PLOS | ONE

Genetic Variation and Reproductive Timing: African American Women from the Population Architecture Using Genomics and Epidemiology (PAGE) Study

Kylee L. Spencer[1,2,9], Jennifer Malinowski[1,9], Cara L. Carty[5], Nora Franceschini[4], Lindsay Fernández-Rhodes[4], Alicia Young[5], Iona Cheng[7], Marylyn D. Ritchie[1,13], Christopher A. Haiman[6], Lynne Wilkens[7], Chunyuan Wu[5], Tara C. Matise[2], Christopher S. Carlson[5], Kathleen Brennan[8], Amy Park[9], Aleksandar Rajkovic[10], Lucia A. Hindorff[11], Steven Buyske[2,3,9], Dana C. Crawford[1*,9]

1 Center for Human Genetics Research, Department of Molecular Physiology and Biophysics, Vanderbilt University, Nashville, Tennessee, United States of America, 2 Department of Genetics, Rutgers University, Piscataway, New Jersey, United States of America, 3 Department of Statistics, Rutgers University, Piscataway, New Jersey, United States of America, 4 Department of Epidemiology, University of North Carolina, Chapel Hill, North Carolina, United States of America, 5 Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington, United States of America, 6 Keck School of Medicine, University of Southern California, Los Angeles, California, United States of America, 7 University of Hawaii Cancer Center, University of Hawaii, Honolulu, Hawaii, United States of America, 8 Department of Obstetrics and Gynecology, University of California Los Angeles, Los Angeles, California, United States of America, 9 Department of Obstetrics and Gynecology, School of Medicine, Georgetown University, Washington, DC, United States of America, 10 Magee-Womens Research Institute, Department of Obstetrics, Gynecology and Reproductive Sciences, University of Pittsburgh, Pittsburgh, Pennsylvania, United States of America, 11 Office of Population Genomics, National Human Genome Research Institute, National Institutes of Health, Bethesda, Maryland, United States of America, 12 Department of Biology and Environmental Science, Heidelberg University, Tiffin, Ohio, United States of America, 13 Biochemistry and Molecular Biology, Penn State University, University Park, Maryland, United States of America

Abstract

Age at menarche (AM) and age at natural menopause (ANM) define the boundaries of the reproductive lifespan in women. Their timing is associated with various diseases, including cancer and cardiovascular disease. Genome-wide association studies have identified several genetic variants associated with either AM or ANM in populations of largely European or Asian descent women. The extent to which these associations generalize to diverse populations remains unknown. Therefore, we sought to replicate previously reported AM and ANM findings and to identify novel AM and ANM variants using the Metabochip (n = 161,098 SNPs) in 4,159 and 1,860 African American women, respectively, in the Women's Health Initiative (WHI) and Atherosclerosis Risk in Communities (ARIC) studies, as part of the Population Architecture using Genomics and Epidemiology (PAGE) Study. We replicated or generalized one previously identified variant for AM, rs1361108/*CENPW*, and two variants for ANM, rs897798/*BRSK1* and rs769450/*APOE*, to our African American cohort. Overall, generalization of the majority of previously-identified variants for AM and ANM, including *LIN28B* and *MCM8*, was not observed in this African American sample. We identified three novel loci associated with ANM that reached significance after multiple testing correction (*LDLR* rs189596789, $p = 5 \times 10^{-08}$; *KCNQ1* rs79972789, $p = 1.9 \times 10^{-07}$; *COL4A3BP* rs181686584, $p = 2.9 \times 10^{-07}$). Our most significant AM association was upstream of *RSF1*, a gene implicated in ovarian and breast cancers (rs11604207, $p = 1.6 \times 10^{-06}$). While most associations were identified in either AM or ANM, we did identify genes suggestively associated with both: *PHACTR1* and *ARHGAP42*. The lack of generalization coupled with the potentially novel associations identified here emphasize the need for additional genetic discovery efforts for AM and ANM in diverse populations.

Competing Interests: Co-author Dana C. Crawford is a PLOS ONE Editorial Board member. This membership does not alter the authors' adherence to all the PLOS ONE policies on sharing data and materials.

* E-mail: dana.c.crawford@vanderbilt.edu

⑨ These authors contributed equally to this work.

Introduction

Age at menarche (AM) and age at natural menopause (ANM) are components of the reproductive lifespan in women. Timing of these reproductive milestones is associated with various diseases and cancers such as type 2 diabetes, cardiovascular disease, endometrial and breast cancers, as well as with fertility issues [1–9].

Both cross-sectional and longitudinal studies have shown an overall decline in age of menarche in US girls from the 1960s to the 1990s [10–16]. These studies have also shown clear differences in age of sexual maturation of European Americans compared to African Americans, with African American girls attaining menarche earlier than European American girls [13]. Childhood obesity, higher in African American adolescents than other groups, has been linked to the earlier timing of menarche observed compared with European Americans [13,17–19]. A genetic component for the timing of menarche has been investigated in numerous twin and large population studies, with heritability estimates ranging from 0.49 in the Fels Longitudinal Study to 0.72 in the Breakthrough Generations Study [11,20–22].

Similar to the timing of age at menarche, the age at which natural menopause occurs is affected by multiple factors [23]. Active smoking is consistently associated with earlier menopause; however, the effects of exposure to other carcinogens and endocrine disruptors have not been completely elucidated [24–26]. Diet and obesity are also suggested to impact the timing of natural menopause [27,28]. Based on twin studies and mother-daughter pairs, the heritability of age at natural menopause has been estimated to be between 44–63% [29–31]. Family history of the timing of these reproductive events is a strong predictor of both AM and ANM [29–31].

Genetic and environmental factors that determine AM and ANM have been considered in numerous studies, but many of these studies have conflicting or unreplicated results [32–34]. Furthermore, the majority of these studies have been performed in cohorts of largely European or Asian descent [35–39]. As a result, generalization of genetic associations with AM/ANM to other race/ethnicities is lacking. A recent review has noted the absence of studies with non-European-descent ethnicities and suggests expanding future studies to include other race/ethnicities [40] to identify genetic factors that influence AM/ANM across all populations. Replication of known ANM loci identified in European-descent women has been demonstrated in Hispanic women in the Women's Health Initiative (WHI); however, to our knowledge, there has been no genome-wide association study (GWAS) or generalization study published to date on AM or ANM with an African American cohort [41].

In this study, we used data from the Metabochip genotyping array to characterize previously identified variants associated with menarche and menopause in African Americans in a combined cohort of African-American women from the Women's Health Initiative (WHI) and Atherosclerosis Risk in Communities (ARIC) studies [42] as part of the Population Architecture using Genomics and Epidemiology (PAGE) Study [43]. The Metabochip array is based on the Illumina iSelect platform and contains approximately 200,000 single nucleotide polymorphisms (SNPs) consisting of GWAS index variants and fine-mapping common and less common variants for GWAS-identified regions relevant to metabolic and cardiovascular traits [43,44]. Using current GWAS and candidate gene literature as a guide, we attempted to generalize previously identified menarche and menopause SNPs and gene regions identified in European-descent populations to African Americans in the PAGE Study. We then sought to identify novel SNPs associated with AM and/or ANM.

Materials and Methods

Study Participants

Women participants from two cohorts of the PAGE Study [42], Atherosclerosis Risk in Communities Study (ARIC) and the Women's Health Initiative (WHI), were included in these analyses. ARIC is a population-based prospective study of cardiovascular diseases and their causes in ~16,000 men and women aged 45–64 at baseline [45]. Participants were recruited in Forsyth County, N.C., Jackson, M.S., Minneapolis, M.N., and Washington County, M.D. From this group, 2,070 women, all of self-reported African American ancestry and with information on reproductive timing, were selected for study. The WHI is a long term national health study investigating the leading causes of mortality and frailty in post-menopausal women in the United States, including heart disease, breast and colorectal cancer, and osteoporotic fractures [46]. A subset of 2,455 self-reported African American women selected based on consent to use DNA and availability of DNA, blood lipids, and glucose and insulin measurements were included in this study. The appropriate institutional review board at each participating study site approved all procedures, and written informed consent was obtained from all participants.

Definition of Age at Menarche and Age at Natural Menopause

Age at menarche was defined as the age when menstrual periods started in years, with extreme values pooled in groups of 9 years or less and 17 years or older. Age at natural menopause was defined as the age at which cessation of regular menstrual periods due to the body's natural aging process occurred. In ARIC, women were asked, "Was your menopause natural or the result of surgery or radiation?" Only women who indicated natural menopause were included. Women in WHI who underwent hysterectomy, oophorectomy, or hormone replacement therapy before the onset of natural menopause were excluded. In both studies, women reporting age at natural menopause <40 years were excluded; women reporting age at natural menopause >60 years were censored at age 60. All women included in the present study were post-menopausal.

Genotyping

Genotyping was performed on the Metabochip, a custom Illumina iSelect genotyping chip designed to genotype SNPs associated with metabolic traits and cardiovascular disease [43,44]. The array also includes 2,207 SNPs associated at genome-wide significance to any trait published in the NHGRI GWAS catalog as of August 1, 2009. For each of these GWAS-identified SNPs, an additional proxy SNP with $r^2>0.90$ in the CEU HapMap II dataset, plus up to four additional SNPs with $r^2>0.5$ in the YRI HapMapII dataset were also included on the array. Lastly, SNPs selected to fine-map regions of interest related to metabolic traits, copy number variant-tagging SNPs, Major Histocompatibility Complex (MHC) SNPs, SNPs on the X and Y chromosomes, mitochondrial DNA SNPs, and "wildcard" SNPs were also targeted, for a total of approximately 200,000 SNPs. Of these, 161,098 (81.9%) passed quality control filters for tests of Hardy-Weinberg Equilibrium ($>1\times10^{-7}$) and genotyping efficiency (>95% call rate). There was no filter for minor allele frequency. The design and performance of this genotyping chip in this African American sample has been described in detail elsewhere [43].

Statistical Analysis

All participants self-reported African American ancestry. To adjust for potential population stratification, we used the principal components method implemented in Eigenstrat [47]. We excluded any ancestry outliers further than eight standard deviations away from the mean for the first ten principal components determined by EIGENSOFT.

Linear regression was performed assuming an additive genetic model to test for associations between individual SNPs and the outcomes of age at menarche in years. We examined two models for menarche: 1) a minimally adjusted model that accounted only for study sites and principal components, and 2) a fully adjusted model that included study site, year of birth, principal components, and body mass index at ascertainment, with the understanding that BMI at ascertainment may be a poor proxy for BMI at age of menarche. Age at menarche was self-reported many years later at time of examination, which has been shown to be fairly accurate [48]. We studied one model for natural menopause using Cox's proportional hazards for time-to-event (natural menopause) analysis, which adjusted for study site, principal components, and year of birth. Women with a missing age at menopause, an age at menopause <40 years, or hysterectomy, oophorectomy, or hormone replacement therapy after age 40 but prior to menopause, were excluded from the study. Women who had menopause >60 years had their ANM set as censored at age 60. A fixed effects meta-analysis was then performed using METAL to obtain effect size and standard error (SE) estimates [49]. All analyses were carried out in either METAL or the R software package, and data were plotted using LocusZoom [50,51]. Statistical power to detect an expected association was estimated in Quanto [52] assuming the observed sample size and coded allele frequency in this African American cohort and the genetic effect size previously reported in the literature.

The overall goal of the study was to test for SNPs associated with AM and/or ANM using the Metabochip in African Americans from the WHI and ARIC studies. We looked to generalize to our population of African American women genes, gene regions (400 kb upstream and downstream of a gene of interest), and SNPs described in previous GWAS and candidate gene studies associated with AM and ANM. We tested all SNPs in the regions regardless of linkage disequilibrium (LD) with the index SNP, although we only considered a test of association generalized if the tested SNPs were identical to the index SNP or in strong LD with the index variant in HapMap CEU samples. For each candidate gene, we plotted results of single SNP tests of association using LocusZoom and examined regions 400 kb upstream and downstream of the gene/gene region of interest. Tests of association were considered significant for generalization at a liberal threshold of p<0.05. For previously reported variants not genotyped in our study, we identified SNPs in LD with our directly genotyped SNPs [53] and reported results from our minimally adjusted model (Model 1) for the proxy SNPs.

In addition to generalization, we sought to discover novel SNP-trait associations using the entire Metabochip. Significance in this discovery phase was defined as $p < 3.1 \times 10^{-07}$, after Bonferroni correction (0.05/161,098). Because this threshold is highly conservative given the correlation among the SNPs on the Metabochip, we also defined an arbitrary suggestive significance level as $p < 1 \times 10^{-4}$ in the discovery phase.

Results

Study Population

A total of 4,159 and 1,860 African American female participants met the study definitions for AM and ANM, respectively, and both PAGE studies were represented roughly equally (Table 1). In ARIC, the mean age at menarche was 12.9 years, which was slightly greater than the mean age at menarche in WHI (12.6 years) (Table 1). For ANM, the WHI group had a slightly later average onset than ARIC (Table 1).

Age at Menarche: Generalization to PAGE African Americans

To generalize previously-associated genetic variants in our African American population, we examined regions/genes previously associated with AM from either published candidate gene studies or GWAS: *CYP19A1, CYP17, CYP1B1, FTO, LIN28B*, 9q31.2 region, *IGF1, TNFSF11, TNFRSF11A*, and *LHCGR* [35–37,54–59]. We also evaluated forty-two SNPs associated with AM identified in a recent meta-analysis by Elks *et al.* of >87,000 European-descent women from forty-nine studies [35].

Overall, 11/21 (52%) SNPs previously identified for AM from earlier studies and 15/42 (36%) from the Elks *et al.* meta-analysis were directly genotyped or in strong ($r^2 > 0.70$) LD in the CEU panel of HapMap with those genotyped (Table 2 and Table S1, respectively), and one generalized to this African American cohort: rs9385399, in LD with previously reported rs1361108 ($r^2 = 1.00$, p = 0.01) (Table S1). Representative results of tests of association and LD in this African American sample are given for *CYP19A1*, *FTO*, *LIN28B*, and *CYP1B1*– genes previously associated with AM (Figure 1) [35,36,54,60]. Three SNPs in *LIN28B* were included on the Metabochip (rs314277, rs4946651, and rs7759938), and while the direction of genetic effect was consistent with previous reports, all failed to reach statistical significance in this sample (p>0.30). Four additional SNPs in LD with these *LIN28B* SNPs were also not significant. At the 9q31 locus, rs7861820 and rs4452860, both located downstream of *TMEM38B*, had betas opposite to prior reports [36,37]. Neither SNP nor their proxy SNPs were significant at p<0.05. Similarly, SNPs in LD (rs1856142 and rs605765) with previously associated variants in and around *FSHB* were not significantly associated with AM in this African American sample, though rs605765 ($\beta = -0.06$) had the same direction of effect and comparable magnitude as rs1782507 ($\beta = -0.07$) [59]. Results obtained under our fully adjusted model (Model 2) were similar to those of Model 1 and are available in Table 2.

We also examined SNPs associated with AM that were reported in a recent meta-analysis performed by Elks *et al.* for the ReproGen Consortium [35]. Of the forty-two SNPs associated with AM in Elks *et al.*, we detected an association with rs9385399 (p = 0.01), located downstream of *CENPW*, which is a perfect proxy ($r^2 = 1.00$) for previously associated variant rs1361108, and the only SNP to generalize to our African American sample. We also identified an association with rs2947411 (p = 0.02) with AM (Table S1), though the directions of effect were opposite. One additional SNP, rs4929923 (p = 0.06), nearly reached the significance threshold and had a similar magnitude and direction of effect compared with the previous report. Overall, AM SNPs from previously published studies of European-descent women, including the Elks et al. meta-analysis, did not generalize to our PAGE African American population.

Table 1. Study population characteristics of African American women from the PAGE Study.

	Age at Menarche (AM)		Age at Natural Menopause (ANM)	
	Study Population		Study Population	
	ARIC	WHI	ARIC	WHI
Participants (n)	2078	2081	994	866
Age at menarche, yrs/Age at menopause, yrs	12.89 (1.76)	12.56 (1.64)	47.97 (3.83)	50.84 (4.50)
Age at enrollment, yrs	53.36 (5.73)	61.01 (6.87)	53.07 (5.75)	61.30 (6.78)
Body mass index, kg/m²	30.86 (6.63)	31.34 (6.83)	31.29 (6.94)	30.95 (6.76)
Weight, lbs.	181.05 (39.68)	182.87 (41.26)	183.78 (40.80)	181.05 (40.63)
Height, in.	64.24 (2.43)	64.00 (2.63)	64.31 (2.38)	64.05 (2.75)
Decade of birth, #(%) 1910s	–	26 (1.24)	–	12 (1.39)
1920s	504 (24.07)	414 (19.82)	221 (22.23)	183 (21.13)
1930s	1083 (51.72)	981 (46.96)	522 (52.52)	414 (47.81)
1940s	507 (24.21)	668 (31.98)	251 (25.25)	257 (29.68)

Data presented as means (sd) unless otherwise noted. Abbreviations: Atherosclerosis Risk in Communities (ARIC) and Women's Health Initiative (WHI).

doi:10.1371/journal.pone.0055258.t001

Age at Natural Menopause: Generalization to PAGE African Americans

As with AM, to generalize results to our African American population, we examined previously identified 400 kb regions around genes associated with ANM from published candidate gene studies and GWAS: *APOE*, *CYP1B1*, *CYP19A1*, *CYP17A1*, *ESR2*, *BRSK1*, *FSHB*, *IGF2R*, *PPARG*, *TNFSF11*, *TNFRSF11A*, *BMP15*, *AMHR2*, *TMEM224*, *MCM8*, and *IGF1* (Figure 2, Table 3) [36,38,58,59,61–65]. We also examined twenty SNPs associated with ANM that were identified in a recent study by Stolk *et al.* [66] (Table S2).

Overall, 14/23 (40%) SNPs previously identified for ANM via GWAS and 6/20 SNPs from the Stolk *et al.* meta-analysis were directly genotyped on the Metabochip or were in strong LD ($r^2 > 0.70$) in CEU panel of HapMap. 1/12 (8%) of the tested SNPs in these regions/genes generalized to this African American sample: rs8113016 (Table 3). Rs8113016, located in an intron of *TMEM150B/TMEM224* and downstream of *BRSK1*, is in LD with previously reported rs897798 ($r^2 = 0.72$) and was associated with ANM in our sample (p = 0.03). An intronic *APOE* variant, rs769450, was associated with ANM (p = 0.03), though the nonsynonymous *APOE* rs7412 was not (p = 0.55); these SNPs are not in LD with each other ($r^2 = 0.04$). In *BRSK1*, no previously reported SNPs were genotyped in our study; however, directly genotyped intronic *TMEM150B* rs4806660 was in very strong LD with intronic *BRSK1* rs1172822 ($r^2 = 0.98$). *BRSK1* rs1168309, in strong LD with rs2384687 ($r^2 = 0.85$) was not associated with ANM in this African American sample (p = 0.59).

Three of the twenty SNPs recently identified by Stolk *et al.* as associated with ANM were directly genotyped on the Metabochip. Two of the three genotyped SNPs (rs2303369 and rs2153157) had the same directions of effect, though the magnitudes were smaller. Of the remaining 17 SNPs not directly targeted by the Metabochip, three were in strong LD (HapMap CEU r^2 ranging from 0.86 to 0.91) with the SNPs identified by Stolk *et al*: rs1176133, rs4668368, and rs12593363. For seven SNPs, no proxy SNP could be identified on the Metabochip (Table S2). Of the twenty SNPs identified in the Stolk *et al.* meta-analysis and directly or indirectly represented on the Metabochip, none were associated with ANM in this African American sample (Table S2).

Age at Menarche: Discovery

We tested all SNPs genotyped on the Metabochip for an association with AM adjusted for study site and principal components (Model 1) and adjusted for study site, year of birth, principal components, and body mass index (Model 2) (Table 4). After accounting for multiple testing (p < 3.1×10^{-07}), no SNPs were significantly associated with AM in either model (Table S3). The most significant SNP in both models was rs11604207 (Model 1: p = 1.59×10^{-06}; Model 2: p = 1.82×10^{-06}), which is located upstream of *RSF1*, a gene encoding a chromatin remodeling protein implicated in ovarian and breast cancers [67–69] (Table S3).

Age at Natural Menopause: Discovery

We tested all SNPs on the Metabochip for associations with ANM adjusted for study site and principal components. Three SNPs were significant after Bonferroni correction (p < 3.1×10^{-07}): *LDLR* (rs189596789, p = 4.98×10^{-08}), *KCNQ1* (rs79972789, p = 1.90×10^{-07}), and *COL4A3BP* (rs181686584, p = 2.85×10^{-07}). The most significant association was with rs189596789, located approximately 10 kb upstream of the low-density lipoprotein receptor (*LDLR*) gene, which has been associated with familial hypercholesterolemia [70,71]. Several of the most significant SNPs for ANM were located in/near genes previously associated with obesity, type 2 diabetes (T2D), coronary artery disease and lipid metabolism, e.g., *LDLR* (rs189596789), *NOS1AP* (rs76078015), *DGKB* (rs74486449), *LYPLAL1* (rs78696400), and *CDKAL1* (rs114158228) (Table 4). We were unable to generalize the previously reported association between ANM and *PPARG* rs4135280 in this African American sample.

Two genes were suggestively associated with both ANM and AM at a nominal significance threshold. *PHACTR1* was suggestively associated with AM (rs73725617; Table S3) and ANM (rs117124693; Table 4). Though the direction of effects was similar for each SNP in *PHACTR1*, the SNPs are not in LD with each other. Likewise, SNPs in *ARHGAP42*, located at the 11q22.1 locus, were suggestively associated with AM (rs11224447; Table S3) and ANM (rs11224401; Table 4), but are not in LD with each other, though the direction of effects was the same.
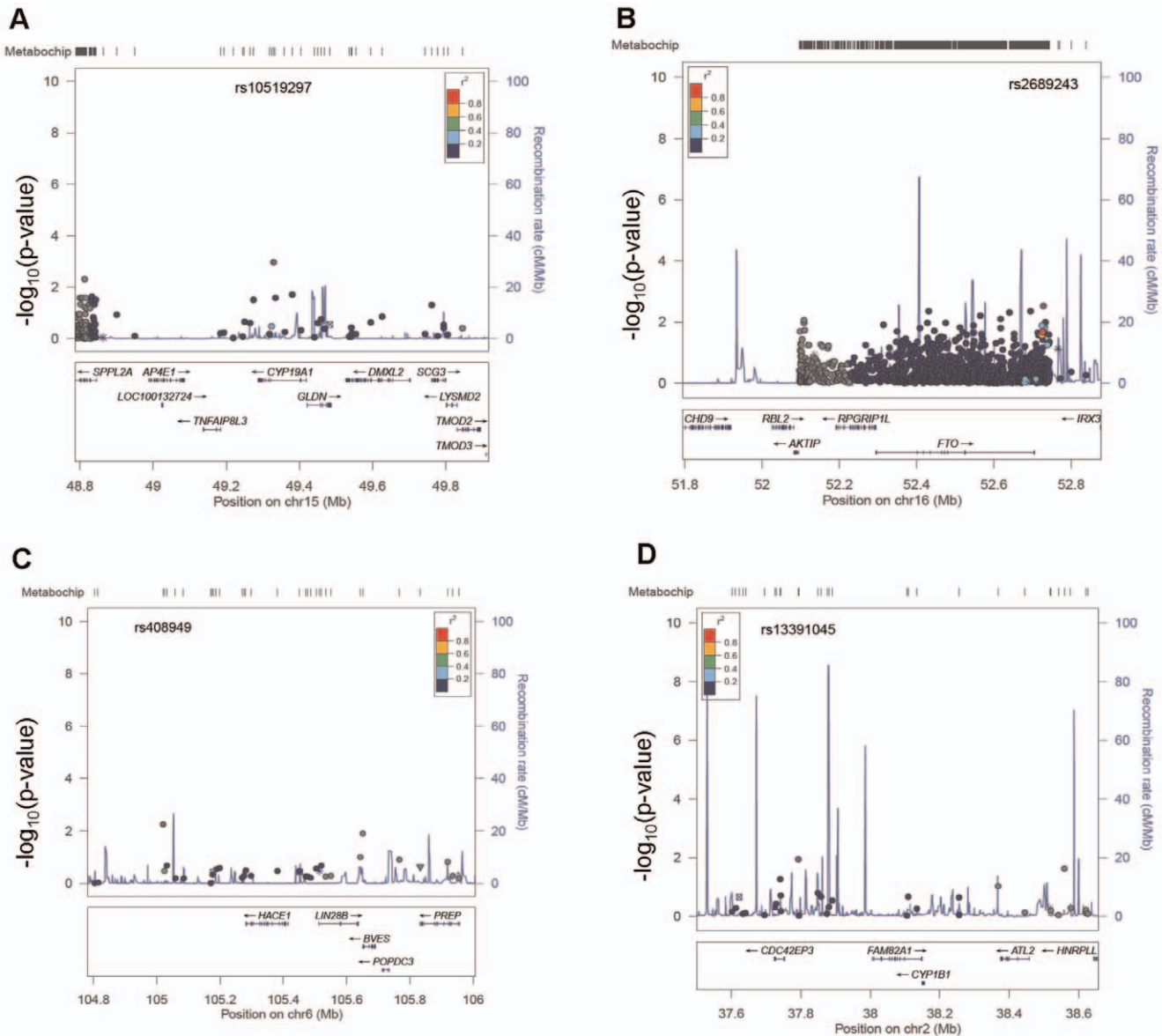
Figure 1. Regional Association Plots for Age at Menarche in African American women in the PAGE Study. Locus Zoom plots for selected gene regions in age at menarche analysis. Vertical axis is $-\log_{10}$ of the p-value, the horizontal axis is the chromosomal position. Each dot represents a SNP tested for association with age at natural menopause in 1,860 African American women from the PAGE Study. Approximate linkage disequilibrium between the most significant SNP, listed at the top of each plot, and the other SNPs in the plot is shown by the r^2 legend in each plot. (A) Locus Zoom plot for the *CYP19A1* region, with rs10519297 the most significant SNP in the region. (B) Locus Zoom plot for the *FTO* region; rs2689243 was the most significant SNP in the plot region. (C) *LIN28B* region Locus Zoom plot; rs408949 was the most significant SNP in the plot region. (D) Locus Zoom plot of the *CYP1B1* region; rs13391045 was the most significant SNP in the plot region.
doi:10.1371/journal.pone.0055258.g001

Discussion

Here we demonstrated the use of the Metabochip genotyping array to identify SNPs associated with AM and ANM in a sample of African American women. Previous GWAS studies for AM and ANM have been performed in primarily European descent populations; generalization to diverse populations has largely been lacking [72]. Our study is the first, to our knowledge, to consider this trait in a large African American cohort. We were able to generalize only one previously identified variant for AM and two variants for ANM to our African American cohort [AM: rs1361108; ANM: rs897798 and rs9385399 (proxy for rs1361108)]. Overall, however, we were unable to generalize the

majority of significant associations for previously identified SNPs associated with AM, including *LIN28B* or the 9q31 locus, or with ANM, including *MCM8* or *TMEM150b/TMEM224*, which have recently been identified in several GWAS of European-descent women. Our inability to replicate earlier findings in our African American sample may have, in part, resulted from scant Metabochip coverage of these regions. The emphasis of the Metabochip on genes involved in lipid metabolism and cardiovascular traits is evident comparing coverage in the *FTO* region (1053 SNPs) to the *LIN28B* region (28 SNPs).

In the discovery phase of our AM analysis, none of our results reached genome-wide significance. However, the ANM analysis

Table 2. Comparison of GWAS-identified AM variants in African American women from the PAGE Study.

| Locus SNP | Chr | Gene/Region | Prior GWAS in European descent women | | | | African American women from the PAGE Study | | | Model 1 | | Model 2 | |
			Coded Allele	Beta	P-value	Ref.	Best Proxy SNP from present study	r² in HapMap CEU/YRI	Coded Allele	Beta (SE)	P-value	Beta (SE)	P-value
rs314277	6	LIN28B	A	0.16	2.7E-13	[36]	rs314277	–	A	0.03(0.04)	0.34	0.03(0.04)	0.36
rs369065	6	LIN28B	C	0.11	2.4E-11	[36]	rs7759938	1.00/0.34	A	−0.02(0.04)	0.61	−0.02(0.04)	0.55
rs7759938	6	LIN28B	C	0.09	7.0E-09	[37]	rs7759938	–	A	−0.02(0.04)	0.61	−0.02(0.04)	0.55
rs314276	6	LIN28B	C	−0.22	1.5E-08	[57]	rs314274	1.00/0.73	A	0.05(0.04)	0.22	0.05(0.04)	0.24
rs314280	6	LIN28B	T	0.09	2.3E-08	[36,39]	rs7759938	0.64/0.28	A	−0.02(0.04)	0.61	−0.02(0.04)	0.55
rs4946651	6	LIN28B	A	0.09	3.1E-08	[36]	rs4946651	–	A	0.03(0.04)	0.55	0.03(0.04)	0.55
rs314262	6	LIN28B	C	0.08	9.7E-08	[36]	rs7759938	0.60/0.29	A	−0.02(0.04)	0.61	−0.02(0.04)	0.55
rs7861820	9	9q31	C	−0.09	3.4E-09	[36]	rs7861820	–	A	−0.10(0.06)	0.10	−0.09(0.06)	0.12
rs12684013	9	9q31	T	−0.10	3.6E-08	[36]	rs4452860	0.81/0.01	A	−0.03(0.04)	0.43	−0.03(0.04)	0.42
rs4452860	9	9q31	G	−0.09	7.9E-08	[36]	rs4452860	–	A	−0.03(0.04)	0.43	−0.03(0.04)	0.42
rs7028916	9	9q31	A	−0.09	9.7E-08	[36]	rs4452860	0.98/0.85	A	−0.03(0.04)	0.43	−0.03(0.04)	0.42
rs2090409	9	9q31	A	−0.10	1.7E-09	[37]	rs4452860	0.83/0.82	A	−0.03(0.04)	0.43	−0.03(0.04)	0.42
rs555621	11	FSHB	C	0.06	0.001	[59]	rs1856142	0.43/0.71	A	0.03(0.04)	0.44	0.03(0.04)	0.36
rs1782507	11	FSHB	T	−0.07	0.006	[59]	rs605765	0.83/0.87	A	−0.06(0.04)	0.14	−0.06(0.04)	0.13
rs4953616	2	LHCGR	T	−0.07	0.006	[59]	rs1589749	0.17/0.05	A	0.002(0.07)	0.97	−0.01(0.07)	0.87
rs7579411	2	LHCGR	T	0.06	0.01	[59]	rs1589749	0.17/0.05	A	0.002(0.07)	0.97	−0.01(0.07)	0.87
rs4374421	2	LHCGR	C	0.06	0.02	[59]	rs17326321	0.19/0.69	A	−0.01(0.06)	0.86	−0.01(0.06)	0.84
rs2470144	15	CYP19A1	G	–	5.9E-06	[54]	rs12148492	0.23/0.01	A	−0.01(0.07)	0.91	−0.02(0.07)	0.73
rs2445761	15	CYP19A1	G	–	1.2E-06	[54]	rs4774585	0.28/0.02	A	0.04(0.05)	0.47	0.03(0.05)	0.58
rs9525641	13	TNFSF11/RANKL	T	–	0.04	[58]	rs931273	0.05/0.03	A	0.11(0.09)	0.24	0.11(0.09)	0.21
rs3826620	18	TNFRSF11A/RANK	A	–	0.02	[58]	rs8092336	0.16/0.22	A	0.16(0.17)	0.33	0.17(0.17)	0.29
rs6214	12	IGF1	G	–	0.02	[56]	rs6214	–	A	−0.01(0.04)	0.71	−0.02(0.04)	0.61

Comparison of previously reported SNPs associated with AM in European descent women to 4,159 African American women from the PAGE Study in a minimally adjusted model for AM (Model 1) and a model adjusted for study site, year of birth, principal components, and body mass index (Model 2). Data presented are for the previously identified SNP. If the previously identified SNP was not directly genotyped in present study, data shown are for the best proxy SNP based on linkage disequilibrium from the International HapMap Project CEU panel.
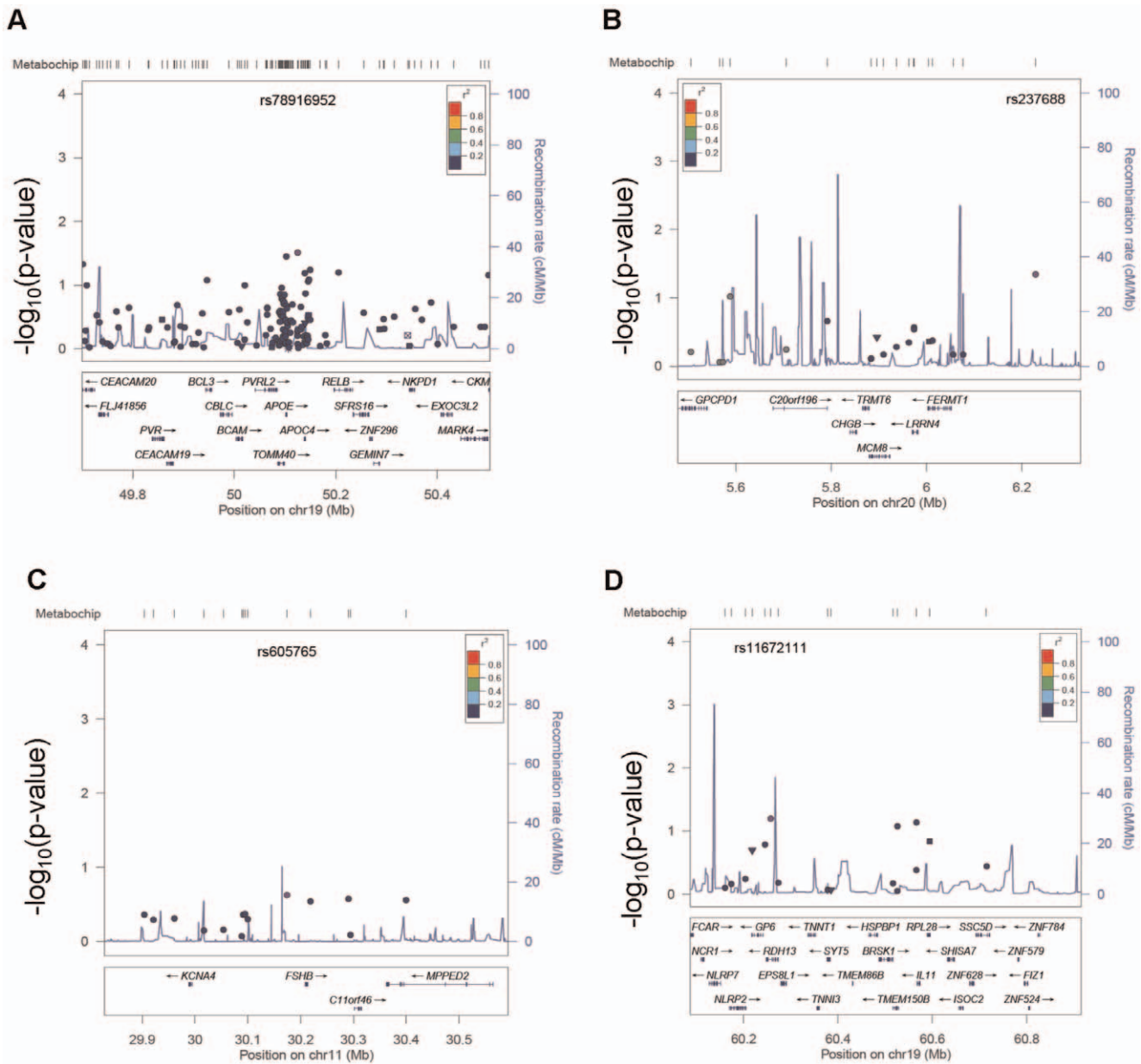doi:10.1371/journal.pone.0055258.t002

Figure 2. Regional Association Plots for Age at Natural Menopause in African American women in the PAGE Study. Locus Zoom plots for selected gene regions in age at natural menopause analysis. Vertical axis is the −log$_{10}$ of the p-value, the horizontal axis is the chromosomal position. Each dot represents a SNP tested for association with age at natural menopause in 1,860 African American women from the PAGE Study. Linkage disequilibrium between the most significant SNP, listed at the top of each plot, and the other SNPs in the plot is shown by the r^2 legend in each plot. (A) Locus Zoom plot for the *APOE* region, with rs78916952 the most significant SNP in the region. (B) Locus Zoom plot for the *MCM8* region; rs237688 is the most significant SNP in the plot region. (C) *FSHB* region Locus Zoom plot; rs605765 is the most significant SNP in the plot region. (D) Locus Zoom plot of the *BRSK1* region with rs11672111 as the most significant SNP in the plot region.
doi:10.1371/journal.pone.0055258.g002

yielded three associations that were significant after multiple testing corrections. Broadly, we demonstrate the ability to potentially uncover new variants associated with age at natural menopause in our African American cohort using the Metabochip.

Several studies have shown relationships between a woman's reproductive milestones (AM, ANM, parity) and menstrual characteristics and risk for breast cancer, endometrial cancer, and ovarian cancer [73–77] and chronic diseases such as diabetes, osteoporosis and cardiovascular disease (briefly [78–80]). The most significant result in the ANM analysis was a SNP located upstream of *LDLR* (rs189596789) which encodes a low density

lipoprotein receptor implicated in familial cholesterolemia. *KCNQ1* (rs79972789) also reached genome wide significance in our ANM analysis. Numerous variants in *KCNQ1* have also been implicated in type 2 diabetes in several populations, though none were in linkage disequilibrium with rs79972789 [81–86]. Recently, Buber *et al.* evaluated the role of menopausal hormonal changes with cardiac events in women with mutations in *KCNQ1* and congenital long-QT syndrome (LQTS) [87] and determined the onset of menopause was associated with an increase in the risk of cardiac events in LQTS women. Though not significant, suggestive AM associations included *LPL* and *CYP4F22*, which are associated with

Table 3. Comparison of GWAS-identified ANM variants in African American women in PAGE Study.

Locus			Prior GWAS in European descent women				African American women from the PAGE Study				
SNP	Chr	Gene/region	Coded Allele	Beta	P-value	Ref.	Best Proxy SNP from present study	r² in HapMap CEU/YRI	Coded Allele	Beta (SE)	P-value
rs16991615	20	MCM8	A	1.07	1.21E-21	[36,62]	rs16991615	–	A	−0.17(0.15)	0.25
rs236114	20	MCM8	A	0.50	9.71E-11	[38]	rs236114	–	A	0.02(0.06)	0.69
rs1172822	19	BRSK1	T	−0.49	1.8E-19	[36,38]	rs4806660	0.98/0.64	A	0.002(0.03)	0.97
rs2384687	19	BRSK1	C	−0.47	2.4E-18	[36]	rs11668309	0.85/0.43	A	0.02(0.04)	0.59
rs897798	19	BRSK1	G	−0.40	1.1E-14	[36]	rs8113016	0.72/0.02	A	0.12(0.05)	0.03
rs1065778	15	CYP19A	A	–	0.05	[61]	rs10519297	0.90/0.32	A	−0.01(0.05)	0.84
rs2255192	15	CYP19A	A	–	0.04	[61]	rs10459592	0.32/0.02	A	−0.02(0.04)	0.52
rs621686	11	FSHB	A	0.32	0.007	[59]	rs1856142	0.27/0.32	A	0.04(0.03)	0.29
rs7951733	11	FSHB	A	−0.32	0.02	[59]	rs7951733	–	A	0.11(0.13)	0.37
rs769450	19	APOE	A	–	0.007	[97]	rs769450	–	A	−0.07(0.03)	0.03
rs7412	19	APOE	–	–	0.001	[98]	rs7412	–	A	−0.03(0.05)	0.55
rs1019731	12	IGF1	C	−0.28	0.005	[59]	rs1019731	–	A	−0.03(0.11)	0.82
rs9457827	17	IGF2R	T	0.37	0.04	[59]	rs9457827	–	A	0.04(0.04)	0.28
rs4135280	3	PPARG	T	0.54	0.005	[59]	rs4135280	–	A	−0.14(0.18)	0.42
rs1256044	14	ESR2	G	–	0.03	[61]	rs1268656	0.08/0.004	A	−0.01(0.06)	0.88
rs1256059	14	ESR2	A	–	0.05	[61]	rs1268656	0.08/0.004	A	−0.01(0.06)	0.88
rs1056836	2	CYP1B1	G	–	0.04	[64]	rs10495874	0.04/0.03	A	−0.03(0.05)	0.60
rs346578	13	TNFSF11	A	–	0.007	[58]	rs6561072	0.07/0.07	A	0.04(0.04)	0.22
rs9525641	13	TNFSF11	T	–	0.01	[58]	rs931273	0.05/0.03	A	−0.02(0.08)	0.81
rs8086340	18	TNFRSF11A	G	–	0.02	[58]	rs8094440	0.10/0.01	A	0.03(0.03)	0.38
rs2002555	12	AMHR2	G	0.30	0.02	[65]	rs7131938	0.59/0.54	A	0.01(0.04)	0.84
rs2384687	19	TMEM224	C	0.38	1.39E-10	[38]	rs11668309	0.85/0.43	A	0.02(0.04)	0.59
rs897798	19	TMEM224	G	0.31	3.91E-08	[38]	rs8113016	0.72/0.02	A	0.12(0.05)	0.03

Comparison of previously reported SNPs associated with ANM in European and Chinese descent women to 1,860 African American women from the PAGE Study. Data presented are for the previously identified SNP. If the previously identified SNP was not directly genotyped in present study, data are shown for the best proxy SNP based on linkage disequilibrium calculated from the International HapMap Project CEU data.
doi:10.1371/journal.pone.0055258.t003

type 2 diabetes and lipid metabolism (rs1372339, rs4922116, rs1273516), and *TMEM18* (rs2947411), associated with obesity and body mass index [88,89]. These ANM associations and suggestive AM associations with genes involved in cardiovascular function, lipid metabolism, and type 2 diabetes concur with research showing later AM lowers obesity and diabetes risk while earlier ANM increases risk for cardiovascular disease, obesity and insulin resistance [90,91].

Different pathways appear to be involved in the initiation and cessation of menses. Prior GWAS and linkage studies performed in European descent or Asian populations for AM and ANM show little concordance with specific genes (reviewed in [5]). Our analysis is consistent with this observation. Only *PHACTR1* and *ARHGAP42* SNPs were suggestively significant in both our AM and ANM analyses. *PHACTR1* is a phosphatase and actin regulator which has been implicated in coronary artery disease [92,93]. Its role in menarche and menopause is yet to be determined. *ARHGAP42*, a Rho GTPase activating protein, has not yet been evaluated for a role in menarche or menopause. A GWAS identified intronic *ARHGAP42* rs633185 is associated with blood pressure [94], but this variant is not in strong LD with *ARHGAP42* variants suggestively associated with either AM or ANM in this study. A recent study by Lu *et al.*, found SNPs in both

TNFSF11 and *TNFRSF11A* significant for AM and ANM [58]. SNPs genotyped on the Metabochip were in weak LD with the reported SNPs and failed to reach significance in this African American sample. Given the role that both *PHACTR1* and *ARHGAP42* play in atherosclerosis, osteoporosis and the development of lactation glands in pregnancy, further investigation on the influence of these genes in AM and ANM is warranted [95,96].

The Metabochip was designed to be a cost-effective method of genotyping approximately 200,000 metabolic and cardiovascular SNPs and SNPs in other useful regions of the genome, such as the HLA region and the X and Y chromosomes. Overall, median SNP density on the Metabochip is approximately one SNP per 370 bases [43]. This coverage appears sufficient to replicate some loci associated with both cardiovascular or metabolic traits and AM/ANM. However, we found instances of previously identified genes for AM/ANM with little/no Metabochip coverage (*CYP1B1*, *LIN28B*, *ESR2*, and *BRSK1*) which may have impacted our results. Additionally, prior studies that identified SNPs associated with AM and ANM were performed primarily in European-descent cohorts. Though our study included over 4,000 African American women, we had limited power to identify significant associations in most previously identified loci, which may explain why we failed to detect the same associations identified in European-descent

Table 4. ANM Discovery–SNPs associated with age at natural menopause (ANM) in African American women from the PAGE Study.

CHR	SNP	GENE	GENE REGION	CODED ALLELE	CAF	BETA	SE	P VALUE
19	rs189596789	LDLR	upstream	A	0.006	1.09	0.20	4.98E-08
11	rs79972789	KCNQ1	intronic	C	0.997	−1.76	0.34	1.90E-07
5	rs181686584	COL4A3BP	intronic	A	0.002	2.35	0.46	2.85E-07
6	rs114158228	CDKAL1	intronic	A	9E-04	3.60	0.73	7.12E-07
21	rs117876865	KCNE1	downstream	A	9E-04	3.58	0.73	8.55E-07
10	rs11195485	ADRA2A	downstream	A	0.002	2.89	0.59	9.63E-07
11	rs11224401	ARHGAP42	intronic	A	0.997	2.20	0.45	1.13E-06
1	rs78937547	SEC16B	downstream	A	0.992	−1.97	0.41	1.89E-06
17	rs75394140	KCNJ2	downstream	A	0.002	−0.93	0.21	6.48E-06
11	rs76988592	KCNJ1	downstream	A	0.702	−0.93	0.21	7.24E-06
3	rs114451007	PPARG	intronic	A	0.253	1.70	0.38	9.30E-06
12	rs10846771	DHX37	downstream	A	0.997	−0.16	0.04	9.43E-06
11	rs12804247	CCDC81	upstream	A	0.655	0.17	0.04	1.45E-05
1	rs76571116	SEC16B	downstream	A	3E-04	−1.54	0.36	1.57E-05
17	rs17634167	TTLL6	cds-synon.	A	6E-04	−0.34	0.08	1.62E-05
7	rs117382431	FKBP6	downstream	A	0.999	4.38	1.03	2.17E-05
6	rs76294174	LOC100130357	intronic	C	3E-04	4.38	1.03	2.17E-05
6	rs74918542	SCGN	intronic	A	0.999	−4.38	1.03	2.17E-05
1	rs76078015	NOS1AP	intronic	A	9E-04	4.38	1.03	2.17E-05
18	rs117454233	MC4R	downstream	A	0.999	−4.38	1.03	2.17E-05
3	rs73025249	PPARG	intronic	A	9E-04	4.38	1.03	2.17E-05
3	rs182857216	ETV5	intronic	A	0.999	−4.38	1.03	2.17E-05
3	rs73027210	PPARG	intronic	A	9E-04	4.38	1.03	2.17E-05
9	rs75220302	CDKN2A	downstream	A	0.999	−4.38	1.03	2.18E-05
9	rs74599268	CDKN2B	upstream	A	3E-04	4.38	1.03	2.18E-05
9	rs3731245	CDKN2A	intronic	A	3E-04	4.38	1.03	2.18E-05
9	rs76774391	CDKN2B	upstream	C	3E-04	4.38	1.03	2.18E-05
2	rs117258126	IRS1	downstream	A	3E-04	4.38	1.03	2.18E-05
9	rs3808846	CDKN2B	5' flanking	A	3E-04	4.38	1.03	2.18E-05
9	rs77706751	CDKN2B	upstream	A	6E-04	4.38	1.03	2.18E-05
9	rs3808845	CDKN2B	5' flanking	A	3E-04	4.38	1.03	2.18E-05
9	rs76810097	CDKN2B	upstream	A	3E-04	4.38	1.03	2.18E-05
9	rs36228836	CDKN2A	5' flanking	A	3E-04	4.38	1.03	2.18E-05
9	rs75039118	ADAMTS13	intronic	A	0.999	−4.38	1.03	2.19E-05
18	rs75914913	MC4R	downstream	A	3E-04	4.38	1.03	2.19E-05
11	rs190060931	BUD13	downstream	A	0.999	−4.38	1.03	2.21E-05
2	rs186397905	IRS1	downstream	C	3E-04	4.38	1.03	2.21E-05
16	rs9934222	JPH3	cds-synon.	A	0.163	−0.19	0.04	2.28E-05
15	rs72751410	MAP2K5	intronic	A	0.998	−1.51	0.36	2.30E-05
15	rs72747452	LOC100506686	intronic	A	0.002	1.51	0.36	2.30E-05
11	rs180751580	NUCB2	missense	C	0.999	−4.36	1.03	2.30E-05
3	rs186437034	SCN5A	intronic	A	0.999	−2.46	0.58	2.45E-05
7	rs78912482	JAZF1	upstream	A	0.012	0.64	0.15	3.04E-05
1	rs116071515	SEC16B	intronic	A	0.002	1.88	0.45	3.06E-05
6	rs1997770	OFCC1	downstream	A	0.970	−0.41	0.10	3.55E-05
7	rs118135044	DGKB	upstream	A	4E-04	4.22	1.02	3.73E-05
11	rs74402657	ARFGAP2	intronic	C	4E-04	2.93	0.72	3.96E-05
1	rs117217277	SEC16B	downstream	A	0.999	−2.97	0.72	3.97E-05
1	rs116881786	SEC16B	downstream	A	0.999	−2.97	0.72	3.97E-05

Table 4. Cont.

CHR	SNP	GENE	GENE REGION	CODED ALLELE	CAF	BETA	SE	P VALUE
1	rs76471454	SEC16B	downstream	A	6E-04	2.97	0.72	3.97E-05
1	rs79775735	SEC16B	downstream	A	6E-04	2.97	0.72	3.97E-05
1	rs79468804	SEC16B	downstream	A	6E-04	2.97	0.72	3.97E-05
1	rs74703854	SEC16B	downstream	A	0.999	−2.97	0.72	3.97E-05
1	rs116923068	SEC16B	downstream	C	0.999	−2.97	0.72	3.97E-05
1	rs117674205	SEC16B	downstream	C	0.999	−2.97	0.72	3.97E-05
1	rs117260315	SEC16B	downstream	A	6E-04	2.97	0.72	3.97E-05
1	rs76020919	SEC16B	downstream	A	6E-04	2.97	0.72	3.97E-05
11	rs2306034	LRP4	UTR-3′	A	4E-04	2.94	0.72	3.99E-05
2	rs189110944	IRS1	downstream	A	4E-04	4.17	1.02	4.72E-05
5	rs1976311	KCNN2	upstream	C	0.996	−1.02	0.25	4.98E-05
7	rs13245084	LOC100507421	intronic	A	4E-04	4.14	1.02	5.07E-05
6	rs115178932	LRRC16A	intronic	A	4E-04	4.14	1.02	5.07E-05
1	rs77353590	SYF2	downstream	A	0.009	0.74	0.18	5.42E-05
2	rs111826230	APOB	upstream	A	0.984	−0.58	0.14	5.47E-05
11	rs193030163	DDB2	upstream	C	0.999	−4.11	1.02	5.57E-05
11	rs114702513	KCNQ1	intronic	A	0.996	−1.23	0.31	5.60E-05
6	rs117124693	PHACTR1	intronic	A	0.999	−4.11	1.02	5.62E-05
6	rs181947983	SLC17A3	upstream	A	4E-04	4.11	1.02	5.62E-05
15	rs183951867	CHRNB4	upstream	A	9E-04	4.11	1.02	5.62E-05
9	rs191930498	CDKN2B	upstream	C	4E-04	4.10	1.02	5.83E-05
17	rs192656758	CCT6B	downstream	A	4E-04	4.10	1.02	5.86E-05
7	rs740259	JAZF1	5′ flanking	A	4E-04	4.09	1.02	5.97E-05
1	rs114389068	GPR153	cds-synon.	A	0.005	0.93	0.23	6.07E-05
11	rs185476610	KCNQ1	intronic	A	0.999	−4.08	1.02	6.24E-05
16	rs246192	NDRG4	intronic	C	0.256	0.15	0.04	6.25E-05
7	rs192457106	JAZF1	intronic	A	0.999	−4.08	1.02	6.35E-05
7	rs73702566	WBSCR22	intronic	A	0.999	−4.08	1.02	6.35E-05
6	rs187190790	TAP2D	upstream	A	0.999	−4.08	1.02	6.38E-05
7	rs74984879	DGKB	upstream	C	0.999	−2.04	0.51	6.40E-05
11	rs184056970	ARAP1	intronic	A	4E-04	4.07	1.02	6.53E-05
3	rs76909367	COLQ	intronic	A	4E-04	4.06	1.02	6.89E-05
10	rs11187795	PLCE1	intronic	A	4E-04	4.06	1.02	6.93E-05
6	rs186129489	TFAP2D	intronic	A	4E-04	4.05	1.02	7.12E-05
2	rs73923981	BRE	intronic	A	9E-04	4.05	1.02	7.32E-05
15	rs180807356	ADAMTS7	upstream	A	0.999	−4.04	1.02	7.52E-05
5	rs10062135	NPR3	intronic	A	0.009	0.73	0.19	7.85E-05
12	rs17568045	C12orf42	intronic	A	0.993	−0.86	0.22	8.11E-05
1	rs116411856	WARS2	upstream	A	0.003	1.32	0.34	8.16E-05
1	rs78696400	LYPLAL1	downstream	A	0.985	−0.58	0.15	8.96E-05
15	rs74979292	C15orf39	upstream	A	0.002	1.49	0.38	9.29E-05
11	rs144204188	TRIM66	intronic	A	0.002	2.79	0.72	9.39E-05
1	rs78411379	TBX15	intronic	A	0.999	−2.27	0.58	9.62E-05
15	rs190893945	ADAMTSL3	intronic	A	0.998	−1.76	0.45	9.67E-05
9	rs12555547	CDKN2B	upstream	C	0.998	−2.30	0.59	9.69E-05
2	rs10932320	C2orf67	intronic	A	0.807	−0.17	0.04	9.93E-05

Tests of association at p≤1E-04 from single SNP linear regressions adjusted for study site and principal components in 1,860 African American women from the PAGE Study are shown. For each significant test of association, the chromosome, rs number, nearest gene, location, coded allele, beta, standard error (SE), and p-value are given. Genes listed are nearest genes to the SNP as measured from the transcription start site for upstream SNPs or the transcription stop site for downstream SNPs. Abbreviations: CAF, coded allele frequency.
doi:10.1371/journal.pone.0055258.t004

GWAS. For specific tests of association, our power was impacted by sample size and by minor allele frequencies. For example, the allele frequency for rs7861820 in this African American cohort was 0.11 compared to a higher frequency observed in HapMap CEU (0.57; Table S4). Interestingly, we were adequately powered (>98%) to generalize the intronic *LIN28B* SNP, rs314277, with AM in our sample, yet failed to find an association with this SNP or with SNPs in strong LD with it.

Metabochip performance in non-European populations was recently evaluated in a pilot study in African American PAGE participants [43]. In this pilot study, Buyske *et al.* demonstrated that the majority (89%) of SNPs targeted by the Metabochip passed rigorous quality control with high call rates [43]. Using lipid traits as an example, Buyske *et al.* demonstrated that Metabochip data can be used to replicate known GWAS-identified SNP-trait relationships. Furthermore, the pilot study demonstrated that Metabochip data can be used to fine-map GWAS-identified regions to uncover potential novel index SNPs specific to African Americans in an established locus for that trait. Fine-mapping data for AM/ANM was not included in the Metabochip content. While we were able to use the Metabochip to identify potentially novel SNP-trait relationships for AM/ANM, additional fine-mapping efforts of other loci already implicated for these traits are needed. Furthermore, additional studies in general are warranted for diverse (non-European descent) populations using Metabochip or other arrays designed for fine-mapping. Admixture in the African American population and its associated decreased LD compared to European Americans challenge identification of trait-associated SNPs. Targeted fine mapping, such as use of the Metabochip, may be more appropriate in some circumstances than GWAS to evaluate specific SNPs and regions associated with particular traits.

Although the Metabochip was designed for genotyping of cardiovascular and metabolic SNPs, this study demonstrates the feasibility of utilizing such a targeted chip to identify SNP associations with age at menarche and age at natural menopause. We identified potentially novel associations with AM/ANM at loci implicated in cardiovascular traits, obesity and cancer. This may result from pleiotropic loci or may suggest that the AM/ANM timing mechanisms influence underlying disease process. With numerous genes implicated in both metabolic and cardiovascular

phenotypes and both AM and ANM, further studies will allow us to consider how specific genes may influence the reproductive lifespan in women.

Supporting Information

Table S1 Comparison of SNPS in Elks et al. meta-analysis for AM to African American women in PAGE Study.
(DOCX)

Table S2 Comparison of SNPs in Stolk et al. meta-analysis for ANM to African American women in PAGE Study.
(DOCX)

Table S3 AM Discovery—SNPs associated (p<1e−4) with AM in African American women from the PAGE Study.
(DOCX)

Table S4 Minor allele frequency comparisons of African American women in PAGE Study to HapMap CEU Panel.
(DOCX)

Author Contributions

Conceived and designed the experiments: KLS CLC NF LFR AY IC MDR CAH LW CW TCM CSC KB AP AR LAH SB DCC. Performed the experiments: KLS CLC NF LFR AY IC MDR CAH LW CW TCM CSC KB AP AR LAH SB DCC. Analyzed the data: KLS JM CLC NF LFR AY IC MDR CAH LW CW TCM CSC KB AP AR LAH SB DCC. Contributed reagents/materials/analysis tools: KLS JM CLC NF LFR AY IC MDR CAH LW CW TCM CSC KB AP AR SB DCC. Wrote the paper: KLS JM CLC NF LFR LAH SB DCC.

References

1. Hsieh CC, Trichopoulos D, Katsouyanni K, Yuasa S (1990) Age at menarche, age at menopause, height and obesity as risk factors for breast cancer: associations and interactions in an international case-control study. Int J Cancer 46: 796–800.
2. He C, Zhang C, Hunter DJ, Hankinson SE, Buck Louis GM, et al. (2010) Age at menarche and risk of type 2 diabetes: results from 2 large prospective cohort studies. Am J Epidemiol 171: 334–344. kwp372 [pii];10.1093/aje/kwp372 [doi].
3. Lakshman R, Forouhi NG, Sharp SJ, Luben R, Bingham SA, et al. (2009) Early age at menarche associated with cardiovascular disease and mortality. J Clin Endocrinol Metab 94: 4953–4960. jc.2009-1789 [pii];10.1210/jc.2009-1789 [doi].
4. Dossus L, Allen N, Kaaks R, Bakken K, Lund E, et al. (2010) Reproductive risk factors and endometrial cancer: the European Prospective Investigation into Cancer and Nutrition. Int J Cancer 127: 442–451. 10.1002/ijc.25050 [doi].
5. Hartge P (2009) Genetics of reproductive lifespan. Nat Genet 41: 637–638. ng0609-637 [pii];10.1038/ng0609-637 [doi].
6. Lisabeth LD, Beiser AS, Brown DL, Murabito JM, Kelly-Hayes M, et al. (2009) Age at natural menopause and risk of ischemic stroke: the Framingham heart study. Stroke 40: 1044–1049. STROKEAHA.108.542993 [pii];10.1161/STROKEAHA.108.542993 [doi].
7. Atsma F, Bartelink ML, Grobbee DE, van der Schouw YT (2006) Postmenopausal status and early menopause as independent risk factors for cardiovascular disease: a meta-analysis. Menopause 13: 265–279. 10.1097/01.gme.0000218683.97338.ea [doi];00042192-200613020-00017 [pii].
8. Mondul AM, Rodriguez C, Jacobs EJ, Calle EE (2005) Age at natural menopause and cause-specific mortality. Am J Epidemiol 162: 1089–1097. kwi324 [pii];10.1093/aje/kwi324 [doi].
9. Remsberg KE, Demerath EW, Schubert CM, Chumlea WC, Sun SS, et al. (2005) Early menarche and the development of cardiovascular disease risk factors in adolescent girls: the Fels Longitudinal Study. J Clin Endocrinol Metab 90: 2718–2724. jc.2004-1991 [pii];10.1210/jc.2004-1991 [doi].
10. Deardorff J, Ekwaru JP, Kushi LH, Ellis BJ, Greenspan LC, et al. (2011) Father absence, body mass index, and pubertal timing in girls: differential effects by family income and ethnicity. J Adolesc Health 48: 441–447. S1054-139X(10)00389-7 [pii];10.1016/j.jadohealth.2010.07.032 [doi].
11. Morris DH, Jones ME, Schoemaker MJ, Ashworth A, Swerdlow AJ (2011) Familial concordance for age at menarche: analyses from the Breakthrough Generations Study. Paediatr Perinat Epidemiol 25: 306–311. 10.1111/j.1365-3016.2010.01183.x [doi].
12. Carter R, Caldwell CH, Matusko N, Antonucci T, Jackson JS (2011) Ethnicity, perceived pubertal timing, externalizing behaviors, and depressive symptoms among black adolescent girls. J Youth Adolesc 40: 1394–1406. 10.1007/s10964-010-9611-9 [doi].
13. Salsberry PJ, Reagan PB, Pajer K (2009) Growth differences by age of menarche in African American and White girls. Nurs Res 58: 382–390. 10.1097/NNR.0b013e3181b4b921 [doi].
14. Adams Hillard PJ (2008) Menstruation in adolescents: what's normal? Medscape J Med 10: 295.
15. Casazza K, Goran MI, Gower BA (2008) Associations among insulin, estrogen, and fat mass gain over the pubertal transition in African-American and European-American girls. J Clin Endocrinol Metab 93: 2610–2615. jc.2007-2776 [pii];10.1210/jc.2007-2776 [doi].
16. Kaplowitz P (2006) Pubertal development in girls: secular trends. Curr Opin Obstet Gynecol 18: 487–491. 10.1097/01.gco.0000242949.02373.09 [doi];00001703-200610000-00003 [pii].

17. Anderson SE, Must A (2005) Interpreting the continued decline in the average age at menarche: results from two nationally representative surveys of U.S. girls studied 10 years apart. J Pediatr 147: 753–760. S0022-3476(05)00682-7 [pii];10.1016/j.jpeds.2005.07.016 [doi].

18. Adair LS, Gordon-Larsen P (2001) Maturational timing and overweight prevalence in US adolescent girls. Am J Public Health 91: 642–644.

19. Ogden CL, Carroll MD, Kit BK, Flegal KM (2012) Prevalence of obesity and trends in body mass index among US children and adolescents, 1999–2010. JAMA 307: 483–490. jama.2012.40 [pii];10.1001/jama.2012.40 [doi].

20. Anderson CA, Zhu G, Falchi M, van den Berg SM, Treloar SA,et al. (2008) A genome-wide linkage scan for age at menarche in three populations of European descent. J Clin Endocrinol Metab 93: 3965–3970. jc.2007–2568 [pii];10.1210/jc.2007–2568 [doi].

21. Pan L, Ober C, Abney M (2007) Heritability estimation of sex-specific effects on human quantitative traits. Genet Epidemiol 31: 338–347. 10.1002/gepi.20214 [doi].

22. Towne B, Czerwinski SA, Demerath EW, Blangero J, Roche AF, Siervogel RM (2005) Heritability of age at menarche in girls from the Fels Longitudinal Study. Am J Phys Anthropol 128: 210–219. 10.1002/ajpa.20106 [doi].

23. Gold EB (2011) The timing of the age at which natural menopause occurs. Obstet Gynecol Clin North Am 38: 425–440. S0889-8545(11)00066-0 [pii];10.1016/j.ogc.2011.05.002 [doi].

24. Reynolds RF, Obermeyer CM (2001) Age at natural menopause in Beirut, Lebanon: the role of reproductive and lifestyle factors. Ann Hum Biol 28: 21–29.

25. Reynolds RF, Obermeyer CM (2003) Correlates of the age at natural menopause in Morocco. Ann Hum Biol 30: 97–108. BLX4XMEWNT9HYFT9 [pii].

26. van Noord PA, Dubas JS, Dorland M, Boersma H, te VE (1997) Age at natural menopause in a population-based screening cohort: the role of menarche, fecundity, and lifestyle factors. Fertil Steril 68: 95–102. S0015028297814823 [pii].

27. Elias SG, van Noord PA, Peeters PH, den T, et al. (2005) Childhood exposure to the 1944–1945 Dutch famine and subsequent female reproductive function. Hum Reprod 20: 2483–2488. dei090 [pii];10.1093/humrep/dei090 [doi].

28. Kinney A, Kline J, Kelly A, Reuss ML, Levin B (2007) Smoking, alcohol and caffeine in relation to ovarian age during the reproductive years. Hum Reprod 22: 1175–1185. del496 [pii];10.1093/humrep/del496 [doi].

29. Snieder H, MacGregor AJ, Spector TD (1998) Genes control the cessation of a woman's reproductive life: a twin study of hysterectomy and age at menopause. J Clin Endocrinol Metab 83: 1875–1880.

30. Kok HS, van Asselt KM, van der Schouw YT, Peeters PH, Wijmenga C (2005) Genetic studies to identify genes underlying menopausal age. Hum Reprod Update 11: 483–493. dmi024 [pii];10.1093/humupd/dmi024 [doi].

31. van Asselt KM, Kok HS, Pearson PL, Dubas JS, Peeters PH, et al. (2004) Heritability of menopausal age in mothers and daughters. Fertil Steril 82: 1348–1351. S0015-0282(04)02243-5 [pii];10.1016/j.fertnstert.2004.04.047 [doi].

32. Cerne JZ, Pohar-Perme M, Cerkovnik P, Gersak K, Novakovic S (2011) Age at menarche and menopause is not associated with two common genetic variants in the methylenetetrahydrofolate reductase (MTHFR) gene. Eur J Contracept Reprod Health Care 16: 241–247. 10.3109/13625187.2011.575481 [doi].

33. Liu P, Lu Y, Recker RR, Deng HW, Dvornyk V (2010) Association analyses suggest multiple interaction effects of the methylenetetrahydrofolate reductase polymorphisms on timing of menarche and natural menopause in white women. Menopause 17: 185–190. 10.1097/gme.0b013e3181aa2597 [doi].

34. Lunetta KL, D'Agostino RB, Sr., Karasik D, Benjamin EJ, Guo CY, et al. (2007) Genetic correlates of longevity and selected age-related phenotypes: a genome-wide association study in the Framingham Study. BMC Med Genet 8 Suppl 1: S13. 1471–2350-8-S1-S13 [pii];10.1186/1471-2350-8-S1-S13 [doi].

35. Elks CE, Perry JR, Sulem P, Chasman DI, Franceschini N, et al. (2010) Thirty new loci for age at menarche identified by a meta-analysis of genome-wide association studies. Nat Genet 42: 1077–1085. ng.714 [pii];10.1038/ng.714 [doi].

36. He C, Kraft P, Chen C, Buring JE, Pare G, et al. (2009) Genome-wide association studies identify loci associated with age at menarche and age at natural menopause. Nat Genet 41: 724–728. ng.385 [pii];10.1038/ng.385 [doi].

37. Perry JR, Stolk L, Franceschini N, Lunetta KL, Zhai G, et al. (2009) Meta-analysis of genome-wide association data identifies two loci influencing age at menarche. Nat Genet 41: 648–650. ng.386 [pii];10.1038/ng.386 [doi].

38. Stolk L, Zhai G, van Meurs JB, Verbiest MM, Visser JA, et al. (2009) Loci at chromosomes 13, 19 and 20 influence age at natural menopause. Nat Genet 41: 645–647. ng.387 [pii];10.1038/ng.387 [doi].

39. Sulem P, Gudbjartsson DF, Rafnar T, Holm H, Olafsdottir EJ, et al. (2009) Genome-wide association study identifies sequence variants on 6q21 associated with age at menarche. Nat Genet 41: 734–738. ng.383 [pii];10.1038/ng.383 [doi].

40. Dvornyk V, Waqar UH (2012) Genetics of age at menarche: a systematic review. Hum Reprod Update. dmr050 [pii];10.1093/humupd/dmr050 [doi].

41. Chen CT, Fernandez-Rhodes L, Brzyski RG, Carlson CS, Chen Z, et al. (2012) Replication of loci influencing ages at menarche and menopause in Hispanic women: the Women's Health Initiative SHARe Study. Hum Mol Genet 21: 1419–1432. ddr570 [pii];10.1093/hmg/ddr570 [doi].

42. Matise TC, Ambite JL, Buyske S, Carlson CS, Cole SA, et al. (2011) The Next PAGE in understanding complex traits: design for the analysis of Population Architecture Using Genetics and Epidemiology (PAGE) Study. Am J Epidemiol 174: 849–859. kwr160 [pii];10.1093/aje/kwr160 [doi].

43. Buyske S, Wu Y, Carty CL, Cheng I, Assimes TL, et al. (2012) Evaluation of the metabochip genotyping array in African Americans and implications for fine mapping of GWAS-identified loci: the PAGE study. PLoS One 7: e35651. 10.1371/journal.pone.0035651 [doi];PONE-D-12-01108 [pii].

44. Voight BF, Kang HM, Ding J, Palmer CD, Sidore C, et al. (2012) The metabochip, a custom genotyping array for genetic studies of metabolic, cardiovascular, and anthropometric traits. PLoS Genet 8: e1002793. 10.1371/journal.pgen.1002793 [doi];PGENETICS-D-11-02644 [pii].

45. The ARIC investigators (1989) The Atherosclerosis Risk in Communities (ARIC) Study: design and objectives. Am J Epidemiol 129: 687–702.

46. The Women's Health Initiative Study Group (1998) Design of the Women's Health Initiative clinical trial and observational study. Control Clin Trials 19: 61–109. S0197245697000780 [pii].

47. Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, et al. (2006) Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 38: 904–909. ng1847 [pii];10.1038/ng1847 [doi].

48. Must A, Phillips SM, Naumova EN, Blum M, Harris S, et al. (2002) Recall of early menstrual history and menarcheal body size: after 30 years, how well do women remember? Am J Epidemiol 155: 672–679.

49. Willer CJ, Li Y, Abecasis GR (2010) METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics 26: 2190–2191. btq340 [pii];10.1093/bioinformatics/btq340 [doi].

50. Pruim RJeal (2010) LocusZoom: regional visualization of genome-wide association scan results. 26.

51. R Development Core Team (2012) R: A language and environment for statistical computing., version R Foundation for Statistical Computing.

52. Gauderman WJ (2002) Sample size requirements for association studies of gene-gene interaction. Am J Epidemiol 155: 478–484.

53. Johnson AD, Handsaker RE, Pulit SL, Nizzari MM, O'Donnell CJ, et al. (2008) SNAP: a web-based tool for identification and annotation of proxy SNPs using HapMap. Bioinformatics 24: 2938–2939. btn564 [pii];10.1093/bioinformatics/btn564 [doi].

54. Guo Y, Xiong DH, Yang TL, Guo YF, Recker RR, et al. (2006) Polymorphisms of estrogen-biosynthesis genes CYP17 and CYP19 may influence age at menarche: a genetic association study in Caucasian females. Hum Mol Genet 15: 2401–2408. ddl155 [pii];10.1093/hmg/ddl155 [doi].

55. Mitchell ES, Farin FM, Stapleton PL, Tsai JM, Tao EY, et al. (2008) Association of estrogen-related polymorphisms with age at menarche, age at final menstrual period, and stages of the menopausal transition. Menopause 15: 105–111. 10.1097/gme.0b013e31804d2406 [doi].

56. Zhao J, Xiong DH, Guo Y, Yang TL, Recker RR, et al. (2007) Polymorphism in the insulin-like growth factor 1 gene is associated with age at menarche in caucasian females. Hum Reprod 22: 1789–1794. dem052 [pii];10.1093/humrep/dem052 [doi].

57. Ong KK, Elks CE, Li S, Zhao JH, Luan J, et al. (2009) Genetic variation in LIN28B is associated with the timing of puberty. Nat Genet 41: 729–733. ng.382 [pii];10.1038/ng.382 [doi].

58. Lu Y, Liu P, Recker RR, Deng HW, Dvornyk V (2010) TNFRSF11A and TNFSF11 are associated with age at menarche and natural menopause in white women. Menopause 17: 1048–1054. 10.1097/gme.0b013e3181d5d523 [doi].

59. He C, Kraft P, Chasman DI, Buring JE, Chen C, et al. (2010) A large-scale candidate gene association study of age at menarche and age at natural menopause. Hum Genet 128: 515–527. 10.1007/s00439-010-0878-4 [doi].

60. Mitchell ES, Farin FM, Stapleton PL, Tsai JM, Tao EY, et al. (2008) Association of estrogen-related polymorphisms with age at menarche, age at final menstrual period, and stages of the menopausal transition. Menopause 15: 105–111. 10.1097/gme.0b013e31804d2406 [doi].

61. He LN, Xiong DH, Liu YJ, Zhang F, Recker RR, et al. (2007) Association study of the oestrogen signalling pathway genes in relation to age at natural menopause. J Genet 86: 269–276.

62. Murray A, Bennett CE, Perry JR, Weedon MN, Jacobs PA, et al. (2011) Common genetic variants are significant risk factors for early menopause: results from the Breakthrough Generations Study. Hum Mol Genet 20: 186–192. ddq417 [pii];10.1093/hmg/ddq417 [doi].

63. Meng FT, Wang YL, Liu J, Zhao J, Liu RY, et al. (2011) ApoE genotypes are associated with age at natural menopause in Chinese females. Age (Dordr). 10.1007/s11357-011-9287-4 [doi].

64. Long JR, Shu XO, Cai Q, Cai H, Gao YT, et al. (2006) Polymorphisms of the CYP1B1 gene may be associated with the onset of natural menopause in Chinese women. Maturitas 55: 238–246. S0378-5122(06)00096-X [pii];10.1016/j.maturitas.2006.03.005 [doi].

65. Voorhuis M, Broekmans FJ, Fauser BC, Onland-Moret NC, van der Schouw YT (2011) Genes involved in initial follicle recruitment may be associated with age at menopause. J Clin Endocrinol Metab 96: E473–E479. jc.2010-1799 [pii];10.1210/jc.2010-1799 [doi].

66. Stolk L, Perry JR, Chasman DI, He C, Mangino M, et al. (2012) Meta-analyses identify 13 loci associated with age at menopause and highlight DNA repair and immune pathways. Nat Genet. ng.1051 [pii];10.1038/ng.1051 [doi].

67. Maeda D, Chen X, Guan B, Nakagawa S, Yano T, et al. (2011) Rsf-1 (HBXAP) expression is associated with advanced stage and lymph node metastasis in ovarian clear cell carcinoma. Int J Gynecol Pathol 30: 30–35. 10.1097/PGP.0b013e3181e9a319 [doi].

68. Choi JH, Sheu JJ, Guan B, Jinawath N, Markowski P, et al. (2009) Functional analysis of 11q13.5 amplicon identifies Rsf-1 (HBXAP) as a gene involved in paclitaxel resistance in ovarian cancer. Cancer Res 69: 1407–1415. 0008-5472.CAN-08-3602 [pii];10.1158/0008-5472.CAN-08-3602 [doi].

69. Brown LA, Kalloger SE, Miller MA, Shih I, McKinney SE, et al. (2008) Amplification of 11q13 in ovarian carcinoma. Genes Chromosomes Cancer 47: 481–489. 10.1002/gcc.20549 [doi].

70. Diakou M, Miltiadous G, Xenophontos SL, Manoli P, Cariolou MA, et al. (2011) Spectrum of LDLR gene mutations, including a novel mutation causing familial hypercholesterolaemia, in North-western Greece. Eur J Intern Med 22: e55–e59. S0953-6205(11)00008-2 [pii];10.1016/j.ejim.2011.01.003 [doi].

71. De Castro-Oros I, Pampin S, Bolado-Carrancio A, De CA, Palacios L, et al. (2011) Functional analysis of LDLR promoter and 5′ UTR mutations in subjects with clinical diagnosis of familial hypercholesterolemia. Hum Mutat 32: 868–872. 10.1002/humu.21520 [doi].

72. Dvornyk V, Waqar UH (2012) Genetics of age at menarche: a systematic review. Hum Reprod Update. dmr050 [pii];10.1093/humupd/dmr050 [doi].

73. Milne FH, Judge DS, Preen DB, Weinstein P (2011) Early life environment, life history and risk of endometrial cancer. Med Hypotheses 77: 626–632. S0306-9877(11)00318-5 [pii];10.1016/j.mehy.2011.07.001 [doi].

74. Opdahl S, Alsaker MD, Janszky I, Romundstad PR, Vatten LJ (2011) Joint effects of nulliparity and other breast cancer risk factors. Br J Cancer 105: 731–736. bjc2011286 [pii];10.1038/bjc.2011.286 [doi].

75. Narod SA (2011) Alcohol and risk of breast cancer. JAMA 306: 1920–1921. 306/17/1920 [pii];10.1001/jama.2011.1589 [doi].

76. Narod SA (2011) Early-onset breast cancer: what do we know about the risk factors?: A Countercurrents Series. Curr Oncol 18: 204–205.

77. Jasen P (2011) Menopause and historical constructions of cancer risk. Can Bull Med Hist 28: 43–70.

78. Dishi M, Enquobahrie DA, Abetew DF, Qiu C, Rudra CB, et al. (2011) Age at menarche, menstrual cycle characteristics and risk of gestational diabetes. Diabetes Res Clin Pract 93: 437–442. S0168-8227(11)00351-2 [pii];10.1016/j.diabres.2011.07.001 [doi].

79. Campbell Jenkins BW, Addison C, Wilson G, Liu J, Fortune M, et al. (2011) Association of the joint effect of menopause and hormone replacement therapy and cancer in African american women: the Jackson Heart Study. Int J Environ Res Public Health 8: 2491–2504. 10.3390/ijerph8062491 [doi];ijerph-08-02491 [pii].

80. Kallen AN, Pal L (2011) Cardiovascular disease and ovarian function. Curr Opin Obstet Gynecol 23: 258–267. 10.1097/GCO.0b013e3283488a21 [doi].

81. Cui B, Zhu X, Xu M, Guo T, Zhu D, et al. (2011) A genome-wide association study confirms previously reported loci for type 2 diabetes in Han Chinese. PLoS One 6: e22353. 10.1371/journal.pone.0022353 [doi];PONE-D-11-06453 [pii].

82. Saif-Ali R, Ismail IS, Al-Hamodi Z, Al-Mekhlafi HM, Siang LC, et al. (2011) KCNQ1 Haplotypes Associate with Type 2 Diabetes in Malaysian Chinese Subjects. Int J Mol Sci 12: 5705–5718. 10.3390/ijms12095705 [doi];ijms-12-05705 [pii].

83. Saif-Ali R, Muniandy S, Al-Hamodi Z, Lee CS, Ahmed KA, et al. (2011) KCNQ1 variants associate with type 2 diabetes in Malaysian Malay subjects. Ann Acad Med Singapore 40: 488–492.

84. Rees SD, Hydrie MZ, Shera AS, Kumar S, O'Hare JP, et al. (2011) Replication of 13 genome-wide association (GWA)-validated risk variants for type 2 diabetes in Pakistani populations. Diabetologia 54: 1368–1374. 10.1007/s00125-011-2063-2 [doi].

85. Yasuda K, Miyake K, Horikawa Y, Hara K, Osawa H, et al. (2008) Variants in KCNQ1 are associated with susceptibility to type 2 diabetes mellitus. Nat Genet 40: 1092–1097. ng.207 [pii];10.1038/ng.207 [doi].

86. Unoki H, Takahashi A, Kawaguchi T, Hara K, Horikoshi M, et al. (2008) SNPs in KCNQ1 are associated with susceptibility to type 2 diabetes in East Asian and European populations. Nat Genet 40: 1098–1102. ng.208 [pii];10.1038/ng.208 [doi].

87. Buber J, Mathew J, Moss AJ, Hall WJ, Barsheshet A, et al. (2011) Risk of recurrent cardiac events after onset of menopause in women with congenital long-QT syndrome types 1 and 2. Circulation 123: 2784–2791. CIRCULA-TIONAHA.110.000620 [pii];10.1161/CIRCULATIONAHA.110.000620 [doi].

88. Jurvansuu JM, Goldman A (2011) Obesity risk gene TMEM18 encodes a sequence-specific DNA-binding protein. PLoS One 6: e25317. 10.1371/journal.pone.0025317 [doi];PONE-D-11-12025 [pii].

89. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, et al. (2010) Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet 42: 937–948. ng.686 [pii];10.1038/ng.686 [doi].

90. Carr MC (2003) The emergence of the metabolic syndrome with menopause. J Clin Endocrinol Metab 88: 2404–2411.

91. Salpeter SR, Walsh JM, Greyber E, Salpeter EE (2006) Brief report: Coronary heart disease events associated with hormone therapy in younger and older women. A meta-analysis. J Gen Intern Med 21: 363–366. JGI389 [pii];10.1111/j.1525-1497.2006.00389.x [doi].

92. Schunkert H, Konig IR, Kathiresan S, Reilly MP, Assimes TL, et al. (2011) Large-scale association analysis identifies 13 new susceptibility loci for coronary artery disease. Nat Genet 43: 333–338. ng.784 [pii];10.1038/ng.784 [doi].

93. Ripatti S, Tikkanen E, Orho-Melander M, Havulinna AS, Silander K, et al. (2010) A multilocus genetic risk score for coronary heart disease: case-control and prospective cohort analyses. Lancet 376: 1393–1400. S0140-6736(10)61267-6 [pii];10.1016/S0140-6736(10)61267-6 [doi].

94. Ehret GB, Munroe PB, Rice KM, Bochud M, Johnson AD, et al. (2011) Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. Nature 478: 103–109. nature10405 [pii];10.1038/nature10405 [doi].

95. Hofbauer LC, Schoppet M (2004) Clinical implications of the osteoprotegerin/RANKL/RANK system for bone and vascular diseases. JAMA 292: 490–495. 10.1001/jama.292.4.490 [doi];292/4/490 [pii].

96. Boyce BF, Xing L (2008) Functions of RANKL/RANK/OPG in bone modeling and remodeling. Arch Biochem Biophys 473: 139–146. S0003-9861(08)00159-8 [pii];10.1016/j.abb.2008.03.018 [doi].

97. He LN, Recker RR, Deng HW, Dvornyk V (2009) A polymorphism of apolipoprotein E (APOE) gene is associated with age at natural menopause in Caucasian females. Maturitas 62: 37–41. S0378-5122(08)00266-1 [pii];10.1016/j.maturitas.2008.10.011 [doi].

98. Meng FT, Wang YL, Liu J, Zhao J, Liu RY, et al. (2011) ApoE genotypes are associated with age at natural menopause in Chinese females. Age (Dordr). 10.1007/s11357-011-9287-4 [doi].