BRIEF REPORT

LIVER TRANSPLANTATION FOR TYPE IV GLYCOGEN STORAGE DISEASE

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TYPE IV glycogen storage disease is a rare autosomal recessive disorder (also called Andersen's disease¹ or amylopectinosis) in which the activity of branching enzyme alpha-1, 4-glucan: alpha-1, 4-glucan 6-glucosyltransferase is deficient in the liver as well as in cultured skin fibroblasts and other tissues.^{2,3} This branching enzyme is responsible for creating branch points in the normal glycogen molecule. In the relative or absolute absence of this enzyme, an insoluble and irritating form of glycogen, an amylopectin-like polysaccharide that resembles plant starch, accumulates in the cells. The amylopectin-like form is less soluble than normal glycogen, with longer outer and inner chains and fewer branch points. The clinical onset of the disease is insidious, with nonspecific

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gastrointestinal symptoms at first, followed by progressive hepatosplenomegaly, portal hypertension, ascites, and hepatic failure. Children with this disorder usually die of hepatic cirrhosis by the age of two to four years. ⁴⁻⁸ In exceptional cases, cardiomyopathy, ^{5-7,9} neurologic syndromes — including tremors, seizures, and dementia ^{10,11} — or variable manifestations of myopathy ^{5,12,13} have been reported. In patients with these unusual symptoms, the clinical onset is frequently later than in typical cases, and death most often results from cardiac failure.

Liver transplantation for Type IV glycogen storage disease was attempted in 1972; the recipient died 110 days later after the rejection of the first liver transplant and attempted retransplantation. Liver transplantation was first performed successfully in September 1984 in Patient 1 of this series; since that time we have treated six more such patients. Our experience with these seven patients forms the basis of this report.

CASE REPORTS

All the patients were boys, including two sets of brothers (Table 1). All had progressive liver failure with massive hepatomegaly, splenomegaly, and ascites. (Because patients with Type IV glycogen storage disease but without progressive liver disease have occasionally been reported, ¹⁶ progressive liver disease was a condition for transplantation in each case.) Two of the seven patients died 7 and 36 days after liver transplantation — from a bowel perforation and thrombosis of the hepatic artery, respectively. The five other recipients (71 percent) are healthy and have normal liver function 16 to 73 months after transplantation. The five transplant recipients who survived were hospitalized for 26 to 49 days (mean, +0). The mean (±SD) cost of transplantation in these five patients, including all professional fees, was \$107,900±30,000. Rehospitalization was

not required except for follow-up percutaneous needle biopsies of the liver, performed 1 to 45 months after transplantation, or transjugular endomyocardial biopsies, performed 3 weeks to 54 months after transplantation (Table 2). The transplantations and follow-up biopsies were undertaken with informed parental consent. The biopsy procedures were considered essential for optimal care of the patients and were not part of a prospective experimental design.

Height, weight, gross and fine motor development, and language ability were retarded in all patients before transplantation. The physical strength and motor skills of the five survivors improved steadily after transplantation. All have had a normal growth rate, two are performing well in elementary school, and the three in preschool have no apparent abnormalities in intellectual development. None has had any cardiac symptoms or any evidence of abnormalities on chest films or electrocardiographic and echocardiographic examinations. A magnetic resonance imaging scan of the heart in one child was normal more than four years after transplantation.

METHODS

Liver Transplantation

The diseased liver was totally excised and replaced with a size-matched cadaveric liver, which was revascularized in an anatomically normal manner and provided biliary drainage with a Rouxen-Y choledochojejunostomy.¹⁷ Immunosuppressive therapy consisted of cyclosporine and prednisone. The daily maintenance dose of prednisone in the five surviving patients was 5 mg or less.

Tissue Studies

Cultured skin fibroblasts and homogenates of liver tissue from the patients were assayed at Washington University for branchingenzyme activity. The phosphorylase-coupled assay measured the rate of formation of inorganic phosphate from glucose-1-phosphate as glucose was polymerized to glycogen by the tissue homogenates. ^{3,6,15} The branching-enzyme activity in skin fibroblasts averaged 0.08 μ mol of inorganic phosphate per minute per milligram of protein, which was less than 10 percent of the activity in normal subjects (Table 1).

Amylopectin was sought on light-microscopical examination in the excised livers, in liver-biopsy samples obtained after transplantation, and in other tissues obtained at the time of transplantation or at autopsy. The characteristic inclusions were positive on periodic acid–Schiff staining after digestion with diastase (PAS-D). Electron-microscopical studies demonstrated the fibrillar aggregations that are typical of amylopectin. 18-20

Morphometric studies were performed on PAS-D-stained myocardial tissue obtained at autopsy from one patient and endomyocardial-biopsy samples from three others obtained at various times

Table 1. Clinical Data on Patients with Type IV Glycogen Storage Disease Who Underwent Liver Transplantation.

Patient No.*	AGE AT TRANS- PLANTATION	Skin-Fibroblast Branching-Enzyme Activity†	DATE OF OPERATION	Length of Survival	
	mo	μmol/min/mg			
1	31	0.10	9/6/84	73 mo	
2	11	0.07	8/28/85	Died 7 days after transplantation	
3	36	0.11	9/5/85	Died 36 days after transplantation	
4	46	0.11	11/25/85	58 mo	
5	22	0.08	10/24/87	35 mo	
6	20	0.06	3/21/89	18 mo	
7	34	0.09	5/26/89	16 mo	

^{*}Patients 1 and 6 were brothers, as were Patients 3 and 5

Table 2. Deposits Positive on Periodic Acid—Schiff Staining after Digestion with Diastase (PAS-D) in the Myocardium of Four Patients with Type IV Glycogen Storage Disease after Liver Transplantation.

PATIENT No.	TIME AFTER TRANSPLAN- TATION	No. of Inclusions	MEAN AREA OCCUPIED BY INCLUSIONS	MEAN SIZE OF INCLU- SIONS
	mo	per 3.6×10 ⁵ μm ²	%	μm^2
l	54.0	24	0.5	140
3	1.2 (at autopsy)	46	2	141
6	0.9 14.0	332 129	13 5.9	146 164
7	1.0 11.0	Too few to quantitate Too few to quantitate	_	

after transplantation (Table 2), with the Bioquant System IV (R&M Biometrics, Nashville) used for automated image analysis. The biopsy samples were also separately evaluated with the point-count technique. To eliminate artifacts caused by vessels or compression, interstitial and vascular spaces in the field were eliminated from the final calculations of the area of amylopectin deposition. Variation within samples was studied in 24 fields from a large piece of left ventricle obtained at autopsy from Patient 3. All the fields had much the same amylopectin distribution, with the single exception of a superficial subendocardial field that contained little muscle.

RESILTS

There was no trace of amylopectin in the graft of the child who died 36 days after transplantation from hepatic-artery thrombosis or in the liver-biopsy specimens obtained from the five surviving children 1 to 45 months after transplantation. The biopsy specimens also showed no evidence of rejection.

Amylopectin deposits were found in all biopsy specimens of skin (including arrectores pilorum muscles), jejunum, and skeletal muscle obtained at the time of transplantation. Postmortem studies in Patient 3, who died 36 days after transplantation at 36 months of age, showed amylopectin in the esophagus, bowel, bladder smooth muscle, skeletal muscle, central nervous system, peripheral nerves, and heart. In this patient, amylopectin occupied about 2 percent of the myocardial area (Table 2).

Myocardial samples were available from three other patients (Table 2). Patient 1, whose liver was replaced when he was 36 months old, had a nearly amylopectin-free myocardial-biopsy specimen 54 months later. His younger brother (Patient 6) had involvement of 13 percent of the myocardial area three weeks after transplantation (Table 2). A year later, the amount of amylopectin had been reduced to 5.9 percent (Fig. 1, Table 2). In Patient 7, the myocardial-biopsy specimen obtained one month after liver transplantation showed very few deposits in the myocytes; 10 months later, there was no change.

DISCUSSION

The absence of amylopectin deposition in the liver grafts was expected, but the freedom from neuromuscular or cardiac morbidity associated with extrahepat-

[†]Normal branching-enzyme activity in skin fibroblasts, 1 to 2 μ mol per minute per milligram of protein, 7.15

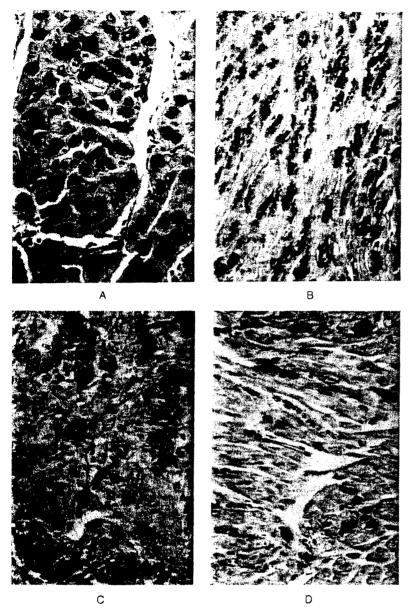


Figure 1. Myocardial Biopsy Specimens from Patient 6 (PAS-D, ×405).

Transverse sections (Panel A) and oblique sections (Panel B) of myocardial fibers in a biopsy specimen obtained 3½ weeks after liver transplantation show abundant amylopectin inclusions (red material). Panels C and D show comparable fields from a biopsy specimen obtained 14 months after transplantation, in which there is evidence of a marked decrease in the number of inclusions.

ic amylopectin deposits was more noteworthy. No neuromuscular complications were seen, and the retarded growth that was characteristic of the patients before transplantation was restored to more nearly normal values in all the survivors. More important, none of the surviving patients has had cardiac complications during follow-up periods as long as six years. Because a wide variety of cardiac and noncardiac tissues collected perioperatively from different patients contained cytoplasmic amylopectin, it must be assumed that all the patients had cardiac involvement, as was demonstrated in the four patients studied.

However, there was little amylopectin in myocardial-biopsy specimens obtained a year or more after liver transplantation. Particularly striking was the decrease in myocardial amylopectin over a period of 13 months after liver transplantation in Patient 6, whose older brother had minimal heart involvement 54 months after transplantation. We speculate that the older brother may have cleared amylopectin from his extrahepatic tissues by the time this first heart biopsy was performed. That amylopectin disappears slowly is illustrated by the cases of our patients and may account for unpublished observations in a 14-month-old Belgian child who died of a respiratory infection superimposed on congestive heart failure 11 months after liver transplantation. Amylopectin was found post mortem in this child's myocardium and in other organs (Otte IB, University of Louvain Medical School, Brussels: personal communication).

These observations could have implications for other enzyme deficiencies that affect multiple organs, particularly if cell-to-cell transfer of the deficient enzyme can be demonstrated. However, any explanation for extrahepatic amylopectin clearance must await a better understanding of the paradoxical enzymologic features of Type IV glycogen storage disease, in which a branching-enzyme defect results in glycogen with fewer than normal branches.² Furthermore, the distribution of the normal and abnormal branching enzymes may be different in various tissues.21

Finally, it is possible that longterm cyclosporine therapy was an ameliorating factor in our patients.

This drug,²² as well as the new immunosuppressive agent FK 506,²³ has hepatotrophic qualities similar to those of insulin,²⁴ including the ability to increase hepatic glycogen stores. Both these immunosuppressive drugs attach to cytosolic binding sites that are rich in the ubiquitous enzyme peptidyl-prolyl isomerase, ^{25,26} and both cause wide-ranging immunologic and nonimmunologic effects, including alterations in carbohydrate, cholesterol, and uric acid metabolism.²³ Thus, it is conceivable that the early treatment of patients with Type IV glycogen storage disease could obviate the need for transplantation.

REFERENCES

- Anderson DH. Studies on glycogen disease with report of a case in which the glycogen was abnormal. In: Najjar VA, ed. Carbohydrate metabolism. Baltimore: Johns Hopkins Press, 1952:28-42.
- Illingworth B, Cori GT. Structure of glycogens and amylopectins. III. Normal and abnormal human glycogen. J Biol Chem 1952; 199:653-60.
- Brown BI, Brown DH. Lack of an α-1,4-glucan:α-1,4-glucan 6-glycosyl transferase in a case of type IV glycogenosis. Proc Natl Acad Sci U S A 1966: 56:725-9.
- Bannayan GA, Dean WJ, Howell RR. Type IV glycogen-storage disease: light-microscopic, electron-microscopic, and enzymatic study. Am J Clin Pathol 1976; 66:702-9.
- Greene GM, Weldon DC, Ferrans VJ, et al. Juvenile polysaccharidosis with cardioskeletal myopathy. Arch Pathol Lab Med 1987; 111:977-82.
- Hers H-G, Van Hoof F, de Barsy T. Glycogen storage diseases. In: Scriver CR, Beaudet AL, Sly WS, Valle D, eds. The metabolic basis of inherited disease. 6th ed. Vol. 1. New York: McGraw-Hill, 1989:425-52.
- Brown BI. Debranching and branching enzyme deficiencies. In: Engel AG, Banker BQ, eds. Myology. New York: McGraw-Hill, 1986:1653-61.
- Ishihara T, Uchino F, Adachi H, et al. Type IV glycogenosis a study of two cases. Acta Pathol Jpn 1975; 25:613-33.
- Servidei S, Riepe R, Langston C, et al. Severe cardiopathy in branching enzyme deficiency. J Pediatr 1987; 111:51-6.
- McMaster KR, Powers JM, Hennigar GR Jr, Wohltmann HJ, Farr GH Jr. Nervous system involvement in type IV glycogenosis. Arch Pathol Lab Med 1979; 103:105-11.
- Ferguson IT, Mahon M, Cumming WJ. An adult case of Andersen's disease
 — Type IV glycogenosis: a clinical, histochemical, ultrastructural and biochemical study. J Neurol Sci 1983; 60:337-51.
- Zellweger H, Mueller S, Ionasescu V, Schochet SS, McCormick WF. Glycogenosis. IV. A new cause of infantile hypotonia. J Pediatr 1972; 80:842-4.
- Guerra AS, van Diggelen OP, Carneiro F, Tsou RM, Simoes S, Santos NT.
 A juvenile variant of glycogenosis IV (Andersen disease). Eur J Pediatr 1986; 145:179-81.

- Starzl TE, Koep LJ, Halgrimson CG, et al. Fifteen years of clinical liver transplantation. Gastroenterology 1979; 77:375-88.
- Brown BI, Brown DH. Branching enzyme activity of cultured amniocytes and chorionic villi: prenatal testing for type IV glycogen storage disease. Am J Hum Genet 1989; 44:378-81.
- Greene HL, Brown BI, McClenathan DT, Agostini RM Jr, Taylor SR. A new variant of Type IV glycogenosis: deficiency of branching enzyme activity without apparent progressive liver disease. Hepatology 1988: 8:302-6.
- Starzl TE, Demetris AJ, Van Thiel D. Liver transplantation. N Engl J Med 1989; 321:1014-22.
- Ishihara T, Yokota T, Yamashita Y, et al. Comparative study of the intracytoplasmic inclusions in Lafora disease and type IV glycogenosis by electron microscopy. Acta Pathol Jpn 1987; 37:1591-601.
- Reed GB Jr, Dixon JFP, Neustein HB, Donnell GN, Landing BH. Type IV glycogenosis: patient with absence of a branching enzyme α-1, 4-glucan: α-1, 4-glucan 6-glycosyl transferase. Lab Invest 1968; 19:546-57.
- Schochet SS Jr, McCormick WF, Zellweger H. Type IV glycogenosis (amylopectinosis): light and electron microscopic observations. Arch Pathol 1970; 90:354-63.
- Brown DH, Brown BI. Studies of the residual glycogen branching enzyme activity present in human skin fibroblasts from patients with type IV glycogen storage disease. Biochem Biophys Res Commun 1983; 111:636-43
- Mazzaferro V, Porter KA, Scotti-Foglieni CL, et al. The hepatotropic effect of cyclosporine. Surgery 1990; 107:533-9.
- Starzl TE, Porter KA, Mazzaferro V, Todo S, Fung J, Francavilla A. Hepatotrophic effects of FK506 in dogs. Transplantation (in press).
- Starzl TE, Watanabe K, Porter KA, Putnam CW. Effects of insulin, glucagon, and insulin/glucagon infusions on liver morphology and cell division after complete portacaval shunt in dogs. Lancet 1976; 1821-5.
- Siekierka JJ, Hung SHY, Poe M, Lin CS, Sigal NH. A cytosolic binding protein for the immunosuppressant FK506 has peptidyl-prolyl isomerase activity but is distinct from cyclophilin. Nature 1989; 341:755-7.
- Harding MW, Galat A, Uehling DE, Schreiber SL. A receptor for the immunosuppressant FK506 is a cis-trans peptidyl-prolyl isomerase. Nature 1989; 341:758-60.

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