

**THE ASSOCIATION BETWEEN GENETIC VARIANTS, BODY COMPOSITION AND
BLOOD PRESSURE IN AFRO-CARIBBEAN MEN FROM TOBAGO**

by

Tracey Samantha Beason

BS, Oakwood University, 2001

MSPH, Meharry Medical College, 2003

Submitted to the Graduate Faculty of
Graduate School of Public Health in partial fulfillment
of the requirements for the degree of
Doctor of Philosophy

University of Pittsburgh

2010

UNIVERSITY OF PITTSBURGH

Graduate School of Public Health

This dissertation was presented

by

Tracey Samantha Beason

It was defended on

April 22, 2010

and approved by

John W. Wilson, Ph.D., Assistant Professor of Biostatistics, Graduate School of Public
Health, University of Pittsburgh

Clareann H. Bunker, Ph.D., Associate Professor of Epidemiology, Graduate School of Public
Health, University of Pittsburgh

Joseph M. Zmuda, Ph.D., Associate Professor of Epidemiology, Assistant Professor of
Human Genetics, Graduate School of Public Health, University of Pittsburgh

Dissertation Advisor: Joel L. Weissfeld, M.D., M.P.H, Associate Professor of Epidemiology,
Graduate School of Public Health, University of Pittsburgh

Copyright © by Tracey Samantha Beason

2010

**THE ASSOCIATION BETWEEN GENETIC VARIANTS, BODY COMPOSITION AND
BLOOD PRESSURE IN AFRO-CARIBBEAN MEN FROM TOBAGO**

Tracey Samantha Beason, PhD

University of Pittsburgh, 2010

Prostate cancer is one of the most common malignancies affecting blacks worldwide. Blacks have a higher incidence and prevalence of prostate cancer than Caucasians. Treatment for prostate cancer usually involves androgen deprivation therapy. Even though androgen deprivation therapy has a high efficacy there are many deleterious side effects such as increase in body fat. Consequently, we investigated the association between androgen deprivation therapy for prostate cancer and the rate of change of percent total body fat in a cohort of Afro-Tobagonian men. Case-control analysis of 1691 men in our study indicated that, men with prostate cancer exposed to ADT had higher increases in percent total body fat over time compared to unexposed men with prostate cancer.

Likewise, obesity and hypertension disproportionately affects individuals of African descent. These complex diseases are multi-factorial in origin and are typically controlled by many genes. $ADRB_2$ and AR CAG repeat lengths have been widely studied in the literature and these genotypes were available for study in the Tobago Prostate Study; consequently, we investigated the association between $ADRB_2$, body composition and blood pressure. We also investigated the association between AR CAG repeats and body composition.

Cross-sectional analysis of 2,584 men in our cohort indicated that, men with shorter AR CAG repeats had higher body fat measurements (DEXA). We found that the direction of the association was opposite what we had anticipated, of men with longer AR CAG repeats having

higher body fat measurements. 1,965 men were investigated cross-sectionally to determine the association between $ADRB_2$, body composition and blood pressure. We found no association between $ADRB_2$, DEXA measures of body fat and blood pressure.

In our cohort of Afro-Tobagonian men *AR* CAG repeats and $ADRB_2$ did not show an association with our outcome of interest. Even though we had non-significant findings, other genes should be evaluated to assess if an association exists with body composition and blood pressure in Tobago population. These studies are of public health relevance or importance because they contribute epidemiologic information to the body of scientific information available especially regarding Tobago men.

TABLE OF CONTENTS

PREFACE.....	XV
1.0 INTRODUCTION.....	1
1.1 SPECIFIC AIMS.....	3
1.1.1 Project 1:β₂-adrenergic receptor (<i>ADRB2</i>) genotype, body composition, and blood pressure in African-Caribbean men from Tobago	3
1.1.2 Project 2: Androgen receptor (<i>AR</i>) genotype and body composition in African-Caribbean men from Tobago	3
1.1.3 Project 3: To examine the association between androgen deprivation therapy (ADT) for prostate cancer and body composition	4
1.2 BACKGROUND.....	4
1.2.1 Epidemiology	4
1.2.2 Anatomy of the Prostate	6
1.2.3 Pathogenesis of the Prostate.....	6
1.2.3.1 Risk factors.....	7
1.2.4 Clinical risk factors for prostate cancer.....	10
1.2.5 Androgens	12
1.2.5.1 Racial differences in sex steroid hormones	13
1.2.5.2 Definition of maleness and androgenicity.....	15

1.2.5.3	Androgen metabolism in the prostate.....	16
1.2.5.4	Serum testosterone, obesity, age, and tumor grade.....	17
1.2.5.5	Androgen deprivation	18
1.2.6	Treatment of prostate cancer	19
1.2.6.1	Orchiectomy	19
1.2.6.2	Prostatectomy	20
1.2.6.3	LHRH agonists.....	20
1.2.6.4	Antiandrogens.....	20
1.2.6.5	Chemotherapy for hormone resistant cancer	21
1.2.6.6	Untreated prostate cancer.....	21
1.2.7	ADT and body composition.....	22
1.2.7.1	ADT and bone loss	23
1.2.7.2	Definition of osteoporosis.....	24
1.2.7.3	Osteoporosis gene polymorphism.....	25
1.2.7.4	Heritability of bone mass	25
1.2.7.5	Racial differences with BMD.....	26
1.2.8	Androgen receptor (<i>AR</i>)	28
1.2.8.1	Ethnic differences in <i>AR</i> CAG repeat length	29
1.2.8.2	<i>AR</i> CAG repeat length and BMD	29
1.2.8.3	<i>AR</i> CAG repeat length and body composition	30
1.2.8.4	β 2Adrenergic Receptor	31
1.2.8.5	β 2 adrenergic receptor and hypertension.....	32
1.2.8.6	β ₂ adrenergic receptor and body composition	33

1.2.9	Obesity in the Caribbean.....	34
1.2.9.1	Causes of weight gain	35
1.2.9.2	Energy output: physical activity.....	35
1.2.10	Distribution of body fat	36
1.2.10.1	Subcutaneous and visceral fat	36
1.2.10.2	Intramuscular fat.....	37
1.2.10.3	Waist circumference (WC) and waist to hip ratio (WHR)	38
1.2.10.4	Body mass index (BMI).....	38
1.2.10.5	Visceral fat as a pro-inflammatory tissue.....	39
1.2.10.6	Race ethnicity and body fat distribution	40
1.2.11	The metabolic syndrome.....	40
1.2.11.1	The impact of ethnicity on the metabolic syndrome.....	41
1.2.11.2	Prevalence of the metabolic syndrome	42
1.2.11.3	Lifestyle modification	43
1.2.12	Measurement of body fat distribution	44
1.2.12.1	Dual X-ray absorptiometry (DEXA).....	44
1.2.12.2	Computed tomography (CT)	45
1.2.12.3	Quantitative computerized tomography (QCT)	45
1.2.12.4	Peripheral quantitative computerized tomography (pQCT).....	45
1.2.13	Diabetes	46
1.2.13.1	Prevalence of diabetes in the United States.....	46
1.2.13.2	Pathogenesis of diabetes	46
1.2.13.3	Obesity and insulin resistance	48

1.2.13.4	Adipocytes, insulin resistance and diabetes.....	48
1.2.13.5	Insulin resistance	49
1.2.13.6	Diabetes in the Caribbean.....	49
1.2.13.7	Economic burden of diabetes.....	50
1.2.13.8	Under report of diabetes in the Caribbean	50
1.2.14	Hypertension.....	51
1.2.14.1	Definition of hypertension.....	52
1.2.14.2	Characteristics of severe hypertension	53
1.2.14.3	Obesity and Hypertension.....	53
1.2.14.4	Environment.....	54
1.2.14.5	Nutrition	54
1.2.14.6	Pathophysiology of hypertension.....	55
1.2.14.7	Reninangiotensin system	56
1.2.14.8	Autonomic nervous system	57
1.2.14.9	Cardiac output and peripheral resistance.....	57
1.2.14.10	Sympathetic over-activity	57
1.2.14.11	Oxidative stress and endothelial dysfunction.....	58
1.2.14.12	Inflammatory mediators	59
1.2.15	Hypertension prevalence in Afro-Caribbeans.....	60
1.2.16	Prevalence of hypertension in Trinidad.....	61
2.0	PUBLIC HEALTH SIGNIFICANCE.....	62
3.0	SYNTHESIS STATEMENT	64

4.0 PAPER 1: ADRB2 GENE VARIANTS, DEXA BODY COMPOSITION, AND HYPERTENSION IN TOBAGO MEN OF AFRICAN DESCENT	65
4.1 ABSTRACT	66
4.2 INTRODUCTION	67
4.3 METHODS AND MATERIAL	68
4.3.1 Outcome measurements.....	70
4.3.2 Statistical analysis	70
4.4 RESULTS.....	71
4.5 DISCUSSION.....	73
4.6 REFERENCES	78
5.0 PAPER 2: ANDROGEN RECEPTOR ARCAG REPEAT LENGTH POLYMORPHISM AND OBESITY IN AFRO-CARIBBEAN MEN: RESULTS FROM THE TOBAGO PROSTATE STUDY.	86
5.1 ABSTRACT	87
5.2 INTRODUCTION	88
5.3 MATERIALS AND METHODS.....	89
5.3.1 Outcome measurements.....	90
5.3.2 Genotyping.....	91
5.3.3 Statistical analyses.....	91
5.4 RESULTS.....	92
5.5 DISCUSSION.....	94
5.5.1 Racial differences in CAG repeat length	95

6.0 PAPER 3: EFFECTS OF ANDROGEN DEPRIVATION THERAPY ON CHANGE IN BODY COMPOSITION IN AFRO-CARIBBEAN MEN: TOBAGO PROSTATE STUDY.....	107
6.1 ABSTRACT	108
6.2 INTRODUCTION	109
6.3 MATERIALS AND METHODS.....	110
6.3.1 Study sample.....	110
6.3.2 Outcome measurements.....	111
6.3.3 Statistical Analyses.....	112
6.4 RESULTS.....	112
6.5 DISCUSSION.....	114
BIBLIOGRAPHY.....	123

LIST OF TABLES

Table 4-1: Body composition measures (means \pm standard deviations) according to <i>ADRB2</i> genotype.....	80
Table 4-2: <i>ADRB2</i> genotype and body composition.....	81
Table 4-3: <i>ADRB2</i> genotype and hypertension.....	82
Table 4-4: Estimated difference (Δ ; and 95% confidence interval) in body composition measure between men homozygous for a specified haplotypelrelative to men homozygous for the Arg-Glnhaplotype.	83
Table 5-1: Body composition measures (mean \pm standard deviations) according to <i>AR</i> CAG genotype.....	98
Table 5-2:Proportion of men with long vs. short <i>AR</i> CAG repeats (N=2584).	98
Table 5-3: Potential confounders.....	99
Table 5-4: Spearman rank correlations between <i>AR</i> CAG repeats and body composition measures.....	100
Table 5-5: Results from standardized univariate linear regression models (n=2584).	101
Table 5-6:A comparison of <i>AR</i> CAGlength between excludedmen (men excluded from analysis because of missing DEXA and missing HH questionnaire) and included men (men included in analysis of association between <i>AR</i> CAG and body composition).....	102

Table 5-7: Unadjusted <i>AR</i> CAG repeat length category (≤ 22 vs. > 22 repeats) associations with age, weight, body mass index (BMI), percent total body fat, percent trunk fat, percent arm fat, percent leg fat, percent limb fat (n=2584).	102
Table 5-8: Association between <i>AR</i> CAG repeat and percent total body fat, by age.	103
Table 5-9: Association between <i>AR</i> CAG repeat and percent total body fat, by BMI category.	103
Table 5-10: Age-adjusted associations between <i>AR</i> CAG repeat length category (≤ 22 vs. > 22 repeats) and eight different measures of body composition (n=2584).	104
Table 5-11: <i>AR</i> CAG repeat length associations with age, BMI, and percent total body fat, in study groups defined differently with respect to the availability of BMI and percent total body fat information.....	105
Table 6-1: Baseline body composition characteristics (mean \pm S.D.).....	118
Table 6-2: Change in percent total body fat (Δ TBFat%) by prostate cancer status and androgen deprivation exposure.	119
Table 6-3: Mean rate of change of percent total body fat (Δ TBFat%) by prostate cancer and androgen deprivation.	120
Table 6-4: Tobago Prostate Study, subject mean age, median interval between sequential DEXA scans, median interval between prostate cancer diagnosis and first DEXA, and median interval between prostate cancer diagnosis and second DEXA, according to prostate cancer status, timing of prostate cancer diagnosis with respect to sequential DEXA, and prostate cancer treatment status (ADT vs. no ADT).....	121
Table 6-5: Type of androgen deprivation, in men reporting an ADT exposure.	122

LIST OF FIGURES

Figure 4-1: Box plots showing body mass index (BMI; upper figure) and DEXA-determined percent body fat (lower figure) in men sub-group according to diplotype.....	84
Figure 5-1: Sample selection for paper 2.....	96
Figure 5-2: Distribution of <i>AR</i> CAG repeat length.....	97
Figure 6-1: Sample selection for Paper 3.....	117

PREFACE

I would like to thank the staff at the Tobago Health Studies Office in Scarborough, Tobago for collecting and providing the information used in my dissertation. I would also like to thank my PhD committee, my family (especially my mother Barbara Francis and Dad Valbert Francis), and friends for their encouragement and support throughout the completion of my Ph.D.

I would also like to say Special thanks to Dr. Zmuda and Dr. Bunker for allowing me to use their dataset and for guiding me through this process. Dr. John Wilson has been a constant source of encouragement; special thanks to Dr. Wilson for his patience, listening ear and training.

I would also like to give special thanks to my mentor and advisor Dr. Joel L. Weissfeld for his continued support, encouragement and training. Dr. Weissfeld, thank you so much for your support and guidance. I would not have completed my doctoral education at the University of Pittsburgh without your help. I want you to know that you have inspired me to prove that I am capable of becoming anything I aspire to be. Thank you for being a positive force in my life, it has made a difference in my experience at the University. I do appreciate the time and effort you spent teaching me.

1.0 INTRODUCTION

Prostate cancer is considered one of the most common carcinomas and is the leading cause of cancer death in American men. African American men have the greatest risk for the disease and there is some suggestion that there might be some ethnic variation in prostate cancer risk.^{1, 2} Androgen deprivation therapy with GnRH is the most common treatment for metastatic prostate cancer. In addition, androgens are important determinants of body composition in men. Serum Testosterone concentrations have been shown to correlate positively with muscle mass and negatively with fat mass.¹ GnRH agonist treatment in men has been shown to reduce bone mineral density, which is an important determinant of fracture risk. Furthermore, the changes in body composition may contribute to the adverse effects of physical function and potentially their quality of life.¹

Lean mass, fat mass and body weight have been shown to be associated with high bone mineral density (BMD). Several studies indicate that a decline in lean body mass, or body weight is associated with declines in BMD.^{3, 4} It is hypothesized that sex steroid hormone levels (testosterone and estradiol) contribute to low BMD and bone loss in older men and vary according to ethnicities.⁵ Variations in BMD are mostly genetic but other factors such as environment, nutrition and physical activity can play an important role⁶. There is still uncertainty regarding the independent risk factors for BMD in men and women. Many of the studies related to BMD have been conducted in women because of bone loss following

menopause¹⁷ Despite lower osteoporotic risk in African Americans the number of individuals afflicted by this disease is expected to increase exponentially in the next few years.

High blood pressure has been shown to be relatively common in the West Indies⁸. Patrick et al.⁹ showed that Tobagonian's have higher blood pressure levels, body bulk and obesity compared to mixed African populations of Jamaica, Guyana, and the European Caucasian populations of rural Wales.⁹In Tobago (2006), hypertension/high blood pressure and its co-morbidities account for approximately 35% of all yearly visits to health care facilities and is considered a significant contributor to morbidity and mortality.¹⁰

The literature is replete with evidence that genes influence obesity, and body fat distribution⁴. The development of many metabolic diseases such as CVD, prostate cancer, high blood pressure and T2DM is influenced by lifestyle and also genes. Obesity and insulin resistance are key risk factors for these diseases. Metabolic diseases such as T2DM are associated with aberrations in the lipid metabolism that typically cause fat accumulation in the liver and muscle.¹¹ Even though many of these studies have indicated a genetic basis for these diseases, many of the variations in these diseases are due to genes that are yet unidentified. Furthermore there is little information in the literature regarding genetic factors influencing body fat, and body fat distribution in populations of African descent.

Tobago would serve as an important population to investigate the impact genetics has on various diseases. Since Tobago is comprised mainly of West African ancestry with miniscule European and Native American admixture, the Tobago population may carry a higher prevalence of high risk alleles of African origin compared to more admixed populations.¹² The objective of this study is to determine the association between CAG AR repeat length polymorphism(Xq11-12), androgen deprivation therapy (ADT), β_2 -adrenergic receptor polymorphism (5q33.1) and

high blood pressure, body composition, and low bone mineral density in a subset of Tobagonian men in the Tobago Bone Follow-up Study.

1.1 SPECIFIC AIMS

1.1.1 Project 1: β_2 -adrenergic receptor (*ADRB2*) genotype, body composition, and blood pressure in African-Caribbean men from Tobago

The Tobago Prostate Cancer Study determined *ADRB2* genotype (single nucleotide polymorphism (SNP) rs1042714), performed dual energy X-ray absorptiometry (DEXA), and measured height, weight, waist circumference, and blood pressure in N=1965 Afro-Caribbean men from Tobago. Research questions include 1) the association between *ADRB2* rs1042714 genotype and obesity and 2) the association between *ADRB2* rs1042714 genotype and blood pressure. Research hypotheses include,

1. Men with one or two copies of a variant *ADRB2* rs1042714 allele have more body fat, expressed as DEXA measured total body fat as a percentage of total body mass, and
2. Men with one or two copies of a variant *ADRB2* rs1042714 allele have higher blood pressure.

1.1.2 Project 2: Androgen receptor (*AR*) genotype and body composition in African-Caribbean men from Tobago

The Tobago Prostate Cancer Study determined *AR* genotype (CAG trinucleotide repeat polymorphism length) and performed dual energy X-ray absorptiometry in N=2,584 Afro-

Caribbean men from Tobago. Research questions include 1) the association between *AR* genotype (expressed as the number of CAG repeats) and obesity. Research hypotheses include,

1. Men with longer *AR* alleles (more CAG repeats) have more body fat, expressed as DEXA measured total body fat as a percentage of total body mass, and

1.1.3 Project 3: To examine the association between androgen deprivation therapy (ADT) for prostate cancer and body composition

The Tobago Prostate Cancer Study performed dual energy X-ray absorptiometry in greater than equal to 40 year-old Afro-Caribbean men from Tobago, before and after androgen deprivation therapy (ADT) for prostate cancer. Research questions include 1) the association between androgen deprivation therapy and temporal change in body composition. Research hypotheses include,

1. ADT-treated prostate cancer men will experience a gain in DEXA body fat mass (expressed as a percentage of DEXA total body mass) more rapidly than ADT-untreated prostate cancer men and men without prostate cancer

1.2 BACKGROUND

1.2.1 Epidemiology

Prostate cancer is the second leading cause of death in American men. It is estimated 186, 320 men will be diagnosed with prostate cancer and 28, 8660 men will die of cancer of the prostate in 2008. The age adjusted incidence rate is 163.0 per 100, 000 per year and the age adjusted death

rate is 26.7 per 100, 000 men per year. This data is based on diagnosed cases /patients who died in 2001-2005 from 17 SEER locations.¹³

In men with prostate cancer androgen deprivation therapy (ADT) is being widely used in patients with prostate carcinoma. Even though, ADT therapy is effective in controlling prostate cancer, hypogonadism as a result of ADT treatment is associated with many adverse effects for instance: impotence, hot flashes, cardiovascular morbidity, and osteoporosis¹⁴. Osteoporosis is no longer a disease that only affects women. There is growing concern, regarding men with osteoporosis. In the United States there is approximately 1.5 million men >65 with osteoporosis and another 3.5 million men with osteopenia (low bone density), potentially increasing their risk of fractures.¹⁵

In the literature there is limited data showing that men with prostate cancer could have preexisting low BMD¹⁶⁻¹⁹ and also showing that ADT causes additional decreases in BMD in men.^{17, 18, 20, 21} In addition to the loss in BMD another problem with ADT is the change in body composition. Changes in body composition are adverse side effects of androgen deprivation.¹⁹

Further more, studies show that body composition is related to other diseases such as diabetes and hypertension^{22, 23} an increase in body mass and decreases in muscle mass is typically a result of reduced androgen levels in men.²⁴⁻²⁷ It is hypothesized that reduced muscle mass will lead to decreased muscle strength, which would increase the risk of bone fractures in men. Even though, the literature is replete with studies emphasizing the effects of hypogonadism on body composition²⁸ research on body composition among men with prostate cancer after receiving ADT therapy is rarely discussed.¹⁴

1.2.2 Anatomy of the Prostate

The prostate gland is an exocrine organ that measures 25cc and is located in the pelvis between the bladder and the urinary sphincter, anterior to the rectum and below the pubis. The prostate is comprised of lobular tubuloalveolar glands that secrete fluids into the prostatic urethra. This fluid is comprised of seminal emissions that are rich in prostate-specific-antigen (PSA). Because of the location of the prostate the treatment of prostate cancer can have significant effects on urinary, sexual and bowel function.²⁹

Most cancers of the prostate seem to develop near the capsule of the prostate in the peripheral zone. Prostate cancer is considered a multifocal disease in which tumors are present throughout the gland. The spread of prostate cancer can occur through defects in the capsule in which the neurovascular structures and the ejaculatory ducts enter the gland or possibly in the region of the bladder neck. The cancer can also progress to the seminal vesicles, bladder neck or the levator muscles. Systemic spread of the disease can also occur through the lymphatics to the external iliac, hypogastric, obturator, and presacral nodes. Prostate cancer has also been shown to have a predilection towards bone because of the bidirectional interaction between tumor cells and the surrounding stroma.²⁹

A number of hereditary and environmental risk factors have been hypothesized in the risk of prostate cancer. These risk factors include, age race and family history which are the generally accepted risk factors in the development of prostate cancer.³⁰

1.2.3 Pathogenesis of the Prostate

Prostate cancer is a heterogeneous disease that is caused by a series of genetic events in the prostate. These genetic events along with environmental risk factors can promote the development of cancer of the prostate.^{31, 32}

1.2.3.1 Risk factors

Increasing Age

Aside from being of male sex age is considered one of the most important risk factors for prostate cancer. Prostate cancer is an age-related adult malignancy. Prostate cancer rarely occurs before the age of 40 years but the incidence seems to rapidly increase there after.³² Bunker et al.,³³ found that prostate cancer rates increased with increasing age in the Tobago population with the highest rates being detected between the ages of 60-69 years (18%) and 70-79 years (28%).

Race and ethnicity

Men of African descent have the highest prostate cancer morbidity and compared to other racial or ethnic groups.^{34, 35} Even though the overall morbidity and mortality from prostate cancer has been declining in whites since 1991 possibly due to improvements in diagnosis techniques, screening, surgical and radiologic treatments the rates in African American men are still high; approximately 2.4 times higher than in White men.³⁴ Furthermore, within the last couple of years, prostate cancer has emerged as the most common cancer in African American men and is considered the second most common cause of cancer related deaths in the western world.^{36, 37}

Black men of African descent have high incidence of prostate cancer and men residing in some regions of Caribbean have rates similar to that of US black men. Glover et al.³⁸ provided some significant data, as prior to his study African American men in the United States were considered to have the highest rates of prostate cancer. The international agency for research in cancer (IARC) in the GLOBOCAN software and database had estimated the incidence rate of prostate cancer in Jamaica to be 42.4 per 100,000³⁹. However, Glovers' retrospective study of

1,000 prostate cancer cases in Kingston between the years of 1989-1994 (by evaluating Jamaican Cancer Registry, hospital, and clinic records) showed that the true prostate cancer incidence in Kingston, Jamaica was 304 per 100,000 which is significantly higher than US blacks and is considered to be the highest incidence in the world. The findings from Glover et al.³⁸ of high rates of prostate cancer was further supported by Brooks and Wolf⁴⁰ who found that prostate cancer is the most prevalent cancer in Jamaican men in St. Andrew and Kingston between the years of 1958-1987. Moreover, in 2001 Hanchard et al.⁴¹ reported that prostate cancer is the main site of cancer in Kingston and St. Andrew based on 1,941 malignant neoplasms.

Bunker et al.^{33, 35, 42} reported on information in three studies relevant to Trinidad and Tobago. In that population based prostate cancer study of men 40-79 years screening for prostate cancer was determined via serum prostate-specific antigen (PSA) and digital rectal exam (DRE). This study determined prostate cancer prevalence to be 11% i.e. (277 cases out of 2,583 men)⁴². These findings were further supported by an earlier study by Bunker et al.,³³ in the same population. In 2002 another study by Bunker et al.³⁵ compared prostate cancer prevalence in men of West African descent residing in Tobago and Asian-Indian men in Trinidad between the ages of 50-64 years. Afro-Tobagonian men had higher prevalence of prostate cancer (8.3%) compared to Asian-Indian men (2.3%). This difference observed was also true for all age categories: 50-54; 55-69; and 60-64 years.

In a study by Phillips et al.⁴³ that evaluated cancer incidence and mortality in the Caribbean showed that Puerto Rican men had a high incidence rate of prostate cancer 100.1 per 100,000 and men in Barbados had similar rates of 99.7 per 100,000. Cuban men had the lowest rate 28.2 per 100,000, however, Barbados men had the highest mortality rate 68.1 per 100,000 in the Caribbean and Haitian men had the lowest mortality rate of 20.0 per 100,000.

Migrant studies also show that people of African descent have high rates of prostate cancer. Black men residing in the United Kingdom (UK) are considered an interesting population to examine because that population is mostly comprised of Afro-Caribbean and West African men³⁴. Typically immigrants bring with them patterns of disease that characterize their country of origin. However, over time immigrant groups or their progeny usually adopt a pattern similar to that of the host country⁴³. Consequently, the UK represents a unique microcosm of people of Afro-Caribbean and West-African populations. *The Prostate Cancer in Ethnic Subgroups* (Process) Study evaluated new prostate cancer diagnosis between January 1, 1999 and December 31, 2000 in the regions of: London Boroughs of Tower Hamlets; Hackney; Newham and the city of London. These parishes are comprised of 600,000 individuals with ethnicities of: white (British, Irish, and other white), black (black Caribbean, black African and other black, white and black African mix, white and black Caribbean mix), Asian (Indian, Pakistani, Bangladeshi, other Asian, white and Asian mix). This study reported 248 European, 91 African-Caribbean, and 20 south Asian prostate cancer cases.⁴⁴

In addition, in a retrospective follow-up study in Process on prostate cancer incidences in North Bristol, South West London, South East London, and North East London from 1997-2001 reported 2,140 cases: 61.4% white, 20% Black Caribbean, 4.8% Black African, and 7% uncoded ethnicity.²

Family History

The incidence of prostate cancer is greater/higher in men with a family history of a relative with prostate cancer. This risk increases with the number of affected family members. First-degree relatives of men with prostate cancer have a two-threefold increased risk of developing prostate

cancer. Men with two or more relatives with prostate cancer have a fivefold increased risk of developing this disease.⁴⁵

Studies have shown that familial prostate cancer constitutes approximately 5-10% of all prostate cancers and approximately 50% of all prostate cancers in men younger than 55 years of age.⁴⁶

Diet and lifestyle

Studies have reported that dietary intakes of high saturated fats, red meats, high total consumption, low fruits, low vegetables, low tomato products, low fish and low soy products increase the risk of prostate cancer. Vitamin D, calcium, lycopene, zinc, omega-3 fatty and alpha-linolic fatty acids, selenium, vitamin E, statins, and non-steroidal anti-inflammatory medications have also been shown to affect risk. In addition, certain anthropomorphics and activity have been shown to be linked to prostate cancer.²⁹

Likewise, higher circulating insulin like growth factor -1(IGF-1) can affect risk. It has also been shown that ejaculatory frequency maybe protective but the number of sexual partners can affect increase one's risk. The association between obesity and prostate cancer is somewhat controversial however obesity is associated with higher grade prostate cancer which could be caused by lowered testosterone levels.²⁹

1.2.4 Clinical risk factors for prostate cancer

Most prostate cancer cases are considered adenoma carcinomas that are derived from glandular epithelial cells. Furthermore, autopsies have shown that middle age individuals have a prevalence of malignant precursor prostatic intraepithelial neoplasia (PIN) and invasive

cancers.³¹ When a biopsy shows PIN but not actual cancer further biopsy is typically warranted.³²

Symptoms

The general symptoms of prostate cancer are urinary frequency, nocturia, and urgency, which are typically caused by the obstruction of the Urethra. However, in some individuals symptoms may come from painful skeletal metastases or anemia caused by bone marrow replacement.³¹

Screening and detection

Various studies have shown that a PSA level >4ng/mL increases the likelihood that prostate cancer will be detected at a prostate biopsy. Programs focused on early detection prostate cancer have reported that approximately 70% of cancer cases can be detected by using a PSA cut-off of 4ng/mL.⁴⁷

PSA test can provide diagnostic lead-time of 5-10 years. Approximately 80% of PSA cancers are biologically significant based on tumor volume and also Gleason grade. The use of PSA, results in earlier detection and diagnosis of organ confined disease.²⁹ Furthermore, experts assert that the use of PSA and DRE has resulted in the downward spiral of cancer mortality in the last few decades. The use of PSA screening method has resulted in increased number of diagnosed cases and the identification of cancer in the early stages.²⁹

Currently, the American Cancer Society recommends annual DRE and PSA test for men 50 years and older who have a life expectancy of over 10 years. For men who are considered high-risk i.e. African American men and men with a family history of prostate cancer screening should commence at 40 years of age.^{32, 48}

Digital rectal exam (DRE)

Typically physical findings are limited to the rectal examination. Attention is usually paid to areas of induration and also to determine if there is extension laterally to the pelvic sidewall, superiorly to the seminal vesicles and inferiorly at the apex. If it is determined that there is urinary obstruction, the bladder might be palpable. This examination is subjective and may not correlate cancer volume or extent of disease. Non-the-less, DRE results are an important facet of the algorithms that are used to determine if cancer is confined to the prostate gland or if it has spread to other regional areas.⁴⁹⁻⁵²

Gleason score

The Gleason score is used to microscopically evaluate prostate tissue and provide careful histologic grading. This is considered one of the most important factors in understanding clinical outcomes and prognosis of this heterogeneous disease.²⁹

Gleason grading evaluates the architectural details of cancer cells under a low-to – medium magnification. There are five patterns of growth from well to poorly differentiated.²⁹ For instance, pattern 1 is the most differentiated with discrete glandular architecture where as pattern 5 is the most undifferentiated, showing loss of glandular architecture. Consequently, the final Gleason score is the sum of the grades of the most common plus the second most common growth patterns. The Gleason score can range from 2(1+1) to 10(5+5).²⁹ The Gleason score correlates to clinical prognosis and is used to stratify individuals.

1.2.5 Androgens

Testicular androgens influence the development of the normal prostate and also play a key role in neoplasia. Testosterone is the main circulating androgen in the blood. Males who reach

castration levels of testosterone before puberty (eunuchs) do not seem to develop benign prostatic hypertrophy or prostate cancer⁵³.

Adrenal androgens androstenedione, dehydroepiandrosterone, and dehydroepiandrosterone sulfate can also be converted to testosterone and DHT but they provide a small proportion of circulating androgens hence they do not support the growth and development of the normal prostate. They however, could play an important role in advanced prostate cancer as a residual source of testosterone and DHT and by binding directly to mutant ARs. DHT is considered an extremely potent intracellular mediator both in the internal and the external genitalia⁵⁹.

1.2.5.1 **Racial differences in sex steroid hormones**

Studies have shown that African American men have higher mean circulating testosterone levels and other androgens compared to European men of comparable age. This difference in testosterone levels is higher during young adulthood (10-20%)^{54, 55} than in mid-adulthood (30%)⁵⁵. There is also greater age decline in testosterone levels in African American men compared to European men until the age of 40 years (6.7% > than white men age 31-34 years; 3.7% by age 35-39; and 0.5% by age 40-50 years)⁵⁵. Epidemiologic studies suggest that testosterone levels decline as part of the natural aging process in men. In addition, to this decline in testosterone there is an age related increase in sex hormone binding globulin (SHBG) which binds to approximately 44% of testosterone. The non-SHBG-bound testosterone fraction seems to decrease with increasing age. Suggested reasons for this decline is, decreased Leydig cell and secretory functions which might contribute to declining serum testosterone levels^{56, 57}. Harman et al.⁵⁶ suggest that modifiable lifestyle factors that are associated with aging could play a key role in the decline in testosterone levels.

In the CARDIA study, 483 African men and 695 European men aged 24-34 years at baseline were evaluated. In this study African American men were found to have 3% higher mean serum testosterone levels compared to European men even after adjusting for age, body mass index at baseline, and change in BMI after three hormone assessment in 8 years⁵⁸. The decline in testosterone with age was similar between African American and European men. Following the adjustment of baseline waist circumference and change in waist circumference racial difference in testosterone levels were eliminated. It should also be noted that in this study the African American men had smaller waist circumference and they had a greater increase in waist circumference over time compared to European men. In addition there were no racial differences in sex hormone binding globulin and free testosterone. Observations made from the CARDIA study suggested that observed racial variability in hormone levels might not be the result of inherent differences across racial groups but rather differences in certain factors that contribute to hormone levels such as: BMI, waist circumference which are inversely correlated with testosterone. On the other hand, racial differences in body habits could be caused by inherent differences in systemic hormone⁵⁹.

Racial differences in androgen variability in utero showed that mean serum testosterone levels were 50% higher in African American mothers in the first trimester compared to age, week of gestation matched European women⁶⁰. Another study observed higher maternal testosterone levels at delivery in African American women even after adjustments for known predictors of maternal testosterone levels. On the other hand, there were no observed differences in cord blood testosterone levels (African American 19.9, European 20.6 ng/dl).⁶¹.

1.2.5.2 **Definition of maleness and androgenicity**

The variations between genders are moderated by sex hormones. This process typically starts in the uterus. A complete insensitivity to testosterone leads to a female phenotype expression. Maleness is described as, “the phenotypical correlation of androgen effects in humans”⁶². Furthermore, maleness is denoted as, the degree of androgenicity. A male phenotype that has not been exposed to adequate androgen will vary according to the time of exposure but the major hallmarks will be during foetal development and also puberty. Moreover, because AR can be found in almost every tissue androgenicity is typically exerted everywhere. The areas that are most affected will be the places with an abundance of androgen receptors such as: prostate, testis, bones, larynx, brain, haematopoietic cells and certain types of hair follicles.⁶²

Persons affected by low androgen levels typically present with small prostates, low spermatogenesis, low bone density, a high pitched voice, anemia, feminized pattern of secondary hair growth, decreased libido, depression, and poor spatial cognition abilities. The effect of androgen has also been detected in lipid and glucose metabolism, even in fat cell physiology.⁶³

Testosterone (T) and its metabolite dihydrotestosterone (DHT) cause gene expression through the androgen receptor (AR). Typically T levels within normal range will saturate existing ARs and their androgenic effects will reach plateau levels depending on the type of tissue^{64, 65}. Consequently, it is reasonable to think that within a particular range of saturation, genetically determined differences in androgen receptor activity would be observed, where as in the condition of hypogonadism, androgenicity is dependent on the levels of T and DHT.⁶²

1.2.5.3 **Androgen metabolism in the prostate**

The pathway for androgen metabolism involves the two steroid hormones testosterone and dihydrotestosterone, the androgen receptor and important genes in the nucleus that are regulated by androgens⁵³.

In males testosterone is primarily synthesized by the testes and to a small extent by the adrenal glands. Approximately 45% of circulating testosterone binds to the sex hormone binding globulin (SHBG), 50% binds to albumin and <4% is unbound free testosterone⁶⁶. Testosterone diffuses into the prostate from blood and then binds directly to the AR⁵³. Testosterone in the prostate is converted irreversibly to 5 α -dihydrotestosterone by the enzyme 5 α -reductase type II which is encoded by the SRD5A2 gene⁶⁶. Even though testosterone and 5 α -dihydrotestosterone can bind to the androgen receptor; the androgen receptor has a higher affinity for the 5 α -dihydrotestosterone than for the testosterone. In addition, the androgen receptor is more transcriptionally active when it is bound to the 5 α -dihydrotestosterone. The 5 α -dihydrotestosterone –androgen receptor–transcription factor is regulated by translocation to the cell nucleus and binding of androgen receptor co-regulators, co-activators, and co-repressors. Furthermore, the 5 α -dihydrotestosterone coregulator complex translocates to the cell nucleus, in which once there it activates the transcription of genes with the help of other hormone responsive elements⁶⁷. The amount of 5 α -dihydrotestosterone in the prostate is determined by: testosterone metabolism, metabolism of androstenedione via 5 α -androstanedione and 5 α -dihydrotestosterone (DHT) inactivation by its reduction to 3 α or 3 β - androstanediol which is a reversible process. The metabolites of 5 α -dihydrotestosterone can also be conjugated into 3 α or 3 β -androstanediol glucuronide which is also an irreversible process⁶⁷. 3 β -androstanediol glucuronide is considered a terminal metabolite of testosterone and is used as a surrogate marker

of tissue androgen levels. 3β -androstenediol glucuronide reflect the activities of 5α -reductase types I and II. Type I is typically expressed in extraprostatic tissue like the skin and type II is expressed in the prostate⁶⁸. Serum concentrations, of 3α -androstenediol glucuronide correlates with 5α -reductase activity in the genital skin. Finasteride a 5α -reductase type II inhibitor show that DHT and 3α -androstenediol glucuronide decrease in men who were treated⁶⁹.

1.2.5.4 Serum testosterone, obesity, age, and tumor grade

There are many unresolved issues with serum androgen. It is well established that serum androgen decreases with increasing age and increasing age is a major risk factor for prostate cancer along with race and family history⁶⁹. In the literature there is evidence that high levels of serum testosterone is associated low grade prostate tumors but a decreased risk for high grade prostate tumors^{70, 71}. There is no evidence showing that total testosterone is associated with a decreased risk of high grade tumors but there is a possible association between free testosterone (unbound testosterone) and an increased risk for low grade tumors⁷⁰.

The deleterious effect of obesity also complicates the relationship between androgens and prostatic tumors. Hsing et al.⁷⁰ observed that obese men have a decreased risk of low grade and an increased risk for high-grade tumors, which is consistent with other findings that these men also have low serum testosterone levels^{72, 73}. This evidence further corroborates the findings that high serum testosterone concentration is associated with increased risk of low-grade prostate cancer and a reduced risk of high-grade prostate cancer. Obesity is associated with increased inflammatory markers, which are associated with a risk of prostate cancer⁷⁴. However, these observations may be modified by other additional modifiers and needs further evaluation.

1.2.5.5 **Androgen deprivation**

The concept that androgen deprivation can affect body composition was first proposed by Charles Edouard Brown-Sequard in 1889 ⁷⁵ he has been credited by many as the founder of endocrinology because he was the first person to suggest the existence of substances that are secreted by a particular organ but affects others ⁷⁶.

Male hormones called androgens (testosterone and dihydrotestosterone) is responsible for determining male secondary sex characteristics and is also responsible for stimulating prostate cell growth. Once prostate cells are deprived of androgens both healthy and cancerous they no longer grow and hence eventually die ⁷⁷.

Androgen deprivation therapy also denoted androgen suppression therapy and hormone therapy uses drugs or surgery to suppress or block male hormones (testosterone and dihydrotestosterone) that cause prostate cell growth. Testosterone is an androgen and androgens are produced by the testis and small amounts are also produced by the adrenal glands located on top of the kidneys. Androgen deprivation therapy blocks the production and the effects of androgen on the body. Androgen deprivation therapy is not considered a cure for prostate cancer but it can control symptoms and also disease progression in individuals. Androgen deprivation therapy is used to treat metastatic cancer and localized prostate cancer depending on PSA levels ⁷⁷.

In the literature there has been some debate as to when to commence androgen deprivation therapy. According to a report in 2007 by American Cancer Society of Clinical Oncology (ASCO) patients with recurrent, progressive or advanced prostate cancer should delay treatment until patients begin to experience symptoms. However, if therapy is deferred patients should have regular visits with their physicians (every 3-6 months) to monitor their condition.

ASCO also recommends bilateral orchiectomy, or injections with luteinizing hormone-releasing hormone as initial androgen deprivation treatments.⁷⁸ ADT treatment is considered palliative care and rarely curative. In the literature there is evidence to suggest that early treatment results in better long-term prognoses for prostate cancer.^{29, 32}

1.2.6 Treatment of prostate cancer

1.2.6.1 Orchiectomy

Orchiectomy is the surgical removal of the testicles. This method is considered the most effective way of reducing androgen hormones and is considered a more permanent solution to androgen deprivation. Orchiectomy in addition to radical prostatectomy can delay the progression of disease in patients with prostate cancer⁷⁹. Most of the testosterone that is produced by a man is produced by the testis in response to luteinizing hormone produced by the anterior pituitary gland. After bilateral orchiectomy, serum testosterone levels typically falls to castration levels (<10ng/ml)⁷⁹.

A reduction in serum testosterone is associated with decreased fat free mass. Mauras et al., reported that when serum testosterone was suppressed by the administration of gonadotropin-releasing hormone in healthy young men it was associated with significant reduction in fat free mass, increased fat mass, and a decrease in fractional muscle protein synthesis. Likewise, an age associated decline in serum testosterone levels was correlated with reduced appendicular muscle mass and reduction in lower extremity strength in Caucasians and African Americans^{80, 81}.

Current data on hypopituitary men suggested that hypogonadism is associated with abnormalities in body composition such as a reduction in lean body mass and elevation in fat mass⁸². Katznelson et al., reported effects of hypogonadism in men who also had

hypopituitarism and observed a higher percent body mass and abdominal subcutaneous fat in hypogonadal men compared to healthy controls even after controlling for BMI. However visceral fat was not greater in the hypogonadal group compared to the eugonadal groups^{83, 84}. Studies have also examined the relationship of androgen in elderly men such as bone mineral density and have found rather weak and inconsistent associations^{85, 86}.

1.2.6.2 **Prostectomy**

Another procedure that is sometimes recommended for the treatment of prostate cancer is radical prostatectomy, which is typically recommended for the treatment of stage A, and B (Whitmore-Jewett system staging system) prostate cancers. This procedure generally requires a surgical cut through the abdomen or the perineal area and is done with general or spinal anesthesia⁸⁷.

1.2.6.3 **LHRH agonists**

The main drugs used for suppressing androgens are called luteinizing hormone-releasing hormones (LHRH) agonists. LHRH drugs work by blocking the pituitary gland from producing hormones that stimulate the production of testosterone. These drugs include Leuprolide (leupron, leuprogel) goserelin (zoladex), and buserelin. Side effects from these drugs include hot flashes, nipple and breast tenderness, gynecomastia (breast enlargement)^{77, 79}.

1.2.6.4 **Antiandrogens**

Anti-androgens are drugs used to block the effects of testosterone. The main anti-androgen drugs are:

1. Flutamide (Eulexin, Drogenil). Flutamide side effects include: diarrhea and liver damage

2. Nilutamide (Nilandron). Nilutamide side effects include reversible interstitial pneumonitis, nausea, alcohol intolerance, and visual disturbances.
3. Bicalutamide (Casodex). Bicalutamide seems to have fewer side effects compared to other anti-androgens
4. Cyproterone combined with estrogen could prevent testosterone surge that typically occurs with LHRH.
5. Other drugs. If patients are not responding to hormonal medications, other drugs that can be tried include estrogen therapy and ketoconazole (Nizoral) which is an anti-fungal drug that blocks testosterone function in men⁷⁷.

1.2.6.5 **Chemotherapy for hormone resistant cancer**

For prostate cancer does not respond to hormone treatment chemotherapy may be use to treat hormone-resistant cancer. Chemotherapy drugs for prostate cancer are: docetaxel (taxotere), mitoxantrone (novantrone), estramustine (Emcyt), andriamycin, paclitaxel and other platinum based drugs such as carboplatin. These drugs are typically combined with other cancer dugs such as 5-fluorouacil, or corticosteroids (e.g. prednisone)⁷⁷.

1.2.6.6 **Untreated prostate cancer**

Information regarding patients who received no therapy for prostate cancer was evaluated in over 3,000 men identified in the Connecticut Tumor Registry. After diagnosis of prostate cancer these men either choose to receive no treatment or were advised by their physicians to receive no treatments. These individuals were followed for a minimum 15 years or until they died. In this study the two major determinants of death because of prostate cancer were: tumor grade and age at diagnosis. This study reported that men with Gleason score of 7 or higher and who were

below the age of 74 years were more likely to die of prostate cancer. On the other hand, men of all ages whose Gleason scores are 6 or lower have a low/minimal risk of dying from prostate cancer. In summary, this study resulted in two important observations namely: Gleason 7 tumors and higher should be considered high grade tumors and that age at diagnosis seems to bear no significant impact on cancer-specific mortality after 15 years in individuals with low grade or high-grade tumors. Age is more of a determinant in men with moderately differentiated tumors.

32, 88

1.2.7 ADT and body composition

It is well documented that androgen deprivation therapy with GnRH (gonadotropin-releasing hormone) agonist is the mainstay treatment for metastatic prostate cancer. Androgens are extremely important determinants of body composition in men. Studies have shown that GnRH agonist treatment increased weight and percentage fat body mass and decreased percentage lean body mass and muscle size in men ¹. Studies have also indicated that low serum testosterone concentrations are associated with decreased muscle mass and increased fat mass ⁸⁹. However, other studies have also shown that testosterone replacement therapy increases lean body mass in men with hypogonadism because of aging, HIV, and other chronic diseases ⁹⁰⁻⁹². In a prospective study that evaluated 48 weeks of GnRH agonist therapy in men with prostate cancer showed that these men experienced a 2.4% increase in weight, 9.4% increase in fat mass, and 2.7% decrease in lean body mass. This increased fatness resulted primarily from the accumulation of subcutaneous rather than intra-abdominal adipose tissue ¹. This effect on body composition could be related to testosterone deficiency because testosterone has been shown to promote lipolysis in visceral adipose tissue ⁹³.

In addition ADT has been shown to produce other metabolic changes for instance: increased serum insulin levels, total cholesterol, low-density lipoprotein, cholesterol, triglycerides and leptin^{1,94,95}. These abnormalities in lipid and glucose metabolism in addition to increases in body mass suggest that ADT could elevate an individual's cardiovascular risk. For example one study observed that after three months of GnRH agonist treatment participants had increased arterial stiffness in the radial artery and this problem resolved after treatment was discontinued⁹⁵. However, because testosterone and estrogen both have direct and indirect effects on the vascular endothelium, it is not quite clear if the hemodynamic effects are related to deficiencies in one hormone or both⁹⁵. Androgen deprivation therapy is also associated with fatigue, loss of energy, emotional distress and a poorer overall quality of life^{96,97}.

1.2.7.1 **ADT and bone loss**

In a study by Greenspan et al.⁹⁸ men with prostate cancer who initiated hormonal suppression therapy had significant BMD loss at a various skeletal sites. In this study there was greater bone loss after the first year of androgen deprivation therapy. The observed reduction in BMD following 12 months of acute ADT was 1.5-4.0% depending on the skeletal sites measured in individuals. Moreover, bone loss at the hips was related to the length of time that individuals were on ADT. On the other hand, men with prostate cancer who had not initiated ADT and healthy controls had no significant bone loss over 12 months. This study also observed an increase in percent body fat and a reduction in lean body mass in 12 months after the initiation of ADT. Studies have shown that reduced muscle strength is an independent predictor of falls in the elderly. Consequently reduced muscle mass may contribute a higher risk of fractures in men on ADT⁹⁹.

Furthermore in a study that was conducted men on ADT had 47% lower estradiol levels than healthy controls and these levels were still considerably lower than those of men with prostate cancer but had not initiated ADT treatment (31% lower). Treatment of prostate cancer with orchiectomy or GnRH agonist causes testosterone and its metabolite estradiol to fall significantly. These hormones are important determinants of peak bone mass in men ¹⁰⁰. However, estrogen is the dominant hormone that prevents bone resorption in men ¹⁰¹.

Current suggested treatments to prevent ADT related bone losses in men are: Calcium and vitamin D supplement of 1200-1500mg daily. In addition, individuals are encourage to abstain from negative habits that would exacerbate bone loss such as: smoking, excessive alcohol consumption, and a sedentary lifestyle ¹⁰². Another potential option to reduce bone toxicity caused by ADT is the use of bisphosphonates. Bisphosphonates are inhibitors of osteoclastic bone resorption¹⁰³.

1.2.7.2 **Definition of osteoporosis**

Osteoporosis is considered a skeletal disorder that is characterized by decreased bone mass and also a deterioration of bone tissue ultimately resulting in fragility of bone and subsequent susceptibility to falls¹⁰⁴. Studies have shown that there is an inverse relationship between bone mass and fractures. Osteoporotic fracture risk increases continuously as BMD decreases with an approximate 1.5 to 3-fold elevated risk of fracture for every standard deviation fall in BMD¹⁰⁵. Osteoporotic fractures account for approximately 0.83% of the global burden of diseases worldwide¹⁰⁶.

According to the World Health Organization, normal, osteopenia, and osteoporosis are based on BMD compared to the mean value in a young person. Normal is defined BMD within 1 standard deviation of the mean of a young adult person. Osteopenia is defined as BMD

between 1.0 and 2.5 standard deviation below the mean of a young adult; and osteoporosis is defined as ≥ 2.5 standard deviations below the mean of a young adult. These measures are based on DEXA forearm BMD measurements and hip fractures in postmenopausal Caucasian women. However, because men have higher BMD and larger bones than women additional studies are needed in this area¹⁰⁷.

1.2.7.3 **Osteoporosis gene polymorphism**

Due to the importance of family history and race genome wide linkage scans have been conducted to determine the genes that influence bone mineral density and osteoporosis. Association studies of the genetic effects of certain candidate genes on BMD and /or fractures have focused on the genes directly involved the metabolism of bone, structure, mineral and regulatory/hormonal pathways¹⁰⁸. Genes that have been involved in association studies are: estrogen receptor (α & β)^{109, 110}, vitamin D receptor¹⁰⁹⁻¹¹¹, PTH receptor type 1¹¹⁰, interleukin 6¹¹²⁻¹¹⁴, insulin like growth factors¹¹⁵, and alpha 2HS glycoprotein¹¹¹.

1.2.7.4 **Heritability of bone mass**

Genetics does play an important role in the determination of bone mineral density (BMD), and osteoporotic risk. For instance in a comparison of monozygotic twins, dizygotic twins, offspring and parents it was determined that genetic factors play a key role in osteoporosis^{116, 117}. Most studies demonstrated a heritability (h^2) of approximately 50-80% for BMD at fracture sites such as the lumbar spine, hips and the radius^{3, 118} but fracture heritability is considered more difficult to estimate with estimates of approximately 50%^{119, 120}. An individual's susceptibility to fractures depends on factors such as ones propensity to fall, non-skeletal factors, physical environment, and diminished soft tissue cushion. In addition, some heritability of fracture is

independent of bone mineral density^{121, 122}. Studies have also shown that SNPs do contribute to variations in BMD but might not contribute to osteoporotic fractures^{123, 124}.

Furthermore, studies have reported differences in the heritability of BMD in persons of the same and opposite genders¹²⁵. This could possibly be explained by in-utero imprinting expressions of osteoporosis genes. Other studies have found 20% greater heritability of BMD in men compared to women^{126, 127} and some did not^{128, 129}. A possible explanation for these gender specific heritability differences in BMD are our ability to account for environmental factors for instance estrogen in men^{130, 131}.

1.2.7.5 **Racial differences with BMD**

Osteoporosis is a serious public health problem that is associated with morbidity and mortality because of the risk of fractures. Fractures are usually associated with pain, limited mobility, and a reduced quality of life. Data suggest that fracture risk is greater among Europeans compared to African Americans living in the United States¹³².

The Baltimore Men's Osteoporosis Study examined racial differences in BMD, which is a longitudinal observational cohort study. The study was comprised of 503 ambulatory white and 191 ambulatory African American men. Men with bilateral hip replacement and excess weight of 300 pounds were excluded from the study. Higher BMD at the femoral neck, lumbar spine, and total body of African American men were observed even with the adjustments for potential confounders. Further analysis also showed that older African American men had narrower bone at the proximal femur and higher bone mineral content which is independently associated with greater bone strength¹³³.

In the Third National Health and Nutrition Survey (NHANES III); osteoporosis was present in 5 and 20 percent of black women and white women age 50 and above at the femoral

neck; 8 and 17 percent at the total femur. However, the prevalence of osteoporosis was 4 and 6 percent in African American and European men at the femoral neck and total femur. In this study prevalence of osteoporosis in men according to race varied depending on if female or male cutoff values were used. Prevalence's tended to be higher based on male cutoffs compared to female cutoffs ^{15, 132}.

Furthermore, in the Men's Osteoporosis study decline in BMD was evaluated in 349 European men and 119 African American men. BMD measurements were taken at the proximal femur and the lumbar spine at approximately 18.8 months apart. The results indicated that the average annual percentage decline in BMD was significantly greater in European men than African American men at the femoral neck and total hip. These observed differences persisted with the adjustment of known confounders: weight, age, and current smoking. In this study current smoking was associated with greater bone loss in blacks and whites even with adjustments of: age, change in weight, and height ^{132, 134}. Studies have also shown that African Americans have lower rate of bone turnover compared to European Americans ^{135, 136}. This lower rate of bone turnover ≈ lower decline in BMD in African Americans is believed to be the product of the combined effect of: nutrition, genetics, hormones, and lifestyle in adulthood ¹³².

Bone strength in individuals is determined by a number of factors namely: mass and geometry of the bone, microarchitecture of the trabecular bone, the porosity of the cortical bone, composition of the bone matrix, and the accumulation of micro-damage in the bone. African Americans seem to have greater mineral density compared to their European peers ¹³². This observed higher bone strength in African Americans could be the result of: a stronger skeleton during early childhood and adolescence, and slower bone loss in adulthood ¹³². Moreover,

studies have shown that African American children had greater bone mass than European children and these results continued into young adulthood in males^{137, 138}.

1.2.8 Androgen receptor (AR)

Androgen receptor (AR) is an intracellular transcription factor that is a member of the steroid nuclear receptor super family^{139, 140}. The androgen receptor gene (AR) contains in N-terminal of exon-1 a polymorphic trinucleotide CAG repeat (encodes polyglutamine), whose length regulates androgen receptor action. Included in this region is another trinucleotide repeat GGC which codes for glycine¹⁴¹. AR is activated by androgens and then translocated to the nucleus where it binds to DNA sequences called androgen response elements located in regulatory region of androgen dependent genes. Furthermore, the binding of androgen-AR complexes does one of two functions it either represses or activates androgen –regulated proteins¹⁴². Hence, ARs are very important molecular switches that control transcription of androgen-dependent proteins during embryogenesis until adulthood. The AR gene, located on the X chromosome at Xq11-12¹⁴³, is encoded in eight exons¹⁴⁴ and has three functional domains namely: the transactivation domain (TAD), DNA binding domain and ligand binding domain¹⁴⁵. Differences in the AR sequence are mostly characterized by polymorphic trinucleotide repeat (CAG) and the normal length of a CAG is approximately 9-37^{146, 147}. The transactivational activity of AR is inversely related to the length of the CAG repeat chain. In addition, because the androgen receptor is located on the X-chromosome males have only one copy¹⁴⁸.

The androgen receptor is sometimes mutated in prostate cancer with varying effects on functionality and on activating steroid and non-steroid ligands. Evidence for the importance of CAG AR repeat lengths are that: men with 40 or more CAG repeats suffer from Kennedy Syndrome (spinobulbar Ataxia) which is characterized by androgen insensitivity, and

progressive muscle weakness^{144, 149}. Men with longer CAG repeats are more likely to have reduced spermatogenesis than men with shorter repeats¹⁵⁰. One study found that men with prostate cancer had shorter CAG repeats which were associated with being African American and a higher and more aggressive prostatic carcinoma¹⁵¹. In addition, shorter repeats are associated with a higher risk of benign prostatic hyperplasia (BPH). BPH is characterized by overgrowth of the tissue of the transition zone and the periurethral area of the prostate¹⁵².

1.2.8.1 **Ethnic differences in *AR* CAG repeat length**

Within the normal range of the AR polyglutamate, ethnic differences have been reported. For instance, in healthy men of African descent the average number of CAG repeats reported is between 18 and 20^{13, 147} and is somewhat shorter in other African populations¹⁵³. In Caucasians the average number of CAG repeats is 21-22^{13, 147} whereas in East Asians a mean of 22-23 was observed^{146, 147, 154}.

1.2.8.2 **AR CAG repeat length and BMD**

Studies have shown that in healthy men, polymorphism of the oestrogen receptor regulate quantity and quality of bone tissue¹⁵⁵ and this is also applicable to CAG repeat polymorphisms found in the AR gene. In 110 males age 20-50 years an increased number of CAG AR repeats was associated with low bone density⁶⁵ and the results of this study was further corroborated by a study conducted in peri-menopausal women¹⁵⁶.

However, results in older men are somewhat conflicting. In a study of 508 Caucasian men 65 and over a negative association between AR CAG repeat length and bone mineral density of the femoral neck was observed¹¹¹. But in this same group, a more pronounced bone loss was

observed at the hip and an increased risk for vertebral fracture was reported in men with longer AR CAG repeat length ¹¹¹.

In a cohort of 273 Belgian men age 71 and 86 years no influence of AR polymorphism was observed ¹⁴¹. It is hypothesized that higher androgenization will cause greater peak bone mass¹³¹, and AR polymorphism effects on bone density will only be evident in healthy younger men. This difference in AR polymorphisms effects could be mitigated because of age-dependent bone loss in older men caused by confounders that exerted some influence on bone mass ⁶². In another study, that evaluated the effects CAG repeat polymorphism on bone density and metabolism, determined that increasing number of CAG repeats in the AR gene influence on bone density and bone metabolism is attenuated in healthy males. In this study CAG repeat polymorphism accounted for 8% of total variation of bone density; which suggests that the number of CAG repeats in the AR is a determinant of bone density in healthy men. Consequently one could speculate that a mild deficiency in testosterone is associated with negative effect on bone density in individuals with long CAG repeats sequence. While persons with shorter CAG triplets show no sign of bone loss for comparable testosterone levels ¹⁵⁷. Hence, longer CAG repeats in the AR gene will lead to lower peak bone density in men. Clearly, more studies are needed to elucidate the relationship between CAG AR and bone mineral density

1.2.8.3 **AR CAG repeat length and body composition**

Testosterone has shown conflicting associations with cardiovascular risk factors. Low levels of testosterone can have unfavorable effect on body composition, insulin sensitivity ⁵⁷ and haemostatic parameters¹⁵⁸. Studies have shown that androgen has a lowering effect on good cholesterol (HDL)¹⁵⁹. Furthermore, studies on the role of testosterone on vascular endothelial

function have shown a negative effect but these results were contradictory and appear to be contingent on endogenous hormone levels and how artificial testosterone was administered⁶⁴.

In a study of 110 men age 20-50 years CAG repeat polymorphisms decreases testosterone effect on HDL cholesterol and flow mediated dilation of the brachial artery. A longer polyglutamine stretch in the AR protein is associated with a higher cholesterol levels and arterial vasoreactivity⁶⁴. Similar findings were also reported in patients who suffered from XSBMA (X-linked spino-bulbar muscular atrophy)¹⁶⁰. It is quite possible that longer CAG repeats in the AR gene could possibly act as a cardio-protective factor but the receptor polymorphism might play a role in determining a person's atherosclerotic risk. It has been reported that in men with longer CAG repeats also had higher body fat and also insulin resistance⁶². Similarly, in patients with XSBMA they had a tendency to develop diabetes mellitus¹⁶¹.

1.2.8.4 **β 2Adrenergic Receptor**

Adrenergic receptors are members of a large super family of cell surface receptors that are involved in signaling via coupling to guanine nucleotide binding proteins (G-proteins). Adrenergic receptors are targets for epinephrine, catecholamines and norepinephrine and are an important component of the sympathetic nervous system for the maintenance of homeostasis and response to disease. Human adrenergic receptors consist of nine subtypes: α_{1A} -, α_{1B} -, α_{1D} AR; α_{2A} -, α_{2B} -, α_{2C} AR; β_1 , β_2 , and β_3 AR^{162, 163}. Four of these results in changes in the amino acid sequence: glycine-for-arginine substitution at codon 16 (Arg16Gly), glutamic acid-for-glutamine substitution at codon 27 (Gln27Glu), the methionine-for-valine substitution at codon 34 (Val34Met) and threonine-for-isoleucine substitution at codon 164 (Thr164Ile)¹⁶⁴. The human β_2 -adrenoceptor is located on chromosome 5q31-32¹⁶⁵.

A variability in a DNA sequence with an allele frequency of >1% in the population is denoted a polymorphism. Polymorphisms may have no effect, or have clinically silent effects, or could modify diseases or alter response to therapy, act as low level risk factors, and have increased prevalence in particular diseases. The most common type of polymorphism is single nucleotide substitution termed SNPs. Within the coding regions of genes, polymorphisms encode different amino acids (nonsynonymous polymorphisms). Polymorphisms occur in the 5' UTR, promoter, 3' UTR and introns¹⁶³.

1.2.8.5 **β_2 adrenergic receptor and hypertension**

Studies have shown that the β_2 -adrenergic receptor is involved in vascular smooth muscle relaxation and vasodilation in response to the release of epinephrine^{85, 166}. For instance, the impairment of β_2 -adrenergic receptor-mediated vasodilation in the presence of normal or elevated vasoconstrictor response causes an increased vascular reactivity and also raises blood pressure. Furthermore, blunted β_2 -adrenergic receptor-mediated vasodilation has been observed in hypertensive individuals¹⁶⁷⁻¹⁶⁹ indicating that attenuated β_2 -adrenergic receptor could be important in the pathogenesis of essential hypertension.

Even though, the mechanism is not fully understood, it is well known that hypertension is typically more severe and associated with higher rates of morbidity and mortality in African Americans compared to Caucasians¹⁷⁰⁻¹⁷².

In addition, it has been denoted that vascular β_2 -adrenergic receptor responses were attenuated in blacks considered normotensive¹⁷³ implying that an attenuated vasodilator response might play a key role in the development of hypertension in Blacks¹⁷⁴.

Genetic studies indicated that β_2 -adrenergic receptor could play a key role in increased hypertension and the risk of hypertension. It was denoted in an association study, that the gene

encoding β_2 -adrenergic receptor was associated with the development of essential hypertension¹⁷⁵. Likewise, salt sensitivity was found to be associated with β_2 -adrenergic receptor in African Americans¹⁷⁶.

However, the relationship between β_2 -adrenergic receptor polymorphism and hypertension still remains somewhat unclear. For instance, Gly16 polymorphism occurred more frequently in hypertensive Afro-Caribbean's¹⁷⁷ while in blacks from South Africa no association was found between Arg16-Gly and/or Gln27-Glu β_2 -adrenergic receptor polymorphism and essential hypertension⁹⁵. However, in a cohort of Germans Arg16 increased the predisposition of hypertension¹⁷⁸.

Consequently, it is evident that the relationship between β_2 -adrenergic receptor polymorphism and ethnicity and hypertension is poorly understood and further studies are needed to elucidate this relationship. Clearly, evaluating this relationship in Afro-Caribbean men would contribute significantly to the literature and to current knowledge and understanding of β_2 -adrenergic receptor polymorphism in hypertension.

1.2.8.6 β_2 adrenergic receptor and body composition

There have been studies showing a putative association between obesity and β_2 -adrenergic receptor. The β_2 -adrenergic receptor genes have been studied because of their key roles in the regulation of energy metabolism and utilization¹⁷⁹. The β_2 -adrenoceptor plays important roles in the regulation of energy homeostasis, for instance the stimulation of glycogen breakdown and lipid mobilization. β_1 co-exist with β_2 and β_3 adrenoceptors in the white adipose tissue^{164, 180}.

Gln27Glu variant influences body fat by changing the lipolytic response of adipose tissue to catecholamines. Catecholamines are lipolysis stimulating hormones^{180, 181}. Gln27Glu

is associated with a relative risk of obesity ≈ 7 and odds ratio of ≈ 10 ¹⁷⁹. The role of β_2 -receptor gene polymorphism for obesity has so far been mainly evaluated in women¹⁷⁹. Women who are homozygous for Glu27 have been found to have 20kg more fat mass compared to controls. Carriers of Glu27 had 50% larger fat cells compared to non-carriers¹⁷⁹. This polymorphism has also been found to be associated with obesity in the Swedes¹⁸² linked to obesity in the Japanese¹⁸³ and is associated with the early onset of obesity in the Danes¹⁸⁴. Interethnic differences have also been observed in allele frequencies. The Glu27 variant has been found to be more common in European Americans than African Americans¹⁷⁴.

1.2.9 Obesity in the Caribbean

The epidemic of obesity in the Caribbean has become a major health care problem. In the Barbados population between 1968 and 1981 obesity increased from 7.0 to 16.2% in males and 31.0 to 37.9% in females. In women over the age of 40 years the prevalence of obesity increased from 32% to 50%. Furthermore, in Jamaica over a 33 year span (1962-1995) the prevalence of obesity increased from 2% to 25% in men and 30 to 60% in women¹⁸⁵.

Obesity in the Caribbean is a major concern and has been shown to be more prevalent in countries with a higher per capita gross national product (GNP) and it seems to disproportionately affect women. Studies conducted in Jamaica, Barbados, Nigeria, Cameroon, St. Lucia and Maywood have demonstrated that hypertension increases in a manner similar to that of obesity¹⁸⁵. The prevalence of these chronic diseases are greater in lower income individuals from poorer countries who start living a westernized lifestyle influenced by sedentism, and caloric dense foods¹⁸⁶. Similar to United States obesity in the Caribbean plays an important role in the pathogenesis of T2DM, hypertension and negative body composition changes. It has also been demonstrated that diabetes mellitus frequently coexists with other

conditions such as abnormal serum cholesterol which is a symptom of the metabolic syndrome¹⁸⁷.

1.2.9.1 **Causes of weight gain**

Obesity typically occurs when there is an increase in energy consumption compared to energy expenditure¹⁸⁸⁻¹⁹⁰. Dietary surveys show an increase in caloric intake in the United States of approximately 200kcal/day in the past 20 years^{191, 192}. Much of this caloric intake corresponds to the increase consumption of fats, sugars, fast foods, snacks, high caloric beverages, larger food portion sizes and eating outside the home¹⁹³. For instance, meals at fast food restaurants¹⁹⁴, full service restaurants¹⁹⁵, and eating between meals have been shown to cause an increase in body weight^{193, 196}.

1.2.9.2 **Energy output: physical activity**

In 2002 the Center for Disease Control (CDC) and Prevention determined that approximately less than 30% of individuals in the United States were engaging in sufficient physical activity. While an additional 30% of persons were active but not sufficiently with the remainder of individuals predominantly sedentary engaging in minimal levels of physical activity^{191, 197}.

The percentage of the population that reports participating in no leisure time physical activity is greater amongst women compared to men, greater in African Americans, and Hispanics compared to European Americans, higher among older adults compared to younger adults and greater in the less affluent compared to the more affluent in society¹⁹⁸. It has been shown that individuals with lower levels of education and income engage in less leisure time physical activity¹⁹⁸. Studies have also demonstrated that individuals who are moderately or

highly active have a longer life expectancy and have more years free of chronic diseases such as diabetes¹⁹⁹.

1.2.10 Distribution of body fat

Obesity related changes are significantly related to an accumulation of fat in the body, and in the literature there is substantial evidence indicating that central obesity is a stronger predictor of morbidity than the accumulation of fat in the lower body^{200, 201}. The accumulation of fat in the abdominal regions has significant implications for metabolism and insulin sensitivity²⁰¹. These co-morbidities are more related to waist circumference than to BMI. Additionally, visceral or abdominal fat is strongly associated with an elevated risk of cardiovascular disease and type 2 diabetes²⁰². Visceral adiposity is considered a risk factor for cardiovascular disease, insulin resistance, metabolic syndrome and T2DM in adults^{203, 204}.

1.2.10.1 Subcutaneous and visceral fat

Intra-abdominal fat has been shown to be linked to an increased risk of developing debilitating diseases. Adipose tissue that is located in the abdomen is comprised of both visceral and subcutaneous fat; and there has been some discrepancy regarding the contribution of these compartments to the development of the metabolic syndrome in overly obese individuals. Studies have shown that there is a positive relationship between abdominal subcutaneous fat and the development of insulin sensitivity²⁰⁵.

Many researchers report that visceral adipose tissue is a stronger determinant of insulin resistance compared to subcutaneous adipose tissue. Studies have shown that both visceral and subcutaneous fat are related to insulin sensitivity²⁰⁶. It has also been shown that the amount of

subcutaneous fat is strongly associated with the level of insulin resistance and increased lipolytic rates compared to the superficial subcutaneous abdominal fat²⁰⁷.

1.2.10.2 **Intramuscular fat**

Studies have revealed that the location of fat within the thigh determines disease risk²⁰⁸. Fat stored in the non-subcutaneous regions (meaning fat located in the muscle or around muscle fibers) was related to insulin resistance in persons considered obese; however, there was no correlation with subcutaneous thigh fat²⁰⁸. In addition, a positive association has also been observed between insulin resistance and intramuscular lipid levels^{209, 210}.

Krssak et al.,²⁰⁹ has shown that intracellular lipid concentrations were correlated with insulin sensitivity using novel proton nuclear magnetic resonance spectroscopy technique (H NMR)²⁰⁹. In addition, Goodpaster et al.,²⁰⁸ revealed that sub-facial adipose tissue and intermuscular adipose tissue are markers of insulin resistance in obese persons and type II diabetics. However, some but not all studies delineated a relationship between skeletal muscle lipid and insulin sensitivity. One study of older normoglycaemic women found that intracellular lipid content was inversely correlated with insulin sensitivity²¹⁰. Intracellular lipid however was shown to be related to waist hip ratio and fasting plasma concentrations of non-esterified fatty acids (NEFA) but not with glucose tolerance or whole body insulin sensitivity²¹⁰. Another study reported an inverse correlation between muscle triglyceride and insulin sensitivity²¹¹.

Fatty acid concentration of the cell membrane in the skeletal muscle affects insulin sensitivity by altering the physicochemical properties of the membranes, which alters receptor function, ion transport over the membrane, cell energy requirement and cell signaling. In addition, elevated levels of saturated fatty acids in the cell membrane disrupts insulin action by altering insulin receptor binding, altered ability to translocate or insert glucose transporters,

changing phospholipid fatty acids-interaction with function of second messenger (protein kinase C) and reduced ion permeability²¹².

1.2.10.3 **Waist circumference (WC) and waist to hip ratio (WHR)**

Waist and hip circumference ratios are important measurements of the abdomen as well as in predicting abdominal obesity²¹³. The values of the waist-to-hip circumference ratio (WHR) used to define abdominal obesity are 0.95 for men and 0.80 for women. There is evidence suggesting that waist circumference (WC) is a better clinical measure of abdominal obesity than WHR²¹⁴. A potential limitation of WHR is, it might not change when the hip and waist circumference increase by comparable ratios. Consequently, WHR is typically inaccurate in the event of significant loss of visceral adipose tissue²¹⁵.

There have been a few studies that have suggested that WC and WHR predicts abdominal adipose tissue equally well. However, current data are more in favor of using WC as a better measure of abdominal fat^{216,217}. WC predicts percentage body fat which correlates well with the development of insulin resistance and the development of cardiovascular risk factors in African Americans, European Americans and Europeans²¹⁸⁻²²¹. Moreover, the interpretation of these WC values depends on the race/ethnicity of the group under evaluation. Caribbean populations, such as Jamaica, Nigeria, St. Lucia, Barbados and Cameroon the predictive values of established cut-offs of WC showed poorer sensitivities for abdominal obesity²²².

1.2.10.4 **Body mass index (BMI)**

Body mass index (BMI) is a statistical measurement, which compares a person's weight and height. The BMI is a useful tool to estimate a healthy body weight based on how tall a person is, however; it does not measure the percentage of body fat. This measure of general adiposity is

defined as weight in kilograms (kg) divided by height in meters². According, to the WHO a BMI <18.5 kg/m² is defined as underweight, 25-29.9 kg/m² is considered overweight and ≥30 kg/m² is defined as obese^{191, 223, 224}. Several studies have shown that abdominal circumference is highly correlated with increased risk of type 2 diabetes than BMI²²⁵. Although, BMI is one of the most common measures of assessing general obesity in adults, Other measures such as dual X-ray absorptiometry (DEXA), Quantitative computerized tomography (QCT), computed tomography (CT) and peripheral quantitative computerized tomography (pQCT) can provide more accurate measures of body fat or the percentage body fat in an individual²²⁶.

1.2.10.5 **Visceral fat as a pro-inflammatory tissue**

Visceral fat shows evidence of accelerated lipolytic activity with an elevated release of free fatty acids that can severely affect insulin action and glucose diffusion in many tissues²²⁷⁻²³⁰. These elevated levels in circulating FFAs may also cause an increase in triglyceride reservoirs in the muscle and liver decreasing insulin action and elevating hepatic very low-density protein output^{231, 232}.

On the other hand, a reduction in visceral adiposity and FFA levels after weight loss has been associated with enhanced insulin sensitivity^{233, 234}. Visceral fat has been shown to be responsible for the release of cytokines, such as tumor necrosis factor α (TNF- α) and interleukin 6 (IL-6)^{235, 236}. TNF- α inhibits tyrosine kinase phosphorylation of the insulin receptor, causing a defect in insulin signaling contributing to insulin resistance and impaired glucose transport²³⁷. An imbalance of proinflammatory cytokines from adipose tissue can trigger C-reactive protein (CRP) secretion. It has been shown that CRP levels are strongly associated with insulin resistance and adiposity, and elevated levels of CRP has been shown to be correlated with impaired endothelial dysfunction and CAD²³⁸.

1.2.10.6 **Race ethnicity and body fat distribution**

There have been several studies that have shown that body fat distribution varies according to population. These differences in body fat distribution were found to be associated with elevated triglycerides, high blood pressure, low HDL cholesterol, and high glucose levels. Studies in the United States and South Africa have indicated that visceral fat is smaller in African Americans compared to Europeans matched on BMI. However, it has been noted that in the black populations, individuals are more insulin resistant compared to European populations²³⁹⁻²⁴¹.

Abdominal fat contributes different degrees of risk for hypertension depending on the ethnicity of the person. For instance, compared to Europeans, black men and women have a 1.58 and 1.39 greater risk of developing hypertension if their waist circumference values are extremely high²⁴². Likewise, the clustering of insulin resistance syndrome associated with abdominal obesity seems to occur more frequently in Black and Hispanic individuals²⁴³. Depress et al.,²⁴⁴ reported that with comparable levels of fatness, Europeans appeared to have more abdominal fat compared to African Americans. African American women appear to have higher levels of total body fat but similar abdominal adipose tissues as their European counterparts.

1.2.11 **The metabolic syndrome**

The term metabolic syndrome is defined as the clustering of cardiovascular risk factors related to hypertension, abdominal obesity, dyslipidemia, and insulin resistance. According to the National Cholesterol Education Program Adult Treatment Panel ATP III guidelines in 2001, metabolic syndrome is the presence of three or more of the five risk factors. Other characteristics of the metabolic syndrome are: hyper-insulinemia, insulin resistance, and higher density of LDL cholesterol. The metabolic syndrome is also associated with elevated levels of inflammatory risk

markers, such as fibrinolysis, plasminogen activator inhibitor-1, oxidative stress, microalbuminuria, irregularity in autonomic regulation, and the activation of the rennin angiotensinaldosterone axis²⁴⁵.

1.2.11.1 **The impact of ethnicity on the metabolic syndrome**

Ethnicity has a major impact on metabolic and anthropometric changes as well as co-morbid diseases. A major component of the metabolic syndrome is obesity. In South Africa, obese Black women matched for BMI and body composition were compared to obese urban European Caucasian women and their metabolic differences and β -cell function examined. The Black women had a higher degree of insulin resistance, lower plasma insulin and C-peptide levels, reduced anti-lipolytic activity, higher post absorptive FFA concentrations, lower fasting and 3-h triglyceride concentrations, and lower plasma cortisol concentrations^{240, 241, 246-251}.

Other studies have shown that black women have less visceral fat and black diabetics have higher fat mass compared to obese black women without diabetes with similar BMIs. These studies also showed that weight loss occurred more swiftly in the gluteofemoral regions in black women, but weight loss in the visceral regions occurred in both groups. This weight loss that occurs in the in black women might be the result of greater lipolytic activity in the gluteofemoral regions^{252, 253}. On the other hand, the visceral fat in obese European women has been shown to be associated with more atherogenic fasting and postprandial lipid profiles^{240, 247, 248, 253}.

Similar findings were also observed in fit men as that observed in obese women. For example plasma insulin and postprandial glucose tolerance were lower in lean black men compared to European Caucasian men. Lipolytic activity was also elevated in lean black men compared to lean European Caucasian men²⁵³.

1.2.11.2 **Prevalence of the metabolic syndrome**

According to the NHANES data (1988-1994) the prevalence of the metabolic syndrome in the United States was estimated to be approximately at 23.7 percent or approximately 47 million people have this problem. In the United States the metabolic syndrome is expected to increase exponentially. Prevalence of the metabolic syndrome was defined according to ATP III of the abnormalities: waist circumference greater than 102 cm in men and 88 cm in women; serum triglycerides level of 150 mg/dL (1.69 mmol/L); high density lipoprotein cholesterol level of less than 40 mg/dL in men and 50 mg/dL in women; blood pressure of at least 130/85 mm Hg; or a serum glucose level of 110 mg/dL. The unadjusted and age adjusted prevalence of the metabolic syndrome were 21.8% and 23.7% percent respectively. The prevalence of the metabolic syndrome increased from 6.7% in adult's age 20 years to 29 years to 43.5% and 42% for individuals 60 to 69 years. Mexican Americans had the highest age adjusted prevalence of the metabolic syndrome (31.9%). The age-adjusted prevalence was similar for men (24.0%) and women (23.4%). But African American women had 57% higher prevalence than men ²⁵⁴.

In men the metabolic syndrome (MetSyn) is associated with a fourfold elevated risk for CHD, a twofold risk of CVD and all cause mortality even with adjustments for age, LDL-cholesterol, smoking, and a family history of CHD. Likewise, the metabolic syndrome in women is associated with an increased risk of CHD. Individuals with the metabolic syndrome are five to nine times more likely to develop diabetes. However, in the PROSPER study, the metabolic syndrome and its individual components were not associated with the development of CVD but the presence of the metabolic syndrome at baseline was associated with a future risk of developing diabetes especially in individuals with a BMI >30kg/m² and a fasting a glucose of >6.1 mmol/L ²⁵⁵. Similarly, in the BRHS study, the metabolic syndrome was weakly associated

with the development of CVD events but its individual's components such as waist circumference, serum triglycerides, HDL cholesterol, elevated blood pressure, and fasting glucose were associated with an increased risk of developing diabetes²⁵⁵. Hence, these results suggest that the metabolic syndrome is strongly associated with the development of diabetes in the middle age and also the elderly but is weakly associated with CVD²⁵⁵.

In the St. James Coronary Risk Factor study, African Trinidadians had lower cholesterol and LDL cholesterol and higher HDL cholesterol than Asian Trinidadians. It is believed that a large percentage of this population has at-least three maybe four of the characteristics considered the deadly quartet or Raven syndrome X: which are obesity, hypertension, diabetes, glucose intolerance and hyperlipidemia²⁵⁶⁻²⁵⁹.

1.2.11.3 **Lifestyle modification**

According to the SMART study regular exercise reduced the risk of developing metabolic syndrome by at least 50% and these results remained significant even without weight loss in individuals. In this cohort insulin resistance was inversely correlated with physical activity on the homeostasis model assessment (HOMA)²⁶⁰. Furthermore, exercise reduces adiposity and weight while improving insulin sensitivity.

Lakka et al.,²⁶¹ reported that in the HERTAGE study physical activity reduced markers of inflammation for instance C-reactive protein. It is believed that physical activity improves the risk factors for insulin resistance and hypertension by inhibiting the production of cytokines and their potential interaction with monocytes and endothelial cells^{261, 262}.

Dietary modifications are also helpful in treating insulin resistance and hypertension. For instance, a heart-healthy diet lowers levels of TNF- α ²⁶³. In the Framingham offspring cohort a

diet high in whole grains and fiber reduced the risk of developing insulin resistance and the metabolic syndrome²⁶⁴.

The ARIC (atherosclerosis risk in communities) study reported that a western diet increases the risk of developing the metabolic syndrome by 18%. This study reported that the consumption of meats, fried foods, and the regular intake of diet sodas was associated with increased risk of the MetSyn²⁶⁵. However, dairy products were inversely associated with MetSyn both cross-sectionally²⁶⁶⁻²⁶⁸ and prospectively²⁶⁷. No association was found between the association of the MetSyn and whole grains, refined grains, fruits, vegetables, nuts, coffee and sweetened drinks²⁶⁵.

Final, the DASH diet, which is considered to be rich in fruits, vegetables, low fat dietary products, sweeteners and reduced consumption of meats, reduced the risk of developing the metabolic syndrome and hypertension²⁶⁹. Exercise does provide some protection against the metabolic syndrome and also hypertension. Individuals who lost weight showed a reduction in their mean arterial pressure, plasma renin activity and aldosterone levels¹⁵⁶.

1.2.12 Measurement of body fat distribution

1.2.12.1 Dual X-ray absorptiometry (DEXA)

DEXA scans are primarily used to evaluate bone mineral density, measure total body composition and fat content. The speed of DEXA, its easy use, relatively low costs and lower radiation compared to computed tomography (CT) makes it good a method for use in clinical settings, research, and also in large population studies. The DEXA machine uses two-X-ray beams with differing energy levels. When soft tissue absorption is subtracted out, the BMD can be determined from the absorption of each beam by bone^{270, 271}.

1.2.12.2 **Computed tomography (CT)**

Computed Tomography Imaging can be used to quantify a cross sectional area of tissue. CT distinguishes tissue in vivo based on the level of which they attenuate the energy of an X-ray. These attenuation processes depend substantially on their density and are expressed in Hounsfield units (HU), positive for muscle and negative for fat. Adipose tissue is not as dense as water and hence displays attenuation values that are of a negative value. However, muscle is denser than water and consequently would have more positive attenuation values²⁷². CT is a very important method for quantification of visceral fat.

1.2.12.3 **Quantitative computerized tomography (QCT)**

Historically quantitative computerized tomography (QCT) was used to measure volumetric bone density of the spine and hip. QCT also analyzes the cortical and trabecular bone separately²⁷⁰. This method has been proven to be quite useful in clinical research settings and also used to evaluate patient progress particularly therapeutic responses to parathyroid hormone therapy. Even though this method has mainly been used to determine bone mineral density of the periphery it can also be used to measure body fat.

1.2.12.4 **Peripheral quantitative computerized tomography (pQCT)**

Peripheral quantitative computerized tomography (pQCT) is used to measure peripheral sites²⁷². pQCT has been used to determine body fat in mice. The accuracy of the values obtained from pQCT was compared to chemical extraction in mice. The coefficient variation for determining percentage body fat in mice using pQCT was 3.9% (± 1.8 SD). pQCT was considered to have good accuracy and also precision in determining percentage body fat and liver fat in animals studies²⁷³. Ranke et al.,²⁷⁴ determined that the use of pQCT was a suitable technique to

determine the size, the structure of muscle, fat and bone in pubertal girls with Turner syndrome who were being evaluated on the effects of growth hormone therapy.

1.2.13 Diabetes

1.2.13.1 Prevalence of diabetes in the United States

Current data show that the prevalence of diabetes in the United States has risen 40% from 4.9% in 1990 to 6.9% in 1999²⁷⁵. Type 2 diabetes mellitus is 10 times more common in adults over the age of 65 years compared to those younger than 45 years of age²⁷⁵.

In addition, T2DM is more common in minority racial groups such as: Native Americans, Hispanics, and African Americans. This disease affects these groups at a rate 2-4 times compared to European American populations. This increased prevalence of T2DM has been reported in adolescence and children, T2DM mellitus tend to occur more frequently than type 1 diabetes²⁷⁵.

In the United States the estimated lifetime risk of developing T2DM in 2000 was 33% for males and 39% for females²⁷⁵. T2DM is also associated with reductions in life expectancy of 11 years in males diagnosed after the age of 40 years²⁷⁶.

1.2.13.2 Pathogenesis of diabetes

Glucose is considered the body's main source of energy and in most mammals there is a cell membrane protein that facilitates the diffusion of glucose from the blood stream following the digestion of food into the cell. A gradient is maintained facilitating the diffusion of glucose into the cell by phosphorylation of sugar once it enters the cytoplasm lowering intracellular glucose

concentration. There are several proteins that facilitate the transportation of glucose and these isoforms and they are termed GLUT1-5 distinguished by the tissues in which they are found²⁷⁷.

Insulin is a hormone produced by the endocrine cells of the pancreas (β -cells) and plays an important role in the maintenance of blood sugar levels. An elevation in blood glucose levels triggers the release of insulin, which stimulates the uptake of glucose into target cells such as the skeletal muscle and adipocytes. When insulin levels rise in response to elevated blood glucose levels this hormone acts on target cells and stimulates the translocation of glucose into the cell. Diabetes is caused by a defect in insulin activity²⁷⁷. The glucose transporter GLUT4 mediates insulin uptake in adipocytes and also in muscle cells by moving glucose from intracellular storage sites into the plasma membrane²⁷⁸.

In persons who are not diabetic normal plasma glucose range is between 4 and 7 mM. This maintenance of normal glucose levels is governed by a balance between glucose absorption from the intestines and its production by the liver and its metabolism by the peripheral tissues. As mentioned before insulin increases glucose uptake in muscle and fat, and inhibits hepatic glucose production consequently acting as the regulator of blood glucose concentrations. Likewise, insulin fuels cell growth and differentiation, and enhances the storage of substrates in fat, liver and muscle through the process of lipogenesis, glycogen, and protein synthesis, and also through the inhibition of lypolysis, glycogenolysis, and the breakdown of proteins²⁷⁹.

T2DM is a non-autoimmune disorder. Patients with T2DM have insulin resistance as well as a significant reduction in insulin response to glucose. Cross-sectional studies have demonstrated that high risk groups have severe reductions in insulin sensitivity and or defects in β -cell function or they may have abnormalities in both path ways²⁸⁰. Studies have indicated that older individuals are at an increased risk of developing type II diabetes. Typically because these

patients have impairments in postprandial rather than fasting glucose²⁸¹. It is believed that a significant number of individuals that were originally diagnosed as type 2 diabetics were cryptic type 1 diabetics who later evolved to a type 1 state denoted as anti- β -cell autoimmunity²⁸².

1.2.13.3 **Obesity and insulin resistance**

Insulin resistance is denoted as, “reduced insulin-mediated glucose uptake in insulin sensitive tissues such as skeletal muscle and liver”²⁸³. In individuals who are obese, insulin resistance is common and is typically related to central abdominal fat. Much of the fat located in the central abdominal region is called visceral adipose tissue (VAT). VAT is related to insulin resistance and the metabolic syndrome. This fat is mostly comprised of mesenteric and omental fat masses that can be measured through a variety of diagnostic measures such as CTs, MRIs and DEXA scans. Visceral adipose tissue accounts for a small proportion of total fat in the body namely 10% whereas 25% to 50% of abdominal fat is subcutaneous. However, many researchers believe that it is VAT that confers a risk for insulin resistance because it has a higher lipolysis rate than SAT and also because of its close proximity to the portal circulatory system²⁸⁴.

1.2.13.4 **Adipocytes, insulin resistance and diabetes**

Adipose tissue is considered an extremely active metabolic and endocrine organ that participates in a variety of homeostatic processes that produces hormones such as leptin, adiponectin, resistin, TNF α , and IL-6 which influence insulin sensitivity and plays a key role in the pathogenesis of insulin resistance, diabetes, dyslipidemia, inflammation and atherosclerosis²⁸⁵⁻²⁸⁸. Likewise, adipocyte derived factors contribute to the development of β -cell dysfunction and also type 2 diabetes²⁸⁹.

Adiponectin is a protein and also an adipose tissue-specific collagen like factor²⁹⁰ that is secreted by adipocytes and can be found in concentrations of 7 to 12mg/L in healthy individuals. It has been shown that adiponectin strongly correlates with decreased fatty acid blood concentrations and a reduction in BMI and body weight. In addition, adiponectin could possibly be the molecular link between obesity, insulin resistance and could potentially serve as the biomarker for the metabolic syndrome²⁹¹.

1.2.13.5 **Insulin resistance**

Insulin resistance is related to diabetes, hypertension and obesity and is characterized by decreases in receptor concentration, kinase activity and also the phosphorylation of IRS-1 and IRS-2, PIK activity, glucose transporter translocation and the activity of intracellular enzymes. In persons who suffer from T2DM the activation of the MAP kinase pathways is not diminished potentially leading to some of the harmful effects of chronic hyperinsulinaemia on cell development in the vasculature^{279, 292}.

Genetic as well as acquired factors can significantly affect insulin sensitivity. Genetic defects in insulin receptor is extremely rare however it can be found in syndromes such as leprechaunism, Rabson Mendenhall, and type A syndrome of insulin resistance. Type 2 diabetes is polygenetic and entails multiple gene polymorphism that encode the proteins involved in insulin signaling, secretion, and also its metabolism²⁷⁹.

1.2.13.6 **Diabetes in the Caribbean**

Diabetes mellitus is considered one of the most prevalent diseases and a leading cause of death in the Caribbean region and has emerged as a major health concern. Diabetes is characterized by chronic hyperglycemia, and other metabolic disorders for instance, atherosclerosis, damage to

target organs such as the heart, kidneys, eyes, brain, and the peripheral nerves. Obesity seems to play a central role in the development of diabetes and other co-morbid conditions such as hypertension and dyslipidemia. Despite the growing concern regarding diabetes in the Caribbean there are still many undiagnosed individuals in that region ¹⁸⁷.

Diabetes mellitus has been shown to be associated with increasing age up to approximately the 6th decade and then eventually leveling off. A denoted reason for this increased prevalence in diabetes mellitus is the aging population in the Caribbean. For example, amongst Barbadian Blacks diabetes was shown to be positively associated with increased age, family history of diabetes, central obesity and a body mass index $\geq 25\text{kg/m}^2$ ^{2187, 293}.

1.2.13.7 **Economic burden of diabetes**

NIDDM causes economic burden to many societies especially bi-ethnic Caribbean countries such as Trinidad and Tobago ²⁹⁴⁻²⁹⁶. This increase in diabetes in the Caribbean seems to be driven by changes in living conditions. For example, diets in the Caribbean have become more like westernized countries high in fat, sugars and low in dietary fibers. Sedentary lifestyle, and labor saving devices have also increased the likelihood of individuals developing diabetes ²⁹⁷.

1.2.13.8 **Under report of diabetes in the Caribbean**

Diabetes in the Caribbean has a high rate of being under-reported. For instance, in a study of the national death certificates in Jamaica, revealed that there was a 30% under-recognition of diabetes ²⁹⁸. In Spanish Town, Jamaica the prevalence of diabetes in persons with an average age of 47 years was 9.8% for men and 15.7% in women with an overall prevalence of 13.4% ²⁹⁷. In Trinidad and Tobago, the prevalence among Indo-Trinidadian men and women 35-69 years of age increased from 11.6% and 18.9% in 1961-62 to 19.5% and 21.6% in 1977. Among Afro-

Trinidadians the rates were 2.5% to 8.2% among men and 5.4% to 14.8% amongst women²⁹⁴,²⁹⁹. This epidemic of diabetes mellitus is expected to continue with a 35% increase in the prevalence from 1995 until 2025 which will be accompanied by growth in population size, which will be largest in developing countries³⁰⁰.

In Trinidad and Tobago type 2 diabetes accounts for approximately 13.6% of hospital admissions and cost the government an estimated US\$ 1.6 million dollars³⁰¹. It is well established that abdominal obesity is associated with insulin resistance and is highly predictive of the development of type 2 diabetes and other cardiovascular diseases³⁰². In the Caribbean, there is also a gender dimorphism in which females have a higher prevalence of diabetes and obesity. The influence of lifestyle factors on prevalent diabetes has been succinctly demonstrated by the International Collaborative Study on Hypertension in Blacks in which a gradient of prevalent diabetes cases was delineated ranging from 2.0% in West Africa to 9.0% in the Caribbean and 11% in African Americans³⁰³.

In this region of the world many of the studies on the prevalence of diabetes in the Caribbean were conducted more than 30 years ago^{299, 304-306} and since then there have been changes in diagnostic criteria which has significantly limited the ability to compare those data to current data. However, Poon-King et al. conducted a prevalence study in approximately 2400 individuals in July 1961 and found that rates of diabetes were 2.4% in East Indian Trinidadians and 1.4% in African descent Trinidadians²⁹⁹. There are no new studies addressing the differences in rates of diabetes in blacks and Indians residing in Trinidad today.

1.2.14 Hypertension

A small percentage of patients have as the main cause of their hypertension renal or adrenal disease approximately 2%-5%. However, in the remaining population there is no single

identifiable cause for this disease called essential hypertension. There are a number of physiologic processes involved in the maintenance of normal blood pressure and their disturbance is possibly a key role in the development of essential hypertension. It is also quite possible that there are a number of inter-related factors involved in the etiology of hypertension. Some of the more intensely studied factors are: salt-intake, obesity, insulin resistance, the rennin-angiotensin system, and sympathetic nervous system³⁰⁷.

1.2.14.1 **Definition of hypertension**

Hypertension is defined as the “consistent elevation of the systemic arterial blood pressure”. Individuals are usually diagnosed as having hypertension after an average of approximately two or more diastolic blood pressure measurements of 90 mm Hg or higher on consecutive visits or when the average systolic blood pressure measurements on consecutive visits is 140 mm Hg or greater²⁴⁵.

Hypertension is typically caused by an increase in cardiac output, peripheral resistance or a combination of both. Increases in cardiac output can be caused by any condition that could increase the heart rate or stroke volume. On the other hand, peripheral resistance is caused by any condition that increases the viscosity of the blood or decreases vessel diameters for instance arteriolar diameter. Individuals who suffer from hypertension can have a combined systolic and diastolic hypertension or some individuals might have isolated systolic hypertension. Usually most cases of combined systolic and diastolic have no clearly delineated causes and so are termed primary hypertension. Primary hypertension is also called essential hypertension or idiopathic hypertension³⁰⁸. Hypertension of unknown origin is termed essential hypertension. This type of hypertension is treatable but not curable and there is a strong hereditary component³⁰⁹.

1.2.14.2 **Characteristics of severe hypertension**

- MAP is increased by 40-60%
- In severe stages renal blood flow is decreased by one half of normal
- Blood flow through the kidneys is increased twofold to four fold
- CO is normal
- The kidneys do not excrete adequate salt and water unless the arterial pressure is high

309

Secondary hypertension is usually caused by altered hemodynamics related to the primary disease such as arteriosclerosis. Secondary hypertension accounts for approximately 5% to 8% of cases. Isolated systolic hypertension is defined as elevated systolic blood pressure accompanied by normal diastolic blood pressure defined as <90 mm Hg. Isolated systolic blood pressure is characterized by increased cardiac output or rigidity of the aorta or both³¹⁰.

1.2.14.3 **Obesity and Hypertension**

The relationship between hypertension and obesity is well documented. Individuals with larger body masses seem to have higher blood pressure and also higher rates of hypertension^{311, 312}. The most significant modifiable risk factors for hypertension are overweight and obesity accounting for nearly 66% of the risk in many populations³¹³. For instance, in the Nurses Health Study (NHS), data showed that BMI values at 18 years and also midlife were positively correlated with hypertension. In this study, there was a significant association between long-term weight loss after the age of 18 years and a decreased risk of hypertension while weight gain after 18 years significantly increased this risk. This association continued to remain significant following the adjustments for current BMI which implied that BMI and history of weight

changes are independent predictors of hypertension³¹⁴. In the Framingham Study Moore et al.,³¹⁵ reported that when modest amounts of weight loss were sustained over an extended period of time it was associated with a 22% to 26% reduced risk of hypertension.

A variety of mechanisms have been postulated to help explain the relationship between obesity and hypertension. One such mechanism states that obesity and overweight causes renal structural changes that lead to tubular re-absorption and also sodium retention^{156, 316}. This further causes an elevation in arterial pressure which damages nephron function causing obesity, hypertension and renal damage¹⁵⁶.

1.2.14.4 **Environment**

Environmental factors can also play an important role in hypertension. Environmental factors that are implicated in the development of hypertension are salt intake, obesity, occupation, alcohol intake, family size and crowding. These listed factors are important for the development of high blood pressure with increasing age in the more affluent societies compared to the decline in blood pressure noticed in the less affluent societies³¹⁷. The environmental factor of all the above listed that has received the most attention is salt sensitivity. The cause of salt sensitivity in individuals is dependent on a number of factors namely: primary aldosterism, bilateral renal artery stenosis, renal parenchymal disease and low-renin essential hypertension which accounts for approximately half of the patients³¹⁷.

1.2.14.5 **Nutrition**

Lifestyle factors can also affect the development of hypertension. These factors include: excessive caloric consumption, excessive sodium consumption, excessive alcohol intake, inadequate potassium, inadequate calcium intake mostly in elderly African Americans,

inadequate physical activity, poor fitness level, and cigarette smoking³¹⁷. Diet related diseases and conditions related to hypertension include: obesity/overweight, diabetes mellitus, and dyslipidemias. In addition, adverse health outcomes that are associated with uncontrolled or poorly controlled hypertension are: cardiovascular disease, coronary heart disease, left ventricular hypertrophy, angina, myocardial infarction, congestive heart failure, nephropathy, cerebrovascular disease, transient ischemic attacks, stroke (CVA), peripheral artery disease (PAD), and retinopathy³¹⁷.

1.2.14.6 Pathophysiology of hypertension

Although the exact pathogenesis of primary hypertension remains to be elucidated, several hypotheses have been proposed. These include (1) increased blood volume, (2) inappropriate autoregulation, (3) overstimulation of sympathetic neural fibers in the heart and vessels, (4) water and sodium retention by the kidneys, and (5) hormonal inhibition of sodium-potassium transportation to kidneys and blood vessels. Researchers hypothesize that any of these mechanisms or a combination of more than one might be at work in the pathogenesis of primary hypertension³¹⁰.

Chronic hypertension can cause changes in the walls of the systemic blood vessels. Extended vasoconstriction along with high pressures within these vessels especially in the arteries and arterioles can cause the vessels to thicken and toughen to handle stress. These arterial smooth muscles further undergo cellular enlargement (hypertrophy) and cellular proliferation (hyperplasia). Later the tunica intima and the tunica media develop fibromuscular thickening that cause significant narrowing of the lumina. The injury of these vessels promotes biochemical mediators of inflammation that increase the permeability of the vascular endothelium. With the increased permeability humoral substances are absorbed into the vessel

walls causing further thickening and smooth muscle contraction ³¹⁰. The contraction of the smooth muscle cells is thought to be caused by a rise in the intracellular calcium concentration, which could possibly explain the vasodilatory effect of certain drugs that block calcium channels. Furthermore, this smooth muscle constriction of the arteriolar vessels causes structural changes leading to an irreversible rise in peripheral resistance ³⁰⁷.

In addition, it has been hypothesized that in the early stages of hypertension the peripheral resistance is not raised and that the elevation of the blood pressure is induced by the elevated cardiac output, which is associated with sympathetic over activity. This rise in peripheral arteriolar resistance could be considered a compensatory mechanism to prevent a raise in pressure being transmitted to the capillary bed where it could affect cell homeostasis ³⁰⁷.

1.2.14.7 **Reninangiotensin system**

The rennin angiotensin system is considered to be one of the most important of the endocrine system that regulates blood pressure. Renin is secreted by the juxtaglomerular system of the kidneys in reaction to glomerular underperfusion or decreased sodium intake. Renin is also emitted in response to stimulation by the sympathetic nervous system ³⁰⁷.

Renin acts on the rennin substrate angiotensinogen converting it to angiotensin I, which is rapidly converted in the lungs by angiotensin converting enzyme to angiotensin II. Angiotensin is a vasoconstrictor and causes a rise in blood pressure. It also stimulates the adrenal gland to release aldosterone which cause a further rise in blood pressure associated with sodium and water retention ³⁰⁷.

1.2.14.8 **Autonomic nervous system**

The stimulation of the sympathetic nervous system can induce constriction and dilatation of the arteriolar. The autonomic nervous system plays an important role in the management of blood pressure. The autonomic nervous system also regulates temporary variations in blood pressure that might be related to increases in stress levels and physical activity. It is postulated that hypertension is associated with the interaction of the autonomic nervous system, the rennin-angiotensin system and other factors for instance, sodium and circulating volume³⁰⁷.

1.2.14.9 **Cardiac output and peripheral resistance**

The ability to maintain a normal blood pressure is dependent on a balance in cardiac output and peripheral vascular resistance. Peripheral resistance is dependent on the walls of the small arterioles, which are comprised of smooth muscle cells. The continued constriction of the smooth muscle can cause structural changes in addition to the thickening of the arteriolar vessel walls possibly mediated by angiotensin leading to an irreversible rise in peripheral resistance. It has been hypothesized that in the very early stages of hypertension elevation of blood pressure is caused by elevated cardiac output that is related to sympathetic over-activity³⁰⁷.

1.2.14.10 **Sympathetic over-activity**

Individuals who were obese had higher serum catecholamine and muscle sympathetic nervous activity (MSNA) than lean individuals. MSNA was significantly higher in persons with central obesity compared to individuals with peripheral obesity³¹⁸. High resting heart rates and baroreflex dysfunction has been cited in the development of hypertension in the metabolic syndrome³¹⁹⁻³²¹.

Individuals who are obese have an activated rennin angiotensin system, which can bring on hypertension³²². This angiotensin system and the sympathetic nervous system are linked by positive feedback³²³. Possible agents involved in the stimulation of sympathetic nervous action in the metabolic syndrome are: elevated leptin, insulin resistance, and non-esterified fatty acids (NEFA)³²⁴. NEFA raises blood pressure, heart rate, and α 1-adrenoceptor vasoreactivity but also reduces baroreflex sensitivity, endothelium-dependent vasodilatation, and vascular compliance³²⁵.

Second, insulin resistance raises plasma leptin levels and it has been delineated that leptin increases sympathetic nervous activity suggesting that leptin-dependent sympathetic nervous activation could contribute to obesity associated hypertension³²⁶. There is also data suggesting that the metabolic syndrome is related to markers of adrenergic overdrive³²⁷.

1.2.14.11 **Oxidative stress and endothelial dysfunction**

In animal studies involving rats with the metabolic syndrome, following the consumption of high fat and refined sugars these animals had hypertension that was related to oxidative stress, nitric oxide inactivation, down regulation of nitric oxide synthase isoforms, and endothelial nitric oxide synthase activator³²⁸. This indicated that oxidative stress and endothelial dysfunction could be related to the development of hypertension in the metabolic syndrome. Moreover, current evidence indicate that oxidative stress, which is elevated in metabolic syndrome is related to sodium retention and sensitivity³²⁹.

In non-diabetics lipid peroxidation was correlated with body mass index and waist circumference suggesting that the accumulation of adipose tissue is correlated to oxidative stress in individuals³³⁰. Likewise, cross-sectional data from the Framingham offspring Study, showed that systemic oxidative stress is related with insulin resistance³³¹.

Insulin resistance causes an impairment in phosphatidylinositol 3-kinase (PI3K) which may cause an imbalance in the production of nitric oxide and the secretion of endothelin-1 further leading to endothelial dysfunction³³². Epidemiological studies show a reciprocal relationship between endothelial dysfunction leading to the development of hypertension and insulin resistance³³².

1.2.14.12 **Inflammatory mediators**

Abnormalities in the inflammatory markers have been reported as being related to the development of hypertension. For instance, in the Women Health Study, a positive relationship existed between elevated serum CRP levels and the development of hypertension³³³. A strong association between inflammation, hypertension, and the metabolic syndrome was likewise proposed by Grundy SM³³⁴.

Second, TNF- α is considered to play a key role in the pathophysiology of hypertension and the metabolic syndrome. TNF- α activates the production of endothelin-1 and angiotensinogen^{335, 336} and, TNF- α gene locus is hypothesized to play a role in insulin resistance mediated hypertension³³⁷. Serum TNF- α has been shown to be positively related to systolic blood pressure and insulin resistance³³⁸ and elevated TNF- α levels has been detected in monocytes from individuals with hypertension³³⁹.

Furthermore, studies have also shown that blood pressure is a significant and independent predictor of IL-6 levels in women³⁴⁰ and that IL-6 stimulates the central and the sympathetic nervous system which could potentially cause hypertension^{341, 342}. In women who were given IL-6 an elevation in heart rate, and serum norepinephrine levels were observed³⁴³. IL-6 causes an increase in plasma angiotensinogen and angiotensin II leading to the development of hypertension³⁴⁴.

1.2.15 Hypertension prevalence in Afro-Caribbeans

In a cohort of individuals from the Barbados Eye Study, more than half of the participants had hypertension at baseline with approximately one in two men and three in five women being affected by this disease. In this cohort hypertension increased with age and was similar for men and women between the age of 40-49; however, age specific rates were higher among women at older ages³⁴⁵. Age specific hypertension increased from 32.7% in men and 34.0% in women age 40-49 to 63.4% in men and 85.5% in women at age 80 and above with an overall prevalence in this population of 55.4 percent. Additionally, in this study hypertension awareness, treatment and control rates were 62.5, 53.8 and 18.5% respectively³⁴⁵.

Bobb-Liverpool et al.,³⁴⁶ reported that, in Jamaica hypertensive women had inadequate knowledge of hypertension, and poor compliance with JNCVI guidelines (Joint National Committee on Prevention, Detection and Evaluation and Treatment of High Blood Pressure). In this population, it was recommended that to improve patient compliance, counseling/education on diet, exercise, medication, and weight management be provided. In Trinidad and Tobago a study that evaluated health behavior patterns of chronic disease patients in relation to diet, weight control, and medication, determined that health professionals did not provide patients with the sufficient knowledge to adopt healthy lifestyles to prevent disease³⁴⁷.

Following age standardization similarly high rates of hypertension were found in other countries of the African Diaspora that mirror levels of western acculturation. In the French speaking country of the Antilles called Guadeloupe, the prevalence of hypertension was 24.7% for men and 22.1% for women. In this population the major contributing factor was once again obesity especially in the disadvantaged cohort that was evaluated³⁴⁸.

Similarly findings were confirmed from a combined study of the Birmingham factory screen, Birmingham INTERSALT volunteers and four West-Midlands churches. In this cohort of N=2853, Afro-Caribbean's had a significantly higher systolic blood pressure with higher diastolic blood pressure in the Afro-Caribbean women. Even following the adjustment of age, BMI, smoking, and alcohol intake the odds ratios and 95% confidence interval for being hypertensive were 1.56 (1.14 to 2.13; P=0.005) and 2.40 (1.51 to 3.81; P=0.0002) in Afro-Caribbean men and women respectively³⁴⁹.

1.2.16 Prevalence of hypertension in Trinidad

In Trinidad the incidence of hypertension and non-insulin dependent diabetes mellitus in African and Indian populations residing in St. James did not differ significantly according to ethnicity. In a cohort comprised of 2,491 men and women age 35 to 69 years of age, incidence rates of hypertension ranged between 33 and 41 per 1000 persons in men and 27 and 32 per 1000 in person years in women³⁵⁰.

Nichols et al.,¹⁰ reported that blood pressure in Tobagonian adolescent population was found to be significantly higher than children residing in the U.S.A using US reference level cut points for systolic and diastolic blood pressure. The prevalence of elevated diastolic blood pressure was almost twice the expected levels using US cut points. The results of the Tobagonian study indicated that Tobagonian adolescents were heavier than adolescence residing in the United States, United Kingdom and Jamaica. Blood pressure in this study was associated with a family history of hypertension and being overweight.

2.0 PUBLIC HEALTH SIGNIFICANCE

Prostate cancer is one of the most common malignancies and the second leading cause of cancer related deaths in the United States and other countries³⁵¹. African American men have been found to have the highest incidence rates for prostate cancer of all racial and ethnic groups³⁵¹. High rates of prostate cancer have also been observed in Afro-Tobgonian men^{35, 42}. Screening detected prostate cancer prevalence rates were 4.9% for individuals aged 50-55, 7.7% for individuals ages 55-59; and 13.3% for individuals aged 60-64 years³⁵. Even though the reasons for the large disparity in prostate cancer risk are not fully understood, one possible explanation is androgen levels³⁵¹. A common treatment for prostate cancer is androgen deprivation therapy (ADT) which reduces the levels of androgen in the body but also has many side effects³⁵². Consequently, prostate cancer is a major public health concern and especially in persons of African ancestry. Further studies are needed to investigate the effects of treatments for prostate cancer such as ADT in at risk populations.

Obesity is also a major public health concern. Individuals who are considered minority/non-white are more prone to increases in weight and its associated complications³⁵³. World wide there are approximately a billion adults that are considered overweight and 300 million that are considered obese³⁵³. In the United States the prevalence of obesity in African Americans between 2003 and 2004 was 76.1%³⁵⁴. According to a 2003-2004 health survey that

was conducted in England Black Caribbean men had the highest prevalence rates of obesity compared to Bangladeshi, Chinese, Pakistani, Black African and Indian men³⁵⁵.

Hypertension is also a disease that affects people of African descent and is of major public health concern. In the United States the prevalence of hypertension is highest amongst black Americans³⁵⁶. Individuals of African descent appear to develop high blood pressure much earlier in life, have higher average blood pressures and experience poorer outcomes³⁵⁷.

Obesity and hypertension are multi-factorial in origin and the development of these diseases requires interplay between genes and the environment^{356, 358, 359}. Therefore, to elucidate the relationship between high blood pressure, obesity and candidate genes, we propose to investigate the association between *ADRB₂*, blood pressure, and obesity in Afro-Tobagonian men. In addition, we also propose to investigate the association between *AR CAG* and obesity in Afro-Tobagonian men. We hope that these studies will provide important clues into the etiology of aforementioned diseases. Moreover, we hope that these studies will increase our knowledge in understanding the role race/ethnicity plays as a risk factor, the identification of treatments that target specific populations, and the selection of appropriate therapies.

3.0 SYNTHESIS STATEMENT

This paper attempts to elucidate complex phenotypes such as body weight and blood pressure in a cohort of Afro-Caribbean men who reside on the island of Tobago. Research on obesity and blood pressure has revealed that genetics, diet, environment, behavior, and social norms play important roles in disease etiology. The multi-factorial nature of obesity and blood pressure and the involvement of many genes have made the search for candidate genes exceedingly difficult. These papers seek to further understand the role *ADRB2* and *AR* CAG repeats plays in the development of obesity related phenotypes and blood pressure.

A common treatment for prostate cancer (Prostate cancer) is androgen deprivation therapy (ADT). ADT is typically considered the treatment of choice for metastatic prostate cancer. Androgens are important for physiologic activity of myriad bodily functions. However, ADT administration has several deleterious side effects one of which is altered body composition. This paper also investigates the effect of ADT administration in Prostate cancer individuals on the rate of change of percent total body fat (DEXA) from baseline to follow-up in Afro-Tobagonian men. We hope that these studies will strength our understanding of the association between genes and our primary outcomes and secondly in explaining whether the rate of change of percent total body fat increases or decreases with ADT exposure.

4.0 PAPER 1: ADRB2 GENE VARIANTS, DEXA BODY COMPOSITION, AND HYPERTENSION IN TOBAGO MEN OF AFRICAN DESCENT

Tracey Samantha Beason MSPH,¹ Clareann H. Bunker PhD,¹ Joseph M. Zmuda PhD,¹ John W. Wilson PhD,² Alan L. Patrick MD FRCP,³ Victor W. Wheeler MBBS MRCOG,³ and Joel L. Weissfeld MD MPH¹

1. Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, PA, USA
2. Department of Biostatistics, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, PA, USA
3. Tobago Health Studies Office, Scarborough, Tobago, Trinidad and Tobago

Acknowledgements

This study was supported by grant R01-AR049747 from the National Institute of Arthritis and Musculoskeletal and Skin Diseases, grant R01-CA84950, U.S. National Cancer Institute, and by support from the Division of Health and Social Services, Tobago House of Assembly, and the University of Pittsburgh Cancer Institute.

Objective: Classic tissue effects of β_2 adrenergic receptor activation include skeletal muscle glycogenolysis and vascular smooth muscle relaxation, factors relevant to obesity and hypertension, respectively. In a population-based study, we examined two common amino acid substitutions in the β_2 adrenergic receptor gene (ADRB2) in relation to body composition and blood pressure.

Design and subjects: Cross-sectional analysis of 1965 African descent men living in Tobago and participating in a prostate cancer screening study.

Measurements: Body mass index (BMI), waist circumference, blood pressure, dual energy X-ray absorptiometry (DEXA) body composition, and ADRB2 (Arg16Gly; Gln27Glu) genotype.

Results: Twenty-five percent were obese (BMI \geq 30 kg/m²) and 50% hypertensive. ADRB2 Arg16Gly and Gln27Glu alleles were in linkage disequilibrium ($r^2=0.96$). ADRB2 16Gly - containing and 27Glu-containing genotypes were equally frequent in low, medium, and high tertiles of percentage body fat mass (16Gly-containing genotypes: 73.2%, 74.4%, 75.1%, $p_{trend}=0.45$; 27Glu-containing genotypes: 27.8%, 24.2%, 26.4%, $p_{trend}=0.58$) and in normal blood pressure, pre-hypertensive, and hypertensive men (16Gly-containing genotypes: 73.6%, 72.5%, 74.6%, $p_{trend}=0.55$; 27Glu-containing genotypes: 26.1%, 24.6%, 27.1%, $p_{trend}=0.51$).

Conclusion: In a high obesity and high hypertension risk population with ancestry in common with African-Americans, genetic variation defined by two common ADRB2 amino acid substitutions was not associated with body composition or hypertension.

Key words: African ancestry, adrenergic β_2 receptor, genetic polymorphism, body composition, blood pressure, hypertension

4.2 INTRODUCTION

Interacting with diet and physical activity, genetic factors contribute to obesity and hypertension.[1-3] Frequently studied candidate genes include the leptin,[4] catecholamine,[5] and peroxisomeproliferator-activated receptors [6] and genes in the renin-angiotensin system.[7] Stimulating β adrenergic receptor function promotes lipolysis in fat cells.[2, 8] The β 2 adrenergic receptor gene (ADRB2) attracts interest because catecholamine activation of the β 2 adrenergic receptor regulates skeletal muscle glycogenolysis and vascular smooth muscle relaxation.[9] The ADRB2 single nucleotide polymorphisms most often studied in human populations substitute glycine for arginine at codon 16 (Arg16Gly) and glutamic acid for glutamine at codon 27 (Gln27Glu).[10]

Despite numerous published studies, the connection, if any, between ADRB2 Arg16Gly or Gln27Glu genotype and metabolic phenotypes, including obesity and hypertension, is tentative. Results from early recombinant cell culture experiments,[11] for example, launched a long standing hypothesis asserting that the ADRB2 16Gly variant promoted obesity and hypertension by causing receptor down regulation and physiologic desensitization in response to chronic agonist stimulation.[12] However, subsequent human experiments showed agonist-induced desensitization only in persons with the ADRB2 16Arg variant.[12-14] A 2008 meta-analysis examined obesity risk in relation to ADRB2 codon 16 variation (13 studies with 6825 subjects) and in relation to ADRB2 codon 27 variation (28 studies with 14,450 subjects).[10] Random effects meta-regression analysis showed statistically insignificant obesity risks in

relation to 16Arg-containing genotype (odds ratio 1.02, 95% confidence interval 0.89-1.18) and in relation to 27Glu-containing genotype (odds ratio 1.11, 95% confidence interval 0.98-1.27). However, meta-analysis restricted to Asian (7 studies with 3575 subjects), Pacific Island (1 study with 1020 subjects), and native South American populations (1 study with 149 subjects) showed statistically significant association between 27Glu-containing genotype and obesity (odds ratio 1.46, 95% confidence interval 1.02-2.10). One study [15] with African-American subjects, but not a second,[16] showed association between obesity measures and ADRB2 genotype.

We defined a large population on the Caribbean island of Tobago with ancestry in common with African-Americans, determined ADRB2 Arg16Gly and Gln27Glu genotypes, measured blood pressure, and used dual energy X-ray absorptiometry (DEXA) to estimate body composition. Analyses examined ADRB2 genotypes in relation to body composition and hypertension.

4.3 METHODS AND MATERIAL

Between September 1997 and September 2007, the Tobago Prostate Study used public service announcements, flyers, local health care workers, and word of mouth to solicit 40-79 year-old men in Tobago for participation in a study of periodic prostate cancer screening.[17] The study excluded non-ambulatory, terminally ill, or cognitively impaired men. Participants signed written informed consent. The Institutional Review Boards of the University of Pittsburgh and Tobago Division of Health and Social Services approved the research protocol.

The Tobago Prostate Study enrolled 3837 men. Standardized entry questionnaires provided demographic and basic cancer risk factor information. To support studies of obesity,

hypertension, diabetes, and musculoskeletal health, men enrolling or returning after January 2004, also completed a detailed standardized staff-administered health history questionnaire. We defined, for the current analysis, a race-eligible study group that consisted of 3452 (90.0% of 3837) men who reported having at least three grandparents of African descent.

Procedures included periodic musculoskeletal tests (DEXA and/or quantitative computed tomography) and, for the subset of men who completed the health history questionnaire and accepted musculoskeletal testing, genetic tests. Information from DEXA was available for 2850 (82.6%) and health history questionnaires for 2259 (65.4%) of 3452 race-eligible men and ADRB2 Arg16Gly and/or Gln27Glu genotypes for 1960 of 2133 (91.9%) race-eligible men with both health history questionnaires and DEXA. Data analyses included 1965 (1960 with and 5 without DEXA) race-eligible ADRB2 genotyped men with health history questionnaires. When compared against the group of men not available for the health history questionnaire (n=1193) or not otherwise genotyped (n=294), the 1965 men included in analyses enrolled at younger age (median 52 vs. 56 years, $p(\text{Wilcoxon}) < 0.0001$). Genotyped and non-genotyped race-eligible men available for the health questionnaire had statistically similar body mass index (BMI), waist circumference, hypertension prevalence, and body composition (percent body fat).

ADRB2 Arg16Gly (rs1042713) and Gln27Glu (rs1042714) genotype determinations used a TaqMan allelic discrimination assay performed on an Applied Biosystems (Foster City, CA) 7900HT Fast Real-Time PCR System. Call rates for the ADRB2 Arg16Gly and Gln27Glu polymorphisms were 97.0% and 94.5%, respectively. ADRB2 Arg16Gly, Gln27Glu, and both Arg16Gly and Gln27Glu genotypes were available for 1856 (94.5%), 1871 (95.2%), and 1762 (89.7%) men included in data analyses, respectively. Genotype distributions for ADRB2 Arg16Gly and Gln27Glu satisfied Hardy-Weinberg equilibrium conditions ($p > 0.05$). Allele

frequencies (16Gly 49.6%, standard error 0.8% and 27Glu 14.1%, standard error 0.6%) estimated for the 1965 men included in data analyses agreed with values reported by the International HapMap Project (www.hapmap.org) for the Yoruba population (50.0% and 17.5%, respectively). ADRB2 Arg16Gly and Gln27Glu alleles were in linkage disequilibrium ($D=0.96$).

4.3.1 Outcome measurements

Outcome measurements included height (measured without shoes to the nearest 0.1 cm on a wall-mounted stadiometer), weight (measured without shoes to the nearest 0.1 kg on a balance beam scale), BMI (calculated as weight in kg divided by height in meters-squared), and waist circumference (measured in cm at the umbilicus with an inelastic tape measure). Percent body fat was acquired on a Hologic QDR 4500W DEXA operated in array beam mode and analyzed with QDR software version 8.26a (Hologic Inc. Bedford, MA). After seating and resting subjects for 5 minutes, technicians selected an appropriate cuff size and used an automated OMRON HEM705CP sphygmomanometer (Omron Healthcare, Inc., Vernon Hills, IL) to obtain three consecutive blood pressure measurements.

4.3.2 Statistical analysis

Using the second and third measurements, if available, or any two available measurements, otherwise, analyses calculated average values for systolic (SBP) and diastolic blood pressure (DBP). Using these measurements, we classified subjects into three mutually exclusive hypertension categories, hypertension (SBP > 140 mmHg or DBP > 90 mmHg or current antihypertensive medication use), pre-hypertension (SBP 120-139 mmHg or DBP 80-89 mmHg and current antihypertensive medication nonuse), and normal (SBP < 120 mmHg and DBP < 80 mmHg and current antihypertensive medication nonuse). Sixty-six (3.3%) men were not

classified because of missing blood pressure measurements (n=8) and/or missing or inconsistent self-reports of antihypertensive medication use (n=64). BMI classification used World Health Organization cutpoints (underweight <18.5 kg/m², normal weight 18.5- 24.9 kg/m², overweight 25-29.9 kg/m², and obese ≥30 kg/m²).

Statistical analyses (SAS 9.2; SAS Institute Inc., Cary, NC) used conventional methods (chi-square or Fisher exact tests for discrete data and t-tests, ANOVA, or Wilcoxon rank-sum tests for continuous data) to evaluate the significance of group differences. Haplotype-based analyses used the EM algorithm implemented in SAS Genetics (PROC HAPLOTYPE) to estimate group-level haplotype frequencies and to generate subject-level haplotype probability weights.¹ We then used traditional linear models (implemented in SAS PROC GENMOD with normal probability distribution and identity link function) to estimate independent associations between the haplotype probability weights and continuous outcomes (BMI and total body percent body fat) and logistic regression (implemented in SAS PROC LOGISTIC) to estimate independent associations between the haplotype probability weights and binary outcomes (hypertension or pre-hypertension vs. normal blood pressure).

4.4 RESULTS

Study men were mean age 58.9 years (standard deviation (SD) 10.4 years). Over half (50.4%) were hypertensive, 32.5% pre-hypertensive, 44.1% overweight, and 25.5% obese. One in 6 (16.5%) self-reported a physician diagnosis of diabetes.

¹http://support.sas.com/documentation/cdl/en/geneug/59659/HTML/default/geneug_haplotype_sect021.htm

Genotype frequencies were 25.9% Arg/Arg, 49.1% Arg/Gly, and 25.0% Gly/Gly at codon 16 (n=1856) and 74.0% Gln/Gln, 23.8% Gln/Glu, and 2.2% Glu/Gly at codon 27 (n=1871). The prevalence of obesity (BMI ≥ 30 kg/m²) did not vary according to genotype (27.3% of 480, 24.6% of 908, 25.2% of 465 men with the codon 16 Arg/Arg, Arg/Gly, and Gly/Gly genotype, $p=0.5331$, and 25.9% of 1382, 26.1% of 445, and 24.4% of 41 men with the codon 27 Gln/Gln, Gln/Glu, and Glu/Glu genotype, $p= 0.9728$). Height, weight, and waist circumference were slightly higher in men with a 16Arg allele (Table 4-1). Otherwise, weight, waist circumference, BMI, and percent body fat were unrelated to ADRB2 genotype (Table 4-1). In the obese subset, body measurements did not vary in any systematic or meaningful way with respect to ADRB2 genotype (Table 4-1).

ADRB2 16Gly-containing and 27Glu-containing genotypes were equally frequent in low, medium, and high tertiles of percent body fat (Table 4-2; 16Gly-containing genotypes: 73.2%, 74.4%, 75.1%, $ptrend=0.45$; 27Glu-containing genotypes: 27.8%, 24.2%, 26.4%, $ptrend=0.58$) and in normal blood pressure, pre-hypertensive, and hypertensive men (Table 4-3; 16Gly-containing genotypes: 73.6%, 72.5%, 74.6%, $ptrend=0.55$; 27Glu-containing genotypes: 26.1%, 24.6%, 27.1%, $ptrend=0.51$). In men not taking antihypertensive medication, systolic, diastolic, and mean arterial blood pressure values were unrelated to ADRB2 genotype (data not shown).

Estimated ADRB2 Arg16Gly Gln27Glu haplotype frequencies were Arg-Gln 50.0% (standard error 0.8%), Gly-Gln 35.9% (standard error 0.8%), Gly-Glu 13.8% (standard error 0.6%), and Arg-Glu 0.3% (standard error 0.1%). The Figure 4-1 uses box plots to summarize BMI and percent body fat distributions in men classified according to most probable haplotype combination. In linear models, the less common Gly-Gln and Gly-Glu haplotypes associated with lower BMI and lower percent body fat (Table 4-4). In obese men, these haplotypes associated

with higher percent body fat and lower waist circumference (Table 4-4). However, none of these associations, including the apparent interaction between haplotype and obesity grouping (data not shown), was statistically significant. Finally, logistic regression did not show statistically significant ADRB2 haplotype associations with either pre-hypertension or hypertension (data not shown).

4.5 DISCUSSION

Mediating fat cell lipolysis [2, 8] and vascular smooth muscle relaxation,[9] ADRB2 gene variants that code for functionally altered receptors could promote weight gain or high blood pressure. In a study of 1965 Tobago men of African ancestry, however, we observed no significant body composition or blood pressure associations with the two most commonly studied ADRB2 gene variants.

Different investigations have reached different conclusions about the significance of ADRB2 genotype in relation to obesity and related phenotypes [10, 18-20]. Variability related to sex may explain inconsistencies in the published literature. Some authors, including Corbalan et al. [21] and GonzálezSánchez et al. [22], suggest that ADRB2 genotype determines not obesity, but obese subtype, perhaps in only one sex group. For example, in a Spanish clinic-based study with 40 men and 199 women, Corbalan et al. [21] compared the ADRB2 Gln27Glu genotypes of subjects with either abdominal obesity (BMI >30 kg/m² and waist-to-hip ratio >0.85) or normal body mass (BMI <25 kg/m² and waist-to-hip ratio <0.85). In men, but not women, 27Glu-containing genotypes were more frequent in the group with abdominal obesity. In a Spanish population-based study with 319 white men and 347 white women, GonzálezSánchez et al. [22]

noted a statistically non-significant trend in men (but not women) between ADRB2 Gln27Glu genotype and obesity prevalence (20.0% of 130, 27.7% of 155, and 29.4% of 34 men with the codon 27 Gln/Gln, Gln/Glu, and Glu/Glu genotype, respectively, $p=0.2572$). In the obese subset, men with the rare Glu/Glu genotype had significantly higher mean BMI (34.1 kg/m², $n=10$) than men with either the Gln/Glu (31.9 kg/m², $n=46$, $p=0.013$) or Gln/Gln genotype (32.0 kg/m², $n=26$, $p=0.023$). To support specific association with abdominal obesity, GonzálezSánchez et al. [22] noted that obese men with the rare Glu/Glu genotype also had significantly higher mean sagittal abdominal diameter (27.8 cm, $n=10$) than obese men with either the Gln/Glu (24.9 cm, $n=46$, $p=0.037$) or Gln/Gln genotype (24.9 cm, $n=26$, $p=0.062$). Among obese women, sagittal abdominal diameter was not related to ADRB2 Gln27Gly genotype. In agreement with GonzálezSánchez et al., we observed no statistical association between obesity prevalence and ADRB2 genotype. Among obese Afro-Caribbean men, BMI, waist circumference, and body fat values were unrelated to ADRB2 genotype (Table 4-1). Simply, our results do not support a relationship, specifically in obese men, between ADRB2 genotype and obesity subtype.

Excluding persons treated for high blood lipids, high blood pressure, or diabetes, Meirhaeghe et al. [23] studied a population-based sample of 836 35-64 year-old urban-dwelling persons from northern France (419 men and 417 women, 14.2% obese overall) and observed sex-specific associations between ADRB2 genotype and body composition. Body weight, BMI, waist circumference, hip circumference, and waist-to-hip ratio mean values were significantly higher in men with the codon 16 Arg/Arg genotype than men with either the Arg/Gly or Gly/Gly genotype and significantly higher in men with the codon 27 Glu/Glu genotype than men with either the Gln/Glu or Gln/Gln genotype. In women, differences were not statistically significant. Observing linkage disequilibrium between 16Arg and 27Gln, Meirhaeghe et al. [23] compared,

in men, body composition measures according to combined genotypes at Arg16Gly and Gln27Gly. With the GlyGluhaplotype serving as reference, higher body weight, BMI, and waist-to-hip ratio associated with the ArgGlnhaplotype, not with the GlyGlnhaplotype. In Afro-Caribbean men, body weight and waist circumference were slightly higher in men with a 16Arg allele (Arg/Arg or Arg/Gly genotype) than men without a 16Arg allele (Gly/Gly genotype; Table 1). However, BMI and percent body fat did not vary according to Arg16Gly genotype. Body weight, waist circumference, BMI, and percent body fat were completely independent of Gln27Glu genotype.

Only two studies [15, 16] to date have included a meaningful number of African-Americans. The Insulin Resistance and Atherosclerosis (IRAS) Family Study [15] genotyped 272 African Americans and 720 Hispanic Americans from 18 and 45 families, respectively, and used single-slice computed tomography to measure visceral and subcutaneous fat. Obesity measures associated with Gln27Glu, specifically the Glu/Glu genotype, but not with Arg16Gly genotype. High visceral fat area associated with the Glu/Glu genotype, even after control for BMI. Results for men and women and for African and Hispanic Americans were statistically indistinguishable. The Heritage Family Study included 274 African-Americans (31.8% obese) and 502 whites (19.3% obese) and used under water weighing to measure fat mass, single-slice computed tomography to measure visceral and subcutaneous fat, and skin calipers to measure skin-fold thickness[16]. In white obese ($BMI \geq 30$ kg/m²) subjects only, the Heritage Family Study observed lower fat mass in obese white men with 27Glu-containing genotypes and lower fat mass in obese white women with 16Gly-containing genotypes. In African-American subjects, the Heritage Family Study did not report any statistically significant cross-sectional associations between ADRB2 genotype and any body composition measure.

One study of ADRB2 genotype included Afro-Caribbeans recruited from primary care clinics located on St. Vincent [24]. The case group included 136 patients (19.9% men) with high blood pressure (DBP > 95 mmHg or antihypertensive medication use) and family history of hypertension. The control group included 81 patients (46.9% men) with normal blood pressure (DBP < 85 mmHg and antihypertensive medication nonuse) and no family history of hypertension. The 16Gly allele was much more frequent in the case group (84.6% vs. 66.7%; $p=0.000014$). Our community-based study of 1965 Afro-Caribbean men from Tobago included 331 and 557 persons who satisfied the St. Vincent case and control definitions. In Tobago, 16Gly was also more frequent in cases, 51.5% vs. 48.7%. However, the difference was not statistically significant ($p(\text{allele chi-square})=0.2594$). In the St. Vincent study, genotype frequencies in both the case and control groups violated Hardy-Weinberg equilibrium. The control frequency (66.7%) of the putative risk allele (16Gly) in St. Vincent was high when compared against reference frequencies in the HapMap Yoruba (50.0%) and Tobago populations (49.5%). Therefore, selection bias and genotyping error plausibly explain the anomalous St. Vincent results.

Our study participants volunteered for prostate cancer screening, in many instances survived a variable duration after study entry, and agreed to extra visits that included measurements, such as DEXA. Therefore, absence of meaningful association between ADRB2 genotype and body composition could occur if these study selection factors preferentially excluded men with certain genotype-phenotype combinations. However, selection bias deriving from factors related to initial study participation can be discounted to the extent that Tobago Prostate Study, as a whole, enrolled a high proportion (60%) of age-eligible men (approximately 5000) living in Tobago. To address possible survival bias, we compared the 1727 and 238 men

who had body composition measurements after study entry and coincident with study entry, respectively, and found statistically similar relationships between ADRB2 genotype and body composition (data not shown). Finally, study procedures captured a reasonable proportion (1965 of 3452, 56.9%) of race-eligible Tobago Prostate Study enrollees.

Results from some studies suggest specific ADRB2 association with regional fat distribution [15, 21-23]. Our primary body composition measures, BMI and DEXA-derived percent body fat, do not distinguish between visceral and subcutaneous fat. However, our data set included waist circumference, an indirect measure of central obesity. Waist circumference and ADRB2 genotype were statistically independent (Table 4-1).

Finally, we evaluated only two ADRB2 genetic variants. These variants change the amino acid sequence of the β 2 adrenergic receptor protein. However, these changes are not generally believed to alter agonist binding or signal transduction [12]. The Arg16Gly and Gln27Glu variants may affect function secondarily, perhaps through agonist-induced receptor protein down regulation or linkage disequilibrium with other less common but more influential variants [12].

In a large racially homogenous population of men with black African ancestry and high prevalence of obesity and hypertension, single variant and haplotype-based analyses did not show meaningful or consistent association between ADRB2 Arg16Gly and Gln27Glu variation and phenotypes related to obesity and hypertension.

1. Liu Z-q, Mo W, Huang Q, Zhou H-h. Genetic polymorphisms of human beta-adrenergic receptor genes and their association with obesity. *Zhong Nan daXueXueBao Yi Xue Ban*. Jun 2007;32(3):359-367.
2. Arner P. Genetic variance and lipolysis regulation: implications for obesity. *Ann Med*. Nov 2001;33(8):542-546.
3. Marti A, Martinez-Gonzalez MA, Martinez JA. Interaction between genes and lifestyle factors on obesity. *Proc Nutr Soc*. Feb 2008;67(1):1-8.
4. de Silva AM, Walder KR, Boyko EJ, et al. Genetic variation and obesity in Australian women: a prospective study. *Obes Res*. Dec 2001;9(12):733-740.
5. Arner P, Hoffstedt J. Adrenoceptor genes in human obesity. *J Intern Med*. Jun 1999;245(6):667-672.
6. Lindi V, Sivenius K, Niskanen L, Laakso M, Uusitupa MI. Effect of the Pro12Ala polymorphism of the PPAR-gamma2 gene on long-term weight change in Finnish non-diabetic subjects. *Diabetologia*. Jul 2001;44(7):925-926.
7. Strazzullo P, Iacone R, Iacoviello L, et al. Genetic variation in the renin-angiotensin system and abdominal adiposity in men: the Olivetti Prospective Heart Study.[Summary for patients in *Ann Intern Med*. 2003 Jan 7;138(1):I26; PMID: 12513065]. *Ann Intern Med*. Jan 7 2003;138(1):17-23.
8. Large V, Hellstrom L, Reynisdottir S, et al. Human beta-2 adrenoceptor gene polymorphisms are highly frequent in obesity and associate with altered adipocyte beta-2 adrenoceptor function. *J Clin Invest*. Dec 15 1997;100(12):3005-3013.
9. Westfall TC, Westfall DP. Chapter 6. Neurotransmission: The Autonomic and Somatic Motor Nervous Systems (Table 6-6). *Goodman & Gilman's The Pharmacological Basis of Therapeutics, 11e* [<http://www.accessmedicine.com/content.aspx?aID=954433>. Accessed February 19, 2010].
10. Jalba MS, Rhoads GG, Demissie K. Association of codon 16 and codon 27 beta 2-adrenergic receptor gene polymorphisms with obesity: a meta-analysis. *Obesity*. Sep 2008;16(9):2096-2106.
11. Green SA, Turki J, Innis M, Liggett SB. Amino-terminal polymorphisms of the human beta 2-adrenergic receptor impart distinct agonist-promoted regulatory properties.[Erratum appears in *Biochemistry* 1994 Nov 29;33(47):14368]. *Biochemistry*. Aug 16 1994;33(32):9414-9419.
12. Brodde O-E. Beta1- and beta2-adrenoceptor polymorphisms and cardiovascular diseases. *Fundam Clin Pharmacol*. Apr 2008;22(2):107-125.
13. Bruck H, Leineweber K, Park J, et al. Human beta2-adrenergic receptor gene haplotypes and venodilation in vivo. *Clin Pharmacol Ther*. Sep 2005;78(3):232-238.
14. Dishy V, Sofowora GG, Xie HG, et al. The effect of common polymorphisms of the beta2-adrenergic receptor on agonist-mediated vascular desensitization. *N Engl J Med*. Oct 4 2001;345(14):1030-1035.

15. Lange LA, Norris JM, Langefeld CD, et al. Association of adipose tissue deposition and beta-2 adrenergic receptor variants: the IRAS family study. *Int J Obes*. May 2005;29(5):449-457.
16. Garenc C, Perusse L, Chagnon YC, et al. Effects of beta2-adrenergic receptor gene variants on adiposity: the HERITAGE Family Study. *Obes Res*. May 2003;11(5):612-618.
17. Bunker CH, Patrick AL, Konety BR, et al. High prevalence of screening-detected prostate cancer among Afro-Caribbeans: the Tobago Prostate Cancer Survey. *Cancer Epidemiol Biomarkers Prev*. Aug 2002;11(8):726-729.
18. Hayakawa T, Nagai Y, Kahara T, et al. Gln27Glu and Arg16Gly polymorphisms of the beta2-adrenergic receptor gene are not associated with obesity in Japanese men. *Metabolism*. Sep 2000;49(9):1215-1218.
19. Hellstrom L, Large V, Reynisdottir S, Wahrenberg H, Arner P. The different effects of a Gln27Glu beta 2-adrenoceptor gene polymorphism on obesity in males and in females. *J Intern Med*. Mar 1999;245(3):253-259.
20. Ishiyama-Shigemoto S, Yamada K, Yuan X, Ichikawa F, Nonaka K. Association of polymorphisms in the beta2-adrenergic receptor gene with obesity, hypertriglyceridaemia, and diabetes mellitus. *Diabetologia*. Jan 1999;42(1):98-101.
21. Corbalan MS, Marti A, Forga L, Martinez-Gonzalez MA, Martinez JA. Beta(2)-adrenergic receptor mutation and abdominal obesity risk: effect modification by gender and HDL-cholesterol. *Eur J Nutr*. Jun 2002;41(3):114-118.
22. Gonzalez Sanchez JL, Proenza AM, Martinez Larrad MT, et al. The glutamine 27 glutamic acid polymorphism of the beta2-adrenoceptor gene is associated with abdominal obesity and greater risk of impaired glucose tolerance in men but not in women: a population-based study in Spain. *ClinEndocrinol (Oxf)*. Oct 2003;59(4):476-481.
23. Meirhaeghe A, Helbecque N, Cottel D, Amouyel P. Impact of polymorphisms of the human beta2-adrenoceptor gene on obesity in a French population. *Int J ObesRelatMetabDisord*. Mar 2000;24(3):382-387.
24. Kotanko P, Binder A, Tasker J, et al. Essential hypertension in African Caribbeans associates with a variant of the beta2-adrenoceptor. *Hypertension*. Oct 1997;30(4):773-776.

Table 4-1: Body composition measures (means \pm standard deviations) according to *ADRB2* genotype.

Body composition measure	Arg16Gly				Gln27Glu			
	Arg/Arg	Arg/Gly	Gly/Gly	p-value [1]	Gln/Gln	Gln/Glu	Glu/Glu	p-value [1]
<i>All men</i>								
n [2]	477–480	905–910	461–465		1379–1383	441–446	41	
Height (cm)	175.7 \pm 7.1	175.0 \pm 6.9	174.5 \pm 6.6	0.0175	174.8 \pm 6.8	175.4 \pm 7.1	175.5 \pm 6.6	0.2534
Weight (kg)	84.9 \pm 15.0	84.8 \pm 17.1	82.8 \pm 15.6	0.0528	84.6 \pm 16.2	84.7 \pm 17.3	84.0 \pm 14.5	0.9733
Waist circumference (cm)	93.4 \pm 12.7	93.5 \pm 11.3	92.1 \pm 11.3	0.0798	93.3 \pm 11.6	93.1 \pm 11.3	92.2 \pm 8.8	0.7911
Body mass index (BMI; kg/m ²)	27.5 \pm 4.4	27.7 \pm 5.1	27.2 \pm 4.7	0.1894	27.7 \pm 4.9	27.5 \pm 5.1	27.2 \pm 3.9	0.6795
Total body fat (%) [3]	20.9 \pm 6.4	21.3 \pm 6.3	20.7 \pm 6.3	0.1963	21.2 \pm 6.3	21.0 \pm 6.5	21.5 \pm 6.0	0.7455
<i>Obese men (BMI \geq 30 kg/m²)</i>								
n [2]	129-131	221-223	116-117		354-358	114-116	10	
Height (cm)	174.9 \pm 6.6	175.3 \pm 6.2	174.0 \pm 6.7	0.2235	174.6 \pm 6.3	175.3 \pm 7.2	177.5 \pm 6.7	0.2709
Weight (kg)	101.2 \pm 12.4	104.8 \pm 17.2	101.1 \pm 12.8	0.0307	103.0 \pm 14.7	103.8 \pm 17.8	102.0 \pm 8.3	0.8557
Waist circumference (cm)	105.5 \pm 9.8	106.6 \pm 9.5	104.8 \pm 8.2	0.2082	106.1 \pm 9.6	105.9 \pm 8.2	101.6 \pm 4.0	0.3093
Body mass index (BMI; kg/m ²)	33.1 \pm 3.1	34.1 \pm 5.3	33.4 \pm 3.3	0.0704	33.8 \pm 4.5	33.7 \pm 4.9	32.4 \pm 1.5	0.6315
Total body fat (%) [3]	25.9 \pm 5.2	26.6 \pm 4.8	26.1 \pm 4.4	0.3392	26.3 \pm 4.7	26.6 \pm 5.0	25.7 \pm 4.0	0.7848

1. Statistical significance (ANOVA) of differences in mean values across genotype categories
2. Sample numbers vary because of missing attribute data
3. Total body (except head) fat mass expressed as a percentage of total body (except head) mass

Table 4-2: *ADRB2* genotype and body composition.

<i>ADRB2</i> genotype	Percent Total Body Fat [1]						
	Low tertile N=653		Middle tertile N=654		High tertile N=653		
	N	%	N	%	N	%	
Arg16Gly							
Arg/Arg	165	26.8	159	25.6	153	24.9	$P_{\text{global}}=0.72$
Arg/Gly	289	46.9	307	49.4	313	51.0	
Gly/Gly	162	26.3	155	25.0	148	24.1	
Arg/Gly or Gly/Gly	451	73.2	462	74.4	461	75.1	$P_{\text{trend}}=0.45$
Gln27Glu							
Gln/Gln	439	72.2	471	75.8	469	73.6	$P_{\text{global}}=0.31$
Gln/Glu	153	25.2	142	22.9	151	23.7	
Glu/Glu	16	2.6	8	1.3	17	2.7	
Gln/Glu or Glu/Glu	169	27.8	150	24.2	168	26.4	$P_{\text{trend}}=0.58$

1. Percent total body fat (total body (except head) fat mass expressed as a percentage of total body (except head) mass) -- Low tertile: 4.769-18.824; Middle tertile: 18.825-23.819; High tertile: 23.820-44.352

Table 4-3: *ADRB2* genotype and hypertension.

<i>ADRB2</i> genotype	Normal blood pressure N=324		Pre-hypertensive N=617		Hypertensive N=958		
	N	%	N	%	N	%	
Arg16Gly							
Arg/Arg	81	26.4	160	27.5	229	25.4	$P_{\text{global}}=0.90$
Arg/Gly	146	47.6	281	48.3	445	49.3	
Gly/Gly	80	26.1	141	24.2	229	25.4	
Arg/Gly or Gly/Gly	226	73.6	422	72.5	674	74.6	$P_{\text{trend}}=0.55$
Gln27Glu							
Gln/Gln	224	73.9	447	75.4	667	72.9	$P_{\text{global}}=0.73$
Gln/Glu	75	24.8	134	22.6	228	24.9	
Glu/Glu	4	1.3	12	2.0	20	2.2	
Gln/Glu or Glu/Glu	79	26.1	146	24.6	248	27.1	$P_{\text{trend}}=0.51$

Table 4-4: Estimated difference (Δ ; and 95% confidence interval) in body composition measure between men homozygous for a specified haplotyperelative to men homozygous for the Arg-Glnhaplotype.

Body composition measure	n	Gly-Gln			Gly-Glu		
		Δ	95% CI	p-value	Δ	95% CI	p-value
BMI (kg/m ²)	1962	-0.28	-0.97, 0.41	0.4261	-0.52	-1.47, 0.43	0.2828
Total body fat % [1]	1960	-0.16	-1.04, 0.73	0.7290	-0.40	-1.62, 0.82	0.5213
Total body fat % [1], in obese men	497	0.17	-1.14, 1.48	0.7983	0.26	-1.57, 2.09	0.7815
Waist circumference (cm), in obese men	499	-0.57	-3.21, 2.08	0.6751	-2.87	-6.58, 0.83	0.1281

1. Total body (except head) fat mass expressed as a percentage of total body (except head) mass

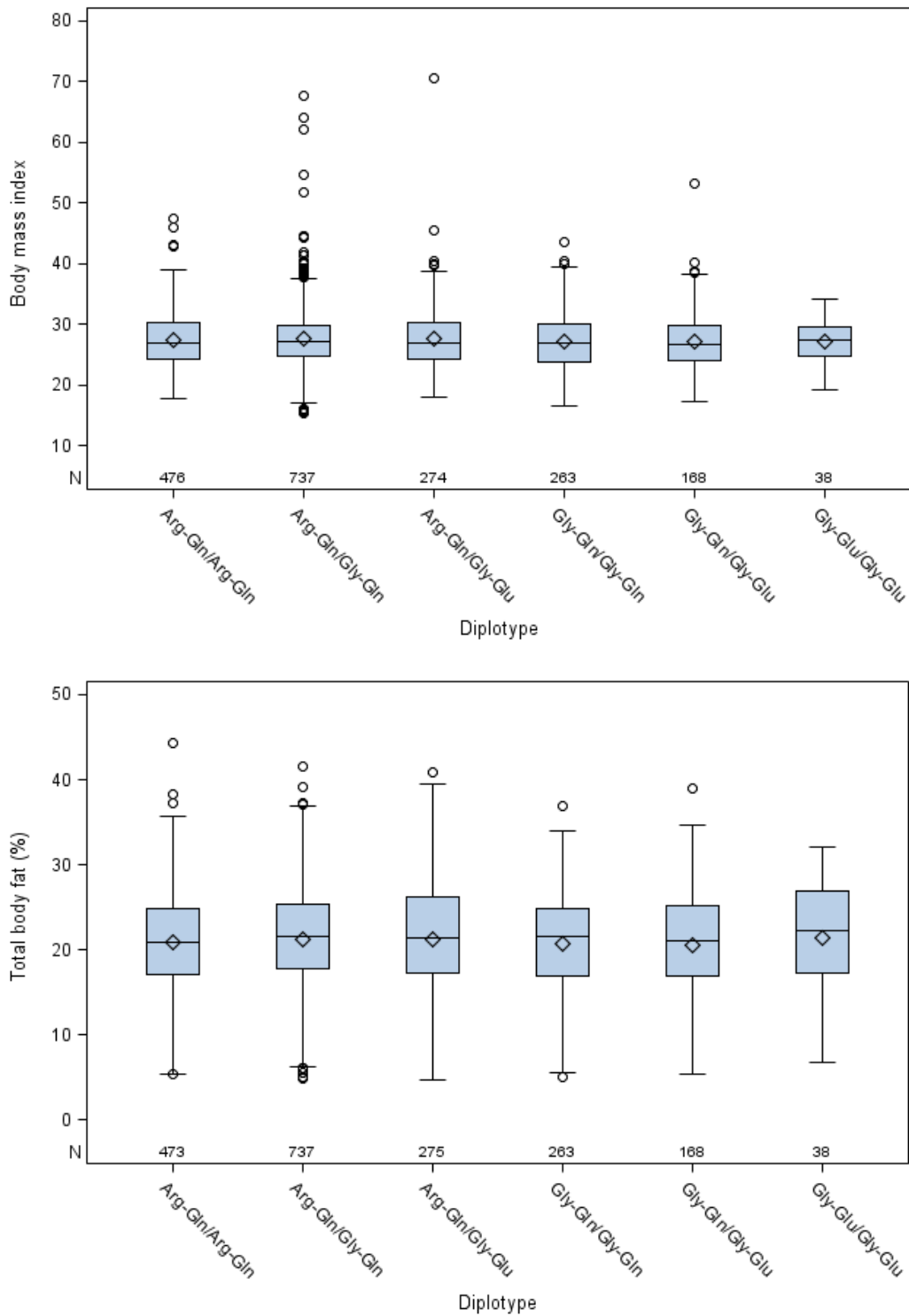


Figure 4-1: Box plots showing body mass index (BMI; upper figure) and DEXA-determined percent body fat (lower figure) in men sub-group according to diplotype.

Figure Footnote: The number (N) in each subgroup appears above the diplotype label. Analyses exclude six men with an ArgGluhaplotype-containing diplotype. The line segment and diamond symbol within each box identify the median and mean, respectively. The lower and upper borders of each box identify the 25th and 75th percentiles, respectively. The lower whisker extends to the minimum observation greater than or equal to the 25th percentile minus 1.5 times the inter-quartile range. The upper whisker extends to the maximum observation less than or equal to the 75th percentile plus 1.5 times the inter-quartile range. Open circle symbols identify individual observations less than the 25th percentile minus 1.5 times the inter-quartile range or greater than the 75th percentile plus 1.5 times the inter-quartile range.

**5.0 PAPER 2: ANDROGEN RECEPTOR ARCAG REPEAT LENGTH
POLYMORPHISM AND OBESITY IN AFRO-CARIBBEAN MEN: RESULTS FROM
THE TOBAGO PROSTATE STUDY.**

Tracey Samantha Beason MSPH,¹ Clareann H. Bunker PhD,¹ Joseph M. Zmuda PhD,¹
John W. Wilson PhD,² Alan L. Patrick MD FRCP³, Victor W Wheeler MBBS, MRCOG,³ and
Joel L. Weissfeld MD MPH¹

¹Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh

²Department of Biostatistics, Graduate School of Public Health, University of Pittsburgh

³Tobago Health Studies Office, Scarborough, Tobago, Trinidad & Tobago

Acknowledgements

This study was supported by grant R01-AR049747 from the National Institute of Arthritis and Musculoskeletal and Skin Diseases, grant R01-CA84950, U.S. National Cancer Institute, and by support from the Division of Health and Social Services, Tobago House of Assembly, and the University of Pittsburgh Cancer Institute.

Objective: Expansion of *AR* CAG repeat has been shown to lower transcriptional activity causing lower AR protein production and androgen signaling. In a population based study we investigated AR CAG repeat length (CAG)_n and its association with body fat parameters in Afro-Caribbean men.

Design and Subjects: Cross-sectional analysis of 2584 Afro-Caribbean men residing on the island of Tobago.

Measurements: Body mass index (BMI), waist circumference, dual energy x-ray absorptiometry (DEXA), AR CAG repeat length genotype.

Results: Mean age was 59 years. *AR* CAG repeats lengths ranged from 9 to 33 with a median of 19. Standardized univariate regression analyses revealed no significant associations between the number of CAG repeats and percent total body fat (β =change in percent total body fat per 3.3 standard deviation change in *AR* CAG repeats), (β =-0.052, P =0.690), BMI (β =-0.184, P =0.085), waist circumference (β =-0.236, P =0.352) and weight (β =-0.647, P =0.068).

Conclusion: Longer *AR* CAG repeat length was not associated with higher body fat measurements.

5.2 INTRODUCTION

Obesity is a major health problem and public health concern³⁶¹. Data suggest that genetics in conjunction with environmental factors may play a pivotal role in the development of obesity. Obesity is associated with a plethora of diseases and increases in risk with increasing age³⁶². Obesity phenotypes in individuals appear to be dimorphic. Men seem to develop an android shape (distribution of fat around the abdomen) whereas women distribute fat mainly around the hips (gluteofemoral regions)^{362, 363}.

Exogenous and endogenous testosterone can affect body composition⁵⁷. Testosterone is important in regulating adiposity, insulin resistance and type II diabetes. Men with type II diabetes have lower testosterone (TT) and sex hormone binding globulin (SHBG) compared to normal men³⁶⁴. Furthermore, lower than normal testosterone levels seem to determine an individual's future risk of developing the metabolic syndrome and type 2 diabetes^{63, 365, 366}. Studies have consistently demonstrated that low levels of testosterone and sex hormone-binding globulin (SHBG) in men are associated with type 2 diabetes, visceral adiposity, and insulin resistance^{365, 366}. Singh et al, delineated that androgens affect the fat cells ability to store lipids by interrupting or blocking signal transduction pathways that are important in adipocyte function³⁶⁷. The actions of androgen in developing obesity are thought to be mediated by the androgen receptor (AR)³⁶⁸.

The androgen receptor is important for the regulation of androgen in men. Dysfunctional receptors may cause variations in body composition³⁶². The AR gene is located on the X-chromosome (Xq11.2-12), highly polymorphic, and comprised of 8 exons³⁶². Most

studies have focused on two polymorphic trinucleotide repeats (microsatellite) CAG² coding for polyglutamine and GGN ((GGT)₃(GGG)(GGT)₂(GGC)_n) coding for glycine^{351, 362}. CAG repeats range between 8-35³⁶⁹ with an average of 20-23 repeats³⁷⁰. In vitro studies have demonstrated that CAG length is inversely associated with AR transactivation (increased rate of gene expression triggered by biological processes)³⁷¹. Longer CAG repeats are associated with lower transcriptional activity, Kennedy's disease, androgen insensitivity and infertility¹⁵¹. Polymorphisms of the AR gene are associated with prostate cancer, metabolic, and cardiovascular diseases³⁷⁰ and these polymorphic repeats (CAG) can cause variations in serum hormone levels leading to hormone-dependent diseases^{62, 372}.

Studies have also shown that the frequency of the *AR* gene CAG repeats varies amongst different racial and ethnic groups¹⁵¹. In a population-based study of Afro-Caribbean men we investigated the association of *AR* CAG repeat lengths on body composition parameters. We hypothesize that men with longer *AR* CAG repeats will have more total body fat mass when expressed as percent total body mass.

5.3 MATERIALS AND METHODS

During 1997 through September 2007, the Tobago Prostate Study recruited 40-79 year old men by public service announcements, flyers, local health care workers, and word of mouth to participate in a study assessing prostate cancer screening. Men were excluded if they were found to be non-ambulatory, terminally ill or cognitively impaired. All participants signed a written

² <http://www.androgendb.mcgill.ca/>

informed consent approved by the Institutional Review Boards of the University of Pittsburgh and the Tobago Division of Health and Social Services.

The Tobago Prostate cohort enrolled 3837 Afro-Caribbean men. Demographic and cancer information were provided by standardized baseline questionnaires. To facilitate genetic studies a more detailed staff-administered Health History questionnaire (HH) was used to record information on newly recruited or follow-up men beginning January 2004. Our study population consisted of 3452 (90.0% of 3837) race-eligible Afro-Caribbean men with ≥ 3 Afro -Caribbean grandparents.

Procedures consisted of body composition tests (DEXA), and genetic testing. DEXA measurements were obtained on 2850 (82.6%) and health history questionnaires on 2259 (65.4%) of 3452 race eligible men, and *AR* CAG genotyping information on 2816 (81.6) of 3452 race eligible men (2126 with health history information and 690 with no health history information). Two hundred and thirty two (232) individuals were excluded because they did not provide any health history (HH) or DEXA information. Participants with DEXA were pulled from both baseline (566 or 22.9%) and follow-up (1902 or 77.1%). Our final sample size for this study was 2584 (Figure 5-1).

5.3.1 Outcome measurements

Key outcome variables from the Health History Questionnaire included: height (measured with shoes to the nearest 0.1 cm on a wall mounted stadiometer), weight (measured to the nearest 0.1 kg on a beam balance), body mass index (BMI, calculated as weight in kg divided by height in meters-squared), waist circumference (measured in cm at the umbilicus with an inelastic tape measure). Percent total body fat was ascertained on a Hologic QDR 4500W DEXA operated in array beam mode and analyzed by QDR software version 8.26a (Hologic Inc. Bedford, MA).

5.3.2 Genotyping

High molecular weight genomic DNA was isolated from peripheral blood samples using the salting out procedure. To determine the AR CAG repeat length, we constructed a set of fluorescent dye labeled oligonucleotide primers that flanked the CAG repeat region in exon 1: Forward Primer = TET dye-5'-ACCGAGGAGCTTTCCAGAAT-3'; Reverse Primer = 5'-AGAACCATCCTCACCCTGCT-3'. The DNA was amplified by polymerase chain reaction (PCR) to produce fragments which varied in length by the number of CAG repeats. PCR consisted of an initial 4-minute denaturation at 95°C, followed by 37 cycles of denaturation (95°C, 60 seconds), annealing (58°C, 30 seconds), and elongation (72°C, 60 seconds), and a final elongation step (72°C, 10 minutes). Amplified samples were analyzed by automated high-throughput scanning fluorescent detectors that have the ability to detect and separate the TET dye. To determine the exact number of CAGs for each fragment length, we subsequently sequenced a series of PCR products of varying length using standard Sanger sequencing. PCR clean up was performed using Exo-SAP according to manufacturer instructions. Sequencing buffer and a 1:4 dilution of BigDye 3.1 were added and thermocycling performed according to manufacturer recommendations. Removal of unincorporated sequencing reagents was performed using CleanSeq magnetic beads according to manufacturer instructions (Beckman Coulter, Inc; Brea, CA). Sanger sequencing was completed using an Applied Biosystems 3730xl DNA Analyzer

5.3.3 Statistical analyses

Statistical analyses were carried out using (SAS 9.2; SAS Institute Inc., Cary NC). All continuous descriptive data are expressed as mean±S.D. To determine the association with androgen receptor CAG repeat length correlations were assessed and expressed as Spearman's Rank Correlation coefficient with the associated P-value. AR CAG data was standardized using the formula (CAG repeats-mean of CAG)/Standard Deviation of CAG before univariate linear

regression. Univariate linear regression techniques were used to assess the independent association between standardized AR CAG and body fat parameters. Results are expressed using the regression coefficient (β) (beta coefficients are expressed in terms of change in the independent variable per one standard deviation unit change in the number of CAG repeats) and the associated P-value. Stratified regression analyses were also performed according to BMI and age categories using linear regression.

5.4 RESULTS

Table 5-1 summarizes characteristics by (mean \pm SD) of the study population according to long vs. short repeats. In Table 5-1 we see that the means were the same in the long and short AR CAG repeat groups except for BMI which showed that the means were statistically different ($p=0.03$). These variables used in this analysis were pulled from the Health History Questionnaire. The number of AR CAG repeats ranged from 9-33 with a median of 19 (Figure 5-2). The average age of participants was 59 years. To determine the frequency of long versus short repeats, AR CAG repeat length was split into low (≤ 22) and high groups (>22) according to Campbell et al³⁷³. Approximately 81.4% or 2103 of (2584) had short repeats and 18.6% or 481 participants had long repeats (Table 5-2). Table 5-3 describes potential confounders (Health History Questionnaire) by long and short repeats. The proportion of men with heart attack (dichotomous variable) was significantly different ($P=0.0302$) according to long versus short repeats.

In our analysis multiple body fat parameters were explored including: percent trunk fat, percent limb fat, percent arm fat, and percent leg fat. Spearman-Rank order correlation analysis

revealed no significant association between *AR* gene CAG repeat polymorphism as a continuous variable and body fat parameters (Table 5-4). Univariate regression analyses revealed no significant associations between *AR* gene CAG repeat length as standardized continuous variable and body fat parameters, percent total body fat ($\beta=-0.052$, $P=0.690$), BMI ($\beta=-0.184$, $P=0.085$), waist circumference ($\beta=-0.236$, $P=0.352$) and weight ($\beta=-0.647$, $P=0.068$) (Table 5-5). The negative β -coefficient indicates that men with shorter CAG repeats had higher body fat measurements. In addition, comparison of included (2584) and excluded (232) participants revealed that the median number of *AR* CAG repeats was the same for both groups (Table 5-6).

Table 5-8 presents stratified regression analysis by age category. This analysis did not detect, in any age group, a statistically significant association between the number of *AR* CAG repeats and percent total body fat. Table 5-9 presents stratified analysis by BMI category. Likewise, this analysis did not detect, in any BMI category, a statistically significant association between the number of *AR* CAG repeats and percent total body fat. Moreover, the negative association observed between different measures of body fat and *AR* CAG repeats shown in Table 5-10 was not significant even after adjusting for age for all body fat parameters except BMI which was statistically significant with a $P=0.04$ (Table 5-10). In addition, Table 5-7 also shows a significant negative association unadjusted for age ($P=0.04$). Therefore regardless of how the analyses were performed, age adjusted or not, the associations observed in this cohort were not altered. We also do recognize that different sample sizes were used in the analyses presented so far and that caution should be used in making inferences. Hence, in an effort to show a more complete analysis various non-missing sample sizes were used in analyses shown in Table 5-11. Even in these analyses, a persistent non-significant negative association was observed between *AR* CAG and percent total body fat. On the other hand BMI continued to

show a statistically significant negative association with *AR* CAG length. In this population having shorter *AR* CAG means that men may be fatter opposite what was originally hypothesized.

5.5 DISCUSSION

Our study revealed that longer *AR* CAG was not associated with higher body fat parameters. Instead our study revealed that men with shorter *AR* CAG had higher measures of body fat.

Studies elucidating an association between *AR*-CAG repeat length and body composition have produced inconsistent results. For instance, Lapauw et al³⁷⁴, did not find an association between *AR* CAG repeat length and body composition parameters. This non-significant finding in Belgium men persisted even after the adjustment of potential confounders (age, weight, total testosterone and sex hormone binding globulin (SHBG)). However, Campbell et al³⁷³ reported an association between *AR* CAG repeats and body composition parameters contrary to findings reported in the aforementioned study. In Campbell's study amongst the Ariaal men of northern Kenya *AR* CAG repeats were positive predictors of height ($\beta=0.420$, $P<0.05$), fat free mass ($\beta=0.383$, $P<0.05$), percent body fat ($\beta=0.329$, $P<0.05$), waist circumference ($\beta=0.385$, $P<0.05$), and suprailiac skinfold ($\beta=0.760$, $P<0.001$). Both Campbell et al³⁷³ and Lapauw et al³⁷⁴ report a regulatory effect of the *AR* CAG repeats on testosterone and consequently the relationship between testosterone (serum testosterone concentration) and body composition (Lapauw et al³⁷⁴ $\beta=0.12$, $P<0.1$ modeled an interaction between CAG repeat and testosterone). Possible explanations for our results of no association include: Campbell et al³⁷³ study consisted of a younger cohort of men 20+ compared to our study in which age ranged from 33-91 years.

Other explanations for the divergent results observed may be explained by gene-environment interaction, and differences in study design. Moreover, it has been postulated that *AR* CAG repeats may function as low penetrance alleles that require the presence of additional genetic and environmental factors to cause an increase in disease susceptibility³⁷⁵. It is quite possible that *AR* CAG may be in linkage disequilibrium with other mutations in the *AR* gene (on the X-chromosome) such as (GGN) that are in close proximity at a particular locus or other unknown genes³⁷⁰ that may affect body composition in men. Furthermore, obesity is a multifactorial disease that may be affected by other hormones, diet and lifestyle that were not accounted for in the present study.

Non-the-less, the results presented in this paper are of interest because based on our knowledge there have been relatively few studies evaluating the *AR* gene in populations of African descent with a large sample size.

5.5.1 Racial differences in CAG repeat length

The distribution of CAG repeats has been reported to differ amongst different racial and ethnic groups. African Americans have been found to have shorter CAG repeats compared to Caucasians (19.8 vs. 21.9)¹⁵¹. Sartor et al³⁷⁶ reported almost twice as many black men had shorter CAG repeats (<20 vs. >20) compared to non-Hispanic white men (56.92% vs. 28.46%) with mean repeats of 19.0 vs. 21.0. The findings of shorter CAG repeats in previous studies are inline with the findings from our study of a mean of 19.7 and a median of 19.

Limitations of our study include: our study was based on a single time point DEXA measurements where longitudinal data might have provided additional information.

Flow chart of sample selection for aim 2 (AR CAG repeats)

Tobago Prostate Study

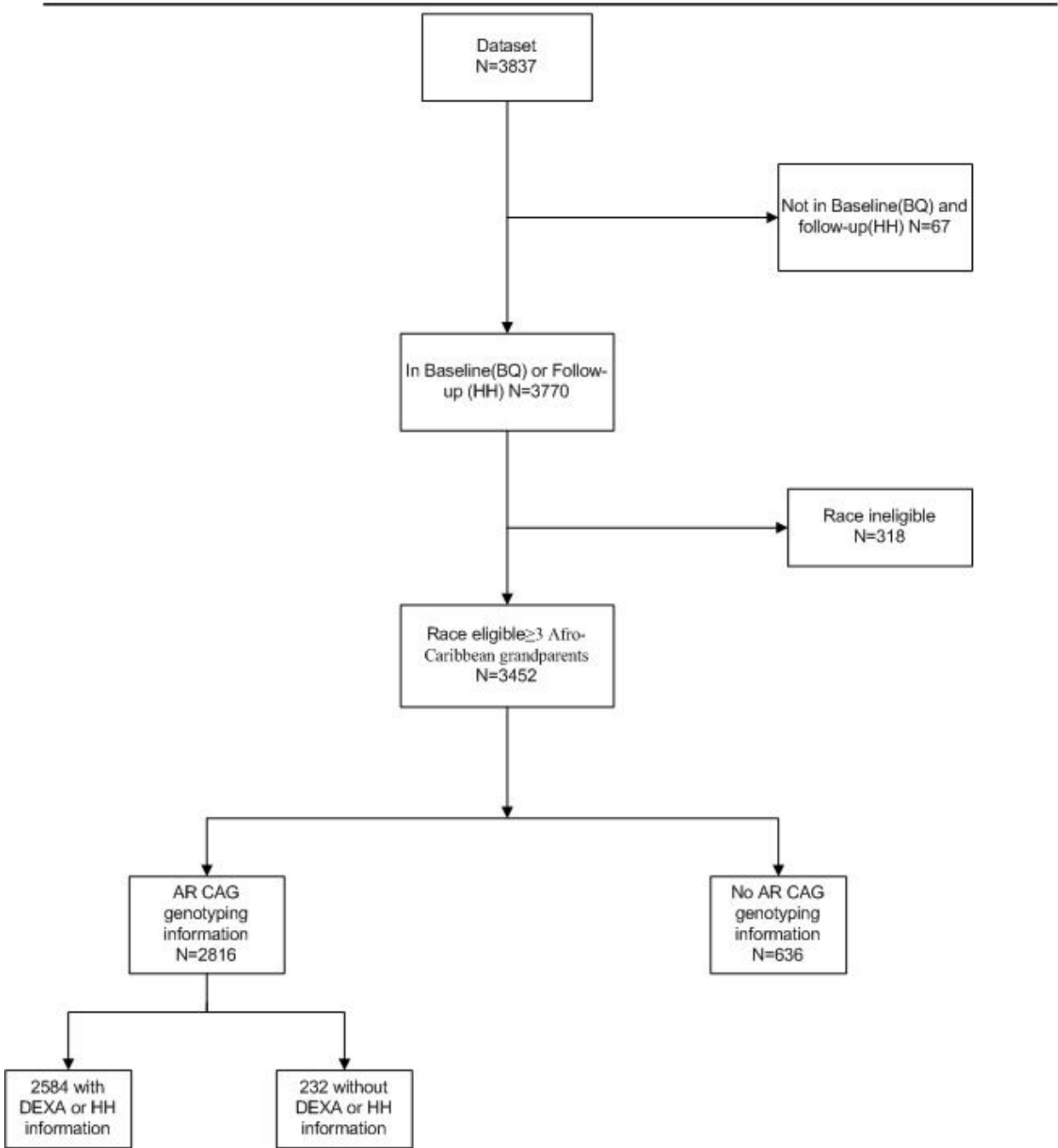


Figure 5-1: Sample selection for paper 2.

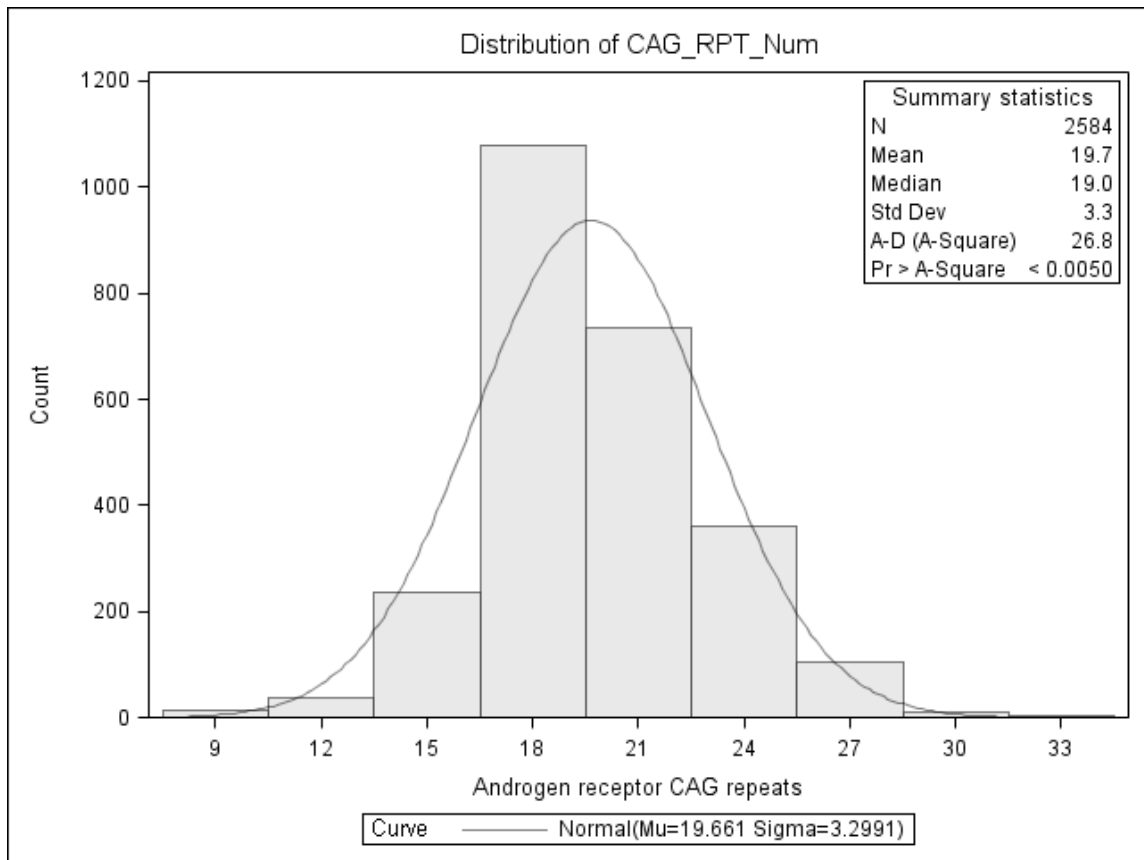


Figure 5-2: Distribution of *AR* CAG repeat length.

Table 5-1: Body composition measures (mean±standard deviations) according to *AR* CAG genotype.

Body composition measure	<i>AR</i> CAG repeat length								P-value [1]
	≤22				>22				
	n	mean	±	SD	n	mean	±	SD	
Age (years)	1733	58.98	±	10.49	393	58.94	±	10.25	0.95
Age at 2nd DEXA (years)	2013	59.20	±	10.94	455	59.28	±	10.63	0.78
Weight (kg)	1726	84.85	±	16.71	392	83.15	±	15.03	0.09
Body mass index (kg/m ²)	1724	27.68	±	5.04	392	27.10	±	4.48	0.03
Waist circumference (cm)	1717	93.40	±	11.77	393	92.66	±	11.40	0.23
Percent total body fat (%)	2013	21.06	±	6.42	455	20.71	±	6.67	0.31

1. Wilcoxon-Mann-Whitney test

Data presented as mean±S.D

Data restricted to individuals with non-missing *AR* CAG repeat length genotype and ≥3 Afro-Caribbean grandparents

Percent total body fat calculated as total fat 100*mass/total body mass.

Table 5-2: Proportion of men with long vs. short *AR* CAG repeats (N=2584).

<i>AR</i> CAG Repeats	Men (N)	Men (%)
≤22	2103	81.4
>22	481	18.6

Table 5-3: Potential confounders.

Attribute		AR CAG repeat length (n=2584)				P-value
		<=22 (n=2103)		>22 (n=481)		
		N	%	N	%	
Demographic						
Education	Primary only	1287	75.2	293	75.7	0.9627
	Post primary	368	21.5	82	21.2	
	University graduate	57	3.3	12	3.1	
Marital status	Married	1197	69.6	261	67.3	0.3450
	Widowed	101	5.9	19	4.9	
	Divorced	96	5.6	18	4.6	
	Separated	88	5.1	27	7.0	
	Never married	239	13.9	63	16.2	
Risk factors						
Smoking status	Current smoker	162	9.4	49	12.5	0.1452
	Ex-smoker	393	22.7	92	23.4	
	Never smoker	1175	67.9	252	64.1	
Alcohol (drinks/wk)	None	679	39.3	158	40.2	0.3072
	<1	735	42.5	156	39.7	
	1-3	144	8.3	26	6.6	
	4-7	71	4.1	22	5.6	
	8-14	56	3.2	16	4.1	
	>14	44	2.5	15	3.8	
Physical activity (walking outside)	Never	52	3.0	9	2.3	0.457
	Seldom (1-2 days/wk)	196	11.4	44	11.3	
	Sometimes (3-4 days/wk)	413	23.9	81	20.8	
	Often (5-7 days/wk)	1065	61.7	255	65.6	
Health history						
Diabetes	Yes	287	16.6	60	15.4	0.5582
	No	1442	83.4	330	84.6	
Heart attack	Yes	14	0.8	8	2.0	0.0302
	No	1716	99.2	385	98.0	

Chi-square test

Table 5-4: Spearman rank correlations between *AR* CAG repeats and body composition measures.

	Weight	Waist circum- ference	Body mass index	Total body fat	Trunk body fat	Arm body fat	Leg body fat	Limb body fat
CAG repeat length	-0.04	-0.02	-0.04	-0.01	-0.02	0.00	0.00	0.00
Weight		0.83*	0.89*	0.59*	0.60*	0.58*	0.51*	0.55*
Waist circumference			0.84*	0.76*	0.76*	0.73*	0.65*	0.70*
Body mass index				0.68*	0.68*	0.68*	0.58*	0.63*
Total body fat					0.97*	0.92*	0.92*	0.96*
Trunk body fat						0.86*	0.81*	0.86*
Arm body fat							0.83*	0.92*
Leg body fat								0.98*

Entries correlation coefficients. *P<0.001.

Data based on restrictions to individuals with CAG repeat length genotyping information, and ≥ 3 afro -Caribbean grandparents.

The number of observations used to estimate individual correlations range between 1998 to 2468

Table 5-5: Results from standardized univariate linear regression models (n=2584).

Attribute	β	SE	t-value	P-value
Age (years)	0.155	0.219	0.71	0.48
Body weight (kg)	-0.647	0.354	-1.830	0.068
Waist circumference (cm)	-0.236	0.253	-0.930	0.352
Body mass index (kg/m ²)	-0.184	0.107	-1.720	0.085
Percent total body fat (%)	-0.052	0.130	-0.400	0.690
Percent trunk fat (%)	-0.109	0.141	-0.780	0.438
Percent arm fat (%)	0.003	0.152	0.020	0.984
Percent leg fat (%)	0.010	0.124	0.080	0.935
Percent limb fat (%)	0.005	0.128	0.040	0.969

Entries are standardized linear regression coefficients

Data presented are participants who are race eligible (≥ 3 Afro-Caribbean grandparents) with AR CAG genotyping

Table 5-6: A comparison of AR CAG length between excluded men (men excluded from analysis because of missing DEXA and missing HH questionnaire) and included men (men included in analysis of association between AR CAG and body composition).

Study	N	Q ₂₅	Median	Q ₇₅	P-value
Excluded	232	18	19	22	0.45
Included	2584	17	19	22	

Q₂₅, 25th Percentile; Q₇₅, 75th percentile

Table 5-7: Unadjusted AR CAG repeat length category (<=22 vs. >22 repeats) associations with age, weight, body mass index (BMI), percent total body fat, percent trunk fat, percent arm fat, percent leg fat, percent limb fat (n=2584).

Attribute	β	SD	t-value	P-value
Age (years)	0.07	0.56	0.13	0.90
Weight (kg)	-1.70	0.92	-1.85	0.06
Average standing height (cm)	0.06	0.38	0.17	0.87
Waist circumference (cm)	-0.73	0.65	-1.12	0.26
Body Mass Index (kg/m ²)	-0.58	0.28	-2.10	0.04
Percent trunk fat (%)	-0.27	0.36	-0.73	0.46
Percent arm fat (%)	-0.41	0.39	-1.04	0.30
Percent leg fat (%)	-0.44	0.32	-1.39	0.16
Percent limb fat (%)	-0.44	0.33	-1.35	0.18
Percent total body fat (%)	-0.35	0.34	-1.04	0.30

Entries linear regression coefficients

Data presented are participants who are race eligible (≥ 3 Afro-Caribbean grandparents) with AR CAG genotyping and DEXA

Table 5-8: Association between AR CAG repeat and percent total body fat, by age.

Age category (years)	N	β (%TBF/CAG repeat)	P-Value
33-50	652	-0.08	0.32
51-57	570	0.09	0.29
58-66	572	-0.04	0.59
67-92	674	-0.05	0.55

Age in years

%TBF; change in percent total body fat

CAG repeat; change in CAG repeat length

Estimated regression coefficients

Table 5-9: Association between AR CAG repeat and percent total body fat, by BMI category.

BMI category	N	β (%TBFAT/CAG Repeat)	P-Value
Underweight	29	-0.04	0.83
Normal weight	609	-0.01	0.83
Overweight or obese	1478	0.04	0.33

%TBF; change in percent total body fat

CAG repeat; change in CAG repeat length

Estimated regression coefficients

Table 5-10: Age-adjusted associations between *AR* CAG repeat length category (≤ 22 vs. > 22 repeats) and eight different measures of body composition (n=2584).

		Dependent variables							
		PcnTBFat		Weight		Waist circumference		Body mass index	
		β	<i>P</i>	β	<i>P</i>	β	<i>P</i>	β	<i>P</i>
CAG repeat ^a	≤ 22 vs. > 22	-0.36	0.28	-1.71	0.056	-0.73	0.26	-0.58	0.034

		Dependent variables							
		PcnTarmFat		PcnTlimbFat		PcnTlegFat		PcntTrunkFat	
		β	<i>P</i>	β	<i>P</i>	β	<i>P</i>	β	<i>P</i>
CAG repeat ^a	≤ 22 vs. > 22	-0.42	0.28	-0.45	0.16	-0.45	0.15	-0.27	0.45

^aAdjusted for age

BMI, body mass index; PcnTBFat, percent total body fat; PcnTarmFat, percent arm fat; PcnTlimbFat, percent limb fat; PcntlegFat, percent leg fat; PcnTtrunkFat, percent trunk fat;

Table 5-11: AR CAG repeat length associations with age, BMI, and percent total body fat, in study groups defined differently with respect to the availability of BMI and percent total body fat information.

Table 5-11a: TBFat% not missing AND BMI not missing at the visit with the non-missing DEXA, n=2330.

Parameters	n	Rank order correlation with AR CAG repeat number		Average difference in the dependent variable, comparing men with long (23-33 repeats) vs. short (9-22 repeats) AR CAG			
		r	p-value	β	S.E.	t-value	p-value
Age at DEXA visit (years)	2330	0.02	0.40	0.09	0.58	0.16	0.87
BMI at DEXA visit (kg/m ²)	2330	-0.05	0.02	-0.54	0.25	-2.14	0.03
TBFat%	2330	-0.01	0.73	-0.37	0.35	-1.07	0.29

Age, BMI, and TBFat% pulled from Visit 1 and 3 in 435 and 1895, respectively
TBFat%; percent total body fat

Table 5-11b: TBFat% not missing, n=2468.

Parameters	n	Rank order correlation with AR CAG repeat number		Average difference in the dependent variable, comparing men with long (23-33 repeats) vs. short (9-22 repeats) AR CAG			
		r	p-value	β	S.E.	t-value	p-value
Age at DEXA visit (years)	2468	0.01	0.52	0.07	0.56	0.13	0.90
BMI at DEXA visit (kg/m ²)	2330	-0.05	0.02	-0.54	0.25	-2.14	0.03
TBFat%	2468	-0.01	0.68	-0.35	0.34	-1.04	0.30

Age, BMI, and TBFat% pulled from Visit 1 and 3 in 566 and 1902, respectively
TBFat%; percent total body fat

Table 5-11c: BMI not missing, n=2466.

Parameters	n	Rank order correlation with AR CAG repeat number		Average difference in the dependent variable, comparing men with long (23-33 repeats) vs. short (9-22 repeats) AR CAG			
		r	p-value	β	S.E.	t-value	p-value
Age at BMI visit (years)	2466	0.02	0.37	-0.04	0.55	-0.08	0.94
BMI (kg/m ²)	2466	-0.05	0.01	-0.62	0.25	-2.44	0.01
TBFat%	2355	-0.01	0.63	-0.43	0.34	-1.25	0.21

BMI pulled from Visit 3, if not missing, from Visit 1, otherwise
Age and BMI pulled from Visit 1 and 3 in 350 and 2116, respectively
TBFat% (available in n=2355) pulled from Visit 1 and 3 in 455 and 1900, respectively
TBFat%; percent total body fat

Table 5-11d: TBFat% not missing OR BMI not missing, n=2585.

Parameters	n	Rank order correlation with <i>AR</i> CAG repeat number		Average difference in the dependent variable, comparing men with long (23-33 repeats) vs. short (9-22 repeats) <i>AR</i> CAG			
		r	p-value	β	S.E.	t-value	p-value
Age at DEXA visit (years)	2468	0.01	0.52	0.07	0.56	0.13	0.90
BMI at Visit 3 (kg/m ²)	2116	-0.04	0.08	-0.58	0.28	-2.10	0.04
TBFat%	2468	-0.01	0.68	-0.35	0.34	-1.04	0.30

N=2468 with non-missing TBFat% and 2116 with non-missing Visit 3 BMI.

Age and TBFat% pulled from Visit 1 and 3 in 566 and 1902, respectively

TBFat%; percent total body fat

**6.0 PAPER 3: EFFECTS OF ANDROGEN DEPRIVATION THERAPY ON
CHANGE IN BODY COMPOSITION IN AFRO-CARIBBEAN MEN: TOBAGO
PROSTATE STUDY**

Tracey Samantha Beason, MSPH¹, Clareann H. Bunker, PhD¹, Joseph M. Zmuda, PhD¹, John, W. Wilson, PhD², Alan L Patrick, MD³, Victor W. Wheeler, MBBS, MRCOG,³Joel L. Weissfeld, MD, MPH¹.

¹Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh

²Department of Biostatistics, Graduate School of Public Health, University of Pittsburgh

³The Tobago Health Studies Office, Scarborough, Tobago, Trinidad & Tobago, West Indies

Funding: Funding for this study was provided by grant R01-AR049747 from the National Institute of Arthritis and Musculoskeletal and Skin Diseases and by funding from the Division of Health and Social Services, Tobago House of Assembly, the University of Pittsburgh Cancer Institute, and grant R01-CA84950, U.S. National Cancer Institute.

Objective: A common treatment for prostate cancer is androgen deprivation therapy (ADT). ADT affects the body by lowering the concentrations of androgen. This can have adverse side effects of which changes in body composition is a major concern. These changes may influence risk for other chronic diseases

Design and Subjects: Case-control Study analysis of 1691 Afro-Caribbean men residing in Tobago and participating in a prostate cancer screening study. Controls included participants without a history of prostate cancer and individuals with prostate cancer unexposed to ADT.

Measurements: Body mass index (BMI), waist circumference, total fat estimated by dual energy x-ray absorptiometry (DEXA).

Results: Percent total body fat increased over time (mean \pm S.E.) (0.57 ± 0.08) compared in men with prostate cancer treated with ADT compared to men with prostate cancer unexposed to ADT (0.29 ± 0.07) with a p-value for change (0.28 ± 0.11 , $P=0.0111$). The change in men with prostate cancer unexposed to ADT did not differ from controls with prostate cancer

Conclusion: Men with prostate cancer exposed to ADT had greater gains in percent total body fat over time compared to men unexposed to ADT.

6.2 INTRODUCTION

Prostate cancer is known as one of the most common malignancies affecting black men in the United States and other countries around the world³⁵¹. There are considerable variations in prostate cancer incidence across racial/ethnic groups with people of African descent having the highest rates, whites having an intermediate rate and Asians the lowest^{53, 55, 351}. Even though this disparity remains to be clearly elucidated, possible explanations for the overt population differences include androgen levels, and dietary factors^{351, 377}.

The production of androgen is regulated by the hypothalamic–pituitary-gonadal axis. Gonadotropin releasing hormone is secreted by the hypothalamus stimulating the pituitary gland to secrete luteinizing hormone, which acts on the Leydig cells of the testis to produce androgen. Testosterone is the main androgen circulating in males. However, Dihydrotestosterone (DHT) is considered the most potent androgen and the main androgen found in tissue³⁵¹. Testosterone moves from the blood into the prostate where it binds to the Androgen receptor (AR)⁵³. Testosterone is also converted to DHT by the enzyme 5 α -reductase types II which is encoded by the SRD5A2 gene. DHT binds to the Androgen receptor in addition to its co-regulators (AIB1/SRC3) and co-repressors (DAX1 and SHP) (the androgen receptor has a higher affinity for DHT) in the cytoplasm and is then translocated into the nucleus where transcription begins³⁷⁸. Androgens are steroid hormones that influence the development of male secondary sex characteristics. In men androgen are developed in the testes and the adrenal glands and are metabolized in the prostate and skin³⁵¹. Testosterone and DHT exact their androgenic effects through the androgen receptor³⁵¹.

There is evidence suggesting that serum levels of testosterone are associated with the development of prostate cancer. A high level of serum testosterone is associated with an

increased risk of low-grade prostate tumors and a decreased risk of low-grade prostate cancer. Whilst low levels of serum testosterone is associated with an increased risk of high grade prostate cancer and reduced risk of low grade prostate cancer³⁷⁸.

A common treatment of prostate cancer is androgen deprivation therapy (ADT). ADT involves the suppression of gonadal androgen in the body. This can be accomplished through a variety of ways some of which are orchiectomy, and gonadotropin-releasing hormone. ADT has numerous side effects such as sexual side effects, hot flashes, gynecomastia, changes in body composition, metabolism, osteoporosis, the cardiovascular system, cognition problems and an overall decline in one's quality of life³⁷⁹. This paper investigates the association between ADT treatment for prostate cancer and rate of change in percent total body fat in Afro-Caribbean men from Tobago.

6.3 MATERIALS AND METHODS

6.3.1 Study sample

From 1997 until September 2007, the Tobago Prostate Study recruited 40 to 79 year old men to a population-based prostate cancer screening study. The men were recruited by public service announcements, flyers, local health care workers, and by word of mouth to participate in a study evaluating prostate cancer screening. Men were excluded from the study if they were determined to be non-ambulatory, terminally ill, or cognitively impaired. All eligible participants signed a written informed consent approved by the institutional review boards of the University of Pittsburgh and the Tobago Division of Health and Social Services.

The Tobago Prostate study enrolled 3837 Afro-Caribbean men (Five hundred and thirty one men were excluded because 485 were new enrollees and did not have a baseline DEXA and 46 were less than 40 years of age). Demographic and cancer information was provided by staff-administered standardized baseline questionnaire and a more detailed follow-up questionnaire (Health History Questionnaire). The Health History questionnaire was used to record information on newly recruited and follow-up men beginning January 2004. Our study consisted of 3452 (90% of 3837) race eligible Afro-Caribbean men with ≥ 3 Afro-Caribbean grandparents.

Procedures consisted of body composition measurements (DEXA). DEXA measurements were obtained on 2850 (82.6%) and health history (HH) questionnaires on 2259 (65.4%) of 3452 race eligible men. Our final sample size was 1691 (Figure 6-1). Men with prostate cancer were categorized as unexposed to ADT, and ADT exposed based on questions from questionnaires. ADT constituted either treatment with gonadotropin releasing hormone (GnRH) agonists, luteinizing releasing hormone (LHRH), anti-androgens, orchiectomy, or any combination of the aforementioned.

6.3.2 Outcome measurements

Important outcome variables included height (measured with shoes to the nearest 0.1 cm on a wall mounted stadiometer), weight (measured to the nearest 0.1 kg on a beam balance), body mass index (BMI- calculated as weight in kg divided by height in meters-squared), waist circumference (measured in centimeters at the umbilicus with an inelastic tape measure). These key anthropometric measures were ascertained from the HH questionnaire. Percent total body fat was ascertained on a Hologic QDR4500W DEXA operated in array beam mode and analyzed by QDR software version 8.26a (Hologic Inc. Bedford, MA). Prostate specific antigen test prior to year 2000 was the Abbot AxSYM PSA assay (Abbott Laboratories, Abbott Park, IL, USA)

and after 2000 the PSA assay was the Advia (Siemen's) Centaur (Siemens Healthcare Diagnostic Advia Centaur, Deerfield, IL).

6.3.3 Statistical Analyses

All statistical analyses were performed using SAS[®] software, version 9.2 (SAS Institute, Inc., Cary, NC, USA). Descriptive statistics were used to characterize the six groups in the study sample. Three hundred individuals and two were determined to have prostate cancer and these individuals were placed in particular groups depending on when they were diagnosed with prostate cancer. 143 individuals were diagnosed with prostate cancer before initial DEXA (80 unexposed to ADT and 60 ADT exposed), 98 were diagnosed with prostate cancer between initial and repeat DEXA (59 unexposed to ADT and 39 ADT exposed), and 61 were diagnosed with prostate cancer after repeat DEXA (59 unexposed to ADT, and 2 ADT exposed). Analysis of variance (one-way-ANOVA) was used to compare the mean difference among the groups. Adjusted comparisons controlled for baseline percent body fat measurements, age and PSA as potential confounders.

6.4 RESULTS

Of the individuals enrolled in the study, 143 individuals were diagnosed with prostate cancer before initial DEXA (80 unexposed to ADT and 60 ADT exposed), 98 were diagnosed with prostate cancer between initial and repeat DEXA (59 unexposed to ADT and 39 ADT exposed), and 61 were diagnosed with prostate cancer after repeat DEXA (59 unexposed to ADT, and 2 ADT exposed). The 2 ADT exposed individuals were excluded from the study because this

group was extremely small to use in making any reliable conclusions. Our control group consisted of, 1391 individuals without a history of prostate cancer and unexposed to ADT (21 individuals indicated that they were ADT exposed on questionnaires but they had no history of prostate cancer; these individuals were excluded (Figure 6-1). One person was also excluded from the final analysis because they did not have all the relevant repeat DEXA measurements). The majority of the study samples were Afro-Caribbean and participants were more likely to be married and have at least a primary school level education. Table 6-1 shows the mean and standard deviation for a number of clinical characteristics. Men had significantly different age ($P=0.0001$), percent total body fat ($P=0.0076$) and Prostate specific antigen ($P=0.0001$) across groups. Mean body mass index was not significantly different across groups ($P=0.6691$).

Table 6-2 shows the mean change in percent total body fat in men according to prostate cancer status and ADT exposure. In men exposed and unexposed to ADT who had their prostate cancer diagnosed before the 1st DEXA had significant mean difference in percent total body fat unadjusted ($P=0.0056$), adjusted for age ($P=0.0036$) and adjusted for age and 1st DEXA percent total body fat ($P=0.0021$). Similar findings were also observed in men exposed and unexposed to ADT with prostate cancer diagnosed between the 1st and 2nd DEXA scans; unadjusted ($P=0.0041$), adjusted for age ($P=0.0032$) and adjusted for age and 1st DEXA percent total body fat ($P=0.0021$). Men with prostate cancer diagnosed after repeat (2nd) DEXA and unexposed to ADT had similar mean difference in percent total body fat (prostate cancer diagnosed after 2nd DEXA (0.82 ± 0.31), compared to men without prostate cancer (0.80 ± 0.07). Finally Table 6-3 shows the rate of change of percent body fat in the six groups. Overall table 4 indicates that the rate of change of percent total body fat was greater in persons exposed to ADT than in persons unexposed the ADT. In the prostate cancer diagnosed after 2nd DEXA and Healthy controls the

mean rate of change of percent total body fat was similar 0.17 ± 0.09 and 0.18 ± 0.02 . The difference in the two mean rate of change of percent total body fat 0 ± 0.09 , $p=0.9626$.

Table 6-4 shows the median years between initial and repeat DEXA, which was approximately 4.5 years. Table 6-5 shows the frequency of the type of androgen deprivation therapy used. Antiandrogen, GnRH, and LHRH was the treatment of choice in this cohort. Antiandrogen, GnRH, and LHRH was used in 74.6% (47) in prostate cancer diagnosed after 1st DEXA and 76.9% (30) in the group prostate cancer diagnosed between the 1st and 2nd DEXA. Nineteen percent (12) persons had orchiectomy only, in the group prostate cancer diagnosed after 1st DEXA and 12.8% (5) persons had orchiectomy in the group prostate cancer diagnosed between 1st and 2nd DEXA. Four (6.3% and 10.3% respectively) had combined treatments in both groups.

6.5 DISCUSSION

Androgen suppression or androgen ablation or androgen deprivation is chemical treatment to suppress or block the production or action of the male sex hormone to treat prostate cancer in men. Two common methods by which this can be done is by removing the testicles or by taking anti-androgens³⁸⁰. There are several different types of anti-androgen drugs available. In our study evaluating the effect of ADT on percent total body fat we determined that percent total body fat increased in men with prostate cancer exposed to ADT. In comparison men without prostate cancer and men with prostate cancer who did not initiate ADT therapy had much lower increases in percent body fat.

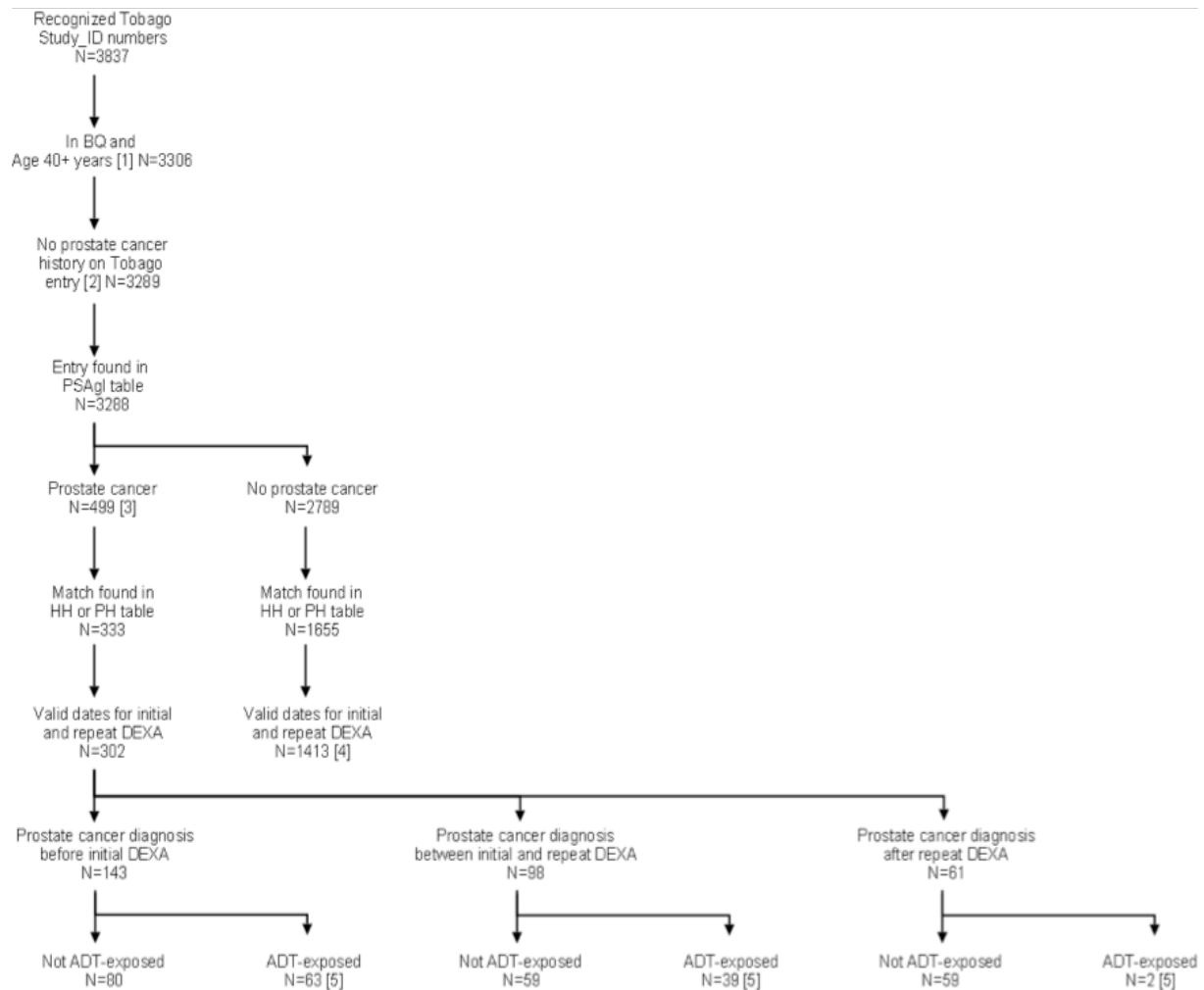
Androgens are considered important determinants of body composition in men. ADT has been shown to increase serum triglycerides and cholesterol³⁸⁰. A reduction in muscle mass in men leads to reduced strength and increased risk of falls and fractures (sarcopenia)¹⁴. The replacement of androgen in the body reverses the effects of androgen deprivation²⁸. In men with prostate cancer the use of androgen antagonist affects the body by reducing physical function³⁸⁰.

Our results support an increase in percent total body fat measured by DEXA following ADT administration in Afro-Caribbean men with prostate cancer. However, there were several limitations with our study. First we were unable to determine the duration of ADT treatment. It is possible that men who had a longer duration of ADT gained more fat. Hence, our study even though we found an increase in percent body fat may have underestimated the response. We also encountered possible information bias. There were 21 individuals who did not have prostate cancer but reported that they were taking ADT. ADT exposure variable was determined from self-report and was not independently verified. These individuals were of course excluded from the study. There was also the issue of channeling bias; beyond PSA clinical indications for ADT treatment may have confounded observed associations between ADT and changes in percent total body fat. Moreover, some individuals with prostate cancer were prescribed ADT while others were not. One can only speculate about possible reasons why some individuals received ADT treatment and others did not such as the cases were more advanced or had metastasized, or maybe it was not affordable. Another limitation was selection bias; the requirement that participants have a second DEXA may have excluded some men according to ADT exposure and outcome. Finally we had small sample sizes for the various groups.

Nonetheless, our study had important strengths. It prospectively observed body composition changes following ADT administration. Two different control groups were used for

comparison (healthy controls and prostate cancer individuals unexposed to ADT). Finally we used dual-energy absorptiometry machine to determine percent total body fat measurements.

In summary, the results of our study should be interpreted with caution because of the small sample size. Non-the-less, in men with prostate cancer percent total body fat increased over time, on average, more so in men exposed than in men unexposed to ADT.



1. Age according to BRTHDATE and INDATE on Wave 1 Tobago BQ.
2. No 31 cancer code entered in CANCER_1, CANCER_2, or CANCER_3 fields of Wave 1 Tobago BQ.
3. GLEASDX>0 in PSAgl table.
4. Includes N=21 (1.5%) with ADT exposure. See Footnote [5].
5. RMTE=Yes ("Have you had your testicles removed (castration, orchiectomy) for prostate cancer?") on HH table or TESTR=Yes on PH table ("Since your last Tobago Prostate Survey visit, have you had your testicles removed?") or MDPRO=Yes on HH table ("Have you ever taken Lucrin (Lucopride), Zolodex, Androcur, Stilbesterol, or Casodex?") or MDFLU=Yes on HH table ("Have you ever taken Flutamide?") or MDCAR=Yes on HH table ("Have you ever taken Proscar or Finasteride?") or MEDIP=Yes on PH table ("Since your last Tobago Prostate Survey visit, have you taken any medication(s) for your prostate?")

Figure 6-1: Sample selection for Paper 3.

Table 6-1: Baseline body composition characteristics (mean ±S.D.).

Parameter	Prostate cancer before 1st DEXA		Prostate cancer diagnosis between 1st and 2nd DEXA		Prostate cancer after 2nd DEXA	No Prostate cancer	P-Value (across groups)
	ADT (n=63)	No ADT (n=80)	ADT (n=39)	No ADT (n=59)	No ADT (n=59)	No ADT (n=1391)	
Age (years)	67.9±6.9	63.58±8.7	64.5±6.97	61.76±9.0	58.3±7.85	53.71±9.9	0.0001
BMI (kg/m ²)	26.89±4.3	28.2±4.1	27.3±4.15	27.2±3.03	27.42±4.02	27.57±4.2	0.6691
TBF (%)	22.75±5.7	21.7±6.8	21.26±5.76	20.52±5.1	20.01±6.3	20.17±6.2	0.0076
PSA (ng/ml)	23.64±34.6	9.69±18.35	10.14±10.52	15.54±45.34	8.92±24.61	2.21±10.69	0.0001

Abbreviations: TBF (%); Percent total body fat; ADT; Androgen deprivation therapy

PSA; Prostate specific antigen

Age used in this table was the age at 1st DEXA

Table 6-2: Change in percent total body fat (Δ TBFat%) by prostate cancer status and androgen deprivation exposure.

Categories	Androgen deprivation treatment						Δ meanTBFat%		p- value
	Yes			No					
	N	Mean	S.E.	N	Mean	S.E.	Delta	S.E.	
<i>Prostate cancer diagnosed before 1st DEXA scan</i>									
Unadjusted	63	2.56	0.43	80	1.25	0.39	1.30	0.47	0.0056
Age-adjusted	63	2.50	0.44	80	1.30	0.39	1.37	0.47	0.0036
Age and 1st DEXA scan TBFat%-adjusted	63	2.85	0.36	80	1.43	0.31	1.42	0.46	0.0021
Age, psa, and 1st DEXA scan TBFat%-adjusted	63	2.75	0.36	80	1.40	0.31	1.35	0.46	0.0038
<i>Prostate cancer diagnosed between 1st and 2nd DEXA scans</i>									
Unadjusted	39	3.16	0.45	59	1.50	0.37	1.66	0.58	0.0041
Age-adjusted	39	3.08	0.45	59	1.55	0.37	1.70	0.58	0.0032
Age and 1st DEXA scan TBFat%-adjusted	39	3.29	0.44	59	1.55	0.36	1.74	0.56	0.0021
Age, psa, and 1st DEXA scan TBFat%-adjusted	39	3.28	0.44	59	1.49	0.36	1.78	0.57	0.0017
<i>Prostate cancer diagnosed after 2nd DEXA scan</i>									
Unadjusted				59	0.82	0.31			
<i>No prostate cancer</i>									
Unadjusted				1391	0.80	0.07			

Legend:

Δ TBFat% = 2nd DEXA scan TBFat% – 1st DEXA scan TBFat%

Delta = mean TBFat% in the group that received androgen deprivation treatment – mean TBFat% in the group that did not receive androgen deprivation treatment

PSA; prostate specific antigen

Values for comparison groups, no prostate cancer and prostate cancer diagnosed after 2ndDEXA (Δ meanTBF%, 0.0153), (S.E., 0.37076) and (P-value=0.9672)

Table 6-3: Mean rate of change of percent total body fat (Δ TBFat%) by prostate cancer and androgen deprivation.

Categories	Androgen deprivation treatment						Delta		p-value
	Yes			No					
	N	Mean	SE	N	Mean	SE	Mean	S.E.	
<i>Prostate cancer diagnosed before 1st DEXA scan</i>									
Unadjusted	63	0.57	0.08	80	0.29	0.07	0.28	0.11	0.0111
Age-adjusted	63	0.61	0.09	80	0.31	0.07	0.30	0.11	0.0077
Age and 1st DEXA scan TBF%-adjusted	63	0.63	0.08	80	0.32	0.07	0.31	0.11	0.0047
Age, psa and 1st DEXA scan TBF%-adjusted	63	0.61	0.09	80	0.32	0.07	0.29	0.11	0.0084
<i>Prostate cancer diagnosed between 1st and 2nd DEXA scans</i>									
Unadjusted	39	0.68	0.11	59	0.33	0.09	0.35	0.14	0.0094
Age-adjusted	39	0.71	0.11	59	0.35	0.09	0.36	0.14	0.0077
Age and 1st DEXA scan TBF%-adjusted	39	0.71	0.10	59	0.34	0.08	0.37	0.13	0.0054
Age, psa and 1st DEXA scan TBF%-adjusted	39	0.70	0.11	59	0.32	0.09	0.38	0.13	0.0047
<i>Prostate cancer diagnosed after 2nd DEXA scan</i>									
Unadjusted				59	0.17	0.09			
<i>No prostate cancer</i>									
Unadjusted				1391	0.18	0.02			

Legend:

rate of Δ TBFat% = (2nd DEXA scan TBFat% – 1st DEXA scan TBFat%)/ years between 1st and 2nd DEXA

Delta = mean rate Δ TBFat% in the group that received androgen deprivation treatment -mean rate Δ TBFat% in the group that did not receive androgen deprivation treatment

PSA; prostate specific antigen

Values for comparison groups, no prostate cancer and prostate cancer diagnosed after 2ndDEXA (rate of Δ meanTBF%, 0.00), (S.E., 0.09) and (P-value=0.9626)

Table 6-4: Tobago Prostate Study, subject mean age, median interval between sequential DEXA scans, median interval between prostate cancer diagnosis and first DEXA, and median interval between prostate cancer diagnosis and second DEXA, according to prostate cancer status, timing of prostate cancer diagnosis with respect to sequential DEXA, and prostate cancer treatment status (ADT vs. no ADT).

Category	N	Age Tobago entry (years)		Age at initial DEXA (years)		Years between initial and repeat DEXA			Years from diagnosis to initial DEXA			Years from diagnosis to repeat DEXA		
		Mean	S.D.	Mean	S.D.	Median	Q ₂₅	Q ₇₅	Median	Q ₂₅	Q ₇₅	Median	Q ₂₅	Q ₇₅
No prostate cancer	1391	52.55	9.69	53.71	9.92	4.45	3.95	4.77						
Prostate cancer before initial DEXA, no ADT	80	62.28	8.62	63.59	8.77	4.50	4.05	4.76	0.71	0.38	1.11	5.25	4.72	5.53
Prostate cancer before initial DEXA, ADT	63	66.64	6.85	67.89	6.91	4.36	4.08	4.88	0.88	0.42	1.50	5.45	5.07	6.26
Prostate cancer between initial and repeat DEXA, no ADT	59	60.88	9.15	61.76	9.01	4.65	4.33	5.04	-1.04	-1.78	-0.27	3.73	2.95	4.50
Prostate cancer between initial and repeat DEXA, ADT	39	57.15	7.69	64.49	6.97	4.56	4.31	4.91	-0.84	-1.89	-0.21	3.37	2.51	4.82
Prostate cancer after repeat DEXA	59	52.55	9.69	58.31	7.86	4.61	4.24	4.94	-5.62	-6.75	-5.03	-0.76	-2.19	-0.27

Legend:

Q₂₅; 25 percentile; Q₇₅; 75 percentile

Table 6-5: Type of androgen deprivation, in men reporting an ADT exposure.

Type of androgen deprivation	Prostate cancer before 1st DEXA and ADT exposed (N=63)		Prostate cancer diagnosis between 1st and 2nd DEXA and ADT exposed (N=39)	
	N	%	N	%
Medical treatment only (anti-androgen or LHRH agonist)	47	74.6	30	76.9
Surgical treatment (orchiectomy) only	12	19.0	5	12.8
Medical and surgical treatment	4	6.3	4	10.3

Legend:

ADT, androgen deprivation therapy

LHRH, luteinizing hormone releasing hormone

Bibliography

1. Smith, M. R.; Finkelstein, J. S.; McGovern, F. J.; Zietman, A. L.; Fallon, M. A.; Schoenfeld, D. A.; Kantoff, P. W., Changes in body composition during androgen deprivation therapy for prostate cancer. *J Clin Endocrinol Metab* **2002**,*87* (2), 599-603.
2. Ben-Shlomo, Y.; Evans, S.; Ibrahim, F.; Patel, B.; Anson, K.; Chinegwundoh, F.; Corbishley, C.; Dorling, D.; Thomas, B.; Gillatt, D.; Kirby, R.; Muir, G.; Nargund, V.; Popert, R.; Metcalfe, C.; Persad, R., The risk of prostate cancer amongst black men in the United Kingdom: the PROCESS cohort study. *Eur Urol* **2008**,*53* (1), 99-105.
3. JM Ordovas; EJ Schaefer, Genes, variation of cholesterol and fat intake and serum lipids. *Current Opinion in Lipidology* **1999**,*10* (1), 15-22.
4. Jose M Ordovas; Schaefer, E., treatment of dyslipidemia: genetic interactions with diet and drug therapy. *Current Artherosclerosis Reports* **1999**,*1* (1), 16-23.
5. K. E. Ensrud; Lewis, C. E.; Lambert, L. C.; Taylor, B. C.; Fink, H. A.; Cauley, J. A.; M.L.Stefanick; E.Orwoll, Endogeneous sex steroids, weight change and rates of hip bone loss in older men: the MrOS Study. *Osteoporos Int* **2006**,*17*, 1329-13336.
6. Heaney, R. P.; Abrams, S.; Dawson-Hughes, B.; Looker, A.; Marcus, R.; Matkovic, V.; Weaver, C., Peak bone mass. *Osteoporos Int* **2000**,*11* (12), 985-1009.
7. Van Langendonck, L.; Claessens, A. L.; Lefevre, J.; Thomis, M.; Philippaerts, R.; Delvaux, K.; Lysens, R.; Vanden Eynde, B.; Beunen, G., Association between bone mineral density (DXA), body structure, and body composition in middle-aged men. *Am J Hum Biol* **2002**,*14* (6), 735-42.
8. Ashcroft, M., Prevalence of hypertension and associated electrocardiographic abnormalities in Jamaica and West Africa. *West Indian Med J* **1977**,*26*, 24.
9. Patrick, A.; Vaughan, J.; Boyd-Patrick, H., Cardiovascular risk factors in Tobagonians comparison with other African populations. *West Indian Med J* **1986**,*35*, 149.
10. Nichols, S.; Cadogan, F., Blood pressure and its correlates in Tobagonian adolescents. *West Indian Med J* **2006**,*55* (5), 305-12.
11. Eva Warensjo; Ingelsson, E.; Landmark, P.; Lanfelt, L.; Syvanen, A.-C.; Vessby, B., Polymorphisms in the SCD1 gene: associations with body fat distribution and insulin sensitivity. *Genetics* **2007**,*15* (7), 1732-1740.
12. Miljkovic-Gacic, I.; Ferrell, R. E.; Patrick, A. L.; Kammerer, C. M.; Bunker, C. H., Estimates of African, European and Native American ancestry in Afro-Caribbean men on the island of Tobago. *Hum Hered* **2005**,*60* (3), 129-33.
13. Reis, L.; Melbert, D.; Krapcho, M.; Stinchcomb, D.; Howlander, N.; Horner, M.; Mariotto, A.; Miller, B.; Feuer, E.; Alterkruse, S.; Lewis, D.; Clegg, L. X.; Eisner, M.; Reichman, M.; Edwards, B. *SEER Cancer Statistics Review 1975-2005* National Cancer Institute: Bethesda, MD, 2007.
14. Chen, Z.; Maricic, M.; Nguyen, P.; Ahmann, F. R.; Bruhn, R.; Dalkin, B. L., Low bone density and high percentage of body fat among men who were treated with androgen deprivation therapy for prostate carcinoma. *Cancer* **2002**,*95* (10), 2136-44.
15. Looker, A. C.; Orwoll, E. S.; Johnston, C. C., Jr.; Lindsay, R. L.; Wahner, H. W.; Dunn, W. L.; Calvo, M. S.; Harris, T. B.; Heyse, S. P., Prevalence of low femoral bone density in older U.S. adults from NHANES III. *J Bone Miner Res* **1997**,*12* (11), 1761-8.

16. Wei, J. T.; Gross, M.; Jaffe, C. A.; Gravlin, K.; Lahaie, M.; Faerber, G. J.; Cooney, K. A., Androgen deprivation therapy for prostate cancer results in significant loss of bone density. *Urology* **1999**,*54* (4), 607-11.
17. Diamond, T.; Campbell, J.; Bryant, C.; Lynch, W., The effect of combined androgen blockade on bone turnover and bone mineral densities in men treated for prostate carcinoma: longitudinal evaluation and response to intermittent cyclic etidronate therapy. *Cancer* **1998**,*83* (8), 1561-6.
18. Eriksson, S.; Eriksson, A.; Stege, R.; Carlstrom, K., Bone mineral density in patients with prostatic cancer treated with orchidectomy and with estrogens. *Calcif Tissue Int* **1995**,*57* (2), 97-9.
19. Smith, M. R.; McGovern, F. J.; Fallon, M. A.; Schoenfeld, D.; Kantoff, P. W.; Finkelstein, J. S., Low bone mineral density in hormone-naive men with prostate carcinoma. *Cancer* **2001**,*91* (12), 2238-45.
20. Goldray, D.; Weisman, Y.; Jaccard, N.; Merdler, C.; Chen, J.; Matzkin, H., Decreased bone density in elderly men treated with the gonadotropin-releasing hormone agonist decapeptyl (D-Trp6-GnRH). *J Clin Endocrinol Metab* **1993**,*76* (2), 288-90.
21. Daniell, H. W.; Dunn, S. R.; Ferguson, D. W.; Lomas, G.; Niazi, Z.; Stratte, P. T., Progressive osteoporosis during androgen deprivation therapy for prostate cancer. *J Urol* **2000**,*163* (1), 181-6.
22. Kroke, A.; Bergmann, M.; Klipstein-Grobusch, K.; Boeing, H., Obesity, body fat distribution and body build: their relation to blood pressure and prevalence of hypertension. *Int J Obes Relat Metab Disord* **1998**,*22* (11), 1062-70.
23. Pi-Sunyer, F., Comorbidities of overweight and obesity: current evidence and research issues. *Med Sci Sports Exerc* **1999**,*31*, S602-8.
24. van den Beld, A. W.; de Jong, F. H.; Grobbee, D. E.; Pols, H. A.; Lamberts, S. W., Measures of bioavailable serum testosterone and estradiol and their relationships with muscle strength, bone density, and body composition in elderly men. *J Clin Endocrinol Metab* **2000**,*85* (9), 3276-82.
25. Demark-Wahnefried, W.; Conaway, M. R.; Robertson, C. N.; Mathias, B. J.; Anderson, E. E.; Paulson, D. F., Anthropometric risk factors for prostate cancer. *Nutr Cancer* **1997**,*28* (3), 302-7.
26. Katznelson, L.; Rosenthal, D. I.; Rosol, M. S.; Anderson, E. J.; Hayden, D. L.; Schoenfeld, D. A.; Klibanski, A., Using quantitative CT to assess adipose distribution in adult men with acquired hypogonadism. *AJR Am J Roentgenol* **1998**,*170* (2), 423-7.
27. Ongphiphadhanakul, B.; Rajatanavin, R.; Chailurkit, L.; Piaseu, N.; Teerarungsikul, K.; Sirisriro, R.; Komindr, S.; Puavilai, G., Serum testosterone and its relation to bone mineral density and body composition in normal males. *Clin Endocrinol (Oxf)* **1995**,*43* (6), 727-33.
28. Katznelson, L.; Finkelstein, J. S.; Schoenfeld, D. A.; Rosenthal, D. I.; Anderson, E. J.; Klibanski, A., Increase in bone density and lean body mass during testosterone administration in men with acquired hypogonadism. *J Clin Endocrinol Metab* **1996**,*81* (12), 4358-65.
29. Zelefsky, M. J.; Eastham, J. A.; Sartor, O. A.; Kantoff, P., Cancer of the prostate. In *Devita, Hellman & Rosenberg's cancer: principles & practice of oncology, 8th edition*, Devita, V. T.; Lawrence, T. S.; Rosenberg, S. A., Eds. Lippincott Williams & Wilkins: 2008; Vol. 1.

30. Ross, R. K.; Pike, M. C.; Coetzee, G. A.; Reichardt, J. K.; Yu, M. C.; Feigelson, H.; Stanczyk, F. Z.; Kolonel, L. N.; Henderson, B. E., Androgen metabolism and prostate cancer: establishing a model of genetic susceptibility. *Cancer Res* **1998**,*58* (20), 4497-504.
31. Adami, H.; Hunter, D.; Trichopoulos, D., *Textbook of cancer epidemiology second edition*. Oxford University Press: Oxford, New York, 2008.
32. Kantoff, P. W., Prostate cancer. In *ACP Medicine*, WebMD Corporation: New York, NY, 2008.
33. Bunker, C. H.; Patrick, A. L.; Konety, B. R.; Dhir, R.; Brufsky, A. M.; Vivas, C. A.; Becich, M. J.; Trump, D. L.; Kuller, L. H., High prevalence of screening-detected prostate cancer among Afro-Caribbeans: The Tobago Prostate Cancer Survey. *Cancer Epidemiology, Biomarkers & Prevention* **2002**,*11*, 726-729.
34. Odedina, F. T.; Akinremi, T. O.; Chinegwundoh, F.; Roberts, R.; Yu, D.; Reams, R. R.; Freedman, M. L.; Rivers, B.; Green, B. L.; Kumar, N., Prostate cancer disparities in Black men of African descent: a comparative literature review of prostate cancer burden among Black men in the United States, Caribbean, United Kingdom, and West Africa. *Infect Agent Cancer* **2009**,*4 Suppl 1*, S2.
35. Bunker, C. H.; Patrick, A. L.; Maharaj, G.; Keenan, H. A.; Ramnarine, S.; Belle, A.; Richard, J. R.; Dhir, R., Prostate cancer risk is three-fold higher among men, aged 50-64, of African descent compared with men of Asian-Indian descent in Trinidad and Tobago. *Ethn Dis* **2002**,*12* (4), S3-30-3.
36. Ogunbiyi, J. O.; Shittu, O. B., Increased incidence of prostate cancer in Nigerians. *J Natl Med Assoc* **1999**,*91* (3), 159-64.
37. Haas, G. P.; Sakr, W. A., Epidemiology of prostate cancer. *CA Cancer J Clin* **1997**,*47* (5), 273-87.
38. Glover, F. E., Jr.; Coffey, D. S.; Douglas, L. L.; Cadogan, M.; Russell, H.; Tulloch, T.; Baker, T. D.; Wan, R. L.; Walsh, P. C., The epidemiology of prostate cancer in Jamaica. *J Urol* **1998**,*159* (6), 1984-6; discussion 1986-7.
39. Ferlay, J. B. F.; Pisani, P.; Parkin, D. M. *GLOBOCAN 2002: cancer incidence, mortality and prevalence worldwide*. In: IARC Cancer Base No.5. version 2.0 ed. : 2004.
40. Brooks, S. E.; Hanchard, B.; Wolff, C.; Samuels, E.; Allen, J., Age-specific incidence of cancer in Kingston and St. Andrew, Jamaica, 1988-1992. *West Indian Med J* **1995**,*44* (3), 102-5.
41. Hanchard, B.; Blake, G.; Wolff, C.; Samuels, E.; Waugh, N.; Simpson, D.; Ramjit, C.; Mitchell, K., Age-specific incidence of cancer in Kingston and St Andrew, Jamaica, 1993-1997. *West Indian Med J* **2001**,*50* (2), 123-9.
42. Bunker, C. H.; Patrick, A. L.; Miljkovic-Gacic, I.; Konety, B. R.; Belle, A.; Richard, J. R.; Dhir, R., Prostate cancer screening parameters in a high-risk African-Caribbean population. *Urology* **2004**,*63* (4), 737-41.
43. Phillips, A. A.; Jacobson, J. S.; Magai, C.; Consedine, N.; Horowicz-Mehler, N. C.; Neugut, A. I., Cancer incidence and mortality in the Caribbean. *Cancer Invest* **2007**,*25* (6), 476-83.
44. Chinegwundoh, F.; Enver, M.; Lee, A.; Nargund, V.; Oliver, T.; Ben-Shlomo, Y., Risk and presenting features of prostate cancer amongst African-Caribbean, South Asian and European men in North-east London. *BJU Int* **2006**,*98* (6), 1216-20.
45. Steinberg, G. D.; Carter, B. S.; Beaty, T. H.; Childs, B.; Walsh, P. C., Family history and the risk of prostate cancer. *Prostate* **1990**,*17* (4), 337-47.

46. Carter, B. S.; Beaty, T. H.; Steinberg, G. D.; Childs, B.; Walsh, P. C., Mendelian inheritance of familial prostate cancer. *Proc Natl Acad Sci U S A* **1992**,*89* (8), 3367-71.
47. Catalona, W. J.; Smith, D. S.; Ratliff, T. L.; Dodds, K. M.; Coplen, D. E.; Yuan, J. J.; Petros, J. A.; Andriole, G. L., Measurement of prostate-specific antigen in serum as a screening test for prostate cancer. *N Engl J Med* **1991**,*324* (17), 1156-61.
48. Ross, K. S.; Carter, H. B.; Pearson, J. D.; Guess, H. A., Comparative efficiency of prostate-specific antigen screening strategies for prostate cancer detection. *JAMA* **2000**,*284* (11), 1399-405.
49. In *Cancer: Principles & Practice of Oncology*, DeVita, V. T.; Hellman, S.; Rosenberg, S. A., Eds.
50. In *Kelley's Textbook of Internal Medicine*, Humes, H. D.; DuPont, H. L.; Gardner, L. B.; Griffin, J. W.; Harris, E. D.; Hazzard, W. R.; King, T. E.; Loriaux, D. L.; Nabel, E. G.; Todd, R. F.; Traber, P. G., Eds.
51. In *Primary Care*, Singleton, J. K.; Sandowski, S. A.; Green-Hernandez, C.; Horvath, T. V.; DiGregorio, R. V.; Holzemer, S. P., Eds.
52. In *5-Minute Clinical Consult, The (2007 edition)*. Domino, F. J., Ed.
53. Pettaway, C. A., Racial differences in the androgen/androgen receptor pathway in prostate cancer. *J Natl Med Assoc* **1999**,*91* (12), 653-60.
54. Ross, R. K.; Bernstein, L.; Lobo, R. A.; Shimizu, H.; Stanczyk, F. Z.; Pike, M. C.; Henderson, B. E., 5-alpha-reductase activity and risk of prostate cancer among Japanese and US white and black males. *Lancet* **1992**,*339* (8798), 887-9.
55. Ellis, L.; Nyborg, H., Racial/ethnic variations in male testosterone levels: a probable contributor to group differences in health. *Steroids* **1992**,*57* (2), 72-5.
56. Harman, S. M.; Metter, E. J.; Tobin, J. D.; Pearson, J.; Blackman, M. R., Longitudinal effects of aging on serum total and free testosterone levels in healthy men. Baltimore Longitudinal Study of Aging. *J Clin Endocrinol Metab* **2001**,*86* (2), 724-31.
57. Zmuda, J. M.; Cauley, J. A.; Kriska, A.; Glynn, N. W.; Gutai, J. P.; Kuller, L. H., Longitudinal relation between endogenous testosterone and cardiovascular disease risk factors in middle-aged men. A 13-year follow-up of former Multiple Risk Factor Intervention Trial participants. *Am J Epidemiol* **1997**,*146* (8), 609-17.
58. Gapstur, S. M.; Gann, P. H.; Kopp, P.; Colangelo, L.; Longcope, C.; Liu, K., Serum androgen concentrations in young men: a longitudinal analysis of associations with age, obesity, and race. The CARDIA male hormone study. *Cancer Epidemiol Biomarkers Prev* **2002**,*11* (10 Pt 1), 1041-7.
59. Field, A. E.; Colditz, G. A.; Willett, W. C.; Longcope, C.; McKinlay, J. B., The relation of smoking, age, relative weight, and dietary intake to serum adrenal steroids, sex hormones, and sex hormone-binding globulin in middle-aged men. *J Clin Endocrinol Metab* **1994**,*79* (5), 1310-6.
60. Henderson, B. E.; Bernstein, L.; Ross, R. K.; Depue, R. H.; Judd, H. L., The early in utero oestrogen and testosterone environment of blacks and whites: potential effects on male offspring. *Br J Cancer* **1988**,*57* (2), 216-8.
61. Troisi, R.; Potischman, N.; Roberts, J.; Siiteri, P.; Daftary, A.; Sims, C.; Hoover, R. N., Associations of maternal and umbilical cord hormone concentrations with maternal, gestational and neonatal factors (United States). *Cancer Causes Control* **2003**,*14* (4), 347-55.
62. Zitzmann, M.; Nieschlag, E., The CAG repeat polymorphism within the androgen receptor gene and maleness. *Int J Androl* **2003**,*26* (2), 76-83.

63. Oh, J. Y.; Barrett-Connor, E.; Wedick, N. M.; Wingard, D. L., Endogenous sex hormones and the development of type 2 diabetes in older men and women: the Rancho Bernardo study. *Diabetes Care* **2002**,*25* (1), 55-60.
64. Zitzmann, M.; Brune, M.; Nieschlag, E., Vascular reactivity in hypogonadal men is reduced by androgen substitution. *J Clin Endocrinol Metab* **2002**,*87* (11), 5030-7.
65. Zitzmann, M.; Brune, M.; Vieth, V.; Nieschlag, E., Monitoring bone density in hypogonadal men by quantitative phalangeal ultrasound. *Bone* **2002**,*31* (3), 422-9.
66. Chu, L. W.; Reichardt, J. K.; Hsing, A. W., Androgens and the molecular epidemiology of prostate cancer. *Curr Opin Endocrinol Diabetes Obes* **2008**,*15* (3), 261-70.
67. Dehm, S. M.; Tindall, D. J., Molecular regulation of androgen action in prostate cancer. *J Cell Biochem* **2006**,*99* (2), 333-44.
68. Thigpen, A. E.; Silver, R. I.; Guileyardo, J. M.; Casey, M. L.; McConnell, J. D.; Russell, D. W., Tissue distribution and ontogeny of steroid 5 alpha-reductase isozyme expression. *J Clin Invest* **1993**,*92* (2), 903-10.
69. Chokkalingam, A. P.; Stanczyk, F. Z.; Reichardt, J. K.; Hsing, A. W., Molecular epidemiology of prostate cancer: hormone-related genetic loci. *Front Biosci* **2007**,*12*, 3436-60.
70. Hsing, A. W.; Sakoda, L. C.; Chua, S., Jr., Obesity, metabolic syndrome, and prostate cancer. *Am J Clin Nutr* **2007**,*86* (3), s843-57.
71. Platz, E. A.; Leitzmann, M. F.; Michaud, D. S.; Willett, W. C.; Giovannucci, E., Interrelation of energy intake, body size, and physical activity with prostate cancer in a large prospective cohort study. *Cancer Res* **2003**,*63* (23), 8542-8.
72. Gapstur, S. M.; Kopp, P.; Gann, P. H.; Chiu, B. C.; Colangelo, L. A.; Liu, K., Changes in BMI modulate age-associated changes in sex hormone binding globulin and total testosterone, but not bioavailable testosterone in young adult men: the CARDIA Male Hormone Study. *Int J Obes (Lond)* **2007**,*31* (4), 685-91.
73. Kupelian, V.; Page, S. T.; Araujo, A. B.; Travison, T. G.; Bremner, W. J.; McKinlay, J. B., Low sex hormone-binding globulin, total testosterone, and symptomatic androgen deficiency are associated with development of the metabolic syndrome in nonobese men. *J Clin Endocrinol Metab* **2006**,*91* (3), 843-50.
74. Platz, E. A.; Leitzmann, M. F.; Rifai, N.; Kantoff, P. W.; Chen, Y. C.; Stampfer, M. J.; Willett, W. C.; Giovannucci, E., Sex steroid hormones and the androgen receptor gene CAG repeat and subsequent risk of prostate cancer in the prostate-specific antigen era. *Cancer Epidemiol Biomarkers Prev* **2005**,*14* (5), 1262-9.
75. Brown-Sequard, C., Note on the effects produced on man by sebcutaneous injections of a liquid obtained from the testicles of animals. *Lancet* **1889**,*2*, 105-107.
76. Freeman, E.; Bloom, D.; McGuire, E., A brief history of testosterone. *J Urol* **1965**, 371-373.
77. The New York Times, prostate cancer: hormone therapy and chemotherapy. *The New York Times* **2009**.
78. Laino, C., Androgen deprivation may raise risk of cardiovascular death, adding toremifene may improve lipid and bone profiles. *Prostate cancer symposium* **2007**, 42-44.
79. Andrew D. Loblaw; Mendelson, D. S.; Talcott, J. A.; Virgo, K. S.; Somerfield, M. S.; Ben-Josef, E.; Middleton, R., American Society of Clinical Oncology Recommendations for initial hormonal management of androgen-sensitive metastatic, recurrent, or progressive prostate cancer. *Journal of Clinical Oncology* **2004**,*22* (14), 2927-2941.

80. Morley, J.; Perry, H.; Kaiser, F.; Kraenzle, D.; Jenson, J.; Houston, K., Effects of testosterone replacement therapy in old hypogonadal males : a preliminary study. *Journal of American Geriatric Society* **1993**,*41*, 149-152.
81. Morley, J.; Kasier, F.; Raum, W.; Perry, H.; Flood, J.; Jenson, J.; Silver, A.; Roberts, E., Potentially predictive and manipulable blood serum correlates of aging in the healthy male : progressive decreases in bioavailable testosterone dehydroepiandrosterone sulfate, and the ratio of insulin like growth factor 1 to growth hormone. *PNAS* **1997**,*94*, 7537-7542.
82. Woodhouse, L. J.; Gupta, N.; Bhasin, M.; Singh, A. B.; Ross, R.; Phillips, J.; Bhasin, S., Dose-dependent effects of testosterone on regional adipose tissue distribution in healthy young men. *J Clin Endocrinol Metab* **2004**,*89* (2), 718-26.
83. katznelson, L.; Finkelstein, J.; Schoenfeld, D.; Rosenthal, D.; Anderson, E., Increase in bone density and lean body mass during testosterone administration in men with acquired hypogonadism. *J Clin Endocrinol Metab* **1996**,*81*, 4358-4365.
84. Katznelson, L.; Rosenthal, D.; Rosol, M., Using quantitative CT to assess adipose distribution in adult men with acquired hypogonadism. *AJR Am J Roentgenol* **1997**,*170*, 423-427.
85. Greendale, G. A.; Edelstein, S.; Barrett-Connor, E., Endogenous sex steroids and bone mineral density in older women and men: the Rancho Bernardo Study. *J Bone Miner Res* **1997**,*12* (11), 1833-43.
86. Slemenda, C. W.; Longcope, C.; Zhou, L.; Hui, S. L.; Peacock, M.; Johnston, C. C., Sex steroids and bone mass in older men. Positive associations with serum estrogens and negative associations with androgens. *J Clin Invest* **1997**,*100* (7), 1755-9.
87. Herbert Lepor, A review of surgical techniques for radical prostatectomy. *Reviews in Urology* **2005**,*7 suppl. 2*, S11-S17.
88. Albertsen, P. C.; Hanley, J. A.; Gleason, D. F.; Barry, M. J., Competing risk analysis of men aged 55 to 74 years at diagnosis managed conservatively for clinically localized prostate cancer. *JAMA* **1998**,*280* (11), 975-80.
89. Vermeulen, A.; Goemaere, S.; Kaufman, J. M., Testosterone, body composition and aging. *J Endocrinol Invest* **1999**,*22* (5 Suppl), 110-6.
90. Snyder, P. J.; Peachey, H.; Hannoush, P.; Berlin, J. A.; Loh, L.; Lenrow, D. A.; Holmes, J. H.; Dlewati, A.; Santanna, J.; Rosen, C. J.; Strom, B. L., Effect of testosterone treatment on body composition and muscle strength in men over 65 years of age. *J Clin Endocrinol Metab* **1999**,*84* (8), 2647-53.
91. Bhasin, S.; Storer, T. W.; Javanbakht, M.; Berman, N.; Yarasheski, K. E.; Phillips, J.; Dike, M.; Sinha-Hikim, I.; Shen, R.; Hays, R. D.; Beall, G., Testosterone replacement and resistance exercise in HIV-infected men with weight loss and low testosterone levels. *JAMA* **2000**,*283* (6), 763-70.
92. Grinspoon, S.; Corcoran, C.; Stanley, T.; Baaj, A.; Basgoz, N.; Klibanski, A., Effects of hypogonadism and testosterone administration on depression indices in HIV-infected men. *J Clin Endocrinol Metab* **2000**,*85* (1), 60-5.
93. Xu, X.; De Pergola, G.; Bjorntorp, P., The effects of androgens on the regulation of lipolysis in adipose precursor cells. *Endocrinology* **1990**,*126* (2), 1229-34.
94. Nowicki, M.; Bryc, W.; Kokot, F., Hormonal regulation of appetite and body mass in patients with advanced prostate cancer treated with combined androgen blockade. *J Endocrinol Invest* **2001**,*24* (1), 31-6.

95. Smith, J. C.; Bennett, S.; Evans, L. M.; Kynaston, H. G.; Parmar, M.; Mason, M. D.; Cockcroft, J. R.; Scanlon, M. F.; Davies, J. S., The effects of induced hypogonadism on arterial stiffness, body composition, and metabolic parameters in males with prostate cancer. *J Clin Endocrinol Metab* **2001**,*86* (9), 4261-7.
96. Herr, H. W.; O'Sullivan, M., Quality of life of asymptomatic men with nonmetastatic prostate cancer on androgen deprivation therapy. *J Urol* **2000**,*163* (6), 1743-6.
97. Potosky, A. L.; Knopf, K.; Clegg, L. X.; Albertsen, P. C.; Stanford, J. L.; Hamilton, A. S.; Gilliland, F. D.; Eley, J. W.; Stephenson, R. A.; Hoffman, R. M., Quality-of-life outcomes after primary androgen deprivation therapy: results from the Prostate Cancer Outcomes Study. *J Clin Oncol* **2001**,*19* (17), 3750-7.
98. Greenspan, S. L.; Coates, P.; Sereika, S. M.; Nelson, J. B.; Trump, D. L.; Resnick, N. M., Bone loss after initiation of androgen deprivation therapy in patients with prostate cancer. *J Clin Endocrinol Metab* **2005**,*90* (12), 6410-7.
99. Luukinen, H.; Koski, K.; Laippala, P.; Kivela, S. L., Factors predicting fractures during falling impacts among home-dwelling older adults. *J Am Geriatr Soc* **1997**,*45* (11), 1302-9.
100. Khosla, S.; Melton, L. J., 3rd; Riggs, B. L., Clinical review 144: Estrogen and the male skeleton. *J Clin Endocrinol Metab* **2002**,*87* (4), 1443-50.
101. Taxel, P.; Fall, P. M.; Albertsen, P. C.; Dowsett, R. D.; Trahiotis, M.; Zimmerman, J.; Ohannessian, C.; Raisz, L. G., The effect of micronized estradiol on bone turnover and calciotropic hormones in older men receiving hormonal suppression therapy for prostate cancer. *J Clin Endocrinol Metab* **2002**,*87* (11), 4907-13.
102. Smith, M. R., Diagnosis and management of treatment-related osteoporosis in men with prostate carcinoma. *Cancer* **2003**,*97* (3 Suppl), 789-95.
103. Santini, D.; Gentilucci, U. V.; Vincenzi, B., The antineoplastic role of bisphosphonates: from basic research to clinical evidence. *Ann Oncol* **2003**,*14*, 1468-1476.
104. Holroyd, C.; Cooper, C.; Dennison, E., Epidemiology of osteoporosis. *Best Pract Res Clin Endocrinol Metab* **2008**,*22* (5), 671-85.
105. Marshall, D.; Johnell, O.; Wedel, H., Meta-analysis of how well measures of bone mineral density predict occurrence of osteoporotic fractures. *Bmj* **1996**,*312* (7041), 1254-9.
106. Johnell, O.; Kanis, J. A., An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. *Osteoporos Int* **2006**,*17* (12), 1726-33.
107. Bilezikian, J. P., Osteoporosis in men. *J Clin Endocrinol Metab* **1999**,*84* (10), 3431-4.
108. Gennari, L.; Becherini, L.; Falchetti, A.; Masi, L.; Massart, F.; Brandi, M. L., Genetics of osteoporosis: role of steroid hormone receptor gene polymorphisms. *J Steroid Biochem Mol Biol* **2002**,*81* (1), 1-24.
109. Gennari, L.; Becherini, L.; Masi, L.; Mansani, R.; Gonnelli, S.; Cepollaro, C.; Martini, S.; Montagnani, A.; Lentini, G.; Becorpi, A. M.; Brandi, M. L., Vitamin D and estrogen receptor allelic variants in Italian postmenopausal women: evidence of multiple gene contribution to bone mineral density. *J Clin Endocrinol Metab* **1998**,*83* (3), 939-44.
110. Duncan, E. L.; Brown, M. A.; Sinsheimer, J.; Bell, J.; Carr, A. J.; Wordsworth, B. P.; Wass, J. A., Suggestive linkage of the parathyroid receptor type 1 to osteoporosis. *J Bone Miner Res* **1999**,*14* (12), 1993-9.

111. Zmuda, J. M.; Eichner, J. E.; Ferrell, R. E.; Bauer, D. C.; Kuller, L. H.; Cauley, J. A., Genetic variation in alpha 2HS-glycoprotein is related to calcaneal broadband ultrasound attenuation in older women. *Calcif Tissue Int* **1998**,*63* (1), 5-8.
112. Ota, N.; Hunt, S. C.; Nakajima, T.; Suzuki, T.; Hosoi, T.; Orimo, H.; Shirai, Y.; Emi, M., Linkage of interleukin 6 locus to human osteopenia by sibling pair analysis. *Hum Genet* **1999**,*105* (3), 253-7.
113. Ota, N.; Nakajima, T.; Nakazawa, I.; Suzuki, T.; Hosoi, T.; Orimo, H.; Inoue, S.; Shirai, Y.; Emi, M., A nucleotide variant in the promoter region of the interleukin-6 gene associated with decreased bone mineral density. *J Hum Genet* **2001**,*46* (5), 267-72.
114. Zmuda, J. M.; Cauley, J. A.; Danielson, M. E.; Theobald, T. M.; Ferrell, R. E., Vitamin D receptor translation initiation codon polymorphism and markers of osteoporotic risk in older African-American women. *Osteoporos Int* **1999**,*9* (3), 214-9.
115. Takacs, I.; Koller, D. L.; Peacock, M.; Christian, J. C.; Evans, W. E.; Hui, S. L.; Conneally, P. M.; Johnston, C. C., Jr.; Foroud, T.; Econs, M. J., Sib pair linkage and association studies between bone mineral density and the interleukin-6 gene locus. *Bone* **2000**,*27* (1), 169-73.
116. Peacock, M.; Turner, C. H.; Econs, M. J.; Foroud, T., Genetics of osteoporosis. *Endocr Rev* **2002**,*23* (3), 303-26.
117. Slemenda, C. W.; Christian, J. C.; Williams, C. J.; Norton, J. A.; Johnston, C. C., Jr., Genetic determinants of bone mass in adult women: a reevaluation of the twin model and the potential importance of gene interaction on heritability estimates. *J Bone Miner Res* **1991**,*6* (6), 561-7.
118. Ferrari, S. L.; Chevalley, T.; Bonjour, J. P.; Rizzoli, R., Childhood fractures are associated with decreased bone mass gain during puberty: an early marker of persistent bone fragility? *J Bone Miner Res* **2006**,*21* (4), 501-7.
119. Kannus, P.; Palvanen, M.; Kaprio, J.; Parkkari, J.; Koskenvuo, M., Genetic factors and osteoporotic fractures in elderly people: prospective 25 year follow up of a nationwide cohort of elderly Finnish twins. *Bmj* **1999**,*319* (7221), 1334-7.
120. Michaelsson, K.; Melhus, H.; Ferm, H.; Ahlbom, A.; Pedersen, N. L., Genetic liability to fractures in the elderly. *Arch Intern Med* **2005**,*165* (16), 1825-30.
121. Deng, H. W.; Mahaney, M. C.; Williams, J. T.; Li, J.; Conway, T.; Davies, K. M.; Li, J. L.; Deng, H.; Recker, R. R., Relevance of the genes for bone mass variation to susceptibility to osteoporotic fractures and its implications to gene search for complex human diseases. *Genet Epidemiol* **2002**,*22* (1), 12-25.
122. Kanis, J. A.; Oden, A.; Johnell, O.; Johansson, H.; De Laet, C.; Brown, J.; Burckhardt, P.; Cooper, C.; Christiansen, C.; Cummings, S.; Eisman, J. A.; Fujiwara, S.; Gluer, C.; Goltzman, D.; Hans, D.; Krieg, M. A.; La Croix, A.; McCloskey, E.; Mellstrom, D.; Melton, L. J., 3rd; Pols, H.; Reeve, J.; Sanders, K.; Schott, A. M.; Silman, A.; Torgerson, D.; van Staa, T.; Watts, N. B.; Yoshimura, N., The use of clinical risk factors enhances the performance of BMD in the prediction of hip and osteoporotic fractures in men and women. *Osteoporos Int* **2007**,*18* (8), 1033-46.
123. Andrew, T.; Antoniadou, L.; Scurrah, K. J.; Macgregor, A. J.; Spector, T. D., Risk of wrist fracture in women is heritable and is influenced by genes that are largely independent of those influencing BMD. *J Bone Miner Res* **2005**,*20* (1), 67-74.
124. van Meurs, J. B.; Schuit, S. C.; Weel, A. E.; van der Klift, M.; Bergink, A. P.; Arp, P. P.; Colin, E. M.; Fang, Y.; Hofman, A.; van Duijn, C. M.; van Leeuwen, J. P.; Pols, H.

- A.; Uitterlinden, A. G., Association of 5' estrogen receptor alpha gene polymorphisms with bone mineral density, vertebral bone area and fracture risk. *Hum Mol Genet* **2003**,*12* (14), 1745-54.
125. Naganathan, V.; Macgregor, A.; Snieder, H.; Nguyen, T.; Spector, T.; Sambrook, P., Gender differences in the genetic factors responsible for variation in bone density and ultrasound. *J Bone Miner Res* **2002**,*17* (4), 725-33.
126. Karasik, D.; Cupples, L. A.; Hannan, M. T.; Kiel, D. P., Age, gender, and body mass effects on quantitative trait loci for bone mineral density: the Framingham Study. *Bone* **2003**,*33* (3), 308-16.
127. Ralston, S. H.; Galwey, N.; MacKay, I.; Albagha, O. M.; Cardon, L.; Compston, J. E.; Cooper, C.; Duncan, E.; Keen, R.; Langdahl, B.; McLellan, A.; O'Riordan, J.; Pols, H. A.; Reid, D. M.; Uitterlinden, A. G.; Wass, J.; Bennett, S. T., Loci for regulation of bone mineral density in men and women identified by genome wide linkage scan: the FAMOS study. *Hum Mol Genet* **2005**,*14* (7), 943-51.
128. Brown, L. B.; Streeten, E. A.; Shuldiner, A. R.; Almasy, L. A.; Peyser, P. A.; Mitchell, B. D., Assessment of sex-specific genetic and environmental effects on bone mineral density. *Genet Epidemiol* **2004**,*27* (2), 153-61.
129. Kammerer, C. M.; Schneider, J. L.; Cole, S. A.; Hixson, J. E.; Samollow, P. B.; O'Connell, J. R.; Perez, R.; Dyer, T. D.; Almasy, L.; Blangero, J.; Bauer, R. L.; Mitchell, B. D., Quantitative trait loci on chromosomes 2p, 4p, and 13q influence bone mineral density of the forearm and hip in Mexican Americans. *J Bone Miner Res* **2003**,*18* (12), 2245-52.
130. Amin, S.; Zhang, Y.; Sawin, C. T.; Evans, S. R.; Hannan, M. T.; Kiel, D. P.; Wilson, P. W.; Felson, D. T., Association of hypogonadism and estradiol levels with bone mineral density in elderly men from the Framingham study. *Ann Intern Med* **2000**,*133* (12), 951-63.
131. Khosla, S.; Melton, L. J., 3rd; Robb, R. A.; Camp, J. J.; Atkinson, E. J.; Oberg, A. L.; Rouleau, P. A.; Riggs, B. L., Relationship of volumetric BMD and structural parameters at different skeletal sites to sex steroid levels in men. *J Bone Miner Res* **2005**,*20* (5), 730-40.
132. Hochberg, M. C., Racial differences in bone strength. *Trans Am Clin Climatol Assoc* **2007**,*118*, 305-15.
133. Beck, T. J.; Ruff, C. B.; Warden, K. E.; Scott, W. W., Jr.; Rao, G. U., Predicting femoral neck strength from bone mineral data. A structural approach. *Invest Radiol* **1990**,*25* (1), 6-18.
134. Tracy, J. K.; Meyer, W. A.; Flores, R. H.; Wilson, P. D.; Hochberg, M. C., Racial differences in rate of decline in bone mass in older men: the Baltimore men's osteoporosis study. *J Bone Miner Res* **2005**,*20* (7), 1228-34.
135. Han, Z. H.; Palnitkar, S.; Rao, D. S.; Nelson, D.; Parfitt, A. M., Effects of ethnicity and age or menopause on the remodeling and turnover of iliac bone: implications for mechanisms of bone loss. *J Bone Miner Res* **1997**,*12* (4), 498-508.
136. Parfitt, A. M.; Han, Z. H.; Palnitkar, S.; Rao, D. S.; Shih, M. S.; Nelson, D., Effects of ethnicity and age or menopause on osteoblast function, bone mineralization, and osteoid accumulation in iliac bone. *J Bone Miner Res* **1997**,*12* (11), 1864-73.
137. Bell, N. H.; Shary, J.; Stevens, J.; Garza, M.; Gordon, L.; Edwards, J., Demonstration that bone mass is greater in black than in white children. *J Bone Miner Res* **1991**,*6* (7), 719-23.

138. Bell, N. H.; Gordon, L.; Stevens, J.; Shary, J. R., Demonstration that bone mineral density of the lumbar spine, trochanter, and femoral neck is higher in black than in white young men. *Calcif Tissue Int* **1995**,*56* (1), 11-3.
139. Tilley, W. D.; Marcelli, M.; Wilson, J. D.; McPhaul, M. J., Characterization and expression of a cDNA encoding the human androgen receptor. *Proc Natl Acad Sci U S A* **1989**,*86* (1), 327-31.
140. Quigley, C. A.; De Bellis, A.; Marschke, K. B.; el-Awady, M. K.; Wilson, E. M.; French, F. S., Androgen receptor defects: historical, clinical, and molecular perspectives. *Endocr Rev* **1995**,*16* (3), 271-321.
141. Van Pottelbergh, I.; Lumbroso, S.; Goemaere, S.; Sultan, C.; Kaufman, J. M., Lack of influence of the androgen receptor gene CAG-repeat polymorphism on sex steroid status and bone metabolism in elderly men. *Clin Endocrinol (Oxf)* **2001**,*55* (5), 659-66.
142. Choong, C. S.; Wilson, E. M., Trinucleotide repeats in the human androgen receptor: a molecular basis for disease. *J Mol Endocrinol* **1998**,*21* (3), 235-57.
143. Brown, C. J.; Goss, S. J.; Lubahn, D. B.; Joseph, D. R.; Wilson, E. M.; French, F. S.; Willard, H. F., Androgen receptor locus on the human X chromosome: regional localization to Xq11-12 and description of a DNA polymorphism. *Am J Hum Genet* **1989**,*44* (2), 264-9.
144. La Spada, A. R.; Wilson, E. M.; Lubahn, D. B.; Harding, A. E.; Fischbeck, K. H., Androgen receptor gene mutations in X-linked spinal and bulbar muscular atrophy. *Nature* **1991**,*352* (6330), 77-9.
145. Mangelsdorf, D. J.; Thummel, C.; Beato, M.; Herrlich, P.; Schutz, G.; Umesono, K.; Blumberg, B.; Kastner, P.; Mark, M.; Chambon, P.; Evans, R. M., The nuclear receptor superfamily: the second decade. *Cell* **1995**,*83* (6), 835-9.
146. Hsing, A. W.; Gao, Y. T.; Wu, G.; Wang, X.; Deng, J.; Chen, Y. L.; Sesterhenn, I. A.; Mostofi, F. K.; Benichou, J.; Chang, C., Polymorphic CAG and GGN repeat lengths in the androgen receptor gene and prostate cancer risk: a population-based case-control study in China. *Cancer Res* **2000**,*60* (18), 5111-6.
147. Danforth, K. N.; Hayes, R. B.; Rodriguez, C.; Yu, K.; Sakoda, L. C.; Huang, W. Y.; Chen, B. E.; Chen, J.; Andriole, G. L.; Calle, E. E.; Jacobs, E. J.; Chu, L. W.; Figueroa, J. D.; Yeager, M.; Platz, E. A.; Michaud, D. S.; Chanock, S. J.; Thun, M. J.; Hsing, A. W., Polymorphic variants in PTGS2 and prostate cancer risk: results from two large nested case-control studies. *Carcinogenesis* **2008**,*29* (3), 568-72.
148. Mhatre, A. N.; Trifiro, M. A.; Kaufman, M.; Kazemi-Esfarjani, P.; Figlewicz, D.; Rouleau, G.; Pinsky, L., Reduced transcriptional regulatory competence of the androgen receptor in X-linked spinal and bulbar muscular atrophy. *Nature genetics* **1993**,*5* (2), 184-8.
149. Liu, C. S.; Chang, Y. C.; Chen, D. F.; Huang, C. C.; Pang, C. Y.; Lee, H. C.; Cheng, C. C.; Horng, C. J.; Wei, Y. H., Type IV hyperlipoproteinemia and moderate instability of CAG triplet expansion in the androgen-receptor gene. Lipid, sex hormone and molecular study in a Chinese family with Kennedy-Alter-Sung disease. *Acta Neurol Scand* **1995**,*92* (5), 398-404.
150. Tut, T. G.; Ghadessy, F. J.; Trifiro, M. A.; Pinsky, L.; Yong, E. L., Long polyglutamine tracts in the androgen receptor are associated with reduced trans-activation, impaired sperm production, and male infertility. *J Clin Endocrinol Metab* **1997**,*82* (11), 3777-82.
151. Bennett, C. L.; Price, D. K.; Kim, S.; Liu, D.; Jovanovic, B. D.; Nathan, D.; Johnson, M. E.; Montgomery, J. S.; Cude, K.; Brockbank, J. C.; Sartor, O.; Figg, W. D., Racial

variation in CAG repeat lengths within the androgen receptor gene among prostate cancer patients of lower socioeconomic status. *J Clin Oncol* **2002**,*20* (17), 3599-604.

152. Price, H.; McNeal, J. E.; Stamey, T. A., Evolving patterns of tissue composition in benign prostatic hyperplasia as a function of specimen size. *Hum Pathol* **1990**,*21* (6), 578-85.

153. Kittles, R. A.; Young, D.; Weinrich, S.; Hudson, J.; Argyropoulos, G.; Ukoli, F.; Adams-Campbell, L.; Dunston, G. M., Extent of linkage disequilibrium between the androgen receptor gene CAG and GGC repeats in human populations: implications for prostate cancer risk. *Hum Genet* **2001**,*109* (3), 253-61.

154. Wang, G.; Chen, G.; Wang, X.; Zhong, J.; Lu, J., [The polymorphism of (CAG)_n repeats within androgen receptor gene among Chinese male population]. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* **2001**,*18* (6), 456-8.

155. Sapir-Koren, R.; Livshits, G.; Landsman, T.; Kobylansky, E., Bone mineral density is associated with estrogen receptor gene polymorphism in men. *Anthropol Anz* **2001**,*59* (4), 343-53.

156. Aneja, A.; El-Atat, F.; McFarlane, S. I.; Sowers, J. R., Hypertension and obesity. *Recent Prog Horm Res* **2004**,*59*, 169-205.

157. Zitzmann, M.; Brune, M.; Kornmann, B.; Gromoll, J.; Junker, R.; Nieschlag, E., The CAG repeat polymorphism in the androgen receptor gene affects bone density and bone metabolism in healthy males. *Clin Endocrinol (Oxf)* **2001**,*55* (5), 649-57.

158. Zitzmann, M.; Junker, R.; Kamischke, A.; Nieschlag, E., Contraceptive steroids influence the hemostatic activation state in healthy men. *J Androl* **2002**,*23* (4), 503-11.

159. Whitsel, E. A.; Boyko, E. J.; Matsumoto, A. M.; Anawalt, B. D.; Siscovick, D. S., Intramuscular testosterone esters and plasma lipids in hypogonadal men: a meta-analysis. *Am J Med* **2001**,*111* (4), 261-9.

160. Dejager, S.; Bry-Gaillard, H.; Bruckert, E.; Eymard, B.; Salachas, F.; LeGuern, E.; Tardieu, S.; Chadarevian, R.; Giral, P.; Turpin, G., A comprehensive endocrine description of Kennedy's disease revealing androgen insensitivity linked to CAG repeat length. *J Clin Endocrinol Metab* **2002**,*87* (8), 3893-901.

161. Arbizu, T.; Santamaria, J.; Gomez, J. M.; Quilez, A.; Serra, J. P., A family with adult spinal and bulbar muscular atrophy, X-linked inheritance and associated testicular failure. *J Neurol Sci* **1983**,*59* (3), 371-82.

162. Liggett, S. B., Polymorphisms of adrenergic receptors: variations on a theme. *Assay Drug Dev Technol* **2003**,*1* (2), 317-26.

163. Small, K. M.; McGraw, D. W.; Liggett, S. B., Pharmacology and physiology of human adrenergic receptor polymorphisms. *Annu Rev Pharmacol Toxicol* **2003**,*43*, 381-411.

164. Garland, E. M.; Biaggioni, I., Genetic polymorphisms of adrenergic receptors. *Clin Auton Res* **2001**,*11* (2), 67-78.

165. Leineweber, K.; Buscher, R.; Bruck, H.; Brodde, O. E., Beta-adrenoceptor polymorphisms. *Naunyn Schmiedebergs Arch Pharmacol* **2004**,*369* (1), 1-22.

166. Feldman, R. D., Beta-adrenergic receptor alterations in hypertension--physiological and molecular correlates. *Can J Physiol Pharmacol* **1987**,*65* (8), 1666-72.

167. Larsson, I.; Berteus Forslund, H.; Lindroos, A. K.; Lissner, L.; Naslund, I.; Peltonen, M.; Sjostrom, L., Body composition in the SOS (Swedish Obese Subjects) reference study. *Int J Obes Relat Metab Disord* **2004**,*28* (10), 1317-24.

168. Feldman, R. D., Defective venous beta-adrenergic response in borderline hypertensive subjects is corrected by a low sodium diet. *J Clin Invest* **1990**,*85* (3), 647-52.

169. Stein, C. M.; Nelson, R.; Deegan, R.; He, H.; Wood, M.; Wood, A. J., Forearm beta adrenergic receptor-mediated vasodilation is impaired, without alteration of forearm norepinephrine spillover, in borderline hypertension. *J Clin Invest* **1995**,*96* (1), 579-85.
170. Burt, V. L.; Whelton, P.; Roccella, E. J.; Brown, C.; Cutler, J. A.; Higgins, M.; Horan, M. J.; Labarthe, D., Prevalence of hypertension in the US adult population. Results from the Third National Health and Nutrition Examination Survey, 1988-1991. *Hypertension* **1995**,*25* (3), 305-13.
171. Ma, J.; Stampfer, M. J.; Giovannucci, E.; Artigas, C.; Hunter, D. J.; Fuchs, C.; Willett, W. C.; Selhub, J.; Hennekens, C. H.; Rozen, R., Methylenetetrahydrofolate reductase polymorphism, dietary interactions, and risk of colorectal cancer. *Cancer Res* **1997**,*57* (6), 1098-102.
172. The HDFP Cooperative Group, Race, education and prevalence of hypertension. *Am J Epidemiol* **1997**,*106*, 351-61.
173. Budai, B.; Hitre, E.; Adleff, V.; Czeglédi, F.; Gyergyay, F.; Lang, I.; Kralovanszky, J., [The clinical importance of methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism in the 5-fluoropyrimidine-based therapy of metastatic colorectal tumours]. *Magy Onkol* **2004**,*48* (3), 253-7.
174. Xie, H. G.; Stein, C. M.; Kim, R. B.; Gainer, J. V.; Sofowora, G.; Dishy, V.; Brown, N. J.; Goree, R. E.; Haines, J. L.; Wood, A. J., Human beta2-adrenergic receptor polymorphisms: no association with essential hypertension in black or white Americans. *Clin Pharmacol Ther* **2000**,*67* (6), 670-5.
175. Svetkey, L. P.; Timmons, P. Z.; Emovon, O.; Anderson, N. B.; Preis, L.; Chen, Y. T., Association of hypertension with beta2- and alpha2c10-adrenergic receptor genotype. *Hypertension* **1996**,*27* (6), 1210-5.
176. Svetkey, L. P.; Chen, Y. T.; McKeown, S. P.; Preis, L.; Wilson, A. F., Preliminary evidence of linkage of salt sensitivity in black Americans at the beta 2-adrenergic receptor locus. *Hypertension* **1997**,*29* (4), 918-22.
177. Kotanko, P.; Binder, A.; Tasker, J.; DeFreitas, P.; Kamdar, S.; Clark, A. J.; Skrabal, F.; Caulfield, M., Essential hypertension in African Caribbeans associates with a variant of the beta2-adrenoceptor. *Hypertension* **1997**,*30* (4), 773-6.
178. Timmermann, B.; Mo, R.; Luft, F. C.; Gerdts, E.; Busjahn, A.; Omvik, P.; Li, G. H.; Schuster, H.; Wienker, T. F.; Hoehe, M. R.; Lund-Johansen, P., Beta-2 adrenoceptor genetic variation is associated with genetic predisposition to essential hypertension: The Bergen Blood Pressure Study. *Kidney Int* **1998**,*53* (6), 1455-60.
179. Arner, P.; Hoffstedt, J., Adrenoceptor genes in human obesity. *J Intern Med* **1999**,*245* (6), 667-72.
180. Arner, P., Obesity--a genetic disease of adipose tissue? *Br J Nutr* **2000**,*83 Suppl 1*, S9-16.
181. Green, S. A.; Turki, J.; Hall, I. P.; Liggett, S. B., Implications of genetic variability of human beta 2-adrenergic receptor structure. *Pulm Pharmacol* **1995**,*8* (1), 1-10.
182. Large, V.; Hellstrom, L.; Reynisdottir, S.; Lonnqvist, F.; Eriksson, P.; Lannfelt, L.; Arner, P., Human beta-2 adrenoceptor gene polymorphisms are highly frequent in obesity and associate with altered adipocyte beta-2 adrenoceptor function. *J Clin Invest* **1997**,*100* (12), 3005-13.

183. Ishiyama-Shigemoto, S.; Yamada, K.; Yuan, X.; Ichikawa, F.; Nonaka, K., Association of polymorphisms in the beta2-adrenergic receptor gene with obesity, hypertriglyceridaemia, and diabetes mellitus. *Diabetologia* **1999**,*42* (1), 98-101.
184. Echwald, S. M.; Sorensen, T. I.; Tybjaerg-Hansen, A.; Andersen, T.; Pedersen, O., Gln27Glu variant of the human beta2-adrenoreceptor gene is not associated with early-onset obesity in Danish men. *Diabetes* **1998**,*47* (10), 1657-8.
185. Wilks, R.; McFarlane-Anderson, N.; Bennett, F.; Fraser, H.; McGee, D.; Cooper, R.; Forrester, T., Obesity in peoples of the African diaspora. *Ciba Found Symp* **1996**,*201*, 37-48; discussion 48-53, 188-93.
186. Rueda-Clausen, C. F.; Silva, F. A.; Lopez-Jaramillo, P., Epidemic of overweight and obesity in Latin America and the Caribbean. *Int J Cardiol* **2008**,*125* (1), 111-2.
187. Wilks, R., Clinical aspects of Obesity and Diabetes Mellitus. *West Indian Med J* **2002**,*51* (Suppl. 1), 26.
188. Papas, M. A.; Alberg, A. J.; Ewing, R.; Helzlsouer, K. J.; Gary, T. L.; Klassen, A. C., The built environment and obesity. *Epidemiol Rev* **2007**,*29*, 129-43.
189. Hill, J. O.; Wyatt, H. R.; Reed, G. W.; Peters, J. C., Obesity and the environment: where do we go from here? *Science* **2003**,*299* (5608), 853-5.
190. Centers for Disease Control and Prevention About Healthy Places. <http://www.cdc.gov/healthyplaces/about.htm>.
191. Caballero, B., The global epidemic of obesity: an overview. *Epidemiol Rev* **2007**,*29*, 1-5.
192. Nielsen, S. J.; Siega-Riz, A. M.; Popkin, B. M., Trends in energy intake in U.S. between 1977 and 1996: similar shifts seen across age groups. *Obes Res* **2002**,*10* (5), 370-8.
193. Drewnowski, A., The real contribution of added sugars and fats to obesity. *Epidemiol Rev* **2007**,*29*, 160-71.
194. Bowman, S. A.; Gortmaker, S. L.; Ebbeling, C. B.; Pereira, M. A.; Ludwig, D. S., Effects of fast-food consumption on energy intake and diet quality among children in a national household survey. *Pediatrics* **2004**,*113* (1 Pt 1), 112-8.
195. McCrory, M. A.; Fuss, P. J.; Hays, N. P.; Vinken, A. G.; Greenberg, A. S.; Roberts, S. B., Overeating in America: association between restaurant food consumption and body fatness in healthy adult men and women ages 19 to 80. *Obes Res* **1999**,*7* (6), 564-71.
196. Zizza, C.; Siega-Riz, A. M.; Popkin, B. M., Significant increase in young adults' snacking between 1977-1978 and 1994-1996 represents a cause for concern! *Prev Med* **2001**,*32* (4), 303-10.
197. Dubbert, P. M.; Carithers, T.; Sumner, A. E.; Barbour, K. A.; Clark, B. L.; Hall, J. E.; Crook, E. D., Obesity, physical inactivity, and risk for cardiovascular disease. *Am J Med Sci* **2002**,*324* (3), 116-26.
198. President's Council on Physical Fitness and Sports *Healthy people 2010: Physical Activity and Fitness*; Washington, D.C., 2001.
199. Jonker, J. T.; De Laet, C.; Franco, O. H.; Peeters, A.; Mackenbach, J.; Nusselder, W. J., Physical activity and life expectancy with and without diabetes: life table analysis of the Framingham Heart Study. *Diabetes Care* **2006**,*29* (1), 38-43.
200. Cefalu, W. T., Insulin resistance: cellular and clinical concepts. *Exp Biol Med (Maywood)* **2001**,*226* (1), 13-26.

201. Yamashita, S.; Nakamura, T.; Shimomura, I.; Nishida, M.; Yoshida, S.; Kotani, K.; Kameda-Takemuara, K.; Tokunaga, K.; Matsuzawa, Y., Insulin resistance and body fat distribution. *Diabetes Care* **1996**,*19* (3), 287-91.
202. Sanchez-Castillo, C. P.; Velasquez-Monroy, O.; Lara-Esqueda, A.; Berber, A.; Sepulveda, J.; Tapia-Conyer, R.; James, W. P., Diabetes and hypertension increases in a society with abdominal obesity: results of the Mexican National Health Survey 2000. *Public Health Nutr* **2005**,*8* (1), 53-60.
203. Weiss, R.; Caprio, S., The metabolic consequences of childhood obesity. *Best Pract Res Clin Endocrinol Metab* **2005**,*19* (3), 405-19.
204. Caprio, S.; Tamborlane, W. V., Metabolic impact of obesity in childhood. *Endocrinol Metab Clin North Am* **1999**,*28* (4), 731-47.
205. Abate, N.; Garg, A.; Peshock, R. M.; Stray-Gundersen, J.; Adams-Huet, B.; Grundy, S. M., Relationship of generalized and regional adiposity to insulin sensitivity in men with NIDDM. *Diabetes* **1996**,*45* (12), 1684-93.
206. Wong, S.; Janssen, I.; Ross, R., Abdominal adipose tissue distribution and metabolic risk. *Sports Med* **2003**,*33* (10), 709-26.
207. Kelley, D. E.; Thaete, F. L.; Troost, F.; Huwe, T.; Goodpaster, B. H., Subdivisions of subcutaneous abdominal adipose tissue and insulin resistance. *Am J Physiol Endocrinol Metab* **2000**,*278* (5), E941-8.
208. Goodpaster, B. H.; Thaete, F. L.; Kelley, D. E., Thigh adipose tissue distribution is associated with insulin resistance in obesity and in type 2 diabetes mellitus. *Am J Clin Nutr* **2000**,*71* (4), 885-92.
209. Krssak, M.; Falk Petersen, K.; Dresner, A.; DiPietro, L.; Vogel, S. M.; Rothman, D. L.; Roden, M.; Shulman, G. I., Intramyocellular lipid concentrations are correlated with insulin sensitivity in humans: a ¹H NMR spectroscopy study. *Diabetologia* **1999**,*42* (1), 113-6.
210. Phillips, D. I.; Caddy, S.; Ilic, V.; Fielding, B. A.; Frayn, K. N.; Borthwick, A. C.; Taylor, R., Intramuscular triglyceride and muscle insulin sensitivity: evidence for a relationship in nondiabetic subjects. *Metabolism* **1996**,*45* (8), 947-50.
211. Pan, D. A.; Lillioja, S.; Kriketos, A. D.; Milner, M. R.; Baur, L. A.; Bogardus, C.; Jenkins, A. B.; Storlien, L. H., Skeletal muscle triglyceride levels are inversely related to insulin action. *Diabetes* **1997**,*46* (6), 983-8.
212. Vessby, B., Dietary fat and insulin action in humans. *Br J Nutr* **2000**,*83* Suppl 1, S91-6.
213. Ashwell, M.; Chinn, S.; Stalley, S.; Garrow, J. S., Female fat distribution-a simple classification based on two circumference measurements. *Int J Obes* **1982**,*6* (2), 143-52.
214. Despres, J. P.; Lemieux, I.; Prud'homme, D., Treatment of obesity: need to focus on high risk abdominally obese patients. *Bmj* **2001**,*322* (7288), 716-20.
215. Armellini, F.; Zamboni, M.; Rigo, L.; Bergamo-Andreis, I. A.; Robbi, R.; De Marchi, M.; Bosello, O., Sonography detection of small intra-abdominal fat variations. *Int J Obes* **1991**,*15* (12), 847-52.
216. Lemieux, S.; Prud'homme, D.; Tremblay, A.; Bouchard, C.; Despres, J. P., Anthropometric correlates to changes in visceral adipose tissue over 7 years in women. *Int J Obes Relat Metab Disord* **1996**,*20* (7), 618-24.
217. Pouliot, M. C.; Despres, J. P.; Lemieux, S.; Moorjani, S.; Bouchard, C.; Tremblay, A.; Nadeau, A.; Lupien, P. J., Waist circumference and abdominal sagittal diameter:

- best simple anthropometric indexes of abdominal visceral adipose tissue accumulation and related cardiovascular risk in men and women. *Am J Cardiol* **1994**,*73* (7), 460-8.
218. Lean, M. E.; Han, T. S.; Deurenberg, P., Predicting body composition by densitometry from simple anthropometric measurements. *Am J Clin Nutr* **1996**,*63* (1), 4-14.
219. Dobbelsteyn, C. J.; Joffres, M. R.; MacLean, D. R.; Flowerdew, G., A comparative evaluation of waist circumference, waist-to-hip ratio and body mass index as indicators of cardiovascular risk factors. The Canadian Heart Health Surveys. *Int J Obes Relat Metab Disord* **2001**,*25* (5), 652-61.
220. Okosun, I. S.; Liao, Y.; Rotimi, C. N.; Choi, S.; Cooper, R. S., Predictive values of waist circumference for dyslipidemia, type 2 diabetes and hypertension in overweight White, Black, and Hispanic American adults. *J Clin Epidemiol* **2000**,*53* (4), 401-8.
221. Patel, S.; Unwin, N.; Bhopal, R.; White, M.; Harland, J.; Ayis, S. A.; Watson, W.; Alberti, K. G., A comparison of proxy measures of abdominal obesity in Chinese, European and South Asian adults. *Diabet Med* **1999**,*16* (10), 853-60.
222. Okosun, I. S.; Rotimi, C. N.; Forrester, T. E.; Fraser, H.; Osotimehin, B.; Muna, W. F.; Cooper, R. S., Predictive value of abdominal obesity cut-off points for hypertension in blacks from west African and Caribbean island nations. *Int J Obes Relat Metab Disord* **2000**,*24* (2), 180-6.
223. Lev-Ran, A., Human obesity: an evolutionary approach to understanding our bulging waistline. *Diabetes Metab Res Rev* **2001**,*17* (5), 347-62.
224. Wang, Y.; Beydoun, M. A., The obesity epidemic in the United States--gender, age, socioeconomic, racial/ethnic, and geographic characteristics: a systematic review and meta-regression analysis. *Epidemiol Rev* **2007**,*29*, 6-28.
225. Wang, Y.; Rimm, E. B.; Stampfer, M. J.; Willett, W. C.; Hu, F. B., Comparison of abdominal adiposity and overall obesity in predicting risk of type 2 diabetes among men. *Am J Clin Nutr* **2005**,*81* (3), 555-63.
226. Yang, W.; Kelly, T.; He, J., Genetic epidemiology of obesity. *Epidemiol Rev* **2007**,*29*, 49-61.
227. Mittelman, S. D.; Van Citters, G. W.; Kirkman, E. L.; Bergman, R. N., Extreme insulin resistance of the central adipose depot in vivo. *Diabetes* **2002**,*51* (3), 755-61.
228. Griffin, M. E.; Marcucci, M. J.; Cline, G. W.; Bell, K.; Barucci, N.; Lee, D.; Goodyear, L. J.; Kraegen, E. W.; White, M. F.; Shulman, G. I., Free fatty acid-induced insulin resistance is associated with activation of protein kinase C theta and alterations in the insulin signaling cascade. *Diabetes* **1999**,*48* (6), 1270-4.
229. Dresner, A.; Laurent, D.; Marcucci, M.; Griffin, M. E.; Dufour, S.; Cline, G. W.; Slezak, L. A.; Andersen, D. K.; Hundal, R. S.; Rothman, D. L.; Petersen, K. F.; Shulman, G. I., Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity. *J Clin Invest* **1999**,*103* (2), 253-9.
230. Boden, G.; Chen, X., Effects of fat on glucose uptake and utilization in patients with non-insulin-dependent diabetes. *J Clin Invest* **1995**,*96* (3), 1261-8.
231. Ginsberg, H. N., Insulin resistance and cardiovascular disease. *J Clin Invest* **2000**,*106* (4), 453-8.
232. Ginsberg, H. N.; Stalenhoef, A. F., The metabolic syndrome: targeting dyslipidaemia to reduce coronary risk. *J Cardiovasc Risk* **2003**,*10* (2), 121-8.
233. Brunzell, J. D.; Ayyobi, A. F., Dyslipidemia in the metabolic syndrome and type 2 diabetes mellitus. *Am J Med* **2003**,*115 Suppl 8A*, 24S-28S.

234. Purnell, J. Q.; Kahn, S. E.; Albers, J. J.; Nevin, D. N.; Brunzell, J. D.; Schwartz, R. S., Effect of weight loss with reduction of intra-abdominal fat on lipid metabolism in older men. *J Clin Endocrinol Metab* **2000**,*85* (3), 977-82.
235. Yudkin, J. S.; Stehouwer, C. D.; Emeis, J. J.; Coppack, S. W., C-reactive protein in healthy subjects: associations with obesity, insulin resistance, and endothelial dysfunction: a potential role for cytokines originating from adipose tissue? *Arterioscler Thromb Vasc Biol* **1999**,*19* (4), 972-8.
236. Fried, S. K.; Bunkin, D. A.; Greenberg, A. S., Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab* **1998**,*83* (3), 847-50.
237. Feinstein, R.; Kanety, H.; Papa, M. Z.; Lunenfeld, B.; Karasik, A., Tumor necrosis factor-alpha suppresses insulin-induced tyrosine phosphorylation of insulin receptor and its substrates. *J Biol Chem* **1993**,*268* (35), 26055-8.
238. Hsueh, W. A.; Quinones, M. J., Role of endothelial dysfunction in insulin resistance. *Am J Cardiol* **2003**,*92* (4A), 10J-17J.
239. Ali, A. T.; Crowther, N. J., Body fat distribution and insulin resistance. *S Afr Med J* **2005**,*95* (11), 878-80.
240. Punyadeera, C.; Crowther, N. J.; van der Merwe, M. T.; Toman, M.; Immelman, A. R.; Schlaphoff, G. P.; Gray, I. P., Metabolic response to a mixed meal in obese and lean women from two South african populations. *Obes Res* **2002**,*10* (12), 1207-16.
241. Buthelezi, E. P.; van der Merwe, M. T.; Lonroth, P. N.; Gray, I. P.; Crowther, N. J., Ethnic differences in the responsiveness of adipocyte lipolytic activity to insulin. *Obes Res* **2000**,*8* (2), 171-8.
242. Okosun, I. S.; Choi, S.; Dent, M. M.; Jobin, T.; Dever, G. E., Abdominal obesity defined as a larger than expected waist girth is associated with racial/ethnic differences in risk of hypertension. *J Hum Hypertens* **2001**,*15* (5), 307-12.
243. Okosun, I. S.; Liao, Y.; Rotimi, C. N.; Prewitt, T. E.; Cooper, R. S., Abdominal adiposity and clustering of multiple metabolic syndrome in White, Black and Hispanic americans. *Ann Epidemiol* **2000**,*10* (5), 263-70.
244. Despres, J. P.; Couillard, C.; Gagnon, J.; Bergeron, J.; Leon, A. S.; Rao, D. C.; Skinner, J. S.; Wilmore, J. H.; Bouchard, C., Race, visceral adipose tissue, plasma lipids, and lipoprotein lipase activity in men and women: the Health, Risk Factors, Exercise Training, and Genetics (HERITAGE) family study. *Arterioscler Thromb Vasc Biol* **2000**,*20* (8), 1932-8.
245. Cuddy, M. L., Treatment of hypertension: guidelines from JNC 7 (the seventh report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure 1). *J Pract Nurs* **2005**,*55* (4), 17-21; quiz 22-3.
246. Joffe, B. I.; Panz, V. R.; Wing, J. R.; Raal, F. J.; Seftel, H. C., Pathogenesis of non-insulin-dependent diabetes mellitus in the black population of southern Africa. *Lancet* **1992**,*340* (8817), 460-2.
247. Punyadeera, C.; van der Merwe, M. T.; Crowther, N. J.; Toman, M.; Immelman, A. R.; Schlaphoff, G. P.; Gray, I. P., Weight-related differences in glucose metabolism and free fatty acid production in two South African population groups. *Int J Obes Relat Metab Disord* **2001**,*25* (8), 1196-205.
248. Punyadeera, C.; van der Merwe, M. T.; Crowther, N. J.; Toman, M.; Schlaphoff, G. P.; Gray, I. P., Ethnic differences in lipid metabolism in two groups of obese South African women. *J Lipid Res* **2001**,*42* (5), 760-7.

249. van der Merwe, M. T.; Crowther, N. J.; Schlaphoff, G. P.; Gray, I. P.; Joffe, B. I.; Lonroth, P. N., Evidence for insulin resistance in black women from South Africa. *Int J Obes Relat Metab Disord* **2000**,*24* (10), 1340-6.
250. van der Merwe, M. T.; Crowther, N. J.; Schlaphoff, G. P.; Boyd, I. H.; Gray, I. P.; Joffe, B. I.; Lonroth, P. N., Lactate and glycerol release from the subcutaneous adipose tissue of obese urban women from South Africa; important metabolic implications. *J Clin Endocrinol Metab* **1998**,*83* (11), 4084-91.
251. van der Merwe, M. T.; Wing, J. R.; Celgow, L. H.; Gray, I. P.; Lonn, L.; Joffe, B. I.; Lonroth, P. N., Metabolic indices in relation to body composition changes during weight loss on Dexfenfluramine in obese women from two South African ethnic groups. *Int J Obes Relat Metab Disord* **1996**,*20* (8), 768-76.
252. van der Merwe, M.; VR, P.; Crowther, N.; Schlaphoff, G. P.; Gray, I. P., Free fatty acids and insulin levels-relationship to leptin levels and body composition in various patient groups in South Africa. *Int J Obes Relat Metab Disord* **1999**,*23*, 909-917.
253. van der Merwe, M. T.; Pepper, M. S., Obesity in South Africa. *Obes Rev* **2006**,*7* (4), 315-22.
254. Ford, E. S.; Giles, W. H.; Dietz, W. H., Prevalence of the metabolic syndrome among US adults: findings from the third National Health and Nutrition Examination Survey. *JAMA* **2002**,*287* (3), 356-9.
255. Sattar, N.; McConnachie, A.; Shaper, A. G.; Blauw, G. J.; Buckley, B. M.; de Craen, A. J.; Ford, I.; Forouhi, N. G.; Freeman, D. J.; Jukema, J. W.; Lennon, L.; Macfarlane, P. W.; Murphy, M. B.; Packard, C. J.; Stott, D. J.; Westendorp, R. G.; Whincup, P. H.; Shepherd, J.; Wannamethee, S. G., Can metabolic syndrome usefully predict cardiovascular disease and diabetes? Outcome data from two prospective studies. *Lancet* **2008**,*371* (9628), 1927-35.
256. Bublely, G. J.; Carducci, M.; Dahut, W.; Dawson, N.; Daliani, D.; Eisenberger, M.; Figg, W. D.; Freidlin, B.; Halabi, S.; Hudes, G.; Hussain, M.; Kaplan, R.; Myers, C.; Oh, W.; Petrylak, D. P.; Reed, E.; Roth, B.; Sartor, O.; Scher, H.; Simons, J.; Sinibaldi, V.; Small, E. J.; Smith, M. R.; Trump, D. L.; Wilding, G.; et al., Eligibility and response guidelines for phase II clinical trials in androgen-independent prostate cancer: recommendations from the Prostate-Specific Antigen Working Group. *J Clin Oncol* **1999**,*17* (11), 3461-7.
257. Reaven, G. M., Insulin resistance, hyperinsulinemia, hypertriglyceridemia and hypertension. Parallels between human disease and rodent models. *Diabetes Care* **1991**,*14*, 195-202.
258. Miller, G. J.; Beckles, G. L.; Byam, N. T.; Price, S. G.; Carson, D. C.; Kirkwood, B. R., Serum lipoprotein concentrations in relation to ethnic composition and urbanisation in men and women of Trinidad, west Indies. *Int J Epidemiol* **1984**,*13*, 413-21.
259. Fraser, H., Hypertension: The silent killer and the deadly quartet. *West Indian Med J* **2000**,*49* (2), 91.
260. Brouwer, B. G.; Visseren, F. L.; van der Graaf, Y., The effect of leisure-time physical activity on the presence of metabolic syndrome in patients with manifest arterial disease. The SMART study. *Am Heart J* **2007**,*154* (6), 1146-52.
261. Lakka, T. A.; Lakka, H. M.; Rankinen, T.; Leon, A. S.; Rao, D. C.; Skinner, J. S.; Wilmore, J. H.; Bouchard, C., Effect of exercise training on plasma levels of C-reactive protein in healthy adults: the HERITAGE Family Study. *Eur Heart J* **2005**,*26* (19), 2018-25.
262. Diamant, M.; Tushuizen, M. E., The metabolic syndrome and endothelial dysfunction: common highway to type 2 diabetes and CVD. *Curr Diab Rep* **2006**,*6* (4), 279-86.

263. Nistala, R.; Stump, C. S., Skeletal muscle insulin resistance is fundamental to the cardiometabolic syndrome. *J Cardiometab Syndr* **2006**,*1* (1), 47-52.
264. McKeown, N. M.; Meigs, J. B.; Liu, S.; Saltzman, E.; Wilson, P. W.; Jacques, P. F., Carbohydrate nutrition, insulin resistance, and the prevalence of the metabolic syndrome in the Framingham Offspring Cohort. *Diabetes Care* **2004**,*27* (2), 538-46.
265. Lutsey, P. L.; Steffen, L. M.; Stevens, J., Dietary intake and the development of the metabolic syndrome: the Atherosclerosis Risk in Communities study. *Circulation* **2008**,*117* (6), 754-61.
266. Mennen, L. I.; Lafay, I.; Feskens, E.; Novak, M.; Lepinay, P.; Balkau, B., Possible protective effect of bread and dairy products on the risk of metabolic syndrome. *Nutr Res* **2000**,*20*, 335-347.
267. Pereira, M.; Jacobs, D. R., Jr.; Van Horn, L.; Slattery, M. L.; Kartashov, A. I., Dairy consumption, obesity, and the insulin resistance syndrome in young adults: the CARDIA Study. *JAMA* **2002**,*287*, 2081-2089.
268. Azadbakht, L.; Mirmiran, P.; Azizi, F., Dairy consumption is inversely associated with the prevalence of the metabolic syndrome in Tehranian adults. *Am J Clin Nutr* **2005**,*82*, 523-530.
269. Chobanian, A. V.; Bakris, G. L.; Black, H. R.; Cushman, W. C.; Green, L. A.; Izzo, J. L., Jr.; Jones, D. W.; Materson, B. J.; Oparil, S.; Wright, J. T., Jr.; Roccella, E. J., The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. *Jama* **2003**,*289* (19), 2560-72.
270. Goodpaster, B. H., Measuring body fat distribution and content in humans. *Curr Opin Clin Nutr Metab Care* **2002**,*5* (5), 481-7.
271. Mazess, R. B.; Barden, H. S.; Bisek, J. P.; Hanson, J., Dual-energy x-ray absorptiometry for total-body and regional bone-mineral and soft-tissue composition. *Am J Clin Nutr* **1990**,*51* (6), 1106-12.
272. Goodpaster, B. H.; Kelley, D. E.; Thaete, F. L.; He, J.; Ross, R., Skeletal muscle attenuation determined by computed tomography is associated with skeletal muscle lipid content. *J Appl Physiol* **2000**,*89* (1), 104-10.
273. Nagy, T. R.; Johnson, m. S., Measurement of body and liver fat in small animals using peripheral quantitative computed tomography. *International Journal of Body Composition Research* **2004**,*1* (4), 155-160.
274. Ranke, M. B.; Schweizer, R.; Martin, D. D.; Trebar, B., Effects of Gh therapy, muscle and fat mass (pQCT) and metabolism in prepubertal children with Turner syndrome University Children Hospital, Paediatric Endocrinology Section, Tuebingen, Germany.
275. Narayan, K. M.; Boyle, J. P.; Thompson, T. J.; Gregg, E. W.; Williamson, D. F., Effect of BMI on lifetime risk for diabetes in the U.S. *Diabetes Care* **2007**,*30* (6), 1562-6.
276. Centers for Disease Control and Prevention National diabetes fact sheet: national estimates on diabetes. <http://www.cdc.gov/diabetes/pubs/estimates.htm#prev>.
277. Karp, G., *Cell and mecular biology: concepts and experiments*. 2nd edition ed.; John Wiley and sons, Inc: 1996.
278. Abel, D. E.; Peroni, O.; Kim, J. K.; Kim, Y.-b.; Boss, O.; Hadro, E.; Minnemann, T.; Shulman, G. I.; Khan, B. B., Adipose-selection targeting of the GLUT4 gene impairs insulin action in muscle and liver. *Nature* **2001**,*409* (8), 729.
279. Saltiel, A. R.; Kahn, C. R., Insulin signalling and the regulation of glucose and lipid metabolism. *Nature* **2001**,*414* (6865), 799-806.

280. Kahn, S. E.; Prigeon, R. L.; Schwartz, R. S.; Fujimoto, W. Y.; Knopp, R. H.; Brunzell, J. D.; Porte, D., Jr., Obesity, body fat distribution, insulin sensitivity and Islet beta-cell function as explanations for metabolic diversity. *J Nutr* **2001**,*131* (2), 354S-60S.
281. Chen, M.; Bergman, R. N.; Pacini, G.; Porte, D., Jr., Pathogenesis of age-related glucose intolerance in man: insulin resistance and decreased beta-cell function. *J Clin Endocrinol Metab* **1985**,*60* (1), 13-20.
282. Mathis, D.; Vence, L.; Benoist, C., beta-Cell death during progression to diabetes. *Nature* **2001**,*414* (6865), 792-8.
283. DeFronzo, R. A.; Ferrannini, E., Insulin resistance. A multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* **1991**,*14* (3), 173-94.
284. Miles, J.; Jensen, M., Counterpoint: visceral adiposity is not causally related to insulin resistance. *Diabetes Care* **2005**,*28*, 2326-2328.
285. Iwashima, Y.; Katsuya, T.; Ishikawa, K.; Ouchi, N.; Ohishi, M.; Sugimoto, K.; Fu, Y.; Motone, M.; Yamamoto, K.; Matsuo, A.; Ohashi, K.; Kihara, S.; Funahashi, T.; Rakugi, H.; Matsuzawa, Y.; Ogihara, T., Hypoadiponectinemia is an independent risk factor for hypertension. *Hypertension* **2004**,*43* (6), 1318-23.
286. Kershaw, E. E.; Flier, J. S., Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab* **2004**,*89* (6), 2548-56.
287. Arner, P., Insulin resistance in type 2 diabetes -- role of the adipokines. *Curr Mol Med* **2005**,*5* (3), 333-9.
288. Fantuzzi, G., Adipose tissue, adipokines, and inflammation. *J Allergy Clin Immunol* **2005**,*115* (5), 911-9; quiz 920.
289. Zhao, Y. F.; Feng, D. D.; Chen, C., Contribution of adipocyte-derived factors to beta-cell dysfunction in diabetes. *Int J Biochem Cell Biol* **2006**,*38* (5-6), 804-19.
290. Arita, Y.; Kihara, S.; Ouchi, N.; Takahashi, M.; Maeda, K.; Miyagawa, J.; Hotta, K.; Shimomura, I.; Nakamura, T.; Miyaoka, K.; Kuriyama, H.; Nishida, M.; Yamashita, S.; Okubo, K.; Matsubara, K.; Muraguchi, M.; Ohmoto, Y.; Funahashi, T.; Matsuzawa, Y., Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* **1999**,*257* (1), 79-83.
291. Schondorf, T., Biological background and role of adiponectin as a marker for insulin resistance and cardiovascular risk. *Clin Lab* **2005**,*51* (9-10), 489-494.
292. Khan, R. C., Causes of insulin resistance. *Nature* **1995**,*373* (2).
293. Hennis, A.; Wu, S. Y.; Nemesure, B.; Li, X.; Leske, M. C., Diabetes in a Caribbean population: epidemiological profile and implications. *Int J Epidemiol* **2002**,*31* (1), 234-9.
294. Beckles, G. L.; Miller, G. J.; Kirkwood, B. R.; Alexis, S. D.; Carson, D. C.; Byam, N. T., High total and cardiovascular disease mortality in adults of Indian descent in Trinidad, unexplained by major coronary risk factors. *Lancet* **1986**,*1* (8493), 1298-301.
295. Miller, G. J.; Kirkwood, B. R.; Beckles, G. L.; Alexis, S. D.; Carson, D. C.; Byam, N. T., Adult male all-cause, cardiovascular and cerebrovascular mortality in relation to ethnic group, systolic blood pressure and blood glucose concentration in Trinidad, West Indies. *Int J Epidemiol* **1988**,*17* (1), 62-9.
296. Gulliford, M. C., Epidemiological transition in Trinidad and Tobago, West Indies 1953-1992. *Int J Epidemiol* **1996**,*25* (2), 357-65.

297. Wilks, R.; Rotimi, C.; Bennett, F.; McFarlane-Anderson, N.; Kaufman, J. S.; Anderson, S. G.; Cooper, R. S.; Cruickshank, J. K.; Forrester, T., Diabetes in the Caribbean: results of a population survey from Spanish Town, Jamaica. *Diabet Med* **1999**,*16* (10), 875-83.
298. Alleyne, S. I.; Cruickshank, J. K.; Golding, A. L.; Morrison, E. Y., Mortality from diabetes mellitus in Jamaica. *Bull Pan Am Health Organ* **1989**,*23* (3), 306-14.
299. Poon-King, T.; Henry, M. V.; Rampersad, F., Prevalence and natural history of diabetes in Trinidad. *Lancet* **1968**,*1*, 842-847.
300. King, H.; Aubert, R.; Herman, W., Global burden of diabetes, 1995-2025. Prevalence, numerical estimates, and projections. *Diabetes Care* **1998**,*21*, 1414-31.
301. Gulliford, M. C.; Ariyanayagam-Baksh, S. M.; Bickram, L.; Picou, D.; Mahabir, D., Counting the cost of diabetic hospital admissions from a multi-ethnic population in Trinidad. *Diabet Med* **1995**,*12* (12), 1077-85.
302. Ezenwaka, C. E.; Offiah, N. V., Abdominal obesity in type 2 diabetic patients visiting primary healthcare clinics in Trinidad, West Indies. *Scand J Prim Health Care* **2002**,*20* (3), 177-82.
303. Cooper, R. S.; Rotimi, C. N.; Kaufman, J. S.; Owoaje, E. E.; Fraser, H.; Forrester, T.; Wilks, R.; Riste, L. K.; Cruickshank, J. K., Prevalence of NIDDM among populations of the African diaspora. *Diabetes Care* **1997**,*20* (3), 343-8.
304. Hugh-Jones, P., Diabetes in Jamaica. *Lancet* **1955**,*269* (6896), 891-7.
305. Tulloch, J. A.; Johnson, H. M., A pilot survey of the incidence of diabetes in Jamaica. *West Indian Med J* **1958**,*7* (2), 134-6.
306. Florey Cdu, V.; McDonald, H.; McDonald, J.; Miall, W. E., The prevalence of diabetes in a rural population of Jamaican adults. *Int J Epidemiol* **1972**,*1* (2), 157-66.
307. Beevers, G.; Lip, G. Y.; O'Brien, E., ABC of hypertension: The pathophysiology of hypertension. *Bmj* **2001**,*322* (7291), 912-6.
308. American Heart Association *Heart and stroke facts: 1996 statistical supplements*; Dallas.
309. Guyton; Ganong physiology II. Pathophysiology of Hypertension. <http://www.hodsonhome.com/mna2001/physiology/physiology2/exam2/phys2.pathohtn.htm>.
310. McCance, K. L.; Huether, S. E., *Pathophysiology : The biologic basis for disease in adults and children*. 1997.
311. Long, A. E.; Prewitt, T. E.; Kaufman, J. S.; Rotimi, C. N.; Cooper, R. S.; McGee, D. L., Weight-height relationships among eight populations of West African origin: the case against constant BMI standards. *Int J Obes Relat Metab Disord* **1998**,*22* (9), 842-6.
312. Hu, F. B.; Wang, B.; Chen, C.; Jin, Y.; Yang, J.; Stampfer, M. J.; Xu, X., Body mass index and cardiovascular risk factors in a rural Chinese population. *Am J Epidemiol* **2000**,*151* (1), 88-97.
313. Garrison, R. J.; Kannel, W. B.; Stokes, J., 3rd; Castelli, W. P., Incidence and precursors of hypertension in young adults: the Framingham Offspring Study. *Prev Med* **1987**,*16* (2), 235-51.
314. Huang, Z.; Willett, W. C.; Manson, J. E.; Rosner, B.; Stampfer, M. J.; Speizer, F. E.; Colditz, G. A., Body weight, weight change, and risk for hypertension in women. *Ann Intern Med* **1998**,*128* (2), 81-8.
315. Moore, L. L.; VISIONI, A. J.; Qureshi, M. M.; Bradlee, M. L.; Ellison, R. C.; D'Agostino, R., Weight loss in overweight adults and the long-term risk of hypertension: the Framingham study. *Arch Intern Med* **2005**,*165* (11), 1298-303.

316. Davy, K. P.; Hall, J. E., Obesity and hypertension: two epidemics or one? *Am J Physiol Regul Integr Comp Physiol* **2004**,*286* (5), R803-13.
317. White, J. V., Hypertension nutrition management for older adults. *American Academy of family physicians*.
318. Grassi, G.; Dell'Oro, R.; Facchini, A.; Quarti Trevano, F.; Bolla, G. B.; Mancia, G., Effect of central and peripheral body fat distribution on sympathetic and baroreflex function in obese normotensives. *J Hypertens* **2004**,*22* (12), 2363-9.
319. Grassi, G.; Dell'Oro, R.; Quarti-Trevano, F.; Scopelliti, F.; Seravalle, G.; Paleari, F.; Gamba, P. L.; Mancia, G., Neuroadrenergic and reflex abnormalities in patients with metabolic syndrome. *Diabetologia* **2005**,*48* (7), 1359-65.
320. Julius, S.; Jamerson, K., Sympathetics, insulin resistance and coronary risk in hypertension: the 'chicken-and-egg' question. *J Hypertens* **1994**,*12* (5), 495-502.
321. Julius, S.; Krause, L.; Schork, N. J.; Mejia, A. D.; Jones, K. A.; van de Ven, C.; Johnson, E. H.; Sekkarie, M. A.; Kjeldsen, S. E.; Petrin, J.; et al., Hyperkinetic borderline hypertension in Tecumseh, Michigan. *J Hypertens* **1991**,*9* (1), 77-84.
322. Grassi, G.; Seravalle, G.; Cattaneo, B. M.; Bolla, G. B.; Lanfranchi, A.; Colombo, M.; Giannattasio, C.; Brunani, A.; Cavagnini, F.; Mancia, G., Sympathetic activation in obese normotensive subjects. *Hypertension* **1995**,*25* (4 Pt 1), 560-3.
323. Grassi, G., Renin-angiotensin-sympathetic crosstalks in hypertension: reappraising the relevance of peripheral interactions. *J Hypertens* **2001**,*19* (10), 1713-6.
324. Egan, B. M., Insulin resistance and the sympathetic nervous system. *Curr Hypertens Rep* **2003**,*5* (3), 247-54.
325. Sarafidis, P. A.; Bakris, G. L., Non-esterified fatty acids and blood pressure elevation: a mechanism for hypertension in subjects with obesity/insulin resistance? *J Hum Hypertens* **2007**,*21* (1), 12-9.
326. Correia, M. L.; Haynes, W. G., Obesity-related hypertension: is there a role for selective leptin resistance? *Curr Hypertens Rep* **2004**,*6* (3), 230-5.
327. Mancia, G.; Bousquet, P.; Elghozi, J. L.; Esler, M.; Grassi, G.; Julius, S.; Reid, J.; Van Zwieten, P. A., The sympathetic nervous system and the metabolic syndrome. *J Hypertens* **2007**,*25* (5), 909-20.
328. Roberts, C.; Barnard, R.; Sindhu, R.; Jurczak, M.; EHdaie, A.; Vaziri, N., A high fat, refined -carbohydrate diet induces endothelial dysfunction and oxidant/antioxidant imbalance and depresses NOS protein expression. *J Appl Physiol* **2005**,*98*, 203-210.
329. Sarafidis, P. A.; Bakris, G. L., The antinatriuretic effect of insulin: an unappreciated mechanism for hypertension associated with insulin resistance? *Am J Nephrol* **2007**,*27* (1), 44-54.
330. Furukawa, S.; Fujita, T.; Shimabukuro, M.; Iwaki, M.; Yamada, Y.; Nakajima, Y.; Nakayama, O.; Makishima, M.; Matsuda, M.; Shimomura, I., Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* **2004**,*114* (12), 1752-61.
331. Meigs, J. B.; Larson, M. G.; Fox, C. S.; Keaney, J. F., Jr.; Vasan, R. S.; Benjamin, E. J., Association of oxidative stress, insulin resistance, and diabetes risk phenotypes: the Framingham Offspring Study. *Diabetes Care* **2007**,*30* (10), 2529-35.
332. Kim, J. A.; Montagnani, M.; Koh, K. K.; Quon, M. J., Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms. *Circulation* **2006**,*113* (15), 1888-904.

333. Sesso, H. D.; Buring, J. E.; Rifai, N.; Blake, G. J.; Gaziano, J. M.; Ridker, P. M., C-reactive protein and the risk of developing hypertension. *Jama* **2003**,*290* (22), 2945-51.
334. Grundy, S. M., Inflammation, hypertension, and the metabolic syndrome. *Jama* **2003**,*290* (22), 3000-2.
335. Kahaleh, M. B.; Fan, P. S., Effect of cytokines on the production of endothelin by endothelial cells. *Clin Exp Rheumatol* **1997**,*15* (2), 163-7.
336. Brasier, A. R.; Li, J.; Wimbish, K. A., Tumor necrosis factor activates angiotensinogen gene expression by the Rel A transactivator. *Hypertension* **1996**,*27* (4), 1009-17.
337. Pausova, Z.; Deslauriers, B.; Gaudet, D.; Tremblay, J.; Kotchen, T. A.; Larochelle, P.; Cowley, A. W.; Hamet, P., Role of tumor necrosis factor-alpha gene locus in obesity and obesity-associated hypertension in French Canadians. *Hypertension* **2000**,*36* (1), 14-9.
338. Zinman, B.; Hanley, A. J.; Harris, S. B.; Kwan, J.; Fantus, I. G., Circulating tumor necrosis factor-alpha concentrations in a native Canadian population with high rates of type 2 diabetes mellitus. *J Clin Endocrinol Metab* **1999**,*84* (1), 272-8.
339. Dorffel, Y.; Latsch, C.; Stuhlmuller, B.; Schreiber, S.; Scholze, S.; Burmester, G. R.; Scholze, J., Preactivated peripheral blood monocytes in patients with essential hypertension. *Hypertension* **1999**,*34* (1), 113-7.
340. Fernandez-Real, J. M.; Vayreda, M.; Richart, C.; Gutierrez, C.; Broch, M.; Vendrell, J.; Ricart, W., Circulating interleukin 6 levels, blood pressure, and insulin sensitivity in apparently healthy men and women. *J Clin Endocrinol Metab* **2001**,*86* (3), 1154-9.
341. Papanicolaou, D. A.; Petrides, J. S.; Tsigos, C.; Bina, S.; Kalogeras, K. T.; Wilder, R.; Gold, P. W.; Deuster, P. A.; Chrousos, G. P., Exercise stimulates interleukin-6 secretion: inhibition by glucocorticoids and correlation with catecholamines. *Am J Physiol* **1996**,*271* (3 Pt 1), E601-5.
342. Besedovsky, H. O.; del Rey, A., Immune-neuro-endocrine interactions: facts and hypotheses. *Endocr Rev* **1996**,*17* (1), 64-102.
343. Torpy, D. J.; Papanicolaou, D. A.; Lotsikas, A. J.; Wilder, R. L.; Chrousos, G. P.; Pillemer, S. R., Responses of the sympathetic nervous system and the hypothalamic-pituitary-adrenal axis to interleukin-6: a pilot study in fibromyalgia. *Arthritis Rheum* **2000**,*43* (4), 872-80.
344. Takano, M.; Itoh, N.; Yayama, K.; Yamano, M.; Ohtani, R.; Okamoto, H., Interleukin-6 as a mediator responsible for inflammation-induced increase in plasma angiotensinogen. *Biochem Pharmacol* **1993**,*45* (1), 201-6.
345. Hennis, A.; Wu, S. Y.; Nemesure, B.; Leske, M. C., Hypertension prevalence, control and survivorship in an Afro-Caribbean population. *J Hypertens* **2002**,*20* (12), 2363-9.
346. Bobb-Liverpool, B.; Duff, E. M.; Bailey, E. Y., Compliance and blood pressure control in women with hypertension. *West Indian Med J* **2002**,*51* (4), 236-40.
347. Dyer-Regis, B., An assessment of health behaviour of chronic disease clients at health centers in St. Georges, West Trinidad. *West Indian Med J* **1996**,*45* (Suppl 2), 24-5.
348. Atallah, A.; Inamo, J.; Lang, T.; Larabi, L.; Chatellier, G.; Rozet, J. E.; Machuron, C.; De Gaudemaris, R., [Prevalence of hypertension in a disadvantaged population in Antilles: a major role for obesity?]. *Arch Mal Coeur Vaiss* **2007**,*100* (1), 22-7.
349. Lane, D.; Beevers, D. G.; Lip, G. Y., Ethnic differences in blood pressure and the prevalence of hypertension in England. *J Hum Hypertens* **2002**,*16* (4), 267-73.

350. Miller, G. J.; Maude, G. H.; Beckles, G. L., Incidence of hypertension and non-insulin dependent diabetes mellitus and associated risk factors in a rapidly developing Caribbean community: the St James survey, Trinidad. *J Epidemiol Community Health* **1996**,*50* (5), 497-504.
351. Ntais, C.; Polycarpou, A.; Tsatsoulis, A., Molecular epidemiology of prostate cancer: androgens and polymorphisms in androgen-related genes. *Eur J Endocrinol* **2003**,*149* (6), 469-77.
352. Miller, K. K., Androgen deficiency: effects on body composition. *Pituitary* **2009**,*12* (2), 116-24.
353. Osei-Assibey, G.; Kyrou, I.; Adi, Y.; Kumar, S.; Matyka, K., Dietary and lifestyle interventions for weight management in adults from minority ethnic/non-White groups: a systematic review. *Obes Rev*.
354. Ogden, C. L.; Carroll, M. D.; Curtin, L. R.; McDowell, M. A.; Tabak, C. J.; Flegal, K. M., Prevalence of overweight and obesity in the United States, 1999-2004. *JAMA* **2006**,*295* (13), 1549-55.
355. Joint Health Surveys Unit; National Centre for Social Research; Department of Epidemiology and Public Health the Royal Free and University College Medical School *Health Survey for England: The health of minority ethnic groups summary of key findings*; 2004.
356. O'Byrne, S.; Caulfield, M., Genetics of hypertension. Therapeutic implications. *Drugs* **1998**,*56* (2), 203-14.
357. Ferdinand, K. C.; Armani, A. M., The management of hypertension in African Americans. *Crit Pathw Cardiol* **2007**,*6* (2), 67-71.
358. Comuzzie, A. G.; Allison, D. B., The search for human obesity genes. *Science* **1998**,*280* (5368), 1374-7.
359. Pomp, D.; Mohlke, K. L., Obesity genes: so close and yet so far. *J Biol* **2008**,*7* (9), 36.
360. Meirhaeghe, A.; Helbecque, N.; Cottel, D.; Amouyel, P., Impact of polymorphisms of the human beta2-adrenoceptor gene on obesity in a French population. *International Journal of Obesity & Related Metabolic Disorders: Journal of the International Association for the Study of Obesity* **2000**,*24* (3), 382-7.
361. Mohr, B. A.; Bhasin, S.; Link, C. L.; O'Donnell, A. B.; McKinlay, J. B., The effect of changes in adiposity on testosterone levels in older men: longitudinal results from the Massachusetts Male Aging Study. *Eur J Endocrinol* **2006**,*155* (3), 443-52.
362. Gustafson, D. R.; Wen, M. J.; Koppanati, B. M., Androgen receptor gene repeats and indices of obesity in older adults. *Int J Obes Relat Metab Disord* **2003**,*27* (1), 75-81.
363. Pausova, Z.; Abrahamowicz, M.; Mahboubi, A.; Syme, C.; Leonard, G. T.; Perron, M.; Richer, L.; Veillette, S.; Gaudet, D.; Paus, T., Functional Variation in the Androgen-Receptor Gene Is Associated With Visceral Adiposity and Blood Pressure in Male Adolescents. *Hypertension*.
364. Andersson, B.; Marin, P.; Lissner, L.; Vermeulen, A.; Bjorntorp, P., Testosterone concentrations in women and men with NIDDM. *Diabetes Care* **1994**,*17* (5), 405-11.
365. Laaksonen, D. E.; Niskanen, L.; Punnonen, K.; Nyysönen, K.; Tuomainen, T. P.; Valkonen, V. P.; Salonen, R.; Salonen, J. T., Testosterone and sex hormone-binding globulin predict the metabolic syndrome and diabetes in middle-aged men. *Diabetes Care* **2004**,*27* (5), 1036-41.

366. Stellato, R. K.; Feldman, H. A.; Hamdy, O.; Horton, E. S.; McKinlay, J. B., Testosterone, sex hormone-binding globulin, and the development of type 2 diabetes in middle-aged men: prospective results from the Massachusetts male aging study. *Diabetes Care* **2000**,*23* (4), 490-4.
367. Singh, R.; Artaza, J. N.; Taylor, W. E.; Braga, M.; Yuan, X.; Gonzalez-Cadavid, N. F.; Bhasin, S., Testosterone inhibits adipogenic differentiation in 3T3-L1 cells: nuclear translocation of androgen receptor complex with beta-catenin and T-cell factor 4 may bypass canonical Wnt signaling to down-regulate adipogenic transcription factors. *Endocrinology* **2006**,*147* (1), 141-54.
368. Stanworth, R. D.; Kapoor, D.; Channer, K. S.; Jones, T. H., Androgen receptor CAG repeat polymorphism is associated with serum testosterone levels, obesity and serum leptin in men with type 2 diabetes. *Eur J Endocrinol* **2008**,*159* (6), 739-46.
369. Guadalupe-Grau, A.; Rodriguez-Gonzalez, G.; Dorado, C.; Olmedillas, H.; Fuentes, T.; Perez-Gomez, J.; Delgado-Guerra, S.; Vicente-Rodriguez, G.; Ara, I.; Guerra, B.; Arteaga-Ortiz, R.; Calbet, J. A.; Diaz-Chico, B. N., Androgen receptor gene polymorphisms lean mass and performance in young men. *Br J Sports Med* **2009**.
370. Vijayalakshmi, K.; Thangaraj, K.; Rajender, S.; Vettriselvi, V.; Venkatesan, P.; Shroff, S.; Vishwanathan, K. N.; Paul, S. F., GGN repeat length and GGN/CAG haplotype variations in the androgen receptor gene and prostate cancer risk in south Indian men. *J Hum Genet* **2006**,*51* (11), 998-1005.
371. Chamberlain, N. L.; Driver, E. D.; Miesfeld, R. L., The length and location of CAG trinucleotide repeats in the androgen receptor N-terminal domain affect transactivation function. *Nucleic Acids Res* **1994**,*22* (15), 3181-6.
372. Zitzmann, M.; Nieschlag, E., Androgens and vascular function. *J Endocrinol Invest* **2003**,*26* (8), 767-9.
373. Campbell, B. C.; Gray, P. B.; Eisenberg, D. T.; Ellison, P.; Sorenson, M. D., Androgen receptor CAG repeats and body composition among Ariaal men. *Int J Androl* **2009**,*32* (2), 140-8.
374. Lapauw, B.; Goemaere, S.; Crabbe, P.; Kaufman, J. M.; Ruige, J. B., Is the effect of testosterone on body composition modulated by the androgen receptor gene CAG repeat polymorphism in elderly men? *Eur J Endocrinol* **2007**,*156* (3), 395-401.
375. Nwosu, V.; Carpten, J.; Trent, J. M.; Sheridan, R., Heterogeneity of genetic alterations in prostate cancer: evidence of the complex nature of the disease. *Hum Mol Genet* **2001**,*10* (20), 2313-8.
376. Sartor, O.; Zheng, Q.; Eastham, J. A., Androgen receptor gene CAG repeat length varies in a race-specific fashion in men without prostate cancer. *Urology* **1999**,*53* (2), 378-80.
377. Wolk, A., Diet, lifestyle and risk of prostate cancer. *Acta Oncol* **2005**,*44* (3), 277-81.
378. Hsing, A. W.; Chu, L. W.; Stanczyk, F. Z., Androgen and prostate cancer: is the hypothesis dead? *Cancer Epidemiol Biomarkers Prev* **2008**,*17* (10), 2525-30.
379. Chen, A. C.; Petrylak, D. P., Complications of androgen deprivation therapy in men with prostate cancer. *Curr Oncol Rep* **2004**,*6* (3), 209-15.
380. Isbarn, H.; Boccon-Gibod, L.; Carroll, P. R.; Montorsi, F.; Schulman, C.; Smith, M. R.; Sternberg, C. N.; Studer, U. E., Androgen deprivation therapy for the treatment of prostate cancer: consider both benefits and risks. *Eur Urol* **2009**,*55* (1), 62-75.