

◎原 著

Release of histamine and leukotriene C₄ from bronchoalveolar cells in patients with bronchial asthma.

A role of histamine in atopic asthma.

Fumihiro Mitsunobu, Takashi Mifune, Kazuhiro Kajimoto, Yasuhiro Hosaki, Satoshi Yokota, Yoshiro Tanizaki, Koji Ochi¹⁾, Hideo Harada¹⁾

Division of Medicine, Misasa Medical Branch, ¹⁾Department of Laboratory Medicine, Okayama University Medical School

Abstract : To clarify the main humoral triggering factor (histamine and/or leukotriene) of the early stage of asthma attacks, the release of histamine and leukotriene C₄ (LTC₄) from bronchoalveolar lavage (BAL) cells stimulated with Ca ionophore A23187 was examined in 7 patients with atopic asthma, and the results were compared to those in 7 nonatopic asthma patients. 1. The proportion of BAL basophilic cells was significantly higher in atopic patients than in nonatopic patients ($p < 0.05$). 2. The content of histamine in BAL fluid was significantly higher in atopic (2.3mcg/ml) compared to that in nonatopic patients (0 mcg/ml) ($p < 0.001$). The content of LTC₄ was high in nonatopic (2.4ng/ml) than in atopic patients (0.5ng/ml), however, this was not significant. 3. The release of histamine from BAL cells was 32.6% in atopic and 0% in nonatopic patients, and this was significant ($p < 0.001$). The release of LTC₄ from BAL cells was significantly higher in nonatopic (11.3ng/10⁶cell) than in atopic (3.5ng/10⁶cell) ($p < 0.02$).

The results demonstrate that histamine play more important role in atopic patients as a main triggering factor of attacks than LTC₄. In contrast, in nonatopic patients, LTC₄ is more predominant than histamine during early stage of asthma attacks.

Key words : Histamine, LTC₄, BAL cells, atopic asthma

Introduction

In the onset mechanism of asthma, humoral factors such as histamine and

leukotrienes in the early stage of asthma attacks¹⁻⁴⁾, and cellular components such as lymphocytes, neutrophils, eosinophils, and basophils in the late stage have been shown

to play important roles⁵⁻¹³. Particularly, release of histamine and leukotrienes in the early stage are important as the early triggering factor of attacks, since pathophysiological changes in the airways of asthma, such as bronchoconstriction, mucus hypersecretion and edema of mucous membrane, are at first introduced by the release of these chemical mediators from tissue mast cells¹⁴.

Asthma is classified into two types, atopic and nonatopic, based on the presence or absence of an IgE-mediated allergic reaction¹⁵. Atopic asthma is often observed in young patients, having specific IgE antibodies to allergens. In contrast, nonatopic asthma is often seen in late onset adult patients, and shows low serum IgE levels and negative IgE antibodies to allergens. This suggests that the main triggering factor of asthma attacks are different between atopic and nonatopic asthma patients.

In the present study, to clarify the main triggering factor of attacks, release of histamine and leukotrienes from inflammatory cells in the airways was examined in atopic asthma patients, and the results were compared to those in nonatopic asthma patients.

Subjects and Methods

Seven patients with atopic asthma were selected in this study to examine the main humoral triggering factor (histamine and/or leukotrienes) of IgE-mediated allergic reaction of atopic asthma. They were all sensitive to house dust mite, showing a positive RAST score (2+ or more) to the allergen. Seven nonatopic asthma patients were selected as control subjects. Their serum IgE levels were under 200 IU/ml and RAST score to various allergens was negative.

Bronchoalveolar lavage (BAL) was per-

formed according to a previously reported method¹²⁻¹⁴ when the subjects were attack-free. Informed consent for this BAL procedure was obtained from all study subjects. The aspirate obtained by BAL were filtered through a sterile steel mesh, and the filtrates were centrifuged at 300 *g* for 10 min at 4 °C. The cell pellet was resuspended in Tris ACM. After the number of cells was adjusted to 10⁶ cells/ml in Tris ACM, Ca ionophore A23187 (1 mcg) was added to the cell suspension. The mixed solution was then incubated for 15 min at 37°C and centrifuged at 300 *g* for 10 min at 4 °C. The histamine content of both the cells and supernatant fluid were analyzed by perchloric acid precipitation and assayed with automated spectrofluorometric histamine analysis system (Technicon Instruments Co). Histamine release was expressed as a percentage of total histamine. The HPLC analysis for extraction and quantification of LTC₄ was performed by a method described by Lam et al¹⁶. The extraction of leukotrienes was performed by a method using a C18 Sep-Pak (Walters Associates). The concentration of LTC₄ was analyzed by an HPLC system, Model 510 (Walters Associate), equipped with an ultraviolet detector. The column used was a 5 mm × 10 cm Radial-Pak cartridge (Shimazu). The results were expressed as ng/10⁶ cells. BAL cytology was performed by observing 500 cells, excluding epithelial cells, on smear preparations which were made from BAL cell suspensions and stained with May-Giemsa. Regarding mast cells and basophils in BAL fluid, 1000 cells were observed and the number of basophilic cells was calculated. The results were expressed as percentages of the total number of cells. In this study, the mean recovery rate at BAL was 28.8 ± 11.8% (± SD). The total

number of cells aspirated into BAL fluid was $8.21 \pm 10.2 \times 10^6$.

The level of serum IgE was measured by the radioimmunosorbent test (RIST), and IgE antibodies against allergens were assessed by the radioallergosorbent test (RAST).

Statistically significant difference of the mean was evaluated using the unpaired Student's t test. A value of $p < 0.05$ was regarded as significant.

Results

Table 1 shows characteristics of atopic and nonatopic asthma patients. Serum IgE level was remarkably higher in atopic asthma than in nonatopic asthma. However, this difference was not significant. Cellular composition of BAL fluid in both atopic and nonatopic subjects was shown in Table 2. The proportions of BAL lymphocytes, neutrophils, and eosinophils were not significantly different between atopic and nonatopic asthma patients. The proportion of BAL basophilic cells was significantly higher in atopic patients than in nonatopic patients ($p < 0.05$).

Table 1. Characteristics of patients with bronchial asthma studied

Asthma type	No of patients	Mean age (years)	RAST 2+< to HD	serum IgE (IU/ml)
Atopic	7	48.0 (36-62)	7	1202 (170-4134)
Nonatopic	7	52.0 (22-63)	0	91 (18-174)

HD; house dust

Table 2. Cellular composition of BAL fluid in patients with asthma studied

Asthma type	No of patients	BAL cells (%)				
		Mac	Lym	Neut	Eos	Bas
Atopic	7	83.5 \pm 11.2	11.6 \pm 8.3	2.4 \pm 2.1	2.3 \pm 1.9	0.15 \pm 0.13 ^a
Nonatopic	7	77.4 \pm 7.9	18.9 \pm 7.9	2.9 \pm 2.6	0.9 \pm 1.0	0.03 \pm 0.05 ^a

Mac; macrophages, Lym; lymphocytes, Neut; neutrophils, Eos; eosinophils, Bas; basophilic cells. ^a $p < 0.05$.

The content of histamine in BAL fluid was significantly higher in atopic patients (mean \pm SD : 2.3 ± 1.5 mcg/ml) than in nonatopic patients (mean 0 mcg/ml)($p < 0.001$)(Fig. 1). In contrast, the content of leukotrienes C 4 (LTC4) was higher in nonatopic patients (2.4 ± 2.6 ng/ml) compared to that in atopic patients (0.5 ± 1.3 ng/ml). However, this was not significant (Fig. 2).

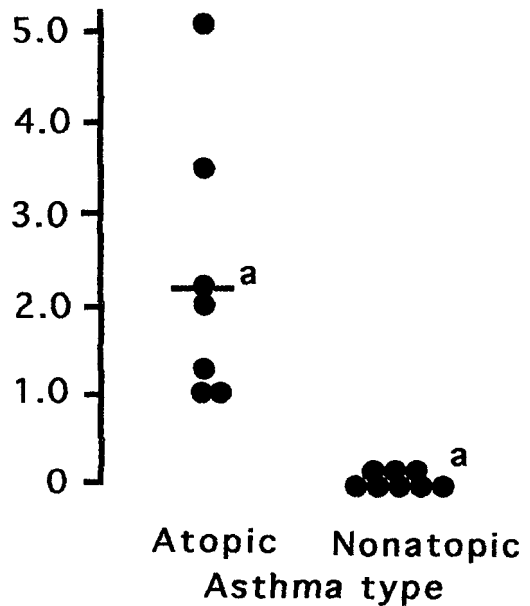


Fig. 1. Histamine content in BAL fluid in patients with asthma. ^a $p < 0.001$.

Discussion

Asthma can be clinically classified into two types, atopic and nonatopic by the onset mechanism of attacks. Although there are some reports suggesting that asthma is almost always associated with some type of IgE-related reaction^{17, 18)}, there are many asthma patients who are clinically regarded as nonatopic, showing negative skin test, low serum IgE levels, and negative RAST score to various allergens. Nonatopic asthma clinically diagnosed is often observed in late onset (over the age of 40 years) adult patients. Their attacks often become severe and intractable^{19, 20)} within a couple of years from onset of asthma. The onset mechanism of attacks, particularly, triggering event in early stage of attacks in nonatopic asthma is still unclear. In contrast, in atopic asthma, IgE-mediated allergic reaction was triggered by bridging of IgE receptors on mast cell membrane, followed by release of chemical mediators such as histamine and leukotrienes^{1-4, 21-23)}.

In recent years, focus has been placed on inflammatory cell infiltration in the airways as a main onset mechanism of asthma⁵⁻¹⁴⁾. However, release of chemical mediators such as histamine and leukotrienes has been considered to be the main mechanism of the onset of early stage of asthma attacks. The mechanism is important as triggering factor of asthma attacks.

In the present study, to clarify whether a main triggering factor in the early stage of attacks is different between atopic and nonatopic asthma, release of histamine and leukotrienes from BAL cells was examined in these two different asthma types. The results revealed that the release of histamine from

BAL cells was significantly higher in atopic than in nonatopic asthma. In contrast, the release of LTC₄ from BAL cells was significantly higher in nonatopic asthma compared to that in atopic asthma. These results suggest that histamine is more important as triggering factor of attacks in atopic asthma than LTC₄, and that LTC₄ is more important than histamine in nonatopic asthma in relation to early stage of attacks.

References

1. Ishizaka T, Ishizaka K, Conrad DH, Froese A : A new concept of mechanism of IgE-mediated histamine release. *J Allergy Clin Immunol* 64 : 320-330, 1978.
2. Tanizaki Y, Komagoe H, Sudo M, Morinaga H, Kitani H, Goda Y, Tada S, Takahashi K, Kimura I : IgE-mediated histamine release from whole blood in atopic asthmatics. *Jpn J Allergol* 32 : 1079-1083, 1983.
3. Tanizaki Y, Komagoe H, Morinaga H, Kitani H, Goda Y, Kimura I : Allergen- and anti-IgE-induced histamine release from whole blood. *Int Arch Allergy Appl Immunol* 73 : 141-145, 1984.
4. Tanizaki Y, Sudo M, Kitani H, Kawauchi K, Mifune T, Takahashi K, Kimura I : Release of heparin-like substance and histamine from basophilic leucocytes separated by counterflow centrifugation elutriation. *Jpn J Med* 29 : 356-361, 1990.
5. Nadel JA : Inflammation and asthma. *J Allergy Clin Immunol* 73 : 651-653, 1984.
6. Wardlaw AJ and Kay AB : The role of the eosinophil in the pathogenesis of asthma. *Allergy* 42 : 321-335, 1987.
7. Kirby JG, Hargreave FE, Gleich GJ, O'Byrne DM : Bronchoalveolar cell profiles of asthmatics and nonasthmatic subjects.

- Am Rev Respir Dis 136 : 379–383, 1987.
8. Lozewicz S, Gomez E, Ferguson H, Davies RJ : Inflammatory cells in the airways in mild asthma. *Br Med J* 297 : 1515–1516, 1988.
 9. Kelly CA, Ward D, Stenton CS, Bird G, Hendrick DJ, Walters CM : Numbers and activity of inflammatory cell in bronchoalveolar lavage in asthma, and their relationship to airway responsiveness. *Thorax* 43 : 684–692, 1988.
 10. Kelly CA, Stenton SC, Ward C, Bird G, Hendrick DJ, Walters EH : Lymphocyte subsets in bronchoalveolar lavage fluid from stable asthmatics, and their correlations with bronchial responsiveness. *Clin Exp Allergy* 19 : 169–175, 1989.
 11. Holgate ST, Djukanovic R, Wilson J, Roche WR, Howarth PH : Inflammatory process and bronchial hyperresponsiveness. *Clin Exp Allergy* 21 : 30–36, 1991.
 12. Tanizaki Y, Kitani H, Okazaki M, Mifune T, Mitsunobu F, Ochi K, Harada H : Cellular composition of fluid in the airways of patients with house dust sensitive asthma, classified by clinical symptoms. *Int Med* 31 : 333–338, 1992.
 13. Tanizaki Y, Kitani H, Okazaki M, Mifune T, Mitsunobu F, Okano T, Honke N, Kimura I : A new modified classification of bronchial asthma based on clinical symptoms. *Int Med* 32 : 197–203, 1993.
 14. Tanizaki Y, Kitani H, Okazaki M, Mifune T, Mitsunobu F, Kimura I : Changes in the proportions of bronchoalveolar lymphocytes, neutrophils and basophilic cells and the release of histamine and leukotrienes from bronchoalveolar cells in patients with steroid-dependent intractable asthma. *Int Arch Allergy Immunol* 101 : 196–202, 1993.
 15. Ostergaard : Non-IgE mediated asthma in children. *Acta Paediatr Scand* 74 : 713–716, 1985.
 16. Lam S, Chan H, LeRiche JC, Chan-Yeung M, Salari H : Release of leukotrienes in patients with bronchial asthma. *J Allergy Clin Immunol* 81 : 711–717, 1988.
 17. Burrows B, Martinez FD, Halonen M, Barbee RA, Cline MG : Association of asthma with serum IgE levels and skin-test reactivity to allergens. *N Engl J Med* 320 : 271–277, 1989.
 18. Tollerud DJ, O'Connor GT, Sparrow D, Weiss ST : Asthma, hay fever, and phlegm production associated with distinct patterns of allergy skin test reactivity, eosinophilia, and serum IgE levels. The normative aging study. *Am Rev Respir Dis* 144 : 776–781, 1991.
 19. Tanizaki Y, Kitani H, Okazaki M, Mifune T, Mitsunobu F, Kimura I : Effects of long-term glucocorticoid therapy on bronchoalveolar cells in adult patients with bronchial asthma. *J Asthma* 30 : 309–318, 1993.
 20. Tanizaki Y, Kitani H, Mifune T, Mitsunobu F, Kajimoto K, Sugimoto K : Effects of glucocorticoids on humoral and cellular immunity and on airway inflammation in patients with steroid-dependent intractable asthma. *J Asthma* 30 : 485–492, 1993.
 21. Wardlaw AJ, Hay H, Cromwell O, Collins JV, Kay AB : Leukotrienes, LTC₄ and LTB₄, in bronchoalveolar lavage in bronchial asthma and other respiratory diseases. *J Allergy Clin Immunol* 84 : 19–27, 1989.
 22. Chan-Yeung M, Chan H, Tse KS, Salari H, Lam B : Histamine and leukotrienes release in bronchoalveolar fluid during

- plicatic acid-induced bronchoconstriction. *J Allergy Clin Immunol* 84 : 762-771, 1989.
23. Tanizaki Y, Kitani H, Okazaki M, Mifune T, Mitsunobu F, Kimura I: Association of asthma with serum IgE levels

and aging. *Bronchoalveolar cells and the release of histamine and leukotrienes, LTC4 and LTB4, from leucocytes.* *Jpn J Clin Immun* 16 : 44-51, 1993.

気管支喘息患者の気管支肺胞細胞からのヒスタミンとロイコトリエンC4遊離-アトピー性喘息におけるヒスタミンの役割について-

光延文裕, 御船尚志, 梶本和宏, 横田 聡,
保崎泰弘, 谷崎勝朗, 越智浩二¹⁾, 原田英雄¹⁾

岡山大学医学部附属病院三朝分院内科,
岡山大学医学部附属病院臨床検査医学¹⁾

気管支喘息発作初期に関与する液性因子(ヒスタミン, ロイコトリエン)の役割を明らかにするために, 気管支肺胞洗浄(BAL)細胞からのカルシウムイオノフォアA23187によるヒスタミンとロイコトリエンC4(LTC4)遊離の検討を行った。対象はアトピー性喘息患者7名, 非アトピー性喘息患者7名とした。1. BAL液中の好塩基性細胞の比率は, アトピー性喘息患者において有

意に高値を示した。2. BAL液中ヒスタミン濃度は非アトピー性喘息患者(0 mcg/ml)に比して, アトピー性喘息患者(2.3mcg/ml)において有意に高値を示した。一方, BAL液中LTC4濃度は, アトピー性喘息患者(0.5ng/ml)に比して非アトピー性喘息患者(2.4ng/ml)において, 高値を示したが, 有意差は見られなかった。3. BAL細胞からのヒスタミン遊離は非アトピー性喘息患者(0%)に比して, アトピー性喘息患者(32.6%)において有意に高値を示した。LTC4遊離はアトピー性喘息患者(3.5ng/10⁶細胞)に比して, 非アトピー性喘息患者(11.3ng/10⁶細胞)において有意に高値を示した。

以上より, 喘息発作初期に関与する液性因子としては, アトピー性喘息患者においてはヒスタミンが重要であり, 非アトピー性喘息患者においてはLTC4が, 重要であることが, 明らかになった。