

Albert B. Zajko, M.D.
Klaus M. Bron, M.D.
Thomas E. Starzl, M.D.
David H. Van Thiel, M.D.
J. Carlton Gartner, M.D.
Shunzaburo Iwatsuki, M.D.
Byers W. Shaw, Jr., M.D.
Basil J. Zitelli, M.D.
J. Jeffrey Malatack, M.D.
Andrew H. Urbach, M.D.

Over 45 months, 119 angiographic examinations were performed in 95 patients prior to liver transplantation, and 53 examinations in 44 patients after transplantation. Transplantation feasibility was influenced by patency of the portal vein and inferior vena cava. Selective arterial portography, wedged hepatic venography, and transhepatic portography were used to assess the portal vein if sonography or computed tomography was inconclusive. Major indications for angiography after transplantation included early liver failure, sepsis, unexplained elevation of liver enzyme levels, and delayed bile leakage, all of which may be due to hepatic artery thrombosis. Other indications included gastrointestinal tract bleeding, hemobilia, and evaluation of portal vein patency in patients with chronic rejection who were being considered for retransplantation. Normal radiographic features of hepatic artery and portal vein reconstruction are demonstrated. Complications diagnosed using results of angiography included hepatic artery or portal vein stenoses and thromboses and pancreaticoduodenal aneurysms. Intrahepatic arterial narrowing, attenuation, slow flow, and poor filling were seen in five patients with rejection.

Index terms: Liver, angiography, 952.122, 957.124 • Liver, transplantation, 761.45

Radiology 1985; 157:305-311

¹ From the Department of Radiology, Presbyterian-University Hospital (A.B.Z., K.M.B.), the Department of Surgery, University Health Center of Pittsburgh (T.E.S., S.I., B.W.S.), the Department of Medicine (Division of Gastroenterology), University of Pittsburgh School of Medicine (D.H.V.T.), and the Department of Pediatrics, Childrens Hospital of Pittsburgh (J.C.G., B.J.Z., J.J.M., A.H.U.), Pittsburgh, Pennsylvania. Received December 13, 1984; accepted and revision requested May 2, 1985; revision received June 4. Presented at the 70th Scientific Assembly and Annual Meeting of the Radiological Society of North America, Washington, D.C., November 25–30, 1984.

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Angiography of Liver Transplantation Patients¹

The first human liver transplantation was performed in 1963 (1). By 1966, six patients had received liver transplants at the University of Colorado Medical Center. Unfortunately, all of them soon died, with the longest survival being 23 days (2). The first extended survival (13 months) was achieved in 1967 (3). As survival rates have improved, transplantation has increasingly become an accepted mode of therapy for irreversible end-stage liver disease in both children and adults (4, 5). The number of transplantations performed each year has steadily increased (2, 4, 5). At the University Health Center of Pittsburgh, 265 patients received 338 orthotopic liver transplants from January 1981 to October 1984.

Possible transplantation candidates must first undergo medical evaluation (4–6), which may include angiography of the portal vein and/or inferior vena cava (IVC).

After liver transplantation, the occurrence of several complications requires angiographic evaluation. We report our experience with pre- and postoperative angiography in the evaluation of pediatric and adult liver transplantation patients.

SUBJECTS AND METHODS

Pretransplantation

During the 45 months from January 1981 to October 1984, 650 patients were evaluated at the University Health Center of Pittsburgh to determine the feasibility of transplantation. Ninety-five patients (14.6%) required a total of 119 angiographic studies, involving the portal vein in 84, the IVC in two, and both vessels in nine. Of the 11 patients requiring evaluation of the IVC, nine were children in whom congenital absence of the cava was suspected. To evaluate the portal vein, selective superior mesenteric, splenic, and/or hepatic arteriography was performed. If selective arterial portography was inconclusive, wedged hepatic venography was performed; three patients underwent direct transhepatic portography. In most cases, wedged hepatic venography was performed on another day so as not to exceed contrast limitations.

Posttransplantation

During the same period, 265 patients (119 children and 146 adults) received a total of 338 orthotopic liver transplants; 53 received two transplants each, and ten received three transplants each. There were 125 males and 140 females, with an age range of 4 months to 56 years.

Following the procedure, 44 patients (23 children and 21 adults) underwent 53 angiographic studies. This included three patients who underwent transplantation in Denver prior to the beginning of the Pittsburgh program. Seven patients were studied twice, and one patient was studied three times. Angiography was performed from 2 days to 11 years and 8 months after transplantation. Approximately 70% of the studies were performed within the first 2 months following transplantation. Indications for angiography are given in Table 1.

Liver homograft revascularization consisted of anastomoses of the hepatic artery, portal vein, and IVC. The hepatic artery was reconstructed in one of several ways. In most cases the end of the donor's celiac axis or common



Figure 1. Homograft arterial revascularization by direct endto-end anastomosis. Selective celiac arteriogram shows a patent anastomosis (curved arrow) between the donor's celiac axis and the recipient's common hepatic artery, the most common method of reconstruction. The ligated donor splenic (straight arrow) and gastroduodenal arteries (arrowhead) are also shown.

hepatic artery was anastomosed to the end of the recipient's common or proper hepatic artery (Fig. 1). Several patients had an iliac artery homograft obtained from the donor (Fig. 2). In several children, the donor's thoracic or abdominal aorta (together with the celiac axis) was anastomosed to the side of the recipient's abdominal aorta (Fig. 3). In a few patients whose native common hepatic artery arose from the superior mesenteric artery, the donor's celiac axis was anastomosed to the end of the common hepatic artery.

Anomalies of the donor's hepatic artery required a different type of anastomosis, depending on the anomaly (7). The most common variant was origin of an accessory right hepatic artery (or the entire right hepatic artery) from the superior mesenteric artery, coupled with origin of the left hepatic artery from the celiac axis. Vascular reconstruction in adults is illustrated in Figure 4.

The portal vein was usually revascularized by end-to-end anastomosis of the extrahepatic portal veins of the donor and recipient (Fig. 5). In a few patients with an abnormally small or occluded portal vein, a venous homograft from the donor's IVC, iliac, or pulmonary vein was interposed between the donor's extrahepatic portal vein and the confluence of the recipient's superior mesenteric and splenic veins (8) (Fig. 6).

Reconstruction of the IVC consisted of anastomoses of the infra- and suprahepatic vena caval segments of both donor and recipient.

RESULTS

Pretransplantation

The portal vein was patent in 78 of 93 patients (84%). In 12 patients, selective superior mesenteric, splenic, and hepatic angiography were inconclusive regarding portal vein patency;

however, wedged hepatic venography confirmed patency in eight (Fig. 7). Three patients underwent direct transhepatic portography, which confirmed patency in one.

Four of nine children evaluated with inferior vena cavography were found to have congenital absence of the IVC with azygos continuation to the heart. Thrombosis of the IVC was found in two adults.

Posttransplantation

Twenty-six of 53 angiograms were normal or showed no significant abnormality (Table 2). Hepatic artery thrombosis occurred in seven children and one adult 10 days to 5 weeks after transplantation. Four of these patients had the hepatic artery reconstructed by end-to-end anastomosis of the donor and recipient hepatic arteries, and four had vascular homografts (two aortic and two iliac). In one child whose hepatic artery became thrombosed 2 weeks after transplantation (Fig. 8a), a follow-up angiogram 6 weeks later showed arterial collaterals to the transplanted liver (Fig. 8b).

Hepatic artery stenosis was observed in three transplants; two occurred at the arterial anastomosis. In one patient, angiography demonstrated a stenosis, which proved at exploratory laparotomy to be a kink in the artery. The artery was straightened, but the stenosis was not believed to be hemodynamically significant. In one patient who had a follow-up angiogram 5 months later, the anastomotic stenosis had essentially resolved. In another patient with an anastomotic stenosis (Fig. 9), percutaneous balloon



Figure 2. Patent iliac artery homograft (small arrows) and hepatic artery. The proximal anastomosis is between the common iliac end of the graft and the recipient's lower abdominal aorta (curved arrow), while the distal anastomosis is from the external iliac end of the graft to the donor's common hepatic artery (straight arrow). The ligated donor internal iliac artery is also shown (arrowhead).

angioplasty reduced the pressure gradient from 40 to 20 mm Hg. However, the liver was rejected, and retransplantation was required.

Three patients had portal vein occlusion and presented with upper gastrointestinal (GI) tract bleeding. Occlusion developed after 8 days in one patient, and retransplantation was required. Angiography demonstrated massive gastroesophageal varices and no filling of intrahepatic portal vessels. In two patients, occlusion developed 1 year after transplantation. Angiographically, both patients demonstrated complete occlusion of the extrahepatic portal vein associated with massive gastroesophageal varices, but the intrahepatic portal veins were reconstituted via numerous periportal collaterals (Fig. 10).

Portal vein stenosis occurred in three children and one adult at 2, 4, 20, and 32 weeks after transplantation, respectively (Fig. 11). All presented with upper GI tract bleeding.

In one patient who was evaluated for hemobilia 10 weeks after trans-

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Figure 3. Homograft arterial revascularization using the donor's aorta. The end of the thoracic or (as in this case) abdominal aorta (straight arrow), together with the celiac axis, is anastomosed to the side of the recipient's lower abdominal aorta (curved arrow).

Table 1
Indications for Angiography
Following Liver Transplantation

	Children	Adults
Possible hepatic	4.5	
artery thrombosis	17	13
Possible portal		Barran Adil
vein thrombosis	11.3 Tells	1
GI tract bleeding	6	3.
Hemobilia	2.79	3
Chronic rejection;		S. Alba
need to evaluate	un allengage	
the portal vein for		
retransplantation	2	3
4	- 20	72
Total		or had a

plantation, multiple aneurysms were observed in pancreaticoduodenal collateral vessels (Fig. 12). At laparotomy, the aneurysms were found to be mycotic with erosion into the common bile duct.

One patient with pathologically proved chronic rejection was evaluated for retransplantation after 8 months. Diffuse and focal areas of narrowing were present within the hepatic arterial tree (Fig. 13). Four other patients with clinical evidence of rejection demonstrated slowing of flow, poor peripheral filling, and attenuation in the size and number of arterial branches within the liver.

Table 2
Angiographic Findings Following
Liver Transplantation

789.432.56	∵.Chi	ldren - Ad	ults
Normal	11956 BBA 1	14 81	2*
Hepatic arter		7	1
Hepatic arter	'y	0	ੋਂ: 4 ਂ
Portal vein		. 4	0
Portal vein s	tenosis	3	Ĭ.
aneurysms Intrahepatic		0	1
narrowing	, alow 🧢 🦈		en Turkens
flow, atten	8	-1 🔻	4
Arterial colla	r after		
hepatic art		1 ***	0.
	romerador distribution from	3.20	7 Sale (447)

Reconstitution of the intrahepatic portal vessels via periportal venous collaterals (cavernomatous transformation) occurred in two patients (Fig. 10).

DISCUSSION

The increased success rate of liver transplantation over the past 20 years is due to many factors. Among the most important are improved surgical technique, changes in patient selection, use of the immunosuppressive drug cyclosporine, and better postoperative care (9, 10).

Each patient with an acceptable indication for liver transplantation undergoes a preoperative medical evaluation, which generally requires a 5-7day hospitalization. The portal vein is examined by sonography and/or dynamic computed tomography (CT) to determine patency. It is considered normal on sonography if the walls are smooth and there are no internal echoes. If the portal vein cannot be seen because of overlying gas or altered venous anatomy as the result of previous surgery, if it is smaller than 4 mm, or if there are internal portal vein echoes suggesting thrombus, angiography should be performed. Angiography is also indicated if there is evidence of portal vein occlusion on dynamic CT. Doppler ultrasound, which will be available in the near future, should be invaluable in examining portal vein blood flow prior to transplantation.

In most patients, selective superior mesenteric or splenic arteriography with venous-phase filming will demonstrate the portal vein. However, many of these patients have severe portal hypertension with hepatofugal portal venous flow. In some, the portal vein may not be visualized after superior mesenteric or splenic arteriography. Occasionally, selective he-

patic arteriography will show the portal vein in patients with complete reversal of flow in the portal vein. If selective arterial studies are inconclusive, wedged hepatic venography should be performed (Fig. 7). Only three of our patients required direct transhepatic portography.

Until recently, patency of the portal vein was a prerequisite for liver transplantation (6) because of the type of end-to-end anastomosis performed for portal venous revascularization (Fig. 5). With improved surgical techniques, however, transplantation is still feasible even if the recipient's portal vein is thrombosed or abnormally small in diameter (8). If it is less than 3 mm or appears occluded on angiography, the confluence between the superior mesenteric and splenic veins is evaluated. If the junction between these two veins is patent and of reasonable size, the donor's portal vein can be anastomosed to it using an interposed vascular homograft (Fig. 6). At the time of donor hepatectomy, segments of the aorta, IVC, and iliac arteries and veins are removed; then if a vascular graft is needed during transplantation, these homograft vessels are readily available for hepatic artery, portal vein, or IVC reconstruction (11).

The other major indication for pretransplantation angiography is evaluation of children with suspected congenital absence of the IVC. The major indication for transplantation in the pediatric age group is biliary atresia (4), which is associated with a higher frequency of congenital absence of the intrahepatic IVC, as well as other vascular and visceral anomalies (12). Although such absence is not an absolute contraindication to transplantation, when combined with other hepatic vascular anomalies it represents a high-risk group of patients who should be identified prior to surgery.

Vascular anomalies in these children include aberrant origin of the hepatic artery and preduodenal portal vein, while visceral anomalies include situs inversus, polysplenia, and GI tract malrotation (12). Prior to transplantation, most patients with biliary atresia have undergone a portoenterostomy (Kasai procedure) for biliary drainage (13), which frequently reveals visceral or vascular anomalies. In addition, an upper GI tract contrast study will have been performed to exclude malrotation. To identify this high risk group of patients, therefore, all patients with biliary atresia and GI tract malrotation or a visceral or vascular anomaly undergo inferior vena cavography to evaluate for possible congenital absence of the IVC.

At our center, the procedure of choice for hepatic artery reconstruction in patients with a normal native hepatic artery of sufficient size is endto-end anastomosis of the donor's celiac axis to the recipient's common hepatic artery (14) (Fig. 1), which assumes a single arterial supply to the donor liver. Other types of anastomoses depend on the vascular anatomy of the donor and recipient (7). In children with very small hepatic arteries in whom a direct hepatic artery anastomosis may be difficult, the donor's thoracic or abdominal aorta, in continuity with the celiac axis, is anastomosed to the recipient's abdominal aorta (Fig. 3). Usually the abdominal aorta is preferred, but it is frequently unavailable because the kidneys and part of the abdominal aorta are harvested for renal transplantation at the time the donor's liver is removed. In children given a liver with a dual blood supply (e.g., left hepatic artery from the celiac axis and right hepatic artery from the superior mesenteric artery), the thoracic or abdominal aorta (in continuity with both vessels) has been used for arterial revascularization. In adults, however, an anastomosis as shown in Figure 4 is usually employed. An iliac artery homograft may be used when

the recipient's hepatic artery is small (or absent), occluded, has poor flow, or if subintimal dissection develops during vascular mobilization (Fig. 2). An iliac homograft may also be placed between the donor and recipient hepatic arteries when they are too short to permit a direct end-to-end anastomosis.

The main reason for posttransplant angiography is suspected hepatic artery thrombosis, which is a major indication for early retransplantation (15). Clinical findings include early hepatic failure, sepsis, unexplained elevation of liver enzyme levels, and delayed bile leakage. Such leakage may be secondary to necrosis of the distal donor bile duct, and revision of the biliary anastomosis is necessary. However, major bile leakage has been associated with complete necrosis of the donor bile duct secondary to hepatic artery thrombosis. In these patients retransplantation, rather than biliary reconstruction, is always necessary (16).

Postoperatively, CT or ultrasound may be helpful in patients with sepsis or fever if there is a clinical suspicion of an abdominal abscess. Patients with focal alterations in the liver parenchyma (infarct or abscess) with these noninvasive imaging tests have an 85% incidence of hepatic artery

thrombosis (17).

While hepatic artery occlusion may be tolerated by non-liver-transplantation patients, following transplantation it can result in hepatic gangrene or even death because of the lack of potential for arterial collateralization to the liver. With proximal hepatic artery occlusion in a nontransplant liver, collaterals develop rapidly, preventing ischemia and infarction (18-20). Because of this, the survival of the liver probably cannot be solely attributable to a patent portal venous system (21). Since all potential collateral pathways are severed at the time of transplantation, occlusion of the hepatic artery after transplantation completely dearterializes the liver, possibly resulting in severe or even fatal hepatic injury.

Once thrombosis is diagnosed, retransplantation is performed as soon as a donor liver is available. Most of our patients died if a new liver was not immediately available. However, three patients did survive, and in one of them, a follow-up angiogram 6 weeks later showed arterial collaterals supplying the liver (Fig. 8). This patient is currently alive after 1 year. The two additional patients surviving following hepatic artery thrombosis did not have repeat angiography, and we presume that they also have collaterals.

In one patient with hepatic artery thrombosis and liver failure, a donor liver was not available, and low-dose

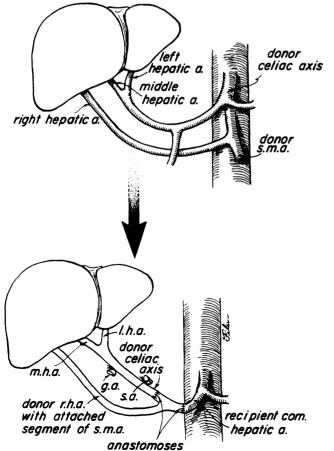




Figure 4. Arterial revascularization of a liver homograft with a dual blood supply. (a) The donor's liver is supplied by both the celiac axis (via the left and middle hepatic arteries) and the superior mesenteric artery (s.m.a.) (via the right hepatic artery). At the time of hepatectomy a small segment of the s.m.a. is removed together with the right hepatic artery. The proximal end of the s.m.a. is anastomosed to the end of the recipient's common hepatic artery, and the distal end is anastomosed to the end of the donor's celiac axis. r.h.a. = right hepatic artery, m.h.a. = middle hepatic artery, l.h.a. = left hepatic artery, g.a. = gastroduodenal artery, s.a. = splenic artery. (b) Selective celiac arteriogram demonstrated patent proximal (curved arrow) and distal anastomoses (straight arrow) to the donor's superior mesenteric artery segment (arrowheads).

streptokinase was infused into the occluded hepatic artery. After several hours, GI tract bleeding occurred, and streptokinase was discontinued. However, the patient continued to bleed and died. At autopsy, there was diffuse hemorrhage into the GI tract but no focal bleeding points. Streptokinase is not routinely used after hepatic artery thrombosis because liver infarction is usually present once the diagnosis has been established. Infection often supervenes in these immunosuppressed patients and may



Figure 5. Portal vein reconstruction. Venous phase of the selective superior mesenteric arteriogram reveals a widely patent end-to-end anastomosis (curved arrow) between the donor and recipient extrahepatic portal veins. Note the minimal "waist" at the anastomotic line.

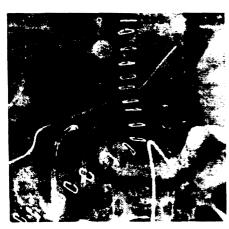


Figure 6. Portal vein reconstruction using a venous homograft. The graft (small arrows) is anastomosed between the donor's extrahepatic portal vein (curved arrow) and the confluence of the recipient's superior mesenteric and splenic veins (straight arrow) as demonstrated on the venous phase of the selective splenic arteriogram. A small segment of the donor's IVC was used for the

lead to septic hepatic gangrene. In such cases, emergency retransplantation is the only hope for survival (15). In addition, these patients have recently undergone surgery and are therefore at a higher risk of bleeding as a result of streptokinase therapy.

Other indications for angiography following transplantation include GI tract bleeding, hemobilia, and evaluation of portal vein patients with chronic rejection who are being considered for retransplantation. In patients with GI tract bleeding in whom endoscopy demonstrates varices, angiography of the portal venous system is indicated. Several of our patients demonstrated either stenosis or occlusion of the portal vein. One patient with stenosis underwent successful revision of the portal vein anastomosis; another patient who had occlusion required retransplantation. Two patients demonstrated typical cavernomatous transformation of the portal vein (Fig. 10). The mechanism by which these venous collaterals develop is unknown, since all potential collaterals are severed during the operation. One of these patients who had persistent upper GI tract bleeding that failed to respond to endoscopic esophageal sclerotherapy was treated with a distal splenorenal shunt for control of portal hypertension. The other patient is being treated by means of endoscopic sclerotherapy.

In the three patients with hemobilia, angiography was performed to evaluate for possible pseudoaneur-vsm at the arterial anastomosis. In

none of these patients was a false aneurysm found. In one case, however, the donor's hepatic artery was anastomosed to the recipient's proper hepatic artery, and the liver was supplied by enlarged pancreaticoduodenal collaterals from the superior mesenteric artery, which had presumably developed in response to stenosis or occlusion of the native celiac axis. In addition, these collateral vessels demonstrated multiple aneurysms (Fig. 12). At exploratory laparotomy, these aneurysms were found to be mycotic in origin and to have eroded into the commom bile duct. These aneurysms were resected, and an iliac artery homograft was placed. Seven days later, the patient died after massive GI tract and intraabdominal bleeding developed due to complete septic disruption of the iliac artery homograft.

The diagnosis of rejection is generally made by exclusion. The patient may present with elevated liver enzyme and serum bilirubin levels; other causes include biliary obstruction, ischemic damage, viral infection, and hepatic artery thrombosis (22). Biliary obstruction can be excluded by cholangiography, and vascular thrombosis can be ruled out by angiography. In our series, several patients with clinical rejection demonstrated slowing of flow, poor peripheral filling, and attenuation in the size and number of the intrahepatic arteries. Analogous findings have been observed in the biliary tree after transplantation (16). In one patient with biopsy-



Figure 7. Patent portal vein (arrows) demonstrated by wedged hepatic venography. The coronary vein is also shown (arrowhead).

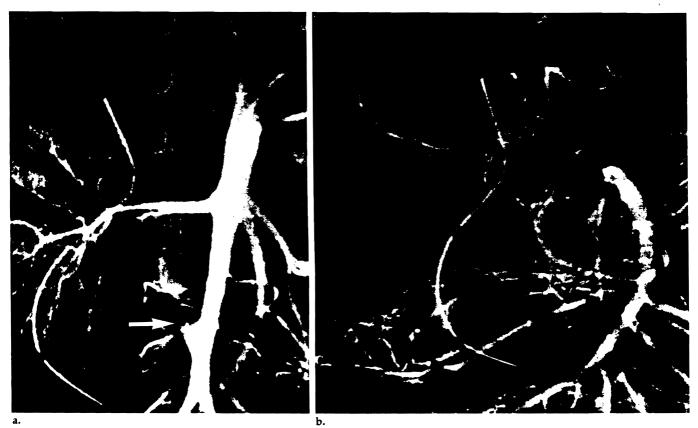


Figure 8. Collateral development following hepatic artery thrombosis. (a) Thrombosis of the iliac artery homograft (arrow) is seen on an abdominal aortogram obtained 2 weeks after transplantation. (b) Six weeks later, a selective superior mesenteric arteriogram demonstrates intrahepatic arterial reconstitution (arrows) via collaterals (arrowheads).



Figure 9. Anastomotic stenosis (arrow) demonstrated by selective hepatic arteriography 10 days after transplantation. A pressure gradient of 40 mm Hg was recorded across the stenosis.

proved chronic rejection, severe diffuse and focal arterial narrowing was demonstrated (Fig. 13). Pathologically, these arterial lesions are due to deposition of subintimal foam cells, intimal sclerosis, and myointimal hyperplasia (23). The end result is severe narrowing or occlusion of the vessels (23). The radiographic-pathologic correlation of rejection will be the subject of a future report.

Acknowledgments: We thank Donna Scahill for manuscript preparation and Ron Filer for artwork.

Send correspondence and reprint requests to: Albert B. Zajko, M.D., Department of Radiology, Presbyterian-University Hospital, DeSoto at O'-Hara St., Pittsburgh, Pennsylvania 15213.

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Figure 10. Collateral development following portal vein thrombosis. Venous phase of the selective superior mesenteric arteriogram I year after transplantation reveals occlusion of the extrahepatic portal vein and reconstitution of the intrahepatic portal vessels (straight arrows) by massive periportal collaterals (arrowheads). Marked gastric varices are also present (curved arrow).



Figure 11. Marked portal vein narrowing (arrows) demonstrated on venous phase of a selective superior mesenteric arteriogram. Gastric and esophageal varices are also present

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Figure 12. Mycotic aneurysms (straight arrows) in pancreaticoduodenal arterial collaterals demonstrated on a selective superior mesenteric arteriogram. The hepatic (arrowhead) and splenic (curved arrow) arteries are also filled.

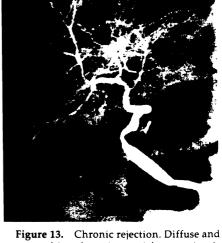


Figure 13. Chronic rejection. Diffuse and segmental intrahepatic arterial narrowing is demonstrated on a selective hepatic arteriogram taken 8 months after transplantation.

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Book Review.

Diagnostic Ultrasound: Principles, Instrumentation, and Exercises. 2d ed. Frederick W. Kremkau, Ph.D.

Orlando, Fla: Grune & Stratton, 1984. Cloth, \$29.50; pp. 278, with 118 figures and tables.

In the second edition of this already highly praised book, the author has kept his original purpose: to provide a comprehensive explanation of the basic physical principles and instrumentation of diagnostic ultrasound (US) to individuals with or without knowledge of advanced physics or mathematics. Dr. Kremkau is a well-recognized authority in the field of diagnostic US imaging, and his outstanding teaching abilities are evident in both editions of this book. The material is presented in a meticulously organized manner and is intermingled with simple drawings, graphs, charts, and tables. All equations are presented in both verbal and symbolic forms. Furthermore, a summary, numerous exercises, and a few crossword puzzles are presented at the end of each chapter for self assessment. Answers are provided at the end of the book, together with an extensive glossary and four appendices summarizing the mathematical principles and physical concepts involved and listing the necessary symbols and equations.

The text is divided into 12 small chapters, each of which builds nicely on the one preceding. Initially, the author explains the basic concepts of US, its reflection and scattering, transducers, and sound beams. Subsequently, he describes the imaging instrumentation, including static, dynamic, and Doppler instruments. The final four chapters include useful information about artifacts, performance measurement, bioeffects, misconceptions, and errors.

The second edition contains major

new and expanded material on digital instrumentation, real-time and Doppler imaging, performance measurements, and artifacts and misconceptions. These topics are all welcome additions, and each is superbly explained. It appears to us that the author has purposely avoided including any clinical images. Although the inclusion of these images would have enhanced many of the book's points, especially in the chapter on artifacts, the author reaches his overall goal nonetheless.

The publication is of excellent quality, with comfortable margins for notes and easy reading. In addition, its cost is reasonable.

It is impossible to work with diagnostic US imaging without some understanding of the basic principles and instrumentation. This book presents this knowledge in a comprehensive and easily understandable form. For those who teach the use of US imaging in a clinical setting, the book contains teaching tips and can serve as an excellent reference. To the student, it is an extremely valuable publication, not only for learning how diagnostic US works but also for preparing for the physics portion of the Registry or Board examinations.

Diagnostic Ultrasound is a classic in its field and should be available for use in all radiology departments and US laboratories.

Reviewed by Vassilios Raptopoulos, M.D., and Andrew Karellas, Ph.D.