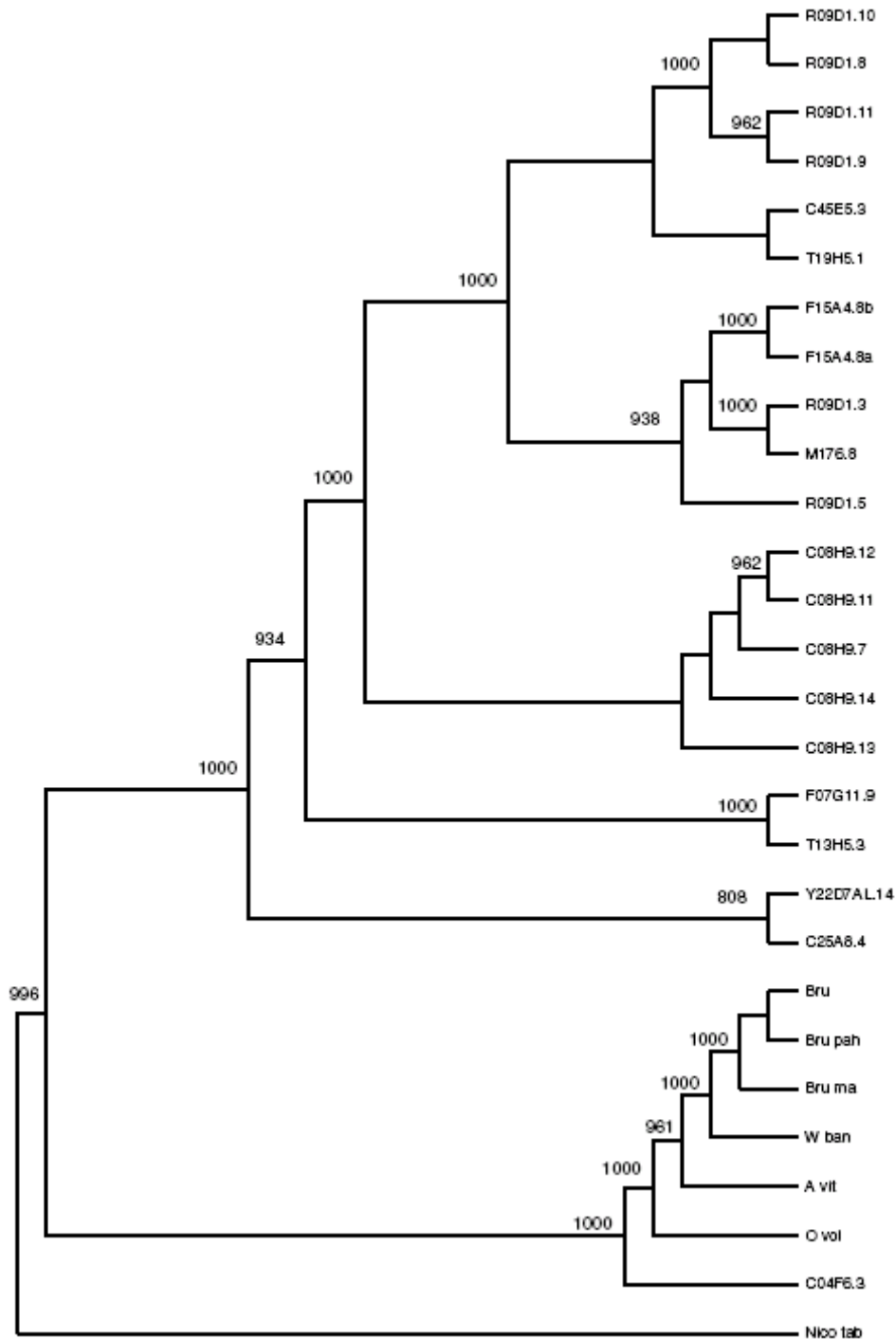


Fig. 14: Phylogenetic relationships of filarial chitinases with *C. elegans* chitinases

The bootstrapped tree was rooted using chitinase from *Nicotiana tabacum* (Nico tab) (Gene bank: X78325) as outgroup. Bootstrap values for 1000 replicates are shown where they are significant (>700). Chitinase sequences are from the following organisms: *Acanthocheilonema viteae* (A vit), *Brugia malayi* (Bru ma), *B. pahangi* (sequence 1= Bru; sequence 2 = Bru pah), *Onchocerca volvulus* (O vol) and *Wuchereria bancrofti* (W ban). All other sequences are from *Caenorhabditis elegans*.

C04F6.3 represents the *C. elegans* chitinase with highest homology and phylogenetic relationship to the filarial chitinases.

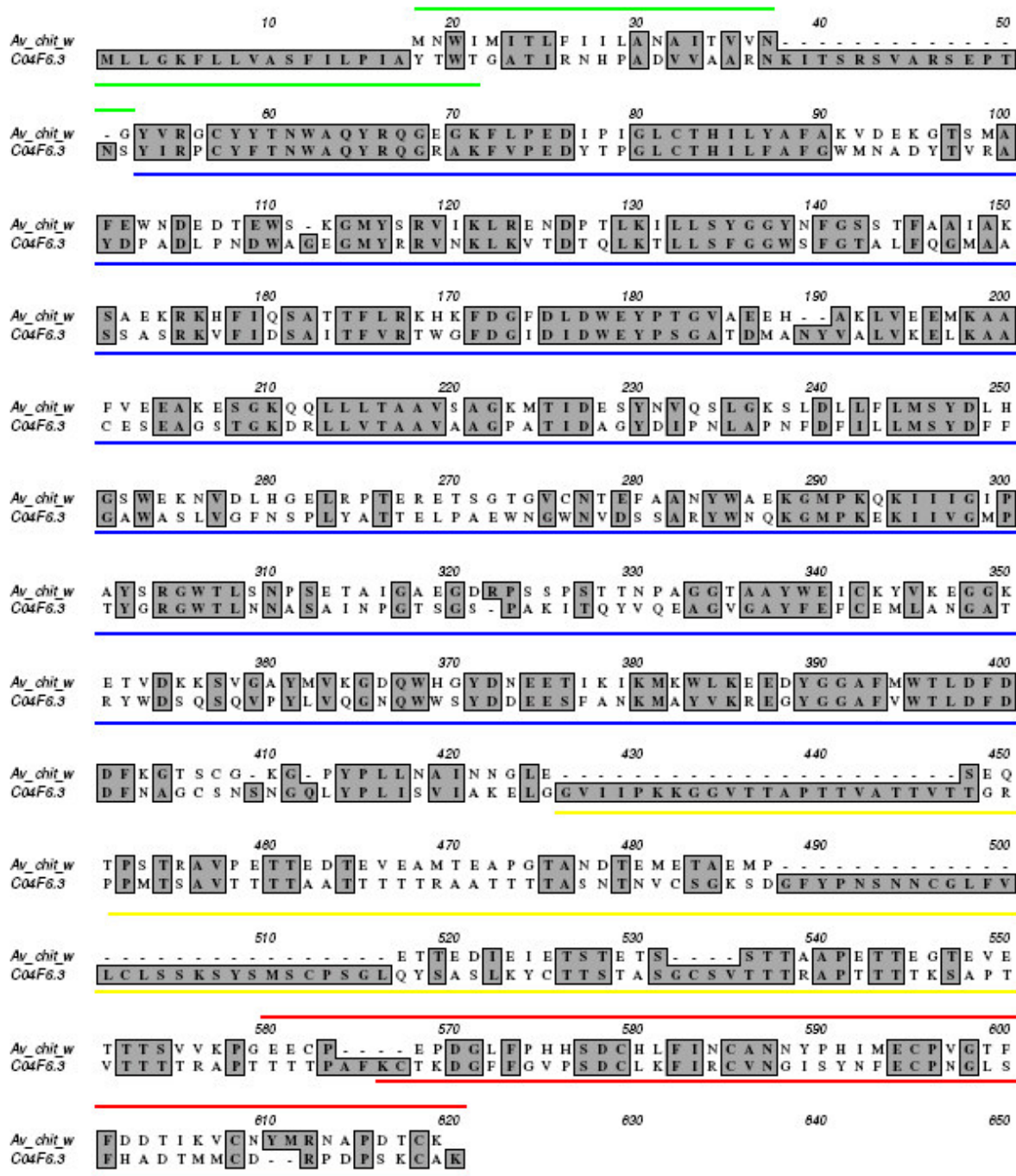


Fig. 15: Amino acid sequence comparison of *Acanthocheilonema viteae* chitinase with *Caenorhabditis elegans* sequence C04F6.3.

Regions corresponding to different domains are indicated with a line closely apposed to the sequence: Green, signal sequence; Blue, glycosyl hydrolase domain (the same for both sequences); Yellow, serine-threonine rich linker; Red, chitin binding domain.

Following phylogenetic analysis of confirmed *C. elegans* chitinase with other filarial chitinase sequences (Figure 14), *C. elegans* sequence C04F6.3 clustered with the filarial chitinases from *A. viteae*, *B. malayi*, *B. pahangi*, *O. volvulus* and *W. bancrofti*.

An alignment of C04F6.3 with *A. viteae* chitinase is shown in Figure 15. There is a great difference in similarity between the two derived protein sequences. Though the overall identity between both sequences is barely 34 %, a general analysis reveals the presence of the four important motifs described for family 18 chitinases: there is a secretory signal sequence, a glycosyl hydrolase domain, a serine-threonine rich linker region, and a substrate binding domain. The important consensus sequence of the active site is fully conserved.

In conclusion, *C. elegans* has several family 18 chitinase genes, one of which is closely related to the filarial chitinases.

2.5 Cloning, expression and purification of recombinant *A. viteae* chitinase

2.5.1 Cloning, periplasmatic/ cytoplasmatic expression and purification of N-terminal portion of *A. viteae* chitinase in the expression vector pET 22 b (+) in *E. coli*

The DNA sequence of the glycosyl hydrolase domain, corresponding to nucleotides 52-1,117 of *A. viteae* L3 chitinase was subcloned into the pET 22 b(+) vector, leading to the expression of a protein with an N-terminal vector-specific bacterial signal sequence and a C-terminal 6- His tag for purification. The bacterial signal sequence allows for potential periplasmatic localisation of the expressed protein and subsequent release of the bacterial signal sequence by bacterial signal peptidases. This construct was transformed into BL 21 (DE3) *E. coli* cells, and expressed as a protein of 43 kDa representing about 40 % (about 7 mg) of the total bacterial biomass, following induction with IPTG as shown in Figure 16. The protein could be exported into the periplasmatic space, released through osmotic shock and purified by affinity chromatography from the osmotic shock fluid (Figure 16). As shown on SDS-PAGE in Figure 16, there is a main band representing N-terminal chitinase without any signal sequence, while there is a minor band that runs slightly higher and should represent the soluble cytoplasmic fraction with the intact bacterial signal sequence. The total soluble protein purified represented about 15 % of the total protein expressed corresponding to 1.1 mg protein from 500 ml bacterial culture.

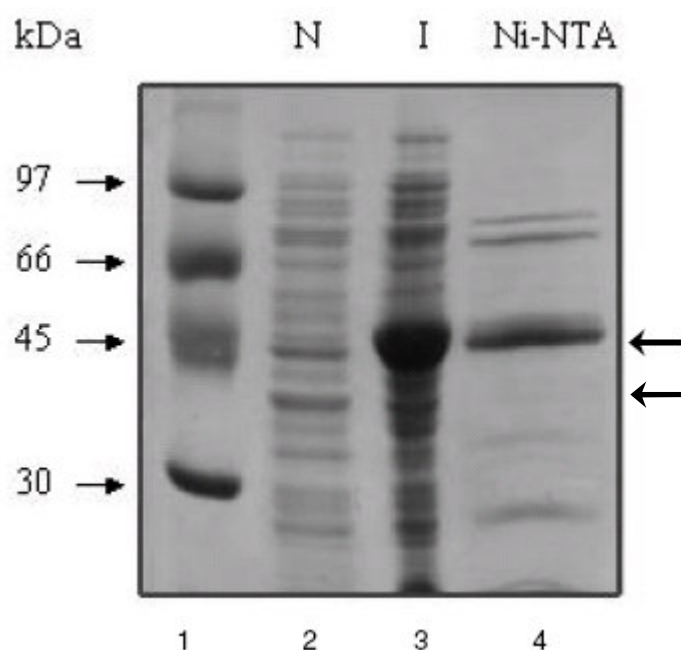


Fig. 16: Expression and purification of *A. viteae* chitinase from the periplasmatic space

Lane 1, low molecular weight marker; lane 2, lysate of non-induced bacteria (N); lane 3, lysate of induced bacteria (I); lane 4, eluted fraction from a Ni-NTA column of *A. viteae* chitinase (arrow)

The insoluble cytoplasmic fraction was solubilised in 8 M urea and also purified by affinity chromatography (Figure 17). The purified protein in urea was then dialysed successively against decreasing concentrations of urea in phosphate buffer. A greater part of the protein (4mg) precipitated during dialysis giving rise to just over 1mg soluble protein per 500 ml bacteria culture.

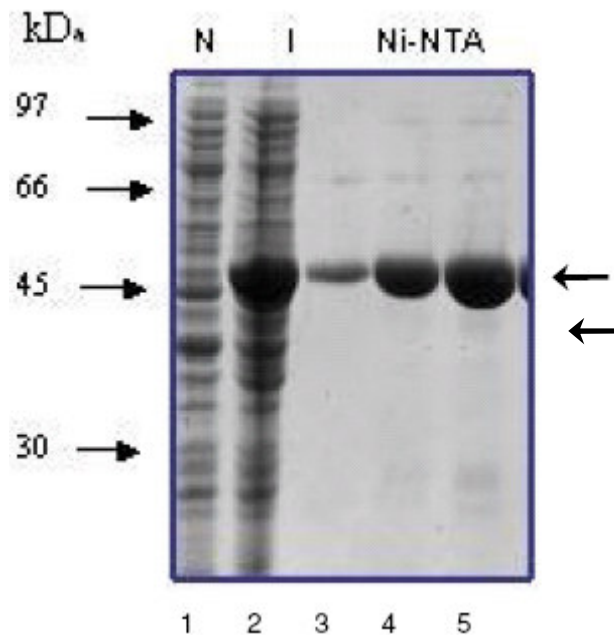


Fig. 17: Expression and purification of *A. viteae* chitinase under denaturing conditions

Lane 1, lysate of non-induced bacteria (N); lane 2, lysate of induced bacteria (I); lanes 3-5, fractions of *A. viteae* chitinase eluted from Ni-NTA column.

2.5.2 Activity of N-terminal chitinases

The activity of the N-terminal fragment of chitinase, purified from the cytoplasmic as well as from the periplasmic space, was assessed using the substrate 4-methylumbelliferyl β -N',N'',N''' chitotrioside that releases the fluorescent product 4-methylumbelliferone upon cleavage by a glycosyl hydrolase. Recombinant N-terminal *A. viteae* chitinase, purified under denaturing conditions and dialysed against phosphate buffer, was shown to have chitinase activity that had about 50 times less specific activity (defined as relative fluorescent units per hour per ng protein) as compared to the positive control, which was Baculovirus expressed *O. volvulus* chitinase (New England Biolabs) (Figure 18, Table 8). The negative control protein, *A. viteae* cystatin (Av17) did not hydrolyse the substrate.

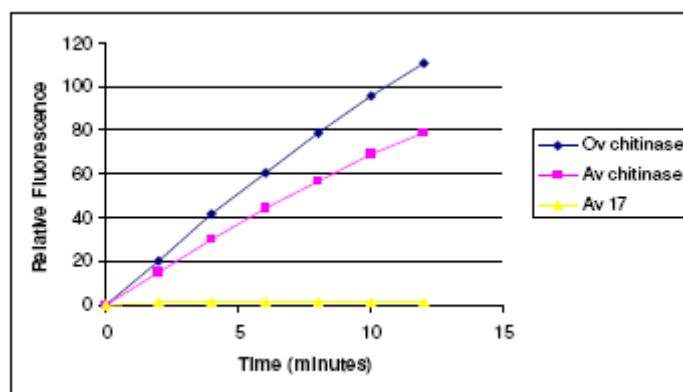


Fig. 18: Activity assay of purified cytoplasmic N-terminal chitinase

Activity assays were done with *A. viteae* chitinase (pink line), *O. volvulus* chitinase as positive control (blue), and Av 17 (filarial cystatin) as negative control. The amounts of protein assayed were 1.043 ng *O. volvulus* chitinase, 38.5 ng *A. viteae* chitinase and 38.5 ng Av 17.

Tab. 9: Specific activity of purified *A. viteae* and *O. volvulus* chitinases

Enzyme	Specific activity (Units*/hr/ng)
<i>A. viteae</i> N-terminal chitinase (Cytoplasm)	11.1
<i>A. viteae</i> N-terminal chitinase (Periplasm)	166.14
<i>O. volvulus</i> chitinase (NEB)	553.8

The specific activities of *A. viteae* and *O. volvulus* chitinases are shown. *The specific activity is reported as relative fluorescence units per hour per ng protein used.

The activity of *A. viteae* N-terminal chitinase purified from the periplasmic space was originally determined using *Serratia marcescens* chitinase (Sigma) as standard. The relative fluorescence units were estimated to have about 30 % of the activity of baculoviral expressed *O. volvulus* chitinase (New England Biolabs) (data not shown).

2.6 Immunisation studies with *A. viteae* chitinase in *Meriones unguiculatus*

For the evaluation of protective potentials of different vaccines, *Meriones* were vaccinated and subsequently challenged with 70 L3s of *A. viteae*. Two preparations were used in this study, namely recombinantly expressed glycosyl hydrolase domain

(N-terminal fragment) of *A. viteae* chitinase, and synthetic peptides designed from the active site of *A. viteae* chitinase.

2.6.1 Immunisation studies with *A. viteae* N-terminal chitinase fragment

Recombinant N-terminal chitinase of *A. viteae* was used in vaccination experiments with STP and Alum as adjuvants. Immunisation experiments were done following the scheme on Figure 19. Ten-week-old *Meriones* were given a primary immunization subcutaneously at the flanks with 25 µg protein, followed by two boosters on the 2nd and 4th weeks before challenge. *Meriones* were bled for sera on weeks -6, 0, 4, 8 and 12, and the microfilaria load was checked on the 8th and 12 weeks following challenge. *Meriones* were sectioned for retrieval of adult worms on the 12th week after challenge (Figure 19).

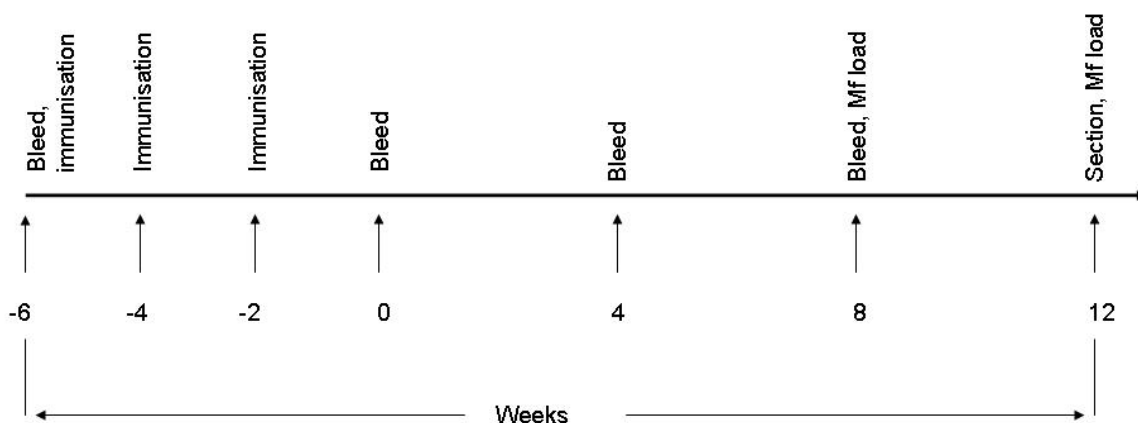


Fig. 19: Immunisation scheme of *Meriones unguiculatus* with experimental vaccines

Recombinant N-terminal *A. viteae* chitinase purified under denaturing conditions was used in the immunization of 10-week-old *Meriones* with STP and Alum as adjuvants. During the course of the experiment, one animal died from the chitinase/ Alum group, two from the chitinase/ STP group, three from the STP group and three from the PBS group (Table 10). Vaccination of *Meriones* with the glycosyl hydrolase domain of chitinase and STP as adjuvants led to a 29% significant overall reduction in worm burden ($p < 0.05$), while vaccination with Alum as adjuvant did not lead to any overall reduction in worm burden (Figure 20, Table 10).

Tab. 10: Summary of biometric data from immunization studies with N-terminal *Acanthocheilonema viteae* chitinase

Antigen/ control	Chitinase/ Alum	Chitinase/ STP	Alum	STP	PBS
Worms recovered	20.16±6.43	17.4±3.43	20.57±3.59	24.5±4.65	24.85±4.84
Female worms (Length)	13.83±4.4 (4.7±0.4)	11 ±2.5 (4.57±0.3)	13.57±3.4 (4.55±0.3)	14.25±2.7 (4.7±0.4)	13.42±3.4 (4.79±0.4)
Male worms (Length)	9.2±5.5 (2.08±0.1)	6.4±4.3 (2.35±0.1)	8.5±4.1 (2.2±0.1)	10.25±2.7 (2.19±0.1)	11.42±2.7 (2.2±0.2)
Mf density / ml blood	285±117.8	216.2±46.2	265±100.2	220±116	432.4±350
Surviving animals	6/7	5/7	7/7	4/7	7/10

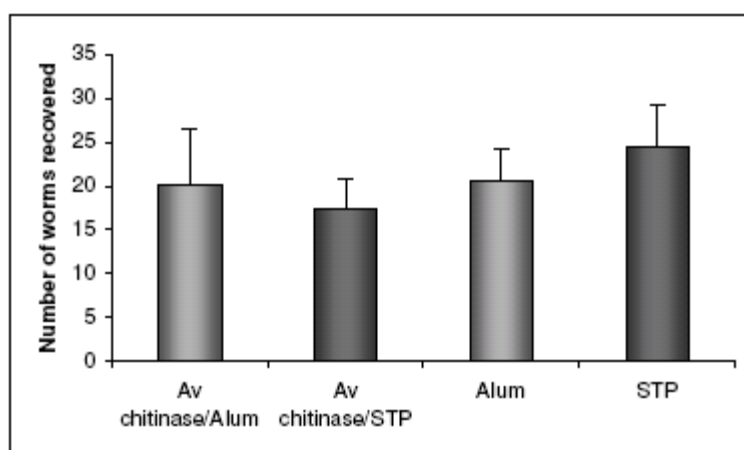


Fig. 20: Immunisation of *Meriones unguiculatus* with N-terminal *Acanthocheilonema viteae* chitinase

The number of worms recovered following immunization with N-terminal chitinase (*rA. viteae* chitinase) and STP and Alum as adjuvants are shown. Controls: Alum and STP.

When the microfilaria load was compared, there was no statistically significant difference in the microfilarial burden between experimental and control groups on the 10th week following challenge infection (Table 10). There was equally no statistically significant difference in the lengths of the worms obtained from experimental and control groups (Table 10).

2.6.2 Immunisation studies with synthetic peptides from the active site of *A. viteae* chitinase

To investigate whether antibodies towards the active site of *A. viteae* chitinase could inhibit chitinase and thus lead to protection against challenge infection, synthetic peptides were used in immunization studies. The synthetic peptides were designed from the active site of *A. viteae* chitinase. For the identification of relevant amino acids, the process of homology modeling was used. *Coccidioides immitis* chitinase, a family 18 chitinase whose structure is known, was used as a template to predict the structure of *O. volvulus* chitinase. The corresponding amino acids were then obtained from *A. viteae* chitinase following alignment with *O. volvulus* chitinase. Homology modeling and synthesis of peptides were done respectively by Professor Höhne and Dr. Volkmer-Engert of the Charité Medical Faculty, Humboldt University of Berlin. Three candidate peptides were identified (see materials) and synthesized, and two turned out to be soluble. To investigate whether the two peptides could generate a specific immune response, they were coupled to a KLH carrier protein and 100 µg of coupled peptide/ KLH was used to immunize BALB/c mice with Alum as adjuvant. The sera of immunized mice could recognize full-length native *A. viteae* chitinase in western blots with *A. viteae* L3 total antigen (Figure 21).

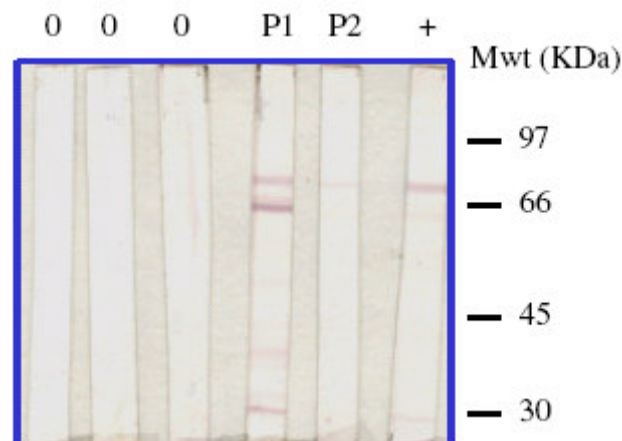


Fig. 21: Recognition of native *Acanthocheilonema viteae* L3 chitinase by sera of mice immunized with peptides.

Western blot reactions are shown for zero bleeds (0) and the reaction of antisera of peptides 1 and 2 to native chitinase. Positive control: Antisera to N-terminal chitinase protein.

The sera from animals immunized with P1 gave stronger reactions than sera from mice immunized with P2. The two synthetic peptides, P1 and P2, were then coupled with KLH and used for immunization. The coupled peptides (100 ug) were administered with alum as adjuvant at the flanks of the animals. Alum was used as adjuvant to shift the immune response towards the Th2 direction since the initial premise was to produce neutralizing antibodies to the active site of chitinase. Boosting, bleeding, challenge and section were done as in section 2.6.1.

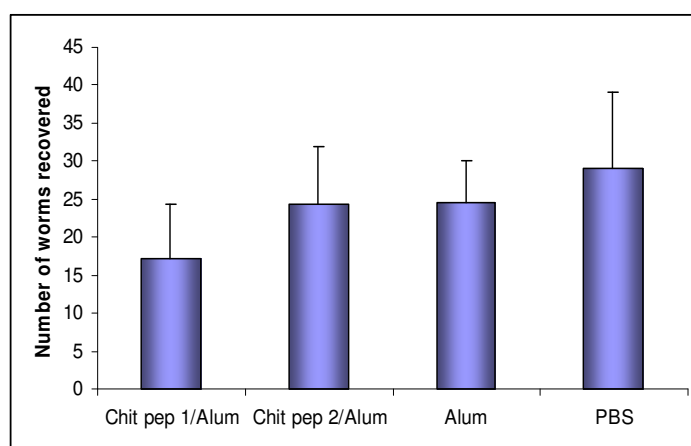
A summary of the biometric data obtained from both experiments is shown in Table 11.

Tab. 11: Summary of biometric data from immunization studies with synthetic peptides from *Acanthocheilonema viteae* chitinase

Experiment A				
Antigen/ control	Peptide 1/ Alum	Peptide 2/ Alum	Control alum	Control PBS
Worms recovered	17.2±7.1	24.3±7.6	24.5±5.6	29.5±10.1
Female worms (Length)	10.7±4.8 (4.99±0.15)	13.9±3.5 (5.16±0.46)	13.3±3.8 (5.08±0.21)	16.4±5.4 (5.25±0.65)
Male worms (Length)	6.5±2.2 (2.29±0.18)	10.4±4 (2.36±0.17)	11.2±2.8 (2.22±0.2)	13±5.2 (2.44±0.22)
Mf density / ml blood	274±146	574±215	439±54	456±152
Surviving animals	4/7	5/7	6/6	9/11

Experiment B			
Antigen/ control	Peptide 1/ Alum	Peptide 2/ Alum	Control PBS
Worms recovered	25±3.2	24.7±5.3	33.1±10.2
Female worms (Length)	15±4.2 (5.02±0.7)	11.8±4.6 (4.58±0.7)	19.1±4.8 (5.15±0.9)
Male worms (Length)	10.3±2.1 (2.32±0.3)	10±2.1 (2.31±0.3)	14.7±7.2 (2.38±0.4)
Mf density / ml blood	443.2±171.8	208.1±58.5	833±384.2
Surviving animals	6/6	6/6	9/11

A:



B:

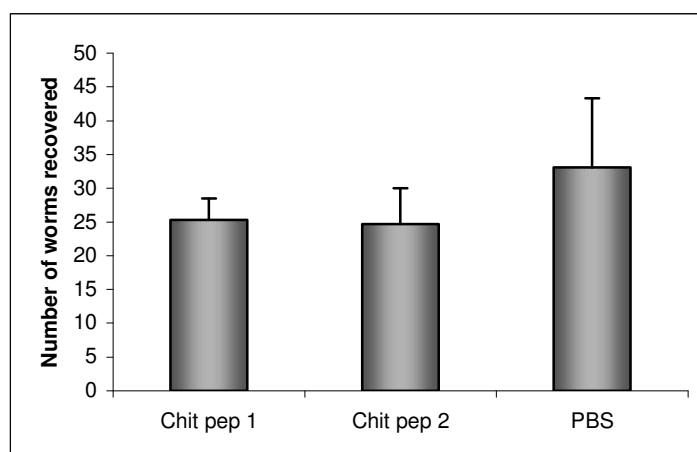


Fig. 22: Vaccination of *Meriones* with KLH-coupled chitinase peptides 1 and 2

The diagram shows the worm burdens following vaccination with 2 different chitinase peptides in two independent experiments, A and B.

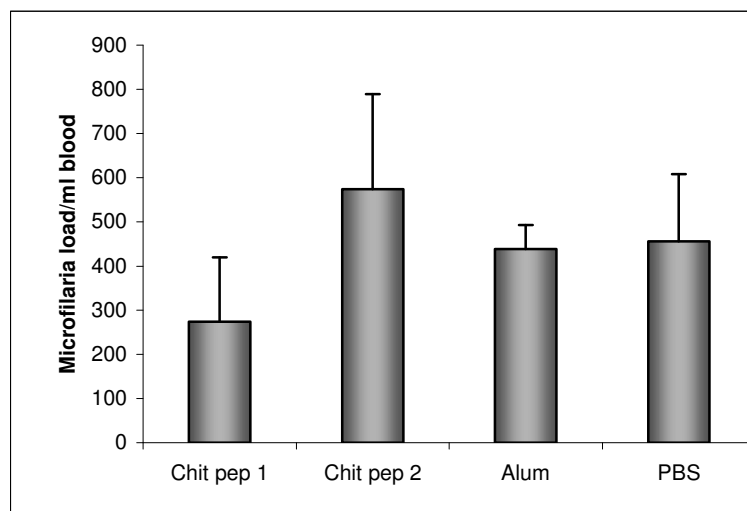
There was no significant difference in the lengths of the worms recovered from both experimental and control groups, indicating that immunization with the peptides may not have influenced the growth of the worms.

In two independent immunization experiments, data from animals vaccinated with peptide 1 suggested a trend towards decrease (33% and 23.4 %) in worm burden as compared to the control groups (Figure 22; Table 11). The first experiment was stopped at week 10 owing to death of animals in different groups (Table 11). Vaccination of *Meriones* with peptide 2 did not lead to an overall decrease in worm burdens in the first experiment, but showed a tendency towards decrease (25.5 %) in

worm burden in the second experiment (Figure 22). In the first experiment, the microfilaria load was significantly reduced ($p < 0.05$) in the group of *Meriones* immunized with chitinase peptide 1 (Figure 23), while the chitinase peptide 2 group showed no tendency towards reduction.

In the second experiment, the microfilaria load was compared between experimental and control groups on the 12th week following challenge, and it could be shown that vaccination with chitinase peptide 1 and peptide 2 led to 45% and 75% significant reductions in microfilaria load ($p < 0.05$), respectively (Figure 23).

A



B

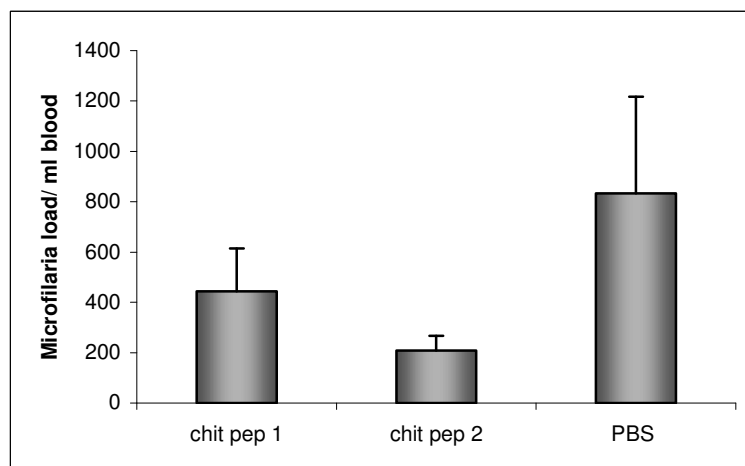


Fig. 23: Microfilaria burden from *Meriones* vaccinated with KLH-coupled chitinase peptides

The microfilaria burden following vaccination with chitinase peptides (chit pep) 1 and 2, as well as Alum and PBS controls are shown.

3 Discussion

3.1 Analysis of *A. viteae* chitinase genes and transcripts

One of the major goals of our study was the characterisation of *A. viteae* chitinase genes. This involves the description of the genomic structures as well as the characterisation of different transcripts of the individual stages of the life cycle of *A. viteae*.

According to our studies, the genome of *A. viteae* contains three different chitinase genes.

Southern blot analysis showed, particularly for the Pvu II digest, that there are at least two different chitinase genes. However, the different band intensities of especially the Pvu II digest could be explained by the fact that the probe bound at least twice more to close gene copies on a single DNA fragment. This indicates that there is a cluster with of a minimum of two chitinase genes. A similar suggestion was made by Arnold and colleagues (1996), who investigated gDNA from *B. malayi* for chitinase genes. The variation in the intensity of individual bands was also explained by the existence of a gene cluster. Further analysis of nine genomic sequences from a gene library of *A. viteae* worms confirmed the existence of three independent chitinase gene sequences. The sequences of two genomic clones (1 and 9) had a cluster of two genes as predicted from Southern hybridisation results, and an identical overlapping region with some minor differences. The corresponding genomic region of two inserts comprises a length of 6,845 bp, of which 93% was identical between two pairs of genes. The main differences arose from the beginning of insert 1, the end of insert 9, a repeating sequence of 25 nucleotides and a non-matching region in the intergenic region of both sequences. An additional argument for the identity of both sequences is the fact that the introns of both pairs of chitinase genes were 100% identical. This strongly indicates that the two pair of sequences are identical because intron sequences should show a higher mutation rate (Neafsey et al., 2005) since they are not under evolutionary pressure as exon sequences coding for particular amino acids responsible for the function of a protein. This hypothesis was confirmed by sequencing two independent clones (13 and 4) with similar restriction maps like clones 1 and 9. Analysis showed that the incomplete gene sequences at the beginning of clone 1 and at the end of clone 9 were truncated versions of gene sequences II and III, respectively. The discrepancies initially

observed could have been due to sequencing errors or to artefacts inherent in the construction of the genomic libraries.

In consequence, there were three independent genomic chitinase sequences that were very similar in the structure, and showed the highest degree of variation in exons for the serine threonine rich domain in the 3' part of the genes. A comparison with the structure of the *B. malayi* MF 1 chitinase gene showed that the whole structure of the genes is identical to the *A. viteae* genes, with the only difference being in the exons coding for the serine threonine rich domain. There are three other annotated chitinase genes in *B. malayi* having a broad range of organisations with some domains and exons fully, partially represented or absent (<http://www.tigr.org>). The absence of some exons in these genomic sequences could be seen as a footprint of unequal recombination occurring between exons of homologous and nonallelic chitinase genes (Maeda and Smithies, 1986). Interestingly, two of these chitinase genes are found in tandem on the same genomic assembly. Taken into account that a similar situation was demonstrated for *A. viteae*, it may be surmised that these genes exist in a cluster in these filarial parasites and are consequently a hot spot for recombination.

A. viteae chitinase gene sequence II, in contrast to sequences I and III, had no first exon with a start ATG and probably no functional transcript. A comparable sequence is found in the genome of *B. malayi*. Both sequences lack a first exon, and have an irregular second exon, a situation which may have arose by unequal crossing over resulting to the loss of the 5' exon with the start ATG (Maeda and Smithies, 1986). While *A. viteae* gene II has a regular stop codon and polyadenylation site, and may thus be considered as part of a true gene, the *B. malayi* sequence is definitely a pseudogene due to the presence of proretrovirus-like elements after the 10th exon (Felder et al., 1994), and to an alteration of the reading frame.

The gene structure for the chitinase gene of the free living nematode *C. elegans* has a different distribution of exons coding for the enzymatic domain and for the serine-threonine rich linker structure. An unusually long exon codes for the glycosyl hydrolase domain responsible for the catalytic activity of the deduced protein, and for most of the serine threonine rich linker region. The exons coding for the secretory signal sequence and the chitin binding domain correspond to those in the filariae.

The *C. elegans* chitinase is an orthologue of the filarial chitinases, despite the lack of similar introns and conservation of all intron positions and phases. This is congruent with the clustering of all of these sequences in a phylogenetic tree, and to the fact that no other *C. elegans* chitinase has a better match to filarial chitinases than this *C. elegans* chitinase. A similar observation was made by comparing an 83 kb syntenic region between *B. malayi* and *C. elegans* and demonstrating that corresponding orthologous genes clearly had differences in gene structure (Guiliano et al., 2002). Apart of this chitinase, there are some other 30 expressed chitinase genes in *C. elegans*, whose implication and function have not been addressed (www.wormbase.org; Popovici et al., 1999).

It is not known why *C. elegans* has many more chitinase genes than the filarial nematodes. The *C. elegans* chitinase gene family is made up of many orthologues and homologues that may have arisen by duplication and diversion of a common ancestor gene (Popovici et al., 1999). Though the function of all the members of this gene family is not yet known, diversion may have led to new copies with different functions as well as tissue or substrate specificities. Some of the genes are very identical with up to 10 nonallelic copies clustered on one chromosome, while others have very low identities and are widely dispersed on different chromosomes. A general conclusion drawn from the *C. elegans* chitinases would be that there may be more members of a gene family than one could suspect on the basis of protein analysis and by the use of probes to fish out identical genes of the same family. This is supported by the fact that many of the chitinase genes share common motifs, but have very low sequence identities. In line with this hypothesis, there may be several other chitinase genes in the filarial nematodes which have not been characterised based on the low identities they share with known family members. Support for this hypothesis comes from the *B. malayi* irradiated L3 chitinase EST that has a bare 32 % identity to the published sequence for *B. malayi* chitinase. Another observation showing that such a phenomenon is present in the filarial genome is the description of two chitin synthases in *Brugia* with a bare 27% identity (Harris et al., 2000; Foster et al., 2005). The existence of orthologous genes with the same function in a parasite makes it difficult to effectively use these genes as vaccine or drug targets. Thus, the further characterisation of filarial chitinases as drug or vaccine targets will entail the identification and/or exclusion of further functional chitinase genes in the nematode of interest.

In keeping with the identification of three chitinase genes in *A. viteae* it was hypothesised that there could be several stage-specific *A. viteae* chitinase genes which may have different substrate specificities or functions. There are two published *A. viteae* chitinase cDNA sequences (Adam et al., and Wu et al., 1996) that differ only in the outmost 5' ends. The Wu et al., sequence begins with a start methionine and three additional amino acids which are absent in the Adam et al., sequence. Within the frame work of this study, data from cDNA and genomic sequences could confirm that the Wu et al., sequence is the variant that is expressed *in vivo*. The exons of *A. viteae* genomic gene I are 100% identical to the sequence of Wu and colleagues, particularly with regards to the 5' end. Transcripts could actually be found for this sequence in uterine microfilaria, blood microfilariae, L3 and L4, while no transcripts could be found for gene sequence III. It would however not be excluded that other *A. viteae* chitinase-like transcripts could be present in *A. viteae*, since western blot data show the existence of four molecules reacting with antibodies to *A. viteae* chitinase. It was shown that sera of *Meriones* protected by vaccination with irradiated L3 were mainly directed against L3 molecules of 205 kDa, 68 kDa and 17 kDa (Lucius et al., 1991). The 205 and 68 kDa molecules were characterised as chitinases (Adam et al., 1996). In addition, monoclonal antibodies produced from sera of animals immunised with irradiated attenuated L3 and Excretory secretory products (ESP) could recognise an array of molecules in different filarial stages in immunoblots (Adam et al., 1996). Amongst others, a 68 kDa chitinase could be shown for L3 and L4, while 220 kDa, 205 kDa and 140 kDa molecules were found in female worms, L3 and microfilariae, respectively. The authors proposed that the 205 kDa L3 molecule could be as a result of oligomers formed from the 68 kDa monomers, or post-synthetic modification of the monomer. The antigens from female worms were shown to be from uterine microfilariae (Adam et al., 1996). The implication of such a finding could be that there are possibly different stage-specific chitinases in microfilariae and L3 of *A. viteae*. While we have demonstrated that the same chitinase molecule (68 kDa) is expressed in mf, L3 and L4, it remains to be shown that there are other chitinase messages in different parasite stages.

Filarial chitinases had been exclusively attributed to the L3 and mf stages of *A. viteae* and *O. volvulus* (Wu et al., 1996; Adam et al., 1996) or to the microfilarial stages of *Brugia* (Fuhrman et al., 1992; Arnold et al., 1996). However, *Brugia* EST data point to

the existence of a novel chitinase transcript in *B. malayi* L3, a stage from which no chitinase gene had previously been characterised. In this study, we showed chitinase transcripts in uterine microfilaria and L3 and L4, but not in the cDNA library from adult female worms, even though the uterine microfilariae were derived from gravid female worms. A similar difficulty was found in the isolation of *B. malayi* chitinase from microfilaria (Southworth et al., 1996) and *O. volvulus* chitinase (Perler, personal communication) from a female cDNA library. In both cases, the reasons advanced were the low representation of chitinase transcripts in the total transcript population of the filarial stages looked into, or suboptimal isolation of targeted transcripts from these filarial stages (Southworth et al., 1996; Perler, personal communication).

A comparison of the deduced amino acid sequences from the three *A. viteae* chitinase gene sequences revealed a very high rate of identity except for the serine threonine rich linker region. The variation in the serine threonine rich linker region of *A. viteae* chitinase is also found when different filarial chitinase sequences are compared. It is not known whether the size and composition of the serine threonine rich linker are of particular relevance to the function of the protein. However, this region has several consensus sequences for glycosylation and when glycosylated will influence protein solubility, secretion and the resistance of the protein against proteases as has been demonstrated for insect chitinases (Arakane et al., 2003). Such properties could be of particular relevance to the host-specificity of parasites and their chitinase molecules.

The function of chitinases in filarial worms is not yet known. However, in protozoan parasites like *Plasmodium* and *Leishmania*, chitinase has been shown to be important for transmission by their role in lysis of the arthropod peritrophic matrix (Langer and Vinetz, 2001; Romalho-Ortigao and Traub-Cseko, 2003). While *Simulium* vectors of *O. volvulus* have been reported to have peritrophic membranes (Ramos et al., 1994; Demanou et al., 2003; Soubey-Alley et al., 2004), such membranes have not been demonstrated in tick vectors of *A. viteae* (Sonenshine, 1991). It is therefore not known whether similar mechanisms operate in these vectors. However, L3 chitinases of *O. volvulus* and *A. viteae* could have a role in the

egress of these parasites from their arthropod vectors since their expression coincides with the transmission of the parasites (Wu et al., 1996).

If chitin is the unique substrate for filarial chitinases, it may be speculated that chitinases play a role in parasite transmission based on two observations. Firstly, *Brugia microfilariae* express chitinase; secondly, it has been shown that *Brugia microfilariae* not expressing chitinase could not penetrate the arthropod vector (Fuhrman et al., 1987). It is hypothesised that lectins or agglutinins in the arthropod midgut binds to the microfilariae and reduces their infectivity. Chitinase promotes infectivity by digesting its substrate and releasing N-acetylglucosamine (GlcNAc) that saturates the lectins and reduces their interaction with microfilariae. Such a hypothesis has been proven in other parasite systems by supplementing the infection blood meal with N-acetylglucosamine, the monomers of chitin, which in turn increases infectivity (Welburn et al., 1993). Such a hypothesis can be tested in the filariae by carrying out similar experiments.

Parallel to the role of insect chitinases in ecdysis (Kramer and Muthukrishnan, 1997), filarial chitinases are speculated to be involved in moulting and hatching, since filarial eggs and adult sheath have been shown to contain chitin (Harris et al., 2000; Brydon et al., 1987). Such a role in ecdysis was suspected for *A. viteae* chitinase by the expression of chitinase on the surface of microfilaria, just before they hatch from their modified eggshell, and the absence of chitinase in blood-borne microfilaria (Adam et al., 1996).

3.2 Expression, purification and activity of N-terminal chitinase

There are three possibilities to control filarial infections: vector control, chemotherapeutic control, and immune control through vaccines. Chitinase is an attractive target for vaccine and drug control, since it is an enzyme that is expressed in two key stages (microfilariae and L3s) in the life cycle of filariae. In addition, knock out experiments with the *C. elegans* chitinase orthologue show that this gene is important for the development of larvae that are the equivalent of microfilariae in the filarial nematodes (www.wormbase.org). Moreover, knowledge on the structure of filarial chitinases will help towards the development of selective inhibitors against these molecules. Chitinase inhibitors might at the same time also target other chitin-containing pathogens like fungi and arthropods. For these reasons, it was necessary to produce large amounts of soluble and active *A. viteae* chitinase.

A bacterial expression system was chosen for the expression of *A. viteae* chitinase. The chitinase gene was cloned into the pET 22 b (+) vector (Novagen) and could be expressed in a suitable *E. coli* host. Expression and purification of large amounts of soluble full-length molecule was not possible, and so the N-terminal portion corresponding to the glycosyl hydrolase domain of the molecule was expressed. Failure to obtain a full-length molecule could have been due to difficulties in disulfide bond in the substrate-binding domain of the molecule that contains 6 cysteins. Amongst other things, the expression of soluble active chitinases from filarial parasites (Southworth et al., 1996; Drabner et al., 2001) and other parasites like *Leishmania* (Razek-Desoukey et al., 2001) has always been difficult due to the 6 cysteins in the chitin binding domain, which disturbs proper folding. To overcome these problems of expressing soluble proteins in heterologous genes in *E. coli*, different strategies are used like reduction in induction temperature (Schein and Noteborn, 1989; Razek-Desoukey, 2001), using different bacteria expression hosts (Vinetz et al., 1999), fusion with a solubility-enhancer tag (Davis et al., 1998), export of the protein into the periplasmic space (Southworth et al., 1996), and expression of truncated forms or independent domains of the molecule (Southworth et al., 1996). The glycosyl hydrolase domain of chitinases folds independently of the substrate-binding domain (Synstad et al., 2004; Fusetti et al., 2003), and both domains are independently stable and active (Henrissat and Bairoch, 1993, 1996; Arakane et al., 2003) (<http://afmb.cnrs-mrsfr/CAZY>). In this perspective, studies on chitinase activity and inhibitor binding could be made with the N-terminal glycosyl hydrolase domain of the molecule. For this reason, the N-terminal glycosyl hydrolase domain of *A. viteae* chitinase was cloned into the vector pET 22 b (+) and expressed in BL 21 (DE3) *E. coli* cells. Most of the protein was insoluble and could be purified only under denaturing conditions. There was a marginal increase in protein solubility when expression was done at lower temperatures. Export into the periplasmic space produced little amounts of soluble material, which was 16 times more active compared to the cytoplasmic protein. A comparable 12-fold increase in the activity of a *B. malayi* MBP fusion chitinase was obtained upon export of the protein into the periplasmic space (Southworth et al., 1996; Venegas et al., 1996). The increased specific activity of chitinase exported into the periplasma could be attributed to the

better folding conditions in the periplasm as opposed to the cytoplasm (Rietsch et al., 1996; Raina and Missiakas, 1997; Sone et al., 1997).

Since most of the chitinase was expressed as insoluble material in the cytoplasm, bacteria hosts that enhance protein solubility in the cytoplasm were used for expression. Bacteria strains like Origami B (DE3) carry the *trxB* and *gor* mutations that enhance the formation of disulfide bonds in the cytoplasm (Derman et al., 1993; Aslund et al., 1999) and increase protein solubility. The use of this strain led to a 4-fold increase in chitinase solubility and a 13-fold increase in activity as compared to chitinase purified under denaturing conditions.

In order to obtain much more active *A. viteae* chitinase, current efforts are directed towards a eukaryotic expression system like insect cells. The reasons for this are two-fold. Firstly, glycosylation may lead to an increase in filarial chitinase activity (Fuhrman et al., 1992; 1995). Secondly, a commercially available eukaryotically expressed *O. volvulus* chitinase was shown to be 4-fold active compared to the *A. viteae* chitinase in this study. The eukaryotic expression system offers better folding conditions and the possibility of post-synthetic modifications, which are absent in the prokaryotic expression system (Vialard et al., 1995).

3.3 Immunological control

Several independent observations and experiments hint to the development of protective immunity against filariae. In natural endemic populations, there are putatively immune (PIs) individuals (Elson et al., 1994; Turaga et al., 2000) with no current or past evidence of filarial infections. In natural and surrogate model systems for filariasis, immunisation with irradiated infective stage larvae led to near-sterile immunity (Lucius et al., 1991; Prince et al., 1992; Lange et al., 1993; Taylor et al., 1994; Johnson et al., 1998; Trees et al., 2000). However, the expression of identified filarial vaccine candidates as *E. coli* expressed recombinant proteins or as DNA vaccines in various models led to varying rates of protection that did not match up to the almost complete protection rates achieved by the use of irradiated third-stage larvae (Lustigman et al., 2002; Cook et al., 2001; Abraham et al., 2002). This failure to mimic vaccination with attenuated parasites can be due amongst others to the absence of non-protein moieties, like in phosphorylcholine and carbohydrates in the recombinant vaccines (Nutman, 2002). In addition, native-protective antigens may have a different tertiary structure and thus a greater capacity to induce protective

immunity as opposed to recombinant proteins. This is consistent with observations in the cestodes for which the initial use of native vaccine antigens has led to the development of successful vaccines (Lightowers et al., 2003). The paucity of starting material however limits such an approach in the filarial nematodes. Thus the use of heterologous nematode protein expression systems may lead to the faithful expression of native proteins.

An understanding of the protective mechanisms in PIs and the irradiated L3 vaccination models will help in targeted development of potential vaccines. The protective mechanisms in PIs is not clearly associated with any of the arms of immunity: antibody responses (Stewart et al., 1995; Boyer et al., 1991) as well as Th1 and Th2 responses (Turaga et al., 2000) have been described. On the other hand, protective immunity associated with vaccination using irradiated larvae was clearly dependent on Th2 responses and involved antibodies and eosinophils (Le Goff et al., 2000; Lange et al., 1994; Bleiss et al., 2002). A role for antibodies in natural systems is provided by the elevated levels of IgG1/ IgG3 in protected subjects (Stewart et al., 1995) and the demonstration that sera from both infected and protected subjects mediate adherence and killing of microfilariae (Brattig et al., 1991) and infective larvae (Johnson et al., 1994).

Filarial chitinases could be considered as targets for immune attack based on two observations: the sera of PIs differentially recognise filarial chitinase molecules as opposed to clinically infected individuals (Rhagavan et al., 1994); and the sera of rodents immunised with irradiated *A. viteae* L3s dominantly recognised L3 chitinase in blots (Adam et al., 1996). Moreover, a monoclonal antibody, that mediated microfilarial clearance upon transfer, recognised *B. malayi* microfilaria chitinases in western blot (Canlas et al., 1984a; Canlas et al., 1984b).

In addition to these observations, confirmatory vaccination studies were done with filarial chitinases from *O. volvulus* leading to a 53% significant reduction in worm burden (Harrison et al., 1999).

It may therefore be surmised that eliciting the right antibody response against chitinase would contribute to immune protection.

The catalytic and substrate binding domains of chitinase have been shown to have different immunological properties (Venegas et al., 1996; Wu et al., 2001). We hypothesized that an increased antibody response to the glycosyl hydrolase domain and/or the active site of *A. viteae* chitinase will lead to the selective inhibition of chitinase activity and function, and an inhibition in the development of stages associated with this enzyme. The use of antibodies to epitopes or peptides from active sites of enzymes has been shown to be a successful tool for vaccination in cases where the enzyme appears at critical points in the life cycle of a parasite. Passive transfer of monoclonal antibodies binding to the active site of glutathione S-transferase of the nematode *Schistosoma bovis* impaired egg development, and this property was in turn correlated with the enzyme inhibition (Da Costa et al., 1999). Using a similar strategy, it has been shown that vaccination of rabbits with synthetic peptides from the active site of *Helicobacter pylori* urease produced neutralising antibodies, and was a useful vaccination tool (Hirota et al., 2001). Synthetic peptides can be obtained from active sites of enzymes by homology modelling using other well characterised enzymes (Fujii et al., 2004).

The N-terminal catalytic domain of chitinase, as well as synthetic peptides designed from the active site of *A. viteae* chitinase 3-D model, was used in immunisation studies. Immunisation with the whole catalytic domain with STP as adjuvant led to a significant reduction in worm burden in a unique experiment. When alum was used as adjuvant, there was no tendency towards reduction in worm burden.

Despite the fact that STP was shown to be the better adjuvant for immunisation, alum was used for immunisation studies involving synthetic peptides, since our premise was to induce neutralising antibodies to the active site of chitinase. Alum is an adjuvant that directs the immune response to the Th2 direction /humoral arm. Immunisation with the synthetic peptides showed a tendency towards an overall decrease in worm burden in two independent experiments. In these same experiments, the group vaccinated with peptide 1 had a consistent significant reduction in microfilaria load, while the group vaccinated with peptide 2 had a significant reduction in microfilaria load in just one experiment.

Immunisation experiments were done just once with recombinant protein, and would therefore have to be repeated to confirm the trend observed. On the other hand, two independent vaccination studies were done using the synthetic peptides. The trend towards reduction in worm / mf burden was inconsistent when both peptides were

used. It can therefore not be definitely concluded that vaccination with either synthetic peptides or recombinant protein conferred protection to challenge infection. Inconsistencies in the protective potential of highly immunogenic filarial proteins have been observed before (Peralta et al., 1999). In their study, Peralta and colleagues used differential screening with sera of a diverse group of filariasis patients to isolate highly immunogenic proteins from a *B. malayi* expression library. However, independent vaccination experiments with the same proteins produced inconsistent protection results (Peralta et al., 1999).

Vaccination with peptide 1 could consistently reduce the microfilaria burden. Vaccination with this peptide may interfere with hatching of eggs to microfilaria or the development and moulting of microfilaria and L3. Such a hypothesis will be consistent with the finding that chitinase is present on the cuticle of post-infective L3 of unsheathed filarial (Harrison et al., 1999) and on the surface of uterine microfilaria (Adam et al., 1996). If chitinase has a role in the the development of filarial worms, any process that interferes with enzyme activity could also distort worm development. Such a case could be expected by the production of antibodies to the active site of the enzyme by vaccination with synthetic peptides. Thus, these molecules may therefore have a role in the development of uterine microfilariae and post-infective L3 which is distorted by vaccination with chitinase. The possible mechanism for reduction in microfilaria burden is not clear, and should be understood by demonstrating the role of antibodies in the observed tendency towards reduction in worm burden.

3.4 Outlook

The prospects for the future are three-fold: determination of the biological function of *A. viteae* chitinase, and its possible influence on host physiology; development of inhibitors to worm chitinases, and use of different immunisation approaches.

The function of *A. viteae* chitinase is not well defined. Therefore, future studies would be aimed at determining the biological function and possible influence on the host physiology. Reverse genetic studies like RNAi would be used to clarify the function of this gene in *A. viteae*. Three chitinase genes were described in this study. Analysis demonstrated transcripts in microfilariae, L3 and L4 larvae of *A. viteae* for gene I only, but not for gene III. To verify whether functional transcripts could actually be

produced from gene III, an *in vitro* transcription / translation system would have to be used. In addition, other *A. viteae* stages will have to be screened for chitinase genes.

Soluble active *A. viteae* is required for the development of inhibitors to worm chitinases, and for crystallisation studies. The expression and purification of soluble active chitinase in prokaryotic systems was difficult since most of the protein was found in inclusion bodies. The expression of *A. viteae* chitinase in eukaryotic systems like yeast or insect cells would provide better alternatives. Moreover, the proteins will be post-synthetically modified in these systems. A comparison of the modification pattern with that of native *A. viteae* chitinase will illustrate the similarities in post-synthetic modification.

Vaccination with prokaryotically expressed *A. viteae* chitinase did not lead to significant reductions in worm burden. Since, native proteins have been shown to be better vaccines in cestodes (Lightowlers et al., 2003), *A. viteae* chitinase expressed in eukaryotic systems would be tested in immunisation experiments along side native and prokaryotically expressed *A. viteae* chitinase. This will give a clue as to whether the modified proteins could have better protective capacities as compared to non-modified proteins.

4 METHODS

4.1 Identification of recombinant cDNA and genomic clones from phage libraries by plaque hybridisation

4.1.1 Preparation of plating bacteria

Host bacteria for bacteriophages (XL 1 blue for lambda zap, and XL 1 blue MRA for lambda dash II) were grown overnight at 30 °C, 150 rpm in 100 ml LB containing 0.2% maltose. Cells were pelleted at 5000 rpm for 10 minutes at 4 °C and re-suspended in 20 ml 10 mM MgSO₄ to an OD₆₀₀ of 2 (about 1.6x10⁹ cells per ml). The cell suspension was incubated at 37 °C, 220 rpm for 1 hr, stored at 4 °C and used for a maximum of three days.

4.1.2 Plating of lambda phage

An appropriate volume of phage suspension was mixed with 100 μ l (for 88 mm Petri dishes) or 200 μ l plating bacteria (for 120 mm petri dishes) and incubated with shaking (100 rpm) for 15 minutes at 37 °C. The suspension was then mixed with 4 or 8 ml of molten Top agar (50 °C) and carefully spread on agar plates. The plates were incubated for 30 minutes at room temperature to allow the top agar to harden, and were then incubated at 37 °C overnight. A uniform bacteria lawn could be observed interspersed with lambda plaques. The plates could be stored at 4 °C for several days.

4.1.3 Titration of phage libraries

To determine the concentration of a phage suspension in Plaque Forming Units (pfu) per ml, dilutions of a phage stock were made from 1:10 to 1:10,000 in SM buffer, and 1 μ l of each dilution was plated as described in section 4.1.2. Plates having between 50 to 350 plaque forming units were used to calculate the phage concentration in pfu.

4.1.4 Amplification and cryopreservation of phage stocks

Confluent plates of 3×10^4 pfu/plate were produced and incubated for a maximum of 8 hours at 37 °C. SM buffer was then added to the plates and the plates were incubated with constant shaking for 2 hours at 4 °C. The lysate was then aspirated and centrifuged for 5 minutes at 10,000 rpm to pellet residual agarose particles and the titre was determined as in section 4.1.3. DMSO was then added to 1 ml aliquots of bacteriophage stock at a concentration of 7% v/v. The tube was mixed gently, plunged into liquid nitrogen and stored at -80 °C until used.

4.2 Screening of libraries

4.2.1 Screening of libraries with a DIG-labelled *A. viteae* chitinase probe

4.2.1.1 Screening of libraries by plaque hybridisation

Up to 2×10^4 independent plaques were plated on a 120 mm petri dish and cultured for 8 to 12 hours. The plates were incubated at 4 °C for 30 minutes to allow the top agar to set. Nylon membranes were rinsed in water and then in 5x SSC and allowed to dry at room temperature. The membranes were carefully placed on the surface of the infected plate so that the entire membrane was in contact with the plate. After 30 minutes of incubation at 4 °C, the membranes were marked with a hot needle for orientation. Plates were again incubated at 4 °C for 30 minutes, after which the

membranes were removed. Replica filters were also used to lift plaques from infected plates.

DNA on the filters was denatured by incubation with 0.5 M NaOH/ 1.5 M NaCl for 15 minutes at room temperature, and subsequently neutralised twice by incubation with 0.5 M Tris, 1.5 M NaCl pH 8.0, after which the filters were allowed to dry on a Whatmann filter paper. DNA on the filters was immobilised by covalently linking the nucleic acids to the membrane by UV cross-linking. Alternatively, the membranes were baked at 80°C for 2 hours. An N-terminal cDNA sequence corresponding to nucleotides 52-1,117 of *A. viteae* L3 chitinase (Adam et al., 1996; Genbank number U14638) was labelled with digoxigenin (DIG) using the PCR labelling and detection kit from Roche. Hybridisation and detection was done as described by the manufacturer (Boehringer, Mannheim). Hybridisation was done with 15 ng/ml of probe in 10 ml of hybridisation buffer at a temperature of 50°C overnight. Stringent washes were done as follows: membranes were washed for 2 x 15 minutes in ample 2x SSC, 0.1% SDS at RT, and then in 0.5x SSC, 0.1% SDS (pre-warmed to wash temperature) for 2x 30 minutes at 65°C under constant agitation. Chemiluminescent detection was done using CSPD (Boehringer Mannheim).

Agarose plugs containing putatively positive plaques were incubated in SM buffer overnight at 4°C to let the phage wander out. The phage suspensions were diluted 1:10 with H₂O and boiled for 5 mins, and 1 µl was used for PCR as in section 4.3.12.1. Primers were vector-specific (T3 or T7) and *A. viteae* chitinase-specific.

To obtain clonal recombinant phages, phage suspensions identified as positive from the primary and PCR screening rounds were subjected to three further rounds of plaque hybridisation. In order to increase the chances of picking single/ independent plaques, 100-fold less independent plaques were used per new screening round as the proportion of positive plaques increased.

4.2.1.2 *In vivo excision*

The cDNA libraries used in this study were constructed in the vector Lambda UniZap XR (Stratagene). This vector is designed to allow *in vivo* excision and recircularization to form a pBluescript phagemid containing the cloned insert. An ExAssist helper phage contains a mutation that prevents replication of the phage genome in the non suppressing *E. coli* host (SOLR) and thus allows only the excised phagemid to replicate (Short et al., 1988). *In vivo* excision was done according to the

manufacturer's instructions. Briefly, 5-ml cultures of XL 1 Blue MRF['] and SOLR cells were grown at 30°C overnight in LB broth. The cells were pelleted and resuspended in 2.5 ml 10 mM MgSO₄ to an OD₆₀₀ of 1.0 (8x10⁸ cells/ml). An excision mix was made of 200 µl of XL 1 bLue MRF['] cells, 250 µl phage stock (1x10⁵ phage particles) and 1 µl ExAssist helper phage (titer: 1 x10⁷ pfu/ml). The mixture was incubated for 3 hours at 37°C with shaking. During this period the excision reaction takes place leading to the formation of pBluescript phagemid packaged as filamentous particles. The tube was heated at 70°C for 20 minutes to lyse the lambda phage particles and cells. Following centrifugation (1000xg for 15 minutes), the supernatant containing the excised phage particles was then stored. To plate the cells, 100 µl of phage supernatant was added to 200 µl SOLR cells (OD₆₀₀=1) and incubated at 37°C for 15 minutes. The cells were then plated on LB-ampicillin plates and incubated overnight at 37°C. Bacteria colonies were isolated containing the pBluescript double-stranded phagemid with the cloned cDNA insert.

4.3 Purification and manipulation of DNA

4.3.1 Isolation of lambda DNA and sub-cloning of genomic inserts

DNA was typically purified from clonal plaques and used for restriction digestion analysis, PCR, Southern and subcloning. The genomic libraries used in this study were constructed in lambda dash II (Stratagene). This vector, unlike lambda UniZap, can not be *in vivo* excised. To circumvent this short-coming, the inserts were released by digestion and cloned into plasmids.

4.3.1.1 Preparation of crude phage lysates

Confluent plates of 3 x 10⁴ pfu/plate were produced and incubated for a maximum of 8 hours at 37°C. SM buffer (10 ml) was then added to the plates and the plates were incubated with constant shaking for 2 hours at 4°C. The lysate was aspirated and centrifuged for 5 minutes at 10, 000 rpm to pellet agarose particles.

4.3.1.2 Isolation of phage DNA

DEAE cellulose columns were prepared and used for isolation of DNA. To prepare these columns, hydrochloric acid (0.05N) was slowly added to DE52 powder (Whartmann DE52 number 4057-050) with continuous stirring until a pH < 4.5 was attained. Concentrated NaOH was then added until the pH was 6.8. The resin was let to settle, the supernatant was aspirated and the ion exchanger was then washed 4 to

5 times with several volumes of LB-Medium. The slurry was adjusted to 60% resin, NaN_3 was added to a final concentration of 0.02% and the slurry was stored at 4°C for up to three months. Columns were prepared by pouring 8 ml DEAE slurry in a 10-ml disposable syringe resulting in a bed height of 6 cm. After the DEAE-cellulose had settled, the column was washed with 5 ml LB and subsequently equilibrated in LB.

The supernatant from two plates was loaded onto a 6-8 ml DEAE column. During this chromatographic step, bacterial DNA and RNA are separated through ion-exchange chromatography; the ion-exchange resin preferentially binds contaminants in the lysate such as *E. coli* DNA, RNA and proteins. The eluate containing the phage particles were then collected and pooled. The column was washed again with 2 ml LB-Medium and the eluate added to the first. The NaCl concentration of the eluate was adjusted to 70 mM and phages were precipitated with 2 volumes of ethanol (20 minutes at -20°C, and then centrifuged for 15 minutes at 12,000 rpm). The pellet was then washed with 2 volumes of 70% ethanol and subsequently dissolved in 2 ml TE with 0.2% SDS to release the phage DNA. The mixture was extracted twice with 2 volumes of Phenol (TE saturated). DNA was precipitated from the aqueous phase with 2 volumes of ethanol without any salt. The phage DNA pellet was then washed twice with 70% ethanol and dissolved in 200 μl TlowE. The yield was about 75 μg per 10 ml phage lysate.

4.3.1.3 Release and sub-cloning of genomic inserts into pBluescript II SK

Purified recombinant lambda DNA and pBluescript II SK (-) were separately digested with Not I and dephosphorylated as described in sections 4.3.8 and 4.3.9. Genomic inserts ranging in size between 10-14 kb were then ligated into pBluescript vector (3kb) using a 1:1 molar ratio of insert and vector (see section 4.3.10.2).

4.3.2 Small scale Plasmid isolation from *E. coli*

Plasmid DNA was isolated as described by Del Sal et al., 1989, with some slight modifications. This technique relies on the property of the detergent cetyl-trimethyl ammonium bromide (CTAB) of preferentially precipitating nucleic acids at low NaCl concentrations (< 0.6 M), while leaving proteins and polysaccharides in solution.

Essentially, a single bacterial colony harbouring a plasmid of interest was used to inoculate 6 ml of LB medium (with an appropriate antibiotic for selection) and grown over night at 37°C. Bacterial cells were recovered by centrifugation at 4,000 rpm for

10 minutes. The pellet was re-suspended in 200µl of STET buffer, 4µl of a 50-mg/ml lysozyme solution was added and the samples were incubated at room temperature for 5 minutes. Samples were boiled for 45 seconds and then centrifuged at 13,000 rpm for 10 minutes in an Eppendorf centrifuge. Pellets were recovered, 5 µl of RNase A (10 mg/ml) were added to the supernatants and the samples were incubated for 10 minutes at 68°C. Nucleic acids were precipitated by the addition of 10 µl of CTAB (5% w/v), followed by centrifugation at 14,000 rpm for 5 minutes. Pellets were re-suspended in 300 µl of 1.2 M NaCl and the plasmids were re-precipitated by the addition of 750 µl ethanol and incubation at - 20°C for 1 hour. Samples were then centrifuged at 14,000 rpm for 15 minutes and the pellets were washed twice in 70% ethanol and allowed to air dry. Plasmids were re-suspended in 40 µl H₂O. The yield was between 20 to 40 µg of plasmid DNA.

4.3.3 Electrophoresis and detection of DNA on agarose gels

Agarose gel electrophoresis was used for the routine analysis of DNA. Agarose was cast in 1x TAE buffer containing 0.5 µg/ml EtBr. The DNA samples (0.1 to 5 µg) were dissolved in DNA loading buffer and separated in agarose gels at 10 volts per cm. The concentration of the agarose gel was relative to the size of DNA fragments to be separated, and was typically between 0.7 and 1.2%.

4.3.4 Isolation of DNA from agarose gels

Following electrophoresis, a DNA fragment was excised from agarose gels using a clean scalpel, and the DNA was purified using the NucleoSpin® Extraction Kit (Clontech) according to the manufacturer's instruction.

4.3.5 Isolation and concentration of DNA from aqueous solutions

4.3.5.1 *Extraction with Nucleospin kit*

The NucleoSpin DNA Extraction Kit (Clontech) was used according to the manufacturer's instruction for the isolation of DNA from PCR reactions, gel fragments and other aqueous DNA solutions.

4.3.5.2 *Phenol chloroform extraction*

To free DNA from protein contaminants, the volume was adjusted to at least 300 µl with TE buffer, and extracted with an equal volume of Tris-equilibrated phenol:chloroform:isoamyl alcohol (25:24:1) (Sambrook et al., 1989). The aqueous

phase was extracted twice with chloroform to remove traces of phenol, and the DNA was concentrated by ethanol precipitation.

4.3.5.3 Precipitation of DNA

Ice cold sodium acetate, pH 4.5 was added to a DNA sample to a final concentration of 0.3 M, followed by the addition of 2.5 volumes of ice cold absolute ethanol. The sample was incubated at -70°C for 15 minutes and centrifuged at 12,000 rpm for 15 minutes at 4°C. The pellet was washed in 70% ethanol, air-dried, and re-suspended in an appropriate buffer.

4.3.6 Isolation of high molecular weight genomic DNA from *A. viteae*

About 200 mg to 1 g of adult *A. viteae* were snap-frozen in liquid nitrogen and ground to powder using a mortar and a pestle. Up to 100 mg of powdered worm material was suspended in 1.2 ml of digestion buffer containing Proteinase K (100 µg/ml), and incubated with shaking at 50°C for 12 to 18 hours until the sample became viscous with a visible sludge. The samples were extracted three times with an equal volume of phenol/chloroform/isoamyl alcohol. The viscous aqueous phase was transferred each time with a wide-bore pipette (0.3-cm-diameter orifice) into a new tube. RNA was digested by the addition of 20 µg/ml RNase A and incubating for 1 hour at 37°C. Genomic DNA was then precipitated by adding ½ volume of 7.5 M ammonium acetate and 2 (original) volumes of 100% ice-cold ethanol. Stringy precipitates of DNA were carefully transferred into new tubes using a blunt spatula, and washed twice with ample amounts of 70% ethanol (2 times the original volume). The DNA pellet was air dried and re-suspended in TE buffer until dissolved. The concentration of the gDNA was determined by photometry and the DNA was analysed on a gel.

4.3.7 Determination of DNA Concentration

The concentration of DNA in solution was determined using a spectrophotometer at OD₂₆₀, and subsequently confirmed by comparing the intensity and size of DNA from a sample to those of standard size markers with known concentrations.

1 unit of absorbance of dsDNA at OD₂₆₀ = 50 µg/ml dsDNA

1 unit of absorbance of ssDNA at OD₂₆₀ = 33 µg/ml ssDNA or RNA

The OD₂₈₀ was also measured and the ratio between the two ODs indicated the purity of the DNA solution. For pure DNA,

$$1.8 \leq \text{absorbance at OD}_{260} / \text{absorbance at OD}_{280} \leq 2.0$$

A value less than 1.8 indicates contamination with proteins or with aromatic substances like phenol, while a value greater than 2.0 indicates possible contamination with RNA.

4.3.8 Restriction digestion of DNA

DNA was digested at the optimal temperature of restriction enzymes according to the pipetting scheme below.

Tab. 12: Pipetting scheme for restriction digestion of DNA

Reagent	Volume (µl)
DNA (0.1 – 5 µg)	5
10 x restriction enzyme buffer	2
10 x Bovine Serum Albumin (BSA)	2
Restriction enzyme (3-20 units)	1
HPLC water	10
Volume	20

Following digestion, the mixture was either heated at 70°C for 15 minutes to inactivate the enzyme, or extracted with phenol chloroform.

4.3.9 5' Dephosphorylation of digested DNA

The removal of 5' phosphate terminals from digested dDNAs ends prevents vector self-ligation and reduces background in ligations, especially when only one restriction site is used. Shrimp alkaline phosphatase (SAP) (0.5 units) was added to the restriction digests and incubated for 15 minutes at 37°C. Enzyme inactivation was achieved by heating the mixture at 65°C for 15 minutes.

4.3.10 Ligation of DNA fragments

4.3.10.1 Ligation of PCR fragments into T-overhang vectors

PCR products were ligated into the pGEM-T vector (Promega, Madison, WI) according to the following protocol.

The reaction mixture was incubated overnight at 16°C. The ligation reaction was used to transform competent bacteria and single clones were picked for the isolation of recombinant plasmid DNA that was sequenced and its insert subcloned into an appropriate vector where necessary.

Tab. 13: Pipetting scheme for ligation of PCR fragments into T-overhang vectors

Reagent	Volume (μ l)
Vector (pGEM-T) DNA (50ng)	1
2X ligation buffer (Promega)	5
Insert DNA	3 (in three molar excess of the vector)
T4 DNA ligase	0.5
HPLC water	0.5
Reaction volume	10

4.3.10.2 Ligation of DNA fragments with sticky ends

Insert DNA was digested out of a vector using appropriate restriction enzymes, while the target vector was also digested with the same enzymes. Both were then ligated following the scheme below.

Tab. 14: Pipetting scheme for ligation of DNA fragments with sticky ends

Reagent	Volume (μ l)
Vector DNA (50ng)	1
10 x ligation buffer (New England Biolabs)	2
Insert DNA	3 (in three molar excess of the vector)
T4 DNA ligase	1
HPLC water	13
Reaction volume	20

4.3.11 mRNA isolation, reverse transcription and RT-PCR

Worms (1g) were homogenised with a plastic pestle in 1000 μ l of RNA isolation reagent (Pepqlab) according to the manufacturer's instruction, and total RNA was isolated by phenol chloroform extraction followed by ethanol washes. Total RNA was reverse transcribed by Moloney murine leukemia virus reverse transcriptase (Promega) in the presence of random hexamer primers.

4.3.12 Polymerase chain reaction (PCR)

Thermostable DNA polymerases were used for the amplification of DNA as described (Saiki et al., 1988, Bej et al., 1991). Two types of PCR amplifications were used in this study: standard PCR for the amplification of DNA fragments up to 3 kb and Mid-range PCR for the amplification of fragments up to 9 Kb in size.

4.3.12.1 Standard PCR

These were done in 50 μ l reaction volumes as shown in Table 15. Optimal annealing temperatures for primer pairs relative to a target template were obtained using the Oligo 4.0 software. PCR was performed on an MJ Research PT-200 DNA machine according to the thermal profile on Table 16.

Tab. 15: Pipetting scheme for standard PCR

Reagent	Volume (μ l)
Forward primer (10 pmol/ μ l)	1
Reverse primer (10 pmol/ μ l)	1
dNTP mix (40 mM)	1
DNA template	1 – 5 (between 100 to 500 ng)
10 X reaction buffer	5
HPLC water	ad 50
Taq polymerase (5 U/ μ l)	0.5

Tab. 16: Thermal profile for standard PCR

Phase	Temperature ($^{\circ}$ C)	Duration	Cycles
Denaturation	94	2 min	1
Annealing	53-60	40 s	
Elongation	72	1 min per 1 kb	
Denaturation	94	20 s	30
Annealing	53-60	40 s	
Elongation	72	1 min per 1 kb	
Denaturation	94	20 s	1
Annealing	53-60	40 s	
Elongation	72	15 min	
	4		

4.3.12.2 Mid-range PCR

This PCR technique was used for the amplification of genomic DNA fragments between 3 and 9 kb (Barnes et al., 1994; Cheng et al., 1994). These were done using the Peq Lab mid-range PCR system as follows:

Tab. 17: Pipetting scheme for Mid range PCR

Reagent	Volume (μ l)
Forward primer (10 pmol/ μ l)	2
Reverse primer (10 pmol/ μ l)	2
dNTP mix (40 mM)	1.75
DNA template	1 – 5 (between 100 to 500 ng)
10 X reaction buffer	5
HPLC water	ad 50
Mid-range polymerase (5 U/ μ l)	0.5

PCR was performed on an MJ Research PT-200 DNA machine as follows:

Tab. 18: Thermal profile for mid range PCR

Phase	Temperature ($^{\circ}$ C)	Duration	Cycles
Denaturation	94	2 min	1
Annealing	53-60	40 s	
Elongation	68	1 min per 1 kb	
Denaturation	94	20 s	30
Annealing	53-60	40 s	
Elongation	68	1.5 min per 1 kb	
Denaturation	94	20 s	1
Annealing	53-60	40 s	
Elongation	68	15 min	
	4		

4.4 Southern blotting

4.4.1 Digestion, electrophoresis, and blotting of genomic DNA

Genomic DNA (20 μ g) were digested with different restriction enzymes in 100 μ l reaction volumes as described in section 4.2.7, and electrophoresed in 0.7% agarose gels (section 4.2.2). The DNA was capillary-blotted onto a nylon membrane at room temperature in 20x SSC overnight (Sambrook et al., 1989). The membrane was then baked at 80 $^{\circ}$ C for 2 hours to fix the DNA onto the membrane.

4.4.2 Radioactive labeling of *A. viteae* chitinase probe

A. viteae chitinase PCR fragments (25 ng) were labelled with 50 μCi $\alpha\text{-}^{32}\text{P}$ dCTP using the RadPrime DNA labelling system (Life Technologies). The pipetting scheme was as follows:

Tab. 19: Pipetting scheme for radioactive labeling of probes

Item	Volume (μl)
<i>A. viteae</i> chitinase PCR fragment (25ng)	5
dATP (500 μM)	1
dGTP (500 μM)	1
dTTP (500 μM)	1
Random primers solution	20
$\alpha\text{-}^{32}\text{P}$ dCTP (3000 Ci/mmol, 10mCi/ml)	5
Distilled water	16
Klenow fragment	1

The mixture was incubated at 37°C for 10 minutes and the reaction stopped. The radioactive probes were purified using SephadexTM MicroSpin G50 columns (Amersham Biosciences Europe GmbH). The labelling efficiency was about 30%.

4.4.3 Hybridisation and detection

Membranes were washed in 6x SSC at room temperature and prehybridised at 65°C in 10 ml hybridisation buffer (6x SSC, 5x Denhardt's reagent, 0.5% SDS, 100 $\mu\text{g}/\text{ml}$ salmon-sperm DNA) for 1 hour. The probe was denatured by boiling for 2 min in a water bath and added to the hybridisation buffer. Membranes were hybridised overnight at 65°C in a roller bottle. Stringent washes were done in 2x SSC, 0.5% SDS for 5 minutes at room temperature, 2x SSC, 0.1% SDS for 15 minutes at room temperature, and 0.1% SSC, 0.1% SDS for 2 hours at 65°C. The membranes were rinsed in 0.1x SSC, covered in Saran Wrap, and exposed to a phosphorimager plate (Fuji Film) for 3 hours, and the bands were scanned using a phosphoimager.

4.5 Microbiological methods

4.5.1 Preparation of competent *E. coli*

Competent *E. coli* cells were prepared essentially as described by Inoue et al. (1990). *E. coli* were streaked on LB agar plates with antibiotics and cultured overnight at 37°C. Twelve large colonies were picked with a sterile toothpick and used to inoculate 125 ml SOB in a 1-liter Erlenmeyer flask. Flasks were incubated at 18°C with vigorous shaking (220 rpm) and the bacteria grown to an OD of 0.6 (mid-log phase). Bacteria cultures were poured into Falcon tubes and incubated on ice for 10 minutes and then centrifuged at 2500x g (3000 rpm) in an Eppendorf 5403 bench top centrifuge. The pellet was resuspended in 40 ml ice-cold TB buffer, incubated on ice for 10 minutes and centrifuged as above. The pellet was then carefully resuspended in 10 ml of TB, and DMSO added drop-wise to a final concentration of 7%. The bacterial suspension was incubated for ten minutes on ice, after which 1 ml aliquots were snap-frozen in liquid nitrogen and stored at – 80°C for up to one month.

4.5.2 Transformation of competent *E. coli*

Eppendorf tubes (1.5 ml) were pre-chilled on ice and 10 ng of purified plasmid DNA in a total volume of 20 µl were pipetted into the tubes. Competent cells were thawed, and 200 µl were dispensed into the tubes on ice. The tubes were flicked gently to mix and incubated on ice for 30 minutes. The cells were then heat-shocked by heating for exactly 45 seconds in a 42°C water bath, and then incubated on ice for 2 minutes. Room temperature SOC medium (800 µl) was added to each tube on ice. The tubes were then incubated at 37°C while shaking vigorously (220 rpm) for one hour. Antibiotic selective agar plates were plated with 50 to 150 µl of the transformation mixture and incubated overnight at 37°C. Positive controls were competent cells transformed with 10 ng pBluescript SK (-) plasmid, while negative controls were competent cells transformed without DNA.

4.5.3 Screening of bacterial colonies for plasmids/ recombinant plasmids

Two methods were routinely employed to identify bacteria colonies that contain the recombinant plasmids or plasmids of interest.

4.5.3.1 Blue-white screening

Most bacteria strains (e.g. JM109, DH5 alpha and XL 1 Blue), used as hosts for initial cloning of a target gene, code for an inactive carboxy-terminal portion of β-

galactosidase due to a mutation in the lac Z gene (lacZdeltaM15). On the other hand, many vectors contain the regulatory sequences and information that could be induced (e.g. by IPTG) to code for the missing amino terminal (alpha region) of the β -galactosidase gene. Both host and plasmid encoded fragments are inactive. However, if a bacterial host is transformed with a plasmid expressing the missing amino terminal of the enzyme, both fragments associate to form an active enzyme in a process called alpha complementation. Such bacteria can be recognised because they metabolise the chromogenic substrate 5-bromo-4-chloro-3-indolyl- β -D-galactoside (X-gal) in the presence of isopropyl- β -D galactopyranoside (IPTG) to form blue colonies. Insertion of a target gene in the alpha peptide region coding for the β -galactosidase enzyme results to insertional inactivation of the alpha peptide. Bacteria carrying plasmids with the target gene will form white colonies since they lack active β -galactosidase. Thus, transformed bacteria were plated on LB-agar plates supplemented with 0.5 mM IPTG, 80 μ g/ml X-gal and antibiotics.

4.5.3.2 Restriction analysis of isolated plasmids

In order to verify constructs of plasmids/ inserts resulting from cloning, plasmid DNA was isolated and analysed. Independently transformed bacterial colonies were picked and grown in 5 ml LB/ antibiotic overnight. Plasmid DNA was isolated (section 4.3.2) and digested (section 4.3.8) at restriction sites used for cloning of the insert DNA. The digested DNA was then analysed by agarose gel electrophoresis (Section 4.3.3).

4.5.4 Bacteria cultures and long-term storage of bacterial stocks

Bacteria host strains used in this study included XL 1 Blue, BL 21 (DE3), and Origami B (DE3). Bacteria were streaked on LB agar plates and grown overnight at 37°C. For long-term storage of bacteria, a single colony was grown to an OD₆₀₀ of 0.6 and 1 part was mixed with 9 parts of Hogness freezing medium (36 mM K₂HPO₄, 13 mM KH₂PO₄, 20 mM sodium citrate, 10 mM MgSO₄ and 40 % glycerol). Tubes with bacteria stocks were snap-frozen in liquid nitrogen and stored at -80°C. Bacteria recovery involved scraping the surface of a bacteria stock with a sterile loop and streaking an LB plate with the appropriate antibiotics.

4.5.5 Expression and purification of recombinant proteins from *E. coli*

The pET expression system was used for prokaryotic expression of recombinant N-terminal *A. viteae* chitinase. In this system, target genes are cloned under the control

of the T7 promoter which is not recognised by *E. coli* RNA polymerase. Some *E. coli* host strains (like BL21 (DE3) and Origami B (DE3)) have a chromosomal copy of the T7 RNA polymerase gene under control of *lacUV5*. Transfer of recombinant pET plasmids into such hosts results in an IPTG-inducible gene expression system. The cDNA encoding the N-terminal domain of chitinase was cloned into the pET 22 b (+) vector and transformed into Origami B (DE3 cells). The pET 22 b (+) vector has a *pelB* signal sequence that leads to the localisation of an expressed protein into the periplasmic space of the *E. coli* host.

A single bacterial colony was picked and used to inoculate LB (100 µg/ml Ampicillin, 12.5 µg/ml Tetracyclin, 15 µg/ml Kanamycin and 1% glucose) and the culture was grown overnight at 37°C, 150 rpm. The overnight culture was diluted 1:50 in fresh LB medium and further grown at 37°C till an OD₆₀₀ of 0.4. The culture was shifted to room temperature and further grown to an OD₆₀₀ of 0.6. Protein expression was induced with 1mM IPTG for 4 hours. Bacteria were pelleted by centrifugation at 6000 rpm for 15 minutes at 4°C. The pellet was then used for the extraction and purification of periplasmatic proteins and proteins in inclusion bodies.

For extraction of periplasmatic fluid, bacteria pellet from a 500 ml culture was resuspended in 30 ml 30mM Tris-HCl pH 8, 20% sucrose. EDTA was added to a final concentration of 1 mM and the suspension stirred for 10 minutes at room temperature. Cells were collected by centrifugation at 10,000xg at 4°C for 10 minutes. The pellet was then re-suspended in 30 ml of ice-cold 5 mM MgSO₄ and stirred slowly on ice for 10 minutes to release the periplasmatic proteins. The suspension was centrifuged at 10,000xg to pellet the cells and the supernatant was used for purification of periplasmatically expressed *A. viteae* chitinase. The insoluble fraction in inclusion bodies was purified as described below.

The pellet was resuspended in lysis buffer A, 100 µg/ml lysozyme and incubated for 30 minutes at 4°C. The suspension was then sonicated for 1.5 min, 30 cycles. After centrifugation (13,000 rpm, 20 min, 4°C), the supernatant was transferred into a falcon tube, incubated with 1 ml Nickel chelate matrix (Qiagen, Hilden) and incubated for 1hr at room temperature with constant gentle shaking. The matrix with bound protein was then loaded onto a column and washes were done with 20 ml of buffers B and C, respectively. Elution was done in 20 ml of buffer E, and collected in 1ml fractions. The fractions were analysed by SDS-PAGE for protein and the positive

fractions were pooled. This pool was then dialysed in phosphate buffers with reducing amounts of urea and the protein concentration was determined. Aliquots of the protein were stored at -20 °C until used.

4.6 Protein analytical methods

4.6.1 Determination of protein concentration

Protein concentration was determined by the BCA test using the BCA kit (Pierce, Rockford, USA). The basis of this reaction is the biuret reaction: reduction of Cu^{2+} to Cu^{1+} by a peptide bond under alkaline conditions. Chelation of two molecules of bicinchoninic acid (BCA) by the cuprous ion (Cu^{1+}) produces a water soluble complex, whose solution has a deep purple colour and an absorbance at 562 nm. A linear standard curve was made with 0.05 to 2mg BSA.

4.6.2 Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE)

Analytical polyacrylamide gel electrophoresis in sodium dodecyl sulfate was performed as described by Laemmli (1970). Samples were heated for 5 minutes at 100°C in sample buffer containing 125 mM Tris-HCL, pH 8.3, 1% SDS, 2.5% β -mercaptoethanol and 10% glycerol. Samples were applied at volumes of 10-30 μl to a 12.5% slab gel cast in a mini-gel apparatus (Hoefer). The electrophoresis was performed at 20mA with electrode buffer (25mM Tris-HCL, pH 8.3, 192mM Glycine and 0.1% SDS). The run was stopped when the bromophenol blue dye front reached the end of the gel. Protein bands were then revealed by staining with Coomassie Brilliant Blue R250, and the gels dried for a permanent record.

4.6.3 Chitinase activity assays

Chitinase assays were performed using 4-methylumbelliferyl β -N, N', N''-triacetylchitotrioside (GlcNAc)₃UMB. (GlcNAc)₃UMB was dissolved in reaction buffer (20mM NaPO_4 , pH 6.0, 200mM NaCl, 1mM EDTA) and 95 μl of 20 μM substrate solution were mixed with 5 μl of enzyme and incubated for 2 to 10 minutes at 25°C . The reactions were stopped by adding 1.9 ml of 0.5 M glycine, pH 10.5. Release of free methylumbelliferone was measured by a fluorescence spectrophotometer at 360 nm excitation and 450 nm emission wavelengths. The fluorometer was scaled at the beginning of the experiments such that 20 nM substrate had a fluorescence reading of 10 units.

4.7 Immunochemical and immunological methods

4.7.1 Western blot

4.7.1.1 *Transfer of proteins onto nitrocellulose membranes*

Proteins separated by SDS-PAGE were immobilised onto nitrocellulose membranes by electrophoretic transfers (Towbin et al., 1979). A transfer cassette was assembled in transfer buffer using a sponge followed progressively by a 3 MM- Whatmann paper, nitrocellulose membrane (pore size 0.2 μM), SDS-PAGE gel, another Whatmann paper and finally a second sponge. The cassette was introduced into a transfer tank between electrodes so that the gel with the protein was towards the negatively charged cathode and the nitrocellulose membrane towards the positively charged anode. A constant current of 70 mA was then applied across the cassette at room temperature overnight. To determine the transfer efficiency, the membrane was reversibly stained in Ponceau S dye (0.1% Ponceau S, 7% trichloroacetic acid in distilled water). After complete washing of the Ponceau stain, unbound sites of the membrane were blocked by incubation for 45 minutes in 5% skimmed milk powder in PBS.

4.7.1.2 *Immunodetection of immobilised proteins*

Nitrocellulose membranes with immobilised proteins were incubated with a primary protein specific antibody for one hour at room temperature, followed by 3 x 5 minutes washes in wash buffer (PBS, 0.02% Tween 20). The membrane was then incubated in a secondary conjugate antibody for one hour followed by washes as above. Detection was done by using a substrate for the alkaline phosphatase enzyme conjugate, 5-Brom-4-chlor-3-indolylphosphate (BCIP) and tetrazolium chloride (NBT) in alkaline phosphatase buffer. All antibodies were diluted in blocking solution (5% skimmed milk powder in PBS).

4.7.2 Coupling of peptides to KLH

Synthetic peptides (20 amino acids long) were synthesised with two extra N-terminal cysteins and coupled to KLH (Keyhole limpet haemocyanin) to make them immunogenic. Coupling was done using the Imject maleimide activated mcKLH kit (Pierce USA) and the coupled proteins were used at a concentration of 100 μg in vaccinations.

4.7.3 Immunisation studies

The protective potential of *A. viteae* N-terminal chitinase was evaluated in the *Meriones/A. viteae* natural host-parasite system. Eight to ten-week-old *Meriones* were anaesthetized with ketamin:xylazin:normal saline (1:1:8) and immunised with 25 µg recombinant *A. viteae* chitinase or 100 µg KLH-coupled synthetic *A. viteae* chitinase peptide in adjuvant. The animals were immunised three times in a two-weekly interval, after which they were challenged with 70 freshly isolated L3s by injecting in the neck region. Microfilaria load was verified at weeks 4, 8 and 12 post infection (pi), and the animals were dissected at week 12 pi for isolation of adult worms and determination of protective potential.

Two adjuvants were used for immunisation, depending on the immune reaction desired: Alum was used for humoral-mediated immune reactions, while STP was used for cell-mediated immune responses.

4.7.3.1 Bleeding of animals for production of sera

Mice and jirds were bled from the retro-orbital sinus using a microhaematocrit capillary, and the blood was stored at 4°C overnight. Blood samples were then centrifuged (10,000 rpm, 20 minutes, 25°C) to separate the sera from blot clots. Sera were stored in 100 µl aliquots at -20°C.

4.8 Parasitological methods

4.8.1 Maintenance of the life cycle of *A. viteae*

The life cycle of *A. viteae* was maintained in *Ornithodoros moubata* essentially as described by Lucius and Textor, 1995.

4.8.2 Quantification of microfilarial load in blood of jirds

Infected *Meriones* were anaesthetized with ketamin:xylazin:normal saline (1:1:8) and bled from the retro-orbital sinus using a heparinised glass capillary tube. 20 µl blood were mixed with 100µl Teepol (10% in H₂O), and the microfilaria load was counted using a Fuchs-Rosenthal counting chamber.

4.8.3 Isolation of filariae

4.8.3.1 Isolation of adult *A. viteae*

Meriones were anaesthetized and fully bled from the retro-orbital sinus. Following dissection of the jirds, adult *A. viteae* were isolated from the subcutaneous and

intramuscular tissues, the iunguinal and subscapular regions and some times in the thoracic chamber. Animal carcasses were then incubated in normal saline (0.9% NaCl) overnight to allow the rest of the worms to wander out.

4.8.3.2 Isolation of L3 from the vector *Ornithodoros moubata*

L3s were isolated using a Baermann apparatus comprising of a funnel with a tap. The inner part of the funnel was covered with bandage and filled with Ringer's solution. Ticks were cut medially and briefly rinsed in a Petri dish with Ringer's solution to remove rests of blood meal and loose tissue. The ticks were incubated in warm Ringer's solution in the funnels for 1 hr for the L3s to migrate into the solution. The tap was then opened to collect the L3s in 50 ml falcon tubes. The larvae were amply washed in Ringer's solution and rinsed in RPMI if they were to be used for culture.

4.8.3.3 Isolation of uterine microfilariae from gravid female worms

Adult female *A. viteae* worms were isolated on day 90 post-infection (average length, 6 cm; embryogenesis starts on day 28 post-infection) and cut into 3mm segments in Ringer's solution. Developmental stages were separated from the worm segments by filtration. The stages obtained included: oocytes attached to the rachis of the ovary, fertilized eggs, multicellular stages, invaginated stages, pretzel and ring stages, and elongated microfilariae.

4.9 Computer analysis and statistical methods

4.9.1 Analysis of DNA sequences

Genomic DNA sequences obtained from primer walking were assembled into contigs by using the program MacVector 7.2 (Accelrys, USA). Assembled sequences were fed into Artemis[®] (www.sanger.ac.uk/Software/Artemis/) for visualisation of sequence features, its six-frame translations and exon-intron prediction. Exons thus predicted were verified by comparison to a cDNA sequence and by further analysis using Netgene (www.cbs.dtu.dk/services/NetGene2/). *Brugia malayi* genomic chitinase DNA sequences were retrieved from the server for the *B. malayi* genome project at <http://www.tigr.org/tdb/e2k1/bma1/> while EST sequence analysis was done at the server of the Filaria genome project (<http://nema.cap.ed.ac.uk/fgp.html>).

4.9.2 Statistical analysis

The worms recovered from immunised and non-immunised *Meriones* in immunisation experiments were compared using the Mann-Whitney U-test at the 95% significance level ($p < 0.05$).

5 Materials

5.1 Commercial Kits and Enzymes

Collagenase A	Biochrom, Berlin
RNase A	Boehringer Mannheim, Mannheim
T4 DNA ligase	New England Biolabs, USA
DNA restriction enzymes	New England Biolabs, USA
Mid-range Polymerase	PeqLab, Erlangen
Sawady Taq Polymerase	PeqLab, Erlangen
Lysosyme	Sigma, Deisenhofen
Qiagen protein purification kit	Qiagen, Hilden
Nucleospin DNA extraction kit	Clontech, Heidelberg
Nucleospin plasmid prep kit	Clontech, Heidelberg
BCA protein assay Kit	Pierce, USA
KLH protein conjugation kit	Pierce, USA
PGEM-T™ Cloning kit	Promega, Madison, USA
Random Primers DNA labeling System	Life Technologies, Karlsruhe
Nick Translation System	Life Technologies, Karlsruhe

5.2 Laboratory Equipment and consumables

Chromatographic columns	Pharmacia, Freiburg
Agarose gel electrophoresis apparatus	AGS, Heidelberg
SDS-PAGE apparatus	Pharmacia, Biotech
Video documentation apparatus E.A.S.Y.RH	Herolab, Wiesloch
Peltier Thermal Cyclers, PTC-200	MJ-Research, Massachusetts
Bandelin Sonoplus HD 200 sonicator	Berlin
Spectrophotometer	Eppendorf
DE52 powder (DE52 number 4057-050)	Whatman, USA
Chromatography paper (3MM)	Whatman, USA
Dialysis membrane	Serva, Heidelberg
Nitrocellulose membrane (0.2 µm)	Schleicher & Schuell
Cell culture plates (Flat & round bottom)	Costar, Bodenheim

HPLC water

Roth, Karlsruhe

CSPD

Boehringer; Mannheim

5.3 Synthetic oligonucleotides

Oligonucleotides were delivered as lyophilised powders and dissolved to final concentrations of 100 pmol/ μ l in water as per manufacturer's instruction. This stock solution was then dissolved 1:10 in water to obtain working solutions with concentrations of 10 pmol/ μ l. A total of 10 pmol primer was used in standard PCRs, while 20 pmol were used in long range PCR.

5.3.1 PCR Primers

5.3.1.1 Primers used in long range PCR

Name	Sense	Clone	Sequence
T7- long		all	GTAATA CGA CTC ACT ATA GGG C
T3-long		all	AAT TAA CCC TCA CTA AAG GGA AC
F3	+	all	CTACGTTTCGCGGATGTTAC
F4	+	all	TGGGCTTGAAAGTGAGGTAAG
R4	-	all	TGTTTGCTCACTTTCAAGCCC
R7	-	12	TCCCAACTGCCGTGTAAATCA
R8	-	12	CATTTCCGACAGTAATACGAT
R5	-	1	CCGGCAGAAATACAATGCTTG

5.3.1.2 Primers used for standard PCR

5.3.2 Sequencing primers

5.3.2.1 *Universal sequencing primers*

5.3.2.1.1 Primers on pGEM-T vector

Name	Sense	Sequence
SP6	-	ATTTAG GTG ACA CTA TAG
T7	+	TAA TAC GAC TCA CTA TAG GG

5.3.2.2 *Primers on pBluescript SK (-) vector*

Name	Sense	Position	Sequence
T3	-	772-791	ATTAACCCTCACTAAAGGGA
T7	+	626-645	TAATACGACTCACTATAGGG
M13 forward	+	603-620	GTA AAACGACGGCCAGT
M13 reverse	-	811-828	CAGGAAACAGCTATGAC

5.3.3 Primers used for sequencing *A. viteae* chitinase genomic clone 1

Name	Sense	Position	Sequence
F3	+	5679-5698	CTACGTTGCGGGATGTTAC
F3walk	+	6107-6127	ACACCTATGCACTCACATTC
F4	+	8698-8719	TGGGCTTGAAAGTGAGGTAAG
F31	+	8075-8094	TCGCCATCGACAACGAATC
F5	+	8653-8674	AAGTTGTGGCAAAGGTCCATA
F6	+	6521-6542	CATCAGTTTTTGCTGTTGCTT
F7	+	8852-8873	GGACCATACGGAAAAATGATT
F7walk	+	9208-9229	CAGATAATACTGAAATGCCAG
R4	-	8698-8709	TGTTTGCTCACTTTCAAGCCC
R5	-	6025-6046	CCGGCAGAAATACAATGCTTG
R5walk	-	5473-5493	CAATAGATGCTACTCGACAG
R6	-	4837-4858	GGAATTCAATGAGTTATGCTG
R6walk	-	4280-4301	GACGTATAACACAAAGTACG
R7	-	7772-7792	TCCCAACTGCCGTGTAAATCA
M13 forward	-	vector	GTA AAA CGA CGG CCA GT
R3n1	-	12355-12376	TCAGGCGCTAAGAAAAGAGAG
R3n1walk	-	11550-11573	TAAGGCTCATTAACTTCGATAT
R3n2	-	10722-10743	TGGACCATTTCCGGTTTTCTG
Rur6	-	3706-3727	ATTCTTTGCACCGCNTGTTTC
Rur6walk	-	2948-2968	TAGGTAAAGCGAAAGAGCTG
Rur6.1	-	2217-2239	GTGAAAATGCGGAAACGGAATA
Rur6.1walk	-	1392-1412	ACTTCCTGAGGCTCTCCCTG

5.3.4 Primers used for sequencing *A. viteae* chitinase genomic clone 9

Name	Sense	Position	Sequence
F4	+	6645-6666	TGGGCTTGAAAGTGAGGTAAG
F>e10_9	+	8187-8208	GGAGAGCCTCAGGAAGTAGTG
K9_4pups	-	9414-9435	ACTAGGAAATTGGGAAAATACAAG
Rb4_4pupstr	+	10265-10286	TGCACTGTCCCATTATTGTC
K9_3pup.	-	10516-10537	CTTTGTACCGCTTGTTTCTTGTT
M13 forward	+	vector	GTA AAACGACGGCCA GT
M13 reverse	-	vector	CAGGAA ACAGCT ATG AC
K9-FV	+	780-801	GGAAATGCAAATACAAACGGC
K9-RV	-	12925-12946	AGGATGTGAGTGCATAGGTG
K9-V1	-	11359-11380	GTA CTTTATCAAACGTACCGT
K9-V2	-	9973-9993	GAAATAAAATCAGGAAGTTGC
K9-V3	-	8632-8650	TCAATTCATTTGACTGCCAC
K9-V4	+	2228-2249	CGAATATTCGTTTATCCATTG
K9-F1	+	3773-3795	GTAGGAAATGTCACATAACATC
K9-F2	+	4408-4427	CACTGATCGATGTAATATGC
K9-F3	+	5227-5240	CTGTGACGATCACTACCAGC
K9-F4	+	6117-6138	GTGAATAGCAACTTCTTTAGC

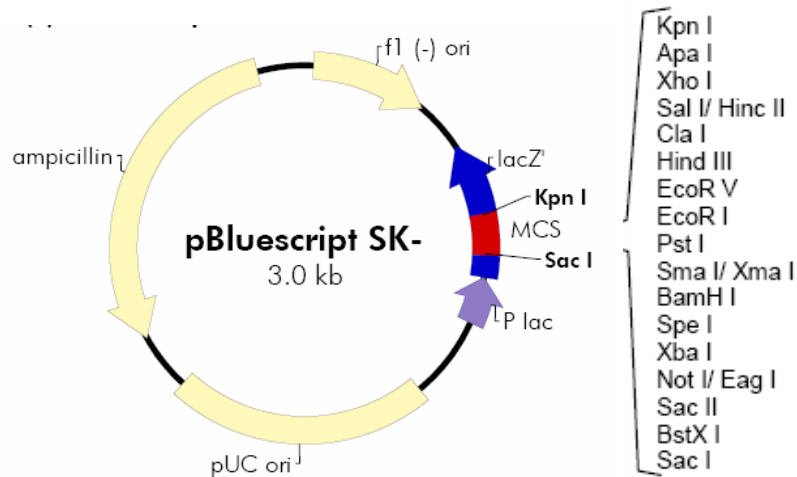
5.3.5 Primers used for sequencing *A. viteae* chitinase genomic clone 12

Name	Sense	Position	Sequence
F3	+	1374-1372	CTACGTTGCGGGATGTTAC
F3.walk12	+	2108-2129	GACGAAGATACCGAATGGTC
F4	+	4587-4607	TGGGCTTGAAAGTGAGGTAAG
F5	+	4542-4562	AAGTTGTGGCAAAGGTCCATA
F5.1	+	5288-5309	TACGTGCATCAAACACAGTC
F8	+	4130-4151	TGGCAAGGAAACGGTGGATAA
R4	-	4588-4608	TGTTTGCTCACTTTCAAGCCC
R7	-	3662-3683	TCCCAACTGCCGTGTAAATCA
R8	-	1709-1729	CATTTCCGACAGTAATACGAT
R13	-	543-564	AACCACGCTGAAGCCAAAATA
R13.1	-	950-971	GTAAGACTTGTACACTTCTG
F13	+	5836-5857	GTTTATTTCCGCATCACAGTG

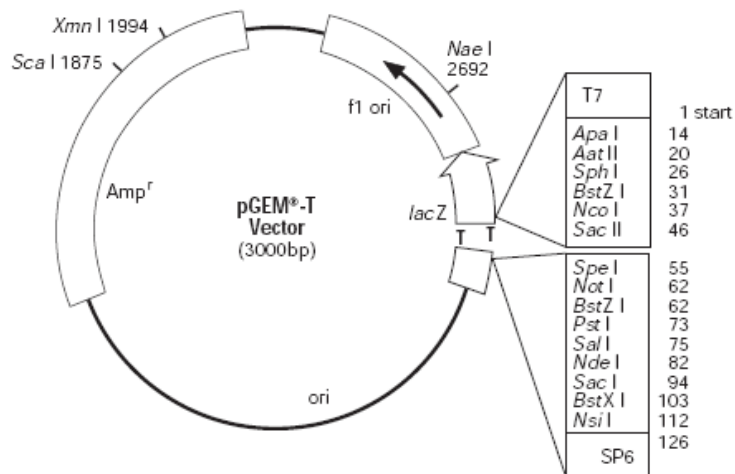
5.4 Plasmids

5.4.1 Cloning plasmids

5.4.1.1 Cloning plasmid for genomic inserts: pBluescript (Stratagen)

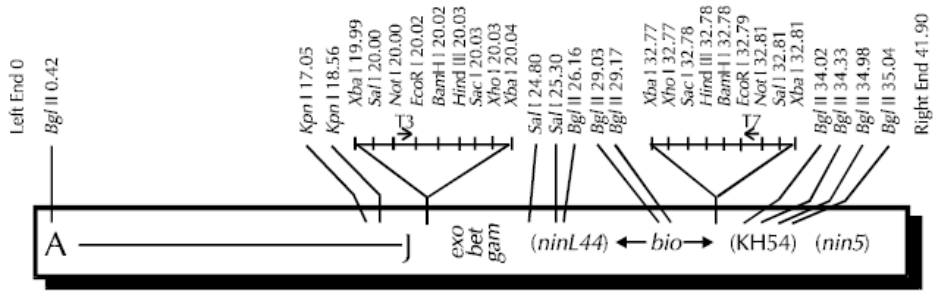


5.4.1.2 TA- overhang vector for cloning PCR products

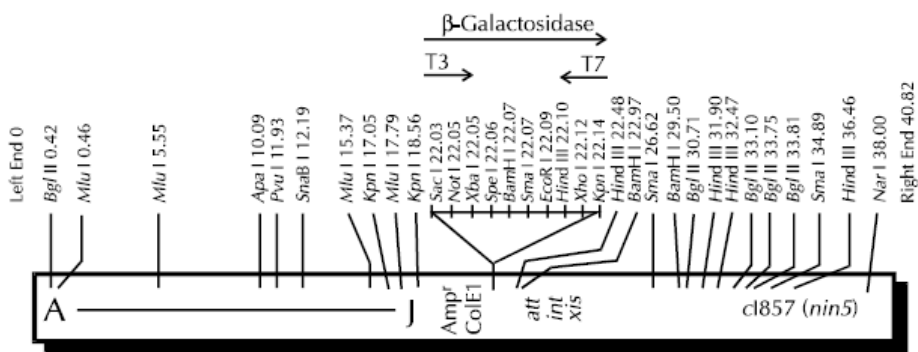


Source: Promega

5.4.2 Plasmids harbouring genomic and cDNA inserts: λ Dash II and λ Zap, respectively

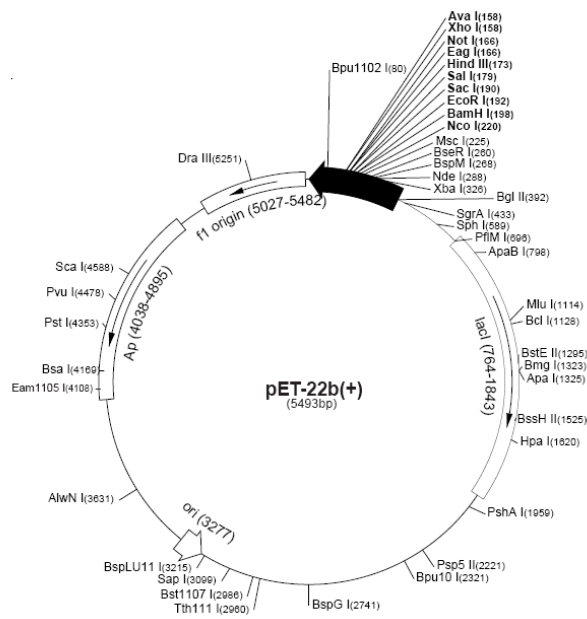


Lambda dash II replacement vector map (source: Stratagene)



Lambda Uni-Zap XR insertion vector

Expression vector: Vector maps of pET 22 b(+) expression vectors (Novagen)



5.5 Buffers and solutions

5.5.1 Molecular biology

5.5.1.1 Buffers for separation, purification and storage of DNA

<u>TAE</u>	<u>10x Sample buffer</u>
40 mM Tris/Acetate pH 8.0	250 mg bromophenol blue
1 mM EDTA	33 ml 150 mM Tris pH 7.6
	60 ml glycerol
	7 ml H ₂ O
<u>TE Buffer</u>	<u>Chloroform Isoamyl alcohol</u>
10 mM Tris/HCl, pH 7.5	96% (v/v) Chloroform
1mM EDTA	4% (v/v) Isoamylalcohol
<u>TlowE Buffer</u>	<u>Ethidium Bromide stock solution</u>
10 mM Tris/HCl, pH 7.5	10 mg/ml in H ₂ O
0.1 mM EDTA	
<u>Phenol chloroform solution</u>	
50 % (v/v) Phenol, pH 7.5	
48% (v/v) Chloroform	
2% (v/v) isoamyl alcohol	

5.5.1.2 Buffers for purification of plasmids: CTAB miniprep

RNase A	10 mg/ml
Lysozyme stock	50 mg/ml

<u>STET</u>	<u>CTAB solution</u>
50 mM Tris-HCl, pH 8	5 % CTAB in H ₂ O
8% sucrose	
0.1% triton X-100	
50 mM EDTA	

5.5.1.3 Buffers for purification of lambda DNA

SM buffer

5.8 g NaCl

2 g MgSO₄·7H₂O

50 ml 1 M Tris.Cl, pH 7.5

5 ml 2% (v/v) gelatine solution

H₂O to 1 l, autoclave

LB medium

see 5.6

5.5.1.4 Buffers and reagents used for screening of libraries

Hybridisation

Blocking solution

10 % (w/v) in MS 1 buffer

MS 1 buffer

0.1 M Maleic acid

0.15 M NaCl

adjust pH 7.5 @ 20 °C; autoclave

20 x SSC

175,2 g NaCl

88.4 g Na-citrate

adjust pH to 7 ; autoclave

Denaturing solution

0.5 M NaOH, 1.5 M NaCl

Neutralisation solution

0.5 M Tris/HCl, pH 7.5, 1.5 M NaCl

Prehybridisation buffer

50 ml 20 x SSC,

100 ml water,

20 ml blocking solution,

2.1 ml 10 % sarcosyl,

0.4 ml 10 % SDS

add dH₂O to 200 ml

Wash buffer I

2 x SSC, 0.5% SDS

Wash buffer II

2 x SSC, 0.1% SDS

Wash buffer III

0.1 x SSC, 0.1% SDS

Antibody reaction and development

Wash buffer:	MS 1 + 0.3 % (v/V) Tween 20
MS 2 buffer:	block solution diluted 1: 10 in MS 1
MS 3 buffer:	0.1 M Tris, 0.1 M NaCl, 0.05 M MgCl, pH 9.5; autoclave
Antibody solution:	dilute 1: 7,500 with MS 2 buffer (from Boehringer)
CSPD solution:	dilute 1:200 in MS 3 buffer

5.5.1.5 Buffers and reagents used for Southern blotting

20 X SSC	see above
Prehybridisation buffer	6 x SSC 5 x Denhardt's reagent 0.5% SDS 100 µg/ml Salmon sperm DNA
Wash buffers	See wash buffers I, II, and III above

5.6 Media and buffers for *E. coli*

<u>LB Medium</u>	<u>Top Agar</u>
10 g Bacto-Tryptone	0.75% agarose in LB
5 g Bacto-Yeast	
10 g NaCl	<u>10x Hogness freezing medium: for dauer cultures</u>
Add H ₂ O to 1l, pH 7.0	36 mM K ₂ HPO ₄ 13 mM KH ₂ PO ₄ 20 mM Sodium citrate
<u>LB Media with antibiotics</u>	10 mM MgSO ₄ 40 % Glycerin
LB supplemented with:	
100µg/ml Ampicillin and/or	
30 µg/ml Kanamycin and/or	
12.5 µg/ml Tetracyclin	
<u>Agar plates</u>	<u>SOB Medium</u>
LB medium supplemented with:	20 g/l Bacto Trypton
15 g/l agar and	0.5 g/l NaCl
Antibiotics (see above)	5g/l Bacto Yeast Extract
Dispensed into Petridishes	pH 7.0 with NaOH
	2.5 mM KCl
	10 mM MgCl ₂ (added before use)

SOC Medium

25 mM Glucose in ready-to-use SOB medium

TB buffer (transformation buffer)

10 mM PIPES

15 mM CaCl₂

250 mM KCl

adjust pH to 6.7 with KOH

add 55 mM MgCl₂

sterile filter

5.7 Protein and immuno- chemistry

5.7.1 SDS-PAGE

Separating gel buffer

1.5 M Tris/HCl, pH 8.8

Stacking gel buffer

1 M Tris/HCl pH 6.8

10x SDS-PAGE running buffer

10 g SDS

30 g Tris

144 g glycine

In 800 ml H₂O

Add H₂O to 1l

Coomassie staining solution

0.2% Brilliant blue G-250

40% Methanol

1% acetic acid

Destaining solution

10% acetic acid

30% Ethanol

2x SDS PAGE sample buffer

10 ml 1.5 M Tris/HCl (pH 6.8)

6 ml 20% SDS

30 ml glycerol

1.8 mg bromophenol blue

Add 168µl 3M DTT stock to 1 ml before use

Preparation of gels

	Separating gel			Stacking gel
	14%	12.5%	10%	4%
Acrylamide stock (30%)	18.7 ml	16.7 ml	13.3 ml	2.7 ml
1 M Tris HCl, pH 6.8	---	---	---	5 ml
1.5 M Tris HCl, pH 8.8	10 ml	10 ml	10 ml	---
10% SDS	400µl	400 µl	400 µl	200 µl
10% APS	200 µl	200 µl	200 µl	300 µl
H ₂ O	10.7 ml	12.7 ml	16.1 ml	12 ml
TEMED	20 µl	20 µl	20 µl	20 µl

5.7.2 Western blot

Transfer buffer (1l)

2.9 g glycine

5.8 g Tris

0.37 SDS

In 200 ml Methanol

Adjust volume to 1l

Ponceau S staining solution

2% Ponceau S

30% Trichloroacetic acid

30% Sulphobenzoic acid

Blocking buffer

5% skimmed milk powder in PBS

Washing buffer

0.2% Triton X 100 in PBS

Alkaline phosphatase buffer

100 mM Tris-HCl, pH 9.5

100 mM NaCl

5 mM MgCl₂Substrate stock solutions

NBT solution: 75 mg/ml in 70% DMF

BCIP solution: 50 mg/ml in 100% DMF

Alkaline phosphatase substrate (working solution)

66 µl NBT solution

33 µl BCIP solution

In 10 ml of alkaline phosphatase buffer

5.7.3 Buffers for native purification of N-terminal chitinase with Ni²⁺-NTA

Lysis buffer

50 mM NaH₂PO₄
300 mM NaCl
10 mM imidazol
pH to 8.0 with NaOH

Elution buffer

50 mM NaH₂PO₄
300 mM NaCl
250 mM imidazole
pH to 8.0

Wash buffer

50 mM NaH₂PO₄
300 mM NaCl
20 mM imidazol
pH to 8.0

5.7.4 Buffers for denaturing purification of N-terminal chitinase with Ni²⁺-NTA

Buffer A (1 liter)

100 mM NaH₂PO₄
10 mM Tris.Cl
8 M urea
adjust pH to 8.0 using NaOH

Buffer C (1 liter)

100 mM NaH₂PO₄
10 mM Tris.Cl
8 M urea
adjust pH to 5.9 using HCl

Buffer B (1 liter)

100 mM NaH₂PO₄
10 mM Tris.Cl
8 M urea
adjust pH to 6.3 using HCl

Buffer D (1 liter)

100 mM NaH₂PO₄
10 mM Tris.Cl
8 M urea
adjust pH to 4.5 using HCl

5.7.5 Synthetic peptides

B1 = Peptide 0: CGLLSYGGYNFGSSTFTAIA

C1 = Peptide 1: CGGGFDLDWEYPTGVAAEEHAK

D1 = Peptide 2: CGGTAAVSAGKMTIDESYNVQ

5.7.6 Buffers and solutions for chitinase enzyme activity assay

10x Enzyme assay buffer

20 mM NaPO₄, pH 6.0

0.2 M NaCl

1 mM EDTA

Stop buffer

0.5 M Glycin buffer, pH 11.0

Substrate

20 mM 4-methylumbelliferyl N',N'',N'''β Chitotrioside in 1x reaction buffer

5.7.7 Stock solutions

10 X PBS

80 g NaCl

2.0 g KCl

2.4 g KH₂PO₄

14.4 g Na₂HPO₄

in 1 l dH₂O, pH 7.4

1 M Tris

121.14 g Tris

In 800 ml H₂O

42 ml conc. HCl

pH to 8.0

ad H₂O to 1l

5.8 Antibiotic stock solutions

Ampicillin sodium salt

100 mg/ml in dH₂O

Kanamycin sulfate

10 mg/ml in dH₂O

Tetracycline

10 mg/ml in 100% ethanol

5.9 *E. coli* host strains and plasmids

XL-1 Blue	Stratagene, Heidelberg
XL-1 Blue MRA	Stratagene, Heidelberg
BL 21 (DE 3)	Novagen, Darmstadt
Origami B (DE 3)	Novagen, Darmstadt
pET 22 b(+)	Novagen, Darmstadt
pET 41 a (+)	Novagen, Darmstadt

5.10 Databanks, softwares and online services

Blast server for ESTs: <http://www.ebi.ac.uk/blast2/parasites>

Sequence analysis: <http://www.ebi.ac.uk/services>

Brugia malayi genome: <http://www.tigr.org/tdb/e2k1/bma1/>

Filaria genome Project: <http://nema.cap.ed.ac.uk/fgp.html>

5.11 Softwares

Genomic DNA analysis, Artemis: www.sanger.ac.uk/Software/Artemis/

Clustalw: <http://www.ebi.ac.uk/clustalw/>

Netgene: www.cbs.dtu.dk/services/NetGene2/

MacVector 7.2: Accelrys, USA

Oligo 4.0: <http://www.oligo.net/>

6 Abbreviations

APS	Ammonium peroxodisulfate
A	Adenosine
APOC	African Programme for Onchocerciasis Control
Amp	Ampicillin
AP	Alkaline phosphatase
bp	Base pairs
BCA	2,2'Bicinchoninic acid
BCIP	5-Bromo-4-chloro-3-indolyphosphate disodium salt
BSA	Bovine serum albumin (Fraction V)
cDNA	Complementary DNA
CTAB	Cetyltrimethylamoniumbromide
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid
dNTP	Deoxyribonucleoside triphosphate
DTT	Dithiothreitol
EDTA	Ethylenediamino tetraacetic acid
ELISA	Enzyme-linked immunosorbent assay
EtBr	Ethidium bromide
EtOH	Ethanol
HPLC	High performance liquid chromatography
IPTG	Isopropyl-thio- β -D-galactopyranoside
IU	International unit
kb	Kilo base
L ₁	First stage larvae
L ₂	Second stage larvae
L ₃	Third stage larvae
L ₄	Fourth stage larvae
LB	Lauria Bertani
LM	Low melting
2-ME	β -Mercaptoethanol
mAb	Monoclonal antibody
Mf	Microfilariae
mRNA	Messenger RNA
NBT	Nitroblue tetrazoliumchloride
NC	Nitrocellulose

OCP	Onchocerciasis Control Program
OD	Optical density
ON	Overnight
PAGE	Polyacrylamide gel electrophoresis
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
PEG	Polyethyleneglycol
PIPES	1,4-Piperazinediethane sulfonic acid
PMSF	Phenylmethylysuöphonylflouride
RNA	Ribonucleic acid
RNase	Ribonuclease
rpm	Revolutions per minute
RT	Room temperature
SDS	Sodium dodecylsulfate
TAE	Tris-acetate-EDTA
TB	Transformation buffer
TBS	Tris-buffered saline
TE	Tris-EDTA
TEMED	N,N,N',N'-Tetramethylethylenediamine
TRIS	2-Amino-2-hydroxymethyl-1,3-propandiol
U	Units
UV	Ultraviolet
X-gal	5-Bromo-4-chloro-3-indoxyl- β -D-galactoside

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8 Appendix

Appendix 1: Insert of clone 1; Sequence Range: 1 to 13963

Gene II: 73...1797 (3' end)

Gene III: 5280...11878

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      10      20      30      40      50      60      70      80      90
CTCACTAAGGGAAACAAAAGCTGGAGCTCCACCGCGGTGGCGGCGCGTAATACGACTCACTATAGGGCGAAGAATTGGATCTTCTAATT
L T K G T K A G A P P R W R P R N T T H Y R A K N S D L L F
S L R E Q K L E L H R G G G R V I R L T I G R R I R I F Y F
H * G N K S W S S T A V A A A * Y D S L * G E E F G S S I

      100     110     120     130     140     150     160     170     180
CTGATGTCGTATGATTACCGGTAGTTGGGAGAAGAACGTCGACTTACCGGAGAGTTGGTCCGACGGAGAGAAACATCCGGAATTG
L M S Y D L P V V G R R T S T Y T E S C V R R R E K H P E L
* C R M I Y R * L G E E R R L T R R V A S D G E R N I R N W
S D V V * F T G S W E K N V D L H G E L R P T E R E T S G I

      190     200     210     220     230     240     250     260     270
GCGTCTGTAATACCGTTAGTTCTTGAATAAAAAATTTCTAATTTTTTCTTGTGGTATGAAAGACGAATTCCTGATGATTCAGGAATTG
A S V I P L V L E I K I S N F F L F A * R R I L D D F R N L
R L * Y R * F L K * K F L I F S C L H E D E F L M I S G I C
G V C N T V S S * N K N F * F F L V C M K T N S * * F Q E F

      280     290     300     310     320     330     340     350     360
CAGCAAATTTATGGGCAGAGAAAGGTATGCCGAAACAGAAAATATCATTGGGATTCCGGCTTACAGTCGGGGATGGACATTAAGTAATC
Q Q I I G Q R K V C R N R K L S L G F R L T V G D G H * V I
S K L L G R E R Y A E T E N Y H W D S G L Q S G M D I K * S
A A N Y W A E K G M P K Q K I I I G I P A Y S R G W T L S N

      370     380     390     400     410     420     430     440     450
CTTCGAAACCGCAATTGGAGCGGAGGGTGGTCCATCATCGCCATCGACAACAAATCCCGCTGGCGGTACTGGGGCTTATGGGAGG
L R K R Q L E R R V V V H H R H R Q Q I P L A V L R L I G R
F G N G N W S G G W S S I I A I D N K S R W R Y C G L L G G
P S E T A I G A E G G R P S S P S T T N P A G G T A A Y W E

      460     470     480     490     500     510     520     530     540
TTTAATTTAAAGTTCTCATTACTTTTGAATATATTTCTTGCTGCTGATGAATTTCTCAAAATCCATTATTAATTAATGTAACCATTTA
F N L K F S F T F E Y I S C C * * I S Q I P L L I N V T I L
L I * S S H L L L N I F L A A D E F L K F H Y * L M * P F *
V * F K V L I Y F * I Y F L L L M N F S N S I I N * C N H F

      550     560     570     580     590     600     610     620     630
AGATATGCAAAATATGTGAAGAAGGTGGCAAGGAAACGTTGGATAAAAAAAGTGTGGAGCGTATATGGTAAAAGGAGATCAGTGGCAGG
R Y A N M * K K V A R K R W I K K V L E R I W * K E I S G T
D M Q I C E R R W Q G N G G * K K C W S V Y G K R R S V A R
K I C K Y V K E G G K E T V D K K S V G A Y M V K G D Q W H

      640     650     660     670     680     690     700     710     720
GTTACGATAATGAGGAAACTATTAATAAATAAAGTAATTTTTATTTCCGCTATTTGTTGGTAGTACAAAAATAAAAAAGTGAITTGATAG
V T I M R K L L K L K * F L F R L L L I V K N K K V I * *
L R * * G N Y * N * S N F Y F A Y C C * * S K I K K * F D S
G Y D N E E T I K I K V I F I S L I V V D S Q K * K S D L I

```


730 740 750 760 770 780 790 800 810
TCAAAAAATAAGAAAAGATAATAAGATAGTAGTCGAAAAATAAAAAATAAAAAATAACGTTGCCAATATATCTTGCCITTAACITTAATT
S K N K K R * * D S S R K I K N * K I T L P I Y L A L T L I
Q K I R K D N K I V V E K * K I K K * R C Q Y I L P * L * F
V K K * E K I I R * * S K N K K L K N N V A N I S C L N F N

820 830 840 850 860 870 880 890 900
TAAAGAGCACCATAAAITTTATAATGTTTATAAGAAGAAATTAATTTGGGAGTTTACTCGTGTAAATGTTTTACTTTAGAAATTAATAATT
* R A P * I L * C L I R R N * F R S L L V * L F Y F R I K I
K E H H K F Y N V * * E E I N F G V Y S C N C F T L E L K F
L K S T I N F I M F N K K K L I S E F T R V I V L L * N * N

910 920 930 940 950 960 970 980 990
TTCAGATGAAATGGCTGAAAGAGAAAGGTTATGGGGTGCCTTCATATGGGCTCTTGATTTTGTATGATTTTAAAGGGTACAAAGTTGTGGCA
F R * N G * K R K V M A V P S Y G L L I L M I L R V Q V V A
S D E M A E R E R L W R C L H M G S * F * * F * G Y K L W Q
F Q M K W L K E K G Y G G A F I W A L D F D D F K G T S C G

1000 1010 1020 1030 1040 1050 1060 1070 1080
AAGGTCCCTATCCACTGTTAAATGCTCTTATAATGAGCTTCGAGACGGCTTCGTATTTTGGGTGAAGGATTTTACAACTTCGTAATGAT
K V P I H C * M L L I M S F E T A S Y F G * R I L Q L R N D
R S L S T V K C S * * * A S R R L R I L G E G F Y N F V M I
K G P Y P L L N A L N N E L R D G F V F W V K D F T T S * *

1090 1100 1110 1120 1130 1140 1150 1160 1170
TTGTTGCATATCATAAAAATGTTATAATCGGTCCGAACCTTAAGGATTTGAAATTTAATCCGGAATCCATAAATTCATAATGTAATGCTG
L L H I I K M L * S V G T * R I E I * S G N P K I H N V M L
C C I S * K C Y N R S E L K G L K F N P E I L K F I M * C C
F V A Y H K N V I I G R N L K D * N L I R K S * N S * C N A

1180 1190 1200 1210 1220 1230 1240 1250 1260
TATTTTAAACAGTATTGCARACGCAACTGGTGCATTTTGGATATTTCTTTGCTTCTTACAAATGGTCCGCAAAAATGAATCAGGACAA
Y F K Q Y C K R N W C I F G Y S L L S Y K W S A K N E S R Q
I L N S I A N A T G A F L D I L C F L T N G P Q K M N H D K
V F * T V L Q T Q L V H F W I F F A F L Q M V R K K * I T T

1270 1280 1290 1300 1310 1320 1330 1340 1350
GCCGAAATGCTATTCAATTAATTTTATATTAATTTGTTCTATACAGATTTTAGATTTAAAAATTAATAAAGCAATCATTTTGTAGTC
A E M S I Q L N F Y I N F V L Y R F * I * K L I K Q S F L V
P K C L F N * I F I L I S F Y T D F R F K N * * S N H F * S
S R N V Y S I K F L Y * F R S I Q I L D L K I N K A I I F S

1360 1370 1380 1390 1400 1410 1420 1430 1440
ATCAATGTTGGCAAAATCAAATAATGAAATATTGAGGAAAACAGGGAGAGCCTCAGGAAGTAGTGAATTTGAAACGATCAAAAGTGGTTT
I N V G K I K * * N I E E N R E S L R K * * N * N D Q K W F
S M L A K S N N E I L R K T I G R A S G S S E I E T I K S G F
H Q C W Q N Q I M K Y * G K Q G E P Q E V V K L K R S K V V

1450 1460 1470 1480 1490 1500 1510 1520 1530
TGAACAGTACGAAGAATCTGGTTTGGTTATCCGCAAGTTAAATGTCGTTCTTGATCTGTTCTTTTGGCTTTAAACACAGCGCAACTTT
* N S T K N F W F V I R K L K C R S * S V L L L * T Q R N F
E T V R R T S G L L S A S * N V V L D L F F C F K H S A T F
L K Q Y E E L L V C Y P Q V K M S F L I C S F A L N T A Q L

1540 1550 1560 1570 1580 1590 1600 1610 1620
TCGAGTTTGGCTTTCATGCAATTTGGATAGAATGGAAATGATTGTGGATAGATGAAAAGAAATTTAGGAAAATCAATGTTCTGCTGTAAAC
S S L L F M H F G * N G N D C G * M K R N L G K S M F C C N
R V C S S C I L D R M E M I V D R * K E I * E N Q C S A V T
F E F A L H A F W I E W K * L W I D E K K F R K I N V L L *

1630 1640 1650 1660 1670 1680 1690 1700 1710
ATTTTCAGATATAAAATGTTTGGAGTCATATGGTCTGTTCCCGCATCCGTGACGATTGCCATTTATTTATTTCATTGGCGAAATGACTATCCA
I S D I K C L E S Y G L F P H P D D C H L F I H C A N D Y P
F Q I * N V W S H M V C S R I L T I A I Y L F I A Q M T I H
H F R Y K M F G V I W S V P A S * R L P F I Y S L R K * L S

1720 1730 1740 1750 1760 1770 1780 1790 1800
TACGTGAAACAATGTCGCGTACATACCTTTTTAAATGATGATATCAAAATTTGGGATCAGTATAGTAAATGCGCGATCAGATGCAAAATGA
Y V K Q C P L H T F F N D D I K I C D H L V N A P I T C K *
T * N N V R Y I P F L M M I S K F A I T * * M R R S H A N D
I R E T M S A T Y L F * * * Y Q N L R S L S K C A D H M Q M

1810 1820 1830 1840 1850 1860 1870 1880 1890
TTTTCATATTACCAAGAATAAAAGTTTAAATGTAAGTGTGGCAGTCAGTAAATGACCATACTCTGTAATAATTCATATTATATAG
F S Y Y Q E * K F N V K V V A V K * I D H T P V I I H I I *
F H I T K N K S L M * K L W Q S N E L T I L L * * F I L Y R
I F I L P R I K V * C K S C G S Q M N * P Y S C N N S Y Y I

1900 1910 1920 1930 1940 1950 1960 1970 1980
GAGAATAAATATGACTATATGAGGACAAATGTCAAAATGAGTCTAAGCGAATTTATTAATAATGAAAAATTTGTTATTATCTGTATTTT
E N K Y D Y M R T N V K N E S N G I I K N E K I C Y Y L Y F
R I N M T I * G Q M S K M S L T E L L K M K K F V I I C I L
G E * I * L Y E D K C Q K * V * R N Y * K * K N L L L S V F

1990 2000 2010 2020 2030 2040 2050 2060 2070
ATTTTGGGTAGCAAAAATCTTTCAACAGTTATCACCCATAACTTAGTAGAAAACATATGAATGATTGCGAAGATGAATAAATCAGATGAA
I L R S K N L S T V I T H N L V E N I * M I R K M N K S H E
F C V A K I F Q Q L S P I T * * K T Y E * F G R * I N H M K
Y F A * Q K S F N S Y H P * L S R K H M N D S E D E * I T *

2080 2090 2100 2110 2120 2130 2140 2150 2160
AAGGATGAAATTCGTATAAAAAATAAGTACTCAATAAAAAAGGATGAATCATTATAAAAAAGGAAATTCGGATAAAAAATAAATACTCAA
K D E I L I K N K Y S I K K D E I I I K R K F R * K I N T Q
R M K F * * K I S T Q * K R M K S L * K G N S D K K * I L K
K G * N S D K K * V L N K K G * N H Y K K E I P I K N K Y S

2170 2180 2190 2200 2210 2220 2230 2240 2250
AGATAAAAATACTGTTCCATTGCATCAGGACTGGAGTATTTCCATCAAGAAATCTTTTATCCGTTTCCGCAATTTTCACTGCTGCATAATT
R * K Y C S I A S G L E Y F H Q E S F I P F P H F H C C I I
D K N T V P L H Q D W S I S I K N L L F R F R I F T A A * F
K I K I L F H C I R T G V F P S R I F Y S V S A F S L L H N

2260 2270 2280 2290 2300 2310 2320 2330 2340
CCITCCITTTTCTTTTGGCAATTCCTTTTGGTTTCCITTTACGACAATTCAGCTTAATTTTGATAAGAGAGTTTGTAGTTGGCTAATGTGATGA
P S F S F G I P F V S F T T I Q L N F D K R V C S W L M * *
L P F L L A F L L F P L R Q F S L I L I R E F V V G * C D D
S F L F F W H S F C F L Y D N S A * F * * E S L * L A N V M

2350 2360 2370 2380 2390 2400 2410 2420 2430
CACTCAATTCGAAAAAGGAAACAGTCAAAGGCGAATTTCAATTTTCAACAATTTCTCATCATTGAATGATATGTGCGATAGAAGGAAT
H S I R K R K Q S K A N F N Y F Q Q F S S L N D M C D R R N
T Q F E K G N S Q R I S I I F N N S H H * M I C A I E G I
T L N S K K E T V K G E F Q L F S T I L I I E * Y V R * K E

2440 2450 2460 2470 2480 2490 2500 2510 2520
TTTCATAATCTCAGATGAAGAAAAGTATTTTGCAGTTGCGCTTCATCTTTCTCTTCCATATTATGATAAAAAGATATAAAAAACTTCCA
F H N L R * R K V F C S C A S S F S S I L * * K I * * K L P
F I I S D E E K Y F A V A L H L S L P Y Y D K R Y N K N F Q
F S * S Q M K K S I L Q L R F I F L F H I M I K D I I K T S

2530 2540 2550 2560 2570 2580 2590 2600 2610
ATAATAAACGGTGGCAGCATTATTTTTCTAGCTTTCTGCTTATTTTTAATGTTTTTCCAGCTTTCTAGCTATTTCTGTATTTCCCAAT
I I N V A A L F F L A F C L F L M F F Q L S S Y F L Y F P N
* * T W Q H Y F F * L S A Y F * C F S S F L A I S C I F P I
N N K R G S I I F S S F L L I F N V F P A F * L F L V F S Q

2620 2630 2640 2650 2660 2670 2680 2690 2700
TTCTAGTGTTTTTCCAGCTTTTGTAGTTGTTTTCTAACTCCTTAATACTATCTACGTCAGCCTAACTTCGGGATTTTAATTAACAAAT
F L V F F Q L L * L F F * L L N T I Y V S L T S G F * L T N
S * C F S S F C S C F S N S L I L S T S A * L R D F N * L I
F P S V F P A F V V V F L T P * Y Y L R Q P N F G I L I N *

2710 2720 2730 2740 2750 2760 2770 2780 2790
TTGGTAAATCAAAATATCTTCGAATCAAAATACCTTTGTAGAGCGCTTATTTTTGACACTCTTAATGTTCTTAAAAAAAACCTTACTTCATTG
L V N Q I S S N Q I L C R A L I F D T L N V L K K N L T S L
W L I K Y L R I K Y F V E R L F L T L L M F L K K T L L H C
F G * S N I F E S N T L * S A Y F * H S * C S * K K P Y F I

2800 2810 2820 2830 2840 2850 2860 2870 2880
TTATGTTTTATGCTTAAACATTTATTAATAAATGCAGAAATTTTTTTTTTTTTTTTTTTGATGTTGCTGAGAACTTTTGAAAA
L C F Y A * H L L I T A E I I F F F Y F W M C S L R T F E K
Y V F M L N I Y * * L Q K L F F F F I F G C V R * E L L K K
V M F L C L T F I N N C R N Y F F F L F L D V F A E N F * K

2890 2900 2910 2920 2930 2940 2950 2960 2970
AHTTGTTGTTATGATTTTGGATTTTTCCACCGTTTTTCATTTTTTCTCTTATACTTATTGTTGTTTTCCAGCTCTTTCCGCTTTACCTAAT
I C C Y D F D F S P F F I F F S Y I L L C F P A L S L Y L I
F V V M I L I F H R F S F F S L I S Y C V F Q L F R F T * L
N L L L * F * F F T V F H F F L L Y L I V F S S S F A L P N

2980 2990 3000 3010 3020 3030 3040 3050 3060
GCAAGATTTTAAATATCTGACGATTTACAAAATGTCAGCAATTTAATTTCTTTATTTGTTAATATGTTTGCATTTTATTTTGCACCTT
A R F * N I * R F T K C A T L L I L Y C * Y C L H F I L Q L
Q D F K I S D D L Q N V Q H Y * F F I V N I V C I L F C N F
C K I L K Y L T I Y K M C N I I N S L L L I L F A F Y F A T

3070 3080 3090 3100 3110 3120 3130 3140 3150
CCTGATTTTATTTCCAGCTATGCAAAACATTTGCTATCTATAAAATCTGTTTCAAACCTTAAGCCTACCAATGTTTTTACAGTAATCCAT
P D F I S K L C K H F V S I K S V S N L S L P M F F T V I H
L I L F P S Y A N I S Y L * N L F Q T * A Y Q C F L Q * S I
S * F Y F Q A M Q T F R I Y K I C F K L K P T N V F Y S N P

3160 3170 3180 3190 3200 3210 3220 3230 3240
TTTTGATTTCCATATAACTCAACCAATTTTCATGATCTACCGACTTTTATTTATATTTTCATTCATTCGTTCCAAATTTCACTTTTATTTCC
F * F P Y N S P Q F H V S T D F Y L Y F H S F V Q F H F Y S
F D S H I T H P N F M Y L P T F I Y I F I H S S N F T F I P
F L I P I * L T P I S C I Y R L L F I F S F I R P I S L L F

3250 3260 3270 3280 3290 3300 3310 3320 3330
AATTTGAGCATATAAATATGAATAAAATTAAGTATTTTGTATGATGATATTAAGATATGAAAAATTTCCAGCTTATATTTAACITTTGAT
N L S I * I * I K L S I L Y D D I K I * K I S S L Y L T F D
I * A Y K Y E * N * V F C M M I L R Y E K F P A Y I * L L I
Q F E H I N M N K I K Y F V * * Y * D M K N F Q L I F N F *

3340 3350 3360 3370 3380 3390 3400 3410 3420
ATGCACAGTATAATTCATTTTCTGATTTCCGACAAAAGATTGAAGATGTGAAAAATATTGACCTTTACCAAAAATGTTGGAGCATTATTATT
M H S I I H F L I S A Q R L K M * K Y L T F T K M W S I I I
C T V * F I F * F P H K D * R C E N I * P L P K C G A L L F
Y A Q Y N S F S D F R T K I E D V K I F D L Y Q N V E H Y Y

3430 3440 3450 3460 3470 3480 3490 3500 3510
CATATTAATTTTTTTCAGTATAATAATGCACITGTOCCATTTATTGTCAACAAAAAATAATCCCAACGGCTGAGCTGCTTTTAATAATA
H I N F F S I I M H C P I Y C Q Q K K K I P N A E L L L I I
I L I F S V * * C T V P F I V N K K K K S Q T L S C F * * Y
S Y * F F Q Y N N A L S H L L S T K K K N P K R * A A F N N

3520 3530 3540 3550 3560 3570 3580 3590 3600
TAATTTAATAATAATTTTCATATATTTCCATTAATTGATTATTTTATTATATTCCTTAAACATAGCAGCGTGTGCTCITTCITCTGAC
* F * * * F H I F P L I D Y F I I F L K H S S V V V F F S D
N F N N N F I Y F H * L I I L L Y S L N I A A L S S S S L T
I I L I I I S Y I S I N * L F Y Y I P * T * Q R C R L L L *

3610 3620 3630 3640 3650 3660 3670 3680 3690
CGTGATAATTTTACGGGTTTGATTGCAATCTTTTCAGTACTGTGAACITCAAACCTTGGTAATTAAGCCATAGCAATGATAGTAAAA
R D N I Y G F D C N L S V T V N F K L W * L K P * Q * * * K
V I I F T G L I A I F Q L L * T S N F G N * S H S N D S K N
P * * Y L R V * L Q S F S Y C E L Q T L V I K A I A M I V K

3700 3710 3720 3730 3740 3750 3760 3770 3780
CACAAATATTAACAAGAACANGCGGTGCAAGAAATAAATCAAACACATAACCTCAACGGGAAGAAATAGTAGAAAGAAATGAAGAGATG
H N I K T R N X R C K E * I K T H N L N G R I S R K N E E M
T I L K Q E T X G A K N K S K H I T S T E E * V E R M K R *
T Q Y * N K K X A V Q R I N Q N T * P Q R K N K * K E * R D

3790 3800 3810 3820 3830 3840 3850 3860 3870
AATGATTTGATGCTTATATAGGCAAGATACGTGCATATCATAACCGGGAAATAAGGAAAGATATTGGAGGATATAAGAGCAAGG
N D L M S Y I G K V Y V H I I T R E N K E R Y W R I * E Q R
M I * C L I * A K Y T C I S * R G K I R K D I G G Y K S K G
E * F D V L Y R Q S I R A Y H N A G K * G K I L E D I R A K

3880 3890 3900 3910 3920 3930 3940 3950 3960
AGATCATGTAGTACATAAATGGAGGCAAGATGGGAACGGATGCGAGTATAGAAATGGATCATTTGAATGTGTTATTTAGTTAGACAGG
R S C S T I I G G K D G N G C E Y R M D H L N V L F S L D R
D H V V Q * L E A K M G T D A S I E W I I * M C Y L V * T G
E I M * Y N N W R Q R W E R M R V * N G S F E C V I * F R Q

3970 3980 3990 4000 4010 4020 4030 4040 4050
AACATCGACATTTGACACGTATTACATGCAGTAATATTCATTATTACAATATGTCTATGAATAAAATATATATATATATATATATC
N I D I * H V L T C S N I H Y Y N Y V Y E * N I Y I Y I Y I
T S T F D T Y * H A V I F I I T I M S M N K I Y I Y I Y I S
E H R H L T R I N M Q * Y S L L Q L C L * I K Y I Y I Y I Y

4060 4070 4080 4090 4100 4110 4120 4130 4140
TTTGGCGCAAAAGGAATTTGCTTTCTCTGAAATTTTAAATCCAAACAAAAAATAAATAAAATTAATTCITCTTTGGTACTTCTGCT
F R D K R N C F S L K F * S K T K K L I K I N S S L V L L A
F A T K G I A F L * N F N P K Q K N * * K L I L L W Y F S L
L S R Q K E L L F S E I L I Q N K K I N K N * F F F G T S R

4150 4160 4170 4180 4190 4200 4210 4220 4230
TTTCACGTATCTTCCATTTAGTGAAGCCATGAATGAAAACATTACCAACGCAACGGTAAGTGTCCAGTAAAGTGACAAATTTTGGTTGG
F H V S S N * * S H E * K H Y Q R K R * V V Q * S D N F G W
F T Y L P I S E A M N E N I T N A N G K L S S K V T I L V G
F S R I F Q L V K P * M K T L P T Q T V S C P V K * Q F W L

4240 4250 4260 4270 4280 4290 4300 4310 4320
ATGATATAGTGGTATATGGAATTTTACACTCTCAGATAAATGAAAAACGTACTTTGTGTTATACGTCACATTTTCTTTTACATATG
M I * V V Y G I F T L S D K * K T Y F V L Y V T F S F H I M
* Y R W Y M E F S H S Q I N E K R T L C Y T S H F L F T L *
D D I G G I W N F H T L R * M K N V L C V I R H I F F S H Y

4330 4340 4350 4360 4370 4380 4390 4400 4410
ATAAAAAGACATAACAAAATTCGACACAGTACGGTATTATTGTGTTTTTCTAACTTTTAGGCTGTGTAAATTTTTCCAGCTTTCCAGC
I K D I T K F S T S T V L F V F F * L L G C C * F F Q L S S
* K T * Q N S R Q V R Y Y L C F S N F * A V V N F S S F P A
D K R H N K I L D K Y G I I C V F L T F R L L L I F P A F Q

4420 4430 4440 4450 4460 4470 4480 4490 4500
TATTTACGTATATTTTTCCATCTTTTTGGCCATTTCCTGTGTTTTTTCAGCTTTCTCGTGTTTTTCTAACTCCTTGAATATTTTTCGTA
Y L R I F F H L F W P F L V F F Q L S R V F L T P * I F F V
I Y V Y F S I F F G H F L C F F S F L V F F * L L E Y F S Y
L F T Y I F P S F L A I S C V F S A F S C F S N S L N I F R

4510 4520 4530 4540 4550 4560 4570 4580 4590
TTTTACAGCTTTTTAGAGGAGATGGTTTACATTTTCTAAAGAACGGTACGTTTGATAAAGTACTTATATAAAAGGCCGTTTATGGCGT
F S Q L F R G D G L H F P K E R Y V * * S T L * K A R L W R
F H S F L E E M V Y I F L K N G T F D K V L Y K R P V Y G V
I F T A F * R R W F T F S * R T V R L I K Y F I K G P F M A

4600 4610 4620 4630 4640 4650 4660 4670 4680
TAAAGACTTCACITTTTCATTTTATATTTTGTCTATCTTTCAACATTCCTCCCGATTACTTTTACAAAAATATCTTTTTTCTATTTTGCACA
* R L H F S F Y I L L S F N I P P D Y F T K I S F F Y F C T
K D F T F H F I F C Y L S T F L P I T L Q K Y L F S I F A H
L K T S L F I L Y F A I F Q H S S R L L Y K N I F F L F L H

4690 4700 4710 4720 4730 4740 4750 4760 4770
TTTATCTTTTTTTTTGAATTTCTTTCTTTTTTGTCTATTTTCTAGATATTTCCAGATTTTTAGTTTTGTATGCAATCTCTGAGAAATGCTT
F I F F F E F L S F L L F S R Y F Q I F S F V C N L * E C L
L S F F L N F F L F C Y F L D I S R F L V L Y A I S E N V L
I Y L F F * I S F F F A I F * I F P D F * F C M Q S L R M S

4780 4790 4800 4810 4820 4830 4840 4850 4860
GCAATTTGGCGAAAAATAGAAAAAGATATTTTCATCGATATTTCTTAATTAGTCAATTTACATTAGCTGCAGCATACTCATTGAATTCAT
A I C A K I E K D I S S I F L N * S F T L A A A * L I E F H
Q F A Q K * K K I F H R Y F L I S H L H * L Q H N S L N S I
C N L R K N R K R Y F I D I S * L V I Y I S C S I T H * I P

4870 4880 4890 4900 4910 4920 4930 4940 4950
ATTTATCCCAAGATTAGTATATCAACATTATATTAACGGTTGATATACTAATCTCATTGGAAACAAATGATATCAAAAACATGAAAA
I Y P K D * Y I N I I L N V D I L I S F G T K * Y Q K H E K
F I P K I S I S T L Y * T L I Y * S H L E Q N D I K N M K N
Y L S Q R L V Y Q H Y I K R * Y T N L I W N K M I S K T * K

4960 4970 4980 4990 5000 5010 5020 5030 5040
TTTGTACTATAGAAAATACTGTACATCTAAGCTTACTTCTCAATTAATTTTTCTTGAATCCTTTGTATGCACCGACAGTTA
F A T Y R K Y C T S K L T S T S Q L I F S * I L C M H R Q L
L L L I E N T V H L S L L L L L N * F F L E S F V C T D S Y
I C Y L * K I L Y I * A Y F Y F S I N F F L N P L Y A P T V

5050 5060 5070 5080 5090 5100 5110 5120 5130
TTCTCACCARTCATTTCAATACAATAATCTCTTATTACTCTGTAACCTCAGCAAAAAAGTATTCCCTAATCGCTGAGCTATGTGTAATTA
F S P I I S I Q * S L I T L * T Q Q K K Y S * S L S Y V * L
S H Q S F Q Y N N L L L L C K L S K K S I P N R * A M C N *
I L T N H F N T I I S Y Y S V N S A K K V F L I A E L C V I

5140 5150 5160 5170 5180 5190 5200 5210 5220
GTTACGGTAATTCATGCATGATATACTGATTGTTTTGCGTATTTTCTTACACATACACTTAATATCTTCCATTAATGAAGGCCAATA
V T V I P C M Y I N * L F R L F L T H T L N I F H * * R Q I
L R * F H A C I L T D C F A Y F L H I H L I S S I N E G K Y
S Y G N S M H V Y * L I V S L I S Y T Y T * Y L P L M K A N

5230 5240 5250 5260 5270 5280 5290 5300 5310
TCAGTTAATCATAGTAATGAATTTTGAATAACAGAAGTGTACAAGTCCCATGAAGTGACATGAATTGGATTATGATAACGCTTTTCATCA
S V N H S N E F * N T E V Y K S H E V T * I G L * * R F S S
Q L I I V M N F E I Q K C T S P M K * H E L D Y D N A F H H
I S * S * * * I L K Y R S V Q V P * S D M N W I M I T L F I

5320 5330 5340 5350 5360 5370 5380 5390 5400
TATTGCGCAATATAAATTACTGTTGCAAAATGGTAAGTTGCTCAATAAAATGACACGTCAGTTGCGTAAGCACGAAATATCATCGCTGAAGA
Y S P I * L L L Q M V S C S I K * H V S C V S T K Y H R * R
I R Q Y N Y C C K W * V A Q * N D T S V A * A R N I I A E D
I F A N I I T V A N G K L L N K M T R Q L R K H E I S S L K

5410 5420 5430 5440 5450 5460 5470 5480 5490
TATAATGAACATGACGGTTGACACTTCAGATCATACTACATCTTAATCATTATATGTTCATTTCATCCTAACTGTCGAGTAGCATCTAT
Y N E H D G * H F R S Y Y I L I I I C H F N P K L S S S I Y
I M N M T V D T S D H T T S * S L Y V I S I L N C R V A S I
I * * T * R L T L Q I I L H L N H Y M S F Q S * T V E * H L

5500 5510 5520 5530 5540 5550 5560 5570 5580
TGAAATTCAAATAGTGGCATATATTGAATTTCAACAGAAGCTCTCACAGAATAAGCAAATGAAGAGACTAAATTTGAAGATAATACAAAT
* N S I V A Y I E F H R S S H K K * A N E E T K L K I I Q I
E I Q * W H I L N F T E A L T R N K Q M K R L N * R * Y K F
L K F N S G I Y * I S Q K L S Q E I S K * R D * I E D N T N

5590 5600 5610 5620 5630 5640 5650 5660 5670
TTTACTTGAAGATTTAGTAACATTGTATTTGAATAAATTCGAAAGAAATAACCACCAATCGATGTTGAACATATCTTCAGGTTGTAGTC
F T * R F S N I V F E * I A K K * P P N R C * T Y L Q V V V
L L E D L V T L Y L N K L Q R N N H Q I D V E H I F R L * S
F Y L K I * * H C I * I N C K E I T T K S M L N I S S G C S

5680 5690 5700 5710 5720 5730 5740 5750 5760
TATTTGAGTTACGTTGCGTGGATGTTACTATATATTAGTTGGGCTGAACGTCACCAAGGTTGAGTGACGTTTGAATAAATTTCAATATAAA
Y F R L R S W M L L Y * L G * T S T R F S D V * N N F I I K
I S G Y V R G C Y Y I S W A E R Q Q G S V T F E I I S L L K
L F Q V T F V D V T I L V G L N V N K V Q * R L K * F H Y *

5770 5780 5790 5800 5810 5820 5830 5840 5850
GAGCTAAATAGTTTCAAGGGGATTTGTGTACAAATTTTGAATAAATAAGTAGAAAACCAATGAAATTTATTTCAAGATTTCCATCTCAAAA
E L N S F K G I V Y K F S N K K * K T N E I I S R F P S Q K
S * I V S R G L C T N F R I K S R K P M K L F Q D F H L K N
R A K * F Q G D C V Q I F E * K V E N Q * N Y F K I S I S K

5860 5870 5880 5890 5900 5910 5920 5930 5940
TGAATTTAAGCCACGAAATATTAGTGGAAAATAAACATCAATAATTAGCAACTTCAACTTCCACATRAACTCAATATTCCCTCATT
* I L S P R N I S G K * T S I I S N F N F Q H K L N I P S F
E F * A H E I L V E N K H Q * L A T S T S N I N S I F P H F
M N F K P T K Y * W K I N I N N * Q L Q L P T * T Q Y S L I

5950 5960 5970 5980 5990 6000 6010 6020 6030
CCTGAATTTCTCATCGTTTTCAGCGTGTGCTTAGCTACCGTAACTAATATGGTGAATAAATAACTAAACAATTAATAAATAAACAAGCA
P E F S S F R R V P * L P * T N M V K I I T K Q I K I K Q A
L N S H R F D V C L S Y R K L I W * K * * L N K L K * N K H
S * I L I V S T C A L A T V N * Y G E N N N * T N * N K T S

6040 6050 6060 6070 6080 6090 6100 6110 6120
TTGTATTTCTGCCGGAATGTAATATACATTTTCATCTTAAGGAGAAAGGCAATTTTGGCCAGAAGATATCCAAAACACCTATGCACT
L Y F C R K C N I H F H L K G E G K F L P E D I P K H L C T
C I S A G N V I Y I F I L K E K A N F C Q K I S Q N T Y A L
I V F L P E M * Y T F S S * R R R Q I F A R R Y P K T P M H

6130 6140 6150 6160 6170 6180 6190 6200 6210
 CACATCTTTTATGCATTACCAACCGTTGAAGGGAAAACGGCGTAAAGTTACAAATGATATTTGCAATGAGAAATGTAAATTAATTTTCGG
H I L Y A F T N V E G K R A * V T N D I C N E K C K L N F R
 T F F M H S P T L K G N A R K L Q M I F A M R N V N * I F G
 S H S L C I H Q R * R E T R V S Y K * Y L Q * E M * I K F S

6220 6230 6240 6250 6260 6270 6280 6290 6300
 AGTTTCTTTTACAAATGCACATTAAAATTATTCTAATTTTTATTGCTGCTCAAGTGATTATAAACCTGAAAACGTTAATATGGGATATTTA
 S F F Y N A H * N Y S N F Y C L L K * L * T * N V N M R Y L
 V S F T M H I K I I L I F I A C S S D Y K P E T L I C D I *
 E F L L Q C T L K L F * F L L P A Q V I I N L K R * Y A I F

6310 6320 6330 6340 6350 6360 6370 6380 6390
 GCTGGTTTTCGATAAAATATTCTTCTGTAACAATTGCTATCAGCAAATGTTATTTATTTCTGTAAATTTCTGATTTTAATAATTTTTTAA
 A G F A * N I L L * Q L L S A N V I Y F C K F L I L I I F K
 L V L H K I F F C N N C Y Q Q M L F I S V N F * F * * F L N
 S W F C I K Y S S V T I A I S K C Y L F L * I S D F N N F *

6400 6410 6420 6430 6440 6450 6460 6470 6480
 TCTTAGAACATTTGAATGGGACGATGAAGATACCGAATGGACGAAAGGGATATATCCACATATGATAAAAGTAAAGAGTACGATCCAAC
 S * N I * M G R * R Y R M D E R D I S T Y D K S K R V R S N
 L R T F E W D D E D T E W T K G I Y P H M I K V K E Y D P T
 I L E H L N G T M K I P N G R K G Y I H I * * K * K S T I Q>

6490 6500 6510 6520 6530 6540 6550 6560 6570
 CCTGAAAATTCCTTTTCCATAGCGACTATAACTCCTACTCATCAGTTTTTGGCTGTTGCTTTTTTTTTTAAACTCTCAGCIATCATTTTT
 P E N S S F L * R L * L S L I S F C C S F F F * T L S Y H F
L K I L L S Y S D Y N S H S S V F A V R F F F K L S A I I F
 P * K F F F P I A T I T L T H Q F L L F V F F L N S Q L S F

6580 6590 6600 6610 6620 6630 6640 6650 6660
 TTTTTTAAAGATTGACACTCAGTTAACGTATTGACTAATAAATTTCAAAGAGACTGCTCATNOGTCATTNTTANACATNAAGATAATG
 F F * R L T L S * R I D * * F Q K R L L X R H X L X X K I M
 F F K D * H S V N V L T N N F K R D C S X V I X X T X R * C
 F F L K I D T Q L T Y * L I I S K E T A H X S F X X H X D N

6670 6680 6690 6700 6710 6720 6730 6740 6750
 CAGGTTAACCTCANACGTTTTCCGGATTACGNNTCATNTTTTTNTTGGCTAAAATATCAACGTAATTTGATGATNTAAAATNTTTGAA
 Q G Y L X R F P D L R X X F X L L K Y Q R K F D D X K X F E
 R V T S X V F R I Y X S X F X C * N I N V N L M X * K X L K
 A G L P X T F S G F T X H X F X A K I S T * I * * X K N X *

6760 6770 6780 6790 6800 6810 6820 6830 6840
 GGCAATGACAAAATCGACAAAAAACCGGAAAAATTTTATTTCAGNCAACTGTAGCATTTTCTCCGTAAGCATAAATTTGANGGATTGATCT
 G N S K I D K K P E K F H S X N C S I S P * A * I * X I R S
A I A K S T K N R K N F I Q X T V A F L R K H K F X G F D L
 R Q * Q N R Q K T G K I S F X Q L * H F S V S I N L X D S I

6850 6860 6870 6880 6890 6900 6910 6920 6930
 CGACTGGGAATATCCAAACCGGNGTAGCAAAGGAACACGCTAAACTTCTTAAGGGGAGTGTAAATTCATATTTCAACTTTTTTTCAGTCCACC
 R L G I S N R X S K G T R * T S * G E C N S I F N F F Q S T
D W E Y P T G V A K E H A K L L K A S V I P Y S T F F S P P
 S T G N I Q P X * Q R N T L N F L R R V * F H I Q L F S V H

6940 6950 6960 6970 6980 6990 7000 7010 7020
 GGAGCAGGCTCATTTCCGAATTGAGCAATACTTGGTGAGCACTGTTTATAGCAATTAATTAATTAATCACTTGGTGAGCACTGTTTATAGC
 G A R S F R I E Q Y L V S T V Y S N * L I N H L V S T V Y S
 E H A H F E L S N T W * A L F I A I N * L I T W * A L F I A
 R S T L I S N * A I L G E H C L * Q L I N * S L G E H C L *

7030 7040 7050 7060 7070 7080 7090 7100 7110
 AATTAATTAATTAATCACTGGTGGAGCACTGTTTATAGCAATTAATTAATTAATCACTTTGTGTATGAAATTAATAGAGAACAAGGAATC
 N * L I N H L V S T V Y S N * L I N H F V Y E I N R E Q G I
 I N * L I T W * A L F I A I N * L I T L C M K L I E N K E S
 Q L I N * S L G E H C L * Q L I N * S L C V * N * * R T R N

7120 7130 7140 7150 7160 7170 7180 7190 7200
 TTCTGGCATTGTTTATAGTGGTTAGATAAGGAGCTGTTGTTGAATATCCGGTAGAGGAAAATAATAATAACAAGAATTACCTGGTCAT
 F W H C L * W L D K E L L L N I R * R K I I I N K E L P G H
 S G I V Y S G * I R S C C * I S G R G K * * * T K N Y L V I
 L L A L F I V V R * G A V V E Y P V E E N N N K Q R I T W S

7210 7220 7230 7240 7250 7260 7270 7280 7290
 CTGATAGGGGAACTGATAGAAAATAAAAGATATTTTCTGTGCAATCACATATCGAAGAACAATAAAGCATATTTTATCACCTGACTG
 L I G G T D R K * K I F S V E S H I E R T N K A Y F I T * L
 * * A E L I E N K R Y F L S N H I S K E Q I K H I L S P D C
 S D R R N * * K I K D I F C R I T Y R K N K * S I F Y H L T

7300 7310 7320 7330 7340 7350 7360 7370 7380
 TTCTATAGCATCGATATCAAGAATCAACTATATATTAAATTATACAAATATTATTATATAAAAATTATTTCTCATTTCGACACTCGTC
 F Y S I D I K N Q L Y I N Y T N I I I L * N Y F S F P T L V
 S I A S I S R I N Y I L I I Q I L L Y Y K I I S H F R H S S
 V L * H R Y Q E S T I Y * L Y K Y Y Y I I K L F L I S D T R

7390 7400 7410 7420 7430 7440 7450 7460 7470
 ARAATTTAATTCACATTCAAAAGAATACTCCACGIGATTTCGTGTAATTTCTCTTCAAAAATGAATTAGTTTCAAAAAGCAAAATTGA
 K F * F T F Q K N T P R D F C N F L F K N E L V S K A K L K
 N F N S H S K R I L H V I S V I F S S K M N * F Q K Q N * N
 Q I L I H I P K E Y S T * F L * F S L Q K * I S F K S K I E

7480 7490 7500 7510 7520 7530 7540 7550 7560
 TATATCAGATCGTACGTGAGAGGATGAAAATTCAAAACGAACTTTGTAGGAAATGAAAGCGGGCTTCGTGGAAGAAGCCAAAGGAATCCG
 Y I R S Y V R G * K I Q N E L C R K * K R R S W K K P R N P
 I S D R T * E D E K F K T N F V G N E S G V R G R S Q G I R
 I Y Q I V R E R M K N S K R T L * E M K A A F V E E A K E S

7570 7580 7590 7600 7610 7620 7630 7640 7650
 GCAAGCAACRAATTAATTCACCGCTGCTGTATCAGCAGTGAAGAATAATTAATGATGAAAGTTACAGTGTTCGATCCCTTGGAAAGTAA
 A S N N Y F L P L L Y Q Q * K K L L M K V T V F D P L E S K
 Q A T I T S Y R C C I S S E R N Y * * K L Q C S I P W K V K
G K Q Q L L L T A A V S A V K E T I D E S Y S V R S L G K *

7660 7670 7680 7690 7700 7710 7720 7730 7740
 GAAATTCATTTTCATACATTGACATTAATTAACACGCAATAAATTAACAATACATTCAAATATATTTTTGTAGCGTGAATTTTCA
 E I A F H T L T L N L T R N K I Y N T S N I F L * R * K F S
 K L L F I H * H * I * H A I K F T I L Q I Y F C S V E N F Q
 R N C F S Y I D I K F N T Q * N L Q Y F K Y I F V A L K I F

7750 7760 7770 7780 7790 7800 7810 7820 7830
 GAAGTTGGATCTTCTATTTCGTATGATGATTTACACGGCAGTGGGAGAAGAAGCTGACTTACACGCAAAAGTTCGCTCCGACGG
 E V W I F Y F * C R M I Y T A V G R R T S T Y T Q S C V R R
 K F G S S I S D V V * F T R Q L G E E R R L T R K V A S D G
 R S L D L L F L M S Y D L H G S W E K N V D L H A K L R P T

7840 7850 7860 7870 7880 7890 7900 7910 7920
 AGAGAGAAACATCCGGAACGCGCTCTTAATACCGGTTAGTCTTGAATATATAAAAATTTCTAATTTTTTCTTGTGCTGATGAGACGA
 R E K H P E L A S L I P L V L E I Y K N F * F F L A C M R R
 E R N I R N W R L * Y R * F L K Y I K I S N F F L L A * D E
E R R E T S G T G V F N T V S S * N I * K F L I F S C L H E T

7930 7940 7950 7960 7970 7980 7990 8000 8010
 ATTCATGATTATTTTCAGAAATTTGCGAGCGAACTATTGGGCAAAGAGAGGTATGCCGAAACAGAAAATTATCATTGGAATTCGACTTACA
 I H D Y F R N L Q R T I G Q R E V C R N R K L S L E F R L T
 F M I I S E I C S E L L G K E R Y A E T E N Y H W N S D L Q
 N S * L F Q K F A A N Y W A K R G M P K Q K I I I G I P T Y

8020 8030 8040 8050 8060 8070 8080 8090 8100
 GTCGGGGATGGACATTAAAGTAATTCITTCGGAACCGGCAATTGGAGCGGAGGGTGGTCCATCATGCCATCGACAAACGAATCCCGTGG
 V G D G H * V I L R K R Q L E R R V V V H H R H R Q R I P W
 S G M D I K * F F G N G N W S G G W S S I I A I D N E S R G
S R G W T L S N S S E T A I G A E G G R P S S P S T T N P V

8110 8120 8130 8140 8150 8160 8170 8180 8190
 CGGTACTGCGGCTTATTGGGAGGTTAAATTTAAAGTTCTCATTTACTTTTAAATACATTTCTTGGTGGTGGATGAATTTCTCAAATCCAT
 R Y C G L L G G L I * S S H L L L N T F L A A D E F L K F H
 G T A A Y W E V * F K V L I Y F * I H F L L L M N F S N S I
A V L R L I G R F N L K F S F T F K Y I S C C * * I S Q I P

8200 8210 8220 8230 8240 8250 8260 8270 8280
 TATTAATGAATGTAACCATTTTAAAGATATGCAAAATATGTGAAAGAAGGTGGCAAGGAAACGGTAGATAAAAAAGGTGTTGGAGCGTATAT
 Y * * M * P F * D M Q I C E R R W Q G N G R * K R C W S V Y
 I N E C N H F K I C K Y V K E G G K E T V D K K G V G A Y M
 L L M N V T I L R Y A N M * K K V A R K R * I K K V L E R I >

8290 8300 8310 8320 8330 8340 8350 8360 8370
 GGTAAAAGGASATCAGTGGCACGGTACGATAATGAGGAAACTATCAAATTAAGTAATTTTATTTGTTCTTGGATGTTATTGATAGTGA
 G K R R S V A R L R * * G N Y Q N * S N F Y C S * C Y * * *
V K G D Q W H G Y D N E E T I K I K V I F I V L D V I D S E
 W * K E I S G I V T I M R K L S K L K * F L L F L M L L I V

8380 8390 8400 8410 8420 8430 8440 8450 8460
 AAAAAAATAAATTTAAGAAATAACGTTGCAAAATATATTTTTTTTATTTTAAATCTAAATAGTTCAAATAAATTTCAATAAAGTTAACGA
 K K N N L R N N V A K Y I F F Y F * S K * F N K F H K V * R
 K K I I * E I T L Q N I F F F I F N L N S S I N F I K F N E
 K K K * F K K * R C K I Y F F L F L I * I V Q * I S * S L T

8470 8480 8490 8500 8510 8520 8530 8540 8550
 GAAGAAATTAATTTAAGAAATTTGCGGTAATACGGTTTAAATTAATTAATTAATTCAGTTTGTGTTTTATGTTTGAACCAACATCTAAAAAGT
 E E I N F R I I G V I T V Y * L N S V C V L C L K Q H L K S
 K K L I L E L L A * L R F I N * I Q F V F Y V * N N I * K V
 R R N * F * N Y W R N Y G L L I K F S L C F M F E T T S K K

8560 8570 8580 8590 8600 8610 8620 8630 8640
 GATTGGTTGCGCATGATTGAACATTTTCAGATGAGATGGTTAAAAAAGAACGGTTATGGCGGTGCTTTTATGTTGGACTCTTGATTTTGAIGA
 D W F A * L K L S D E M V K K E R L W R C L Y V D S * F * *
 I G S H D * N F Q M R W L K K N G Y G G A F M W T L D F D D
 * L V R M I E T F R * D G * K R T V M A V P L C G L L I L M

8650 8660 8670 8680 8690 8700 8710 8720 8730
 TTTCAAAGGTACAAGTTTGGCAAAGGTCCATATCCACTGTTAAATGCTATCAATAATGGGCTTGAAAGTGAGGTAAGACATGTTTCAA
 F Q R Y K L W Q R S I S T V K C Y Q * W A * K * G K T C F K
F K G T S C G K G P Y P L L N A I N N G L E S E V R H V S X
 I S K V Q V V A K V H I H C * M L S I M G L K V R * D M F Q

8740 8750 8760 8770 8780 8790 8800 8810 8820
 GAAATATTTCTAACAAATTCGTTTTCTTTCAATAAAGTTTGGATTAAATTTATGCGTGATATTTGTAATAAATTAATAAAAAAAGG
 E I F L T I L F F F Q * V W I * I M R D I C K * I N K K K R
 K Y F * Q F C F S F N K F G F K L C V I F V N K L I K K K G
 R N I S N N S V F L S I S L D L N Y A * Y L * I N * * K K K

8830 8840 8850 8860 8870 8880 8890 8900 8910
CGGATGGATGAATCANAATAAACTTTCTATGGGACCATACGGAAAAATGATTTTTATGAATTGTAGGATAAATAGAATTTCTGTCGGAAGA
R M D E S X * T F Y G T I R K N D F M N C R I N R I S V R R
G W M N X N K L S M G P Y G K M I L * I V G * I E F L S E D
A D G * I X I N F L W D H T E K * F Y E L * D K * N F C P K

8920 8930 8940 8950 8960 8970 8980 8990 9000
TTATGACTCACTAACTTTTGAATTAGCTGGATGGGACGTCAAATTTCTATATTGATCATGACGTTGATCAAACAAAATCCITAAAATTTTAAA
L * L T N F * L A G W D V N F Y I D H D V D Q T K S L N F K
Y D S L T F D * L D G T S I S I L I M T L I K Q N P * I L N
I M T H * L L I S W M G R Q F L Y * S * R * S N K I L K F *

9010 9020 9030 9040 9050 9060 9070 9080 9090
TTATCCCTTGAAGGTTAATTTATAAAAATTAACACACATTTTTAGCAAACACCATCAACAACAGCAGCACCAGAAACCCAGAGAAGCACT
L S L E S * F I K F N T H F * Q T P S T T A A P E T T E D T
Y P L K V N L * N L T H I F S K H H Q Q Q Q H R K P Q K T L
I I P * K L I Y K I * H T F L A N T I N N S S T G N H R R H

9100 9110 9120 9130 9140 9150 9160 9170 9180
GAAGTTGAAACRAATGACGGGAACCCGGGAACTGCAAAATGACACTGAAATGGAAACAACRAATATACCAGAAATTACAGAAGATACTGAA
E V E T M T E A P G T A N D T E M E T T N I P E I T E D T E
K L K Q * R K H R E L Q M T L K W K Q Q I Y Q K L Q K I L K
* S * N N D G S T G N C K * H * N G N N K Y T R N Y R R Y *

9190 9200 9210 9220 9230 9240 9250 9260 9270
ACTGAAACATCGGAAATGCCGGAACACTACAGATAAATACTGAAATGCCAGGAACCTGCAGATGATACTGAAACTGAAACATCGGAAATGCCG
T E T S E M P E T T D N T E M P G T A D D T E T E T S E M P
L K H R K C R K L Q I I L K C Q E L Q M I L K L K H R K C R
N * N I G N A G N Y R * Y * N A R N C R * Y * N * N I G N A

9280 9290 9300 9310 9320 9330 9340 9350 9360
GAACTACAGATAAATACTGAAATGCCGGAACACTGCAGATGATACTGACGAAACATCGGAAATGCCGGAACACTACAGATAAATACTGAAATG
E T T D N T E M P G T A D D T D E T S E M P E T T D N T E M
K L Q I I L K C R E L Q M I L T K H R K C R K L Q I I L K C
G N Y R * Y * N A G N C R * Y * R N I G N A G N Y R * Y * N

9370 9380 9390 9400 9410 9420 9430 9440 9450
CCAGGAACCTGCGGATGATACTGAAATGGAAACAACAGAATGCCAGAAATTACAGAGGATACTGAAACTGAAACATGGGAAATGCCGGAA
P G T A D D T E M E T T E M P E I T E D T E T E T W E M P E
Q E L R M I L K W K Q Q K C Q K L Q R I L K L K H G K C R K
A R N C G * Y * N G N N R N A R N Y R G Y * N * N M G N A G

9460 9470 9480 9490 9500 9510 9520 9530 9540
ACTACAGATGATATTGAAATGCCAGGAACCTGCAGATGATACTGAAACTGAAACATCGGAAATGCCGGAATTTACAGATAAATACTGAAATG
T T D D I E M P G T A D D T E T E T S E M P E I T D N T E M
L Q M I L K C Q E L Q M I L K L K H R K C R K L Q I I L K C
N Y R * Y * N A R N C R * Y * N * N I G N A G N Y R * Y * N

9550 9560 9570 9580 9590 9600 9610 9620 9630
CCGGGAACCTGCGGATGATACTGAAATGGAAACAACAGAATGCCAGAAATTACAGAGSATACTGACACTAAAACATCGACCCGATGCATCT
P G T A D D T E M E T T E M P E I T E D T D T K T S T D A S
R E L R M I L K W K Q Q K C Q K L Q K I L T L K H R P M H L
A G N C G * Y * N G N N R N A R N Y R R Y * H * N I D R C I

9640 9650 9660 9670 9680 9690 9700 9710 9720
GAACTACAGAAGATACTGACAAATGAAACATCGGAAATGCCGGAACACTACAGATAAATACTGCAATGCCAGGAACCTGCAGATGATACTGAA
E T T E D T D N E T S E M P E T T D N T A M P G T A D D T E
K L Q K I L T M K H R K C R K L Q I I L Q C Q E L Q M I L K
* N Y R R Y * Q * N I G N A G N Y R * Y C N A R N C R * Y *

9730 9740 9750 9760 9770 9780 9790 9800 9810
ACTGAGATATCGGAAATGCCGGAACACTACAGATAGTAGTGAATGCCAGGAACACTGCAGATGATACTGAAACTGAAACATCGGAAATGCCG
7 E I S E M P E T T D S S E M P G T A D D T E T E T S E M P >
L R Y R K C R K L Q I V V K C Q E L Q M I L K L K H R K C R
N * D I G N A G N Y R * * * N A R N C R * Y * N * N I G N A

9820 9830 9840 9850 9860 9870 9880 9890 9900
GAACTACAGATAATACTGAAATGCCAGGAACACTCGGATGATACTGAAATGGAACACAGAAATGCCAGAAATTACAGAAGATACTGAA
E T T D N T E M P G T S D D T E M E T T E M P E I T E D T E
K L Q I I L K C Q E L R M I L K W K Q Q K C Q K L Q K I L K
G N Y R * Y * N A R N F G * Y * N G N N R N A R N Y R R Y *

9910 9920 9930 9940 9950 9960 9970 9980 9990
ACTGAAGCATCAGAAATGCCGGAACACTACAGATAATACTGAAATGCCAGGAACACTCGGATGATGCTGAAATGGAACACAGAAATGCCA
7 E A S E M P E T T D N T E M P G T A D D A E M E T T E M P
L K H Q K C R K L Q I I L K C Q E L R M M L K W K Q Q K C Q
N * S I R N A G N Y R * Y * N A R N C G * C * N G N N R N A

10000 10010 10020 10030 10040 10050 10060 10070 10080
GAAATTACAGAGGATATTGAAACTGAAACATCGGAAATGCCGGAACACTACAGATAATACTGAAATGCCAGGAACACTGCAGATGATACTGAA
E I T E D I E T E T S E M P E T T D N S E M P G T A D D T E
K L Q R I L K L K H R K C R K L Q I I V K C Q E L Q M I L K
R N Y R G Y * N * N I G N A G N Y R * * * N A R N C R * Y *

10090 10100 10110 10120 10130 10140 10150 10160 10170
ACTGAAACATCGGAAATGCCGGAACACTACAGATAATACTGAAATGCCAGGAACACTCGGATGATACTGAAATGGAACACAGAAATGCCA
7 E T S E I P E T T D N T E M P G T A D D T E M E T T E M P
L K H R K F R K L Q I I L K C Q E L R M I L K W K Q Q K C Q
N * N I G N S G N Y R * Y * N A R N C G * Y * N G N N R N A

10180 10190 10200 10210 10220 10230 10240 10250 10260
GAAATTACAGAGGATATTGAAACTGAAACATCGGAAATGCCGGAACACTACAGATAATACTGAAATGCCAGGAACACTGCAGATGATACTGAA
E I T E D I E T E T S E M P E T T D N S E M P G T A D D T E
K L Q R I L K L K H R K C R K L Q I I V K C Q E L Q M I L K
R N Y R G Y * N * N I G N A G N Y R * * * N A R N C R * Y *

10270 10280 10290 10300 10310 10320 10330 10340 10350
ACTGAAACATCGGAAATGCCGGAACACTACAGATAATACTGAAATGCCAGGAACACTCGGATGATACTGAAATGGAACACAGAAATGCCA
7 E T S E M P E T T D N T E M P G T S D D T E M E T T E M P
L K H R K C R K L Q I I L K C Q E L R M I L K W K Q Q K C Q
N * N I G N A G N Y R * Y * N A R N F G * Y * N G N N R N A

10360 10370 10380 10390 10400 10410 10420 10430 10440
GAAATTACAGAAGATACTGAAACTGAAAGCATCAGAAATGCCGGAACACTACAGAGGGGGCTGAAACTGAAACACAGAACCGAAGCATCAGAA
E I T E D T E T E A S E M P E T T E G A E T E T T T E A S E
K L Q K I L K L K H Q K C R K L Q R G L K L K Q R P K H Q K
R N Y R R Y * N * S I R N A G N Y R G G * N * N N D R S I R

10450 10460 10470 10480 10490 10500 10510 10520 10530
ACTACGGAAACTATTAAACCCCAAAAACCTTCTGGTTTGTATTTTCAAGTTAAAARAGCCACTTTTCTGCTTTCTGATTTGGAA
T T E T I K P T K K P S G L L F S S * K S H F S V F L I W K
L R K L L N P P K N L L V C Y F Q V K K A T F L S F * F G K
N Y G N Y * T H Q K T F W F V I F K L K K P L F C L S D L E

10540 10550 10560 10570 10580 10590 10600 10610 10620
ATATACAGTCACCTAAGTTTACTTTTTATGCACTTTAGACAGAACAAAAATACTTACATACAGTTGAAARAGAAGCCTTAAGAAATGTTCA
I Y S H L S L L F M H F R Q N K K Y L H T V E K K L K K C S
Y T V T * V Y F L C T L D R T K N T Y I Q L K R S L R N V Q
N I Q S P K F T F Y A L * T E Q K I L T Y S * K E A * E M F

10630 10640 10650 10660 10670 10680 10690 10700 10710
GTACTCTGAAAGCTAAATGACTCTTCCCAAAGTAAATGTTTACTAAAATTTGCGAAGTACATCTGATATTAATCGTAGAAAAATTAITTCG
V L * K L N D S S Q S N V Y * N F A S T S D I N R R K I I S
Y S E S * M T L P K V M F T K I S Q V H L I L I V E K L F R
S T L K A K * L F P K * C L L K F R K Y I * Y * S * K N Y F

10720 10730 10740 10750 10760 10770 10780 10790 10800
AATATTGATATCAGAAAACCCGAAATGGTCCATAATGAGTCCGTTGAAATTTGAAGAATACITAAATGCATTTTTTTTTTAGTATTGAGC
N I * Y Q K T R N G P * * V R * N * R I L K C I F F L V L S
I F D I R K P E M V H N E S V E I E E Y L N A F F F * Y * A
E Y L I S E N P K W S I M S P L K L K N T * M H F F F S I E

10810 10820 10830 10840 10850 10860 10870 10880 10890
AATGAAATCATGAAAACATGTCAGTAACTGCATAGTTAAATTTGGGATTTGTACTGTATTGAACATAACATCGTTTTATGCTTTGTGTAA
N * I M K T C Q * L H S * I W D L Y C I E H N I V L C F V *
T E S * K H V S N C I V K F G I C T V L N I T S F Y A L C K
Q L N H E N M S V T A * L N L G F V L Y * T * H R F M L C V

10900 10910 10920 10930 10940 10950 10960 10970 10980
AACCGAATCCAAAAAATACGAAAGGAAAAATTTGTTGTTCTATTTCAAATTTGAGCATAGTCTATATGGCAATGTGGCAAAAAAGACGT
N R I Q K N T K G K I V V L F Q I S S I V Y M A M W Q K R R
T E S K K I R K E K L L F Y F K F R A * S I W Q C G K K D V
K P N P K K Y E R K N C C S I S N F E H S L Y G N V A K K T

10990 11000 11010 11020 11030 11040 11050 11060 11070
CATGGTCTTTTTAAGAAAAAATAATTTACCGAATCAGAATTCAAATTTGGTGTAAAGATTTTGAAAAATGAAAAAATGAGGAGAAAAATTA
H G L F K K K L I Y R N Q N S N W C K I L K M K K M R R K L
M V F L R K N * F T E I R I Q I G V R F * K * K K * G E N *
S W S F * E K T N L P K S E F K L V * D F E N E K N E E K I

11080 11090 11100 11110 11120 11130 11140 11150 11160
GTCCAGTATTATTACAGAAATCTTTAAAAAGAAAGTTTTGGAATTTACCGATGGAAAAATATTTAGCAAAATAGTCAAAATTTGCTGAGAAC
V P V L L Q K S L K R K F W N Y G W K I F * Q I V K I A E N
S Q Y Y Y R N L * K E S F G I T D G K Y F S K * S K L L R T
S P S I I T E I F K K K V L E L R M E N I L A N S Q N C * E

11170 11180 11190 11200 11210 11220 11230 11240 11250
AATATCTGGACTTTACATGCATAAAAAGAGAAATTTACCGGTTACAAATCACTGTTAATTGAGTAAGATTGATCGGAAATGACGATTAT
N Y L D F H M H K R E I I T V T I T V N * V R L I G N D D Y
I I W T F T C I K E K L S R L Q S L L I E * D * S E M T I I
Q L S G L S H A * K R N Y H G Y N H C * L S K I D R K * R L

11260 11270 11280 11290 11300 11310 11320 11330 11340
AAATTAGCGAATGGATATTAATTGAACAAACCTAAACAAAAGGAATAACTAAGCAGGAATAAGCAAAATGGATGTATAAGCAACAATGGG
K L A E W I L I E Q T * T K G I T K Q E * A N G C I S N N G
N * P N G Y * L N K P K Q K E * L S R N K Q M D V * A T M G
* I S R M D I N * T N L N K R N N * A G I S K W M Y K Q Q W

11350 11360 11370 11380 11390 11400 11410 11420 11430
AAATGACAATTTACAGAGTTGAAGAAATCGACTGAAAAAGTCAGAAATTAGACGATGGCAATAAATGCTCAATCCAATAACCATGCTTAAT
K * T I T E L K K S T E K V R I R R W Q * M L N P I T M L N
N E Q L Q S * R N R L K K S E L D D G N K C S I Q * P C L I
E M N N Y R V E E I D * K S Q N * T M A I N A Q S N N H A *

11440 11450 11460 11470 11480 11490 11500 11510 11520
AAAGCCTAATTTCAATAATTTATGCTTATGCTGAAATAAATACGAAAAATTTATGCCATACCGACAAAATTAATCATAATCTTTGAGAAAT
K A * F N N Y A S Y A E I I R K I M P Y R Q N Y S * S F R N
K P N S I I M L L M L K * Y E K L C H T D K I T H N L S E M
* S L I Q * L C F L C * N N T K N Y A I P T K L L I I F Q K

11530 11540 11550 11560 11570 11580 11590 11600 11610
GAGACAGAAATTTTGGATTGCCAAAAAATAATATCGAAGTTAANTGAGCCCTTAAAAGCCGATATTGTTTATCTCTCTTTACTACATATAAT
E T E I L I A K K * Y R S * M S L K S R Y C L S L F T T Y N
R Q K F * L P K N N I E V K * A L K A D I V Y L S L L H I I
* D R N F D C Q K I I S K L N E P * K P I L F I S L Y Y I *

11620 11630 11640 11650 11660 11670 11680 11690 11700
TTATCTCTCTTTTGTAGTCCAAAAAATCTTCTAATCTTACGATAGATAATCTAGAAAACAATGAATTATCTTGTATGATATTGAAATCTAT
L F F F L V Q K T F * S Y D R * S R N N E L S C Y D I E I Y
Y S S F * S K K L S N L T I D N L E T M N Y L V M I L K S I
F I L L F S P K N F L I L R * I I * K Q * I I L L * Y * N L

11710 11720 11730 11740 11750 11760 11770 11780 11790
TTCAGAGITGAAATGTCCCGGCCCTTCAGGTCTGTTCCGCATCCCGACGATTGCCATTGTTCCCTTCATTGTGCACACAACTATCCACA
F R V E M S P A F R S V S A S R R L P F V P S L C T Q L S T
S E L K C P R P S G L F P H P D D C H L F L H C A H N Y P H
F Q S * N V P G L Q V C F R I P T I A I C S F I V H T T I H

11800 11810 11820 11830 11840 11850 11860 11870 11880
TGTGATGGAATGTCCGGCAGCAACTTTCTTCAATGAGAAATACAAAGTTTGGCATCATCAGAGAAATGCTCCAGAAGGATGCGTATGATT
C D G M S G S N F L Q * E I Q S L R S S E K C S R R M R M I
V M E C P A A T F F N E K Y K V C D H Q R N A P E G C V * F
M * W N V R Q Q L S S M R N T K F A I I R E M L Q K D A Y D

11890 11900 11910 11920 11930 11940 11950 11960 11970
TGTATTATTTCCATGATAATATATGCAATTGTACATGTTGTGTAATAAATAAGAATTTGAAGTAAATTTTTTGGCAGCACAATGAA
C I I S M I I Y A I V H V V V K K I R I * S K I F W Q H N E
V L F P * * Y M Q L Y M L L * K K * E F E V K F F G S T M N
L Y Y F H D N I C N C T C C C K K N K N L K * N F L A A Q *

11980 11990 12000 12010 12020 12030 12040 12050 12060
TGGTACTTACTCGTATGAAAATTTGTGATATGTTCAAAAAATTATAAAGATAGAAGTAAAGAAATGAGTCAACTGAATCAITTAATG
L V L T R M K I C D M F K K L * R * K * R N E S T E S F K M
W Y L L V * K F V I C S K N Y K D R S K E M S Q L N H L K *
I G T Y S Y E N L * Y V Q K I I K I E V K K * V N * I I * N

12070 12080 12090 12100 12110 12120 12130 12140 12150
AAGATATTTGTTATTTGCAATTTCTTACATCTCAGCAATCCATAAATAACTTCAGATAGACATTAAATGCATTCAACGACGGTAATGGA
K I F V I C I S S H L S K S I N N F R * T L M H S T N G N *
R Y L L F A F L H I S A N P * I T S D R H * C I Q R T V I D
E D I C Y L H F F T S Q Q I H K * L Q I D I N A F N E R * L

12160 12170 12180 12190 12200 12210 12220 12230 12240
TAATCAAAAAAAGTAGAGATAAATGCTTAAAAATACCCTAGCTTTGCACCATAGTTAAAAATTTTACTCAAGAAATCAATCCATT
* S K K K K * R * M L K N T T S F A P * V K I L L K N Q S I
N Q K K K S R D K C L K I P L A L H H K L K F Y S R I N P L
I I K K K K V E I N A * K Y H * L C T I S * N F T Q E S I H

12250 12260 12270 12280 12290 12300 12310 12320 12330
GAAAAATTTTCCATTTCCGATACTTTTAGCTGTGACATCAGCTAATACATAATGGTTTGAAGCCACTTCCATGTACATTTTGAAT
E K F I P F P I L L A V T S R * Y I M V * S H L P C T F * N
K N L F H F R Y F * L * H H A N T * W F E A T F H V H F E I
* K I Y S I S D T F S C D I T L I H N G L K P P S M Y I L K

12340 12350 12360 12370 12380 12390 12400 12410 12420
TTGCTGTTTATGAAATAATTTCTACTCTCTTTTCTAGCGCCCTGAATTTGCAAAAAAATAATTTCTCCATATTATTCACCTCATATTTTCTT
L L F M K * F L L S F L S A * I A K K * F L H I I H S Y F L
C C L * N N F Y S L F L A P E L Q K N N F S I L P T H I F F
F A V Y E I I S T L F S * R L N C K K I I S P Y Y S L I F S

12430 12440 12450 12460 12470 12480 12490 12500 12510
CGGCAGCTTCCTGGAATGTAACACATTCTCTTTCTCTCTACGACTATTCGGCCTAAATTTTTCTCTTTCTCTATGACTATTGGC
R Q L P G M * N I P F L F Y D Y S A * F F S F F P L * L F G
G S F L E C K T F L F S S T T I R P N F F P F F L Y D Y S A
S A A S W N V K H S F S L L R L F G L I F F L F S S M T I R

12520 12530 12540 12550 12560 12570 12580 12590 12600
TTAAAAATTATAATAGAAATGATGTGAGCACATATGTCAAGTCAACGGATAGTGAATTTCTGCTGTTTTCTGTTTCAAGTATAAAATGAA
L K I I I E M M * A H M S S Q R I V K F C S F S V S R I N E
* K L * * K * C E H I C Q V N G * * N S A R F L F H V * M K
L K N Y N R N D V S T Y V K S T D S E I L L V F C F T Y K *

12610 12620 12630 12640 12650 12660 12670 12680 12690
GAAATGTTACGCCACATCTTTTTATTATCCATTGACGATCAATATTACCGAAAATAAAAGTTATTGGTAAATTGAGAATTGAAGCATTAT
E M L R H I F L L S I * R S I L P K I K L L V N * E L K H Y
K C Y A T S F Y Y P F D D Q Y Y R K * S Y W * I E N * S I I
R N V T P H L F I I H L T I N I T E N K V I G K L R I E A L

12700 12710 12720 12730 12740 12750 12760 12770 12780
TAATTAGCTCATTTCATCGTTTCAACAAGGATTACGGGACAAAATTTCAAGAAATCAAAAAGTGAATTTCTATTGTTTTCTATTTCACG
* L A H S S F I N K D Y G T N F K K S K S E I L F V F Y F T
N * L I H R S S T R I T G Q I S R N Q K V K F Y S F S I S R
L I S S F I V H Q Q G L R D K F Q E I K K * N S I R F L F H

12790 12800 12810 12820 12830 12840 12850 12860 12870
TATAAAATATCACACAGAAATGAATTTCTATCCAAACATGAGGCTCGAAATTTGAATGTATATGAAATTGAGCACAAATTTGGTGGGAACA
Y K I S H R N E F Y P N M R L E I * N V Y E I E H N L V R T
I K Y H T E M N S I Q T * G S K F E M Y M K L S T I W C E Q
V * N I T Q K * I L S K H E A R N L K C I * N * A Q F G A N

12880 12890 12900 12910 12920 12930 12940 12950 12960
ATATATTCTGATGTCGATAACTCTGCTTCTGCTTTAGCATGCAAAATTCATTTCCGATATTAATTAGCATTTTTTTTTTATAATATGATTAA
I Y S D V D N S C L L A C K I H F R Y * L A F F F Y N M Y *
Y I L M S I T P A F * H A K F I S D I N * H F F F I I C I N
N I F * C R * L L P F S M Q N S F P I L I S I F F L * Y V L

12970 12980 12990 13000 13010 13020 13030 13040 13050
TGGAAGAGTCTATCTTAAAAATTTTTACTTCTTATTTTTACTTACTGTAAAGTTTTTTAGTTTGTGGGGGGGGGGGGGAAAACGT
W K K S I L K I F Y F L F F T Y C K V F * F V G G G G G K R
G R S L S * K F F T S Y F L L T V K F F S L W G G G G E N V
M E E V Y L K N F L L L I F Y L L * S F L V C G G G G G K T

13060 13070 13080 13090 13100 13110 13120 13130 13140
TTATACGAAACAAATCAAAATTTGATAAAGATTGATTTGCTTATGGAGAAGTTTTTGTITTTAGGACGTTACACACCTGTCAAAATAAAG
L Y * T I K Y L I K I D L L M E K F L F * D V T H L S K * K
Y T E Q S N I * * R L I C L W R S F C F R T L H T C Q N K S
F I L N N Q I F D K D * F A Y G E V F V L G R Y T P V K I K

13150 13160 13170 13180 13190 13200 13210 13220 13230
TAATCAACTAATAGCTAACAAGAATTATGTGTACTTAATATTTTTATTATTATCTCCAAAATTTATCTCCACACATATGCAAAATGAAT
* S T N S * Q E L C V L N I F I I I L Q K L F S T H M Q M N
N Q L I A N K N Y V Y L I F L L L F S K N Y S P H I C K * I
V I N * * L T R I M C T * Y F Y Y Y S P K I I L H T Y A N E

13240 13250 13260 13270 13280 13290 13300 13310 13320
TTATTTATGATCAAGCGGAGCTTCAATATTTCTAACGATTCCTATCAGTGGTGGTGTACAAATGTACGGAATTTTCAGCATTATGA
L F M I K G E L Q L F L T I P I T G W C V Q C T E F S A L *
Y L * S K A S F N Y F * R F L S L D G V Y N V R N F Q H Y D
F I Y D Q R R A S I I S N D S Y H W M V C T M Y G I F S I M

13330 13340 13350 13360 13370 13380 13390 13400 13410
 TAAAAGATATGATGAAAACCTTTTAATAATAAAAACCTCTCGACAAATATAATAGCATTATTACTTATATTTTTCCAGCTTTCCGCTTAATT
 * K I * * K L L I I K T L D K Y N S I I T Y I F P A F R L I
 K R Y D E N F * * * K L S T N I I A L L L I F F Q L S V * F
 I K D M M K T F N N K N S R Q I * * H Y Y L Y F S S F P S N

13420 13430 13440 13450 13460 13470 13480 13490 13500
 TTAATGTTTTTTCAGCTTTCGAGTTATTTGTTTTTTCAGCTTTCAGCTATTCTTGTTTTTTCIACTTCTTAATATTATTATCTC
 L M F F Q L S S Y L F F S S F P A I S C F F * L L N I I Y L
 * C F F S F R V I C F F P A F Q L F L V F S N F L I L F I S
 F N V F S A F E L F V F F Q L S S Y F L F F L T S * Y Y L S

13510 13520 13530 13540 13550 13560 13570 13580 13590
 ATTCCAACTTCGGATTTTAATTAATCAATTTAATTAATCTAATATCTTCTAATCAAAAATATTTTTCAATTACATGACTTGTAGCTGAAAG
 I P T S D F N * S I * L I * Y L L I K N I F H Y M T C S * K
 F Q L R I L I N Q F N * S N I F * S K I F F I T * L V A E R
 H S N F G F * L I N L I N L I S S N Q K Y F S L H D L * L K

13600 13610 13620 13630 13640 13650 13660 13670 13680
 GCCCTTGCATATGAAAAGTTTTGTTTGGGGTTTATAGGACAATTATCAGAGTCTTATCAATAATACAGTAGATTACTTTTCGATACTC
 A L A Y E K V L F G V Y R T I I R V L S I I Q * I T F S I L
 P L H M K K F C L G F I G Q L S E S Y Q * Y S R L L S R Y S
 G P C I * K S F V W G L * D N Y Q S L I N N T V D Y F L D T

13690 13700 13710 13720 13730 13740 13750 13760 13770
 GAAAAATGTTGTTTTTTCCTATTCATCGGAGTGAATTTTTTGAAGCATATGATTGTTTGTGTTGATTGACAAAATACTTTGCGAG
 E K C C F F P I F S S E * N F F E S I * L F V * L T N T L Q
 K N V V F F P F S H R S E I F L K A Y D C L F D * Q I L C R
 R K M L F F S H F L I G V K F F * K H M I V C L I D K Y F A

13780 13790 13800 13810 13820 13830 13840 13850 13860
 AATGCTTATTGAATATTTCTGGAAAAAGCCCTGATCCGAATTCGTTCCCTTTAGTGAGGGTTAATTCGGCGCCGCTCTAGAACTAGTGG
 N A Y L N I L E K S L I R I R S L * * G L I P R P L * N * W
 M L I * I F W K K A * S E F V P F S E G * F R G R S R T S G
 E C L F E Y S G K K P D P N S F P L V R V N S A A A L E L V

13870 13880 13890 13900 13910 13920 13930 13940 13950
 ATCCCCGGGCTGCAGGAATTCGATATCAAGCTTATCGATACCGTCGACCTCGAGGGGGGGCCCGGTACCCAATTCGCCCTATAGTGAGT
 I P R A A G I R Y Q A Y R Y R R P R G G A R Y P I R P I V S
 S P G L Q E F D I K L I D T V D L E G G P G T Q F A L * * V
 D P P G C R N S I S S L S I P S T S R G G P V P N S P Y S E

13960
 CGTATTACAATTC
 R I T I X
 V L Q F
 S Y Y N S

Appendix 2: Insert of Clone 9; Sequence Range: 1 to 13753

Gene II: 3952...8592

Gene III: 12096...13750

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      10      20      30      40      50      60      70      80      90
CAGTGAATTGTAATACGACTCACTATAGGGCGAATTGGGTACCGGGGCCCCCTCGAGGTCGACGGTATCGATAAGCTTGTATCGAATT
Q * I V I R L T I G R I G Y R A P P R G R R Y R * A * Y R I
S E L * Y D S L * G E L G T G P P L E V D G I D K L D I E F
V N C N T T H Y R A N W V P G P P S R S T V S I S L I S N

      100     110     120     130     140     150     160     170     180
CCTGCAGCCCGGGGATCCACTAGTTCTAGAGCGGCGCGGAATTAACCCCTCACTAAAGGGAAACGAATTCGGATCAAAAAATCGACGAA
P A A R G I H * F * S G R G I N P H * R E R I R I K K I D E
L Q P G G S T S S R A A A E L T L T K G N E F G S K K S T K
S C S P G D P L V L E R P R N * P S L K G T N S D Q K N R R

      190     200     210     220     230     240     250     260     270
ATTAATTGATTATGATATCAGTACAACGTAAGTATGATACATATAATCATATAATATAATCACAATAATATAATCACACAATAT
I N * L * Y Q Y N * R R * S Y N I I I * Y N H I N I I T Q Y
L I D Y D I S T T N V D N H T I * S Y N I I T * I * S H N I
N * L I M I S V Q L T * I I I Q Y N H I I * S H K Y N H T I

      280     290     300     310     320     330     340     350     360
CTATAATCTCTACATCCCAATTTGATCTTTGATGCGCAGGTTCTCAAAAAGGACAAAAAAGCTTTGAATCACTAAATGATGCCGAAA
L * Y L Y I P I V F D A Q V L K K D K K K A L N H * M M P K
Y N I S T F P L S L M R R F S K R T K K K L * I T K * C R K
S I I S L H S H C L * C A G S Q K G Q K K S F E S L N D A E

      370     380     390     400     410     420     430     440     450
AACATTAAATTAATAGATTAAAGTATACCTCTTTTGGGACAGTAATAAAAAATGACAATGTTCTGTAGGTAAGAAATAGTCAACTG
N I K L I R F K S I P L F G T V I K N D N V P V G K N S Q L
T L N * * D L K V Y L F L G Q * * K M T M F L * V R I V N *
K H * I N K I * K Y T S F W D S N K K * Q C S C R * E * S T

      460     470     480     490     500     510     520     530     540
ATTTCATGTTTCATCGTCATAACTATCTCGATCATCGTAGATTGTATTTCGATCATCGTTGTCACCTCTCCATAAGCGTCTCAACATCTTTC
I H V H R H N Y L D H R R L Y F D H R C H L S I T L S T S F
F M F I V I T I S I I V D C I S I I V V T S P * R S Q H L S
D S C S S S * L S R S S * I V F R S S L S P L H N A L N I F

      550     560     570     580     590     600     610     620     630
TCTAACGAATGTATGGTACAATGTTATTATAATAATAAATAAAGCATAAATAAGTTATACTATAAAAAATGTTTATGTTTTGATCG
S N E C M V T M L L * * * I I K A * I S Y T I K M F M F * S
L T N V W L Q C Y Y N N K * * K H K * V I L * K C L C F D R
L * R M Y G Y N V I I I I N N K S I N K L Y Y K N V Y V L I

      640     650     660     670     680     690     700     710     720
ACTTATTAGGAAAAATCCCGGATTAGAACCCCTCAATCGTGTAAAGAAAACATCCACTCCTGACGTGCTAATAGGCAATATATTTTCTAC
T Y * E K S R I R T L N R V R K T S T P D V L I G N I I F Y
L I R K N P G L E P S I V * E K H P L L T C * * A I L F S T
D L L G K I P D * N P Q S C K K N I H S * R A N R Q Y Y F L

      730     740     750     760     770     780     790     800     810
AATATAATATGGCCCTATCATGAAACTATCATCTAAATTTGAAAAGCAACGTTTGTGGAAATGCAAAATACAAAACGGCAATATAAGTA
N N N M A L S * K L S S * I E K Q R L L E M Q I Q T A I * V
I I I W P Y H E N Y H P K L K S N V C W K C K Y K R Q Y K *

```


L Q * N F K L Y F K P I L K I C H I R K M S I N K L S T L S

2620 2630 2640 2650 2660 2670 2680 2690 2700
TCTTCTTAATTAATTTACTTTCAACTATTATTCACTACTTTTTTCCCAATTCCTTTTTGTTTATCGTTTATATCATTTAAATAATTT
S S Y L I F T F N Y Y S L L F F P I L F C L S F I S F K * F
L L I * Y L L S T I I H Y F F F Q F F F V Y R L Y H L N N F
F F L S N I Y F Q L L F T T F F S N S F L F I V Y I I * I I

2710 2720 2730 2740 2750 2760 2770 2780 2790
TATATTCAGTTGAATGTTTCTAAAATTTTCAACTGTTTTTAAAAAATTTAATAATGAAAATGATCGTACTTAGGTATGACGATAATCA
Y I Q V E C F * N F Q L F F K K F N N E N D R T * V * R * S
I F K L N V S K I F N C F L K N L I M K M I V L R Y D D N Q
L Y S S * M F L K F S T V F * K I * * * K * S Y L G M T I I

2800 2810 2820 2830 2840 2850 2860 2870 2880
ATTTCAITTAGCATATTTCCGTTAACTAATATTTTCAGATTTTTTTTTCCITATATTTATATAAAGCGCAGGTAAAAAACTGGTTAAC
I S F S I F P L T N I S D F F F L I F I * S A G K K N W L T
F H L A Y F R * L I F Q I F F S L Y L Y K A Q V K K T G * P
N F I * H I S V N * Y F R F F F P Y I Y I K R R * K K L V N

2890 2900 2910 2920 2930 2940 2950 2960 2970
TTGTTAATAAATAAATAATATTTGGGAAATTTTGATATATTTGAGTTAACAAAAAGAAGCTACATTTTAAAGGAAGACAGATGCTAA
L L I N K I Y L G N F D I F E L T K K K L H Y L R K T D V *
C * * I K Y I W E I L I Y L S * Q K R S Y I I * G R Q M S N
L V N K * N I F G K F * Y I * V N K K E A T L F K E D R C L

2980 2990 3000 3010 3020 3030 3040 3050 3060
CAAATATTTTATAGTAAATATTTAATGATGCAGAAGTGGACATAATTAACGGTTAAGTGGTATATATTTATACATATATGATGAATAA
Q I F Y S K Y L M M Q K W T I I N G * V V Y I Y T Y M Y E *
K Y F I V N I * * C R S G Q * L T V K W Y I F I H I C M N N
T N I L * * I F N D A E V D N N * R L S G I Y L Y I Y V * I

3070 3080 3090 3100 3110 3120 3130 3140 3150
TTACACGATGGTACTATTAACITTTIATTAACITTTGGTGTACAATTTTCCITATACAGGTAAGTTAATTTTTTAATTTCTAGCACITGT
L H D G T I N F Y Y I W C Y N F S Y T R * V N F L I S S T C
Y T M V L L T F I T F G V T I F P I Q G K L I F * F L A L V
I T R W Y Y * L L L H L V L Q F F L Y K V S * F F N F * H L

3160 3170 3180 3190 3200 3210 3220 3230 3240
TTAACATTAATTTTACTAAGTTGCAAAAAAAGGGGATAATTTGAAGTAATGTAAGAAATGTTAGCTAGACTTTTCTATGTAATAATATTA
L T L N F T K L Q K K A I I * S N V R N V S * T F L C K I L
* H * I L L S C K K R R * F E V M * E M L A R L F Y V K Y *
F N I K F Y * V A K K G D N L K * C K K C * L D F S M * N I

3250 3260 3270 3280 3290 3300 3310 3320 3330
GAAAGTGAATACTAAATTTGATAGTATAAATAACAAGCAAAATTTTCTCATTTCATCGTTTATCTATCATACCCAGTAATATCAAATGAACG
E S V I L N C S I K Y K Q I F L I H R L S I I P S N I K * T
K V * Y * I V V * N T S K F F S F I V Y L S Y P V I S N E R
R K C N T K L * Y K I Q A N F S H S S F I Y H T Q * Y Q M N

3340 3350 3360 3370 3380 3390 3400 3410 3420
AAATCTTGAAGCCACTTCTCTCATTATCAAAATAGTGATAATTCACTTCAATTTATCCAAACATCTTGATACCTCTTTTTAATCATTATA
K S * N P L L S L S K * * * F T S I I P T S * Y L F L I I I
N L E T H F S H Y Q N S D N S L Q L F Q H L D T S F * S L *
E I L K P T S L I I K I V I I H F N Y S N I L I P L F N H Y

3430 3440 3450 3460 3470 3480 3490 3500 3510
AAAATTTTGATAACTCGATTTAATCTTTATTGAATCTTAGTAATCGCTTTAATCGCTATCAAAATCTTGATAACTCGTTTCAATCAAT
K I L I T R F N L Y * N L S N R F N R Y Q N L D N S F Q S I
K F * * L D L I F I E I L V I A L I A I K I L I T R F N Q F

K N F D N S I * S L L K S * * S L * S L S K S * * L V S I N
3520 3530 3540 3550 3560 3570 3580 3590 3600
TTGGTAAATTTAATCATTGCTAACATATTTTTGATAATGATTGAAGAAGATGATATATAATAGCAGTTGCATGTGATTGCTATTATCGTAA
L V I * S L L T Y F * * * L K K M I Y N S S C M * L L L S *
W * F N H C * H I F D N D * R R * Y I I A V A C D C Y Y R N
F G N L I I A N I F L I M I E E D D I * * Q L H V I A I I V
3610 3620 3630 3640 3650 3660 3670 3680 3690
TTATATACACTTAGTGAATAAACGAAAAAATGGGATTTATGATGGAAGATTTGAAAAAGATAGCGAAAAATTACTGTGCGATTTCGATT
L Y T L S E * T K K W D L * W K I * K K I A K N Y C R I S I
Y I H L V N K R K N G I Y D G R F E K R * R K I T V A F R F
I I Y T * * I N E K M G F M M E D L K K D S E K L L S H F D
3700 3710 3720 3730 3740 3750 3760 3770 3780
CCITTTATGATACTTGTAAATTAATCAATGCATCTTATCTTGCACCTCATATACTTCAATCACTATAGIATAACATAAATATGTAGGAAA
P F M I T S L I I N A S Y L A P H I L Q S L * Y N I I C R K
L L * * L R * L S M H L I L H L I Y F N H Y S I T * Y V G N
S F Y D N F V N Y Q C I L S C T S Y T S I T I V * H N M * E
3790 3800 3810 3820 3830 3840 3850 3860 3870
TGTCACATAACATCAAAAGCAAAATGCATTTTTTAAATTAATAAAAAAAGTGTGTTGAAAAAGTAATGAAAAATTAATCATAAATAAATTT
C H I T S K D K C I F * I K K N C V E K V M K N * I I N K F
V T * H Q K T N A F F K L K K T V L K K * * K I K S * I N L
M S H N I K R Q M H F L N * K K L C * K S N E K L N H K * I
3880 3890 3900 3910 3920 3930 3940 3950 3960
GAATAAAATATCAATTAATAAATAAATTTTTGGAGTAACAATAAATTTGGGAAATGCGCAAATTTAATGACACTGTTTAGCAGTGAAT
E * K Y Q L K * N F G V N K K I W E N A Q I * * H C L A V N
N K N I N * N K I L E * T K K F G K M R K F N D T V * Q * I
* I K I S I K I K F W S K Q K N L G K C A N L M T L F S S E
3970 3980 3990 4000 4010 4020 4030 4040 4050
TATGCGCAATTTTTGACATTTGTCGAAATACGTTTCGCGGATGTTTATTACACCAATTTGGGCTCAATATCGATATGGTTTCAGTGATGTT
Y A A I F A H L S K Y V R G C Y Y T N W A Q Y R Y G S V M F
M P Q F L H I C R N T F A D V I T P I G L N I D M V Q * C L
L C R N F C T F V E I R S R M L L H Q L G S I S I W F S D V
4060 4070 4080 4090 4100 4110 4120 4130 4140
GAAATAATTTCAATTTTTAATAAATAAATAGITGAATAGACTCTAAGGAATGTTTACATTTTTTGAATAAATAAATAGAAAGCTTATGTAAA
E I I S F L K L N S * N R L * G M F T F F E * K L E S L C K
K * F H F * N * I V E I D S K E C L H F L N K N * K A Y V K
* N N F I F K I K * L K * T L R N V Y I F * I K I R K L M *
4150 4160 4170 4180 4190 4200 4210 4220 4230
GAACTTAAATCGAAAAAAGCAAGCTCTAAGCCATATAAATAATGAGTAGAAAAATAAATAATTTTCAGAAAAATTAATAAATGACAGTTTG
E L K S K K S K L * A I * N M S R K * K F S E N * I I D S L
N L N R K K A S S K P Y K I * V E N K N F Q K I K * L T V *
R T * I E K K Q A L S H I K Y E * K I K I F R K L N N * Q F
4240 4250 4260 4270 4280 4290 4300 4310 4320
ACTTCCATTTTTAGTTCCTGATTAGTTCCTTACATTAATCTCTTTTCGTAATGATATATCTTGAATATACATCATTTCGACGTGTTTCTT
T S I F * F L I S S Y I N S L S * * Y I L N I H H F D V F L
L P F F S S * L V L T L I L F R N D I S * I Y I I S T C F L
D F H F L V L D * F L H * F S F V M I Y L E Y T S F R R V S
4330 4340 4350 4360 4370 4380 4390 4400 4410
AGCACCGTAAACTAATGTGGCGCAAGAAAAAAGTAACCAAGTTAAGTAAAAAAGGATTTGCTTCTACTAGTAAGTGAAGTTGCAACAC
S T V N * C G A R K K * P K L S K I R I V L L L V S E L Q H
A P * T N V A Q E K S N Q S * V K * G L C F Y * * V S C N T

* H R K L M W R K K K V I K V K * N K D C A S T S K * V A T

4420 4430 4440 4450 4460 4470 4480 4490 4500
TGATCGAIGTAATATGCATTTTCATGTTAAAGGAGAAGGCARATTTCTTACCGGAGGATATTCCAAAAGGCCCTGTGCACTCACATTTCTTTA
* S M * Y A F S C * R R R Q I L T G G Y S K R P V H S H S L
D R C N M H F H V K G E G K F L P E D I P K G L C T H I L Y
L I D V I C I F M L K E K A N S Y R R I F Q K A C A L T F F

4510 4520 4530 4540 4550 4560 4570 4580 4590
TGCATTGCCCCAAGTGAACAGAAAGGCACGTCATTGTAAAGTTGTAATAATATTTCAGTGAAGAAATATGCTCGAAAATTTTATTGGGC
C I R Q S E P E R H V I V S C K * Y L Q * E I C S K F L L G
A F A K V N Q K G T S L * V V N N I C S E K Y A R N F Y W A
M H S P K * T R K A R H C K L * I I F A V R N M L E I F I G

4600 4610 4620 4630 4640 4650 4660 4670 4680
TATTCATATGTAATCGTGAATAATTTTGTATTGAGTAGTTTTTGTTCATAGACAAAATTTGTTGTAAGAGTTAATCTTTTGTGGAAGATT
Y S Y V S * K I F D * V V F V S * T K I V V K S * S L W K I
I H M Y R E K F L I E * F L F H R Q K L L * R V N L C G R L
L F I C I V K N F * L S S F C F I D K N C C E E L I F V E D

4690 4700 4710 4720 4730 4740 4750 4760 4770
AAITGGTAATACAGTATGACAAATTTTGTAGTACGAAAATTTCTTGTATTATGAGATTGAAATAATCATTTTCTAAGTTGGACGGAAATTTGAAC
N G N T V * Q F * * R K F L D Y * D * N N H F * V G R N L N
M V I Q Y D N F S D E N F L I I E I E I I I S K L D G I * T
* W * Y S M T I L V T K I S * L L R L K * S F L S W T E F E

4780 4790 4800 4810 4820 4830 4840 4850 4860
GGAATTTGATTGAATCTCCGACCCAGITCCGAACAGTAATTAATCGCGCAACCGGAAGAGTGAAGCTGTTGAATGTGACCCGTTGATGTAGACAA
G I D * I S D Q F R T V I N R A T E E * A V E C D R * C R Q
E L I E S P T S S E Q * L I A Q R K S E L L N V T V D V D N
R N * L N L R P V P N S N * S R N G R V S C * M * P L M * T

4870 4880 4890 4900 4910 4920 4930 4940 4950
TAGGGGTGACCACTAAGAGCAATGGACATTATATATAGCCTGGTTGGACAGTACTCGGTGTGCCTTAATGTGACAGACAGTTTTGTATT
* G * P L R A M D I I Y S L V G Q Y S V C T * C D R Q F C I
R G D H * E Q W T L Y I A W L D S T R C A L N V T D S F V F
I G V T T K S N G H Y I * P G W T V L G V H L M * Q T V L Y

4960 4970 4980 4990 5000 5010 5020 5030 5040
TGAACAAAGAAATGACAGTCAATCGATGGATGAACAAGGATTATGCAAAATTCAGTTGAGAGATGAAACGAATTTGCAACAAAGAAATCGTC
* T K K * Q S F D G * T K D Y A N S V E R * N E L Q Q R I V
E Q R N D S H S M D E Q R I M Q I Q L R D E T N C N K E S S
L N K E M T V I R W M N K G L C K F S * E M K R I A T K N R

5050 5060 5070 5080 5090 5100 5110 5120 5130
TACACATGCAACAGACAAAATAATCTTCTCAACTATTTAGACATAGAGCAGATAAAGCCAATGGCTAACTAGTGATTACATAGTTCTTA
Y T C N R Q K I P S Q L F R H R A D K A N G * L V I T * F L
T H A T D K K F L L N Y L D I E Q I K P M A N * * L H S S *
L H M Q Q T K N S F S T I * T * S R * S Q W L T S D Y I V L

5140 5150 5160 5170 5180 5190 5200 5210 5220
ACACAAATTTATCACATTTTTCGGCTTTCCGCGCAAAAGTAGTCACAGATTTGAGGCATCCATATGCGGCATTTAAGCCGATTTTGTGTAA
T Q F I T F F G F R R Q S S H R F E A S I C G I * A D F V *
H N L S H F S A F A D K V V T D L R H P Y A A F K P I L C N
N T I Y H I F R L S P T K * S Q I * G I H M R H L S R F C V

5230 5240 5250 5260 5270 5280 5290 5300 5310
TATATTCTGTGACGATCACTACCAGCAAAATATTATTCTGTAATTTAAACATTTAATAAATTTTAACTTTTAACTTTAGGGCATTGGAATGGAATG
Y I L * R S L P A N I I S V N L T F K * L L I L G H L N G M
I F C D D H Y Q Q I L F L * I * H L N N F * S * G I * M E *

I Y S V T I T T S K Y Y F C K F N I * I T F N L R A F E W N

5320 5330 5340 5350 5360 5370 5380 5390 5400
ATGAAGATACCGAATGGTCAAAAGGGATGTACTCACGGTGAATAAACTGAGAGAGAACGATCCAACTGAAAATCCTTCTTTCTTATG
M K I P N G Q K G C T H V * * N * E R T I Q H * K S F F L M
* R Y R M V K R D V L T C N K T E R E R S N T E N P S F L W
D E D T E W S K G M Y S R V I K L R E N D P T L K I L L S Y

5410 5420 5430 5440 5450 5460 5470 5480 5490
GTGGCTATAACTTCGGATCGTCAACTTTTACTGTTTGTTTCATTTTTTAAAGACTTACACTCGCTCAACTCATGAATTTTCTTTGATT
V A I T S D R Q L L L F V S F F * R L T L A Q L M N F L L I
W L * L R I V N F Y C L F H F F K D L H S L N S * I F F * F
G G Y N F G S S T F T V C F I F L K T Y T R S T H E F S F D

5500 5510 5520 5530 5540 5550 5560 5570 5580
CGGGAAAGCAGATTGGTATCCTCGATAATTATATTGAACAGGTTATTTCGGGAATAGAGTGGAAATATTACGGGAGAACAGTAATGAATAG
R E S R L V F L D N Y I E Q V I R N R V E I L R E N S N E *
G K A D W Y S S I I I L N R L F G I E W K Y Y G R T V M N S
S G K Q I G I P R * L Y * T G Y S E * S G N I T G E Q * * I

5590 5600 5610 5620 5630 5640 5650 5660 5670
TACTATTTCTGACGATTGATTGTGAAACTTTGACGAGTCACTTTTCCCTTTTGATCTGTATATATATATAGAGTGTCCAACTGTAAT
Y Y F * R L I C E T L T S H F S L L I C I Y I * S V Q R N Y
T I S D D * F V K L * R V T F P F * S V Y I Y R V S N V T I
V L F L T I D L * N F D E S L F P F D L Y I Y I E C P T * L

5680 5690 5700 5710 5720 5730 5740 5750 5760
ACATATACCAATATTGGGCGATAACAGGATAGTTATTTAGATACATAAAATCAACATAAAGCAGTAGAGATACGTTGTGATAAGTCACA
T Y T K Y W A I T G * L F R Y I K S T * S T * R Y V * * V T
H I P N I G R * Q D S Y L D T * N Q H K A R R D T C D K S Q
Y I Y Q I L G D N R I V I * I H K I N I K H V E I R V I S H

5770 5780 5790 5800 5810 5820 5830 5840 5850
AAGGATACATACAGATATCAGTCATATAATAAATCAATAGAAATCACGTTAGAGATACCGGTGATAAGTTACAGAATGACGCGAATTGGG
K D H T D I S H I I N Q * K S R R D T R D K L Q E * R R F A
R I I Q I S V I * * I N R N H V E I R V I S Y K N D A D L R
K G S Y R Y Q S Y N K S I E I T * R Y A * * V T R M T Q I C

5860 5870 5880 5890 5900 5910 5920 5930 5940
CAGGATAAATGGGTAGCCGACATGCAGACATATTACAGATAATAGAATAGTTAGGCAATGTAATGAAAAGAAAGAAAGAGCGATGGGGA
Q D K W V A D M Q T Y Y R * * N S * A M * M K R K K E R W G
R I N G * P T C R H I T D N R I V R Q C K * K E R K S D G E
A G * M G S R H A D I L Q I I E * L G N V N E K K E R A M G

5950 5960 5970 5980 5990 6000 6010 6020 6030
AGAAGGAGACTGTTTACTTACTTGGCTAGATTGTTGTTATAGATAAAAAGAAAGATAGAGAACATGTAATAGCGGGAAATGTTGGC
R R R L F D Y S L A R F V V I D K K K D R E H V I A G N V G
E G D C L I T H W L D L L L * I K R K I E N M * * R E M L A
K K E T V * L L I G * I C C Y R * K E R * R T C N S G K C W

6040 6050 6060 6070 6080 6090 6100 6110 6120
GGGTAATTTACTGTGAATGACATTCAATAGCACAAATATAAATAACGGATGGCAGGAAATTAATTTTACATTGAAGAAGATTGTGA
G * F T V N D I Q I A Q I * N N G W Q E I K F H I E E R L *
G N L L * M T F K * H K Y K I T D G R K L N F T L K K D C E
R V I Y C E * H S N S T N I K * R M A G N * I S H * R K I V

6130 6140 6150 6160 6170 6180 6190 6200 6210
ATAGCAACTTCTTTAGCTTGGCTGTCAATTATACAGTCATAAACTTCGGGTTTCCCGACACTAACACATTGACAAATGATTTCAAAAA
I A T S L A W L S I I Q V I K L R V S R H * H I D K * F Q K
* Q L L * L G C Q L Y K S * N F G F P D T N T L T N D F K K

N S N F F S L A V N Y T S H K T S G F P T L I H * Q M I S K
 6220 6230 6240 6250 6260 6270 6280 6290 6300
 GAGCTGCAAAACACGTGATGCAITGTATCTCAAAATGTTCTCCAACITTAAGTCTCACITTTTTCTTGTGTAATATTTGAAGGCAATAGCA
 E L Q T R D A L Y L K C S P T L R L T F F L L K Y L K A I A
 S C K H V M H C I S N V L Q L Y V S L F S C * N I * R Q * Q
 R A A N T * C I V S Q M F S N F T S H F F L A E I F E G N S
 6310 6320 6330 6340 6350 6360 6370 6380 6390
 AAGTCGGCGGAAAAACGGAAGCATTTCATTCACTCAGTCACTACAACATTTCTTCGCAAGCATAAAATTTGATGGATTGATCTCGACTGGGAA
K S A E K R K H F I Q S A T T F L R K H K F D G F D L D W E
 S R R K N G S I S F S Q L Q H F F A S I N L M D S I S T G N
 K V G G K T E A F H S V S Y N I S S Q A * I * W I R S R L G
 6400 6410 6420 6430 6440 6450 6460 6470 6480
 TATCCAAACCGGTGTAGCAGAGGAACATGCCAAAACITTTGTTGAGGTGAGTGTAAATCCCATCACCGACTAACGTTTAAATGTTTGTITACTTG
Y P T G V A E E H A K L V E V S V I P S P T N V * C F V Y L
 I Q P V * Q R N M P N L L R * V * F H H R L T F N V L F T *
 I S N R C S R G T C Q T C * G E C N S I T D * R L M F C L L
 6490 6500 6510 6520 6530 6540 6550 6560 6570
 AAATCTCTGGCACTATTTGTAGCAATCAGATATATGCCATCAAATTTTAAATTTGACATCCAAAAGAGACAATTTCTCGCTTCAAAAATGAGT
 K S L A L L * Q S D I C H Q I L I D I P K E T I L A S K M S
 N L W H Y C S N Q I Y A I K F * L T F Q K R Q F S L Q K * V
 E I S G T I V A I R Y M P S N F N * H S K R D N S R F K N E
 6580 6590 6600 6610 6620 6630 6640 6650 6660
 TGGTTTCAAAAATAAAACTGAAATATATTACATCCATCTGAAAGAGGGAGAAAATTTCAAAAACGAATTTTGTAGGAATGAAAGCGCGGT
 W F Q K * N * N I L H P S E R G R K I Q N E F C R K * K R R
 G F K N K T E I Y Y I H L K E G E K F K T N F V G N E S G V
 L V S K I K L K Y I T S I * K R E K N S K R I L * E M K A A
 6670 6680 6690 6700 6710 6720 6730 6740 6750
 TCGTGAAGAAGCCAAAGGAATCCGGCAAGCAACAATTACTTCTTACCGCTGCTGTATCAGCCGGGAAAATGACTATTGATGAAAGTTACA
 S W K K P R N P A S N N Y F L P L L Y Q P G K * L L M K V T
 R G R S Q G I R Q A T I T S Y R C C I S R E N D Y * * K L Q
F V E E A K E S G K Q Q L L L T A A V S A G K M T I D E S Y
 6760 6770 6780 6790 6800 6810 6820 6830 6840
 ATGTTCAATCCCTTGGAAAGTAAAAAATTTGCTTTTCAATACAGACATTAATTTGACACGCAATAAAATTTACAACTCTTCAAATAT
 M F N P L E S K K N C F S Y T D I K F D T Q * N L Q Y F K Y
 C S I P W K V K K I A F H I Q T L N L T R N K I Y N T S N I
N V Q S L G K * K K L L F I Y R H * I * H A I K F T I L Q I
 6850 6860 6870 6880 6890 6900 6910 6920 6930
 AATTTTGTAGCATTGAAATTTTCAGAAAGTTGGATCTTCTAATTTCTGATGTCGTATGATTACACGGTAGTTGGGAGAAGAACGTCGACT
 N F V A L K F S E V W I F Y F * C R M I Y T V V G R R T S T
 I L * H * N F Q K F G S S I S D V V * F T R * L G E E R R L
 * F C S I E I F R S L D L L F L M S Y D L H G S W E K N V D
 6940 6950 6960 6970 6980 6990 7000 7010 7020
 TACACGGAGAGTTGGCTCCGACGGAGAGAGAAACATCCGGAATTTGGGCTCTGTAATACCGTTAGTTTGTAAATAAAAATTTCTAAATTT
 Y T E S C V R R R E K H P E L A S V I P L V L E I K I S N F
 T R R V A S D G E R N I R N W R L * Y R * F L K * K F L I F
L H G E L R P T E R E T S G I G V C N T V S S * N K N F * F
 7030 7040 7050 7060 7070 7080 7090 7100 7110
 TCTTGTGTTGATGAAAGCAATTTCTGATGATTCAGGAATTTGCAGCAAATTTATGGGAGAGAAAGGTATGCCGAAACAGAAAATTA
 F L F A * R R I L D D F R N L Q Q I I G Q R K V C R N R K L
 S C L H E D E F L M I S G I C S K L L G R E R Y A E T E N Y

F L V C M K T N S * * F Q E F A A N Y W A E K G M P K Q K I
 7120 7130 7140 7150 7160 7170 7180 7190 7200
 TCATTGGGATTCCGGCTTACAGTCGGGGATGGACATTAAGTAATCCTTCGGAAACGGCAATTGGAGCGGAGGGTGGTCCATCATCGC
 S L G F R L T V G D G H * V I L R K R Q L E R R V V V H H R
 H W D S G L Q S G M D I K * S F G N G N W S G G W S S I I A
I I G I P A Y S R G W T L S N P S E T A I G A E G G R P S S
 7210 7220 7230 7240 7250 7260 7270 7280 7290
 CATCGACAACAAATCCCGCTGGCGGTACTGCGGCTTATTGGGAGGTTTAAATTTAAAGTTCTCATTTACTTTTGAATATATTTCTTGCTGC
 H R Q Q I P L A V L R L I G R F N L K F S F T F E Y I S C C
 I D N K S R W R Y C G L L G G L I * S S H L L L N I F L A A
P S T T N P A G G T A A Y W E V * F K V L I Y F * I Y F L L
 7300 7310 7320 7330 7340 7350 7360 7370 7380
 TGATGAATTTCTCAAATTCATTATTAATTAATGTAACCATTTTAAGATATGCAAAATATGTGAAAGGAGGTGGCAAGGAAACGGTGGATA
 * * I S Q I P L L I N V T I L R Y A N M * K K V A R K R W I
 D E F L K F H Y * L M * P F * D M Q I C E R R W Q G N G G *
 L M N F S N S I I N * C N H F K I C K Y V K E G G K E T V D
 7390 7400 7410 7420 7430 7440 7450 7460 7470
 AAAAAAGTGTGGAGCGTATATGGTAAAAGGAGATCAGTGGCAGCGTTACGATAATGAGGAAACTATTAAAATTAAGTAATTTTTATT
 K K V L E R I W * K E I S G T V T I M R K L L K L K * F L F
 K K C W S V Y G K R R S V A R L R * * G N Y * N * S N F Y F
K K S V G A Y M V K G D Q W H G Y D N E E T I K I K V I F I
 7480 7490 7500 7510 7520 7530 7540 7550 7560
 CGCTTATTGTTGTTGATAGTCAAAAATAAAAAAGTGGATTTGATAGTCAAAAATAAGAAAAGATAATAAGATAGTCGAAAAATAAAAAAT
 R L L L L I V K N K K V I * * S K N K K R * * D S R K I K N
 A Y C C * * S K I K K * F D S Q K I R K D N K I V E K * K I
 S L I V V D S Q K * K S D L I V K K * E K I I R * S K N K K
 7570 7580 7590 7600 7610 7620 7630 7640 7650
 TAAAAATAACGTTGCCAATATATCTTGGCCTTAACITTAATTTAAAGAGCACCATAAATTTTATAATGTTTAAATAAGAAGAAATTAATTT
 * K I T L P I Y L A L T L I * R A P * I L * C L I R R N * F
 K K * R C Q Y I L P * L * F K E H H K F Y N V * * E E I N F
 L K N N V A N I S C L N F N L K S T I N F I M F N K K K L I
 7660 7670 7680 7690 7700 7710 7720 7730 7740
 CGGAGTTTACTCGTGAATTTGTTTTACTTTTACAATTAAAATTTTCAGATGAAAATGGCTGAAAGAGAAAGGTTATGGCGGTGCTTCATAT
 R S L L V * L F Y F R I K I F R * N G * K R K V M A V P S Y
 G V Y S C N C F T L E L K F S D E M A E R E R L W R C L H M
 S E F T R V I V L L * N * N F Q M K W L K E K G Y G G A F I
 7750 7760 7770 7780 7790 7800 7810 7820 7830
 GGGCTCTTGATTTTGGATGATTTTAAAGGTACAAGTTTGGCAAAGGTCCTATCCACTGTTAAATGCTCTTAATAATGAGCTTCGAGACG
 G L L I L M I L R V Q V V A K V P I H C * M L L I M S F E T
 G S * F * * F * G Y K L W Q R S L S T V K C S * * * A S R R
W A L D F D D F K G T S C G K G P Y P L L N A L N N E L R D
 7840 7850 7860 7870 7880 7890 7900 7910 7920
 GCTTCGATTTTGGGTGAAGGATTTTACAACCTTCGTAATGATTTTGGCATAATCAAAAATGTTATAATCGGTCGGAACCTTAAGGATT
 A S Y F G * R I L Q L R N D L L H I I K M L * S V G T * R I
 L R I L G E G F Y N F V M I C C I S * K C Y N R S E L K G L
 G F V F W V K D F T T S * * F V A Y H K N V I I G R N L K D
 7930 7940 7950 7960 7970 7980 7990 8000 8010
 GAAATTTAATCCGAAATCCATAAATTCATAATGTAATGCTGATTTTAAACAGTATTGCAAAACGCAACTGGTGCATTTTGGATATTCI
 E I * S G N P K I H N V M L Y F K Q Y C K R N W C I F G Y S
 K F N P E I L K F I M * C C I L N S I A N A T G A F L D I L

* N L I R K S * N S * C N A V F * T V L Q T Q L V H F W I F

8020 8030 8040 8050 8060 8070 8080 8090 8100
 TTGCTTICTTACAAATGGTCCGCCAAAAATGAATCAGCACAAGCCGAAATGTCTATTCAATTAATTTTTATATTAATTTCCGTTCTATAC
 L L S Y K W S A K N E S R Q A E M S I Q L N F Y I N F V L Y
 C F L T N G P Q K M N H D K P K C L F N * I F I L I S F Y T
 F A F L Q M V R K K * I T T S R N V Y S I K F L Y * F R S I

8110 8120 8130 8140 8150 8160 8170 8180 8190
 AGATTTAGATTTAAAAATTAATAAAGCAATCAITTTTGTGTCATCAATGTTGGCAAAATCAAATAATGAAATATTGAGGAAAACAGGGAG
 R F * I * K L I K Q S F L V I N V G K I K * * N I E E N R E
 D F R F K N * * S N H F * S S M L A K S N N E I L R K T G R
 Q I L D L K I N K A I I F S H Q C W Q N Q I M K Y * G K Q G

8200 8210 8220 8230 8240 8250 8260 8270 8280
 AGCCTCAGGAAGTAGTGAATTAAGCAATCAAAAGTGGTTTTGAAACAGTACGAAACTTCTGGTTTGTATCCGCAAGTTAAAAATG
 S L R K * * N * N D Q K W F * N S T K N F W F V I R K L K C
 A S G S S E I E T I K S G F E T V R R T S G L L S A S * N V
 E P Q E V V K L K R S K V V L K Q Y E E L L V C Y P Q V K M

8290 8300 8310 8320 8330 8340 8350 8360 8370
 CGTCTTGATCTGTTCTTTTGTCTTAAACACAGCGCAACTTTTGGAGTTTGTCTTTCATGCAATTTTGGATAGAATGAAATGATTGTGGA
 R S * S V L L L * T Q R N F S S L L F M H F G * N G N D C G
 V L D L F F C F K H S A T F R V C S S C I L D R M E M I V D
 S F L I C S F A L N T A Q L F E F A L H A F W I E W K * L W

8380 8390 8400 8410 8420 8430 8440 8450 8460
 TAGATGAAAGAAATTTAGGAAAATCAATGTTCTGCTGTAACATTTTCAGATATAAAATGTTTGGAGTCATATGGTCTGTTCCCGCATCT
 * M K R N L G K S M F C C N I S D I K C L E S Y G L F P H P
 R * K E I * E N Q C S A V T F Q I * N V W S H M V C S R I L
 I D E K K F R K I N V L L * H F R Y K M F G V I W S V P A S

8470 8480 8490 8500 8510 8520 8530 8540 8550
 GACGATTGCCATTTATTTATTCATTGGCGCAATGACTATCCATACGTAACAATGTCGCGTACATACCTTTTTAATGATGATAACAA
D D C H L F I H C A N D Y P Y V K Q C P L H T F F N D D I K
 T I A I Y L F I A Q M T I H T * N N V R Y I P F L M M I S K
 * R L P F I Y S L R K * L S I R E T M S A T Y L F * * * Y Q

8560 8570 8580 8590 8600 8610 8620 8630 8640
 AATTGGCATCACTTAGTAAATGGCGCATCACATGCAAAATGATTTTTCATATTACCAAGAATAAAAGTTTAAATGTAAGTTGTGGCAGTC
I C D H L V N A P I T C K * F S Y Y Q E * K F N V K V V A V
 F A I T * * M R R S H A N D F H I T K N K S L M * K L W Q S
 N L R S L S K C A D H M Q M I F I L P R I K V * C K S C G S

8650 8660 8670 8680 8690 8700 8710 8720 8730
 AAATGAATTGACCATACTCCGTGTAATAATTCATATTATATAGGAGAATAAATATGACTATATGAGGACAAATGTCAAAATGAGTCTAAC
 K * I D H T P V I I H I I * E N K Y D Y M R T N V K N E S N
 N E L T I L L * * F I L Y R R I N M T I * G Q M S K M S L T
 Q M N * P Y S C N N S Y Y I G E * I * L Y E D K C Q K * V *

8740 8750 8760 8770 8780 8790 8800 8810 8820
 GGAATTATTAATAAATGAAAAATTTGTTATTATCTGTATTTTATTTTGGGTAGCAAAATCTTTCAACAGTTATCACCATAACTTAGTA
 G I I K N E K I C Y Y L Y F I L R S K N L S T V I T H N L V
 E L L K M K K F V I I C I L F C V A K I F Q Q L S P I T * *
 R N Y * K * K N L L L S V F Y F A * Q K S F N S Y H P * L S

8830 8840 8850 8860 8870 8880 8890 8900 8910
 GAAAACATATGAATGATTCGGAAGATGAATAAATCACATGAAAAGGATGAAATCTGTATAAAAAATAAGTACTCAATAAAAAAGGATGAA
 E N I * M I R K M N K S H E K D E I L I K N K Y S I K K D E
 K T Y E * F G R * I N H M K R M K F * * K I S T Q * K R M K

R K H M N D S E D E * I T * K G * N S D K K * V L N K K G *

8920 8930 8940 8950 8960 8970 8980 8990 9000
ATCATTATAAAAAGGAAATTCGGATAAAAAATAAATACTCAAAAGATAAAAAATACTGTTCCATTGCATCAGGACTGGAGTATTTCCATCAA
I I I K R K F R * K I N T Q R * K Y C S I A S G L E Y F H Q
S L * K G N S D K K * I L K D K N T V P L H Q D W S I S I K
N H Y K K E I P I K N K Y S K I K I L F H C I R T G V F P S

9010 9020 9030 9040 9050 9060 9070 9080 9090
GAATCTTTTATTCGGTTTCGGCATTTCCTGCTGCATAATTCCTTCCTTTTCCTTTGGCATTTCCTTTGTTTCCTTTACGACAATTCAG
E S F I P F P H F H C C I I P S F S F G I P F V S F T T I Q
N L L F R F R I F T A A * F L P F L L A F L L F P L R Q F S
R I F Y S V S A F S L L H N S F L F F W H S F C F L Y D N S

9100 9110 9120 9130 9140 9150 9160 9170 9180
CTTAATTTTGATAAGAGAGTTTGTAGTGGCTAATGTGATGACCOCTCAATTCGAAAAAGGAAACAGTCARAGGCGAATTTCAATTATTTT
L N F D K R V C S W L M * * P S I R K R K Q S K A N F N Y F
L I L I R E F V V G * C D D P Q F E K G N S Q R R I S I I F
A * F * * E S L * L A N V M T L N S K K E T V K G E F Q L F

9190 9200 9210 9220 9230 9240 9250 9260 9270
CAACAATTCATCATTTGAATGATATGTGGGATAGAAGGAATTTTCATAATCTCAGATGAAGAAAAGTATTTTGCAGTCTCAGATGAAGA
Q Q F S S L N D M C D R R N F H N L R * R K V F C S L R * R
N N S H H * M I C A I E G I F I I S D E E K Y F A V S D E E
S T I L I I E * Y V R * K E F S * S Q M K K S I L Q S Q M K

9280 9290 9300 9310 9320 9330 9340 9350 9360
AAAGTATTTTGCAGTTGGGCTTCATCTTTCTCTCCATATTATGATAAAAGATATAATAAAAACTTCCAAATAAAGCGTGGCAGCATTAA
K V F C S C A S S F S S I L * * K I * * K L P I I N V A A L
K Y F A V A L H L S L P Y Y D K R Y N K N F Q * * T W Q H Y
K S I L Q L R F I F L F H I M I K D I I K T S N N K R G S I

9370 9380 9390 9400 9410 9420 9430 9440 9450
TTTTTCTAGCTTTCTGCTTATTTTTAATGTTTTTCCAGCTTTCTAGCTATTTCTGTATTTTCCCAATTTCCCTAGTGTTTTTCCAGCTT
F F L A F C L F L M F F Q L S Y F L Y F P N F L V F F Q L
F F * L S A Y F * C F S S F L A I S C I F P I S * C F S S F
I F S S F L L I F N V F P A F * L F L V F S Q F P S V F P A

9460 9470 9480 9490 9500 9510 9520 9530 9540
TTGTAGTGTTTTTCTAAGCTTAAATACTATCTACGTCAGCCTAACTTCGGGATTTTAAATTAAGTAAATTTGGTTAATCAAATATCTTCG
L * L F F * L L N T I Y V S L T S G F * L T N L V N Q I S S
C S C F S N S L I L S T S A * L R D F N * L I W L I K Y L R
F V V V F L T P * Y Y L R Q P N F G I L I N * F G * S N I F

9550 9560 9570 9580 9590 9600 9610 9620 9630
AATCAAACTTTGTAGAGCGCTTATTTTTGACACTCTTAATGTTCTTAAAAAAAACCTTACTTCATTGTTATGTTTTATGCTTAAACAT
N Q I L C R A L I F D T L N V L K K N L T S L L C F Y A * H
I K Y F V E R L F L T L L M F L K K T L L H C Y V F M L N I
E S N T L * S A Y F * H S * C S * K K P Y F I V M F L C L T

9640 9650 9660 9670 9680 9690 9700 9710 9720
TTATTAATAACTGCAGAAATTTATTTTTTTTTATTTTTGGATGTGTTGCTGAGAACTTTTGA AAAAATTTGTTGTTATGATTTTGGAT
L L I T A E I I F F F Y F W M C S L R T F E K I C C Y D F D
Y * * L Q K L F F F F I F G C V R * E L L K K F V V M I L I
F I N N C R N Y F F F L F L D V F A E N F * K N L L L * F *

9730 9740 9750 9760 9770 9780 9790 9800 9810
TTTTCAACGTTTTTTCATTTTTTCTCTTATATCTATTTGTTTTCAGCTCTTTTCGCTTTACCTAATTCGAAGATTTTAAAAATATCTGA
F S P F F I F F S Y I L L C F P A L S L Y L I A R F * N I *
F H R F S F F S L I S Y C V F Q L F R F T * L Q D F K I S D

F F I V F H F F L L Y L I V F S S S F A L P N C K I L K Y L
9820 9830 9840 9850 9860 9870 9880 9890 9900
CGATTACAAAATGTGCAACATTATTAATTCCTTTATTTGTTAATATTGTTTGCATTTTATTTTGCRACTTCCTGATTTTATTTCCRAAGCTA
R F T K C A T L L I L Y C * Y C L H F I L Q L P D F I S K L
D L Q N V Q H Y * F F I V N I V C I L F C N F L I L F P S Y
T I Y K M C N I I N S L L L I L F A F Y F A T S * F Y F Q A
9910 9920 9930 9940 9950 9960 9970 9980 9990
TGCAAAACATTCGATCTATAAAAATCTGTTTCAAACTTAAGCCTACCAATGTTTTTACAGTAATCCATTTTGTATCCCATATAACTCA
C K H F V S I K S V S N L S L P M F F T V I H F * F P Y N S
A N I S Y L * N L F Q T * A Y Q C F L Q * S I F D S H I T H
M Q T F R I Y K I C F K L K P T N V F Y S N P F L I P I * L
10000 10010 10020 10030 10040 10050 10060 10070 10080
CCCCAATTCATGATCTACCGACTTTTATTTATATTTTTCATTCCTGTCACATTTTCACTTTTATCCAAATTTGAGCATATAAATATGA
P Q F H V S T D F Y L Y F H S F V Q F H F Y S N L S I * I *
P N F M Y L P T F I Y I F I H S S N F T F I P I * A Y K Y E
T P I S C I Y R L L F I F S F I R P I S L L F Q F E H I N M
10090 10100 10110 10120 10130 10140 10150 10160 10170
ATAAAATTAAGTATTTTGTATGATGATATTAAGATATGAAAAATTTCCAGCTTATATTTAATCTGTGATATGCACAGTATAATTCATTTT
I K L S I L Y D D I K I * K I S S L Y L T C D M H S I I H F
* N * V F C M M I L R Y E K F P A Y I * L V I C T V * F I F
N K I K Y F V * * Y * D M K N F Q L I F N L * Y A Q Y N S F
10180 10190 10200 10210 10220 10230 10240 10250 10260
CTGATTTCCGCACAAAGATTGAAGATGTGAAAATATTTGACCTTACCAAAATGTGGAGCATTATTATTCATATTAATTTTTTCAGTATA
L I S A Q R L K M * K Y L T F T K M W S I I I H I N F F S I
* F P H K D * R C E N I * P L P K C G A L L F I L I F S V *
S D F R T K I E D V K I F D L Y Q N V E H Y Y S Y * F F Q Y
10270 10280 10290 10300 10310 10320 10330 10340 10350
ATAATGCACTGTCCCAATTTATTTGTCACAAAAAATCCCAACCGCTGAGCTGCTTCTTAATATATAATGTTAATAATAATTTCA
I M H C P I Y C Q Q K K K I P N A E L L L N N I M L I I S
* C T V P F I V N K K K K S Q T L S C F L I I * C * * * F H
N N A L S H L L S T K K K N P K R * A A S * * Y N V N N N F
10360 10370 10380 10390 10400 10410 10420 10430 10440
TATATTTCCAGTAATTTGATTTTATTTATATTCCTTAAACATACAGTTGTCGCTTCTTCTGACCCGATATAATTTAGGGTTTGA
Y I S S N * L F Y Y I P * T Y T L S S S S L T V I I F R V *
I F P V I D Y F I I F L K H T R C R L L L * P * * Y L G F D
I Y F Q * L I I L L Y S L N I H V V V F F S D R D N I * G L
10450 10460 10470 10480 10490 10500 10510 10520 10530
TTGCAATCTTTTCACTTACTGTAAGTTCAAACTGGTAATTAACCATAGCAATGATAGTAAACACCAATATTAACCAAGAAACAAGCG
L Q S F S Y C E L Q T G N * T I A M I V K T Q Y * N K K Q A
C N L S V T V N F K L V I K P * Q * * * K H N I K T R N K R
I A I F Q L L * T S N W * L N H S N D S K N T I L K Q E T S
10540 10550 10560 10570 10580 10590 10600 10610 10620
GTACAAAGATAAATCAAAACACATAACCTCAACGGAAGAATAAGTAGAAGAATGAAGAGATGAATGATTTGATGCTTATATAGGCAA
V Q R I N Q N T * P Q R K N K * K E * R D E * F D V L Y R Q
Y K E * I K T H N L N G R I S R K N E E M N D L M S Y I G K
G T K N K S K H I T S T E E * V E R M K R * M I * C L I * A
10630 10640 10650 10660 10670 10680 10690 10700 10710
AGTATACGTGCATATCATAACCGGGAATAAAGSAAAGATATGGAGGATATAAGAGCAAAGGAGATCATGTAGTACAATTAATGGAGG
S I R A Y H N A G K * G K I L E D I R A K E I M * Y N N W R
V Y V H I I T R E N K E R Y W R I * E Q R R S C S T I I G G

K Y T C I S * R G K I R K D I G G Y K S K G D H V V Q * L E

10720 10730 10740 10750 10760 10770 10780 10790 10800
CAAAGATGGGAACGGATGCGAGTATAGAATGGATCATTGGAATGTGTTATTAGTTTAGACAGGAACATCGACATTGACACGTATTAAC
Q R W E R M R V * N G S F E C V I * F R Q E H R H L T R I N
K D G N G C E Y R M D H L N V L F S L D R N I D I * H V L T
A K M G T D A S I E W I I * M C Y L V * T G T S T F D T Y *

10810 10820 10830 10840 10850 10860 10870 10880 10890
ATGCGATAATATTTCATTATTACAAATTATGTCTATGAATAAAAATATATATATATATATATATCTTTGCGACAAAAGGAATTGCTTTTC
M Q * Y S L L Q L C L * I K Y I Y I Y I Y L S R Q K E L L F
C S N I H Y Y N Y V Y E * N I Y I Y I Y I F R D K R N C F S
H A V I F I I T I M S M N K I Y I Y I Y I S F A T K G I A F

10900 10910 10920 10930 10940 10950 10960 10970 10980
TCTGAATTTTAAATCCAAACAAAATAAATAAATAAATAAATTCTTCTTTGGTACTTCTCGCTTTTCACGATCTTCCAATTAGTGAAG
S E I L I Q N K K I N K N * F F F G T S R F S R I F Q L V K
L K F * S K T K K L I K I N S S L V L L A F H V S S N * * S
L * N F N P K Q K N * * K L I L L W Y F S L F T Y L P I S E

10990 11000 11010 11020 11030 11040 11050 11060 11070
CCATGAATGAAAACATTACCAACGGCAACGGTAAGTTGTCCAGTAAAGTGACAATTTTGGTTGGATGATATAGGTGGTATATGGAATTTT
P * M K T L P T Q T V S C P V K * Q F W L D D I G G I W N F
H E * K H Y Q R K R * V V Q * S D N F G W M I * V V Y G I F
A M N E N I T N A N G K L S S K V T I L V G * Y R W Y M E F

11080 11090 11100 11110 11120 11130 11140 11150 11160
CACACTCTCAGATATAATGAAAACGGTACTTTGTGTTATACGTCACATTTTCTTTTCACATTATGATAAAGACATAACAAAATTCGAC
H T L R * M K N V L C V I R H I F F S H Y D K R H N K I L D
T L S D K * K T Y F V L Y V T F S F H I M I K D I T K F S T
S H S Q I N E K R T L C Y T S H F L F T L * * K T * Q N S R

11170 11180 11190 11200 11210 11220 11230 11240 11250
AAGTACGGTATATTTTGTGTTTTCTAACTTTTAGGCTGTTGTTAATTTTTTCCAGCTTTCCAGCTATTTACGTATATTTTTTCCATCTTTT
K Y G I I C V F L T F R L L L I F P A F Q L F T Y I F P S F
S T V L F V F F * L L G C C * F F Q L S S Y L R I F F H L F
Q V R Y Y L C F S N F * A V V N F S S F P A I Y V Y F S I F

11260 11270 11280 11290 11300 11310 11320 11330 11340
TIGGCCATTTCTGTGTTTTTTCAGCTTTCTCGTGTTTTTTCTAACTCCITGAAATATTTTTTGGTATTTTCACAGCTTTTTAGAGGAGATGG
L A I S C V F S A F S C F S N S L N I F R I F T A F * R R W
W P F L V F F Q L S R V F L T P * I F F V F S Q L F R G D G
F G H F L C F F S F L V F F * L L E Y F S Y F H S F L E E M

11350 11360 11370 11380 11390 11400 11410 11420 11430
TTTACATTTTCTAAAGAACGGTACGTTTGATAAAGTACTTTATAAAGGCCCGTTTATGGCGTTAAAGACTTCACTTTTTCATTTTATAT
F T F S * R T V R L I K Y F I K G P F M A L K T S L F I L Y
L H F P K E R Y V * * S T L * K A R L W R * R L H F S F Y I
V Y I F L K N G T F D K V L Y K R P V Y G V K D F T F H F I

11440 11450 11460 11470 11480 11490 11500 11510 11520
TTTGTATCTTTTCAACATTCCTCCCGATTACTTTACAAAATATCTTTTTTCTATTTTTGACATTTATCTTTTTTTTTGAAATTTCTTTTC
F A I F Q H S S R L L Y K N I F F L F L H I Y L F F * I S F
L L S F N I P P D Y F T K I S F F Y F C T F I F F F E F L S
F C Y L S T F L P I T L Q K Y L F S I F A H L S F F L N F F

11530 11540 11550 11560 11570 11580 11590 11600 11610
TTTTTTGCTATTTTCTAGATATTTCCAGATTTTGTATGCAATCTCTGAGAATGTCTTGGCAATTTGCGCAAAAATAGAAAAAGA
F F A I F * I F P D F * F C M Q S L R M S C N L R K N R K R
F L L F S R Y F Q I F S F V C N L * E C L A I C A K I E K D

L F C Y F L D I S R F L V L Y A I S E N V L Q F A Q K * K K
 11620 11630 11640 11650 11660 11670 11680 11690 11700
 TAITTCATCGATATTTCTTAATTAGTCATTACATTAGCTGCAGCATAACTCATTGAATTCATATTTATCCCAAAGATTAGTATATCAA
 Y F I D I S * L V I Y I S C S I T H * I P Y L S Q R L V Y Q
 I S S I F L N * S F T L A A A * L I E F H I Y P K D * Y I N
 I F H R Y F L I S H L H * L Q H N S L N S I F I P K I S I S
 11710 11720 11730 11740 11750 11760 11770 11780 11790
 CATTATATAACGGTTGATATACTAAATCTCATTGGAAACAAAATGATATCAAAAACATGAAAAATTTGCTACTTATAGAAAATCTGTAC
 H Y I K R * Y T N L I W N K M I S K T * K I C Y L * K I L Y
 I I L N V D I L I S F G T K * Y Q K H E K F A T Y R K Y C T
 T L Y * T L I Y * S H L E Q N D I K N M K N L L L I E N T V
 11800 11810 11820 11830 11840 11850 11860 11870 11880
 ATCTAAGCTTACTTCTACTTCTCAATTAATTTTTCTTGAATCCTTTGTATGCACCGACAGTTATTCTCACCAATCAATTTCAATACAATA
 I * A Y F Y F S I N F F L N P L Y A P T V I L T N H F N T I
 S K L I S T S Q L I F S * I L C M H R Q L F S P I I S I Q *
 H L S L L L L L N * F F L E S F V C T D S Y S H Q S F Q Y N
 11890 11900 11910 11920 11930 11940 11950 11960 11970
 ATCTCTTATTACTCTGTAAACTCAGCAAAAAAGTATTCCTAATCGGCTAGCTATGIGTAATTAGTTACGGTAATTCOCATGCATGTATAT
 I S Y Y S V N S A K K V F L I A E L C V I S Y G N S M H V Y
 S L I T L * T Q Q K K Y S * S L S Y V * L V T V I P C M Y I
 N L L L L C K L S K K S I P N R * A M C N * L R * F H A C I
 11980 11990 12000 12010 12020 12030 12040 12050 12060
 TTAAGTATGTTTTCGCTTATTTCTTACACATCCACTTAATATCTTCCATTAATGAAGGCAAAATATCAGTTAATCATAGTAATGAATTT
 L T D C F A Y F L T H P L N I F H * * R Q I S V N H S N E F
 * L I V S L I F L H I H L I S S I N E G K Y Q L I I V M N F
 F N * L F R L F S Y T S I * Y L P L M K A N I S * S * * * I
 12070 12080 12090 12100 12110 12120 12130 12140 12150
 TGAATACAGAAGTGTACAAGTCCCATGAAGTGACATGAATGGATTATGATAACGCTTTTCATCATATTCGCCAATATAATTACTGTTG
 * N T E V Y K S H E V T * I G L * * R F S S Y S P I * L L L
 E I Q K C T S P M K * H E L D Y D N A F H H I R Q Y N Y C C
 L K Y R S V Q V P * S D M N W I M I T L F I I F A N I I T V
 12160 12170 12180 12190 12200 12210 12220 12230 12240
 CAAATGGTAAGTTGCTCAATAAAAATGACACGTCAGTTGCGTAAGCAGCAAAATATCATCGCTGAAGATATAATGAACATGACGGTTGACAC
 Q M V S C S I K * H V S C V S T K Y H R * R Y N E H D G * H
 K W * V A Q * N D T S V A * A R N I I A E D I M N M T V D T
A N G K L L N K M T R Q L R K H E I S S L K I * * T * R L T
 12250 12260 12270 12280 12290 12300 12310 12320 12330
 TTCAGATCATACTACATCTTAATCAATTATATGTCATTTCAATCCTAAACTGTGAGTAGCATCTATTGAAATTCATAGTGGCATATATT
 F R S Y Y I L I I I C H F N P K L S S S I Y * N S I V A Y I
 S D H I T S * S L Y V I S I L N C R V A S I E I Q * W H I L
 L Q I I L H L N H Y M S F Q S * T V E * H L L K F N S G I Y
 12340 12350 12360 12370 12380 12390 12400 12410 12420
 GAATTCACAGAAGCTCTCACAGAATAAGCAAAATGAAGAGACTAAATGAAGATAATCAAAATTTTTACTTGAAGATTTAGTAACATT
 E F H R S S H K K * A N E E T K L K I I Q I F T * R F S N I
 N F T E A L T R N K Q M K R L N * R * Y K F L L E D L V T L
 * I S Q K L S Q E I S K * R D * I E D N T N F Y L K I * * H
 12430 12440 12450 12460 12470 12480 12490 12500 12510
 GTATTTGAATAAATTTGCAAGAAATAACCCCAAAATCGATGTTGAACATATCTTCAGGTTGTAGTCTATTTTCAGGTTACGTTTCGTTGGATG
 V F E * I A K K * P P N R C * I Y L Q V V V Y F R L R S W M
 Y L N K L Q R N N H Q I D V E H I F R L * S I S G Y V R G C

C I * I N C K E I T T K S M L N I S S G C S L F Q V T F V D

12520 12530 12540 12550 12560 12570 12580 12590 12600
T T A C T A T A T T A G T T G G G C T G A A C G T C A A C A A G S I T C A G T G A C G T T T G A A A T A A T T T C A T T A T T A A A G A G C T A A A T A G T T T C A A G G G G A I T
L L Y * L G * T S T R F S D V * N N F I I K E L N S F K G I
Y Y I S W A E R Q Q G S V T F E I I S L L K S * I V S R G L
V T I L V G L N V N K V Q * R L K * F H Y * R A K * F Q G D

12610 12620 12630 12640 12650 12660 12670 12680 12690
G T G T A C A A A T T T T C G A A T A A A A A G T A G A A A A C C A A T G A A A T T A T T T C A A G A T T T C C A T C T C A A A A A T G A A T T T T A A G C C C A C G A A A T A T T
V Y K F S N K K * K T N E I I S R F P S Q K * I L S P R N I
C T N F R I K S R K P M K L F Q D F H L K N E F * A H E I L
C V Q I F E * K V E N Q * N Y F K I S I S K M N F K P T K Y

12700 12710 12720 12730 12740 12750 12760 12770 12780
A G T G G A A A A T A A A C A T C A A T A A T T A G C A A C T T C A A C T T C C A A C A T A R A C T C A A T A T T C C C T C A T T T C C T G A A T T T C A T C G T T T C G A C G T
S G K * T S I I S N F N F Q H K L N I P S F P E F S S F R R
V E N K H Q * L A T S T S N I N S I F P H F L N S H R F D V
* W K I N I N N * Q L Q L P T * T Q Y S L I S * I L I V S T

12790 12800 12810 12820 12830 12840 12850 12860 12870
G T G C C T T A G C T A C C G T A A A A C A A T A T G G T G A A A A T A A T A C T A A A C A A A T T A A A A T A A A A C R A G C A T T G T A T T T C T G C C G G A A A T G T A A T
V P * L P * N N M V K I I T K Q I K I K Q A L Y F C R K C N
C L S Y R K T I W * K * * L N K L K * N K H C I S A G N V I
C A L A T V K Q Y G E N N N * T N * N K T S I V F L P E M *

12880 12890 12900 12910 12920 12930 12940 12950 12960
A T A C A T T T T C A T C T T A A A G G A G A G G C A A A T T T T T G C C A G A A G A T A T C C C A A A A C A C C T A T G C A C T C A C A T C C T T T A T G C A T T C A C C A A C
I H F H L K G E G K F L P E D I P K H L C T H I L Y A F T N
Y I F I L K E K A N F C Q K I S Q N T Y A L T S F M H S P T
Y T F S S * R R R Q I F A R R Y P K T P M H S H P L C I H Q

12970 12980 12990 13000 13010 13020 13030 13040 13050
G T T G A A G G G A A A C G C G G T A A G T T A C A A T G A T A T T T G C A A T G A G A A A T G T A A A T T A A A T T T T C G G A G T T T C T T T T A C A A T G C A C A T T A A
V E G K R A * V T N D I C N E K C K L N F R S F F Y N A H *
L K G N A R K L Q M I F A M R N V N * I F G V S F T M H I K
R * R E T R V S Y K * Y L Q * E M * I K F S E F L L Q C T L

13060 13070 13080 13090 13100 13110 13120 13130 13140
A A T T A T T C T A A T T T T T A T T G C C T G C T C A A G T G A T T A T A A A C C T G A A A C G T T A A T A T G C G A T A T T A G C T G G T T T T G C A T A A A A T A T T C T T
N Y S N F Y C L L K * L * T * N V N M R Y L A G F A * N I L
I I L I F I A C S S D Y K P E T L I C D I * L V L H K I F F
K L F * F L L P A Q V I I N L K R * Y A I F S W F C I K Y S

13150 13160 13170 13180 13190 13200 13210 13220 13230
C T G T A A C A A T T G C T A T C A G C A A A T G T T A T T T A T T T C T G T A A A T T T C T G A T T T T A A T A T T T T A A A T C T T A G A A C A T T T G A A T G G G A C G A
L * Q L L S A N V I Y F C K F L I L I I F K S * N I * M G R
C N N C Y Q Q M L F I S V N F * F * * F L N L R T F E W D D
S V T I A I S K C Y L F L * I S D F N N F * I L E H L N G T

13240 13250 13260 13270 13280 13290 13300 13310 13320
T G A A G A T A C C G A A T G G A C G A A A G G G A T A T A T C C A C A T A T G A T A A A A G T A A A A G A G T A C G A T C C A A C C C T G A A A A T T C T T C T T C T A T A G
* R Y R M D E R D I S T Y D K S K R V R S N P E N S S F L *
E D T E W T K G I Y P H M I K V K E Y D P T L K I L L S Y S
M K I P N G R K G Y I H I * * K * K S T I Q P * K F F F P I

13330 13340 13350 13360 13370 13380 13390 13400 13410
C G A C T A T A A C T C T C A C T C A T C A G T T T T T G C T G T T G T T T T T T T T T T T T A A A C T C T C A G C T A T C A T T T T T T T T T T T T T T A A A G A T T G A C A C T C A G C
R L * L S L I S F C C S F F F * T L S Y H F F F * R L T L S
D Y N S H S S V F A V R F F F K L S A I I F F F K D * H S A

A T I T L T H Q F L L F V F F L N S Q L S F F F L K I D T Q

13420 13430 13440 13450 13460 13470 13480 13490 13500
 TAACGTATTGACTAATAATTTCAAAAGAGACTGCTCATACGTCATTTCCTTAGACATCAAGATAATGCAAGTGTACCTCAGACGTTTTCCG
 * R I D * * F Q K R L L I R H F L D I K I M Q C Y L R R F P
 N V L T N N F K R D C S Y V I S * T S R * C S V T S D V F R
 L T Y * L I I S K E T A H T S F L R H Q D N A V L P Q T F S

13510 13520 13530 13540 13550 13560 13570 13580 13590
 GATTTACGTCATCTTTTTCTTCTGCTAAAATATCAACGTAATTTGATGATATAAAAATATTTGAAAGGCAATAGCAAAATCGACAAAAA
 D L R L I F F L L K Y Q R K F D D I K I F E G N S K I D K K
 I Y V S S F S C * N I N V N L M I * K Y L K A I A K S T K N
 G F T S H L F L A K I S T * I * * Y K N I * R Q * Q N R Q K

13600 13610 13620 13630 13640 13650 13660 13670 13680
 CCGGAAAAATTTCAATTCAGTCAACTGTAGCATTTCCTCGTAAGCATAAATTTGATGGATTGATCCGAATTCCTCGCCCTATAGTGAGTC
 P E K F H S V N C S I S P * A * I * W I R S E F F A L * * V
R K N F I Q S T V A F L R K H K F D G F D P N S S P Y S E S
 T G K I S F S Q L * H F S V S I N L M D S I R I L R P I V S

13690 13700 13710 13720 13730 13740 13750
 GTATTACGCGGCCCGCCACCGCGGTGGAGCTCCAGCTTTTGTTCCTTTAGTGAGGGTTAATTTGAGCTGGCG
 V L R G R H R G G A P A F V P F S E G * F R A G X
Y Y A A A T A V E L Q L L F P L V R V N F E L A
 R I T R P P P R W S S S F C S L * * G L I S S W R

Appendix 3: Partial insert of clone 12. Sequence Range: 1 to 6506

Gene I: 1011...5984

10 20 30 40 50 60 70 80 90
 ACGTTTTAACTGGGTAACATGTACAGCATAACGGAATTTTCATATTCTCATAAGAAAAATTTTGTTCATTCTCTGTTACGGTAAAAGACATA
 T F * L G N M Y S I R N F H I L I R K I L F I P V I V K D I
 R F N W V T C T A Y G I F I F S * E K F C S F L L R * K T *
 V L T G * H V Q H T E F S Y S H K K N F V H S C Y G K R H

100 110 120 130 140 150 160 170 180
 ACAAAAACCTCCAGACAATAGAATTCCTTGGCAAAATATGGTAGCAGTGTATTATAAGGATTTTTAGCTTTCTGCATATTTTAAATGTTTTTC
 T K T P D N R I L G K Y G S S V I K D F L A F C I F L M F F
 Q K L Q T I E F L A N M V A V L L R I F * L S A Y F * C F S
 N K N S R Q * N S W Q I W * Q C Y * G F F S F L H I F N V F

190 200 210 220 230 240 250 260 270
 CAGCTTTTTAGCTAATTCCTTGTTTTTCCAGCCTTTTAGCCATTTCTTGTGTTTTTCCAGCTTTCTATCTGTTTTTGTTTTTTTTTAA
 Q L F S Y F L V F P A F * P F L V F F Q L S I C F S F F F *
 S F L A I S L F F Q P F S H F L C F S S F L S V F R F F F N
 P A F * L F P C F S S L L A I S C V F P A F Y L F F V F F L

280 290 300 310 320 330 340 350 360
 CTCCTTAAAAATTATTCATGTCATCCCAACATGAAAATTTGACTAATTAATTCAGTTAGCCAGTATCCGTCATCAAAATACGCGTTTT
 L L K I I H V I P T L K F R L I N S V S P V S V N Q I T R F
 S L K L F M S S Q H * N F D * L I Q L A Q Y P S I K L R V F
 T P * N Y S C H P N I E I S T N * F S * P S I R Q S N Y A F

370 380 390 400 410 420 430 440 450
CAGCTTTTGGCTTCACTCTTTGCAAAAGTGTGAAATATATGATTTACAAAGTGTGCAACATTATTAATATTAGATTGTTGATTTGTTT
Q L F G F T L C K V * K Y M I Y K V C N I I N I R L L I L F
S F L A S L F A K C E N I * F T K C A T L L I L D C * Y C F
S A F W L H S L Q S V K I Y D L Q S V Q H Y * Y * I V D I V

460 470 480 490 500 510 520 530 540
TCATTTTATTTCCAACTTTCTTGTGTTTTATTTGAAAGTGTGCAAAACATTCCATATCTATAAAAAATCTGTATAAAAATTCATCTGATTG
S F Y F Q L S C F I * K L C K H S I S I K I C I K F N P D S
H F I S N F L V L F E S C A N I P Y L * K S V * N S I L I R
F I L F P T F L F Y L K A V Q T F H I Y K N L Y K I Q S * F

550 560 570 580 590 600 610 620 630
TCTATTTTGGCTTCAGGGTGGTTTACTTAAATCCATATCTATCCAAATTTTAGCATATAAACAATAAATAAATAAATAAATATAGGATTA
S I L A S A W F T * F H I Y S N F S I * T K I K L N I G L K
L F W L Q R G L L N S I S I P I L A Y K Q K * N * I * D * N
V Y F G F S V V Y L I P Y L F Q F * H I N K N K I K Y R I K

640 650 660 670 680 690 700 710 720
CATGAAAACTTTCCATTTTGTATTATAAAGCAGACATTTTAAAGTTCATCTTCAAGTTTCATCTAGTTTTCTAATGAATGTTTTCTT
H E K L S I L L L K R H F K V L I F K F H L V F L M N V F L
M K N F P F C Y * N D I L K F S S S S F I * F F * * M F F F
T * K T F H F V I K T T F * S S H L Q V S S S F S N E C F S

730 740 750 760 770 780 790 800 810
CAACCTTTGATATGTAGCAACAATTTCTTACTAACTCCCTTAAATCAATGCCGCTATTGTTCCCATTTAATTGTTAAAATTCATAAT
Q P L I C S N N Y S Y * L L * Y N A A I V P H L I V K I H N
N L * Y V A T I I L T N S F N T M P L L F P I * L L K F I I
S T F D M * Q Q L F L L T P L I Q C R Y C S P F N C * N S *

820 830 840 850 860 870 880 890 900
CATCAAAATTCATATAAATGATAGTAATTTCCATAATTTCCATTAACATGATTGTTTATTTTATTCCTAACGCATATGCATATGACGC
H Q I T Y N Y D S N F I I F H * L I V L F Y S * R I C I * R
I K L H I I M I V I S * F S I N * L S Y F I P N A Y A Y D A
S S N Y I * L * * * F H N F P L T D C L I L F L T H M H M T

910 920 930 940 950 960 970 980 990
CGTATCTTCAATAATGAAATTTACTGAAAACAAATATCAGTTAGTCACAGTAATAAATGTTGAAATGCAGAAGTGTACAAGTCTTAC
R I F * * * N Y Y * K Q I S V S H S N K C * N A E V Y K S Y
V S S N N E I I T E N K Y Q L V T V I N V E M Q K C T S L T
P Y L L I M K L L L K T N I S * S Q * * M L K C R S V Q V L

1000 1010 1020 1030 1040 1050 1060 1070 1080
AAGGTGACATGAAGTGGATTATGATAACGCTTTTCAATATACTTGCCCAACGAATTAATGTTGTAACGGTAAGTTGCTCAATAAATGA
K V T * T G L * * R F S L Y L P T Q L L L * T V S C S I K *
R * H E L D Y D N A F H Y T C Q R N Y C C K R * V A Q * N D
Q G D M N W I M I T L F I I L A N A I T V V N G K L L N K M

1090 1100 1110 1120 1130 1140 1150 1160 1170
CACGTCAGTTGGCTTAACACGAAATATCATCGCTGAAGATATAATAAGCATAACACAGATCATACATCCTAATCATTACATGCCATTGC
H V S C V N T K Y H R * R Y N K H N N R S Y I L I I T C H C
T S V A L T R N I I A E D I I S I T T D H T S * S L H A I A
T R Q L R * H E I S S L K I * * A * Q Q I I H P N H Y M P L

1180 1190 1200 1210 1220 1230 1240 1250 1260
AATCCTAAATTTGTCAGATAATGATATACATTGAATTTACAGAAAGCTTTCCAAAAATGAAGAGGCTAAGTTGAGATGAACAAATTTT
N P K L S R * * Y T L N F T E A F T K * R G * V E D E T N F
I L N C R D N D I H * I S Q K L S Q N E E A K L K M K Q I L
Q S * I V E I M I Y I E F H R S F H K M K R L S * R * N K F

1270 1280 1290 1300 1310 1320 1330 1340 1350
ATTTTAAAAAATTGGTAATACTGTTGACTAAATTTTAAAGAAATAACGACGAAATCGATTTTGAATATCTTCAAGCATTAGCCTACTTC
I L K N W * Y C * L N F K E I T T K S I L K Y L Q A L A Y F
F * K I G N T V D * I L K K * R R N R F * N I F K H * P T S
Y F K K L V I L L T K F * R N N D E I D F E I S S S I S L L

1360 1370 1380 1390 1400 1410 1420 1430 1440
AGGTTACGTTCCGGATGTTACTACACAAATTGGGCTCAATATCGACAAGGTTCACTGATGTTTGAATAATTTTGTATTAAATAGTCA
R L R S R M L L H K L G S I S T R F T D V * N N F V I K * S
G Y V R G C Y Y T H W A Q Y R Q G S L M F E I I L L L N S Q
Q V T F A D V T T Q I G L N I D K V H * C L K * F C Y * I V

1450 1460 1470 1480 1490 1500 1510 1520 1530
ATAATTTAAAGGATTGTACAGTTTTTCAATAAAAATTAGAAAACGATGGAATCATTTCAGATTTTCGATCTTAAAGTGAATTTTAA
I I * K D C T V F Q I K I R K L M E S F Q D F D L K S E F *
* F K R I V Q F F K * K L E N * W N H F K I S I L K V N F K
N N L K G L Y S F S N K N * K T D G I I S R F R S * K * I L

1540 1550 1560 1570 1580 1590 1600 1610 1620
ACTATACGAAATATTAGTGGAAAATAAACATCGATTAATGGTTCAAGTTTCAACATAAACTCAAATTTTATTATATTGATATTTCCGTA
I I R N I S G K * T S I N G S S F N I N S N F I Y I D I S *
L Y E I L V E N K H R L M V Q V S T * T Q I L F I L I F P E
N Y T K Y * W K I N I D * W F K F Q H K L K F Y L Y * Y F L

1630 1640 1650 1660 1670 1680 1690 1700 1710
GTGTACATCGTTTCAACGTTGCTTTTTTTTTTCTATCATAACTAATGTGCTGAAAAAATAAATGAGCTAATTAAGTAAAACAAATAT
V Y I V S T C A F F F Y H K L M C * K K * M S * L K * N K Y
C T S F Q R V L F F S I I N * C A E K N K * A N * S K T N I
S V H R F N V C F F F L S * T N V L K K I N E L I K V K Q I

1720 1730 1740 1750 1760 1770 1780 1790 1800
CGTATTACTGTCGGAATGTAATATACATTTTCATCTTAAAGGAGAAGGCAAAATTCITGCGGGAAGATATCCCAATAGGCCCTGTGCACCTC
R I T V G N V I Y I F I L K E K A N S C R K I S Q * A C A L
V L L S E M * Y T F S S * R R R Q I L A G R Y P N R P V H S
S Y Y C R K C N I H F H L K G E G K F L P E D I P I G L C T

1810 1820 1830 1840 1850 1860 1870 1880 1890
ACATTTCTTATGTCATTGCGCAAGTTGACGAGAAAGGCACATCAATGTAAGTTACAATAATACTTGTAAATGAGAAACATAAACTAAAT
T F F M H S P K L T R K A H Q C K L Q I I L V M R N I N * I
H S L C I R Q S * R E R H I N V S Y K * Y L * * E T * T K F
H I L Y A F A K V D E K G T S M * V T N N T C N E K H K L N

1900 1910 1920 1930 1940 1950 1960 1970 1980
TTGCGATTTTGTGCTGACAGTGGACATTCAGACTATTTAAATTTTTTGTGAGCTATTCAAGTTGTACATACTGGCAGCATTTCATGTTGGA
F A F C S Y S G H S D Y L K F L S A I Q V V T Y W Q H S C G
S H F A R T V D I Q T I * N F C Q L F K L S H T G S I H V E
F R I L L V Q W T F R L F K I F V S Y S S C H I L A A F M W

1990 2000 2010 2020 2030 2040 2050 2060 2070
ACATTCAGTAAAGATAGGCGGTTTTGCATAATATGCTCTTCTATGATGACCATATCAGCAAATATAATCTATCTGTAAATTTCCCTATT
I F S K D R R F C I I C S S M M T I I S K Y N L S C K F P I
H S V K I G G F A * Y A L L * * P L S A N I I Y P V N F L F
N I Q * R * A V L H N M L F Y D D H Y Q Q I * S I L * I S Y

2080 2090 2100 2110 2120 2130 2140 2150 2160
TCAATAACTTTTAACTTAGGGCATTGAAATGGAATGACGAAGATACCGAATGGTCAAAAGGGATGTAATCAGTGAATAAACTGAGA
S I T F N L R A F E W N D E D T E W S K G M Y S R V I K L R
Q * L L I L G H L N G M T K I P N G Q K G C T H V * * N * E
F N N F * S * G I * M E * R R Y R M V K R D V L T C N K T E

2170 2180 2190 2200 2210 2220 2230 2240 2250
GAGAACGATCCAACTGAAAATCTTCTTCTTATGGTGGCTATAACTTCGGATCGTCAACTTTTACTGTTTGGTTTCATTITTTTTAAAG
E N D P T L K I L L S Y G G Y N F G S S T F T V C F I F L K
R T I Q H * K S F F L M V A I T S D R Q L L L F V S F F * R
R E R S N T E N P S F L W W L * L R I V N F Y C L F H F F K

2260 2270 2280 2290 2300 2310 2320 2330 2340
ATTTACGCTCGCTCAACTCATGAATTTTCTTTTATTGGGAAAGCAGATTGGTATTCTCGATAATTATATTGAACAGGTTATTCGGAA
I Y A R S T H E F S F D S G K Q I G I P R * L Y * T G Y S E
F T L A Q L M N F L L I R E S R L V F L D N Y I E Q V I R N
D L R S L N S * I F F * F G K A D W Y S S I I I L N R L F G

2350 2360 2370 2380 2390 2400 2410 2420 2430
TAGAGTGGAAATATTACGGGAGAACAGTAATGAATAGTACTATTTCTGACGATTGATTTGTGAAACTTTGACGAGTCACTTTTCCCTTTT
* S G N I T G E Q * * I V L F L T I D L * N F D E S L F P F
R V E I L R E N S N E * Y Y F * R L I C E T L T S H F S L L
I E W K Y Y G R T V M N S T I S D D * F V K L * R V T F P F

2440 2450 2460 2470 2480 2490 2500 2510 2520
GATCTGTATATATATATAGAGTGTCCAACTGTAACATACATATAACAAATATTGGGGGATAACAGGATAGTTATTTAGATACATAAAA
D L Y I Y I * S V Q R N Y T Y T K Y W A I T G * L F R Y I K
I C I Y I Y R V S N V T I H I P N I G R * Q D S Y L D T * N
* S V Y I Y I E C P T * L Y I Y Q I L G D N R I V I * I H K

2530 2540 2550 2560 2570 2580 2590 2600 2610
TCAACATGAACACGATATATCAGTCATATAATAAATCAATAGAAATCAGGTAGAGATACGGGTGATAAGTTACAAGAATCACACAGATTT
S T * N T Y I S H I I N Q * K S R R D T R D K L Q E S H R F
Q H E T R I S V I * * I N R N H V E I R V I S Y K N H T D L
I N M K H V Y Q S Y N K S I E I T * R Y A * * V T R I T Q I

2620 2630 2640 2650 2660 2670 2680 2690 2700
GCSCAGGATAAAATGGGTAGCCGACATGCAGACATATTACAGATAATAGAATAGTTAGGCAATGTAATGAAAAGAAAGAAAGAGCGATG
A Q D K M G S R H A D I L Q I I E * L G N V N E K K E R A M
R R I K W V A D M Q T Y Y R * * N S * A M * M K R K K E R W
C A G * N G * P T C R H I T D N R I V R Q C K * K E R K S D

2710 2720 2730 2740 2750 2760 2770 2780 2790
GGGAAGAGGAGACTGTTTATTACTCATTGGCTAGATTTGTTGTTATAGATAAAAAGAAAGATAGAGAACATGTAATAGCGGGAATGT
G K K E T V * L L I G * I C C Y R * K E R * R T C N S G K C
G R R R L F D Y S L A R F V V I D K K K D R E H V I A G N V
G E E G D C L I T H W L D L L L * I K R K I E N M * * R E M

2800 2810 2820 2830 2840 2850 2860 2870 2880
TGGCGGGTAAATTTACTGTGAATGACATTCAAATAGCACAAATATAAAAATAACGGATGGCAGGAAATTAATTTTACATTGAAGAAAGATT
W R V I Y C E * H S N S T N I K * R M A G N * I S H * R K I
G G * F T V N D I Q I A Q I * N N G W Q E I K F H I E E R L
L A G N L L * M T F K * H K Y K I T D G R K L N F T L K K D

2890 2900 2910 2920 2930 2940 2950 2960 2970
GTGAATAGCAACTTCTTTAGCTTGGCTGTCAATTATACAAGTCATAAAACTTCGGGTTTCCCGACACTAACACATTGACAAATGATTTC
V N S N F F S L A V N Y T S H K T S G F P T L T H * Q M I S
* I A T S L A W L S I I Q V I K L R V S R H * H I D K * F Q
C E * Q L L * L G C Q L Y K S * N F G F P D T N T L T N D F

2980 2990 3000 3010 3020 3030 3040 3050 3060
AAAAGAGCTGCAAAACAGCGATGCAATGTATCTCAATGTCTCCAACTTTTACGTCCTCACTTTTTCTTCTGCTGAAATATTGAAGGCAAT
K R A A N T * C I V S Q M F S N F T S H F F L A E I F E G N
K E L Q T R D A L Y L K C S P T L R L T F F L L K Y L K A I
K K S C K H V M H C I S N V L Q L Y V S L F S C * N I * R Q

3070 3080 3090 3100 3110 3120 3130 3140 3150
 AGCAAAGTCGGGGGAAAAACCGGAAGCATTTCATTTCAGTCAGCTACAAACATTTCCTTCGCAAGCATAAAATTTGATGGATTTCGATCTCGACTG
 S K V G G K T E A F H S V S Y N I S S Q A * I * W I R S R L
A K S A E K R K H F I Q S A T T F L R K H K F D G F D L D W
 * Q S R R R K N G S I S F S Q L Q H F F A S I N L M D S I S T

3160 3170 3180 3190 3200 3210 3220 3230 3240
 GGAATATCCAAACCGGTGTAGCAGAGGAACATGCCAAACTTGTGAGGTGAGTGTAAATTCATCACCAGACTAACGTTTAATGTTTTGTTTA
 G I S N R C S R G T C Q T C * G E C N S I T D * R L M F C L
E Y P T G V A E E H A K L V E V S V I P S P T N V * C F V Y
 G N I Q P V * Q R N M P N L L R * V * F H H R L T F N V L F

3250 3260 3270 3280 3290 3300 3310 3320 3330
 CTTGAAATCTCTGGCACTATTGTAGCAATCAGATATATGCCATCAAAATTTTAATTGACATTCCAAAAGAGACAATTCTCGCTTCAAAAAT
 L E I S G T I V A I R Y M P S N F N * H S K R D N S R F K N
 L K S L A L L * Q S D I C H Q I L I D I P K E T I L A S K M
 T * N L W H Y C S N Q I Y A I K F * L T F Q K R Q F S L Q K

3340 3350 3360 3370 3380 3390 3400 3410 3420
 GAGTTGGTTTCAAAAATAAAAAGTAAATATATTACATCCATCTGAAAGAGGGGAGAAAAATTCAAAAAGAAATTTTGTAGGAAATGAAAGCG
 E L V S K I K L K Y I T S I * K R E K N S K R I L * E M K A
 S W F Q K * N * N I L H P S E R G R K I Q N E F C R K * K R
 * V G F K N K I E I Y Y I H L K E G E K F K T N F V G N E S

3430 3440 3450 3460 3470 3480 3490 3500 3510
 GGGTTCGGGAAAGCCAAAGGAATCCGGCAAGCAACAATTACTTCTTACCGCTGCTGATCAGCCGGGAAAATGACTATTGATGAAAGT
A F V E E A K E S G K Q Q L L L T A A V S A G K M T I D E S
 R S W K K P R N P A S N N Y F L P L L Y Q P G K * L L M K V
 G V R G R S Q G I R Q A T I T S Y R C C I S R E N D Y * * K

3520 3530 3540 3550 3560 3570 3580 3590 3600
 TACAATGTTCAATCCCTTGGAAAGTAAAAAAATTTGCTTTTTTTCATATACTGACATTAAATTTGACACGCAATAAAAATTTACAATACITCA
Y N V Q S L G K * K K L L F H I L T L N L T R N K I Y N T S
 T M F N P L E S K K N C F F I Y * H * I * H A I K F T I L Q
 L Q C S I P W K V K K I A F S Y T D I K F D T Q * N L Q Y F

3610 3620 3630 3640 3650 3660 3670 3680 3690
 AATATAATCTTGTAGCAATTCGAAATTTTCAGAAAGTTTGGATCTTCTATTTCTGATGTCGTATGATTTACACGGTAGTTGGGAGAAGAAACG
 N I I L * H S K F S E V W I F Y F * C R M I Y T V V G R R T
 I * S C S I R N F Q K F G S S I S D V V * F T R * L G E E R
 K Y N L V A F E I F R S L D L L F L M S Y D L H G S W E K N

3700 3710 3720 3730 3740 3750 3760 3770 3780
 TCGACTTACACGGAGAGTTGCGTCCGACGGAGAGAGAAACATCCGGAACCTGGCGTCTGTAATACCGTTAGTTCTTGAATATATAAAAAAT
 S T Y T E S C V R R R E K H P E L A S V I P L V L E I Y K N
 R L T R R V A S D G E R N I R N W R L * Y R * F L K Y I K I
V D L H G E L R P T E R E T S G T G V C N T V S S * N I * K

3790 3800 3810 3820 3830 3840 3850 3860 3870
 TTCTAATTTTTTCTTGTGTTGCAAGAGCGAATTTCTTGTGATTTTCAGGAATTTGCAGCAAATTTATGGGAGAGAAAGGTATGCCGAAA
 F * F F L V C M K T N S * * F Q E F A A N Y W A E K G M P K
 S N F F L F A * R R I L D D F R N L Q Q I I G Q R K V C R N
 F L I F S C L H E D E F L M I S G I C S K L L G R E R Y A E

3880 3890 3900 3910 3920 3930 3940 3950 3960
 CAGAAAATATCATTGGGATTCGGCTTACAGTCGGGGATGGACATTAAGTAATCCCTTCGGAAAACGGCAATTGGAGCGGAGGGGTGATCGT
Q K I I I G I P A Y S R G W T L S N P S E T A I G A E G D R
 R K L S L G F R L T V G D G H * V I L R K R Q L E R R V I V
 T E N Y H W D S G L Q S G M D I K * S F G N G N W S G G * S

3970 3980 3990 4000 4010 4020 4030 4040 4050
CCATCATCGCCATCGACAAACAATCCCGCNGGGCTACTGCGGCTTATTGGGAGGTTTAAATTTAAAGTTCTCATTCTTTAAATACAT
P S S P S T T N P A G G T A A Y W E V * F K V L I Y F * I H
H H R H R Q Q I P X A V L R L I G R F N L K F S F T F K Y I
S I I A I D N K S R X R Y C G L L G G L I * S S H L L L N T

4060 4070 4080 4090 4100 4110 4120 4130 4140
TTCITGCTGCTGATGAATTTCTCAAATTCATTATTAATGAATGTAACCCATTTAAGATATGCAAAATATGTGAANGAAGGTGGCAAGGAA
F L L L M N F S N S I I N E C N H F K I C K Y V K E G G K E
S C C * * I S Q I P L L M N V T I L R Y A N M * K K V A R K
F L A A D E F L K F H Y * * M * P F * D M Q I C E R R W Q G

4150 4160 4170 4180 4190 4200 4210 4220 4230
ACGGTGGATAAAAAAGTGTGGAGCGTATATGGTAAAAGGAGATCAGTGGCAGGTTACGATAATGAGGAACTATTAAAATTAAGTA
T V D K K S V G A Y M V K G D Q W H G Y D N E E T I K I K V
R W I K K V L E R I W * K E I S G T V T I M R K L L K L K *
N G G * K K C W S V Y G K R R S V A R L R * * G N Y * N * S

4240 4250 4260 4270 4280 4290 4300 4310 4320
ATTTTATTTGCTTATTGTTGATAGTCAAAAATAAAAAAGTGAATTTGATAGTCAAAAATAAGAAAAGATAATAAGATAGTCGAAA
I F I S L I V V D S Q K * K S D L I V K K * E K I I R * S K
F L F R L L L L I V K N K K V I * * S K N K K R * * D S R K
N F Y F A Y C C * * S K I K K * F D S Q K I R K D N K I V E

4330 4340 4350 4360 4370 4380 4390 4400 4410
AATAAAAAATTAAAAAATACGTTGCAATATATCTTGCCCTTAACITTAATTTAAAGAGCACCATATAATTTTATAATGTTTAATAAGAG
N K K L K N N V A N I S C L N F N L K S T I N F I M F N K K
I K N * K I T L Q I Y L A L T L I * R A P * I L * C L I R R
K * K I K K * R C K Y I L P * L * F K E H H K F Y N V * * E

4420 4430 4440 4450 4460 4470 4480 4490 4500
AAATTAGTTTCGGAGTTTACTCGTGAATTTGTTTACTTTAGAAATTAATTTTTCAGATGAAATGGCTGAAAGAGGAGGATTANGGCGGT
K L V S E F T R V I V L L * N * N F Q M K W L K E E D X G G
N * F R S L L V * L F Y F R I K I F R * N G * K R R I X A V
E I S F G V Y S C N C F T L E L K F S D E M A E R G G L X R

4510 4520 4530 4540 4550 4560 4570 4580 4590
GCCITATGTGGACTCTTGATTTTGATGATTTCAAGGTACAAAGTTTGGCCAAAGGTCCATATCCACTGTTAAATGCTATCAATAATGGG
A F M W T L D F D D F K G T S C G K G P Y P L L N A I N N G
P L C G L L I L M I S K V Q V V A K V H I H C * M L S I M G
C L Y V D S * F * * F Q R Y K L W Q R S I S T V K C Y Q * W

4600 4610 4620 4630 4640 4650 4660 4670 4680
CTTGAAAGTGAGGTAAGACATGTTTCAAGAAATATTTTAAACAATTCIGTTTTTCTTTCAATACGTTTGGATTTAAATTTATGCTGATGT
L E S E V R H V S K K Y F * Q F C F S F N T F G F K L C * C
L K V R * D M F Q R N I F N N S V F L S I R L D L N Y A D V
A * K * G K T C F K E I F L T I L F F F Q Y V W I * I M L M

4690 4700 4710 4720 4730 4740 4750 4760 4770
TTGTAATAAATTAATAAAAAAAGGCGAATGGATGAATCAGAATAGACTTTTCGGTGGGACCATACGGAAAAATGACTCTATGAATTAT
L * I N * * K K R R M D E S E * T F G G T I R K N D S M N Y
C K * I N K K K G E W M N Q N R L S V G P Y G K M T L * I I
F V N K L I K K K A N G * I R I D F R W D H T E K * L Y E L

4780 4790 4800 4810 4820 4830 4840 4850 4860
AGGATAAATAGAAATTTCTGTCGAAAGTATGACTACTTAACITTTGATTAGCTGGATGGGGCATCAATTTCTATATTGACGTTGTTCAA
R I N R I S V R K * * L T N F * L A G W G I N F Y I D V V Q
G * I E F L S E S D D S L T F D * L D G A S I S I L T L F K
* D K * N F C P K V M T H * L L I S W M G H Q F L Y * R C S

4870 4880 4890 4900 4910 4920 4930 4940 4950
ACAAAATCTTAAATTTTAAATTTATCTCTTGAATAATTTTAAAAATTTAACACATTTTGTAGCAAACACCATCAACAAGAGCAGTACCG
T K S * I F N Y L L K I N F * N L T H F * Q T P S T R A V P
Q N L K F L I I S * K L I F K I * H I F S K H H Q Q E Q Y R
N K I L N F * L S L E N * F L K F N T F L A N T I N K S S T

4960 4970 4980 4990 5000 5010 5020 5030 5040
GAAACTACAGAAGACACTGAAAGTTGAACAATGACGGGAAGCACCGGGAAGTGCATAATGATACTGAAATGGAAACAGCAGAAATGCCAGAA
E T T E D T E V E T M T E A P G T A N D T E M E T A E M P E
K L Q K T L K L K Q * R K H R E L Q M I L K W K Q Q K C Q K
G N Y R R H * S * N N D G S T G N C K * Y * N G N S R N A R

5050 5060 5070 5080 5090 5100 5110 5120 5130
ACTACGGAAGATATTGAAATGAAACATCGACCGGACATCCGACTTTTTCAACAAATTTGGGATTAAATCCITTCATGATATTTATAAAA
T T E D I E I E T S T E A S D F F N K F G I K S F M I F I K
L R K I L K L K H R P R H P T F S T N L G L N P S * Y L * N
N Y G R Y * N * N I D R G I R L F Q Q I W D * I L H D I Y K

5140 5150 5160 5170 5180 5190 5200 5210 5220
TTAGTAAAAATGTAATCGGCAGATGAAGAGATGTACACAGGATCATACGGAGAATTGACTCGATGATTTGTGGCATAAATGGAATTT
L V K N V K S A D E E M Y T G S Y G E L T R * F V A * M E F
* * K M * N R Q M K R C T Q D H T E N * L D D L W H K W N F
I S K K C K I G R * R D V H R I I R R I D S M I C G I N G I

5230 5240 5250 5260 5270 5280 5290 5300 5310
CTGCCGAAAGTAATGACTCATAACTTTTACTACCTAGATGGAACGTCRAATTTCTATATTGATCATTACGTGCATCAAACACAGTCTTT
L P E S N D S * L L I T * M E R Q F L Y * S L R A S N T V F
C R K V M T H N F * L P R W N V N F Y I D H Y V H Q T Q S L
S A G K * * L I T F D Y L D G T S I S I L I I T C I K H S L

5320 5330 5340 5350 5360 5370 5380 5390 5400
AAATCTGAAATTTATCTCTTGAATTTAATTTATAAAATTTAACAAACATTTTGTAGCAGACATCATCAACACAGCAGCACCAGGAAACAC
K S * I I S * N L I Y K I * Q T F L A D I I N N S S T G N H
N L E L S L E I * F I K F N K H F * Q T S S T T A A P E T T
* I L N Y L L K F N L * N L T N I F S R H H Q Q Q Q H R K P

5410 5420 5430 5440 5450 5460 5470 5480 5490
AGAAGGCACCTGAAAGTTGAACAACAGCAGCGTGTGTAACCCGGTTTGTATTTTCAAGTTGAAAAAGCTATTCCGTTTCTCTTATTAT
R R H * S * N N D E R C * T R F V I F K L K K L F R F S Y Y
E G T E V E T T T S V V K P G L L F S S * K S Y S V S L I I
Q K A L K L K Q R R A L L N P V C Y F Q V E K A I P F L L L

5500 5510 5520 5530 5540 5550 5560 5570 5580
CATTGGGATAATGGTCAGTCATCTGAGTTACTGCTTGTGTCATTCTAGGCGGTACAGAAAATACTCATGCAAGATGAGAAATACTTAGG
H L D N G Q S S E L L L V H S R Q Y R K Y S C K D E K * L R
I W I M V S H L S Y C L C I L G S T E N T H A K M R N N L G
S F G * W S V I * V T A C A F * A V Q K I L M Q R * E I T *

5590 5600 5610 5620 5630 5640 5650 5660 5670
AAATCTTCAGTATTTTTTAAATGAATATTTATAAAATTTGATAAGCACACCGACATGGATCACTGGCAATCATTTAAAAATATTC
K S S V F F L K * I F I K I * * A H R H G S L A N H L K Y S
N L Q Y F F * N E Y L L K F D K H T D M D H W Q I I * N I Q
E I F S I F F K M N I Y * N L I S T P T W I T G K S F K I F

5680 5690 5700 5710 5720 5730 5740 5750 5760
ATATGTTTATTATTTTTCATTGCCACATATGCCGCATACAATTTCTTCATTCGTTTTTTTGTCTCGAAAACCTTCTCGATTCTACGGTAT
I C L L F L H C H I C R I Q F L H S F F V S K N F S I L R Y
Y V Y Y F F I A T Y A A Y N F F I R F L S R K T S R F Y G M
N M F I I S S L P H M P H T I S S F V F C L E K L L D S T V

5770 5780 5790 5800 5810 5820 5830 5840 5850
 GAGAACCCTGAAACAAATGAATGCACACTACTGAGATCTGAAATCTATTTTAAAGGTGAGGAATGTCCGGAGCCCGATGGTTTATTTCCGCATC
 E N P E T M N A L L R S E I Y F K V R N V R S P M V Y F R I
 R T L K Q * M H Y * D L K S I L R * G M S G A R W F I S A S
 * E P * N N E C T T E I * N L F * G E E C P E P D G L F P H

5860 5870 5880 5890 5900 5910 5920 5930 5940
 ACAGTGATTGCCATTGTTCATTAAITGGCGAAATAACTATCCACACATAATGGAATGTCCGGTGGGCACCTTTTTTGATGACACAATCA
 T V I A I C S L I A Q I T I H I * W N V R W A P F L M T Q S
 Q * L P F V H * L R K * L S T H N G M S G G H L F * * H N Q
H S D C H L F I N C A N N Y P H I M E C P V G T F F D D T I

5950 5960 5970 5980 5990 6000 6010 6020 6030
 AAGTTTGAATTATATGAGGAACGCGCCGGATACATGCAAAATGATTGGCATTITATGTGATATATATATTTGTTGGAATAAAAATCTAG
 K F A I I * G T R R I H A N D L A F Y V I Y I F V G I K I *
 S L Q L Y E E R A G Y M Q M I W H F M * Y I Y L L E * K S R
K V C N Y M R N A P D T C K * F G I L C D I Y I C W N K N L

6040 6050 6060 6070 6080 6090 6100 6110 6120
 GGTAAAAGTTCTAGAACAAAAAAGACCGCATAAATCATATTAAATAATCTGCAAAAAATTGCAITAAAAATTTATTGAATTTCAAGG
 G K S S R T K K K R R I N H I K * S A K I A L K I Y * I S R
 V K V L E Q K K K D A * I I L N N L Q K L H * K F I E F Q G
 G * K F * N K K K K T H K S Y * I I C K N C I K N L L N F K

6130 6140 6150 6160 6170 6180 6190 6200 6210
 AACGAGTTTGCATGTCATTGTCTCTCTAAAGTATTATTTCAAACCCACTCCTGAATCTTTTCAAAATTTAAAAATTTCTTTTCAGCGAAAT
 N E F A C H L L S L K Y Y F K P T P E S F T I * K F F Q R N
 T S L H V I C S L * S I I S N P L L N L S Q F K N S F S E I
 E R V C M S F A L S K V L F Q T H S * I F H N L K I L S A K

6220 6230 6240 6250 6260 6270 6280 6290 6300
 ATAAGCTATAAATGAATAATATTGATTCAATTTTATTAGTTAATCATCCAACTAAATCTTGAAGCATAAAGATGTTCAATTTATGAAAC
 I S Y K * I I L I Q F Y * L I I Q L N L E S I K M F N L * N
 * A I N E * Y * F N F I S * S S N * I L K A * R C S I Y E T
 Y K L * M N N I D S I L L V N H P T K S * K H K D V Q F M K

6310 6320 6330 6340 6350 6360 6370 6380 6390
 TTTTTTAATAAAAAATTAACATAAATTTACATAAAAAAGTAATGATACGCATTAAACAATTTAATACATTATTTAACATTTATAAAT
 F F * * K I K T * F T * K S N D T H * T I * Y I I * H L * I
 F F N K K L K H N L H K K V M I R I K Q F N T L F N I Y K L
 L F L I K N * N I I Y I K K * * Y A L N N L I H Y L T F I N>

6400 6410 6420 6430 6440 6450 6460 6470 6480
 ATATCAGTACAATGCAAGCATCCTCCAAATGAATAAACAAACAACCGCTAAAGAAAGAAAAGTCCCAACAATCATTIAGTCATTCAATA
 I S V Q C K H P P N E * T N N R * R R K S P N K S F S H S I
 Y Q Y N A S I L Q M N K Q T T A K E E K V P T N H L V I Q *
 Y I S T M Q A S S K * I N K Q P L K K K K S Q Q I I * S F N

6490 6500
 AACTACTCAGCCAAATGTAATTTCA
 N Y S R Q M * F X
 T T H A K C N F X
 K L L T P N V I S


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L  --ACACCAT-CAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  GCAGATGATACTGAAACTGAGATATCGGAAATGCCGGAAACTACAGATAGTAGTGAATG--- 720
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  -----CCAGGAACTGCAGATGATACTGAAACTGAAACATCGGAA 759
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  -----
Z  ATGCCGGAAACTACAGATAAT 780

L  ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  -----ACTGAAATGCCAGGAACTTCGGATGATACTGAAATGGAAACAACAGAA 828
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  -----
Z  ATGCCAGAAAT 840

L  -----ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  ACAGAAGATACTGAAACTGAAGCATCAGAAATGCCGGAAACTACAGATAATACTGAAATG-----900
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  ----ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  CCAGGAACTGCGGATGAT-GCTGAAATGGAAACAACAGAAATGCCAGAAATTACAG--AGGAT- 960
   ** * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  ---ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  ATTGAAACTGAAACATCGGAAATGCCGGAAACTACAGATAATAGTGAATGCCAGGAACT--- 1020
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  --ACACCAT-CAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  GCAGATGATACTGAAACTGAAACATCGGAAATTCGGGAAACTACAGATAATACTGAAATG--- 1080
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  ----ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  CCAGGAACTGCGGATGATA-CTGAAATGGAAACAACAGAAATGCCAGAAATTACAG--AGGAT- 1140
   ** * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  ---ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATGACG 60
Z  ATTGAAACTGAAACATCGGAAATGCCGGAAACTACAGATAATAGTGAATGCCAGGAACT--- 1180
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  -----ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTT 48
Z  GCAGATGATACTGAAACTGAAACATCGGAAATGCCGGAAACTACAGATAATACTGAAATG 1240
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
L  GAAGCAATGACG 60
Z  -----
L  ---ACACCATCAACAAGAGCAGTACCGGAAACTACAGAAGACACTGAAGTTGAAGCAATG 57
Z  CCAGGAACTTCGGATGATACTGAAATGGAAACAACAGAAATGCCAGAAATTACAGAAGAT 1300
   * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
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2) Introns and Exons of identical regions of clones 1 and 9 showing discrepancies

I versus III

Exon	Identities	Intron	Identities	Exon	Identities	Intron	Identities
		f	37	A	100	a	100
G	95*	g	100	B	100	b	100
H	100	h	100	C	100	c	100
I	100	i	100	D	100	d	100
J	100	j	100	E	71	e	53
M	100						

9 Publications and conference abstracts

Manuscript from this study

Babila J. Tachu, Smitha Pillai, Richard Lucius, and Thomas Pogonka: *Acanthocheilonema viteae* Chitinase Genes: Structure and Expression. (In preparation).

Conference abstracts from this study

Babila J. Tachu, Richard Lucius, Jörg Hirzmann, Vincent P.K. Titanji, Thomas Pogonka, (2003): Molecular characterisation of *Acanthocheilonema viteae* chitinase. Meeting of the Federation of African Societies for Biochemistry and Molecular Biology held in Yaounde, Cameroon.

Babila J. Tachu, Richard Lucius, Jörg Hirzmann, Vincent P.K. Titanji, Thomas Pogonka (2004): Molecular characterisation of *Acanthocheilonema viteae* chitinase genes. 21st Meeting of the German Society for Parasitology meeting held in Würzburg, Germany.

Michel J. Sereda, Smitha Pillai, Babila J. Tachu, Susanne Hartmann, Thomas Pogonka, and Richard Lucius (2005): Studies on subunit vaccines against infection with the parasitic nematode *Acanthocheilonema viteae* in a rodent model. Vaccine Congress Berlin 2005