# STUDIES ON DEXTRANSUCRASE

II. Factors Affecting the Formation of Riboflavinylglucoside in Growing Cultures of *Leuconostoc mesenteroides* 

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Riboflavinylglucoside was isolated at first by Whitby (1950, 1952, 1954) from the incubation mixture of riboflavin with homogenates of rat liver and with aqueous extracts of acetone dried powder of rat liver. Not only riboflavinylglucoside, but also various kinds of sugar compounds of riboflavin were obtained by Katagiri, Tachibana and Yamada with enzyme preparations from a mutant of Aspergillus oryzae (Katagiri and Tachibana, 1953; Tachibana and Katagiri, 1955), Escherichia coli (Katagiri, Imai and Yamada, 1954, 1957) and from Clostridium acetobut yricum (Tachibana, 1955).

In the previous paper (Suzuki and Katagiri, 1964), it was found that *Leuconostoc mesenteroides* produced a remarkable amount of riboflavinylglucoside in addition to dextran, when it was grown in a medium containing sucrose and riboflavin. The present paper deals with cultural conditions on the formation of riboflavinylglucoside by the bacterium.

#### METHODS

Two strains of *Leuc. mesenteroides*, IFO. 3426 and L. 20, were used. The stock cultures were maintained by frequent transfers on agar slants (A) and (B). The compositions of the agar slants are as follows; (A) is a malt-extract agar containing 1% glucose, 0.1% yeast extract, 2% CaCO<sub>3</sub> and 2% agar-powder and adjusted pH to 7.0, (B) is a surose agar containing 4% sucrose, 0.5% K<sub>4</sub>HPO<sub>4</sub>, 0.1% NaCl, 0.02% MgSO<sub>4</sub>· 7H<sub>3</sub>O, 2 mg% FeSO<sub>4</sub>· 7H<sub>2</sub>O, 2mg% MnSO<sub>4</sub>, 0.06% (NH<sub>4</sub>)<sub>3</sub>SO<sub>4</sub>, 0.25% yeast extract, 0.05% polypeptone, 4% CaCO<sub>3</sub> and 2% agar-powder and adjusted pH to 7.0. To prepare the inoculum, one loopful of the bacterium from an agar subculture was suspended in 10 ml of a sterilized seed medium and incubated at 30°C for one day under stationary conditions. About 2 ml of this seed culture were used as the inoculum. The estimation of riboflavin-compounds was carried out by the method reported previously (Suzuki and Katagiri, 1964).

#### RESULTS

## 1. Effect of Components of Seed Media

Two kinds of seed media were used in the preparation of the inoculum and

their components were similar to the stock cultures (A) and (B), but both calcium carbonate and agar-powder were omitting. Fifty ml of a medium containing 10% sucrose, 0.5% KH<sub>2</sub>PO<sub>4</sub>, 0.1% NaCl, 0.02% MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.03% (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.05% polypeptone, 0.15% yeast extract and 40 mg% riboflavin adjusted pH to 7.0 with sodium hydroxide were inoculated with 2 ml of a 24-hours' culture of *Leuc. mesenteroides* L. 20, grown in the seed medium and then incubated at 30°C for 2 days. At certain intervals portions (5 ml) of fermentation liquid were withdrawn and the amount of riboflavinylglucoside produced was determined. It will be seen in Table 1 that the seed medium (B) containing sucrose was much

(1). Alte	Type of	B2-glucoside	B2-oligo saccharides			
Condition	seed medium #	Ο. D. at 530 mμ	initial	final	(mg/50ml)	produced (mg/50ml)
Shaking	(B)	0.185	20.0	11.5	6.5	0
Stationary	(B)	0.170	20.0	11.4	7.5	0
Shaking	(A)	0.176	20.0	18.8	+	0
Stationary	(A)	0.169	20.0	18.7	+	0

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	Effect of	components	of	seed	media		
rs'	incubation						

(II). After	II). After 18 hours' incubation								
Condition	Type of	Growth	B <sub>2</sub> present (	(mg/50 ml)	B2-glucoside produced (mg/50 ml)	B2-oligo saccharides			
Condition	seed medium #	530 mµ	initial	final		(mg/50 ml)			
Shaking	(B)	0.311	20.0	2.9	15.6	0.3			
Stationary	(B)	0.300	20.0	1.2	14.8	2.9			
Shaking	(A)	0.287	20.0	11.8	6.1	1.0			
Stationary	(A)	0.268	20.0	11.2	5.8	1.9			

(III). After 48 hours' incubation

C	Type of	Growth	B <sub>2</sub> present	(mg/50ml)	B2-glucoside	B2-oligo saccharides
Condition	seed medium #	0. D. at 530 mµ	initial	final	(mg/50 ml)	produced (mg/50 ml)
Shaking	(B)	0.390	20.0	0.5	13.7	4.6
Stationary	(B)	0.379	20.0	0	10.9	7.7
Shaking	(A)	0.388	20.0	+	15.1	3.8
Stationary	(A)	0.392	20.0	+	12.7	6.0

# (A): malt-extract medium

# (B): sucrose medium

favorable for the formation of riboflavinyl glucoside than the malt-extract medium (A), since with the medium (A) some lag periods were necessary before the formation of riboflavinylglucoside.

### 2. Effect of Cultural Condition

Fifty ml of 10 % sucrose medium with riboflavin mentioned above were taken in 200 ml shaking flask or in 100 ml Erlenmeyer flask. After inoculation, the flask was incubated at 30 °C for 22 hours under shaking or stationary conditions. Table 2 shows a comparison of yields of sugar compounds of riboflavin. No remarkable difference in the yield of riboflavinylglucoside was detected between shaking and stationary conditions, however the amount of riboflavin-compounds of oligosaccharides was found to be somewhat higher on the latter condition.

Condition	Shaking	Stationary
Type of seed medium	Sucrose	Sucrose
Growth (O. D. at 530 m $\mu$ )	0.324	0.312
B <sub>2</sub> present $(mg/50 ml)$		
ſ initial	20.0	20.0
l final	3.4	0.9
B2-glucoside produced (mg/50ml)	16.1	16.2
B2-oligosaccharides produced (mg/50 ml)	0.5	2.9

TABLE 2 Effect of cultural condition

## 3. Effect of Temperature

Leuc. mesenteroides, L. 20, was inoculated on the 10% sucrose medium with riboflavin and incubated for 2 days at various temperatures. As is shown in Table 3, higher yield of riboflavinylglucoside was attained at 25-30°C. At higher temperature as 37°C, both the riboflavin consumption and riboflavinylglucoside production were decreased.

Temperature (C°)	25°		30	0	37°	
Condition	Shal	king	Statio	nary	Static	mary
Type of seed medium*	( <del>B</del> )	(À)	( <b>B</b> )	(À)	( <b>B</b> )	(A)
Growth (O. D. at 530 m $\mu$ )	0.337	0.316	0.394	0.380	0.192	0.188
B <sub>2</sub> present (mg/50ml)						
∫ initial	25.0	25.0	25.0	25.0	25.0	25.0
l final	2.9	3.8	0.9	1.8	11.8	13.8
B2-glucoside produced (mg/50 ml)	14.7	16.0	13.9	15.5	10.4	9.1
B2-oligosaccharides produced (mg/50 ml)	5.5	2.6	9.4	5.2	0.9	+

TABLE 3 Effect of temperature

\* (A): malt-extract medium

(B): sucrose medium

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## 4. Effect of Concentration of Sucrose

Various amounts of sucrose were added to solutions containing 0.5% KH<sub>2</sub>PO<sub>4</sub>, 0.1% NaCl, 0.02% MgSO<sub>4</sub>·7H<sub>2</sub>O, 0.03% (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.05% polypeptone, 0.15% yeast extract, and 20—30 mg% riboflavin, as to get its final concentrations to 0.5%, 1%, 2%, 3%, 10% and 50%. All solutions were dispensed in 50 ml quantities into 100 ml Erlenmeyer flasks and sterilized. Two strains of *Leuc. mesenteroides*, L. 20 and IFO. 3426, were used and cultured at 23°C under stationary conditions. Almost the same yield of riboflavinylglucoside was obtained despite the concentrations of sucrose in the range of 2—10%, but the yield was decreasing at such low concentration of sucrose less than one percent, as will be seen in Table 4. And also a decrease in the yield was observed at such high level of sucrose as 50 percent.

Effect of	concentration	of	sucrose
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(I). After 7 hours' incubation

o	Concentration	Growth	B <sub>2</sub> present (mg/50 ml)		B <sub>2</sub> -glucoside	B2-oligo saccharides
Strain	of sucrose (%)	0. D. at 530 mµ	initial	final	(mg/50 ml) (mg/50 ml)	produced (mg/50 ml)
	( 0.5	0.188	10.0	8.8	0.7	0
IFO. 3426	2.0	0.184	10.0	8.1	1.3	(+)
	10.0	0.190	10.0	7.8	1.6	(+)
	( 0.5	0.178	10.0	8.4	0.9	0
L. 20	2.0	0.180	10.0	7.4	2.0	+
	10.0	0.179	10.0	7.2	2.1	+

## (II). After 14 hours' incubation

	Ci	Concentration	Growth	B <sub>2</sub> present	(mg/50ml)	B2-glucoside	B2-oligo saccharides
_	Stram	or sucrose (%)	530 mµ	initial	final	(mg/50 ml)	produced (mg/50ml)
		( 1.0	0.235	15.0	11.0	3.6	+
	IFO. 3426	3.0	0.257	15.0	8.7	5.9	+
		10.0	0.267	15.0	8.4	6.1	+
		50.0	0.135	15.0	13.0	1.7	—
		( 1.0	0.237	15.0	10.0	4.6	0.4
	T 00	3.0	0.274	15.0	8.1	6.2	0.7
	L. 20	10.0	0.241	15.0	7.2	7.0	0.8
		50.0	0.120	15.0	10.5	4.0	-

# 5. Effect of Kinds of Carbon Source

The media containing 0.03 % (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.15 % polypeptone, 0.15 % yeast extract, 30 mg % riboflavin, other mineral matters and various kinds of carbon sources described in Table 5 were inoculated with the malt-extract culture of *Leuc*.

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Carbon source (10%)	pH	Growth (O. D. at 530 mµ)	B2-glucoside produced (mg/50ml)
Glucose	4.4	0.387	(±)
Fructose	4.4	0.379	0
Galactose	5.8	0.222	0
Xylose	4.6	0.354	0
Glycerol	6.8	0.010	0
Sucrose	4.2	0.394	21.3
Lactose	5.0	0.173	(±)
Maltose	4.4	0.380	(土)
Soluble starch	6.8	0.010	0
Dextran	6.8	0.010	0
*Glucose + Fructose	4.4	0.377	(±)

TABLE 5 Effect of carbon source

\* mixture of 5% glucose + 5% fructose

mesenteroides, L. 20, and incubated at 30°C for 2 days under stationary conditions. Relations of the kinds of carbon source to the formation of riboflavinylglucoside in a growing culture were examined by the determination of bacterial growth, final pH and the amount of riboflavin-compounds as are shown in Table 5. It was found that sucrose was the only active source of carbon for the formation of riboflavinylglucoside, while D-glucose, D-fructose and their mixture or other carbon sources were ineffective.

6. Fluctuation of the Amounts of Riboflavin and of Its Sugar Compounds During the Fermentation

The fermentations were carried out at 30 °C with the same 10%sucrose medium after inoculation with 3 ml of a 24-hours' culture of *Leuc. mesenteroides*, IFO. 3426. grown on the seed media (B) and (A). Aliquots were removed at the requisite periods of fermentation and the amounts of riboflavin and of its sugar compounds were examined, and the results are illustrated in Figures 1 and 2. The amount of riboflavinylglucoside attained a ma-



Fig. 1. Fluctuation of riboflavin and its sugar compounds in a stationary culture.



Fig. 2. Fluctuation of riboflavin and its sugar compounds in a stationary culture.

ximum value in the earlier stage of incubation and then gradually decreased, while riboflavin-compounds of oligosaccharide were successively increasing during the decrease of riboflavinylglucoside. These fluctuations seemed to show that riboflavinylglucoside once produced was consumed gradually in order to form riboflavin-compounds of oligosaccharides by receiving glucosyl group again from sucrose; that is to say, riboflavinylglucoside played the role as the second acceptor in the successive glucosyl transfer, as was already demonstrated with maltose and ribo-

flavin by the action of enzymes of Asp. oryzae, E. coli and of Cl. acetobutyricum.

#### SUMMARY

Leuconostoc mesenteroides was found to produce a remarkable amount of riboflavinylglucoside when grown on sucrose medium containing riboflavin, but not on any other kinds of sugar media. The formation of riboflavinylglucoside was observed to depend largely on the kinds of seed culture and on the temperature of fermentation and also on the concentration of sucrose. Riboflavin-compounds of oligosaccharides were observed to be formed by successive transfers of D-glucosyl group of sucrose to riboflavin during the fermentation.

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#### REFERENCES

- Katagiri, H., Imai, K. and Yamada, H. 1954. Biosynthesis of riboflavinylglucoside by Escherichia coli. (in Japanese) Vitamin 7: 303.
- Katagiri, H. and Tachibana, S. 1953. On the mutation of Aspergillus oryzae. VI. The substance of transglucosidase. (in Japanese) Vitamin 6: 842.
- Katagiri, H., Yamada, H. and Imai, K. 1957. On the transglycosidation relating to riboflavin by *Escherichia coli*. 1. Formation of riboflavinylglucoside. J. Vitaminol. 3: 264–273.
- Suzuki, Y. and Katagiri, H., 1964. Studies on dextransucrase. 1. Formation of riboflavinylglucoside in dextran-producing cultures of *Leuconostoc mesenteroides*. J. Vitaminol. in press.

Tachibana, S. 1955. On the transglycosidation of Clostridium acetobutyricum. (in Japanese) Vita-

min 8: 363-365.

Tachibana, S. and Katagiri, H. 1955. On sugar compounds of riboflavin. 1. Biosynthesis of riboflavinylglucoside by *Aspergillus oryzae*. (in Japanese) Vitamin 8: 304-308.

Whitby, L. G. 1950. Enzymic formation of a new riboflavin derivative. Nature 166: 479-480.

Whitby, L. G. 1952. Riboflavinylglucoside: A new derivative of riboflavin. Biochem. J. 50: 433 ---438.

Whitby, L.G. 1954. Transglucosidation reaction with flavines. Biochem. J. 57: 390-396.