

1 Title

2 Natural genetic variation in fluctuating asymmetry of wing shape in *Drosophila melanogaster*

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17

18 **Abstract**

19 Fluctuating asymmetry (FA), defined as random deviation from perfect symmetry, has been used to
20 assay the inability of individuals to buffer their developmental processes from environmental
21 perturbations (i.e., developmental instability). In this study, we aimed to characterize the natural
22 genetic variation in FA of wing shape in *Drosophila melanogaster*, collected from across the
23 Japanese archipelago. We quantified wing shapes at whole wing and partial wing component levels
24 and evaluated their mean and FA. We also estimated the heritability of the mean and FA of these
25 traits. We found significant natural genetic variation in all the mean wing traits and in FA of one of
26 the partial wing components. Heritability estimates for mean wing shapes were significant in two
27 and four out of five wing traits in males and females, respectively. On the contrary, heritability
28 estimates for FA were low and not significant. This is a novel study of natural genetic variation in
29 FA of wing shape. Our findings suggest that partial wing components behave as distinct units of
30 selection for FA, and local adaptation of the mechanisms to stabilize developmental processes occur
31 in nature.

32

33 **Keywords**

34 Animal model · Geometric morphometrics · Heritability · Natural genetic variation · Wing shape

35

36 **Introduction**

37 Fluctuating asymmetry (FA), defined as random deviations from perfect symmetry, has been
38 observed in many organisms. Because corresponding body parts on the left and right sides of a
39 bilaterally symmetric organism presumably share the same genetic and physical environments, FA is
40 believed to reflect the inability of individuals to buffer their developmental processes from
41 environmental perturbations (i.e., developmental instability) (Whitlock 1996; Palmer and Strobeck
42 1997; Lens et al. 2002; Fuller and Houle 2003; Klingenberg 2003; Van dongen 2006). The ability to
43 stabilize developmental processes and produce morphological traits with high reproducibility, i.e.,
44 smaller FA, is expected to be adaptive under disruptive and fluctuating selection (Pelabon et al.
45 2010). FA has been reported to relate to a wide range of genetic and environmental stresses (Leary
46 and Allendorf 1989; Lens and Van Dongen 2000), and is a popular tool to estimate fitness of
47 organisms (Clarke 1998; Møller and Thornhill 1998), although some inconsistent FA-fitness
48 relationships have been pointed out (Fowler and Whitlock 1994; Vollestad et al. 1999; Bjorksten et
49 al. 2000). The genetic basis of FA has been studied in various organisms, such as plants, insects, and
50 mammals (Møller and Thornhill 1997; Leamy et al. 1998; Leamy and Klingenberg 2005);
51 *Drosophila* wings are the most intensively studied model system (Debat et al. 2009).

52 Although no significant additive genetic variation was estimated for FA of wing shape in
53 *D. melanogaster* (Woods et al. 1998), Carter et al. (2009) observed a significant increase in FA of

54 wing shape by inbreeding, suggesting the existence of genetic factors controlling FA. HSP90, a
55 molecular chaperone, was suggested to buffer developmental fluctuations in morphological traits in
56 diverse species such as *Drosophila*, *Arabidopsis* and zebrafish (Rutherford and Lindquist 1998;
57 Queitsch et al. 2002; Yeyati et al. 2007). However, in most experimental settings, the reduction of
58 HSP90 activity did not affect FA of wing shape in *D. melanogaster*. Debat et al. (2006) concluded
59 that Hsp90 is not the major regulator of FA. Takahashi et al. (2010) recently identified another heat
60 shock protein gene *Hsp67Ba* as having a significant effect on FA of wing shape. In addition, some
61 genomic regions of *D. melanogaster* showed potential to affect FA of wing shape (Breuker et al.
62 2006; Takahashi et al. 2011; Takahashi et al. in press). The genetic basis for FA has been so far
63 investigated using mutant analysis or RNAi approach targeting candidate genes, while little is known
64 about natural genetic variation in FA of wing shape.

65 Natural genetic variation in FA of wing shape has been investigated in a few studies but is
66 not confirmed (Woods et al. 1998; Debat et al. 2008). A possible reason for this is that the range of
67 sample collection was too limited to cover regions with different degrees of environmental stresses
68 where local adaptations of developmental stability have occurred. Wings of *Drosophila* can be
69 subdivided into several compartments that are subjected to different genetic controls and behave as
70 distinct units of selection (Cavicchi et al. 1985; Cavicchi et al. 1991; Garcia-Bellido et al. 1994;
71 Guerra et al. 1997; Pezzoli et al. 1997). These compartments may have independent molecular

72 mechanisms for stabilizing developmental processes. Comparison of wild *D. melanogaster* strains
73 from widespread geographical locations and measurement of relevant morphological traits of wings
74 may have a greater potential to uncover natural genetic variations in FA of wing shape.

75 In this study, we aimed to characterize the natural genetic variation in FA of wing shape in
76 *D. melanogaster*. We used 20 wild strains of *D. melnaogaster* collected from across the Japanese
77 archipelago (latitudes from 24°N to 43°N). We quantified wing shape traits at whole wing and
78 partial wing component levels, the stability of which may be regulated by different mechanisms, and
79 evaluated mean and FA of these traits for each strain. We also estimated the heritability of the mean
80 and FA of these traits for the wild strains. Significant genetic variation was found in all the mean
81 wing traits in both males and females. We found significant natural genetic variation only in FA of
82 "crossvein position," the relative position of the posterior crossvein. Heritability estimates for mean
83 shapes were significant in two and four out of five indices in males and females, respectively. On the
84 contrary, heritability estimates for FA were extremely low and not significant. Our findings suggest
85 that partial wing components behave as distinct units of selection for FA, and local adaptation of the
86 mechanisms to stabilize developmental processes occur in nature.

87

88

89 **Materials and Methods**

90 **Flies**

91 The flies used in this study were derived from 20 wild strains of *D. melanogaster* collected from
92 across the Japanese archipelago (latitudes from 24°N to 43°N; Table 1) and maintained in
93 EHIME-Fly, the laboratory for *Drosophila* resources at Ehime University. All the wild strains used
94 in this study were established as iso-females lines, and the generations maintained before our
95 experiments ranged from about 60 to 260, indicating that they were highly inbred strains (M. Watada,
96 personal communication). After we obtained the strains, they were kept under constant light at 23°C
97 in incubators in plastic vials (95 mm height, 24 mm diameter) containing 10 ml of fly medium
98 comprising dried yeast, soy flour, cornmeal, agar, malt extract, and dextrose.

99

100 **Among-strain genetic variation in wing shape**

101 **Experimental conditions**

102 To evaluate natural genetic variation in wing shape, we measured wing shape of each wild strain and
103 calculated among-strain variation. Because larval density is known to affect wing shape in *D.*
104 *melanogaster* (Bitner-Mathe and Klaczko 1999), we introduced 100 eggs into each vial to control
105 the density effect under constant light at 23°C with the same food medium described above. We set
106 up three replicate vials for each strain and collected emerging adults 10 days after eclosion, and took
107 photographs of wings as described below.

108 **Wing imaging**

109 To quantify wing shape using a landmark-based morphometric approach, we captured wing images
110 and obtained landmark coordinates. First, we anesthetized the flies and immobilized their one wing
111 between a slide glass and a cover slip using a simple suction device, wing grabber (Houle et al.
112 2003). The wing images were then captured with a digital camera, DP25 (Olympus Corporation,
113 Tokyo, Japan), attached to a microscope, SZ61TR (Olympus Corporation, Tokyo, Japan). Right
114 wing images were horizontally flipped to align the orientation of the right and left wing images. We
115 captured wing images of 15 individuals from each strain and each sex. The x and y coordinates for
116 18 landmarks on a wing (Fig. 1a) were obtained with an automated image-analysis system,
117 Wingmachine (Houle et al. 2003). In this system, a priori B-spline model was fitted to each of the
118 wing images using the pixel brightness of the reversed and filtered images (Lu and Milios 1994;
119 Houle et al. 2003). For the B-spline fitting, Wingmachine requires the x and y coordinates of the
120 basal two landmarks (landmark 9 and 14 in Fig. 1a). Because the acquisition of those landmarks
121 needs to be done manually, this process can be a major source of a measurement error. To evaluate
122 the measurement error, we repeated this landmark acquisition procedure twice. A Procrustes
123 ANOVA (Klingenberg and McIntyre 1998) was performed to assess the relative amount of
124 directional asymmetry (DA), FA and measurement error in wing shape variation. In this analysis, we
125 used individuals, sides, and their interaction term, and measurement error as independent variables,

126 and added sums of squares across all the landmarks coordinates, assuming equal and isotropic
127 variation at each landmark. In the current study, the B-spline model fitting on an image was
128 conducted twice and the average coordinates were used in subsequent analyses to minimize the
129 measurement error.

130 **Shape analysis**

131 Because the development of partial wing components of *D. melanogaster* are regulated by partially
132 independent molecular mechanisms (Trotta et al. 2005), the degree of natural genetic variation differ
133 among the partial components of a wing. To evaluate natural genetic variation at whole wing and
134 partial wing component levels separately, we quantified wing shape with all the landmarks and
135 subsets of the landmarks.

136 **Whole wing analysis**

137 In the wing shape analysis based on all the landmarks, we performed the Procrustes generalized least
138 squares procedure to eliminate the effect of translation, scaling, and rotation from the landmark
139 configurations, and to extract the non-allometric effect of the shape change in the dataset. In short,
140 the procedure can be described as follows (Klingenberg and McIntyre 1998). First, all the landmark
141 configurations were scaled to a unit size. Then, the centroids (or center of gravity) of the
142 configurations were superimposed. The configurations were then rotated to minimize the sum of
143 squared deviations of the landmarks of each of the configurations from the homologous landmarks

144 of the overall consensus (mean) configuration. The resulting Procrustes coordinates were used for
145 the whole wing shape analysis.

146 To evaluate the among-strain variation of each landmark, we performed principal
147 component analysis (PCA). This analysis extracts features of shape variation as a set of new shape
148 variables, the principal components (PCs), which are uncorrelated to one another and successively
149 account for maximal amounts of variation. Because a small subset of PCs may be sufficient to make
150 up most of the total variation, PCA is an effective method for data reduction, which is particularly
151 important for shape analysis because of the large number of variables (twice the number of
152 landmarks for 2D data; 36 variables in this study). The landmarks with strong correlation with
153 dominant PCs would be candidates of representative landmarks in wing shape variation. For the
154 purpose of visualization of the shape variation, we used the first and second PCs. Since the first PC
155 (PC1) explained most of the variation in the original landmark configurations (20.4% and 25.5% of
156 the variation in males and females, respectively), we used the PC1 score as a whole wing shape
157 index.

158 **Partial wing component analysis**

159 To quantify partial wing shape components, we used four wing shape indices using subsets of the
160 landmarks. The first index, "elongation index" (Debat et al. 2008), represents the ratio of wing
161 length to width (Fig. 1b) and was computed as follows,

162 $I_1 = \frac{d[2,13]}{d[1,14]},$

163 where, $d[a, b]$ is the linear distance between landmarks a and b. This trait has often been used in
 164 previous studies (Debat et al. 2008) because of its relative ease of measurement. The second index,
 165 "crossvein position" that represents the relative position of the posterior crossvein (Fig. 1c; Pelabon
 166 et al. 2006) was computed as follows,

167 $I_2 = \frac{d[11,6]/d[11,3]+d[10,5]/d[10,4]}{2}.$

168 The third and fourth indices represent the proportion of wing compartments relative to the
 169 whole wing area. The third trait, "anterior compartment size," represents the proportion of anterior
 170 compartment area (surrounded by the landmarks 1, 2, and 16; Fig. 1d) relative to the whole wing
 171 area (surrounded by the landmarks 1, 2, 3, 4, 9, and 14; Fig. 1e). The fourth trait, "posterior
 172 compartment size," represents the proportion of posterior compartment area (surrounded by the
 173 landmarks 3, 4, 5, and 6; Fig. 1d) relative to the whole wing area. The area surrounded by landmarks
 174 1, 2, 3, ...n was calculated:

175 $S = \frac{|(x_n-x_2)y_1+\sum_{k=2}^{n-1}(x_{k-1}-x_{k+1})y_k+(x_{n-1}-x_1)y_n|}{2},$

176 where, x_k and y_k are the x and y coordinates of landmark k . Third and fourth indices were expressed
 177 as follows:

178 $I_3 = S_{\text{anterior}}/S_{\text{total}}$ and $I_4 = S_{\text{posterior}}/S_{\text{total}}.$

179 **Fluctuating asymmetry**

180 Prior to the calculation of FA, we checked for the presence of DA, directional deviations from
181 bilateral symmetry (Klingenberg and Zaklan 2000), and antisymmetry (AS), the two sides are
182 always different but without a predictable direction to the differences. We performed
183 Kolmogorov-Smirnov tests to examine whether the distribution of the signed asymmetry (difference
184 between index values on the left and right wings) of each index deviated from normal distribution
185 with mean zero. As a result, we observed no significant deviation from normal distribution with
186 mean zero, indicating that the signed asymmetry could be treated as FA rather than a mixture of FA,
187 DA and AS. In both whole wing and partial wing component analyses, FA was evaluated as absolute
188 difference between index values on the left and right wings.

189 **Analysis of among-strain variation**

190 Diversification in the mean and FA of the wing shapes among strains was investigated using
191 one-way ANOVA with strain as a random effect. Although the strains were from wide latitudinal
192 range across Japanese archipelago, latitude was not considered as the source of variation. This is
193 because latitudinal cline in mean and FA of the wing shapes was not detected by regression analyses
194 (correlation coefficients ranged from -0.2 to 0.16 and not significant in all the cases). In the present
195 analysis, the following model was used:

$$196 \quad w_{ij} = \mu + \alpha_i + \epsilon_{ij},$$

197 where w_{ij} is the response variable (whole wing shape, elongation index, crossvein position, anterior

198 compartment size or posterior compartment size) of the j th replicate observations (individual) from
199 the i th strain, μ is the overall mean, α_i is an effect of the i th strain, and ε_{ij} is an unexplained error
200 associated with the j th replicate observation from the i th strain. A total of 10 analyses were
201 performed, two sexes and five indices, for mean trait or FA. To retain an experimentwise error rate
202 of $\alpha = 0.05$, a significance level for each test was determined by setting the comparison-wise error
203 rate at $\alpha' = 0.005$, based on the Bonferroni procedure.

204 **Correlation analysis among wing traits**

205 If the major source of shape variation at the whole wing level comes from a partial wing component,
206 a significant correlation between the whole wing and a partial wing trait may be detected. In addition,
207 shared regulatory mechanisms between partial wing components may cause correlation of their
208 variation. To examine these possibilities, pairwise correlations among five indices (whole wing
209 shape index and four partial wing component indices) were tested by using randomization procedure.
210 For each of the trait pairs, we randomized one of the trait vectors, and calculated a correlation
211 coefficient. We repeated the procedure for 1000 times and generated the null distribution of the
212 correlation coefficients. The observed correlation coefficient was judged as significant at $p = \alpha/500$
213 if it was smaller or larger than the bottom or top $\alpha\%$ of the null distribution. A total of 20 analyses
214 were performed, two sexes and 10 combinations of indices, for mean and FA of wing traits. To
215 retain an experimentwise error rate of $\alpha = 0.05$, a significance level for each test was determined by

216 setting the comparison-wise error rate at $\alpha' = 0.0025$, based on the Bonferroni procedure.

217

218 **Heritability experiment**

219 **Experimental conditions**

220 To estimate the heritability of the wing traits, we used mass bred populations initiated from 20 wild
221 strains *D. melanogaster* as described above. Two males and two females from each strain was used
222 to set up each mass bred population (initiated with 40 males and 40 females). Each mass bred
223 population was maintained in four 250 ml plastic bottles (with 50 ml of the food medium) containing
224 100-300 individuals to ensure total population size was more than 1000 individuals per generation to
225 maintain the original genetic variation. These populations were maintained for seven generations
226 prior to the heritability experiment under constant light at 23°C in incubators.

227 Three generations were assayed to obtain heritability estimates for wing traits.
228 Experimental flies for the parental generation were reared at a standard density (100 eggs per vial),
229 and emerging flies were anesthetized with CO₂ and collected as virgins. Wing shapes of 36 male
230 flies and 36 females were measured and then used to establish 36 pair matings (families). Each pair
231 was placed into a vial containing 10 ml of the food medium and allowed to lay eggs under constant
232 light at 23°C. The density of the eggs was checked to prevent overcrowding and the parental flies
233 were removed from the vials 24 hours after introduction. Parental pairs were allowed to lay eggs for

234 an extra 12 hours when egg density was too low. Emerging adults from each vial were collected as
235 virgins, and their wings were measured before the mating for the third generation. The mating pairs
236 were chosen from different families to avoid sib matings. Finally, the emerging grand-offspring
237 generation was collected and their wings were measured.

238 **Estimation of heritability with animal model**

239 Additive genetic variance for wing shape was estimated using a three-generation design. The animal
240 model method (Kruuk 2004) was adopted to estimate narrow-sense heritability (h^2) using all known
241 kin relationships among individuals. The animal model can divide the phenotypic variance into
242 additive genetic, environmental, and other fixed and random variances. We used a univariate animal
243 model of the form:

$$244 \quad y = Xa + e,$$

245 where, y is a vector of phenotypic values on all individuals, a is a vector of the additive genetic
246 effect, e is a vector of residual errors, and X is the corresponding design matrix (of 0s and 1s) that
247 relates the appropriate effects to y . The model was run under the Wombat program (ver. 1.0; Meyer
248 2007).

249 Heritability estimation for the mean and FA of the whole and partial wing components was
250 performed separately for males and females. Narrow-sense heritability was estimated as $h^2 = VA /$
251 $(VA + VE)$ (Houle 1992). In the analysis, significance of the heritability estimate was tested by using

252 the mean and the standard error. To retain an experimentwise error rate of $\alpha = 0.05$, a significance
253 level for each test was determined by setting a comparison-wise error rate at $\alpha' = 0.01$ based on the
254 Bonferroni procedure. The heritability estimate was considered significant if the approximate 99%
255 confidence interval (Wilson et al. 2010), the mean ± 2.58 SE, does not include zero.

256 **Genetic correlation**

257 To evaluate whether different wing traits share common morphogenic mechanisms, we
258 estimated genetic correlations between traits. The genetic correlation was estimated based on the
259 cross-variance obtained from the product of the trait A score in parents and the trait B score in
260 offspring, and the covariances of offspring and parents for each of the characters (Falconer and
261 Mackay 1996). In the current study, the cross-variance was calculated as the arithmetic mean of the
262 reciprocal cross-variances between traits. To test the significance of the observed genetic correlation,
263 we performed randomization test. We randomized one of the trait vectors, and calculated a genetic
264 correlation using the randomized dataset. We repeated the procedure for 1000 times and generated
265 the null distribution of the genetic correlation for each trait-pair. The observed genetic correlation
266 was judged as significant at $p = \alpha / 500$ if it was smaller or larger than the bottom or top α % of the
267 null distribution. A total of 20 analyses were performed, two sexes and 10 combinations of indices,
268 for mean and FA of wing traits. To retain an experimentwise error rate of $\alpha = 0.05$, a significance
269 level for each test was determined by setting the comparison-wise error rate at $\alpha' = 0.0025$, based on

270 the Bonferroni procedure.

271

272

273 **Results**

274 **Measurement error**

275 All the main factors in the Procrustes ANOVA were statistically significant (Table 2). This result
276 indicates that there were significant DA and FA in our dataset although significant DA was not
277 detected in the wing traits calculated based on the dataset as described above. The contribution of
278 measurement error to the overall shape variation was small in both sexes.

279

280 **Patterns of variation in landmarks**

281 PCA extracted features of wing shape variation, indicating that most variation was concentrated in a
282 few dimensions. In both males and females, the first five PCs accounted for 70% of the total
283 variance. Fig. 2 displays the features of variation associated with the first and second PCs, as plots of
284 the PC coefficients superimposed onto a drawing of the wing. PC1 was primarily affected by the
285 large variability of anterior crossvein position, associated with the movement of landmarks 7 and 8,
286 moved along the proximo-distal axis and also associated with the variation of landmark 1 in males
287 (Fig 2a). PC2 was primarily affected by the variability of the posterior crossvein position, associated

288 with the movement of landmarks 5 and 6, moved along the proximo-distal axis and also associated
289 with the movement of landmark 1 in males (Fig 2b). In females, PC1 was primarily affected by the
290 variability of anterior crossvein position (landmarks 7 and 8), moved along the proximo-distal axis
291 and also associated with the variation of landmarks 1 and 3 (Fig 2c). PC2 was primarily affected by
292 the proximo-distal movement of posterior crossvein position (landmarks 5 and 6) and also associated
293 with the proximo-distal movement of anterior crossvein (landmarks 7 and 8; Fig 2d).

294

295 **Patterns of variation in wing traits**

296 In both males and females, the mean of all the wing shape indices showed highly significant
297 diversification among strains (Table 3). Significant correlations were detected between the whole
298 wing shape and partial wing components: whole wing shape-crossvein position, and whole wing
299 shape-posterior compartment size in both sexes, and whole wing shape- anterior compartment size in
300 only males (Table 4). Several significant correlations were found among partial wing components:
301 crossvein position-anterior compartment size, crossvein position-posterior compartment size, and
302 anterior compartment size-posterior compartment size were all significantly correlated in both sexes,
303 and elongation index-posterior compartment size was significantly correlated in females alone
304 (Table 4).

305 In both males and females, no significant diversification in FA of the whole wing shape

306 was detected (Table 5). As for partial wing component FAs, only crossvein position in females
307 showed significant diversification among strains (Table 5). No significant correlation was found
308 between FAs of whole wing shape and partial wing components (Table 6). Several significant
309 correlations were found among FAs of partial wing components: elongation index-posterior
310 compartment size and crossvein position-anterior compartment size were correlated in males, and
311 crossvein position-posterior compartment size and anterior compartment size-posterior compartment
312 size were correlated in females (Table 6).

313

314 **Heritability of wing traits**

315 The heritability estimate for mean whole wing shape was significantly larger than zero in females
316 but not in males (Table 7). For the mean partial wing components, heritability estimates for
317 crossvein position and posterior compartment size in males and crossvein position, anterior
318 compartment size and posterior compartment size in females were significantly larger than zero
319 (Table 7). The significant heritability estimates for mean traits ranged from 0.426 to 0.827
320 depending on the trait and the sex (Table 7). The estimates of genetic correlation among the mean
321 wing traits were not significant in all the cases. In contrast, the heritability estimates for FA of the
322 wing shape traits were small, and not significantly different from zero for all indices in both males
323 and females (Table 8). The estimates of genetic correlation among the FA of wing traits were not

324 significant in all the cases.

325

326

327 **Discussion**

328 In this study, we investigated whether there was natural genetic variation in FA of wing shape in *D.*

329 *melanogaster*. All the means of wing traits showed highly significant diversification among wild

330 strains in both males and females, indicating large natural genetic variation in these traits. Although

331 the measures of wing morphology were somewhat different, previous studies also observed similar

332 natural genetic variation in the wing traits (Pezzoli et al. 1997; Woods et al. 1998; Debat et al. 2008).

333 A recent expression study reported that 164 of 1,335 genes changed their expression significantly

334 during wing morphogenesis and differentiation (Butler et al. 2003), suggesting that a large number

335 of genes are potentially involved in wing morphogenesis, and could be a source of natural genetic

336 variation. In contrast, significant diversification in FA among wild strains for FA was only detected

337 for crossvein position in females. This result, which is consistent with previous results (Woods et al.

338 1998; Debat et al. 2008), suggests that natural genetic variation in FA was limited to a partial wing

339 component, and could not be detected only by assessing the whole wing shape FA.

340 In the correlation analyses for mean traits, we found a couple of significant correlations

341 between the means of whole wing shape and partial wing components and also among partial wing

342 components. On the other hand, we found no significant correlation between FAs of the whole wing
343 shape and partial wing components, and a smaller number of significant correlations among partial
344 wing components than for mean traits. In contrast to the result from among-strain genetic variation
345 experiments, we could not find significant genetic correlation among the means and FAs of these
346 traits in the heritability experiment. These results indicate that some of the partial wing
347 components of *Drosophila* are subjected to at least partially different genetic control (Garcia-Bellido
348 et al. 1994; Guerra et al. 1997; Pezzoli et al. 1997). The results also suggest that the genetic
349 regulation of FA was more independent among partial wing components than of mean traits,
350 resulting in no significant correlation between the FAs of whole wing shape and partial wing
351 components. Although we found several significant correlations between partial wing components,
352 the correlations between crossvein position and posterior compartment size in mean and FA may be
353 an artifact due to shared landmarks on the posterior crossvein (landmarks 5 and 6). The significant
354 correlation between the anterior and posterior compartment sizes, found both in mean and FA in
355 females, suggests that they may share not only morphogenic but also developmental buffering
356 mechanisms. The significant correlation between elongation index and posterior compartment size,
357 found for FA in males, but not for mean traits, suggests that morphogenic and stabilizing factors
358 were independent in this case. These results emphasize that some partial wing components of
359 *Drosophila* wings are distinct units of natural selection, subjected to different genetic control. So far,

360 no gene has been found to affect wing shape FA in a wing compartment-specific manner. Genes that
361 show restricted expression patterns in multiple wing compartments are potential candidates for such
362 an effect. Future investigation of such genes may elucidate how *Drosophila* wings respond to natural
363 selection of developmental stability in nature.

364 In the current study, results of the heritability estimates did not always support the results
365 of the among-strain diversification (e.g., significant among-strain diversification detected in mean
366 whole wing shape, elongation index, and anterior compartment size in males, and elongation index
367 in females, but no significant heritability estimates for them). As for FA of the crossvein position,
368 we found significant among-strain diversification, but no significant heritability. These discrepancies
369 might come from the three-generation approach of the heritability estimation, which allows
370 recombination between homologous chromosomes from different strains. If multiple genes
371 contributed to the genetic diversification in these traits, recombination during the experimental
372 crosses might disrupt a set of coadapted alleles, and reduce the additive effect of these alleles below
373 the limit of detection. Based on a simulation model, Fuller and Houle (2002) suggest that artificial
374 selection for increased FA is the most powerful approach for the detection of genetic variation in
375 developmental instability. In the future, performing artificial selection on the partial wing
376 components may be necessary to estimate genetic variation in FA at higher resolution.

377 In this study, we found significant natural genetic variation in FA of a wing trait for the

378 first time. Our finding that only one component of the wing showed significant genetic variation in
379 FA suggests that partial wing components behave as distinct units of selection for FA in nature.
380 Further investigation on how FA of the wing trait is regulated, and subjected to natural selection may
381 facilitate understanding of the evolution of developmental stability.

382

383

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389

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391 **References**

392 Bitner-Mathe BC, Klaczko LB (1999) Plasticity of *Drosophila melanogaster* wing morphology:
393 Effects of sex, temperature and density. *Genetica* 105: 203-210
394 Bjorksten TA, Fowler K, Pomiankowski A (2000) What does sexual trait FA tell us about stress?
395 *Trends Ecol Evol* 15: 163-166

396 Breuker CJ, Patterson JS, Klingenberg CP (2006) A single basis for developmental buffering of
397 *Drosophila* wing shape. PloS One 1: e7

398 Butler MJ, Jacobsen TL, Cain DM, Jarman MG, Hubank M, Whittle JR, Phillips R, Simcox A (2003)
399 Discovery of genes with highly restricted expression patterns in the *Drosophila* wing disc using
400 DNA oligonucleotide microarrays. Development 130: 659-670

401 Carter AJ, Weier TM, Houle D (2009) The effect of inbreeding on fluctuating asymmetry of wing
402 veins in two laboratory strains of *Drosophila melanogaster*. Heredity 102: 563-572

403 Cavicchi S, Giorgi G, Natali V, Guerra D (1991) Temperature-related divergence in experimental
404 populations of *Drosophila melanogaster*. III. Fourier and centroid analysis of wing shape and
405 relationship between shape variation and fitness. J Evol Biol 4: 141-159

406 Cavicchi S, Guerra D, Giorgi G, Pezzoli C (1985) Temperature-related divergence in experimental
407 populations of *Drosophila melanogaster*. I. Genetic and developmental basis of wing size and shape
408 variation. Genetics 109: 665-689

409 Clarke GM (1998) The genetic basis of developmental stability. V. Inter- and intra-individual
410 character variation. Heredity 80: 562-567

411 Debat V, Cornette R, Korol AB, Nevo E, Soulet D, David JR (2008) Multidimensional analysis of
412 *Drosophila* wing variation in Evolution Canyon. J Genet 87: 407-419

413 Debat V, Debelle A, Dworkin I (2009) Plasticity, canalization, and developmental stability of the

414 *Drosophila* wing: joint effects of mutations and developmental temperature. *Evolution* 63:
415 2864-2876

416 Debat V, Milton CC, Rutherford S, Klingenberg CP, Hoffmann AA (2006) HSP90 and the
417 quantitative variation of wing shape in *Drosophila melanogaster*. *Evolution* 60: 2529-2538

418 Falconer DS, Mackay TFC (1996) Introduction to quantitative genetics, 4th edn. Pearson, Harlow.

419 Fowler K, Whitlock MC (1994) Fluctuating asymmetry does not increase with moderate inbreeding
420 in *Drosophila melanogaster*. *Heredity* 73: 373-376

421 Fuller RC, Houle D (2002) Detecting genetic variation in developmental instability by artificial
422 selection on fluctuating asymmetry. *J Evol Biol* 15: 954-960

423 Fuller RC, Houle D (2003) Inheritance of developmental instability. In: Polak M (ed) *Developmental*
424 *instability: causes and consequences*. Oxford university press, New York, pp 157-183

425 Garcia-Bellido A, Cortes F, Milan M (1994) Cell interaction in the control of size in *Drosophila*
426 wings. *Proc Natl Acad Sci USA* 91: 10222-10226

427 Guerra D, Pezzoli MC, Giorgi G, Garoia F, Cavicchi S (1997) Developmental constraints in the
428 *Drosophila* wing. *Heredity* 79: 564-571

429 Houle D (1992) Comparing evolvability and variability of quantitative traits. *Genetics* 130: 195-204

430 Houle D, Mezey J, Galpern P, Carter A (2003) Automated measurement of *Drosophila* wings. *Bmc*
431 *Evol Biol* 3: 12

432 Klingenberg CP, McIntyre GS (1998) Geometric morphometrics of developmental instability:
433 Analyzing patterns of fluctuating asymmetry with Procrustes methods. *Evolution* 52: 1363-1375

434 Klingenberg CP, Zaklan SD (2000) Morphological integration between developmental compartments
435 in the *Drosophila* wing. *Evolution* 54: 1273-1285

436 Klingenberg CP (2003) A developmental perspective on phenotypic variation, canalization, and
437 fluctuating asymmetry. In: Polak M (ed) *Developmental instability: causes and consequences*.
438 Oxford university press, New York, pp 14-34

439 Kruuk LEB (2004) Estimating genetic parameters in natural populations using the 'animal model'.
440 *Phil Trans R Soc B - Biol Sci* 359: 873-890

441 Leamy LJ, Klingenberg CP (2005) The genetics and evolution of fluctuating asymmetry. *Annu Rev*
442 *Ecol Evol Syst* 36: 1-21

443 Leamy LJ, Routman EJ, Cheverud JM (1998) Quantitative trait loci for fluctuating asymmetry of
444 discrete skeletal characters in mice. *Heredity* 80: 509-518

445 Leary RF, Allendorf FW (1989) Fluctuating asymmetry as an indicator of stress: Implications for
446 conservation biology. *Trends Ecol Evol* 4: 214-217

447 Lens L, Van Dongen S (2000) Fluctuating and directional asymmetry in natural bird populations
448 exposed to different levels of habitat disturbance, as revealed by mixture analysis. *Ecol Lett* 3:
449 516-522

450 Lens L, Van Dongen S, Kark S, Matthysen E (2002) Fluctuating asymmetry as an indicator of
451 fitness: can we bridge the gap between studies? *Biol Rev Camb Philos Soc* 77: 27-38

452 Lu F, Milios EE (1994) Optimal spline fitting to planar shape. *Signal Processing* 37: 129-140

453 Meyer K (2007) Wombat: tool for mixed model analyses in quantitative genetics by restricted
454 maximum likelihood (reml). *J Zhejiang Univ - Sci B* 8: 815-821

455 Møller AP, Thornhill R (1997) A meta-analysis of the heritability of developmental stability. *J Evol*
456 *Biol* 10: 1-16

457 Møller AP, Thornhill R (1998) Bilateral symmetry and sexual selection: A meta-analysis. *Am Nat*
458 151: 174-192

459 Palmer RA, Strobeck C (1997) Fluctuating asymmetry and developmental stability: heritability of
460 observable variation vs. heritability of inferred cause. *J Evol Biol* 10: 39-49

461 Pelabon C, Hansen TF, Carter AJR, Houle D (2006) Response of fluctuating and directional
462 asymmetry to selection on wing shape in *Drosophila melanogaster*. *J Evol Biol* 19: 764-776

463 Pelabon C, Hansen TF, Carter AJR, Houle D (2010) Evolution of variation and variability under
464 fluctuating, stabilizing, and disruptive selection. *Evolution* 64: 1912-1925

465 Pezzoli MC, Guerra D, Giorgi G, Garoia F, Cavicchi S (1997) Developmental constraints and wing
466 shape variation in natural populations of *Drosophila melanogaster*. *Heredity* 79: 572-577

467 Queitsch C, Sangster TA, Lindquist S (2002) Hsp90 as a capacitor of phenotypic variation. *Nature*

468 417: 618-624

469 Rutherford S, Lindquist S (1998) Hsp90 as a capacitor for morphological evolution. *Nature* 396:
470 336-342

471 Takahashi KH, Okada Y, Teramura K (2011) Genome-wide deficiency mapping of the regions
472 responsible for temporal canalization of the developmental processes of *Drosophila melanogaster*. *J*
473 *Hered* 102:448-457.

474 Takahashi KH, Okada Y, Teramura K, Tsujino M (in press) Deficiency mapping of the genomic
475 regions associated with effects on developmental stability in *Drosophila melanogaster*. *Evolution*

476 Takahashi KH, Rako L, Takano-Shimizu T, Hoffmann AA, Lee SF (2010) Effects of small Hsp
477 genes on developmental stability and microenvironmental canalization. *BMC Evol Biol* 10: 284

478 Trotta V, Garoia F, Guerra D, Pezzoli MC, Grifoni D, Cavicchi S (2005) Developmental instability
479 of the *Drosophila* wing as an index of genomic perturbation and altered cell proliferation. *Evol Dev*
480 7: 234-243

481 Van dongen S (2006) Fluctuating asymmetry and developmental instability in evolutionary biology:
482 past, present and future. *J Evol Biol* 19: 1727-1743

483 Vollestad LA, Hindar K, Møller AP (1999) A meta-analysis of fluctuating asymmetry in relation to
484 heterozygosity. *Heredity* 83: 206-218

485 Wilson AJ, Reale D, Clements MN, Morrissey MM, Postma E, Walling CA, Kruuk LEB, Nussey

486 DH (2010) An ecologist's guide to the animal model. *J Anim Ecol* 79: 13-26

487 Whitlock M (1996) The heritability of fluctuating asymmetry and the genetic control of
488 developmental stability. *Proc R Soc B* 263: 849-854

489 Woods RE, Hercus MJ, Hoffmann AA (1998) Estimating the heritability of fluctuating asymmetry in
490 field *Drosophila*. *Evolution* 52: 816-824

491 Yeyati PL, Bancewicz RM, Maule J, van Heyningen V (2007) Hsp90 selectively modulates
492 phenotype in vertebrate development. *PLoS Genet* 3: e43

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495 **Figure legends**

496 Fig. 1 Landmark positions and wing indices, a Eighteen landmarks on the wing vein junctions, the
497 wing margin, and on the free ends of wing veins, b elongation index, c crossvein position, d anterior
498 and posterior compartment, and e whole wing area

499

500 Fig. 2 Principal component analysis (PCA) of variation in landmark positions for individual
501 variability. The diagrams visualize the PC coefficients of each landmark in x and y directions by a
502 line originating at the average location of the landmark (circles). a PC1 in males, b PC2 in males, c
503 PC1 in females, d PC2 in females

504

505