
**SECOND VIENNA
SHOCK FORUM**

z 77 359 / 308

Vienna Shock Forum Series

Series Editors: Günther Schlag and
Heinz Redl

First Vienna Shock Forum

**Part A: Pathophysiological Role of Mediators and
Mediator Inhibitors in Shock**

First Vienna Shock Forum

Part B: Monitoring and Treatment of Shock

Second Vienna Shock Forum

SECOND VIENNA SHOCK FORUM

Proceedings of the Second Vienna Shock Forum held May 12–14, 1988

Editors

Günther Schlag

Heinz Redl

Ludwig Boltzmann Institute
for Experimental Traumatology
Vienna, Austria

42

ALAN R. LISS, INC. • NEW YORK

**Address all Inquiries to the Publisher
Alan R. Liss, Inc., 41 East 11th Street, New York, NY 10003**

Copyright © 1989 Alan R. Liss, Inc.

Printed in the United States of America

Under the conditions stated below the owner of copyright for this book hereby grants permission to users to make photocopy reproductions of any part or all of its contents for personal or internal organizational use, or for personal or internal use of specific clients. This consent is given on the condition that the copier pay the stated per-copy fee through the Copyright Clearance Center, Incorporated, 27 Congress Street, Salem, MA 01970, as listed in the most current issue of "Permissions to Photocopy" (Publisher's Fee List, distributed by CCC, Inc.), for copying beyond that permitted by sections 107 or 108 of the US Copyright Law. This consent does not extend to other kinds of copying, such as copying for general distribution, for advertising or promotional purposes, for creating new collective works, or for resale.

The publication of this volume was facilitated by the authors and editors who submitted the text in a form suitable for direct reproduction without subsequent editing or proofreading by the publisher.

Library of Congress Cataloging-in-Publication Data

Vienna Shock Forum (2nd : 1988)
Second Vienna Shock Forum.

(Progress in clinical and biological research ; v. 308)

Includes bibliographies and index.

1. Shock--Congresses. 2. Shock--Pathophysiology--
Congresses. I. Schlag, Günther. II. Redl, Heinz.
III. Title. IV. Series. [DNLM: 1. Monitoring,
Physiologic--congresses. 2. Shock--physiopathology--
congresses. 3. Shock--therapy--congresses.]

W1 PR668E v.308 / QZ 140 V662s 1988]

RB150.S5V54 1988

616'.047

89-2597

ISBN 0-8451-5158-4

**Bayerische
Staatsbibliothek
München**

Contents

Contributors	xix
Preface	
Günther Schlag and Heinz Redl	xli
1. ORGAN FAILURE/MEDIATORS	
1.1. Acute Respiratory Failure	
Lung in Shock—Posttraumatic Lung Failure (Organ Failure)—MOFS	
Günther Schlag and Heinz Redl	3
Adult Respiratory Distress Syndrome. Pathophysiology and Inflammatory Mediators in Bronchoalveolar Lavage	
Jan Modig	17
Morphologic Features of the Lung in the Respiratory Failure Associated With Hypovolemic and Septic Shock	
James C. Hogg	27
Pulmonary Fat Embolism—An Epiphenomenon of Shock or a Proper Mediator Mechanism?	
Ulrich Bosch, Susanne Reisser, Gerd Regel, Gisela Windus, Werner J. Kleemann, and Michael L. Nerlich	37
The Role of C3a in Pulmonary Alveoli Following Trauma	
Udo Obertacke, Theo Joka, Gertrud Zilow, Michael Kirschfink, and Klaus-Peter Schmit-Neuerburg	43
Cytological Changes in Alveolar Cells With ARDS	
Theo Joka, Udo Obertacke, Z. Atay, E. Kreuzfelder, J. Kalotai, and L. Olivier	51
Septic Adult Respiratory Distress Syndrome and Multiple System Organ Failure	
Jesus Villar, Miguel A. Blazquez, Santiago Lubillo, and Jose L. Manzano	57
Septic Shock and Acute Respiratory Failure	
Jesus Villar, Miguel A. Blazquez, Santiago Lubillo, Jose Quintana, and Jose L. Manzano	61

***Escherichia coli* Hemolysin Causes Thromboxane-Mediated Hypertension and Vascular Leakage in Rabbit Lungs**
Werner Seeger, Henrik Walter, Heinz Neuhof, Norbert Suttorp, and
Sucharit Bhakdi 67

Leukocyte Induced Pulmonary Damage Using Intraperitoneal Zymosan
Osvaldo Chiara, Pier P. Giomarelli, Emma Borrelli, Sandra Betti, Pietro Padalino,
and Angelo Nespoli 73

1.2. Endothelial Cells as Target Organ (in Shock)

Reaction of Vascular Intima to Endotoxin Shock
Nikolaus Freudenberg 77

Endotoxin-Induced Pulmonary Endothelial Injury
Barbara Meyrick, J.E. Johnson, and K.L. Brigham 91

Thrombin-Induced Neutrophil Adhesion
Peter J. Del Vecchio and Asrar B. Malik 101

Cellular Interactions in Sepsis Induced Organ Failure
G. Zeck-Kapp, U.N. Riede, and N. Freudenberg 113

Effects of Bacterial Exo- and Endotoxins on Endothelial Arachidonate Metabolism
Norbert Suttorp, Werner Seeger, and Heinz Neuhof 119

Effects of Bacterial Toxins and Calcium-Ionophores on Endothelial Permeability In Vitro
Norbert Suttorp, Thomas Hessz, Thomas Fuchs, Werner Seeger,
Detlev Drenckhahn, and Heinz Neuhof 127

1.3. Microcirculation

Tissue Oxygen Debt as a Determinant of Postoperative Organ Failure
William C. Shoemaker, Paul L. Appel, and Harry B. Kram 133

Is Skeletal Muscle PO₂ Related to the Severity of Multiple Organ Failure and Survival in Critically Ill Patients?
Gerard I.J.M. Beerthuizen, R. Jan A. Goris, and Ferdinand J.A. Kreuzer 137

Phase-Related Vascular Reactivity in Hemorrhagic Shock
Hermann August Henrich, Franz Bäumer, and Rolf Edgar Silber 143

Ultrastructural Study of the Gastric Mucosa After Septic Shock in the Rat
Katerina Kotzampassi, Efthimios Eleftheriadis, Athanasia Alvanou,
Emmanouel Tzartinoglou, Chryssi Foroglou, and Homeros Aletras 151

Do Endotoxemia and Sepsis Impair the Regulatory Functions of Capillary Endothelial Cells?
Anders Gidlöf and David H. Lewis 157

Peripheral Circulation in Septic Shock	
L.G. Thijs, C.E. Hack, J.H. Nuyens, and A.B.J. Groeneveld	163
Pulmonary Pressure-Flow Relationship and Peripheral Oxygen Supply in ARDS Due to Bacterial Sepsis	
Thomas Kloess, Ulrich Birkenhauer, and Bernd Kottler	175
The Relationship Between Oxygen Supply and Oxygen Uptake in Septic Shock: The Possible Role of Endotoxin	
D. De Backer, A. Roman, and J.L. Vincent	181
Pulmonary Venous Hemodynamics and Disturbances of Gas Exchange During E.Coli Bacteremia in the Goettingen Miniature Pig	
Reinhold Fretschner, Thomas Kloess, Heinz Guggenberger, and Bernd Wagener	185
1.4. Cardiovascular System	
Cardiovascular Dysfunction in Human Septic Shock	
Joseph E. Parrillo	191
Cardiopulmonary Response to Endotoxin and the Eicosanoids	
Daniel L. Traber, David N. Herndon, and Lillian D. Traber	201
Cardiac Function Changes Monitored by Radionuclide Ventriculography in the Septic Shock Baboon Model	
I.C. Dormehl, J.P. Pretorius, R.D. Burow, M.F. Wilson, J. Kilian, M. Maree, N. Hugo, and R. de Winter	207
The Influence of Tachycardia During Shock on Changes in Cardiac Volumes	
Jan P. Pretorius, I.C. Dormehl, J.G. Kilian, G. Beverley, M. Maree, N. Hugo, G. Vermaak, and M.F. Wilson	217
Isolated Rabbit Heart Preparation to Evaluate the Inotropic Effect of Endotoxin	
Peter E. Krösl, Zafar Khakpour, Martin Thurnher, Seth W.O. Hallström, and Heinrich M. Schima	225
Negative Inotropic and Cardiovascular Effects of a Low Molecular Plasma Fraction in Prolonged Canine Hypovolemic Traumatic Shock—Papillary Muscle and Isolated Heart Preparation	
Seth Hallström, Christa Vogl, Zafar Khakpour, Martin Thurnher, Peter Krösl, Heinz Redl, and Günther Schlag	231
Evaluation of Heart Performance During Septic Shock in Sheep	
Josef Newald, Kazuro Sugi, Christa Vogl, Peter Krösl, Daniel L. Traber, and Günther Schlag	237
The Cultured Rat Heart Cell: A Model to Study Direct Cardiotoxic Effects of Pseudomonas Endo- and Exotoxins	
Karl Werdan, S.M. Melnitzki, G. Pilz, and T. Kapsner	247

Chemical Characterization of a Positive Inotropic Plasma Factor in Shock Imre Szabó, Botond Penke, József Kaszaki, and Sándor Nagy	253
Pathophysiological Correlates of Cardiac Overperformance in Sepsis and Septic Shock Carlo Chiarla, Ivo Giovannini, Giuseppe Boldrini, and Marco Castagneto	259
 1.5. Mediators Complement System	
Anaphylatoxin Generation and Multisystem Organ Failure in Acute Pancreatitis Lennart Roxvall, Anders Bengtson, and Mats Heideman	265
Is Activated C3 a Premier Factor of DIC Development in Septic Shock? Qixia Wu, Zhenyuan Liu, Ying Dang, Li Chen, and Huacui Chen	271
Complement Activation and Endotoxin in Sepsis P. Padalino, M. Gardinali, J. Pallavicini, O. Chiara, G. Bisiani, and A. Nespoli	277
In-situ Complement Activation, Pulmonary Hypertension, and Vascular Leakage in Rabbit Lungs—the Role of the Terminal Complement Complex Werner Seeger, Ruth Hartmann, Heinz Neuhofer, and Sucharit Bhakdi	283
The Role of the Complement System in the Pathogenesis of Multiple Organ Failure in Shock T. Zimmermann, Z. Laszik, S. Nagy, J. Kaszaki, and F. Joo	291
Quantitation of C3a by Elisa Using a Monoclonal Antibody to a Neoantigenic C3a Determinant Gertrud Zilow, Werner Naser, Arno Friedlein, Andrea Bader, and Reinhard Burger	299
 1.5.1. Granulocytes, Proteinases, Oxygen-Radicals	
Proteases as Mediators of Pulmonary Vascular Permeability H. Neuhofer, Ch. Hoffmann, W. Seeger, N. Suttorp, and H. Fritz	305
Role of Endotoxin and Proteases in Multiple Organ Failure (MOF) Ansgar O. Aasen, Anne-Lise Rishovd, and Jan O. Stadaas	315
Neutrophil Stimulation by PMA Increases Alveolar Permeability in Rabbits Hilmar Burchardi, Notker Graf, Hartmut Volkmann, and Heribert Luig	323
Changes of Ceruloplasmin Activity in Patients With Multiple Organ Failure Reiner Dauberschmidt, Heinz Mrochen, Barbara Griess, Karin Kaden, Christel Dressler, Hans Grajetzki, and Manfred Meyer	331
Chemiluminescence-Inducing Radicals in Experimental Porcine Septic Shock Lung Hubert Reichle, Ulrich Pfeiffer, Peter Wendt, and Günther Blümel	339

Lipidperoxidation in a Canine Model of Hypovolemic-Traumatic Shock Camille Lieners, Heinz Redl, Helmut Molnar, Walter Fürst, Seth Hallström, and Günther Schlag	345
Detection of 4-Hydroxy-Nonenal, a Mediator of Traumatic Inflammation, in a Patient With Surgical Trauma and in the Sephadex Inflammation Model Mohie Sharaf El Din, Günter Dussing, Gerd Egger, Herwig P. Hofer, Rudolf J. Schaur, and Erwin Schauenstein	351
1.5.2. Endotoxin	
Mediators of Acute Lung Injury in Endotoxaemia J.R. Parrat, N. Pacitti, and I.W. Rodger	357
The Overwhelming Inflammatory Response and the Role of Endotoxin in Early Sepsis Ulrich Schoeffel, Martin Lausen, Günther Ruf, Bernd-Ulrich von Specht, and Nikolaus Freudenberg	371
The Effect of Mucosal Integrity and Mesenteric Blood Flow on Enteric Translocation of Microorganisms in Cutaneous Thermal Injury David N. Herndon, Stephen E. Morris, J. Allen Coffey, Jr., Rusty A. Milhoan, Daniel L. Traber, and Courtney M. Townsend.	377
Endogenous Fibrinolysis in Septic Patients Reinhard Voss, Gerhard Borkowski, Daniela Reitz, Heinrich Ditter, and F. Reinhard Matthias	383
Hemodynamic and Proteolytic Responses in Relation to Plasma Endotoxin Concentrations in Porcine Endotoxemia Frode Naess, Olav Røise, Johan Pillgram-Larsen, Tom E. Ruud, Jan O. Stadaas, and Ansgar O. Aasen	389
Functional Determination of tPA, PAI, and Fibrinogen in Endotoxin Shock of the Pig M. Spannagl, H. Hoffmann, M. Siebeck, H. Fritz, and W. Schramm	395
Studies on Interactions of Endotoxin With Factors of the Contact System of Plasma Olav Røise, Bonno N. Bouma, Jan O. Stadaas, and Ansgar O. Aasen	401
Dose Related Effect of Endotoxin on the Reticulo Endothelial System (RES), the Sinusoidal Cells in the Liver, and on Hepatocytes From Rats M.R. Karim, N. Freudenberg, M.A. Freudenberg, and C. Galanos	407
The Trigger for Posttraumatic Multiple Organ Failure: Surgical Sepsis or Inflammation? M.L. Nerlich	413
Endotoxin Does Not Play a Key Role in the Pathogenesis of Multiple Organ Failure. An Experimental Study Ignas P.T. van Bebber, Ron G.H. Speekenbrink, Paul H. M. Schillings, and R. Jan A. Goris	419

1.5.3. Platelet Activating Factor (PAF)

The Potential Role of Platelet-Activating Factor (PAF) in Shock, Sepsis, and Adult Respiratory Distress Syndrome (ARDS)
 Pierre Braquet and David Hosford 425

The Role of Platelet-Activating Factor (PAF) in Immune and Cytotoxic Processes
 Jean Michel Mencia-Huerta, Bernadette Pignol, Monique Paubert-Braquet, and Pierre Braquet 441

Effect of Platelet-Activating Factor (PAF) Administration in Chronically Instrumented Sheep—Analysis of PAF in Plasma
 Harald Gasser, Anna Schiesser, Heinz Redl, Martin Thurnher, Christa Vogl, Eva Paul, Sabine Krautschneider, and Günther Schlag 447

Modulation of Resynthesis of 1-Alkyl-2-Arachidonyl-Glycero-3-Phosphocholine and Phosphatidylinositols for Interception In Vivo of Free Arachidonic Acid, Lyso-PAF, Diacyl-Glycerols, and Phosphoinositides
 J.A. Bauer, K. Wurster, P. Conzen, and H. Fritz 455

1.5.4. Tumor Necrotizing Factor (TNF)

The Role of Tumor Necrosis Factor/Cachectin in Septic Shock
 Joop M.H. Debets, Wim A. Buurman, and Cees J. van der Linden 463

TNF-Induced Organ Changes in a Chronic Ovine Model—Possible Role of Leukocytes
 Heinz Redl, Günther Schlag, Camille Lieners, Eva Paul, Anna Schiesser, Herbert Lamche, Walter Aulitzky, and Christoph Huber 467

The Involvement of Platelet-Activating Factor(PAF)-Induced Monocyte Activation and Tumor Necrosis Factor (TNF) Production in Shock
 B. Bonavida, M.Paubert-Braquet, D. Hosford, and P. Braquet 485

1.6. Trauma(Sepsis)-Induced Changes of the Immune System

Graduation of Immunosuppression After Surgery or Severe Trauma
 Michael W. Holch, Peter J. Grob, Walter Fierz, Werner Glinz, and Stephanos Geroulanos 491

Mediators and the Trauma Induced Cascade of Immunologic Defects
 Eugen Faist, Wolfgang Ertel, Angelika Mewes, Theo Strasser, Alfred Walz, and Sefik Alkan 495

Early Deterioration of the Immune System Following Multiple Trauma
 Mohammad Maghsudi, Michael L. Nerlich, Johannes A. Sturm, Michael Holch, Jochen W. Seidel, and Uwe Schmuckall 507

Monocyte Dependent Suppression of Immunoglobulin Synthesis in Patients With Major Trauma
 Wolfgang Ertel and Eugen Faist 513

The T Lymphocyte-Mediated Immune Reaction in Polytrauma Matthias Cebulla, Peter Kühnl, Knut Frederking, Peter Konold, and Alfred Pannike	517
Serum Mediated Depression of Chemiluminescence Response of Granulocytes in Hemorrhagic Shock Volker Bühren, Oliver Gonschorek, Günther Sutter, and Otmar Trentz	523
Breakdown of C3 Complement and IgG in Peritonitis Exudate— Pathophysiological Aspects and Therapeutic Approach A. Billing, H. Kortmann, D. Fröhlich, and M. Jochum	527
1.7. Metabolic Disorders	
Abnormal Metabolic Control in the Septic Multiple Organ Failure Syndrome: Pharmacotherapy for Altered Fuel Control Mechanisms John H. Siegel, Thomas C. Vary, Avraham Rivkind, Ron Bilik, Bill Coleman, Ben E. Tall, and J. Glenn Morris	535
Alterations in the Metabolic Control of Carbohydrates in Sepsis John J. Spitzer, Gregory J. Bagby, Diane M. Hargrove, Charles H. Lang, and Károly Mészáros	545
Hepatic Dysfunction in Multiple Systems Organ Failure as a Manifestation of Altered Cell-Cell Interaction Frank B. Cerra, Michael West, Timothy R. Billiar, Ralph T. Holman, and Richard Simmons	563
Modification of Protein Kinase C (PKC) Activity and Diacylglycerol (DAG) Accumulation in Hepatocytes in Continuous Endotoxemia Judy A. Spitzer, I.V. Deaciuc, E.B. Rodriguez de Turco, B.L. Roth, J.B. Hermiller, and J.P. Mehegan	575
Influence of Sepsis on Perfused Rat Liver Metabolism E. Kovats, J. Karner, A. Simmel, J. Funovics, and E. Roth	589
Changes of Serum Amino Acid Concentrations in Experimentally Induced Endotoxic Shock. The Significance of Hyperalaninemia in the Prediction of Lethality Birgit Metzler, Albert W. Rettenmeier, Isolde Wodarz, and Friedrich W. Schmahl	595
Metabolism and Function of Septic Kidneys K. Kürten	601
Regional Respiratory Quotients in Sepsis and Shock Ivo Giovannini, Carlo Chiarla, Giuseppe Boldrini, and Marco Castagneto	607
Analysis of the Determinants of CO₂ and O₂ Exchange Ratios in Shock Ivo Giovannini, Carlo Chiarla, Giuseppe Boldrini, Carlo Iannace, and Marco Castagneto	613
Hyperventilation in Trauma and Shock Carlo Chiarla, Ivo Giovannini, Giuseppe Boldrini, and Marco Castagneto	619

2. MONITORING SCORES/BIOLOGICAL MONITORING

The Use of Scoring Systems in Patients With Cardiogenic and Septic Shock
 Günter Pilz, Alexander Stäblein, Elisabeth Reuschel-Janetschek,
 Gernot Autenrieth, and Karl Werdan 625

Prognostic Indices of Sepsis
 Angelo Nespoli, Pietro Padalino, Claudio Marradi, Jacopo Pallavicini,
 Luca Fattori, and Giuliana Bisiani 633

**Efficiency of Sepsis Score, AT III- and Endotoxin Evaluation in Predicting
 the Prognosis of Post-Operative Sepsis in the Intensive Care Unit**
 N. Kipping, R. Grundmann, M. Hornung, and C. Wesoly 637

Risk Factors of the Multiple Organ Failure
 P. Lehmkuhl, A. Schultz, and J. Gebert 643

Biochemical Analysis in Posttraumatic and Postoperative Organ Failure
 Heinz Redl and Günther Schlag 649

Posttraumatic Plasma Levels of Mediators of Organ Failure
 Marianne Jochum, Alexander Dwenger, Theo Joka, and Johannes Sturm 673

Plasma Levels of Granulocyte Elastase and Neopterin in Patients With MOF
 Richard Pacher, Heinz Redl, and Wolfgang Woloszczuk 683

Elastase- α_1 -PI: Early Indicator of Systemic Infections in Pediatric Patients
 Christian P. Speer, Michaela Rethwilm, Friedrich Tegtmeier, and
 Manfred Gahr 689

**Leucocytes, Neutrophilia, and Elastase-a1-Proteinase-Inhibitor-Complex:
 Marker of Different Validity for Monitoring the Perioperative Infection Risk**
 Peter C. Fink, Rolf Erdmann, Friedrich Schöndube, and Ivo Baca 695

Validity of the Elastase Assay in Intensive Care Medicine
 Hermann Lang, Marianne Jochum, Hans Fritz, and Heinz Redl 701

**An Automated Homogeneous Enzyme Immunoassay for Human PMN
 Elastase**
 M. Dreher, G. Gunzer, R. Helger, and H. Lang 707

**Diiodotyrosine (DIT): A New Marker of Leukocyte Phagocytic Activity in
 Sepsis and Severe Infections**
 H.-J. Gramm, H. Meinhold, K. Voigt, and R. Dennhardt 711

Serum Proteins and Cytokines for Prediction of Sepsis?
 A.F. Hammerle, G. Pöschl, R. Kirnbauer, F. Trautinger, M. Micksche, and
 O. Mayrhofer 715

The Prognostic Value of Plasmaproteins in Patients With Abdominal Sepsis
 Michael Rogy, Reinhold Függer, Wolfgang Graninger, Friedrich Herbst,
 Michael Schemper, and Franz Schulz 719

CRP Predicts Complications in Pancreatitis and Peritonitis
 Åke Lassin, Rikard Berling, and Kjell Ohlsson 725

**The PFI-Index According to Aasen for Prognosis and Course of
 Polytraumatized Patients**
 D. Nast-Kolb, Ch. Waydhas, I. Baumgartner, M. Jochum, K.-H. Duswald,
 and L. Schweiberer 731

Components of the Kallikrein-Kinin-System in Patients With ARDS	
G. Fuhrer, W. Heller, W. Junginger, O. Gröber, and K. Roth	737
Biochemical and Hormonal Parameters in Patients With Multiple Trauma	
M. Brandl, E. Pscheidl, W. Amann, A. Barjasic, and Th. Pasch	743
Patterns of Endocrine Secretion During Sepsis	
R. Denhardt, H.-J. Gramm, K. Meinhold, and K. Voigt	751
Phospholipase A in Severely Ill Patients	
Roland M. Schaefer, M. Teschner, and A. Heidland	757
The Clinical Significance of Serum Phospholipase A in Patients With Multiple Trauma	
Ch. Waydhas, I. Baumgartner, D. Nast-Kolb, P. Lehnert, K.H. Duswald, and L. Schweiberer	763
Lymphocyte/Monocyte-Ratio Correlates With Survival From Infections and Multi-Organ Failure Following Polytrauma	
Michael W. Holch, Peter J. Grob, and Werner Glinz	769
A Prospective Study to Evaluate Posttraumatic Liver Function by Scintigraphy as a Possible Predictor of Organ Failure	
G. Regel, M.L. Nerlich, K.F. Gratz, H.P. Friedl, and J.A. Sturm	775
3. GENERAL THERAPY	
Prophylaxis and Therapy of the Multiple Organ Failure Syndrome (MOFS): Early Ventilatory Support	
Herbert Benzer, Wolfgang Koller, Christian Putensen, and Günther Putz	783
The Use of Exogenous Surfactant to Treat Patients With Acute High-Permeability Lung Edema	
Roger G. Spragg, Paul Richman, Nicolas Gilliard, T.Allen Merritt, Bengt Robertson, and Tore Curstedt	791
Exogenous Surfactant in Experimental Aspiration Trauma	
Wolfgang Strohmaier, Heinz Redl, and Günther Schlag	797
Effect of an Altered Fluid Regimen on Extravascular Lung Water in Advanced Septic Shock States	
Ernst Zadrobilek, Vichra Evstatieva, Paul Sporn, and Karl Steinbereithner	803
Effect of Large Volume Replacement With Crystalloids on Extravascular Lung Water in Human Septic Shock Syndrome	
Ernst Zadrobilek, Werner Hackl, Paul Sporn, and Karl Steinbereithner	809
Hydroxyethyl Starch and Lung Lymph Flow in an Ovine Model of Endotoxemia	
Hans J. Lübbsmeyer, Jesse Basadre, Michael Möllmann, Lillian Traber, James Maguire, David N. Herndon, and Daniel L. Traber	815
Can Hemofiltration Increase Survival Time in Acute Endotoxemia—A Porcine Shock Model	
Karl-H. Staubach, H.-G. Rau, A. Kooistra, H.-M. Schardey, G. Hohlbach, and F. W. Schildberg	821

Decontamination of the Gastrointestinal Tract and Prevention of Multiple Organ Failure. An Experimental Study Ignas P.T. van Bebber, Roland M.G.H. Mollen, Joop P. Koopman, and R. Jan A. Goris	827
 3.1. Corticosteroids	
Development of Animal Models for Application to Clinical Trials in Septic Shock Lerner B. Hinshaw	835
Dilemmas of the Clinical Trial; Review and Critique of VA Cooperative Study of Corticosteroid in Systemic Sepsis Michael F. Wilson	847
Corticosteroids for Septic Shock and the Adult Respiratory Distress Syndrome Roger C. Bone	857
Nebulized Corticosteroid in Experimental Respiratory Distress Sten Walther, Ingvar Jansson, Björn Bäckstrand, and Sten Lennquist	867
Influence of Methylprednisolone Pretreatment on Coagulation, Fibrinolysis, Hemodynamics, and Cellular Responses in Porcine Endotoxemia Olav Røise, Frode Naess, Johan Pillgram-Larsen, Tom E. Ruud, Jan O. Stadaas, and Ansgar O. Aasen	873
Prevention of Anaphylatoxin Formation by High-Dose Corticosteroids in Total Hip Arthroplasty Wolfgang Gammer, Anders Bengtson, and Mats Heideman	879
 3.2. Radical Scavengers	
Free Radical Scavengers in the Cardiopulmonary Response to Endotoxin Daniel L. Traber, David N. Herndon, and Lillian D. Traber	885
The 21-Aminosteroid U74006F Reduces Systemic Lipid Peroxidation, Improves Neurologic Function, and Reduces Mortality After Cardiopulmonary Arrest in Dogs JoAnne E. Natale, Robert J. Schott, Edward D. Hall, J. Mark Braughler, and Louis G. D'Alecy	891
Alpha-Mercaptopropionylglycine in Haemorrhagic Shock B. Weidler, B. v. Bormann, M. Kahle, and G. Hempelmann	897
Dynamics of Prostacyclin and Thromboxane During Myocardial Ischemia Elizabeth Röth, Dezső Keleman, Bela Török, Alexander Nagy, and Susan Pollak	907
Protection by Recombinant Human Superoxide Dismutase in Lethal Rat Endotoxemia Johannes Schneider, Elmar Friderichs, and Hubert Giertz	913

3.3. PAF Antagonists

- Effect of a New and Specific PAF-Antagonist, WEB 2086, on PAF and Endotoxin/Tumor Necrosis Factor Induced Changes in Mortality and Intestinal Transit Velocity**
Hubert Heuer 919
- The Pathophysiological Role of PAF in Anaphylactic Lung Reaction in the Guinea Pig and in Endotoxin Shock Evidenced by the Specific PAF-Antagonist WEB 2086**
Hubert Heuer and Jorge Casals-Stenzel 925
- Effect of PAF-Antagonists in Endotoxin Shock—Ovine and Rat Experiments**
Soheyl Bahrami, Heinz Redl, Martin Thurnher, Christa Vogl, Eva Paul, Anna Schiesser, and Günther Schlag 931

3.4. Protease Inhibitors

- Therapeutic Effects of the Combination of Two Proteinase Inhibitors in Endotoxin Shock of the Pig**
M. Siebeck, H. Hoffmann, J. Weipert, and M. Spannagl 937
- Leukocyte Neutral Proteinase Inhibitor of the Pig: Modification by Eglin C and Superoxide Dismutase of the Response to Shock**
M. Siebeck, H. Hoffmann, R. Geiger, and L. Schweiberer 945
- Reasons for the Ineffectiveness of Eglin C to Ameliorate Endotoxin Shock in Sheep**
Wolfgang G. Junger, Camille Lieners, Heinz Redl, and Günther Schlag 953
- Clinical Relevants of the Membrane Protective Action of Aprotinin on the Intraoperative Histamine Liberation**
Henning Harke and Salah Rahman 959
- Antithrombin III and Plasma Substitution in Septic Shock**
Rainer Seitz, Martin Wolf, and Rudolf Egbring 965
- Immunological Determination of Proteinase Inhibitor Complexes (PICs) and Their Behaviour During Plasma Derivate Treatment in Septic Infections**
Rudolf Egbring, Rainer Seitz, Heiner Blanke, T. Menges, R. Südhoff, T. Stober, G. Kolb, and L. Lerch 971
- Therapeutic Modalities to Ameliorate Endotoxin Induced DIC in the Rats**
Soheyl Bahrami, Eva Paul, Heinz Redl, and Günther Schlag 977
- Endotoxin Shock in the Rat: Reduction of Arterial Blood Pressure Fall by the Bradykinin Antagonist B4148**
Joachim Weipert, Hans Hoffmann, Matthias Siebeck, and Eric T. Whalley ... 983

3.5. Immune Therapy

- First Experience With Immunomodulation in Septic Shock**
Ch. Josten, G. Muhr, and R. Sistermann 989

Thymopentin (TP-5) in the Treatment of the Postburn and Postoperative Immunodeficiency Syndrome	
Gerhard Hamilton, Gerald Zöch, Thomas Rath, and Günther Meissl	995
Protection Against the Consequences of Intravascular Coagulation by Reticuloendothelial Stimulation	
George Lázár, Jr., Elizabeth Husztik, and George Lázár	1001
Behavior of Leukocyte Elastase and Immunoglobulins in Septic Toxic Multiorgan Involvement: Observations on 50 Gas Gangrene Cases	
D. Tirpitz	1007
Haemodynamic Effects During Treatment of Sepsis and Septic Shock With Immunoglobulins and Plasmapheresis	
Karl Werdan, Günter Pilz, and Stefan Kääb	1025
Prediction and Prevention, by Immunological Means, of Septic Complications After Elective Cardiac Surgery	
H.G. Kress, C. Scheidewig, W. Engelhardt, H. Wallasch, and O. Elert	1031
Stimulation of Phagocytosis by Immunoglobulins in Animal Experiment	
Stefan W. Frick and Rolf Hartmann	1037
Determination of Antibodies Against Bacterial Lipopolysaccharides and Lipid A by Immunoblotting	
Peter C. Fink, Gert Bokelmann, and Rainer Haeckel	1043
3.6. Inotropic Agents—Calcium Antagonists	
Diltiazem Prevents Endotoxin-Induced Disturbances in Intracellular Ca²⁺ Regulation	
Mohammed M. Sayeed	1053
Calcium Antagonists in Shock—A Minireview of the Evidence	
James R. Parratt	1065
Circulatory Responses to the Sepsis Syndrome	
William J. Sibbald	1075
Therapy of Acute Respiratory Distress Syndrome With Nifedipine	
Peter Hoffmann, Michael Imhoff, and Ralf Gahr	1087
Pharmacological Effects of RA 642 on Cerebrocortical Perfusion in Acute Hemorrhagic Shock in Rats	
Stefan Hergenröder and Richard Reichl	1091
Long Term Administration of Dopamine: Is There a Development of Tolerance?	
G.G. Braun, F. Bahlmann, M. Brandl, and R. Knoll	1097
Use of Systolic Time Intervals to Evaluate the Effect of Dopamine Infusion in Septic and Burn Shock	
Kornél Szabó	1101
Index	1107

Contributors

Ansgar O. Aasen, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [315,389,401,873]

Homerios Aletras, Department of Surgery, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

Sefik Alkan, Department of Surgery, LMU Munich, Klinikum Grosshadern, D-8000 Munich 70, Federal Republic of Germany [495]

Athanasia Alvanou, Department of Histology, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

W. Amann, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [743]

Paul L. Appel, Department of Surgery, King-Drew Medical Center, Los Angeles, CA 90059 [133]

Z. Atay, Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [51]

Walter Aulitzky, Department of Internal Medicine, University of Innsbruck, Innsbruck A-6020, Austria [467]

Gernot Autenrieth, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [625]

Ivo Baca, Department für Chirurgie, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [695]

Björn Bäckstrand, Department of Surgery, Regionsjukhuset, S-581 85 Linköping, Sweden [867]

Andrea Bader, Institute of Immunology, University of Heidelberg, 6900 Heidelberg, Federal Republic of Germany [299]

Gregory J. Bagby, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [545]

F. Bahlmann, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [1097]

Soheyl Bahrami, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [931,977]

A. Barjasic, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [743]

The numbers in brackets are the opening page numbers of the contributors' articles.

Robert E. Barrow, Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [377]

Jesse Basadre, Department of Anesthesiology and Surgery, The University of Texas Medical Branch and Division of Anesthesia Research, Shriners Burns Institute, Galveston, TX 77550 [815]

J.A. Bauer, Chirurg. Klinik Innenstadt und Chirurg. Polikl. der Universität, D-8000 München 2, Democratic Republic of Germany [455]

Franz Bäumer, Chirurgische Universitätsklinik, Experimentelle Chirurgie, D-8700 Würzburg, Federal Republic of Germany [143]

I. Baumgartner, Abt. Klin. Chemie und Klin. Biochemie in der Chir. Klinik Innenstadt, Universität München, 8000 München 2, Federal Republic of Germany [731,763]

Gerard I.J.M. Beerthuisen, Department of General Surgery, University Hospital Nijmegen, 6500 HB Nijmegen, The Netherlands [137]

Anders Bengtson, Department of Anesthesiology, Sahlgren Hospital, University of Göteborg, 41345 Göteborg, Sweden [265,879]

Herbert Benzer, Clinic for Anaesthesia and General Intensive Care Medicine, A-6020 Innsbruck, Austria [783]

Rikard Berling, Department of Anaesthesiology, Malmö General Hospital, University of Lund, S-214 01 Malmö, Sweden [725]

Sandra Betti, Cardiovascular Surgery, University of Siena, 53100 Siena, Italy [73]

G. Beverley, The HA Grové Research Center of the University of Pretoria, Pretoria, South Africa [217]

Sucharit Bhakdi, Department of Microbiology, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [67,283]

Ron Bilik, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

Timothy R. Billiar, Departments of Surgery and Biochemistry, University of Minnesota Medical School, Minneapolis, MN 55455 [563]

A. Billing, Chirurg. Klinik und Poliklinik der Universität München, Klinikum Grosshadern, 8000 München 70, Federal Republic of Germany [527]

Ulrich Birkenhauer, Klinik für Anaesthesiologie und Transfusionsmedizin der Universität Tübingen, D-7400 Tübingen, Federal Republic of Germany [175]

G. Bisiani, Department of Internal Medicine, University of Milan, 20122 Milan, Italy [277]

Giuliana Bisiani, Department of Emergency Surgery, University of Milan, Milan 20122, Italy [633]

Heiner Blanke, Department Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

Miguel Blazquez, Intensive Care Unit, Hospital N.S. del Pino, Las Palmas, Canary Islands, Spain [57,61]

Günther Blümel, Department of Experimental Surgery, Technical University, 8000 Munich 80, Federal Republic of Germany [339]

Gert Bokelmann, Institut für Laboratoriumsmedizin-Zentrallabor, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [1043]

Giuseppe Boldrini, Centro di Studio per la Fisiopatologia dello Shock, CNR, Istituto di Clinica Chirurgica, Università Cattolica, Roma, Italy [259,607,613,619]

B. Bonavida, Department of Microbiology and Immunology, UCLA School of Medicine, Los Angeles, CA 90024 [485]

Roger C. Bone, Rush-Presbyterian-St. Luke's Medical Center, Chicago, IL 60612 [857]

Gerhard Borkowski, Department of Internal Medicine, University of Giessen, 6300 Giessen, Federal Republic of Germany [383]

Emma Borrelli, Cardiovascular Surgery, University of Siena, 53100 Siena, Italy [73]

Ulrich Bosch, Department of Traumasurgery, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37]

Bonno N. Bouma, Department of Hematology, University Hospital Utrecht, Utrecht, The Netherlands [401]

M. Brandl, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [743,1097]

Pierre Braquet, I.H.B. Research Labs., 92350 Le Plessis-Robinson, France [425,441,485]

J. Mark Braugher, CNS Diseases Research, The Upjohn Company, Kalamazoo, MI 49001 [891]

G.G. Braun, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [1097]

K.L. Brigham, Department of Medicine, The Center for Lung Research, Vanderbilt University Medical Center, Nashville, TN 37232 [91]

Volker Bühren, Department Trauma Surgery, University of Saarland, D-6650 Homburg/Saar, Federal Republic of Germany [523]

Hilmar Burchardi, Department of Anaesthesiology, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [323]

Reinhard Burger, Robert Koch Institute, 1000 Berlin, Federal Republic of Germany [299]

R.D. Burrow, Department of Medicine, University of Oklahoma City, Oklahoma, OK 73104 [207]

Wim A. Buurman, Department of General Surgery, University of Limburg, Biomedical Center, 6200 MD Maastricht, The Netherlands [463]

Jorge Casals-Stenzel, Department of Pharmacology, Boehringer Ingelheim KG, D-6507 Ingelheim, Federal Republic of Germany [925]

Marco Castagneto, Centro di Studio per la Fisiopatologia dello Shock, CNR, Istituto di Clinica Chirurgica, Università Cattolica, Roma, Italy [259,607,613,619]

Matthias Cebulla, Department of Surgery, University Hospital Frankfurt/M., D-6000 Frankfurt/M. 70, Federal Republic of Germany [517]

Frank B. Cerra, Departments of Surgery and Biochemistry, University of Minnesota Medical School, Minneapolis, MN 55455 [563]

Huacui Chen, Department of Pathophysiology, Peking Union Medical College, Beijing 100700, China [271]

Li Chen, Department of Pathophysiology, Peking Union Medical College, Beijing 100700, China [271]

Oswaldo Chiara, Department of Emergency Surgery, University of Milan, 20122 Milan, Italy [73,277]

Carlo Chiarla, Centro di Studio per la Fisiopatologia dello Shock, CNR, Istituto di Clinica Chirurgica, Università Cattolica, Roma, Italy [259,607,613,619]

J. Allen Coffey, Jr., Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [377]

Bill Coleman, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

P. Conzen, Chirurg. Klinik Innenstadt und Chirurg. Polikl. der Universität, D-8000 München 2, Federal Republic of Germany [455]

Tore Curstedt, Department of Clinical Chemistry, Karolinska Hospital, Stockholm, Sweden [791]

Louis G. D'Alecy, Departments of Physiology and Surgery, The University of Michigan Medical School, Ann Arbor, MI 48109 [891]

Ying Dang, Department of Pathophysiology, Peking Union Medical College, Beijing 100700, China [271]

Reiner Dauberschmidt, Research Department of Intensive Care Medicine, Friedrichshain Hospital Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

I.V. Deaciuc, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [575]

D. De Backer, Department of Intensive Care, Erasme Hospital, Free University of Brussels, 1070 Brussels, Belgium [181]

Joop M.H. Debets, Department of General Surgery, University of Limburg, Biomedical Center, 6200 MD Maastricht, The Netherlands [463]

Peter J. Del Vecchio, Departments of Ophthalmology and Physiology, The Albany Medical College, Albany, NY 12208 [101]

R. Denhardt, Klinik für Anästhesiologie, Krankenhaus Nordwest, 6000 Frankfurt 90, Federal Republic of Germany [711,751]

E.B. Rodriguez de Turco, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [575]

R. de Winter, Medical Center Veterans Administration, Oklahoma City, OK 73104 [207]

Heinrich Ditter, Department of Internal Medicine, University of Giessen, 6300 Giessen, Federal Republic of Germany [383]

I.C. Dormehl, AEC Institute for Life Sciences, University of Pretoria, Pretoria, South Africa [207,217]

M. Dreher, Diagnostica Forschung, E. Merck, D-6100 Darmstadt, Federal Republic of Germany [707]

Detlev Drenckhahn, Department of Anatomy and Cell Biology (DD), Phillips University, D-3550 Marburg, Federal Republic of Germany [127]

Christel Dressler, Department of Anaesthesiology, Friedrichshain Hospital of Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

Günter Dussing, Institute of Biochemistry, University of Graz, A-8010 Graz, Austria [351]

K.H. Duswald, Chirurgische Klinik Innenstadt, Universität München, 8000 München 2, Federal Republic of Germany [731,763]

Alexander Dwenger, Klinische Biochemie, Medizinischen Hochschule Hannover, D-3000 Hannover 61, Federal Republic of Germany [673]

Rudolf Egbring, Division of Internal Medicine, Department of Hematology, Philipps-University, D-3550 Marburg, Federal Republic of Germany [965,971]

Gerd Egger, Institute of Functional Pathology, University of Graz, A-8010 Graz, Austria [351]

Efthimios Eleftheriadis, Department of Surgery, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

O. Elert, Department of Thoracic and Cardiovascular Surgery, University Hospital, D-8700 Würzburg, Federal Republic of Germany [1031]

W. Engelhardt, Institute of Anaesthesiology, University Hospital, D-8700 Würzburg, Federal Republic of Germany [1031]

Rolf Erdmann, Institut für Laboratoriumsmedizin-Zentrallabor, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [695]

Wolfgang Ertel, Department of Surgery, LMU Munich, Klinikum Grosshadern, D-8000 München 70, Federal Republic of Germany [495,513]

Vichra Evstatieva, Ludwig Boltzmann Institute, Department of Anaesthesia and Intensive Care, A-1090 Vienna, Austria [803]

Eugen Faist, Department of Surgery, LMU Munich, Klinikum Grosshadern, 8000 München 70, Federal Republic of Germany [495,513]

Luca Fattori, Department of Emergency Surgery, University of Milan, Milan 20122, Italy [633]

Walter Fierz, Section of Clinical Immunology, Department of Medicine, University Hospital Zurich, CH-8091 Zurich, Switzerland [491]

Peter C. Fink, Institut für Laboratoriumsmedizin-Zentrallabor, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [695,1043]

Chryssi Foroglou, Department of Histology, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

Knut Frederking, Department of Surgery, University Hospital Frankfurt/M., D-6000 Frankfurt/M. 70, Federal Republic of Germany [517]

Reinhold Fretschner, Klinik für Anaesthesiologie und Transfusionsmedizin der Universität Tuebingen, D-7400 Tuebingen, Federal Republic of Germany [185]

M.A. Freudenberg, Max-Planck-Institut für Immunbiologie, Freiburg/Br., Federal Republic of Germany [407]

Nikolaus Freudenberg, Department of Pathology, University of Freiburg, D-7800 Freiburg, Federal Republic of Germany [77,113,371,407]

Stefan W. Frick, Surgical University Clinic Marienhospital Ruhr-University of Bochum, D-4690 Herne 1, Federal Republic of Germany [1037]

Elmar Friderichs, Department of Pharmacology, Grünenthal GmbH, 5100 Aachen, Federal Republic of Germany [913]

H.P. Friedl, Department of Traumatology, Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [775]

Arno Friedlein, Progen Biotechnik, 6900 Heidelberg, Federal Republic of Germany [299]

Hans Fritz, Department of Surgery, Division of Clinical Chemistry and Clinical Biochemistry, University of Munich, D-8000 Munich 2, Federal Republic of Germany [305,395,455,701]

xxiv / Contributors

D. Fröhlich, Chirurg. Klinik und Poliklinik der Universität München, Klinikum Grosshadern, 8000 München 70, Federal Republic of Germany [527]

Thomas Fuchs, Department of Anatomy and Cell Biology (DD), Phillips University, D-3550 Marburg, Federal Republic of Germany [127]

Reinhold Függer, Department of Surgery 1, University of Vienna Medical School, A-1090 Vienna, Austria [719]

G. Fuhrer, Department of Cardiovascular Surgery, University of Tübingen, D-7400 Tübingen, Federal Republic of Germany [737]

J. Funovics, First Surgical University Clinic, Metabolic Research Laboratory, University Vienna, A-1090 Vienna, Austria [589]

Walter Fürst, Ludwig Boltzmann Institute for Experimental Traumatology, Vienna A-1200, Austria [345]

Manfred Gahr, Department of Pediatrics, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [689]

Ralf Gahr, Unfallchirurgische Klinik, Städtische Kliniken Dortmund, D-4600 Dortmund 1, Federal Republic of Germany [1087]

C. Galanos, Max-Planck-Institut für Immunbiologie, Freiburg/Br., Federal Republic of Germany [407]

Wolfgang Gammer, Department of Orthopaedic Surgery, Ludvika Hospital, 771 00 Ludvika, Sweden [879]

M. Gardinali, Department of Internal Medicine, University of Milan, 20122 Milan, Italy [277]

Harald Gasser, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [447]

J. Gebert, Zentrum für Anästhesie der Medizinischen, Hochschule Hannover, 3000 Hannover 51, Federal Republic of Germany [643]

R. Geiger, Abteilung für Klinische Chemie und Klinische Biochemie in der Chirurgie Innenstadt, Ludwig-Maximilians-Universität München, D-8000 München 2, Federal Republic of Germany [945]

Stephanos Geroulanos, Department of Surgery, University Hospital Zurich, CH-8091 Zurich, Switzerland [491]

Anders Gidlöf, Clinical Research Center, Faculty of Health Sciences, University Hospital, S-581 85 Linköping, Sweden [157]

Hubert Giertz, Department of Pharmacology, Grünenthal GmbH, 5100 Aachen, Federal Republic of Germany [913]

Nicolas Gilliard, Division of Pulmonary and Critical Care Medicine, Department of Medicine, University of California, San Diego, CA 92103 [791]

Pier P. Giomarelli, Cardiovascular Surgery, University of Siena, 53100 Siena, Italy [73]

Ivo Giovannini, Centro di Studio per Fisiopatologia dello Shock, CNR, Istituto di Clinica Chirurgica, Università Cattolica, Roma, Italy [259,607,613,619]

Werner Glinz, Section of Clinical Immunology, Department of Surgery, University Hospital, CH-8091 Zurich, Switzerland [491,769]

Oliver Gonschorek, Department of Trauma Surgery, University of Saarland, D-6650 Homburg/Saar, Federal Republic of Germany [523]

R. Jan A. Goris, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [137,419,827]

Notker Graf, Department of Anaesthesiology, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [323]

Hans Grajetzki, Central Resuscitation and Intensive Care Unit, Friedrichshain Hospital Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

H.-J. Gramm, Klinik für Anästhesiologie und operative Intensivmedizin, Klinikum Steglitz der Freien Universität Berlin, D-1000 Berlin 45, Federal Republic of Germany [711,751]

Wolfgang Graninger, Department of Chemotherapy, University of Vienna Medical School, A-1090 Vienna, Austria [719]

K.F. Gratz, Department of Nuclearmedicine, Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [775]

Barbara Griess, Research Department of Intensive Care Medicine, Friedrichshain Hospital of Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

Peter J. Grob, Section of Clinical Immunology, Department of Medicine, University Hospital, CH-8091 Zurich, Switzerland [491,769]

O. Gröber, Department of Cardiovascular Surgery, University of Tübingen, D-7400 Tübingen, Federal Republic of Germany [737]

A.B.J. Groeneveld, Medical Intensive Care Unit, Free University Hospital, Amsterdam [163]

R. Grundmann, Chirurgische Universitätsklinik Köln-Lindenthal, D-5000 Köln 41, Federal Republic of Germany [637]

Heinz Guggenberger, Klinik für Anaesthesiologie und Transfusionsmedizin der Universität Tuebingen, D-7400 Tuebingen, Federal Republic of Germany [185]

G. Gunzer, Diagnostica Forschung, E.Merck, D-6100 Darmstadt, Federal Republic of Germany [707]

C.E. Hack, Central Laboratory of the Netherlands Red Cross Bloodtransfusion Service and Laboratory for Experimental and Clinical Immunology, University of Amsterdam, Amsterdam [163]

Werner Hackl, Ludwig Boltzmann Institute, Department of Anaesthesia and Intensive Care, Vienna University School of Medicine, A-1090 Vienna, Austria [809]

Rainer Haeckel, Institut für Laboratoriumsmedizin-Zentrallabor, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [1043]

Edward D. Hall, CNS Diseases Research, The Upjohn Company, Kalamazoo, MI 49001 [891]

Seth W.O. Hallström, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [225,231,345]

Gerhard Hamilton, Experimental Surgery, University Clinic, Burn Care Unit, A-1090 Vienna, Austria [995]

A.F. Hammerle, Department of Anaesthesiology and General Intensive Care, University of Vienna, A-1090 Vienna, Austria [715]

Diane M. Hargrove, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [545]

xxvi / Contributors

Henning Harke, Department of Anaesthesia, General Hospital of Krefeld, D-4150 Krefeld, Federal Republic of Germany [959]

Rolf Hartmann, Surgical University Clinic Marienhospital Ruhr-University of Bochum, D-4690 Herne 1, Federal Republic of Germany [1037]

Ruth Hartmann, Department of Internal Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [283]

Mats Heideman, Department of Surgery, Sahlgren Hospital, University of Göteborg, 41345 Göteborg, Sweden [265,879]

A. Heidland, Department of Medicine, University of Wuerzburg, D-8700 Wuerzburg, Federal Republic of Germany [757]

R. Helger, Diagnostica Forschung, E. Merck, D-6100 Darmstadt, Federal Republic of Germany [707]

W. Heller, Department of Cardiovascular Surgery, University of Tübingen, D-7400 Tübingen, Federal Republic of Germany [737]

G. Hempelmann, Department of Anaesthesia and Intensive Care Medicine, Justus-Liebig Universität, D-6300 Giessen, Federal Republic of Germany [897]

Hermann August Henrich, Chirurgische Universitätsklinik, Experimentelle Chirurgie, D-8700 Würzburg, Federal Republic of Germany [143]

Friedrich Herbst, Department of Surgery I, University of Vienna Medical School, A-1090 Vienna, Austria [719]

Stefan Hergenröder, Department of Pharmacology, Boehringer Ingelheim KG, D-6507 Ingelheim/Rhein, Federal Republic of Germany [1091]

J.B. Hermiller, Naval Medical Research Institute, Bethesda, MD 20814 [575]

David N. Herndon, Department of Anesthesiology and Surgery, The University of Texas Medical Branch and Division of Anesthesia Research, Shriners Burns Institute, Galveston, TX 77550 [201,377,815,885]

Thomas Hessz, Department of Anatomy and Cell Biology (DD), Phillips University, D-3550 Marburg, Federal Republic of Germany [127]

Hubert Heuer, Department of Pharmacology, Boehringer Ingelheim KG, D-6507 Ingelheim, Federal Republic of Germany [919,925]

Lerner B. Hinshaw, Oklahoma Medical Research Foundation, and University of Oklahoma Health Sciences Center, Oklahoma City, OK 73104 [835]

Herwig P. Hofer, Surgical Clinic, University of Graz, A-8010 Graz, Austria [351]

Ch. Hoffmann, Department of Internal Medicine, Division of Clinical Pathophysiology and Experimental Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [305]

H. Hoffmann, Abteilung für Klinische Chemie und Klinische Biochemie in der Chirurgie Innenstadt, Ludwig-Maximilians Universität München, D-8000 München 2, Federal Republic of Germany [395,937,945,983]

Peter Hoffmann, Abteilung für Anästhesiologie, Städtische Kliniken Dortmund, D-4600 Dortmund 1, Federal Republic of Germany [1087]

James C. Hogg, Pulmonary Research Laboratory, University of British Columbia, St. Paul's Hospital, Vancouver, Canada V6Z 1Y6 [27]

G. Hohlbach, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Michael Holch, Department of Trauma Surgery, Medical School Hannover, 3000 Hannover 61, Federal Republic of Germany [507]

Michael W. Holch, Section of Clinical Immunology, Department of Medicine, University Hospital, CH-8091 Zurich, Switzerland [491,769]

Ralph T. Holman, Departments of Surgery and Biochemistry, University of Minnesota Medical School, Minneapolis, MN 55455 [563]

M. Hornung, Chirurgische Universitätsklinik Köln-Lindenthal, D-5000 Köln 41, Federal Republic of Germany [637]

David Hosford, I.H.B. Research Labs., 92350 Le Plessis-Robinson, France [425,485]

Christoph Huber, Department of Internal Medicine, University of Innsbruck, A-6020 Innsbruck, Austria [467]

N. Hugo, The HA Grové Research Center of the University of Pretoria, Pretoria, South Africa [207,217]

Elizabeth Husztki, Institute of Medical Biology, Albert Szent-Györgyi Medical University, Szeged, Hungary [1001]

Carlo Iannace, Centro di Studio per la Fisiopatologia dello Shock, CNR, Istituto di Clinica Chirurgica, Università Cattolica, Roma, Italy [613]

Michael Imhoff, Chirurgische Klinik, Städtische Kliniken Dortmund, D-4600 Dortmund 1, Federal Republic of Germany [1087]

Ingvar Jansson, Department of Surgery, Regionsjukhuset, S-581 85 Linköping, Sweden [867]

Marianne Jochum, Klinische Chemie und Klinische Biochemie, Chirurgische Klinik Innenstadt, Universität München, D-8000 München 2, Federal Republic of Germany [527,673,701,731]

J.E. Johnson, Department of Pathology, The Center for Lung Research, Vanderbilt University Medical Center, Nashville, TN 37232 [91]

Theo Joka, Department of Traumatology, University of Essen, D-4300 Essen 1, Federal Republic of Germany [43,51,673]

F. Joo, Biological Research Center of the Academy of Science of Hungary, Hungary [291]

Ch. Josten, Department of Surgery, Berufsgenossenschaftliche Krankenanstalten "Bergmannsheil", 4630 Bochum, Federal Republic of Germany [989]

Wolfgang G. Junger, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [953]

W. Junginger, Department of Cardiovascular Surgery, University of Tübingen, D-7400 Tübingen, Federal Republic of Germany [737]

Stefan Kääb, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [1025]

Karin Kaden, Paediatric Clinic, Friedrichshain Hospital Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

M. Kahle, Department of General and Thoracic Surgery, Justus-Liebig Universität, D-6300 Giessen, Federal Republic of Germany [897]

J. Kalotai, Department of Traumatology, University of Essen, 4300 Essen 1, Federal Republic of Germany [51]

T. Kapsner, Department of Medicine I, Klinikum Grosshadern, University of Munich, 8000 Munich 70, Federal Republic of Germany [247]

M.R. Karim, Chirurgische Klinik, Städtisches Klinikum Karlsruhe, Karlsruhe, Federal Republic of Germany [407]

J. Karner, First Surgical University Clinic, Metabolic Research Laboratory, University Vienna, A-1090 Vienna, Austria [589]

József Kaszaki, Institute of Experimental Surgery, Szent-Györgyi Albert Medical University, H-6720 Szeged, Hungary [253,291]

Dezsö Kelemen, Department of Experimental Surgery, University of Medicine, Pécs, Hungary H-7643 [907]

Zafar Khakpour, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [225,231]

J.G. Kilian, Department of Medicine, University of Pretoria, Pretoria, South Africa [207,217]

N. Kipping, Chirurgische Universitätsklinik Köln-Lindenthal, D-5000 Köln 41, Federal Republic of Germany [637]

R. Kirnbauer, Department of Anaesthesiology and General Intensive Care, University of Vienna, A-1090 Vienna, Austria [715]

Michael Kirschfink, Department of Immunology, University of Heidelberg, D-6900 Heidelberg, Federal Republic of Germany [43]

Werner J. Kleeman, Department of Forensic Medicine, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37]

Thomas Kloess, Klinik für Anaesthesiologie und Transfusionsmedizin der Universitaet Tuebingen, D-7400 Tuebingen, Federal Republic of Germany [175,185]

R. Knoll, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [1097]

G. Kolb, Department of Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

Wolfgang Koller, Clinic for Anaesthesia and General Intensive Care Medicine, A-6020 Innsbruck, Austria [783]

Peter Konold, Department of Surgery, University Hospital Frankfurt/M., D-6000 Frankfurt/M. 70, Federal Republic of Germany [517]

A. Kooistra, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Joop P. Koopman, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [827]

H. Kortmann, Chirurg. Klinik und Poliklinik der Universität München, Klinikum Grosshadern, 8000 München 70, Federal Republic of Germany [527]

Bernd Kottler, Klinik für Anaesthesiologie und Transfusionsmedizin der Universitaet Tuebingen, D-7400 Tuebingen, Federal Republic of Germany [175]

Katerina Kotzampassi, Department of Surgery, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

E. Kovats, First Surgical University Clinic, Metabolic Research Laboratory, University Vienna, A-1090 Vienna, Austria [589]

Harry B. Kram, Department of Surgery, King-Drew Medical Center, Los Angeles, CA 90059 [133]

Sabine Krautschneider, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [447]

H.G. Kress, Institute of Anaesthesiology, University Hospital, D-8700 Würzburg, Federal Republic of Germany [1031]

Ferdinand J.A. Kreuzer, Department of Physiology, University Hospital Nijmegen, 6500 HB Nijmegen, The Netherlands [137]

E. Kreuzfelder, Institute of Virology and Immunology, University of Essen, 4300 Essen 1, Federal Republic of Germany [51]

Peter E. Krösl, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [225,231,237]

Peter Kühnl, Institute of Immunohematology, University Hospital Frankfurt/M., D-6000 Frankfurt/M. 70, Federal Republic of Germany [278]

K. Kürten, Chirurgische Universitätsklinik Köln, 5000 Köln 41, Federal Republic of Germany [601]

Herbert Lamche, Ernst Boehringer Institute, A-1120 Vienna, Austria [467]

Charles H. Lang, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [545]

Hermann Lang, Diagnostic Research, E. Merck Darmstadt, D-6100 Darmstadt, Federal Republic of Germany [701,707]

Åke Lasson, Departments of Surgery and Surgical Pathophysiology, Malmö General Hospital, University of Lund, S-214 01 Malmö, Sweden [725]

Z. Laszik, Institute of Pathology, Medical University of Szeged, Szeged, Hungary [291]

Martin Lausen, Department of Surgery, University of Freiburg, 7800 Freiburg, Federal Republic of Germany [371]

George Lázár, Institute of Pathophysiology, Albert Szent-Györgyi Medical University, Szeged, Hungary [1001]

George Lázár, Jr., Department of Surgery, Albert Szent-Györgyi Medical University, Szeged, Hungary [1001]

P. Lehmkuhl, Zentrum für Anästhesie der Medizinischen Hochschule Hannover, 3000 Hanover 51, Federal Republic of Germany [643]

P. Lehnert, Med. Klinik Innenstadt, Universität München, 8000 München 2, Federal Republic of Germany [763]

Sten Lennquist, Department of Surgery, Regionsjukhuset, S-581 85 Linköping, Sweden [867]

L. Lerch, Department of Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

David H. Lewis, Clinical Research Center, Faculty of Health Sciences, University Hospital, S-581 85 Linköping, Sweden [157]

Camille Lieners, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [345,467,953]

Zhenyuan Liu, Department of Pathophysiology, Peking Union Medical College, Beijing 100700, China [271]

Hans J. Lübbsmeyer, Department of Anesthesiology and Operative Intensive Care, Westfaelian Wilhelms University, D-4400 Münster, Federal Republic of Germany [815]

Santiago Lubillo, Intensive Care Unit, Hospital N.S. del Pino, Las Palmas, Canary Islands, Spain [57,61]

Heribert Luig, Department of Nuclear Medicine, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [323]

Mohammad Maghsudi, Department of Trauma Surgery, Medical School Hannover, 3000 Hannover 61, Federal Republic of Germany [507]

James Maguire, Department of Anesthesiology and Surgery, The University of Texas Medical Branch and Division of Anesthesia Research, Shriners Burns Institute, Galveston, TX 77550 [815]

Asrar B. Malik, Department of Physiology, The Albany Medical College, Albany, NY 12208 [101]

Jose L. Manzano, Intensive Care Unit, Hospital N.S. del Pino, Las Palmas, Canary Islands, Spain [57,61]

M. Maree, The HA Grové Research Center of the University of Pretoria, Pretoria, South Africa [207,217]

Claudio Marradi, Department of Emergency Surgery, University of Milan, Milan 20122, Italy [633]

F. Reinhard Matthias, Department of Internal Medicine, University of Giessen, 6300 Giessen, Federal Republic of Germany [383]

O. Mayrhofer, Department of Anaesthesiology and General Intensive Care, University of Vienna, A-1090 Vienna, Austria [715]

J.P. Mehegan, Naval Medical Research Institute, Bethesda, MD 20814 [575]

H. Meinhold, Abteilung für Nuklearmedizin, Klinikum Steglitz FU Berlin, D-1000 Berlin 45, Federal Republic of Germany [711]

K. Meinhold, Klinik für Anästhesiologie und operative Intensivmedizin, Klinikum Steglitz der Freien Universität Berlin, D-1000 Berlin 45, Federal Republic of Germany [751]

Günther Meissl, I. Surgery, University Clinic, Burn Care Unit, A-1090 Vienna, Austria [995]

S.M. Melnitzki, Department of Medicine I, Klinikum Grosshadern, University of Munich, 8000 Munich 70, Federal Republic of Germany [247]

Jean Michel Mencia-Huerta, I.H.B. Research Labs., 91952 Les Ulis, France [441]

T. Menges, Department of Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

T. Allen Merritt, Department of Pediatrics, University of California, San Diego, CA 92103 [791]

Károly Mészáros, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [545]

Birgit Metzler, Department of Occupational and Social Medicine, University of Tübingen, 7400 Tübingen, Federal Republic of Germany [595]

Angelika Mewes, Department of Surgery, LMU Munich, Klinikum Grosshadern, D-8000 Munich 70, Federal Republic of Germany [495]

Manfred Meyer, Research Department of Intensive Care Medicine, Friedrichshain Hospital Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

Barbara Meyrick, Department of Pathology, The Center for Lung Research, Vanderbilt University, Nashville, TN 37232 [91]

M. Micksche, Institute for Applied and Experimental Oncology, University of Vienna, A-1090 Vienna, Austria [715]

Rusty A. Milhoan, Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [377]

Jan Modig, Department of Anesthesiology and Intensive Care, University Hospital of Uppsala, S-751 85 Uppsala, Sweden [17]

Roland M.G.H. Mollen, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [827]

Michael Möllmann, Department of Anesthesiology and Operative Intensive Care, Westfaelian Wilhelms University, D-4400 Münster, Federal Republic of Germany [815]

Helmut Molnar, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [345]

Glenn Morris, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

Stephen E. Morris, Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [377]

Heinz Mrochen, Research Department of Intensive Care Medicine, Friedrichshain Hospital Berlin, DDR-1017 Berlin, Democratic Republic of Germany [331]

G. Muhr, Department of Surgery, Berufsgenossenschaftliche Krankenanstalten "Bergmannsheil", 4630 Bochum, Federal Republic of Germany [989]

Frode Naess, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [389,873]

Sándor Nagy, Institute of Experimental Surgery, Szent-Györgyi Albert Medical University, H-6701 Szeged, Hungary [253,291,907]

Werner Naser, Progen Biotechnik, 6900 Heidelberg, Federal Republic of Germany [299]

D. Nast-Kolb, Chirurgische Klinik Innenstadt, Universität München, 8000 München 2, Federal Republic of Germany [731,763]

JoAnne E. Natale, Department of Physiology, The University of Michigan Medical School, Ann Arbor, MI 48109 [891]

Michael L. Nerlich, Department of Traumasurgery, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37,413,507,775]

Angelo Nespoli, Department of Emergency Surgery, University of Milan, 20122 Milan, Italy [73,277,633]

Heinz Neuhof, Department of Internal Medicine, Division of Clinical Pathophysiology and Experimental Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [67,119,127,283,305]

Josef Newald, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [237]

J.H. Nuyens, Central Laboratory of the Netherlands Red Cross Bloodtransfusion Service and Laboratory for Experimental and Clinical Immunology, University of Amsterdam, Amsterdam [163]

Udo Obertacke, Department of Traumatology, University of Essen, D-4300 Essen 1, Federal Republic of Germany [43,51]

Kjell Ohlsson, Departments of Surgery and Surgical Pathophysiology, Malmö General Hospital, S-214 01 Malmö, Sweden [725]

L. Olivier, Department of Traumatology, University of Essen, 4300 Essen 1, Federal Republic of Germany [51]

Richard Pacher, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [683]

N. Pacitti, Department of Physiology and Pharmacology, University of Strathclyde, Glasgow G1 1XW, Scotland [357]

Pietro Padalino, Department of Emergency Surgery, University of Milan, 20122 Milan, Italy [73,277,633]

Jacopo Pallavicini, Department of Emergency Surgery, University of Milan, 20122 Milan, Italy [277,633]

Alfred Pannike, Department of Surgery, University Hospital Frankfurt/M., D-6000 Frankfurt/M. 70, Federal Republic of Germany [517]

James R. Parratt, Department of Physiology and Pharmacology, Royal College, University of Strathclyde, Glasgow G1 1XW, Scotland [357,1065]

Joseph E. Parrillo, Critical Care Medicine Department, National Institutes of Health, Bethesda, MD 20892 [191]

Th. Pasch, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [743]

Monique Paubert-Braquet, Centre de Traitement des Brûlés, Hôpital Percy, 92140 Clamart, France [441,485]

Eva Paul, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [477,467,931,977]

Botond Penke, Institute of Medical Chemistry, Szent-Györgyi Albert Medical University, H-6701 Szeged, Hungary [253]

Ulrich Pfeiffer, Department of Experimental Surgery, Technical University, 8000 Munich 80, Federal Republic of Germany [339]

Bernadette Pignol, I.H.B. Research Labs., 91952 Les Ulis, France [441]

Johan Pillgram-Larsen, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [389,874]

Günter Pilz, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [247,625,1025]

Susan Pollak, Department of Experimental Surgery, University of Medicine, Pécs, Hungary H-7643 [907]

G. Pöschl, Department of Anaesthesiology and General Intensive Care, University of Vienna, A-1090 Vienna, Austria [715]

J.P. Pretorius, AEC Institute for Life Sciences, University of Pretoria, Pretoria, South Africa [207,217]

E. Pscheidl, Institute of Anaesthesiology of the FAU Erlangen-Nürnberg, 8520 Erlangen, Federal Republic of Germany [743]

Christian Putensen, Clinic for Anaesthesia and General Intensive Care Medicine, A-6020 Innsbruck, Austria [783]

Günther Putz, Clinic for Anaesthesia and General Intensive Care Medicine, A-6020 Innsbruck, Austria [783]

Jose Quintana, Intensive Care Unit, Hospital N.S. del Pino, Las Palmas, Canary Islands, Spain [61]

Salah Rahman, Department of Anaesthesia, General Hospital of Krefeld, D-4150 Krefeld, Federal Republic of Germany [959]

Thomas Rath, I. Surgery, University Clinic, Burn Care Unit, A-1090 Vienna, Austria [995]

H.-G. Rau, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Heinz Redl, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [xli,3,231,345,447,467,649,683,701,797,931,953,977]

Gerd Regel, Department of Traumasurgery, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37,775]

Richard Reichl, Department of Pharmacology, Boehringer Ingelheim KG, D-6507 Ingelheim/Rhein, Federal Republic of Germany [1091]

Hubert Reichle, Department of Anaesthesiology, Technical University, 8000 Munich 80, Federal Republic of Germany [339]

Susanne Reisser, Department of Traumasurgery, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37]

Daniela Reitz, Department of Internal Medicine, University of Giessen, 6300 Giessen, Federal Republic of Germany [383]

Michaela Rethwilm, Department of Pediatrics, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [689]

Albert W. Rettenmeier, Department of Occupational and Social Medicine, University of Tübingen, 7400 Tübingen, Federal Republic of Germany [595]

Elisabeth Reuschel-Janetschek, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [625]

Paul Richman, Division of Pulmonary and Critical Care Medicine, Department of Medicine, University of California, San Diego, CA 92103 [791]

U.N. Riede, Department of Pathology, University of Freiburg, D-7800 Freiburg, Federal Republic of Germany [113]

Anne-Lise Rishovd, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, Oslo 4, Norway [315]

Avraham Rivkind, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

Bengt Robertson, Departments of Pediatrics and Pediatric Pathology, St. Goran's Children's Hospital, Stockholm, Sweden [791]

I.W. Rodger, Department of Physiology and Pharmacology, University of Strathclyde, Glasgow G1 1XW, Scotland [357]

Michael Rogy, Department of Surgery I, University of Vienna Medical School, A-1090 Vienna, Austria [719]

Olav Røise, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [389,401,873]

A. Roman, Department of Intensive Care, Erasme Hospital, Free University of Brussels, 1070 Brussels, Belgium [181]

B.L. Roth, Naval Medical Research Institute, Bethesda, MD 20814 [575]

E. Roth, First Surgical University Clinic, Metabolic Research Laboratory, University Vienna, A-1090 Vienna, Austria [589]

Elizabeth Röth, Department of Experimental Surgery, University of Medicine, Pécs, Hungary H-7643 [907]

K. Roth, Department of Cardiovascular Surgery, University of Tübingen, D-7400 Tübingen, Federal Republic of Germany [737]

Lennart Roxvall, Department of Surgery, Sahlgren Hospital, University of Göteborg, 41345 Göteborg, Sweden [265]

Günther Ruf, Department of Surgery, University of Freiburg, 7800 Freiburg, Federal Republic of Germany [371]

Tom E. Ruud, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [389,873]

Mohammed M. Sayeed, Department of Physiology, Loyola University, Stritch School of Medicine, Maywood, IL 60153 [1053]

Roland M. Schaefer, Department of Medicine, University of Wuerzburg, D-8700 Wuerzburg, Federal Republic of Germany [757]

H.-M. Schardey, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Erwin Schauenstein, Institute of Biochemistry, University of Graz, A-8010 Graz, Austria [351]

Rudolf J. Schaur, Institute of Biochemistry, University of Graz, A-8010 Graz, Austria [351]

C. Scheidewig, Institute of Anaesthesiology, University Hospital, D-8700 Würzburg, Federal Republic of Germany [1031]

Michael Schemper, Department of Surgery I, University of Vienna Medical School, A-1090 Vienna, Austria [719]

Anna Schiesser, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [447,467,931]

F.W. Schildberg, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Paul H.M. Schillings, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [419]

Heinrich M. Schima, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [225]

Günther Schlag, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [xli,3,231,237,345,447,467,649,797,931,953,977]

Friedrich W. Schmahl, Department of Occupational and Social Medicine, University of Tübingen, 7400 Tübingen, Federal Republic of Germany [595]

Klaus-Peter Schmit-Neuerburg, Department of Traumatology, University of Essen, D-4300 Essen 1, Federal Republic of Germany [43]

Uwe Schmuckall, Department of Immunology, Medical School Hannover, 3000 Hannover 61, Federal Republic of Germany [507]

Johannes Schneider, Department of Pharmacology, Grünenthal GmbH, 5100 Aachen, Federal Republic of Germany [913]

Ulrich Schoeffel, Department of Surgery, University of Freiburg, 7800 Freiburg, Federal Republic of Germany [371]

Friedrich Schöndube, Department für Chirurgie, Zentralkrankenhaus, D-2800 Bremen 1, Federal Republic of Germany [695]

Robert J. Schott, Department of Surgery, The University of Michigan Medical School, Ann Arbor, MI 48109 [891]

W. Schramm, Medizinische Klinik Innenstadt, Ludwig-Maximilians Universität, 8000 Munich, Federal Republic of Germany [395]

A. Schultz, Zentrum für Anästhesie der Medizinischen, Hochschule Hannover, 3000 Hannover 51, Federal Republic of Germany [643]

Franz Schulz, Department of Surgery 1, University of Vienna Medical School, A-1090 Vienna, Austria [719]

L. Schweiberer, Chirurgische Klinik Innenstadt und Chirurgische Poliklinik, Ludwig-Maximilians-Universität München, D-8000 München 2, Federal Republic of Germany [731,763,945]

Werner Seeger, Department of Internal Medicine, Division of Clinical Pathophysiology and Experimental Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [67,119,127,283,305]

Jochen W. Seidel, Department of Immunology, Medical School Hannover, 3000 Hannover 61, Federal Republic of Germany [507]

Rainer Seitz, Division of Internal Medicine, Department of Hematology, Philipps-University, D-3550 Marburg, Federal Republic of Germany [965,971]

Mohie Sharaf El Din, Institute of Biochemistry, University of Graz, A-8010 Graz, Austria [351]

William C. Shoemaker, Department of Surgery, King-Drew Medical Center, Los Angeles, CA 90059 [133]

William J. Sibbald, Critical Care Trauma Centre, The Victoria Hospital Corporation, and the University of Western Ontario, London, Ontario N6A 4G5, Canada [1075]

Matthias Siebeck, Chirurgische Klinik Innenstadt und Chirurgische Poliklinik, University of Munich, D-8000 Munich 2, Federal Republic of Germany [395,937,945,983]

John H. Siegel, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

Rolf Edgar Silber, Chirurgische Universitätsklinik, Experimentelle Chirurgie, D-8700 Würzburg, Federal Republic of Germany [143]

A. Simmel, First Surgical University Clinic, Metabolic Research Laboratory, University Vienna, A-1090 Vienna, Austria [589]

Richard Simmons, Departments of Surgery and Biochemistry, University of Minnesota Medical School, Minneapolis, MN 55455 [563]

R. Sistermann, Department of Surgery, Berufsgenossenschaftliche Krankenanstalten "Bergmannsheil", 4630 Bochum, Federal Republic of Germany [989]

M. Spannagl, Medizinische Klinik Innenstadt, Ludwig-Maximilians Universität München, D-8000 München 2, Federal Republic of Germany [395,937]

Ron G.H. Speekenbrink, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [419]

Christian P. Speer, Department of Pediatrics, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [689]

John J. Spitzer, Department of Physiology, Louisiana State University Medical Center, New Orleans, LA 70112 [545]

Judy A. Spitzer, Department of Physiology, Louisiana State University Medical School, New Orleans, LA 70112 [575]

Paul Sporn, Ludwig Boltzmann Institute, Department of Anaesthesia and Intensive Care, Vienna University School of Medicine, A-1090 Vienna, Austria [803,809]

Roger G. Spragg, Division of Pulmonary and Critical Care Medicine, Department of Medicine, University of California, San Diego, CA 92103 [791]

Alexander Stäblein, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [625]

Jan O. Stadaas, Department of Surgery and Institute for Experimental Medical Research, Ullevaal Hospital, University of Oslo, 0407 Oslo 4, Norway [315, 389,401,873]

Karl H. Staubach, Department of Surgery, University of Luebeck, 2400 Luebeck, Federal Republic of Germany [821]

Karl Steinbereithner, Ludwig Boltzmann Institute, Department of Anaesthesia and Intensive Care, Vienna University School of Medicine, A-1090 Vienna, Austria [803,809]

T. Stober, Department of Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

Theo Strasser, Department of Surgery, LMU Munich, Klinikum Grosshadern, D-8000 Munich 70, Federal Republic of Germany [495]

Wolfgang Strohmaier, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [797]

Johannes Sturm, Unfallchirurgische Klinik, Medizinischen Hochschule Hannover, D-3000 Hannover 61, Federal Republic of Germany [507,673,775]

R. Südhoff, Department of Hematology/Oncology, Philipps-University, D-3550 Marburg/Lahn, Federal Republic of Germany [971]

Kazuro Sugi, The University of Texas Medical Branch and Shriners Burns Institute, Galveston, TX 77550 [237]

Günther Sutter, Department of Trauma, University of Saarland, D-6650 Homburg/Saar, Federal Republic of Germany [523]

Norbert Suttorp, Department of Internal Medicine, Division of Clinical Pathophysiology and Experimental Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [67,119,127,305]

Imre Szabó, Institute of Experimental Surgery, Szent-Györgyi Albert Medical University, H-6701 Szeged, Hungary [253]

Kornél Szabó, Burn Center of Central Hospital H.P.A., 1553 Budapest, Pf 1, Hungary [1101]

Ben E. Tall, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

Friedrich Tegtmeier, Department of Pediatrics, Medical School of Lübeck, D-2400 Lübeck 1, Federal Republic of Germany [689]

M. Teschner, Department of Medicine, University of Wuerzburg, D-8700 Wuerzburg, Federal Republic of Germany [757]

L.G. Thijs, Medical Intensive Care Unit, Free University Hospital, Amsterdam [163]

Martin Thurnher, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [225,231,447, 931]

D. Tirpitz, Department of Surgery I and Center for Hyperbaric Medicine, St. Joseph-Hospital, D-4100 Duisburg 12, Federal Republic of Germany [1007]

Bela Török, Department of Experimental Surgery, University of Medicine, Pécs, Hungary H-7643 [907]

Courtney M. Townsend, Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [377]

Daniel L. Traber, Shriners Burns Institute and the Departments of Surgery and Anesthesiology, The University of Texas Medical Branch, Galveston, TX 77550 [201,237,377,815,885]

Lillian Traber, Department of Anesthesiology and Surgery, The University of Texas Medical Branch, and Division of Anesthesia Research, Shriners Burns Institute, Galveston, TX 77550 [201,815,885]

F. Trautinger, Institute for Applied and Experimental Oncology, University of Vienna, A-1090 Vienna, Austria [715]

Otmar Trentz, Department of Trauma Surgery, University of Saarland, D-6650 Homburg/Saar, Federal Republic of Germany [523]

Emmanouel Tzartinoglou, Department of Surgery, University of Thessaloniki, AHEPA Hospital, Thessaloniki GR-54006, Greece [151]

Ignas P.T. van Bebber, Department of General Surgery, St. Radboud University Hospital, 6500-HB Nijmegen, The Netherlands [419,827]

Cees J. van der Linden, Department of General Surgery, University of Limburg, Biomedical Center, 6200 MD Maastricht, The Netherlands [463]

Thomas C. Vary, Departments of Surgery, Physiology, and Medicine, University of Maryland, Baltimore, MD 21201 [535]

B. v. Borman, Department of Anaesthesiology and Intensive Care Medicine, Justus-Liebig Universität, D-6300 Giessen, Federal Republic of Germany [897]

G. Vermaak, The HA Grové Research Center of the University of Pretoria, Pretoria, South Africa [217]

Jesus Villar, Intensive Care Unit, Hospital N.S. del Pino, Las Palmas, Canary Islands, Spain; present address: Mount Sinai Hospital, Toronto, Ontario M5G 1X5, Canada [57,61]

JL. Vincent, Department of Intensive Care, Erasme Hospital, Free University of Brussels, 1070 Brussels, Belgium [181]

Christa Vogl, Ludwig Boltzmann Institute for Experimental Traumatology, A-1200 Vienna, Austria [231,237,447, 931]

K. Voigt, Klinik für Anästhesiologie und operative Intensivmedizin, Klinikum Steglitz der Freien Universität Berlin, D-1000 Berlin 45, Federal Republic of Germany [751]

K. Voigt, Institut für Normale und Pathologische Physiologie, Marburg, Federal Republic of Germany [711]

Hartmut Volkmann, Department of Anaesthesiology, University of Göttingen, D-3400 Göttingen, Federal Republic of Germany [323]

Bernd-Ulrich von Specht, Department of Surgery, University of Freiburg, 7800 Freiburg, Federal Republic of Germany [371]

Reinhard Voss, Department of Internal Medicine, University of Giessen, 6300 Giessen, Federal Republic of Germany [383]

Bernd Wagener, Klinik für Anaesthesiologie und Transfusionsmedizin der Universität Tuebingen, D-7400 Tuebingen, Federal Republic of Germany [185]

H. Wallasch, Institute of Anaesthesiology, University Hospital, D-8700 Würzburg, Federal Republic of Germany [1031]

Henrik Walter, Department of Internal Medicine, Justus-Liebig University, D-6300 Giessen, Federal Republic of Germany [67]

Sten Walther, Department of Anaesthesia and Intensive Care, Lasarettet, S-601 82 Norrköping, Sweden [867]

Alfred Walz, Department of Surgery, LMU Munich, Klinikum Grosshadern, D-8000 Munich 70, Federal Republic of Germany [495]

Ch. Waydhas, Chirurgische Klinik Innenstadt, Universität München, 8000 München 2, Federal Republic of Germany [731,763]

B. Weidler, Department of Anaesthesiology and Intensive Care Medicine, Justus-Liebig Universität, D-6300 Giessen, Federal Republic of Germany [897]

Joachim Weipert, Abteilung für Klinische Chemie und Klinische Biochemie in der Chirurgie Innenstadt, Ludwig-Maximilians-Universität München, D-8000 München 2, Federal Republic of Germany [937,983]

Peter Wendt, Department of Experimental Surgery, Technical University, 8000 Munich 80, Federal Republic of Germany [339]

Karl Werdan, Department of Medicine I, Klinikum Grosshadern, University of Munich, D-8000 Munich 70, Federal Republic of Germany [247,625,1025]

C. Wesoly, Chirurgische Universitätsklinik Köln-Lindenthal, D-5000 Köln 41, Federal Republic of Germany [637]

Michael West, Departments of Surgery and Biochemistry, University of Minnesota Medical School, Minneapolis, MN 55455 [563]

Eric T. Whalley, Department of Physiological Sciences, University of Manchester, Manchester, England [983]

Michael F. Wilson, Research Service, VA Medical Center and Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City, OK 73104 [207,217,847]

Gisela Windus, Department of Forensic Medicine, University of Hannover Medical School, 3000 Hannover 61, Federal Republic of Germany [37]

Isolde Wodarz, Department of Occupational and Social Medicine, University of Tübingen, 7400 Tübingen, Federal Republic of Germany [595]

Martin Wolf, Division of Internal Medicine, Department of Hematology, Philipps-University, D-3550 Marburg, Federal Republic of Germany [965]

Wolfgang Woloszczuk, Ludwig Boltzmann Institute for Clinical Endocrinology, A-1090 Vienna, Austria [683]

Qixia Wu, Department of Pathophysiology, Peking Union Medical College, Beijing 100700, China [271]

K. Wurster, Chirurg. Klinik Innenstadt und Chirurg. Polikl. der Universität, D-8000 München 2, Democratic Republic of Germany [455]

Ernst Zadrobilek, Ludwig Boltzmann Institute, Department of Anaesthesia and Intensive Care, Vienna University School of Medicine, A-1090 Vienna, Austria [803,809]

G. Zeck-Kapp, Department of Pathology, University of Freiburg, D-7800 Freiburg, Federal Republic of Germany [113]

Gertrud Zilow, Department of Immunology, University of Heidelberg, D-6900 Heidelberg, Federal Republic of Germany [43,299]

T. Zimmermann, Surgical Department of the Medical College Dresden, Dresden, German Democratic Republic [291]

Gerald Zöch, I. Surgery, University Clinic, Burn Care Clinic, A-1090 Vienna, Austria [995]

Validity of the Elastase Assay in Intensive Care Medicine

Hermann Lang ¹⁾, Marianne Jochum ²⁾, Hans Fritz ²⁾ and
Heinz Redl ³⁾ *)

- 1) Diagnostic Research, E. Merck Darmstadt, FRG
 - 2) Department of Clinical Biochemistry, University Munich, FRG
 - 3) Ludwig Boltzmann Institute for Experimental and Clinical Traumatology, Vienna, Austria
- *) With cooperation of H. Dittmer, M. Dreher, A. Heubner, D. Inthorn, Th. Joka, D. Nast-Kolb, R. Pacher and W. Sandtner

Introduction

The role in the so called inflammatory response of the polymorphonuclear granulocyte (neutrophil) has been well established (Lang and Fritz, 1986). Excessive neutrophil stimulation in traumatic or infectious foci may cause deleterious effects to the organism, manifested by the failure of shock sensitive organs. The pathogenetic agents responsible for those actions mainly are lysosomal proteinases and oxidative molecules released by the neutrophil. Therefore, the possible use of neutrophil elastase (PMN Elastase) as a biochemical marker for pathologic granulocyte stimulation has been investigated during recent years (Jochum et. al., 1986).

In this paper, data on the validity of plasma PMN Elastase measurements as a prognostic marker for clinical complications in postoperative and posttraumatic intensive care patients are presented.

Methods

For PMN Elastase measurements, in recent years a heterogeneous sandwich ELISA assay (Neumann et. al. 1984) has been used. Now, a homogenous "Immuno Activation" (IMAC) assay is available (Figure 1). This assay can be used for routine measurements of plasma elastase (inhibitor complex) levels. The characteristics of this assay are presented on page 708. The IMAC assay has been positively correlated to the ELISA assay. Due to improved protein purification techniques,

the elastase calibrator used in the IMAC assay (in form of the elastase- α_1 -proteinase inhibitor complex) has a 3 times higher purity than the calibrator of 1984. The elastase values of the IMAC assay, therefore, are lower than the values of the ELISA assay by the factor of 3.0. All raw data of the studies presented below have been recalibrated to the new calibrator before evaluation.

1. ELISA TEST

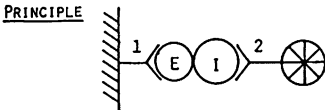
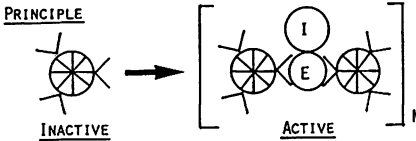


Figure 1. Assays for measurement of plasma concentrations of the elastase- α_1 -PI complex

CHARACTERISTICS

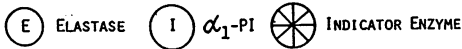
HETEROGENOUS, SOLID PHASE SANDWICH ASSAY
 TIME REQUIREMENT: 4 RESP. 2 HOURS

2. IMAC TEST ("IMMUNO ACTIVATION" TEST)



CHARACTERISTICS

HOMOGENOUS ASSAY, SUITED FOR MECHANIZED ANALYZERS
 TIME REQUIREMENT: MANUAL 25 MIN.; MECHANIZED 10 MIN.



Patients

The studies evaluated in this paper are listed in Figure 2. The results of the different studies will be published in detail by the authors separately.

STUDY CENTER	PATIENTS	NUMBER
1 INTHORN MUNICH	POSTOPERATIVE	47
2 NAST-KOLB MUNICH	POSTTRAUMATIC	33
3 JOKA ESSEN	POSTTRAUMATIC	14
4 PACHER VIENNA	POSTOPERATIVE	12
5 SANDTNER VIENNA	POSTTRAUMATIC	11
6 DITTMER FRANKFURT	POSTTRAUMATIC	8
TOTAL OF PATIENTS		125

Figure 2. Study centers and patients included in the evaluation

Results

The hypothesis to be tested was the question, whether or not plasma elastase measurements in postoperative and posttraumatic patients will yield prognostic information about eventually forthcoming complications in these patients. Daily plasma elastase values (daily mean from 1 to 6 measurements) were correlated, therefore, to the course of the disease on subsequent days.

1. Correlation to Physician's Classification

Plasma elastase values were correlated to the physicians classification, whether or not the patient had suffered complications on the subsequent days. 114 patient reports were accepted as adequately documented for the evaluation. The results of this semi-subjective evaluation are shown in Figure 3: The plasma elastase levels differentiate well between the two groups classified "Complications" vs. "No Complications" during the postoperative/posttraumatic period. A window exists from day 3 to day 7, where the elastase level has predictive quality.

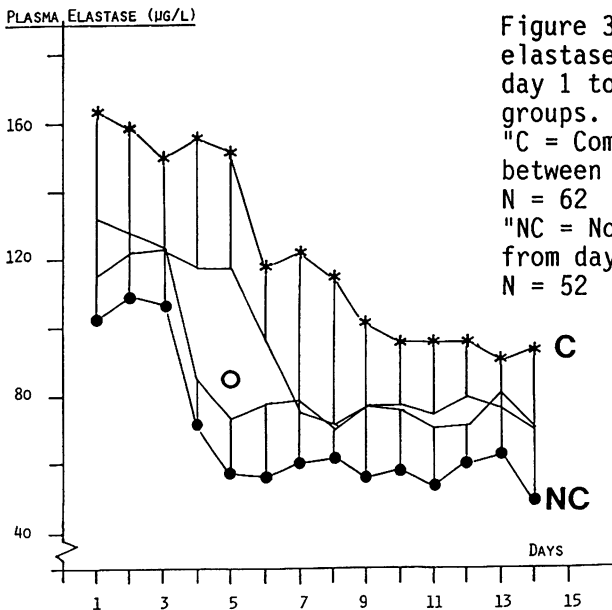


Figure 3. Mean plasma elastase levels from day 1 to 14 of patient groups.
 "C = Complications between days 6 and 12",
 N = 62
 "NC = No Complications from days 6 to 12",
 N = 52

The plasma elastase level on day 5 predicts whether or not a patient will suffer from complications within days 6 to 12, see [Figure 4](#). An elastase discrimination level of 85 $\mu\text{g/l}$ yields the following validity data: Diagnostic sensitivity = 94 %, diagnostic specificity = 79 %, predictive value of the positive test = 84 %, predictive value of the negative test = 91 %.

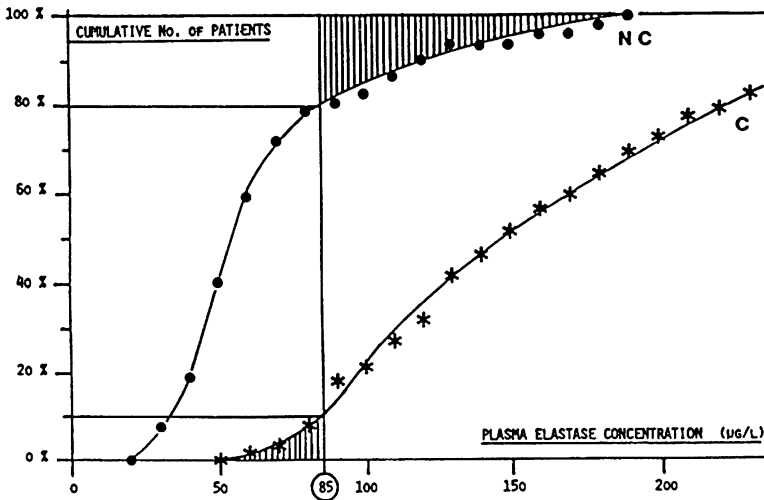


Figure 4. Discrimination on day 5 between the patient groups "Complications" (C) vs. "No Complications" (NC). Discriminatory plasma elastase level = 85 $\mu\text{g/l}$.

2. Correlation to the Multi Organ Failure (MOF) Score

To test fully objective criteria, the plasma elastase values were correlated against the MOF Score of Goris (1). In accordance to the work of Goris, a MOF Score of 5 was selected as discriminator between non critically ill vs. critically ill patients. 86 patient reports were accepted as adequately documented for the evaluation. The results are shown in [Figure 5](#). The elastase value on day 5 discriminates between the patient groups "MOF ≥ 5 on days 7 to 12" vs. MOF < 5 on days 7 to 12. An elastase discrimination level of 135 $\mu\text{g/l}$ (405 $\mu\text{g/l}$ in the sandwich assay) yields the following validity data: Diagnostic sensitivity = 83 %, diagnostic specificity = 84 %, predictive value of the positive test = 66 %, predictive value of the negative test = 93 %.

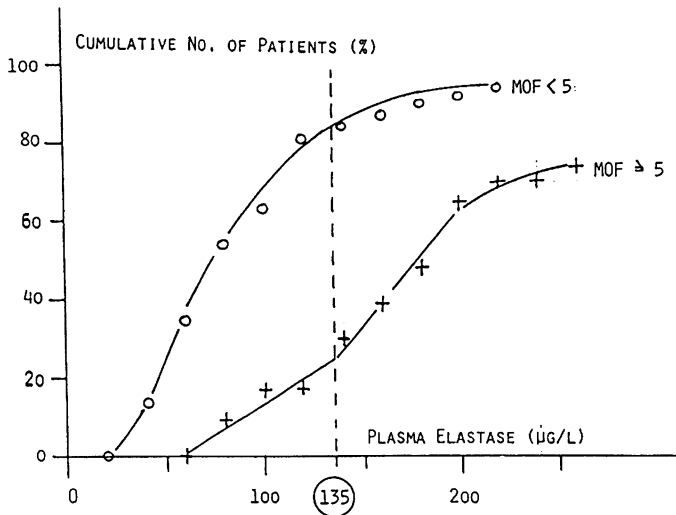


Figure 5. Discrimination between patients groups "MOF ≥ 5 on days 7 to 12" (N = 23) vs. "MOF < 5 on days 7 to 12" (N = 63). Discriminatory elastase level = 135 $\mu\text{g/l}$.

Discussion

This exploratory data analysis is being followed up by a transnational multicenter study. Nevertheless, the data presented already document the diagnostic validity of plasma elastase measurements as a prognostic marker for postoperative/posttraumatic complications. The clinician's classification of a patient's situation is a more sensitive criterium (discriminatory elastase level 85 $\mu\text{g/l}$) than the MOF Score of 5 (discriminatory elastase level 135 $\mu\text{g/l}$). Intensive care patients at risk for complications (e. g. organ failure, septicemia) can be identified from day 5 (4th postoperative/posttraumatic day) on for a period of 1 to 6 days in advance. By selecting a suitable discriminatory elastase level, a predictive value of the positive test of 90 % can be achieved. Using a discriminatory elastase level of 85 $\mu\text{g/l}$, no risk patients can equally be identified with a security of over 90 % (predictive value of the negative test).

References

Goris, R.J.A., te Boekhorst, T.P.A., Nuytinck, J.K.S., Gimbrère, J.S.F. (1985). Multiple-Organ Failure. Arch. Surg. 120: 1109 - 1115

Jochum, M., Witte, J., Duswald, K.-H., Inthorn, D., Welter, H. and Fritz, H. (1986). Pathobiochemistry of Sepsis: Role of Proteinases, Proteinase Inhibitors and Oxidizing Agents. Behring Inst. Mitt. 79: 121 - 130

Lang, H., Fritz, H. (1986). The Role of Phagocyte Proteinases in the Pathobiochemistry of Inflammation. Adv. clin. Enzymol. vol 3: 168 - 178 (Karger, Basel)

Neumann, S., Gunzer, G., Hennrich, N., Lang, H. (1984). "PMN Elastase Assay": Enzyme Immunoassay for Human Polymorphonuclear Elastase complexed with α_1 -Proteinase Inhibitor. J. Clin. Chem. Clin. Biochem. 22: 693 - 697

Index

- A23187, 121
 endothelium permeability and, 129, 131
AA-861, 131
Aasen index (PFI), polytrauma, 731–735
Acetylcholine, bradykinin antagonist B4148
 administration with, 983, 984
Acetylcholinesterase, red blood cell,
 activated C3 in DIC, fulminant
 meningococcal meningitis, 272, 274
Acetylsalicylic acid, 131, 457, 459, 461
 α_1 -Acid glycoprotein, plasma, and sepsis
 prognosis, 634, 635, 716, 717
ACTH, sepsis, 753, 755
Acute phase reactants, 337
ADH. *See* Vasopressin (antidiuretic
 hormone)
Adherence, granulocyte-endothelial cell,
 bacterial endotoxin role, 123–124
ADP
 -induced platelet aggregation, 367
 ribosylation, *Pseudomonas* endotoxin A,
 elongation factor-2, 250
Adrenal glands, animal models for shock,
 838–839
Age, antithrombin III and plasma
 substitution in septic shock, 966, 968
AH 23848, 366
Albumin, plasma, and abdominal sepsis
 prognosis, 720, 721
Allopurinol, 350
Alveolar
 and C3a in ARDS, 43–47, 52
 cell cytological changes, ARDS,
 51–54
 permeability, 323–329
 see also under Permeability
Alveolo capillary interface
 ARDS, 27–28
 corticosteroid, nebulized, in
 experimental respiratory distress,
 867
 membrane permeability, cell interactions
 in septic shock, 116–117
Ambiquitous enzymes, 576
Amino acid clearance, prognostic index in
 sepsis, 634, 635
Amino acid concentrations, serum,
 experimental endotoxin shock,
 595–599
 gabexate mesilate administration, 596,
 597
 glutamine and glutamic acid, 598
 hyperalinemia and lethality, 595,
 597–599
 tyrosine, 598
Amino acid metabolism, respiratory
 quotient (CO_2/O_2 exchange ratio) in
 shock, 619–621
Amino acid release, perfused liver, sepsis
 effect on metabolism, 590–592
21-Aminosteroid U74006F, 891–895
Anaphylactic lung reaction, guinea pig,
 WEB 2086 (PAF antagonist),
 925–929
Anaphylatoxins. *See* C3a; C5a
Angiopathy, diabetic, 1008, 1009
Angiotensin, 379
Animal model development for shock,
 835–840, 843–844, 851
 application to humans, 839–840,
 843–844
 endotoxin shock, history, 836

- methylprednisolone with gentamicin, dogs, *E. coli* shock, 836–837
 adrenal gland role, 838–839
 cf. baboons, 836–840
 rationale, 837–838
- Anipamil, traumatic shock, 1067, 1068
- Antibiotics
 burns, enteric translocation of microorganisms, 377, 378
 and calcium antagonists in endotoxin shock, 1070
- Antibodies, anti-LPS and anti-lipid A, determination with immunoblotting, 1043–1050
- Antichymotrypsin cf. C-reactive protein as prognostic index, 725–727
- Antigen, inhaled, WEB 2086 (PAF antagonist), anaphylactic lung reaction, 926, 928
- Antigen-presenting cells, trauma-induced cascade of CMI effects, 495, 496
- Antioxidant, MTDQ-DA, myocardial ischemia, 907–911; *see also* Free radical scavengers; Oxygen radicals
- Antiplasmin
 kallikrein-kinin system components in ARDS after polytrauma, 738, 741
 methylprednisolone pretreatment, endotoxemia, 874, 875
- α_2 -Antiplasmin
 aprotinin membrane protective action, intraoperative histamine liberation, 961, 963
 plasma, and abdominal sepsis prognosis, 720, 721
- Anti-protease. *See* Protease inhibitor *entries*
- Antithrombin III, 319, 384, 386, 940
 endotoxin and overwhelming inflammatory response of early sepsis, 372
 kallikrein-kinin system components in ARDS after polytrauma, 738, 740–742
 methylprednisolone pretreatment, endotoxemia, 874, 875
 multiple system organ failure, postoperative/posttrauma, biochemical analysis and scoring, 652
 neutrophils, thrombin-induced adhesion with endothelial cells, 103
- plasma
 and abdominal sepsis prognosis, 720
 corticosteroid, nebulized, in experimental respiratory distress, 869–871
 endotoxin-induced DIC, AT III-heparin complex therapy, 979
 plasma substitution in septic shock, humans, 965–968
 prognostic value in sepsis, 637
 lethality, 639, 640
 thrombocyte counts, 637–641
 scintigraphic evaluation of posttraumatic liver function, 776, 777
- Antithrombin III-heparin complex, DIC, endotoxin-induced, 977–981
- α_1 -Antitrypsin
 immune suppression, post-surgical or post-traumatic, 492
 lymphocyte/monocyte ratio in polytrauma survival, 769
 plasma, and abdominal sepsis prognosis, 716, 717, 720, 721
- Aorta, vascular intima in endotoxin shock, 85, 86
- Apache II scoring system, 280, 384, 386, 626–630, 635, 643, 645, 664, 716, 1026–1030
- Aprotinin
 immunoglobulin profiles and PMN-elastase in septic gas gangrene, 1008, 1012, 1014–1016
 membrane protective action, intraoperative histamine liberation, 959–963
- Arachidonate/arachidonic acid metabolism, 21, 351
 in ARDS, pulmonary edema, 306–307, 309–311
 burns, PAF inhibitor effect, scalded pig, 455, 461
 cyclooxygenase metabolites, lung injury in *E. coli* endotoxemia, 358, 362–364
 early ventilatory support, 785
 endotoxin effects, endothelium, 119–121, 121–123

- hemofiltration and survival time, porcine
acute endotoxic shock, 823–825
myocardial ischemia, 911
see also specific metabolites
- ARDS, 7, 8, 13–14, 17–23, 27–33, 787,
788
- alveoli, 31–33, 43–47, 51–54
- alveolocapillary membrane damage,
27–28
- arachidonic acid cascade in, and
pulmonary edema, 306–307,
309–311
- bronchoalveolar lavage, 13, 19, 20, 22,
23
- C3a, 31–33, 43–47, 52, 299, 880
causes, 17–18
air-borne cf. blood-borne, 18, 19
- ceruloplasmin changes, 331–337
C-reactive protein, 335, 336
number of organs involved, 333,
334, 336
- complement activation, 43
pulmonary hypertension and vascular
leakage, 283, 284, 286
- diagnostic markers, 22–23
- dopamine infusion, effect evaluation
with systolic time intervals, 1102
- early cf. late, 18
- exogenous surfactant, 791–795
- exudate organization, 27–29
- hypovolemic shock, 32, 33
- ischemia and circulatory system in
MSOF, 1076, 1078, 1081, 1083
nifedipine, 1087–1090
pathogenesis, 19–22
pathophysiology, 27–28
phospholipase A as prognostic index,
757
- PMN degranulation, 18–20, 31–33
- polytrauma, kallikrein-kinin system,
737–742
- postoperative organ failure, 133–136
- posttraumatic, plasma levels of
mediators, prognosis, 673–679
macrophages, 679
cf. non-ARDS, 676, 677, 679
- septic, and multiple system organ
failure, 57–60
prognosis, 59, 61–65
- clinical conditions, 62
mortality, 63, 65
- septicemia, pulmonary vascular
resistance, 175–179
- septic shock, 30–33
corticosteroids in, 857, 860–864
metabolic abnormality, 535, 537,
538
oxygen supply-uptake relationship,
181
- shock as predisposing factor, 64
surfactant replacement, 29, 797
- Arrhythmias, myocardial ischemia, 907
- Arterial pressure, mean
 α -mercaptopropionylglycine in
hemorrhagic shock, 899–902
RA642, effects on cerebrocortical
perfusion, acute hemorrhagic
shock, 1091–1093
see also Pulmonary artery pressure
- Arthroplasty, total hip, high-dose
corticosteroids to prevent C3a and
C5a formation, 879–882
- Artifact rejection, heart performance during
septic shock, awake sheep, 239–242
- Aspiration trauma, experimental, exogenous
surfactant, 797–800
- Asthma, 486
- ATPase, calcium antagonists in
shock/ischemia, 1065, 1066
- ATP MgCl₂, kidney metabolism in *E. coli*
sepsis, 602, 605
- Atrial pressure, left, pulmonary venous
hemodynamics and gas exchange
disturbances, *E. coli* septicemia,
Gottingen pig, 185, 186
- Autoradiography, in situ, hepatocyte protein
kinase C and diacylglycerol
accumulation, endotoxemia, 579,
583–584, 586
- Autoregulation, hemorrhagic shock, phase-
related vascular reactivity, cats, 147,
148
- B4148, blood pressure maintenance,
endotoxin shock, 983–987
- Bacterial toxins, endothelium permeability
effects, 127–131; *see also specific
bacteria and toxins*

1110 / Index

- Bacterioides fragilis*, 538
- Base excess, respiratory quotient (CO₂/O₂ exchange ratio) in shock, 613, 614, 619
- B cells(s)
- Ig synthesis suppression after multiple trauma, 513–515
 - maturation, trauma-induced cascade of cell-mediated immune effects, 496, 497, 499, 501, 502
 - multiple trauma, early events, 507–509
- B-cell growth factor (IL-4), trauma-induced cascade of cell-mediated immune effects, 497
- Beclometasondipropionate (BDP), nebulized corticosteroid, in experimental respiratory distress, 867, 868, 870, 871
- Bilirubin
- ARDS, posttraumatic, prognosis, 675, 676, 678
 - polytrauma, 743, 745
 - lymphocyte/monocyte ratio in survival, 769
- Biologically active site, C3a, 299
- Biological response modifiers. *See* Immunomodulation
- Blood flow redistribution, septic shock, 164
- Blood volume redistribution, septic shock, 164–165
- BM 13,177, 366, 367
- BN 56,020, 428
- BN 52,021, 426–433, 442–444, 485–488
 - endotoxin shock effects, 931–935
 - PAF effects in sheep, 448
- BN 52,022, 428
- BN 56,203, 429
- Bombesin, 378–380
- Bone, long, fracture, and pulmonary fat embolism, 39–41
- Bradykinin antagonist B4148, blood pressure maintenance, endotoxin shock, 983–987
- Brain, PAF antagonist inhibition of induced shock, 430
- Branched chain amino acids, 617
 - amino acid concentrations, serum, experimental endotoxin shock, 595, 598
 - metabolic abnormalities in sepsis, 539, 540
- Bronchoalveolar lavage, ARDS, 13, 19, 20, 22, 23, 44
 - exogenous surfactant, 795
- Bronchoconstriction, WEB 2086 (PAF antagonist), anaphylactic lung reaction, 925, 926, 928
- Burimamide, 362
- Burn(s)
- dopamine infusion, effect evaluation
 - with systolic time intervals, 1101–1105
 - total burn surface estimation with Nine Rule, 1102–1103
 - endotoxin and overwhelming inflammatory response of early sepsis, 372–374
 - enteric translocation of microorganisms, 377–380
 - scalded rats, 379
 - sheep, 378
 - PAF antagonist inhibition of induced shock, 431
 - scalded pig, 455–461
 - sepsis after, TP-5 immunomodulation, 995–998
- Butyrophenones, 755
- BW 755C, glucose turnover in sepsis, 552, 553
- C3
- activated, DIC in septic shock, 271–274
 - IgG breakdown in peritonitis exudate, 527–532
 - lymphocyte/monocyte ratio in polytrauma survival, 769, 770
 - plasma, and abdominal sepsis prognosis, 720, 721
- C3a, 7, 204
- activation
 - endotoxin role in sepsis, 278–280
 - MSOF pathogenesis in septic shock, dogs, 296
 - pulmonary hypertension and vascular leakage, 283
 - alveoli in, post-trauma, 43–47, 52
 - ARDS, 299, 880
 - biologically active site, 299
 - circulation, peripheral, septic shock, 170

- corticosteroids to prevent formation,
total hip arthroplasty, 879–882
- ELISA quantitation with monoclonal
antibodies, 299–303
- cf. C3, 300–303
- endotoxin and overwhelming
inflammatory response of
early sepsis, 372, 373
- in MSOF, 296, 880
- acute pancreatitis, 265–268
- polytrauma, 414
- postoperative/posttrauma,
biochemical analysis and
scoring, 650, 651
- prognostic index in sepsis, 635
- rheumatoid arthritis, 299
- systemic lupus erythematosus, 299
- C3c, complement activation, MSOF
pathogenesis in septic shock, dogs,
292, 294–296
- C4
complement activation, endotoxin role in
sepsis, 279, 280
- lymphocyte/monocyte ratio in
polytrauma survival, 769, 770
- plasma, and abdominal sepsis prognosis,
720, 721
- C5a, 7, 13, 43, 44, 46, 115, 116, 204, 825
- activation
endotoxin role in sepsis, 278–280
- MSOF pathogenesis in septic shock,
dogs, 292, 296
- pulmonary hypertension and vascular
leakage, 283, 284
- corticosteroids to prevent formation,
total hip arthroplasty, 879–882
- MSOF in acute pancreatitis, 265–268
- terminal complement complexes,
266, 267
- and PAF, 366
- C5b9, 266
- Cachectin. *See* Tumor necrosis factor (TNF,
cachetin) *entries*
- Calcium
capillary endothelial cells, 159
- endothelium permeability effects, 127–
131
- endotoxin-induced intracellular overload,
1053–1061
- hepatocytes, protein kinase C and
diacylglycerol accumulation,
endotoxemia, 575, 578, 584
- isolated heart, effect of LMW plasma
fraction in hypovolemic traumatic
shock, 234
- overload in ischemic cell death,
1065–1066
- and PAF, 366
- rapid influx and toxic action, 68
- Calcium antagonists in shock/ischemia,
1060, 1065–1073
- ATPase, 1065, 1066
- calcium overload in ischemic cell death,
1065–1066
- endotoxin shock, 1068–1071
- glucose deficiency, 1066
- hemorrhagic shock, 1067
- mechanisms of action, 1072
- magnesium, 1072
- mitochondrial function, 1066
- muscle spasm, smooth, 1066
- phospholipase, 1066, 1071
- platelet aggregation, 1066
- traumatic shock, 1067–1068
- see also specific agents*
- Calcium gates, 119, 120
- Calmodulin, 120
- Calvin, John, 850
- Cancer patients, 493–494
- Candida tropicalis*, in vitro phagocytosis
assay, 508, 510
- Candidiasis, systemic, 378
- Capillary endothelium
contractile elements, 157–160
- endotoxemia and shock, impaired
regulation, 157–160
- lung, complement activation, MSOF
pathogenesis in septic shock,
dogs, 292, 294–296
- Capillary permeability. *See under*
Permeability
- Capillary pressure, lung edema, 28
- Capillary surface area reduction, ischemia
and circulatory system in MSOF,
1082
- Carbon dioxide, arterial, metabolic
abnormalities in sepsis, 537, 538; *see*
also Respiratory quotient (CO₂/O₂
exchange ratio) in shock
- Carbon monoxide, α -

- mercaptpropionylglycine in hemorrhagic shock, 899, 900, 902
- Cardiac function
 - heart cells, cultured, effects of
 - Pseudomonas aeruginosa* toxins, rat, 247–251
 - muscle pO₂ role, critically ill patients, 139
 - see also* Heart entries
- Cardiac index, 625, 626
 - extravascular lung water
 - altered fluid regimen, advanced septic shock with acute respiratory failure, 804–806
 - large volume replacement with crystalloids, 810, 811
 - hydroxyethyl starch, volume replacement in ovine endotoxemia, 818, 819
 - immunoglobulin and plasmapheresis therapy, hemodynamic effects during treatment for septic shock, 1026, 1027
 - pulmonary vascular resistance, ARDS in septicemia, 175–179
 - pulmonary venous hemodynamics and gas exchange disturbances, *E. coli* septicemia, Goettingen pig, 185, 186
- Cardiac output
 - cardiopulmonary response to endotoxin, eicosanoids in, sheep, 202
 - circulation, peripheral, septic shock, 163–166
 - hemofiltration and survival time, porcine acute endotoxic shock, 822, 825
 - ischemia and circulatory system in MSOF, 1076–1077, 1080, 1082
 - oxygen supply-uptake relationship, septic shock, 183
 - PAF effects in sheep, 448–449
 - septic shock, chacma baboon (*Papio ursinus*), 207, 210–215, 219–222
- Cardiac work, septic shock, heart overperformance in, 260–262
- Cardiogenic shock, scoring systems, 625–630
 - CI and SVR, 625, 626
- Cardiopulmonary arrest, lipid peroxidation inhibition, 891–895
- Cardiopulmonary response to endotoxin
 - eicosanoids in, sheep, 201–204
 - free radical scavengers, 885–888
- Catalase, 201–202, 887
- Catecholamines, 755, 756
 - glucose turnover in sepsis, 552–553, 557, 559
 - ischemia and circulatory system in MSOF, 1077
 - see also* specific catecholamines
- Cathepsin D, 431, 168
- Cathepsin G, 937, 941, 942
- Cement, methylmethacrylate, 879, 880
- Cerebrocortical perfusion, RA642 effects, hemorrhagic shock, 1091–1094
- Ceruloplasmin, changes in MSOF and ARDS, 331–337
 - C-reactive protein, 335, 336
 - number of organs involved, 333, 334, 336
- C₁-esterase inhibitor
 - circulation, peripheral, septic shock, 170–171
 - complement activation, endotoxin role in sepsis, 278, 280
 - lymphocyte/monocyte ratio in polytrauma survival, 769
 - plasma, and abdominal sepsis prognosis, 720
- Chemiluminescence-inducing radicals, 339–344; *see also* Oxygen radicals
- Chemotactic factors, alveolar macrophages, 20
- Chemotaxis, 4-hydroxy-nonenal, 351
- Chloramphenicol, 852
- Cholecystectomy, 493
- Cholecystokinin, 380
- Cholinesterase, scintigraphic evaluation of posttraumatic liver function, 777
- Chromatography
 - HPLC, PAF effects in sheep, 448–451
 - ion exchange, positive inotropic factor as myocardial stimulant, 254–255
- Chronotropic effect, negative, α -mercaptpropionylglycine in hemorrhagic shock, 902
- Chymase, mast cell, 937, 941, 942
- Cimetidine, 755
- Circulating immune complexes, and abdominal sepsis prognosis, 716
- Circulation, peripheral, septic shock, 163–171

- blood flow redistribution, 164
- blood volume redistribution, 164–165
- cardiac output, 163–166
- oxygen utilization, 163, 167–169
- pathogenesis, 169–171
- peripheral vascular failure, 165–167
- permeability, microvascular, 167
- systemic vascular resistance, 163–166, 168, 170
- Clostridium*, 1008, 1020
 - sordelli*, 1018
- Clotting factor infusion, MSOF prognostic indices, logistic regression analysis, 644
- Coagulation cascade, 31, 318–319
 - fibrinolysis, and kallikrein system, MSOF, postoperative/posttrauma, biochemical analysis and scoring, 652–654
- lung, 6
 - microvascular bed, cell interactions in septic shock, 115
- methylprednisolone pretreatment effects in porcine *E. coli* endotoxemia, 873
- pulmonary vascular permeability, 305–307
- septic patients, 383–387
 - see also* Disseminated intravascular coagulation; *specific components*
- Collagen III propeptide, 665
 - ARDS, posttraumatic, prognosis, 674–679
- Colloid osmotic pressure, plasma, extravascular lung water, large volume replacement with crystalloids, 810–812
- Compensated shock, hemorrhagic shock, phase-related vascular reactivity, cats, 144–148
- Complement, 31
 - ARDS, 43
 - cascade
 - lung, microvascular bed, cell interactions in septic shock, 115, 116
 - MSOF, postoperative/posttrauma, biochemical analysis and scoring, 650–651
 - and endotoxin role in sepsis, 277–281
- Igs, therapeutic, phagocytosis stimulation, peritonitis, 1039, 1041
 - and immune suppression, post-surgical or post-traumatic, 492
- lung in shock, 4, 5, 7, 13
 - hypertension and vascular leakage, rabbit, 283–286
 - vascular permeability, 305, 306
- MSOF pathogenesis, septic shock, dogs, 291–296
- pathway, classical cf. alternative, MSOF pathogenesis in septic shock, dogs, 293
- PMNs, 20
 - prognostic index in sepsis, 633, 634
 - septic shock, 169–170
 - see also specific components*
- Complement complexes, terminal acute pancreatitis, 266, 267
 - pulmonary hypertension and vascular leakage, 283–286
- Contractile elements, capillary endothelial cells, 157–160
- Contractility
 - and cardiac function, *Pseudomonas aeruginosa* toxins, 247, 248
 - heart performance during septic shock, awake sheep, 238
- Coronary artery disease, 1079, 1080
- Corticosteroid(s)
 - nebulized, in experimental respiratory distress, 867–871
 - aveolo-capillary interface, 867
 - antithrombin III, plasma, 869–871
 - beclometasondipropionate (BDP), 867, 868, 870, 871
 - crystalloid infusion, 868
 - superoxide, PMN production, 869, 870
 - for septic shock with ARDS, 857, 860–864
 - dexamethasone, 861
 - methylprednisolone, 861–863
 - severity of underlying disease, 858
- VA comparative study of severe sepsis, 840–844, 847–855, 862–863
 - encephalopathy, 841, 843, 853–855
 - entry criteria, 853
 - inflammation pathways, 850–851
 - mortality, 842

- cf. previous trials, 852
- rationale of therapy, 851, 852
- sepsis criteria, 841
- trial design, 852–853
- see also specific drugs*
- Cortisol, sepsis, 753, 755
- C reactive protein, 665
 - ceruloplasmin changes in MSOF and ARDS, 335, 336
 - immune suppression, post-surgical or post-traumatic, 492
 - lymphocyte/monocyte ratio in polytrauma survival, 769
 - plasma, and abdominal sepsis prognosis, 716, 717
- Creatinine
 - dopamine and renal function, 1098
 - polytrauma, 743, 745
- Crossed immunoelectrophoresis, C3 and IgG breakdown in peritonitis exudate, 529, 531
- Crystalloids
 - corticosteroid, nebulized, in experimental respiratory distress, 868
 - cf. hydroxyethyl starch, volume replacement in ovine endotoxemia, 815–819
 - large volume replacement with, EVLW in septic shock, 809–813
- c-sis*, neutrophils, thrombin-induced adhesion with endothelial cells, 109
- CTP:phosphocholine citidyl transferase, 576
- Cutaneous thermal injury. *See* Burn(s)
- CV-3988, 428, 430, 442, 485
- CV-6209, 426
- Cyclic nucleotides, lung, endotoxin-induced microvascular endothelial injury, 95, 97
- Cycloheximide, 122
 - liver, perfused, sepsis effect on metabolism, 590, 591
- Cyclooxygenase inhibition
 - cardiopulmonary response to endotoxin, eicosanoids in, 202, 203
 - lung injury in *E. coli* endotoxemia, 365–366
- Cysts, honeycomb, 29, 32
- Cytotoxic processes, effects, platelet activating factor antagonists, 443–444
- Cytosan, 377
- Dazoxiben, 365
- D-Dimer, fibrinolysis, 383–386
- Decompensated shock, hemorrhagic shock, phase-related vascular reactivity, cats, 145–148
- Decontamination, GI tract, multiple organ failure prevention, zymosan-induced peritonitis, 827–832
- Delayed-type hypersensitivity
 - Ig prophylactic therapy after cardiac surgery, 1031–1033, 1035
 - T-cell-mediated immune suppression after polytrauma, 518, 519
- 2-Deoxyglucose tracer, metabolic abnormalities in sepsis, 549–551
- Dexamethasone, 121, 122
 - corticosteroids for septic shock with ARDS, 861
- Dextran sulfate, 653
- Diabetic angiopathy, 1008, 1009
- Diabetic microangiopathy, 1009
- Diacylglycerol accumulation, hepatocytes, endotoxemia, rats, 575–586
 - in situ receptor autoradiography, 579, 583–584, 586
 - phorbol ester binding sites, 583–584
- Dialysis, MSOF prognostic indices, logistic regression analysis, 644, 645
- Dichloroacetic acid therapy, metabolic abnormalities in sepsis, 540, 541
- Diiodotyrosine (DIT), sepsis, 752–754, 756
 - leucocyte phagocytic activity, marker for, 711–713
- Diisofluorophosphate- α -thrombin, neutrophils, cf. thrombin-induced adhesion with endothelial cells, 103, 104
- Diltiazem and endotoxin-induced intracellular Ca^{2+} overload, 1053–1061
 - epinephrine, 1057–1059
 - hepatocytes, cytosolic Ca^{2+} in, 1055–1060
 - muscle, skeletal, 1054–1061
- Dimethylthiourea, 887
- Dipyrimadole (RA8), effects on

- cerebrocortical perfusion, acute hemorrhagic shock, 1091–1094
- Disseminated intravascular coagulation, 383, 386
- endotoxin, dose-related effects on RES, 410
- endotoxin-induced, antithrombin III-heparin complex, 977–981
- fibrinolysis syndrome, 971, 972, 974
- reticuloendothelial stimulation to protect against, 1001–1005
- septic ARDS, 59
- and multiple system organ failure, 62, 63
- in septic shock, activated C3, 271–274
- TNF, induction of organ changes in chronic lymph fistula, sheep, 479, 480
- Dopamine
- kidney function, 1097–1099
- long-term administration and tolerance, 1097–1099
- hemodynamics, 1098
- cf. RA642, effects on cerebrocortical perfusion, acute hemorrhagic shock, 1091–1094
- sepsis, 755
- systolic time interval evaluation, 1101–1105
- Doppler flowmetry, laser RA642 effects on cerebrocortical perfusion, acute hemorrhagic shock, 1091–1094
- DPPC:egg PG, surfactant, exogenous, 798
- DPPC in surfactant, C3a and alveoli in, post-trauma ARDS, 44, 45
- DTPA and gamma-scintillation, alveolar permeability increased by PMA-stimulated neutrophils, rabbits, ARDS, 324–327
- Edema, lung. *See* Pulmonary edema
- Eglin C
- endotoxin shock, ineffectiveness in, 953–957
- pulmonary vascular permeability, 311
- septic shock, 945–948
- Eglin C/hirudin, recombinant, proteinase/protease inhibitor therapy, 937–942
- Eicosanoids
- cardiopulmonary response to endotoxin, sheep, 201–204
- pulmonary vascular permeability, mediators, 305, 306, 308, 312
- see also* Arachidonate/arachidonic acid metabolism; *specific eicosanoids*
- Elastase- α_1 -antiproteinase complex, 7, 8
- with antithrombin (EAT), humans, 971–974
- early indicator of pediatric systemic infection, 689–693
- endotoxin and overwhelming inflammatory response of early sepsis, 372, 373
- immunoassay, automated homogenous enzyme immunoassay, 707–710
- cf. coated tube ELISA, 708–710
- intensive care unit assay, validity, 701–705
- correlation to MOF score, 704–705
- correlation to physician's classification, 703–704
- ELISA, 701–702
- IMAC assay, 702
- kallikrein-kinin system components in ARDS after polytrauma, 737, 738, 741, 742
- marker for perioperative infection risk monitoring, validity of ELISA, 695–700
- total leukocyte counts, 699, 700
- mediation of pulmonary vascular permeability, 308–310
- MSOF, polytrauma, 414, 656–659
- and neopterin, plasma levels in MSOF, 683–687
- and prognostic index in sepsis, 634
- TNF, induction of organ changes in chronic lymph fistula, sheep, 470, 477–478, 480
- see also* PMN elastase
- Elebute and Stoner Sepsis Score, 626–630, 638, 664, 1026, 1027, 1029
- Electrical stimulation, efferent, isolated intestinal vascular bed, hemorrhagic shock, phase-related vascular reactivity, cats, 145
- ELISA, 695–700
- coated-tube, 708–710

- proteinase inhibitor, leukocyte neutral, 945–946
- quantitation with monoclonal antibodies, C3a, 299–303
 - cf. C3, 300–303
- Elongation factor-2, ADP ribosylation, *Pseudomonas* endotoxin A, 250
- Embolism, fat, pulmonary, 10, 37–41
 - Hannover Polytrauma Score, 38
 - histologic appearance, 40
 - Injury Severity Score, 38
 - long-bone or pelvis fracture, 39–41
 - respiratory failure, 37, 41
- Encephalopathy, corticosteroids (glucocorticoid), VA comparative study of severe sepsis, 841, 843, 853–855
- Endobulin, 1048, 1050
- Endocrine secretion patterns, sepsis, 751–756
 - ADH, 751, 753, 755, 756
 - prolactin, 751, 753–754, 756
 - thyroid hormones, 751–756
- Endothelial cells, thrombin-induced
 - adhesion with neutrophils, 101–109
- Endothelial injury, endotoxin-induced, 91–97
 - grading, 84–85
- Endothelial proliferation inhibiting capacity, endotoxin and overwhelming inflammatory response of early sepsis, 373, 374
- Endothelial swelling, lung in shock, 8, 9, 11
- Endothelium
 - arachidonate/arachidonic acid metabolism, 119–123
 - permeability in vitro, bacterial toxins and calcium effects, 127–131
 - see also Vascular intima in endotoxin shock
- Endothelium-derived relaxing factors, 157
- Endotoxemia
 - HES volume replacement, 815–819
 - lung injury in, 357–368
 - failure, 12
 - phases I-III, 885–888
 - oxygen free radicals, 886–888
 - recombinant human SOD in, 913–917
 - and shock, impaired regulation, capillary endothelial cells, 157–160
- Endotoxin, 850
 - antithrombin III-heparin complex, DIC, 977–981
 - arachidonate/arachidonic acid metabolism, 119–123
 - complement activation, 277–281
 - MSOF pathogenesis in septic shock, dogs, 291
 - effect, healthy volunteers, heart dysfunction cf. septic shock, 196–197
 - free radical scavengers and cardiopulmonary response, 885–888
 - granulocyte effects, lung, microvascular bed, cell interactions in septic shock, 114–116, 118
 - granulocyte-endothelial cell adherence, 123–124
 - heart cells, cultured, effects of *Pseudomonas aeruginosa* toxins, rat, 247–251
 - inflammatory reaction, GI tract decontamination, MOF prevention, 832
 - inotropic effect in isolated rabbit heart, 225–230
 - oxygen delivery, 227
 - perfusion circuit, 226, 227
 - ventricular pressure cf. perfusion flows, 227, 228
 - metabolic abnormalities in sepsis, 550
 - microvascular endothelial injury, 91–97
 - MSOF, postoperative/posttrauma, biochemical analysis and scoring, 654–655
 - antibody levels, 645–655
 - no role in MSOF, 419–423
 - and overwhelming inflammatory response of early sepsis, 371–375
 - endothelial proliferation inhibiting capacity, 373, 374
 - oxygen supply-uptake relationship, septic shock, 181–183
 - plasma concentrations related to responses, pig, 389–393

- plasma contact system factors, in vitro interactions, 401–405
 - arterial O₂ tension, 391–392
 - hemodynamics, 390–391
 - kallikrein-kinin system, 389, 390, 392, 393
- prognostic value in sepsis, 634, 635, 637
 - lethality, 639, 640
 - thrombocyte counts, 637–641
- proteases in MSOF due to septicemia, 315–321
- renal microthrombosis, 916
- reticuloendothelial system, dose-related effects, 407–411
- structure and biological activity, 79–81 and TNF, 463, 464
 - see also* Antibodies, anti-LPS and anti-lipid A, determination with immunoblotting; *specific bacteria*
- Endotoxin shock
 - amino acid concentrations, serum, 595–599
 - animal model development for shock, 836
 - calcium antagonists in shock/ischemia, 1068–1071
 - eglin C ineffectiveness, 953–957
 - fibrinolytic functional determinants, pig, 395–399
 - hemofiltration and survival time, 821–826
 - hirudin/eglin C, recombinant, 937–942
 - PAF antagonists, 428–429, 931–935
 - proteinase/protease inhibitor therapy, 937–942
 - cf. traumatic shock, 941
 - vascular intima in, 77–87
 - WEB 2086 (PAF antagonist), cf. in anaphylactic lung reaction, 925, 927, 928
- Enteric translocation of microorganisms, burns, 377–380
 - scalded rats, 379
 - sheep, 378
- Enterobacteriaceae, 827–832
- Eosinophil cationic protein, 22
- Epidermal growth factor, 380
- Epinephrine
 - diltiazem and endotoxin-induced intracellular Ca²⁺ overload, 1057–1059
 - metabolic abnormalities in sepsis, 54, 548
- Epithelial lining fluid, ARDS, C3a and alveoli in, post-trauma, 44
- Escherichia coli*, 420, 426, 429, 538, 539, 546, 547, 551, 557, 561, 723, 881, 940
 - animal models for shock, 836–837
 - antibodies, anti-LPS and anti-lipid A, determination with immunoblotting, 1043–1049
 - endotoxin, activated C3 in DIC in septic shock, fulminant meningococcal meningitis, 271, 273
 - hemolysin injury, septicemia in, lung, 67–70
 - transmembrane pores, 69
 - sepsis, kidney metabolism in, 601–605
 - see also* Endotoxemia; Septicemia; Septic shock
- Expired minute volume, respiratory quotient (CO₂/O₂ exchange ratio) in shock, 613, 614, 619–621
- Extravascular lung water (EVLW)
 - altered fluid regimen, advanced septic shock with acute respiratory failure, 803–808
 - cardiac index, 804–806
 - microvascular integrity, 806
 - plasma colloid osmotic pressure, 803–808
 - pulmonary artery pressure, mean, 804
 - pulmonary artery wedge pressure, 803–808
- ARDS, posttraumatic, prognosis, 675 and chemiluminescence-inducing radicals, porcine septic shock, 340, 342
 - hemofiltration and survival time, porcine acute endotoxic shock, 822, 823, 825
 - large volume replacement with crystalloids, septic shock, 809–813

- MSOF, polytrauma, 414, 415
 recombinant hirudin/eglin C, endotoxin shock, 938, 939, 941
- Factor XII (Hageman factor), 384
 and endotoxin
 interactions with plasma contact system factors, 402, 404
 overwhelming inflammatory response of early sepsis, 372
 kallikrein-kinin system components in ARDS after polytrauma, 737, 738, 741
- Factor XIIa, circulation, peripheral, septic shock, 171
- Fat embolism after bone fracture, 10, 37–41
- Fat metabolism
 respiratory quotient (CO_2/O_2 exchange ratio) in shock, 619–621
 in sepsis, 536–539, 545
- Fentanyl, 755
- Fibrin
 endotoxic shock, fibrinolytic functional determinations, pig, 395, 399
 lung organ failure, 10, 11
- Fibrinogen
 consumption, recombinant hirudin/eglin C, endotoxin shock, 937–942
 endotoxic shock, fibrinolytic functional determinations, pig, 395–399
 plasma
 endotoxin-induced DIC, AT III-heparin complex therapy, 979, 980
 recombinant human SOD in endotoxemia, 915, 917
 RES stimulation to protect against DIC, 1002, 1005
 scintigraphic evaluation of posttraumatic liver function, 776
- Fibrinolysis
 aprotinin membrane protective action, intraoperative histamine liberation, 961
 cascade, 318
 lung, microvascular bed, cell interactions in septic shock, 115
 functional determinants, pig, endotoxic shock, 395–399
 kallikrein-kinin system components in ARDS after polytrauma, 737
 methylprednisolone pretreatment effects in porcine *E. coli* endotoxemia, 873, 877
 pulmonary vascular permeability, 305–307
 septic patients, 383–387
 syndrome, DIC, 971, 972, 974
 tests for, 383–386
- Fibrinopeptide A, endotoxin and overwhelming inflammatory response of early sepsis, 372, 373
- Fibrinopeptides, specific, and proteinase inhibitor complex, immunologic determination, humans, 971, 972, 974
- Fibrin split products, 395, 654
- Fibronectin, 886
 capillary endothelial cells, 158
 endotoxin and overwhelming inflammatory response of early sepsis, 372
 plasma, and abdominal sepsis prognosis, 716, 720, 722, 723
- Fluid substitution, hemorrhagic shock, phase-related vascular reactivity, cats, 147–148
- Fluorescent products, lipid peroxidation, hypovolemic-traumatic shock, dogs, 345–349
- Flurbiprofen, 365
- FMLP, 508, 510
- Free radical scavengers
 C3 and IgG breakdown in peritonitis exudate, 527
 cardiopulmonary response to endotoxin, sheep, 885–888
 α -mercaptopyrionyl glycine in hemorrhagic shock, 897–903
- Fructose infusion, kidney function in sepsis, 603
- Gabexate mesilate, 309
 amino acid concentrations, serum, experimental endotoxin shock, 596, 597
- D-Galactosamine, 1070, 1071
- Gamma-scintillation with DTPA, alveolar permeability, 324–327

- Gangrene, gas. *See* Immunoglobulin profiles and PMN-elastase in septic gas gangrene
- Gas exchange
 alveolar permeability increased by PMA-stimulated neutrophils, rabbits, ARDS, 326, 327
 hemofiltration and survival time, porcine acute endotoxic shock, 823, 825
 septicemia (*E. coli*), Goettingen pig, 185–188
- Gas gangrene. *See* Immunoglobulin profiles and PMN-elastase in septic gas gangrene
- Gastric mucosa ulceration
 ultrastructure after septic shock, rat, 151–155
 irreversible changes, 154–155
 parietal cells, 152, 153
 stress ulcer diseases, correlation with, 151
 surface epithelial cells, 153, 154
 WEB 2086 (PAF antagonist) and, 919
- Gastrin, 380
- Gastrointestinal tract
 decontamination, 827–832
 endotoxin-induced damage, and WEB 2086 (PAF antagonist), 919–922
 PAF antagonist inhibition of induced shock, 430
- Gel filtration, positive inotropic factor as myocardial stimulant, 254
- Gentamicin and methylprednisolone, animal models for shock, 836–840
- Glasgow Coma Scale, 644–646
- Global Index, polytrauma, 744–746
- Glucagon, glucose turnover in sepsis, 552–553, 556, 559
- Glucocorticoids, glucose turnover in sepsis, 552–553, 559
- Glucose
 concentrations, kidney metabolism in *E. coli* sepsis, 602, 604
 deficiency, calcium antagonists in shock/ischemia, 1066
 metabolism
 liver dysfunction in MSOF, altered cell-cell interactions, 563–565
 polytrauma, 744
 respiratory quotient (CO_2/O_2 exchange ratio) in shock, 619–621
 oxidation, metabolic abnormalities in sepsis, 536–539, 545
 turnover, metabolic abnormalities in sepsis, 547–552
 mediators, 552–557
- Glucose-insulin-potassium infusions, kidney metabolism in *E. coli* sepsis, 604
- β -Glucuronidase, platelet, activated C3 in DIC in septic shock, fulminant meningococcal meningitis, 272, 273
- Glutamate pyruvate transaminase activity, endotoxin, dose-related effects on RES, 408, 410, 411
- Glutamine and glutamic acid concentrations, serum, experimental endotoxin shock, 598
- alpha₁-acid Glycoprotein, prognostic index in sepsis, 634, 635, 716, 717
- Goris multiple organ failure score, 626–628, 664, 1026, 1027, 1029
- Granulocytes. *See* PMN entries
- Growth hormone, 380
- Gut decontamination, early ventilatory support, 786
- H7, 122
- HA 1004, 122
- Hageman factor. *See* Factor XII (Hageman factor)
- Haldane effect, 615
- Hannover Polytrauma Score, 38
- Haptoglobin, and abdominal sepsis prognosis, 716
- Heart
 inotropic plasma factor
 positive, hypovolemic shock, 253–257
 isolated, effect of LMW plasma fraction, hypovolemic traumatic shock, dog, 231–234
 LV systolic pressure, 233, 234
 negative inotropism, shock plasma ultrafiltrates, 231–234
 isolated, rabbit, endotoxin inotropic effects, 225–230
 overperformance in septic shock, 259–263

- in septic shock, awake sheep, 237–245
 - artifact rejection, 239–242
 - contractility, 238
 - hemodynamic parameters, respiratory influence, 243
 - pressure/volume loop, 238, 244
 - sonomicrometer LV dimension, 237–243
 - see also* Cardiac entries; Myocardial entries
 - Heart cells, cultured, effects of
 - Pseudomonas aeruginosa* toxins, rat, 247–251
 - and cardiac function, 247–251
 - endotoxins, 247, 249
 - type A, 247–251
 - immunoglobulins, *Pseudomonas*, protection, 247–249, 251
 - Heart dysfunction, septic shock, human, 191–197
 - cf. dog, 196
 - end diastolic volume index, 193, 194, 197
 - cf. endotoxin effect on healthy volunteers, 196–197
 - hemodynamic profiles, 192–193
 - interleukin-2, 197
 - left ventricular ejection fraction, 193–196
 - mechanisms, 194–196
 - myocardial depressant substance/factor, 194–196
 - right ventricle, 194
 - stroke volume index, 193, 194, 196, 197
 - TNF, 197
 - Heart function changes, septic shock,
 - chacma baboon (*Papio ursinus*), 207–215, 217–222
 - cardiac output, 207, 210–215, 219–222
 - heart rate, 208, 210–215
 - tachycardia and cardiac volume, 214–215, 217–222
 - left ventricular compliance, 221, 222
 - left ventricular ejection fraction, 207, 210, 211, 213–215
 - left ventricular end diastolic volume, 207, 209–214, 217, 219–222
 - left ventricular end systolic volume, 207, 209–214, 219–222
 - pulmonary capillary wedge pressure, 217, 219–222
 - stroke volume, 207, 210–214, 219, 221–222
 - systemic vascular resistance, 219, 220
 - ventriculography, radionuclide, 209–210, 218, 219
- Heart rate
- heart function changes, septic shock, chacma baboon (*Papio ursinus*), 208, 210–215, 217–222
 - α -mercaptopyropionylglycine in hemorrhagic shock, 899, 900
 - RA 642 effects on, acute hemorrhagic shock, 1094
- Hematocrit, recombinant human SOD in endotoxemia, 914–916
- Hemodynamics
- dopamine infusion, effect evaluation with systolic time intervals, 1104–1105
 - endotoxin, 390–391
 - liver dysfunction in MSOF, altered cell-cell interactions, 565
 - methylprednisolone pretreatment effects in porcine *E. coli* endotoxemia, 874, 876–877
 - PVR, 874, 876–877
 - PAF antagonists in endotoxin shock, 933, 935
 - PAF effects in sheep, 448–449
 - septic shock/septicemia, 192–193
 - immunoglobulin and plasmapheresis therapy, 1025–1030
 - pulmonary venous hemodynamics and gas exchange disturbances, Goettingen pig, 186
 - respiratory influence, awake sheep, 243
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 472, 479
 - see also* specific parameters
- Hemofiltration and survival time, porcine acute endotoxic shock, 821–826
- arachidonic acid metabolites, 823–825
 - cardiac output, 822, 825
 - extravascular lung water, 822, 823, 825

- gas exchange, 823, 825
- 6-keto-PGF₁α, 823, 825
- peripheral resistance, total, 822, 825
- thromboxane, 823–825
- Hemoglobin, respiratory quotient (CO₂/O₂ exchange ratio) in shock, 608
- Hemolysin, *Escherichia coli*, 67–70
- Hemorrhagic shock
 - calcium antagonists in shock/ischemia, 1067
 - cerebrocortical perfusion, RA642 effects, 1091–1094
 - α-mercaptopropionylglycine, 897–903
- Hemorrhagic shock, phase-related vascular reactivity, cats, 143–149
 - autoregulation, 147, 148
 - compensated shock, 144–148
 - decompensated shock, 145–148
 - electrical stimulation, efferent, isolated intestinal vascular bed, 145
 - fluid substitution, 147–148
 - noradrenaline, 146
 - oxygen free radicals, 144, 148, 149
 - permeability, capillary and postcapillary, 143
 - PMNs, 148, 149
 - vascular tone, 143
- Heparin, 940
- Hepatocytes
 - cystolic Ca²⁺ in, diltiazem and endotoxin-induced intracellular Ca²⁺ overload, 1055–1060
 - endotoxin, dose-related effects on RES, 408, 411
 - liver dysfunction in MSOF, altered cell-cell interactions, 563, 567, 568
 - phorbol ester binding sites, 583–584
 - protein kinase C and diacylglycerol accumulation, endotoxemia, rats, 575–586
 - in situ receptor autoradiography, 579, 583–584, 586
 - phorbol ester binding sites, 583–584
 - see also* Liver entries
- Hepatotoxin D-galactosamine, 1070, 1071
- Herniorraphy, 493
- Herniotomy, 1018, 1020
- HETE, cardiopulmonary response to endotoxin, eicosanoids in, sheep, 203
- 5-HETE, 351
- 15-HETE, 351
- Hetrazepine, WEB 2086 (PAF antagonist), 925, 928
- High-density lipoproteins, 81
- Hip arthroplasty, total, high-dose corticosteroids to prevent C3a and C5a formation, 879–882
- Hirudin, neutrophils, thrombin-induced adhesion with endothelial cells, 103
- Hirudin/eglin C, recombinant, endotoxin shock, proteinase/protease inhibitor therapy, 937–942
- Hirudo medicinalis*, 937
- HIS scoring system, 626–628, 645, 1026, 1027, 1029
- Histamine, 941
 - intraoperative liberation, aprotinin membrane protective action, 959–963
 - WEB 2086 (PAF antagonist), anaphylactic lung reaction, 926, 927
- Histologic appearance, pulmonary fat embolism, 40
- Honeycomb cysts, 29, 32
- 5-HT (serotonin)
 - lung injury in *E. coli* endotoxemia, 362, 368
 - platelet, activated C3 in DIC in septic shock, fulminant meningococcal meningitis, 272, 273
- Hyaline membrane disease, 29
 - elastase-α₁-PI as early indicator, 690, 691
- Hydrocortisone, 121, 122
- Hydrogen peroxide, 886, 888, 950
 - eglin C ineffectiveness in endotoxin shock, 954–956
 - lipid peroxidation, hypovolemic-traumatic shock, dogs, 346
- Hydroxyethyl starch, volume replacement in ovine endotoxemia, 815–819
 - cardiac index, 818, 819
 - cf. crystalloids, 815–819
 - lung lymph, 817, 818
 - plasma colloid osmotic pressure, 816–819
 - pulmonary artery pressure, main, 816, 817

1122 / Index

- Hydroxyl radical, 887, 950
 - alveolar permeability increased by PMA-stimulated neutrophils, rabbits, ARDS, 328
 - eglin C ineffectiveness in endotoxin shock, 954–956
 - 4-Hydroxynonenal (HNE), 662, 663
 - inflammation in surgical trauma, human, 351–355
 - lipid peroxidation, hypovolemic-traumatic shock, dogs, 345–348
 - PMNs in Sephadex inflammation model, rats, 351–355
 - chemotaxis, 351
 - superoxide anion production, 354, 355
 - Hydroxyurea, 887, 888
 - Hyperalaninemia and lethality, endotoxin shock, 595, 597–599
 - Hyperbaric oxygenation, 1008, 1010, 1013, 1020
 - Hypersensitivity, delayed-type. *See* Delayed type hypersensitivity
 - Hypertension, pulmonary
 - PAF effects in sheep, 450, 451
 - thromboxane-mediated, *E. coli* hemolysin injury to, septicemia, 67, 70
 - vascular leakage, and complement activation, 283–286
 - cf. pore-forming, *Staphylococcus* alpha-toxin, 286
 - Hyperthermia, metabolic abnormalities in sepsis, 547
 - Hyperventilation, respiratory quotient (CO₂/O₂ exchange ratio) in shock, 615, 619–621
 - Hypovolemic shock
 - ARDS, 32, 33
 - positive inotropic factor as myocardial stimulant, 253–257
 - Hypovolemic traumatic shock, 228, 229
 - lipid peroxidation, dogs, 345–350
 - plasma fraction, LMW, effect on isolated heart, 231–234
 - LV systolic pressure, 233, 234
 - negative inotropism, shock plasma ultrafiltrates, 231–234
 - Hypoxanthine levels, lipid peroxidation, hypovolemia-traumatic shock, dogs, 346–349
 - Hysterectomy, aprotinin membrane protective action, intraoperative histamine liberation, 959–963
 - Ibuprofen, cardiopulmonary response to endotoxin, eicosanoids in, sheep, 202, 203
 - Ileus, MSOF prognostic indices, logistic regression analysis, 644
 - IMAC, 702, 707–710
 - Immune complexes, circulating, plasma, and abdominal sepsis prognosis, 716
 - Immune suppression/dysfunction, post-trauma or surgery, 491–494
 - early events, 507–511
 - lymphocyte/monocyte ratio in survival, 769
 - monocyte-dependent Ig synthesis suppression, 513–516
 - MSOF, biochemical analysis and scoring, 661–662
 - T-cell mediated, polytrauma, 517–521
 - thymopentin for, 995–998
 - trauma-induced cascade of cell-mediated immune effects, 495–505
 - immunorestitution, 504–505
 - schema, 501, 503
- Immunoassay, automated homogenous enzyme immunoassay, elastase- α_1 -antiproteinase complex, 707–710
 - cf. coated tube ELISA, 708–710
 - Immunoelectrophoresis, crossed, C3 and IgG breakdown in peritonitis exudate, 529, 531
 - Immunoglobulin(s)
 - IgA, immunoglobulin profiles and PMN-elastase in septic gas gangrene, 1009, 1011
 - IgG, 427–428
 - breakdown, peritonitis exudate, C3 and, 527–532
 - deficiency substitution, and PMN-elastase in septic gas gangrene, 1007, 1012, 1018–1020

- IgM-enriched Igs (Pentaglobin),
1031–1033, 1035, 1046, 1050
in immune suppression, post-surgical or
post-traumatic, 492
lymphocyte/monocyte ratio in
polytrauma survival, 769, 770
plasma, and abdominal sepsis prognosis,
716
Pseudomonas, protection of cultured
heart cells from effects of
Pseudomonas aeruginosa toxins,
rat, 247–249, 251
synthesis
and plasmapheresis therapy,
hemodynamic effects during
treatment for septic shock,
1025–1030
suppression after multiple trauma,
513–516
trauma-induced cascade of cell-
mediated immune effects,
496, 497, 499, 501, 502
therapeutic
phagocytosis stimulation, peritonitis,
1037–1041
prophylactic, sepsis prevention after
cardiac surgery, 1031–1035
Immunoglobulin profiles and PMN-elastase
in septic gas gangrene, 1007–1022
aprotinin administration, 1008, 1012,
1014–1016
IgA, 1009, 1011
IgG, 1009, 1011, 1012–1021
IgG-deficiency substitution, 1007, 1012,
1018–1020
dosage, 1018
IgM, 1009, 1011
primary cf. secondary, 1008
Immunologic determination, humans,
proteinase inhibitor complexes,
971–974
Immunomodulation
septic shock, 989–993
thymopentin (TP-5), post-burn and
postoperative sepsis, 995–998
Indomethacin, 120, 122, 131, 203, 365,
502
immune/cytotoxic processes, role in, 442
Infarction, myocardial, 910, 972, 973
Infection
DIT marker in, 711–713
lymphocyte/monocyte ratio in
polytrauma survival, 769, 770,
772
Inflammation
autodestructive, complement activation,
MSOF pathogenesis in septic
shock, dogs, 296
early, schema, 456, 457
pharmacologic intervention points,
457
overwhelming, endotoxin role, 371–375
pathways, corticosteroids, VA
comparative study of severe
sepsis, 850–851
vs. sepsis as trigger, MSOF, 413–416
in surgical trauma, 4-hydroxy-nonenal,
351–355
see also specific cell types and mediators
Inflammatory cell activation, multiple
system organ failure,
postoperative/posttrauma,
biochemical analysis and scoring,
655–661
Inhaled antigen, WEB 2086 (PAF
antagonist), anaphylactic lung
reaction, 926, 928
Injury Severity Score, 731
dopamine infusion, effect evaluation
with systolic time intervals,
1102
embolism, fat, pulmonary, 38
phospholipase A as prognostic index,
764
polytrauma, 744, 745
scintigraphic evaluation of posttraumatic
liver function, 776
Insulin, 1060–1061, 1066
infusion, kidney function in sepsis, 604
metabolic abnormalities in sepsis, 538,
547
Intensive care unit assay, validity, elastase- α_1 -antiproteinase complex,
701–705
correlation to MOF score, 704–705
correlation to physician's classification,
703–704
ELISA, 701–702

- IMAC assay, 702
- γ -Interferon, 772
- trauma-induced cascade of cell-mediated immune effects, 495, 497–504
- Interleukin-1, 11, 13, 114, 368, 428, 432, 443, 467, 479, 486, 536, 850
- Ig synthesis suppression after multiple trauma, 513
 - liver dysfunction in MSOF, altered cell-cell interactions, 564, 568, 569
 - pulmonary vascular permeability, 305
 - serum, and sepsis, prognosis/prognostic indices, 715–718
 - trauma-induced cascade of cell-mediated immune effects, 495, 497–503
- Interleukin-2, 772, 990, 995, 996
- heart dysfunction, septic shock, human, 197
 - Ig synthesis suppression after multiple trauma, 513, 515
 - immune/cytotoxic processes, role in, 442
 - trauma-induced cascade of cell-mediated immune effects, 496–503
- Interleukin-2 receptors
- immune suppression, post-surgical or post-traumatic, 492, 494
 - multiple trauma, early events, 508, 509
 - trauma-induced cascade of cell-mediated immune effects, 496, 501, 502
- Interleukin-4 (BCGF), trauma-induced cascade of cell-mediated immune effects, 497
- Intestinal transit velocity, WEB 2086 (PAF antagonist), 919–922
- Intima. *See* Vascular intima in endotoxin shock
- Intraglobin, 1047, 1050
- Intra-tracheal pressure, lung injury in *E. coli* endotoxemia, 360, 361, 365
- Inverse ratio ventilation
- early ventilatory support, 788, 789
 - nifedipine for ARDS, 1087, 1090
- IP₃, hepatocytes, protein kinase C and diacylglycerol accumulation, endotoxemia, 575
- Iron-dependent lipid peroxidation in cardiopulmonary arrest, 895
- Ischemia, myocardial
- acute, dynamics of prostacyclin and thromboxane, 907–911
 - arrhythmias, 907
- Ischemia and circulatory system in MSOF, 1075–1083
- ARDS, 1076, 1078, 1081, 1083
- pulmonary hypertension, 1078
- catecholamines, 1077
- central mechanisms, 1075–1079
- cardiac output, 1076–1077, 1080, 1082
 - myocardial depressant factor, 1076, 1077
 - myocardium, reperfused, 1077
 - oxygen delivery, 1075–1083
- peripheral mechanisms, 1079–1082
- capillary surface area reduction, 1082
 - oxygen extraction, 1079–1083
 - regional blood flow, 1080–1081, 1083
- see also* Calcium antagonists in shock/ischemia
- Isoprinosine, 502
- Kadsurenone, 428, 485
- Kallikrein, 738, 741, 983
- circulation, peripheral, septic shock, 171
 - endotoxin interactions with plasma contact system factors, 401–404
- TNF, induction of organ changes in chronic lymph fistula, sheep, 469, 474, 479
- Kallikrein-kinin system, 318
- in ARDS after polytrauma, 737–742
 - endotoxin, 389, 390, 392, 393
 - pulmonary vascular permeability, 305–307
- Ketanserin, 362
- Kidney
- complement activation, MSOF pathogenesis in septic shock, dogs, 294–296
 - and dopamine, 1097–1099
 - plasma flow, 1098
 - failure, postoperative, 133–136
 - metabolism in *E. coli* sepsis, 601–605
 - ATP MgCl₂, 602, 605
 - glucose concentrations, 602, 604

- glucose-insulin-potassium infusions, 604
- lactate or fructose infusion, 603
- TAN concentrations, 601, 602, 604
- microthrombosis, endotoxin-induced, 916
- PAF antagonist inhibition of induced shock, 429
- RES stimulation to protect against DIC, 1002, 1003
- Kininase II, 941
- Kininogen, 983, 986
- Kinins, 986–987
 - cascade, lung microvascular bed, cell interactions in septic shock, 115
 - see also* Kallikrein-kinin system
- Kupffer cells, 78, 81, 294, 779, 1002, 1004
 - endotoxin, dose-related effects on RES, 408
 - liver dysfunction in MSOF, altered cell-cell interactions, 563, 564, 566–571
 - zymosan-induced MSOF, endotoxin plays no key role, 422
- L-652,731, 426, 428, 442, 485
- L-653,150, 428
- Lactate
 - or fructose infusion, kidney metabolism in *E. coli* sepsis, 603
 - hypovolemic-traumatic shock, dogs, 346–349
 - metabolism, liver, 744
 - polytrauma, 743, 745, 747
- Lactate dehydrogenase, lung, endotoxin-induced microvascular endothelial injury, 93, 96
- Lactoferrin, pulmonary vascular permeability, 307, 308
- Laser Doppler flowmetry, RA642, effects on cerebrocortical perfusion, acute hemorrhagic shock, 1091–1094
- Lavage therapy, C3 and IgG breakdown in peritonitis exudate, 530, 531
- Leukocyte(s)
 - count
 - methylprednisolone pretreatment, endotoxemia, 875, 876
 - recombinant human SOD in endotoxemia, 914, 915, 917
 - total, 699, 700
 - induced injury with zymosan, lung, 73–76
 - phagocytic activity in sepsis/infection, diiodotyrosine (DIT) as marker for, 711–713
 - and TNF, induction of organ changes in chronic lymph fistula, sheep, 469, 470, 475–477, 480
 - see also* specific types
- Leukocyte neutral proteinase inhibitor, 945–950
- Leukopenia, WEB 2086 (PAF antagonist), anaphylactic lung reaction, 926
- Leukostasis
 - alveolar permeability increased by PMA-stimulated neutrophils, rabbits, ARDS, 325, 326
 - lipid peroxidation in hypovolemic shock, 349–350
 - lung in shock, 4–5, 7–8
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 473, 474, 480
- Leukotrienes, 70, 351
 - cardiopulmonary response to endotoxin, eicosanoids in, sheep, 203, 204
 - complement activation, pulmonary hypertension and vascular leakage, 286
 - early ventilatory support, 785
 - LTB₄, sepsis, 535, 536
 - LTC₄, 432
 - lung injury in *E. coli* endotoxemia, 362
 - synthesis, RA642 effects on cerebrocortical perfusion, acute hemorrhagic shock, 1094
- Lipid A. *See* Antibodies, anti-LPS and anti-lipid A, determination with immunoblotting
- Lipid peroxidation, 662, 663
 - hypovolemic-traumatic shock, dogs, 345–350
 - inhibition in cardiopulmonary arrest, dogs, 891–895
 - see also* Oxygen radicals
- Lipocortin, 123
- Lipooxygenase inhibition, AA-861, 131
- Lipopolysaccharide, *S. abortus equi*, 119, 121; *see also* Antibodies, anti-LPS

- and anti-lipid A, determination with immunoblotting
- Lipoprotein, high-density, 81
- Lipoprotein lipase suppression, TNF, 464
- Liver
 - complement activation, MSOF
 - pathogenesis in septic shock, dogs, 293–294, 296
 - enzymes, serum, scintigraphic evaluation of posttraumatic liver function, 776–779
 - failure, complement activation and endotoxin role, sepsis, 277
 - lactate metabolism, 744
 - leukostasis, TNF induction of organ changes in chronic lymph fistula, 473, 474, 480
 - perfused, sepsis effect on metabolism, 589–592
 - scintigraphic evaluation of posttraumatic function, 775–780
 - SGOT, TNF induction of changes in chronic lymph fistula, sheep, 469, 474, 475, 480
 - sinusoidal macrophages, endotoxin, dose-related effects on RES, 408
 - see also* Hepatocytes; Kupffer cells
- Liver dysfunction in MSOF, altered cell-cell interactions, 563–571
 - glucose metabolism, 563–565
 - hemodynamics, 565
 - hepatocytes, 563, 567, 568
 - interleukin-1, 564, 568, 569
 - Kupffer cells, 563, 564, 566–571
 - paracrine amplification, 563, 569
 - polyunsaturated fatty acids, 569–571
 - prostaglandins, 564, 568, 569
 - protein metabolism, 565, 567, 568
 - thromboxane, 569
 - TNF, 564, 568–570
- Low-flow states, respiratory quotient (CO₂/O₂ exchange ratio) in shock, 615, 616
- Luminol-dependent chemiluminescence, 339–344
 - cf. zymosan-activated, 341
- Lung(s), 3–10
 - anaphylactic reaction, WEB 2086 (PAF antagonist), 925–929
 - endothelial swelling, 8, 9, 11
 - endotoxin-induced microvascular endothelial injury, 91–97
 - Escherichia coli* hemolysin injury, septicemia in, 67–70
 - transmembrane pores, 69
 - injury, PMA, SOD after, 945–949
 - isolated, pulmonary vascular permeability, 309–311
 - leukocyte-induced injury with zymosan, 73–76
 - leukostasis, 4–5, 7–8
 - microvascular bed, cell interactions in septic shock, 113–118
 - organ failure, 10–14; *see also* ARDS
 - PAF antagonist inhibition of induced shock, 429
 - perfusion, decreased, 5–6
 - platelet activation, 6, 7
 - zymosan-induced MSOF, endotoxin plays no key role, 421–423
- Lung edema. *See* Pulmonary edema in shock
- Lung epithelial lining fluid, 792, 793
- Lung injury in *E. coli* endotoxemia, 357–368
 - cats, vagotomized, 361
 - intra-tracheal pressure, 360, 361, 365
 - mediators, 358, 362–366
 - pulmonary artery pressure, 360, 364, 365, 368
 - pulmonary compliance, 358–361, 367
 - pulmonary resistance, 358, 359
 - transpulmonary pressure, 358–361
- Lung lymph flow
 - cardiopulmonary response to endotoxin, eicosanoids in, sheep, 202–204
 - free radical scavengers and cardiopulmonary response to endotoxin, 885
 - hydroxyethyl starch, volume replacement in ovine endotoxemia, 817, 818
 - PAF antagonists in endotoxin shock, 933–935
 - PAF effects in sheep, 447, 450
 - permeability, pulmonary vascular, mediators, 308
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 469, 473, 475, 479

- Lung water, extravascular. *See*
Extravascular lung water (EVLW)
- Lupus erythematoses cells, 52, 53
- Lymph fistula. *See* TNF, induction of organ
changes in chronic lymph fistula,
sheep
- Lymphocyte(s)
counts, immune suppression, post-
surgical or post-traumatic, 492
/monocyte ratio in polytrauma survival,
prognosis/prognostic indices,
769–772
see also B cell(s); T cell(s)
- Lysolecithin acyl-transferase (LAT), 122,
123, 455
- α_2 -Macroglobulin, plasma, and abdominal
sepsis prognosis, 716, 717, 720
- Macrophage(s)
activation/induction
elastase- α_1 -PI complex and
neopterin, plasma levels, 687
and TNF productin in shock, PAF,
485–488
alveolar, chemotactic factors, 20
ARDS, posttraumatic, prognosis, 679
hepatic. *See* Kupffer cells
multiple system organ failure,
postoperative/posttrauma,
biochemical analysis and scoring,
657, 659, 660
origins of, lung, microvascular bed, cell
interactions in septic shock,
113–114, 118
phospholipase A source, 757, 763, 767
pulmonary intravascular
endotoxin-induced microvascular
endothelial injury, 92
endotoxin shock, 78, 80, 81
vascular intima in endotoxin shock, 78,
80, 81
- Macrophage-activating factor, trauma-
induced cascade of cell-mediated
immune effects, 497
- Magnesium and calcium antagonists in
shock/ischemia, 1072
- Magnesium chloride, ATP-, kidney function
in sepsis, 602, 605
- Major basic protein, 432, 488
- Malondialdehyde, 662, 663, 910
leukocyte-induced lung injury, 74, 75
lipid peroxidation, hypovolemic-
traumatic shock, dogs, 346–348
- Mannheim Peritonitis Index, 723
- Mast cell chymase, 937, 941, 942
- Meclophenamate, 203
- Meconium aspiration, elastase- α_1 -PI as
early indicator, 690, 691
- Membrane protective action, intraoperative
histamine liberation, aprotinin,
959–963
- Meningitis
elastase- α_1 -PI as early indicator,
690–692
fulminant meningococcal, activated C3
in DIC in septic shock, 271–274
- Mepacrine, 120
- Mepyramine, 362, 926
- α -Mercaptopropionyl glycine, hemorrhagic
shock, 897–903
- Mesenteric blood flow, burns, enteric
translocation of microorganisms,
377–380
- Metabolic abnormalities in sepsis,
535–542, 545–559
ARDS, 535, 537, 538
branched-chain amino acids, 539, 540,
617
carbon dioxide, arterial, 537, 538
dichloroacetic acid therapy, 540,
541
glucose turnover, 547–552
mediators, 552–557
hyperthermia, 547
insulin, 538, 547
LTB₄, 535, 536
mitochondrial pyruvate dehydrogenase,
538–540, 549, 550
MOSF, 535–542
oxidation of glucose cf. fats, 536–539,
545
PGF₂ α /PGE₂ ratio, 535, 536
proteolysis, excessive, 540–542
rat experimental model, 546–547
superoxides, 535, 536
TPN, 537, 538
- Metabolic imbalance, multiple system organ
failure, postoperative/posttrauma,
biochemical analysis and scoring,
663–664

- Metabolic rate, septic shock, heart overperformance in, 260–262
- Methylmethacrylate cement, 879, 880
- Methylprednisolone, pretreatment effects in porcine *E. coli* endotoxemia, 873–877
- coagulation, 873
 - fibrinolysis, 873, 877
 - hemodynamics, 874, 876–877
 - PVR, 874, 876–877
 - proteolysis cascades, 873, 877
 - VA study, 873
- Methylprednisolone for septic shock with ARDS, 861–863
- with gentamicin, dogs, *E. coli* shock, animal model development for shock, 836–837
 - adrenal gland role, 838–839
 - cf. baboons, 836–840
 - rationale, 837–838
- Methysergide, 362
- Microangiopathy, diabetic, 1009
- Microatelectasis, early ventilatory support, 787
- α_2 -Microglobulin
- immune suppression, post-surgical or post-traumatic, 492
 - kallikrein-kinin system components in ARDS after polytrauma, 738, 739, 742
- Microvascular integrity, extravascular lung water, altered fluid regimen, advanced septic shock with acute respiratory failure, 806
- Microvascular permeability. *See under* Permeability
- Milano Sepsis Score, 634–636
- complement activation, endotoxin role in sepsis, 278–280
- Minimal pulmonary dysfunction, 52
- Mitochondrial function
- calcium antagonists in shock/ischemia, 1066
 - pyruvate dehydrogenase, metabolic abnormalities in sepsis, 538–540, 549, 550
- Monocyte(s), 989, 991, 993
- count, multiple trauma, early events, 508, 509
 - dependent Ig synthesis suppression after multiple trauma, immune suppression/dysfunction, 513–516
 - /lymphocyte ratio as prognostic factor, polytrauma, 769–772
 - synthesis, α_1 -protease inhibitor, 948
 - see also* Macrophage(s)
- Monokine synthesis factor, 499
- Monolayer-filter membrane system, endothelium permeability and, 128–131
- MPP
- pulmonary vascular resistance, ARDS in septicemia, 175–178
 - pulmonary venous hemodynamics and gas exchange disturbances, *E. coli* septicemia, Goettingen pig, 185, 186
- MTDQ-DA antioxidant, myocardial ischemia, 907–911
- Mucosal integrity, burns, enteric translocation of microorganisms, 377–380
- Multiple system organ failure (MOF, MOSF, MSOF)
- in acute pancreatitis, C3a and C5a, 265–268
 - terminal complement complexes, 266, 267
 - ARDS, septic, 57–60
 - prognosis, 59
 - C3a, 880
 - ceruloplasmin changes, 331–337
 - C-reactive protein, 335, 336
 - number of organs involved, 333, 334, 336
 - complement activation in, septic shock, 291–296
 - elastase- α_1 -PI complex and neopterin, plasma levels, 683–687
 - endotoxin and proteases, septicemia, 315–321
 - diagnostic criteria for MSOF, 316
 - PFI index, 317–321
 - epidemiology, 783–784
 - Goris score, 1026, 1027, 1029
 - ischemia and circulatory system in, 1075–1083

- liver dysfunction, 563–571
- lymphocyte/monocyte ratio in
 - polytrauma survival, 769, 770
- metabolic abnormalities in sepsis, 535–542
- muscle pO₂ role, critically ill patients, 138–140
- phospholipase A as prognostic index, 766
- prevention, zymosan-induced peritonitis, decontamination, GI tract, 827–832
- prognostic indices, 643–647
- sepsis vs. inflammation as trigger, polytrauma, 413–416
- ventilatory support, early, 784–789
- zymosan-induced, no endotoxin role, 419–423
- Multiple system organ failure,
 - postoperative/posttrauma, biochemical analysis and scoring, 649–665
 - coagulation cascade, fibrinolysis, and kallikrein system, 652–654
 - complement cascade, 650–651
 - endotoxin, 654–655
 - immune suppression/dysfunction, 661–662
 - inflammatory cell activation, 655–661
 - metabolic imbalance, 663–664
 - organ function parameters, 664–665
 - stages, 649
 - target structure degradation, 662–663
- Multiple trauma. *See* Polytrauma; Trauma
- Muscle
 - pO₂ role, critically ill patients, MOF, 137–142
 - MOF scores, 138–140
 - survival, 139, 141
 - skeletal, diltiazem and endotoxin-induced intracellular Ca²⁺ overload, 1054–1061
 - smooth, spasm, calcium antagonists in shock/ischemia, 1066
- Mycostatin, 378
- Mycotoxin-induced shock, PAF antagonist inhibition of induced shock, 431
- Myeloperoxidase, pulmonary vascular permeability, 307, 308
- Myocardial contractility, depressed, dopamine infusion effect evaluation with systolic time intervals, 1101
- Myocardial depressant factor, 253, 261, 262, 431, 1068
 - heart dysfunction, septic shock, human, 194–196
 - ischemia and circulatory system in MSOF, 1076, 1077
- Myocardial infarction, 910, 972, 973
- Myocardial ischemia
 - acute, dynamics of prostacyclin and thromboxane, 907–911
 - arrhythmias, 907
- Myocardium, reperfused, ischemia and circulatory system in MSOF, 1077
- Negative chronotropic effect, α -mercaptopyronylglycine in hemorrhagic shock, 902
- Neonatal respiratory distress syndrome, exogenous surfactant, 797
- Neopterin, 477, 657, 659, 660
 - ARDS, posttraumatic, prognosis, 674–679
 - and elastase- α_1 -PI complex, plasma levels in MSOF, 683–687
 - immune suppression, post-surgical or post-traumatic, 492–494
 - polytrauma, 745, 747
- Neuroleptics, 755
- Neurologic function and lipid peroxidation inhibition in cardiopulmonary arrest, 891–895
- Neutrophils. *See* PMN entries
- Nicardipine, endotoxin shock, 1071
- Nifedipine, 1060
 - ARDS, 1087–1090
 - endotoxin shock, 1070, 1071
- Nimodipine, traumatic shock, 1067, 1068
- Nine Rule, total burn surface estimation, 1102, 1103
- Nitrogen mustard, 887
- Nivadipine, endotoxin shock, 1070
- NK cells, 989, 991
 - PAF antagonist effects, 443–444
- Noradrenaline/norepinephrine
 - hemorrhagic shock, phase-related vascular reactivity, cats, 146

1130 / Index

- metabolic abnormalities in sepsis, 547, 548
- ONO, 6240, 428, 931
- Opsonins, C3 and IgG breakdown in peritonitis exudate, 527–532
- Organ failure, multiple. *See* Multiple system organ failure (MOF, MOSF, MSOF)
- Organ failure, postoperative, tissue oxygen debt (VO₂ deficit) as determinant, 133–136
 - survivors cf. nonsurvivors, 136
- Osmotic pressure. *See* Plasma colloid osmotic pressure
- Oxidation of glucose cf. fats, metabolic abnormalities in sepsis, 536–539, 545
- Oxygen
 - arterial tension, endotoxin effect, 391–392
 - delivery
 - endotoxin, inotropic effect in isolated rabbit heart, 227
 - ischemia and circulatory system in MSOF, 1075–1083
 - extraction
 - ischemia and circulatory system in MSOF, 1079–1083
 - peripheral, septic shock, heart overperformance in, 260–262
 - see also* Respiratory quotient (CO₂/O₂ exchange ratio) in shock
 - hypoxia, early ventilatory support, 784–787
 - and leukocyte-induced lung injury, 74, 75
 - skeletal muscle pO₂ and multiple organ failure, 137–142
 - supply
 - peripheral, pulmonary vascular resistance, ARDS in septicemia, 175, 177, 179
 - uptake relationship, septic shock, 181–183
 - tissue oxygen debt (VO₂) deficit and postoperative organ failure, 133–136
 - survivors cf. nonsurvivors, 136
 - utilization, circulation, peripheral, septic shock, 163, 167–169
- Oxygen, hyperbaric, 1008, 1010, 1013, 1020
- Oxygen radicals
 - ARDS, exogenous surfactant, 791, 793
 - ceruloplasmin changes in MSOF and ARDS, 331–332, 337
 - endotoxemia, 886–888
 - hemorrhagic shock, phase-related vascular reactivity, cats, 144, 148, 149
 - leukocyte-induced lung injury, 73, 75, 76
- lung
 - endotoxin-induced microvascular endothelial injury, 95
 - microvascular bed, cell interactions in septic shock, 114
- multiple organ failure,
 - postoperative/posttrauma, biochemical analysis and scoring, 656, 661
 - permeability, pulmonary vascular, mediators, 305–306, 311, 312
 - PMN activation, 850
 - scavenging. *See* Free radical scavengers
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 470, 476, 477
 - see also* Lipid peroxidation; *specific types*
- Pancreatic cysts, C-reactive protein as prognostic index, 726
- Pancreatitis
 - C3a and C5a in MSOF, 265–268
 - terminal complement complexes, 266, 267
 - C-reactive protein as prognostic index, 725–728
 - extravascular lung water, large volume replacement with crystalloids, 809
- Papillary muscle, guinea pig, 231–233
- Papio ursinus*, septic shock, heart function changes, 207–215, 217–222
- Paracrine amplification, liver dysfunction in MSOF, altered cell-cell interactions, 563, 569
- Parietal cells, gastric mucosa ulceration,

- ultrastructure after septic shock, rat, 152, 153
- Passive sensitization, WEB 2086 (PAF antagonist), anaphylactic lung reaction, 926, 928
- Pediatric systemic infection, elastase- α_1 -PI as early indicator, 689–693
- PEEP
 - nifedipine for ARDS, 1087, 1090
 - ventilatory support, early, 788–789
- Pelvis fracture and pulmonary fat embolism, 39–41
- Pentaglobin, 1031–1033, 1035, 1046, 1050
- Perfusion, decreased, lung in shock, 5–6
- Perfusion circuit, endotoxin, inotropic effect in isolated rabbit heart, 226, 227
- Peripheral circulation. *See* Circulation, peripheral, septic shock
- Peripheral resistance, total
 - hemofiltration and survival time, porcine acute endotoxic shock, 822, 825
 - septic shock, heart overperformance in, 260, 262
- Peritoneal exudate cells, therapeutic Igs, phagocytosis stimulation in peritonitis, 1038
- Peritonitis
 - C-reactive protein as prognostic index, 725–728
 - endotoxin and overwhelming inflammatory response of early sepsis, 372–374
 - exudate, C3 and, immunoglobulin G breakdown, 527–532
 - therapeutic Ig stimulation of phagocytosis, exudate cells, 1038
 - zymosan-induced, decontamination, GI tract, 827–832
- Peritonitis Index, 643
- Permeability
 - alveolar
 - cf. capillary permeability, 323, 328–329
 - gamma-scintillation with DTPA, detection, 324–327
 - PMA-stimulated neutrophils, rabbit ARDS, 323–329
 - capillary
 - endothelial cells, 157, 159
 - and postcapillary, hemorrhagic shock, phase-related vascular reactivity, cats, 143
 - complement and pulmonary hypertension, 283–286
 - endothelial, bacterial toxins and calcium effects, 127–131
 - microvascular, 92–93, 96
 - circulation, peripheral, septic shock, 167
 - free radical scavengers and cardiopulmonary response to endotoxin, 885, 886, 888
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 472, 473, 479–481
- Permeability, pulmonary vascular, mediators, 305–312
 - E. coli* hemolysin injury, septicemia, 67, 69–70
 - eicosanoids/arachidonic acid cascade, 305, 306, 308, 312
 - lung lymph flow, 308
 - thromboxane, 310
 - granulocytes (PMNs) and macrophages as source, 307–311
 - isolated rabbit lung, 309–311
 - oxygen radicals, 305–306, 311, 312
 - PAF effects, sheep, 447, 451
 - PMN-elastase- α_1 -protease inhibitor, 308–310
 - proteases, 306, 309, 312
 - inhibitor Eglin C, 311
- PFI index, 317–321, 653, 731–735
- pH
 - isolated heart, effect of LMW plasma fraction in hypovolemia traumatic shock, 234
 - respiratory quotient (CO_2/O_2 exchange ratio) in shock, 608, 609, 615–617
- Phagocytic index, RES stimulation to protect against DIC, 1004
- Phagocytosis assay, 508, 510
- Phagocytosis stimulation, peritonitis, therapeutic immunoglobulins, 1037–1041
 - IgG, 1041
 - IgM, 1039, 1041
- Phentolamine, 553, 555, 557, 558

- Phenylephrine, protein kinase C and diacylglycerol accumulation in hepatocytes, endotoxemia, 579–581
- Phorbol esters, protein kinase C and diacylglycerol accumulation in hepatocytes, endotoxemia, 576, 578, 579, 582, 586
- Phorbol myristate acetate, 123
lung injury, superoxide dismutase, 945–949
-stimulated neutrophils, rabbit ARDS, permeability, alveolar, 323–329
- Phosphatidate phosphohydrolyase, 576
- Phosphatidylcholine and phosphatidyl glycerol, exogenous surfactant, 792
- Phospholipase A
lethality correlation, 759–761, 764, 765
severely ill patients, prognosis/prognostic indices, 757–761, 763–768
sources, 757, 763, 767
- Phospholipase A₂, 123, 432
C, protein kinase C and diacylglycerol accumulation in hepatocytes, endotoxemia, 575, 576
calcium antagonists in shock/ischemia, 1066, 1071
PMN activatin, 850, 851, 855
- Phospholipid reacylation, burns, PAF inhibitor effect, scalded pig, 455, 459, 461
- Pia arterioles, RA642, effects on cerebrocortical perfusion, acute hemorrhagic shock, 1093
- PIP₂, hepatocytes, protein kinase C and diacylglycerol accumulation, endotoxemia, 575
- Plasma colloid osmotic pressure and extravascular lung water
altered fluid regimen, advanced septic shock with acute respiratory failure, 803–808
large volume replacement with crystalloids, 810–812
hydroxyethyl starch, volume replacement in ovine endotoxemia, 816–819
- Plasma contact system factors, in vitro interactions, endotoxin, 401–405
- Plasma flow, renal, dopamine and renal function, 1098
- Plasma fraction, LMW, effect on isolated heart, hypovolemic traumatic shock, dog, 231–234
LV systolic pressure, 233, 234
negative inotropism, shock plasma ultrafiltrates, 231–234
- Plasmapheresis, 1025–1030
- Plasma proteins, abdominal sepsis, prognosis/prognostic indices, 719–724
- Plasma substitution in septic shock, humans, antithrombin III, 965–968
- Plasma suppressive activity, thymopentin (TP-5) in post-burn and postoperative sepsis and immunodeficiency syndrome, 996–998
- Plasmin, methylprednisolone pretreatment, endotoxemia, 874, 875
- Plasmin- α_2 -antiplasmin complex, immunologic determination, humans, 971
- Plasminogen
consumption, aprotinin membrane protective action, intraoperative histamine liberation, 961, 962
kallikrein-kinin system components in ARDS after polytrauma, 738, 740
- Plasminogen activator, 654
inhibitor, and fibrinolysis, 383–386, 395–399
urokinase-type, neutrophils, thrombin-induced adhesion with endothelial cells, 109
- Plasminogen activator, tissue (tPA), 576, 578, 579, 582, 586, 938
binding sites, hepatocytes, 583–584
endotoxic shock, fibrinolytic function determinations, pig, 395–399
fibrinolysis, 383–386
neutrophils, thrombin-induced adhesion with endothelial cells, 109
- Platelet(s)
activation, lung in shock, 6, 7
aggregation, calcium antagonists in shock/ischemia, 1066
aprotinin membrane protective action, intraoperative histamine liberation, 960, 961

- count
 - methylprednisolone pretreatment, endotoxemia, 875, 876
 - RES stimulation to protect against DIC, 1002, 1005
- endotoxin-induced DIC, AT III-heparin complex therapy, 980
- β -glucuronidase, activated C3 in DIC in septic shock, fulminant meningococcal meningitis, 272, 273
- 5-HT, activated C3 in DIC in septic shock, fulminant meningococcal meningitis, 272, 273
- recombinant human SOD in endotoxemia, 914–915, 917
- Platelet activating factor (PAF), 13, 14, 481, 660
 - antagonists, inhibition of induced shock, 427–433
 - C5a and, 366
 - calcium, 366
 - chronically instrumented sheep, effect on, 447–451
 - generation during shock, 426–427
 - glucose turnover in sepsis, 556–558
 - immune/cytotoxic processes, role in, 441–443
 - infusion in animals, cf. shock, 425–426
 - lung injury in *E. coli* endotoxemia, 358, 366–368
 - macrophage/monocyte induction and TNF production in shock, 485–488
 - PAF antagonist effects, 485, 486
 - neutrophil aggregation, 108, 109, 367
- Platelet activating factor antagonists
 - burns, 455–461
 - cytotoxic processes, effects, 443–444
 - endotoxic shock, 931–935
 - inhibition of induced shock, inhibition of PAF-generated feedback cycles, 432–433
 - neutrophils, thrombin-induced adhesion with endothelial cells, 108
 - ONO-6240, 428, 931
 - see also* BN 52021; WEB 2086 (PAF antagonist); *specific antagonists*
- Platelet-derived growth factor, neutrophils, thrombin-induced adhesion with endothelial cells, 109
- PMN(s) (neutrophils, granulocytes), 20–22, 972
 - activation, 21, 850
 - and leukostasis, lipid peroxidation, hypovolemic-traumatic shock, dogs, 349–350
 - see also specific activation products*
 - aggregation, PAF, 367
 - ARDS, 18–20, 31–33
 - posttraumatic, prognosis, 673, 674, 678, 679
 - ceruloplasmin changes in MSOF and ARDS, 331
 - complement, 20
 - MSOF pathogenesis in septic shock, dogs, 293, 296
 - pulmonary hypertension and vascular leakage, 283, 284
 - early ventilatory support, 784
 - eglin C ineffectiveness in endotoxin shock, 954–956
 - endothelial cell adherence, 157, 159
 - bacterial endotoxin role, 123–124
 - proadherent factor, 106–107, 109
 - thrombin-induced, 101–109
 - generation, chemiluminescence-inducing radicals, porcine septic shock, 339, 341
 - hemorrhagic shock, phase-related vascular reactivity, cats, 148, 149
 - 4-hydroxy-nonenal, 351–355
 - Igs, therapeutic, phagocytosis stimulation in peritonitis, 1039–1041
 - lung injury, 11–12, 74, 75
- MSOF
 - polytrauma, 414
 - postoperative/posttrauma,
 - biochemical analysis and scoring, 653, 655, 657–660
 - multiple trauma, early events, 507, 508, 510, 511
 - phospholipase A source, 757, 763, 767
 - and pulmonary vascular permeability, 307–311
 - superoxide radical production by, recombinant human SOD in endotoxemia, 913, 917

- zymosan-induced MSOF, endotoxin plays no key role, 422
- see also* Elastase- α_1 -antiproteinase complex; Oxygen radicals; PMN elastase
- PMN elastase, 331, 731–735, 937, 941, 942
- ARDS, post-traumatic, prognosis, 674–679
- eglin C ineffectiveness in endotoxin shock, 953–957
- porcine shock, proteinase inhibitor, leukocyte neutral, 945–950
- eglin C in septic shock, 945–948
- PMA lung injury, SOD after, 945–949
- prognostic index in sepsis, 634, 635
- see also* Elastase- α_1 -antiproteinase complex; Immunoglobulin profiles and PMN-elastase in septic gas gangrene
- PMN elastase- α_1 -antitrypsin complex, antithrombin III and plasma substitution in septic shock, 966–968
- Pneumocytes, type II, 52, 53
- Pneumonia
 - Ig prophylactic therapy after cardiac surgery, 1034
 - pediatric, elastase- α_1 -PI as early indicator, 690–692
 - phospholipase A as prognostic index, 765–768
- Polyphloretin, 363
- Polytrauma
 - ARDS, kallikrein-kinin system, 737–742
 - biochemical and hormonal parameters, 743–745, 746–749
 - monocyte/lymphocyte ratio as prognostic factor, 769–772
 - PFI index, 731–735
 - phospholipase A as prognostic index, 763–768
- Polytrauma Score, T-cell-mediated immune suppression after polytrauma, 517
- Polyunsaturated fatty acids, liver dysfunction in MSOF, altered cell-cell interactions, 569–571
- Positive inotropic factor as myocardial stimulant, ion exchange chromatography, 254–255
- Potassium
 - and cardiac function, *Pseudomonas aeruginosa* toxins, 247–249
 - infusion, kidney, function in sepsis, 604
- PR 1501, 443
- PR 1502, 443
- Prealbumin, plasma, and abdominal sepsis prognosis, 720, 722
- Predictors. *See* Prognosis/prognostic indices
- Prednisone, 377
- Preallikrein, 986
 - in ARDS after polytrauma, 737, 738
 - and endotoxin
 - interactions with plasma contact system factors, 402–404
 - and overwhelming inflammatory response of early sepsis, 372
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 469, 474, 475, 479
- Pressure/volume loop, heart performance during septic shock, awake sheep, 238, 244
- Proadherent factor, neutrophils, thrombin-induced adhesion with endothelial cells, 106–107, 109
- Proenzyme Functional Inhibition (PFI) Index, 317–321, 653, 731–735
- Progesterone, 122
- Prognosis/prognostic indices
 - ARDS, posttraumatic, plasma levels of mediators, 673–679
 - macrophages, 679
 - cf. non-ARDS, 676, 677, 679
 - C-reactive protein in pancreatitis and peritonitis, 725–728
 - elastase- α_1 -PI, early indicator of pediatric systemic infection, 689–693
 - interleukin-1, serum, and sepsis, 715–718
 - lymphocyte/monocyte ratio in polytrauma survival, 769–772
 - MSOF, logistic regression analysis, 643–647
 - PFI index, polytrauma, 731–735
 - cf. elastase, 731–735
 - phospholipase A in severely ill patients, 757–761, 763–768

- plasma proteins, abdominal sepsis, 719–724
- scintigraphic evaluation of posttraumatic liver function, 775–780
- sepsis, 633–636
TNF, serum, 715–718
- Prolactin, sepsis, 751, 753, 754, 756
- Promethazine, 1070
- Properdin factor B, plasma, and abdominal sepsis prognosis, 720
- Propranolol, 553, 555, 557, 558
- Prostacyclin (PGI₂), 119–123
cardiopulmonary response to endotoxin, eicosanoids in, sheep, 202, 203
circulation, peripheral, septic shock, 168
complement activation, pulmonary hypertension and vascular leakage, 285–286
liver dysfunction in MSOF, altered cell-cell interactions, 589
lung injury, endotoxin-induced, 366
microvascular endothelium, 92–94, 96, 97
myocardial ischemia, 907–911
neutrophils, thrombin-induced adhesion with endothelial cells, 109
- Prostaglandin(s), 380, 660–661
burns, PAF inhibitor effect, scalded pig, 455, 459, 460
- D₂, lung injury in *E. coli* endotoxemia, 362, 365, 366
- E₂, 429, 661, 662
Ig synthesis suppression after multiple trauma, 513, 515
liver dysfunction in MSOF, altered cell-cell interactions, 564, 568, 569
lung, endotoxin-induced injury, 92–94, 96, 97, 359, 363
trauma-induced cascade of cell-mediated immune effects, 497–504
- F_{2 α}
lung injury in *E. coli* endotoxemia, 359, 362–366
/PGE₂ ratio, metabolic abnormalities in sepsis, 535, 536
glucose turnover in sepsis, 552
- H₂, 122
- 6-keto-PGF_{1 α} , hemofiltration and survival time, porcine acute endotoxin shock, 823, 825
- Prostaglandin endoperoxide synthetase, 910
- Prostaglandin synthetase, 1071
- Protease(s)
in MSOF due to septicemia, endotoxin, 315–321
permeability, pulmonary vascular, mediators, 306, 309, 312
inhibitor Eglin C, 311
see also specific proteases
- α ₁-Protease inhibitor, 331, 725–726
ARDS, posttraumatic, prognosis, 674, 676
kallikrein-kinin system components in ARDS after polytrauma, 738, 740
monocyte synthesis, 948
and PAF antagonist in induced shock, 431–432
- Proteinase inhibitor complexes
immunologic determination, humans, 971–974
leukocyte neutral, ELISA, 945–946
PMN elastase complex, porcine shock, 945–950
PMN-derived, eglin C ineffectiveness in endotoxin shock, 954–956
see also Elastase- α ₁-antiproteinase complex; specific proteinase inhibitors
- Proteinase/protease inhibitor therapy, hirudin/eglin C, recombinant, endotoxin shock, 937–942
- Protein C, 654
- Protein kinase C, 122, 123
and diacylglycerol accumulation, endotoxemia, rat hepatocytes, 575–586
in situ receptor autoradiography, 579, 583–584, 586
phorbol ester binding sites, 583–584
- Protein metabolism, liver
dysfunction in MSOF, altered cell-cell interactions, 565, 567, 568
perfused, sepsis effect on metabolism, 589–592
- Proteolysis
cascades, methylprednisolone, pretreatment effects in porcine *E. coli* endotoxemia, 873, 877

- excessive, metabolic abnormalities in sepsis, 540–542
- Prothrombin, 319
 - methylprednisolone pretreatment, endotoxemia, 874, 875
- Providencia pectgeri*, 420
- Pseudomonas*, 831
 - aeruginosa*
 - cytotoxin, 68, 119, 120, 128, 129
 - cytotoxin, effect on cultured heart cells, rat, 247–251
 - protection by *Pseudomonas* Igs, 247–249, 251
 - septic shock, chemiluminescence-inducing radicals, pig, 339–334
 - immunoglobulin and plasmapheresis therapy, hemodynamic effects during treatment for septic shock, 1025–1029
 - oxygen supply-uptake relationship, septic shock, 182
- Pulmonary. *See also* Cardiopulmonary entries; Lung entries
- Pulmonary artery, sheep, neutrophils, thrombin-induced adhesion with endothelial cells, 102
- Pulmonary artery pressure
 - cf. blood flow, pulmonary vascular resistance, ARDS in septicemia, 175–179
 - cardiopulmonary response to endotoxin, eicosanoids in, sheep, 202
 - extravascular lung water, altered fluid regimen, advanced septic shock with acute respiratory failure, 804
 - hydroxyethyl starch, volume replacement in ovine endotoxemia, 816, 817
 - lung injury in *E. coli* endotoxemia, 360, 364, 365, 368
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 471
- Pulmonary artery wedge pressure and extravascular lung water
 - altered fluid regimen, advanced septic shock with acute respiratory failure, 803–808
 - large volume replacement with crystalloids, 809–812
- Pulmonary capillary pressure, pulmonary venous hemodynamics and gas exchange disturbances, *E. coli* septicemia, Goettingen pig, 185, 186
- Pulmonary capillary wedge pressure, heart function changes, septic shock, chacma baboon (*Papio ursinus*), 217, 219–222
- Pulmonary circulation, ARDS C3a and alveoli in, post-trauma, 45
- Pulmonary contusion, ARDS, C3a and alveoli in, post-trauma, 46, 47
- Pulmonary dysfunction, minimal, 52
- Pulmonary edema in shock, 9, 28–29, 31
 - arachidonic acid cascade in ARDS, 306–307, 309–311
 - capillary pressure, 28
 - see also* Extravascular lung water; Permeability
- Pulmonary failure prediction, elastase- α_1 -PI complex and neopterin, plasma levels, 683, 684, 687
- Pulmonary function, muscle pO₂ role, critically ill patients, 139
- Pulmonary hypertension. *See* Hypertension, pulmonary
- Pulmonary resistance, lung injury in *E. coli* endotoxemia, 358, 359
- Pulmonary vascular pressure, nifedipine for ARDS, 1089
- Pulmonary vascular resistance
 - ARDS in septicemia, 175–179
 - CI, 175–179
 - MPP, 175–178
 - oxygen supply, peripheral, 175, 177, 179
 - pulmonary artery pressure cf. blood flow, 175–179
 - endotoxin response, 390
 - α -mercaptopropionylglycine in hemorrhagic shock, 901, 902
 - recombinant hirudin/eglin C, endotoxin shock, 939, 941
- Pulmonary venous hemodynamics and gas exchange disturbances, *E. coli* septicemia, Goettingen pig, 185–188
 - atrial pressure, left, 185, 186
 - CI, 185, 186
 - hemodynamic parameters, 186

- MPP, 185, 186
 pulmonary capillary pressure, 185, 186
- Quin 2, 1055–1056
- RA642, acute hemorrhagic shock, rats,
 cerebrocortical perfusion, 1091–1904
- Radicals, chemiluminescence-inducing,
 septic shock, pigs, 339–344; *see also*
 Oxygen radicals
- Reanimation, MSOF prognostic indices,
 logistic regression analysis, 644, 646
- Receptor autoradiography, in situ,
 hepatocyte protein kinase C and
 diacylglycerol accumulation,
 endotoxemia, 579, 583–584, 586
- Red blood cell acetylcholinesterase,
 activated C3 in DIC in septic shock,
 fulminant meningococcal meningitis,
 272, 274
- Regional blood flow, ischemia and
 circulatory system in MSOF,
 1080–1081, 1083
- Relaxing factors, endothelium-derived, 157
- Respiratory distress, nebulized
 corticosteroid, 867–871
- Respiratory distress syndrome, neonatal,
 exogenous surfactant, 797; *see also*
 ARDS
- Respiratory failure
 early ventilatory support, 784–789
 embolism, fat, pulmonary, 37, 41
 extravascular lung water, altered fluid
 regimen, 803–808
- Respiratory quotient (CO₂/O₂ exchange
 ratio) in shock, 607–610, 613–617,
 619–621
 amino acid metabolism, 619–621
 base excess, 613, 614, 619
 expired minute volume, 613, 614,
 619–621
 fat metabolism, 619–621
 glucose metabolism, 619–621
 hemoglobin, 608
 hyperventilation, 615, 619–621
 low-flow states, 615, 616
 pH, 608, 609, 615–617
 RRE, 613–617
 TBRE, 613–617
- Reticuloendothelial system
 endotoxin, dose-related effects, 407–411
 liver clearance, scintigraphic evaluation
 of posttraumatic function,
 775–780
 stimulation to protect against DIC,
 1001–1005
- Rheumatoid arthritis, 332
 C3a, 299
- Right ventricle, heart dysfunction, septic
 shock, human 194
- RO-193,430, 704
- Ronipamil, traumatic shock, 1067, 1068
- Salmonella*
abortus equi, 408
 antibodies, anti-LPS and anti-lipid A,
 determination with
 immunoblotting, 1044, 1046
 LPS, 119, 121
enteriditis, 426–428, 430, 1054
minnesota, antibodies, anti-LPS and
 anti-lipid A, determination with
 immunoblotting, 1044–1049
typhimurium, 428
- Sanarelli-Schwartzman reaction, 1003, 1005
- SAPS scoring system, 626–628, 1026,
 1027, 1029
 dopamine infusion, effect evaluation
 with systolic time intervals, 1102
- Scintigraphic evaluation of posttraumatic
 liver function, prognosis/prognostic
 indices, 775–780
- Scoring systems
 cardiogenic/septic shock, 625–630
 CI and SVR, 625, 626
 elastase α_1 -PI complex and neopterin,
 plasma levels in MSOF, 683–687
 prognostic value of antithrombin III and
 endotoxin in sepsis, 637
 lethality, 639, 640
 thrombocyte counts, 637–641
see also Multiple system organ failure,
 postoperative/posttrauma,
 biochemical analysis and scoring;
 Prognosis/prognostic indices;
specific systems
- S-creatinine, MSOF prognostic indices,
 logistic regression analysis, 644, 645

- SDZ 63-441, 429
- Secretin, 380
- Sensitization, passive, WEB 2086 (PAF antagonist), anaphylactic lung reaction, 926, 928
- Sensorium, altered, corticosteroids in severe sepsis, 841, 843, 853-855
- Sephadex inflammation model, rats, 4-hydroxy-nonenal, 351-355
- Sepsis
- DIT marker in, 711-713
 - early, overwhelming inflammatory response, endotoxin role, 371-375
 - elastase- α_1 -PI as early indicator, 690-692
 - endocrine secretion patterns, 751-756
 - ADH, 751, 753, 755, 756
 - prolactin, 751, 753-754, 756
 - thyroid hormones, 751-756
 - endotoxin in complement activation, 277-281
 - kidney metabolism in, 601-605
 - metabolic abnormalities, *See* Metabolic abnormalities in sepsis
 - multiple organ system failure, 413-416
 - ARDS, 61-65
 - prognostic indexes, 633-636
 - IL-1, 715-718
 - plasma proteins, 719-724
 - proteinase inhibitor complex, immunologic determination, humans, 972-974
 - scintigraphic evaluation of posttraumatic function, liver, 775-780
 - severe, corticosteroids in, 840-844, 847-855
 - see also* Immunoglobulin profiles and PMN-elastase in septic gas gangrene
- Septicemia
- antibodies, anti-LPS and anti-lipid A, determination with immunoblotting, 1048-1050
 - ARDS, pulmonary vascular resistance, 175-179
 - coagulation cascade, 383-387
 - E. coli* hemolysin injury to lung, 67-70
 - transmembrane pores, 69
 - fibrinolysis, 383-387
 - gas exchange, disturbed, 185-188
 - MSOF and, endotoxin and proteases in, 315-321
 - T cell changes, 989, 990
- Septic shock
- antithrombin III and plasma substitution, 965-968
 - ARDS, 30-33
 - corticosteroids in, 857, 860-864
 - PMN migration, 30-31
 - cardiac dysfunction, 191-197, 207-215, 217-222, 237-245, 259-263
 - cardiac work, 260-262
 - chemiluminescence-inducing radicals, pig, 339-344
 - complement activation in MSOF, 291-296
 - DIC in, and C3, 271-274
 - dopamine infusion, effect evaluation with systolic time intervals, 1101-1105
 - eglin C, 945-948
 - extravascular lung water
 - altered fluid regimen, 803-808
 - large volume replacement with crystalloids, 809-813
 - immunoglobulin therapy
 - after cardiac surgery, 1034
 - and plasmapheresis, 1025-1030
 - immunomodulation, 989-993
 - lung microvascular bed, cell interactions, 113-118
 - metabolic rate, 260-262
 - oxygen extraction, peripheral, 260-262
 - oxygen supply-uptake relationship, 181-183
 - peripheral circulation, 163-171
 - peripheral resistance, total, 260, 262
 - scoring systems, 625-630
 - CI and SVR, 625, 626
 - TNF, 463-465
- Septic syndrome, definition, 857-860
- Serotonin. *See* 5-HT (serotonin)
- Serratia marcescens*, anti-LPS and anti-lipid A antibodies, determination with immunoblotting, 1044-1048
- SGOT, liver, TNF induction of organ changes in chronic lymph fistula, 469, 474, 475, 479

- SH-groups, α -mercaptopyrionylglycine in hemorrhagic shock, 897, 902, 903
- Silver sulfadiazine, 378
- Simplified Acute Physiologic Score, 280, 635
- c-sis*, neutrophils, thrombin-induced adhesion with endothelial cells, 109
- Small intestine, complement activation, MSOF pathogenesis in septic shock, dogs, 293–294, 296
- Sodium excretion, dopamine and renal function, 1097–1099
- Sodium polyanethol sulfonate-induced shock, 1001–1005
- Somatostatin, polytrauma, 743, 745, 747
- Somatostatin, glucose turnover in sepsis, 553, 556
- Sonomicrometer LV dimension, heart performance during septic shock, awake sheep, 237–243
- Spleen, zymosan-induced MSOF, endotoxin plays no key role, 421
- SRI 63-072, 428, 485
- SRI 63-441, 368, 428, 485
- SRI 63-675, 557
- SSS scoring system, 626–628, 664, 1026, 1027, 1029
- dopamine infusion, effect evaluation with systolic time intervals, 1102
- polytrauma, 744–746
- Staphylococcus aureus*, 420, 868
- alpha toxin, 68, 69, 119, 120, 123, 124, 127–130, 286
- xlyosus*, 420
- Streptococcus, 831
- Streptomycin, GI tract decontamination, MOF prevention, 828–832
- Stress ulcer diseases, correlation with, gastric mucosa ulceration, ultrastructure after septic shock, rat, 151
- Stroke volume
- heart function changes, septic shock, chacma baboon (*Papio ursinus*), 207, 210–214, 219, 221–222
- α -mercaptopyrionylglycine in hemorrhagic shock, 900, 902
- RA642, effects on cerebrocortical perfusion, acute hemorrhagic shock, 1094
- septic shock, human, 193, 196, 197
- Superoxide anion, 886
- generation, PAF and, 488
- 4-hydroxy-nonenal, 354, 355
- lipid peroxidation, hypovolemic-traumatic shock, dogs, 346
- metabolic abnormalities in sepsis, 535, 536
- PMN production
- corticosteroid, nebulized, in experimental respiratory distress, 869, 870
- recombinant human SOD in endotoxemia, 913, 917
- Superoxide dismutase, 887
- PMA lung injury, 945–949
- recombinant human, in *E. coli* endotoxemia, rat, 913–917
- Surface epithelial cells, gastric mucosa ulceration, ultrastructure after septic shock, rat, 153, 154
- Surfactant, ARDS C3a and alveoli in, post-trauma, 44, 45
- Surfactant, exogenous
- acute high-permeability lung edema, 791–795
- ARDS, 29
- cf. neonatal RDS, 797
- aspiration trauma, experimental, rabbit, 797–800
- phosphatidylcholine and phosphatidyl glycerol, 792, 798
- porcine, 798
- Surfactometer, bubble, 792
- Systemic lupus erythematosus, C3a, 299
- Systemic vascular resistance, 625, 626 and heart function changes, septic shock, chacma baboon (*Papio ursinus*), 219, 220
- immunoglobulin and plasmapheresis therapy, hemodynamic effects during treatment for septic shock, 1026–1029
- α -mercaptopyrionylglycine in hemorrhagic shock, 900–902
- septic shock, 163–166, 168, 170
- Tachycardia and cardiac volume, heart

- function changes, septic shock, chacma baboon (*Papio ursinus*), 214–215, 217–222
- TAN concentrations, kidney metabolism in *E. coli* sepsis, 601, 602, 604
- Target structure degradation, multiple system organ failure, postoperative/posttrauma, biochemical analysis and scoring, 662–663
- T cell(s)
 - immune suppression/dysfunction, polytrauma, 517–521
 - multiple trauma, early events, 507, 508
 - septicemia/septic shock, 989, 990
 - subsets, 508, 510, 518–520
 - Ig synthesis suppression after multiple trauma, 513–514, 516
 - MSOF, postoperative/posttrauma, biochemical analysis and scoring, 661
 - PAF and, 442
 - in septic shock, 989, 991, 993
 - thymopentin (TP-5), post-burn and postoperative sepsis and immunodeficiency syndrome, 995, 998
 - trauma-induced cascade of cell-mediated immune effects, 496–505
- T-cell replacing factor, 497
- Tebonin, effect in burns, 455–461
- Terminal complement complex
 - acute pancreatitis, 266, 267
 - pulmonary hypertension and vascular leakage, 283–286
- Theophylline, 755
- Thiobarbituric acid reactive material, lipid peroxidation, hypovolemic-traumatic shock, dogs, 345, 346
- Thiol groups, α -mercaptopropionylglycine in hemorrhagic shock, 897, 902, 903
- Thrombin, 367, 937, 941, 942
 - MSOF, postoperative/posttrauma, 653
- Thrombin-antithrombin III complex
 - antithrombin III and plasma substitution in septic shock, 966–968
 - immunologic determination in humans, 971–974
- Thrombin-induced adhesion with endothelial cells, neutrophils, 101–109
- Thrombocyte counts, prognostic value in sepsis, 637–641
- Thromboplastin, aprotinin membrane protective action, intraoperative histamine liberation, 961, 962
- Thromboxane, 429
 - alveolar permeability increased by PMA-stimulated neutrophils, rabbits, ARDS, 328
 - burns
 - enteric translocation of microorganisms, 379
 - PAF inhibitor effect, scalded pig, 455, 459, 460
 - cardiopulmonary response to endotoxin, eicosanoids in, sheep, 201–204
 - circulation, peripheral, septic shock, 168
 - complement activation, pulmonary hypertension and vascular leakage, 285
 - early ventilatory support, 786
 - eglin C ineffectiveness in endotoxin shock, 955
 - hemofiltration and survival time, porcine acute endotoxic shock, 823–825
 - liver dysfunction in MSOF, altered cell-cell interactions, 569
 - lung, endotoxin-induced microvascular endothelial injury, 92
 - lung injury in *E. coli* endotoxemia, 359, 362–367
 - and hypertension, 67, 70
 - myocardial ischemia, 907–911
 - PAF antagonists in endotoxin shock, 935
 - permeability, pulmonary vascular, mediators, 310
 - TNF, induction of organ changes in chronic lymph fistula, sheep, 469, 474, 475, 479
- Thromboxane receptor blockade, lung injury in *E. coli* endotoxemia, 366, 367
- Thromboxane synthetase inhibition, lung injury in *E. coli* endotoxemia, 365–366
- Thymopentin (TP-5) immunomodulation, post-burn and postoperative sepsis and immunodeficiency syndrome, 995–998

- Thymostimulin (TP-1), immunodulation in septic shock, 990–993
- Thyroid hormones
and leukocyte phagocytic activity, DIT and
T₃, 712
T₄, 711–712
polytrauma, 743, 745
T3, 745, 748
T4, 744, 745, 748
TBG, 744, 745, 749
TSH, 744, 745, 749
sepsis, 751–756
T3, 751–756
T4, 752–754
TSH, 752, 753, 755
- TISS, 643, 645
- Tissue oxygen debt (VO₂ deficit) as determinant, organ failure, postoperative, 133–136
survivors cf. nonsurvivors, 136
- Total parenteral nutrition, 377
metabolic abnormalities in sepsis, 537, 538
- Transfer factor, immunomodulation in septic shock, 990–993
- Transferrin, plasma, and abdominal sepsis prognosis, 720, 721
- Transpulmonary pressure, lung injury in *E. coli* endotoxemia, 358–361
- Trauma
endotoxin and overwhelming inflammatory response of early sepsis, 372–374
-induced cascade of cell-mediated immune effects, immune suppression/dysfunction, 495–505
immunorestitution, 504–505
schema, 501, 503
cf. multiple system organ failure, 58–60
see also Multiple system organ failure, postoperative/posttrauma, biochemical analysis and scoring; Polytrauma
- Trauma score, 643
- Traumatic shock
calcium antagonists in shock/ischemia, 1067–1068
hypovolemic, 228, 229
- PAF antagonist inhibition of induced shock, 431
- Trifluoroperazine, 120, 122, 1070
- Trimethoprim, GI tract decontamination, MOF prevention, 828–832
- Tumor necrosis factor (TNF, cachectin), 13, 14, 114, 368, 432, 444, 536, 661, 850, 851
administration, metabolic abnormalities in sepsis, 550, 551
circulation, peripheral, septic shock, 169
discovery, 463
endotoxin as stimulus, 463, 464
glucose turnover in sepsis, 555
heart dysfunction, septic shock, human, 197
lipoprotein lipase suppression, 464
liver dysfunction in MSOF, altered cell-cell interactions, 564, 568–570
lung injury, endotoxin-induced microvascular endothelial, 91, 95–96
production and macrophage/monocyte induction in shock, PAF, 485–488
PAF antagonist effects, 485, 486
pulmonary vascular permeability, 305
in septic shock, 463–465
serum, and sepsis, prognosis/prognostic indices, 715–718
WEB 2086 (PAF antagonist), 919–921
window phenomenon with PAF, 921–922
- Tumor necrosis factor, induction of organ changes in chronic lymph fistula, sheep, 467–481
disseminated intravascular coagulation, 479, 480
elastase-a₁-anti-proteinase complex, 470, 477–478, 480
hemodynamics, 472, 479
cf. human, 470, 477
cf. in vitro, 470, 477
kallikrein, 469, 474, 479
leucocyte role, 469, 470, 475–477, 480
leucostasis, liver, 473, 474, 480
liver SGOT, 469, 474, 475, 480
lung lymph, 469, 473, 475, 479
oxygen radicals, 470, 476, 477

1142 / Index

- permeability, microvascular, 472, 473, 479–481
- prekallikrein, 469, 474, 475, 479
- pulmonary artery pressure, 471
- thromboxane, 469, 474, 475, 479
- Typhoid fever, 852
- Tyrosine, amino acid concentrations, serum, experimental endotoxin shock, 598
- U46619, 362, 368
- U74006F, 891–895
- Ulceration, gastric mucosa
 - ultrastructure after septic shock, 151–155
 - and stress ulcer disease, 151
 - and WEB 2086 (PAF antagonist), 919
- Ultraviolet-absorption spectra, positive inotropic factor as myocardial stimulant, 255, 256
- Urea, MSOF prognostic indices, logistic regression analysis, 644–646
- Urokinase-type plasminogen activator, neutrophils, thrombin-induced adhesion with endothelial cells, 109
- U.S. Veterans Administration, sepsis studies, corticosteroids, 840–844, 847–855, 873
- Vascular intima in endotoxin shock, 77–87
 - cell origins and replacement, 78–80
 - endothelial injury, grading, 84–85
 - endotoxin transport and elimination, 81
 - generalized inflammation, 85–87
 - aorta, 85, 86
 - non-endothelial cells, 77–78
 - macrophages, 78, 80, 81
 - ultrastructural alterations, early, 82–84
- Vascular permeability. *See* Permeability
- Vascular tone, hemorrhagic shock, phase-related vascular reactivity, cats, 143
- Vasopressin (antidiuretic hormone), 379
 - hepatocytes, protein kinase C and diacylglycerol accumulation in endotoxemia, 576, 579–581, 584–586
 - sepsis, 751, 753, 755, 756
- Ventilatory support, early
 - inversed ratio ventilation, 788, 789
 - multiple organ failure with acute respiratory failure, 784–789
- PEEP, 788–789
- Ventricular pressure cf. perfusion flows, endotoxin, inotropic effect in isolated rabbit heart, 227, 228
- Ventriculography, radionuclide, heart function changes in septic shock, chacma baboon (*Papio ursinus*), 209–210, 218, 219
- Verapamil, 1060
 - endotoxin shock, 1068, 1069
 - pretreatment, calcium antagonists in shock/ischemia, 1067
- Virchow's triad, 851
- Vitamin E
 - leukocyte-induced lung injury, 74–76
 - lipid peroxidation inhibition in cardiopulmonary arrest, 893–895
- W7, 120
- WEB 2086 (PAF antagonist), 426, 428, 429, 442, 485
 - anaphylactic lung reaction, guinea pig, 925–929
 - endotoxin shock effects, 931–933, 935
 - gastrointestinal tract damage, endotoxin-induced, 919–922
 - hetrazepine, 925
- WEB 2170, 928
- Weibel-Palade bodies, 83
- White blood cell count, 725, 728
- Wilhelmy tensiometer, 800
- Xanthine-oxidase, 349
- Zymosan
 - induced peritonitis, decontamination, GI tract, 827–832
 - leukocyte-induced lung injury 73–76