H. Müller, J. Zierski, R. D. Penn (Editors) Local-spinal Therapy of Spasticity

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## The Neuropharmacology of Baclofen

W. Zieglgänsberger , J. R. Howe, B. Sutor

### Introduction

Gamma-aminobutyric acid (GABA) is one of the major inhibitory neurotransmitters in the mammalian central nervous system [see: 36, 69, 92, 98]. GABAergic neurons have been identified throughout the central nervous system with histochemical techniques [see: 75]. GABA is, however, of no therapeutical value because the amino acid does not pass the blood-brain barrier in sufficient amounts to affect neuronal excitability [9, 64].

The GABA analogue baclofen ( $\beta$ -[4chlorophenyl]-gamma-aminobutyric acid; Lioresal<sup>®</sup>) was designed to act as a GABA mimetic that because of its lipophilicity would distribute into the central nervous system after systemic application. Baclofen effectively reduces exaggerated stretch reflexes and muscle tone after oral and intravenous administration and is widely used in the treatment of spasticity caused by traumatic spinal lesions, degenerative, neoplastic or infectious diseases of the spinal cord, and multiple sclerosis. It is less effective in ameliorating the spasticity after stroke or cerebral palsy [see: 109] (see also various authors, this volume).

Baclofen reduces muscle tone in patients with spinal transections and reduces muscle rigidity and tonic stretch reflexes in decerebrated animals [see: 13]. It is therefore suggested that the therapeutically relevant effect of baclofen results from a direct action at the spinal level. In addition to its antispastic activity, however, baclofen can cause muscle weakness, ataxia, drowsiness, insomnia, nausea, hypertension, a decrease in growth hormone release and an increase in prolactin secretion. Also, antinociceptive actions of baclofen have been reported [107; 110] (see also Yaksh, this volume). Particularly in elderly patients, the sudden withdrawal of baclofen after chronic use is occasionally associated with the appearance of psychotic symptoms, including dysphoric episodes and even hallucinations [see: 12]. These latter effects clearly indicate that baclofen acts also on receptors remote from the spinal cord.

This chapter examines the neuronal processes affected by baclofen, with an emphasis on electrophysiological findings. The enormous literature precludes an exhaustive documentation of all aspects.

### The GABA<sub>B</sub> Receptor

The inhibitory actions of GABA are mediated in most neurons studied until now through an increase in chloride conductance of the postsynaptic membrane [see: 39, 69, 101]. These actions are mimicked by muscimol and THIP (4,5,6,7-tetrahydroisoxazolo-[5,4-c]pyridine-3[2H]-one) and can be antagonized by bicuculline and picrotoxin [26; see: 36]. These two antagonists seem to block the actions of GABA via different mechanisms. Whereas bicuculline competes for the GABA binding site [74], picrotoxin seems to interact with the chloride-channel in a more direct manner [see: 36]. Because GABA also evokes bicuculline-insensitive responses which are mimicked by baclofen (e.g. on transmitter release) [15, 67], two classes of GABA receptors,  $GABA_A$  and  $GABA_B$ , have been proposed. The bicuculline-reversible actions of GABA and the GABA mimetic muscimol are mediated through GABA<sub>A</sub> receptors, whereas baclofen is considered as a prototypical bicuculline-insensitive agonist at GABA<sub>B</sub> receptors [15, 54]. This novel GABA<sub>B</sub> receptor mediates ionic processes which are clearly distinct from those sensitive to bicuculline (see below). GABA<sub>A</sub> receptor-mediated inhibition is potentiated by barbiturates and by the benzodiazepines [see: 48, 49] and is blocked by some convulsants like penicillin or pentetrazol [see: 36, 68]. Unfortunately, a selective  $GABA_B$ receptor antagonist is not yet available.

Data from binding studies also support the existence of different types of GABA receptors. Various studies have corroborated the initial finding that baclofen does not displace <sup>3</sup>(H)-GABA from its binding sites on neuronal membranes [111]. Furthermore, the binding of GABA is enhanced by benzodiazepines [see: 48], whereas the binding of baclofen is unaffected by these compounds [108].

Autoradiographic studies with  ${}^{3}$ (H)baclofen have shown that, with a few exceptions, the distribution of GABA<sub>A</sub> and GABA<sub>B</sub> binding sites overlap in most regions of the brain [46]. There are indications, however, that the neuronal distribution of these bindng sites are not identical. For example, GABA<sub>A</sub> binding and high affinity GABA<sub>B</sub> binding were unchanged by interruption of forebrain noradrenergic projections, whereas low affinity GABA<sub>B</sub> binding was reduced [62]. This suggests that these low affinity  $GABA_B$  receptor binding sites are located presynaptically on noradrenergic terminals. Baclofen reduces the evoked release of noradrenaline [15, 16, 40] and several other neurotransmitters (see below).

### Stereoselectivity of GABA<sub>B</sub> Actions

Therapeutically used baclofen (LIO-**RESAL**<sup>®</sup>) is a racemic mixture of the two isomers. In a number of behavioral studies and electrophysiological investigations in vivo, it has been shown that the (-)isomer of baclofen is more potent than the (+)isomer [4, 52, 53, 80, 81, 107] (see also Yaksh, this volume). Several in vitro electrophysiologic studies have shown that the (-)isomer is at least 100-fold more potent than the (+) isomer [5-7, 20, 47, 56, 59, 78]. Similar stereoselectivity was demonstrated in experiments where baclofen reduced the in vitro release of exogeneously loaded <sup>3</sup>(H)-neurotransmitters [15, 61] and in studies of baclofen binding to bicuculline-insensitive receptors on synaptic membranes [54].

It was reported by various authors that the (+)isomer can antagonize the actions of the (-)isomer [94, 104, 105] (see also Yaksh, this volume). In recent electrophysiological experiments in which intracellular recording techniques were employed however, no such antagonism could be demonstrated [4, 58]. Thus (+)-baclofen does not appear to be an antagonist at all GABA<sub>B</sub> receptors.

## Mode of Action of Baclofen

Numerous electrophysiological studies have shown that baclofen has profound inhibitory effects on synaptic transmission in the spinal cord and many other regions of the mammalian central nervous system. Until recently, the majority of the results suggested that baclofen acted presynaptically to selectively reduce excitatory synaptic transmission, a conclusion consistent with reports that baclofen directly reduces the release of excitatory neurotransmitters. It is now clear, however, that baclofen can also reduce inhibitory synaptic transmission and that baclofen directly increases the postsynaptic potassium conductance of many central neurons. In addition, there are several reports that baclofen depresses the firing of central neurons induced by excitatory substances. The following sections review the evidence for each these actions of baclofen and their relevance to baclofen's effects on central nervous function.

## Actions of Baclofen and Synaptic Transmission

In the first study employing intracellular recording techniques, Pierau and Zimmermann [85] reported that baclofen depressed excitatory postsynaptic potentials (EPSPs) evoked in cat motoneurons at doses that did not affect inhibitory processes. Since neither the membrane potential  $(E_m)$ , input resistance  $(R_N)$ , nor the direct excitability (action potentials evoked by intracellular current injection) of these cells were affected by baclofen, these authors concluded that baclofen exerts its action through presynaptically located receptors. The occasionally observed slight hyperpolarizations were interpreted as a disfacilitatory effect caused by inhibition of excitatory interneurons.

In several subsequent in vivo and in vitro electrophysiological studies employing extracellular recording techniques in various structures, further evidence was provided that baclofen preferentially reduces excitatory synaptic transmission by a presynaptic mechanism [5-7, 29, 30, 42, 43, 52, 53, 63, 70, 82-84, 86, 88]. Thus baclofen was shown to produce profound reductions of responses to orthodromic stimulation of excitatory afferent pathways without significantly affecting responses to antidromic responses or presnyaptic fiber volleys [5, 6, 29, 42, 70, 82]. Synaptically evoked responses were shown to be reduced at doses or concentrations of baclofen that had little or no effect on chemically evoked excitation or spontaneous firing [30, 42, 52, 53, 82].

The selectivity of baclofen's depressions of synaptically evoked responses in some structures was also interpreted as being inconsistent with a postsynaptic depressant action of baclofen. Electrophysiological experiments in slice preparations of the hippocampus suggest that baclofen selectively inhibits transmission at putative glutamatergic synapses (Lanthorn and Cotman 1981). In this study, only the CA3-projections to pyramidal cells in the CA1 region (Schaffer collaterals) and mossy fiber synaptic transmission were inhibited [cf. 60]. The excitatory synaptic transmission from the lateral perforant path was insensitive to baclofen. A selective action of baclofen on synaptic tranmission mediated by excitatory amino acids is also suggested by the findings of Ault and Evans [3]. These authors described that baclofen reduced dorsal root potentials recorded from the neonatal isolated spinal cord, potentials which are also reduced by excitatory amino acid antagonists [38], whereas the excitatory responses of cervical ganglion neurons to preganglionic stimulation is not affected even by much higher concentrations. Both excitatory afferent inputs from descending pathways employing still unknown transmitters and cholinergic inputs to motoneurons are not affected by baclofen [11, 63].

In addition to these various extracellular studies, there are now several reports from investigations in which intracellular recordings were obtained that baclofen depresses EPSPs in mammalian central neurons [14, 42, 47, 55, 56, 60, 65, 73, 85, 102]. Although in some of these studies baclofen also hyperpolarized the cells (see below), it was shown that the depressions of EPSPs were not a direct consequence of these hyperpolarizations [14, 60]. In neocortical neurons, baclofen's reductions of EPSP amplitudes were independent of membrane potential over the range of values  $\pm 30 \text{ mV}$ from resting  $E_m$  [56]. In our studies, baclofen applications that produced 70 to 100% depressions of EPSPs typically produced only 20 to 30% decreases in  $R_N$ ; baclofen's depressions of EPSPs typically outlasted its effects on  $E_m$  and  $R_N$  for several minutes. Thus in neocortical neurons, baclofen's reductions of EPSP amplitudes do not appear to be solely the consequence of its action to increase postsynaptic conductance. Another possible interpretation of such findings, however, is that baclofen preferentially increases dendritic conductance, and that only a portion of this conductance increase is detected from intracellular recordings at the neuronal soma. Such an interpretation has been made of results obtained in the hippocampus [47].

The most detailed analyses of baclofen's effects on excitatory synaptic transmission have been performed on nonmammalian preparations. Shapovalov and Shiriaev [97] studied the effect of baclofen on monosynaptic EPSPs in the frog motoneuron. These EPSPs are composed of both a chemically mediated and an electrically mediated component. Baclofen produced marked reductions of the chemically mediated component, but had no effect on the electrically mediated component of these EPSPs. From statistical analyses of singlefiber EPSPs, Shapovalov and Shiriaev concluded that baclofen acted presynaptically to reduce the release of the transmitter which generates chemically mediated EPSPs at these synapses. At the crayfish neuromuscular junction, Barry [8] found that baclofen depressed excitatory transmission without affecting the input resistance of the muscle. She was further able to show that baclofen reduced the frequency of spontaneous excitatory junction potentials without affecting their size, results which clearly indicate a presynaptic site of action. At present, the complexity of mammalian preparations has prohibited these sort of detailed analyses.

Fox et al. [42] noted that, in addition to baclofen's consistent reductions of EPSPs, baclofen also reduced the amplitude of IPSPs recorded in some cells. That baclofen reduces stimulation-evoked GABAergic inhibition in the olfactory cortex and the hippocampus was suggested from extracellular recordings of field potentials [6, 24]. Intracellular studies in vitro have shown that baclofen reduces the amplitude of short-latency GABA<sub>A</sub> receptor-mediated IPSPs evoked in various hippocampal neurons [14, 60, 73], neurons in the olfactory cortex [96], and neurons in the frontal neocortex [55, 56, 102]. These depressions of IPSP amplitudes are not due to the concomitant action of baclofen to hyperpolarize these neurons. Blaxter and Carlen [14] reported that baclofen's depressions of short-latency GABAergic IPSPs in hippocampal neurons persisted when the membrane potential was returned to its resting value by direct current injection. Short-latency IPSPs in rat neocortical neurons are also GABA<sub>A</sub> receptor-me-

diated [100, 101], and baclofen's depressions of these IPSPs were independent of membrane potential between values of -50 and -110 mV [56]. Importantly, baclofen's reductions of short-latency GABAergic IPSPs in olfactory cortical and neocortical neurons were shown to be accompanied by reductions in the conductance increases measured during these IPSPs [56, 96]. Because baclofen does not reduce responses to exogenously applied GABA or muscimol in these same neurons (see below), a postsynaptic blockade of the IPSP conductance can be excluded and these results argue strongly that baclofen reduces GABA<sub>A</sub> receptor-mediated IPSPs by a presynaptic action.

In addition to depressing chloride-dependent GABA<sub>A</sub> receptor-mediated IPSPs, baclofen has also been reported to decrease the amplitude of potassium-dependent long-latency IPSPs (slow IPSPs) in hippocampal neurons [14, 60]. Similar potassium-dependent long-latency IPSPs (times to peak of 150 to 250 ms) are also evoked in neocortical neurons [57], and baclofen consistently and markedly reduces the amplitude of these IPSPs [56, 57]. As for reductions of GABA<sub>A</sub> receptor-mediated IPSPs in these neurons, baclofen's reductions of long-latency IPSPs are independent of membrane potential and are accompanied by a reduction of the conductance increases associated with these IPSPs [56, 57].

Despite baclofen's action to reduce IPSPs, in most neurons studied baclofen's action to depress EPSPs appears to predominate and baclofen causes an increase in the stimulation intensity required to produce a synaptically evoked action potential [42, 47, 57, 85]. These baclofen-induced increases in action potential stimulation thresholds are consistent with the many extracellular studies cited above in which it was found that baclofen decreased synaptic excitability. There are reports, however, that baclofen can either increase or deaction potential crease stimulation thresholds, depending on the cell population investigated and the concentration of baclofen applied [60, 73]. Although baclofen increased the stimulation threshold of synaptically evoked action potentials in

virtually every neocortical neuron tested, baclofen often caused an increase in the number of action potentials produced by suprathreshold stimulation intensities due to baclofen's blockade of IPSPs [56]. Thus there are circumstances in which baclofen can in fact produce increases rather than decreases in synaptic excitability. The action of baclofen is clearly distinguishable from GABA<sub>A</sub> receptor antagonists such as bicuculline, however, which commonly produces significant reductions in action potential stimulation thresholds and promotes the generation of epileptiform bursts of action potentials in mammalian central neurons. In contrast, baclofen has been shown block bicuculline-induced to epileptiform activity in in vitro preparations of the hippocampus [6, 7, 20; but see: 73] and the frontal neocortex [56]. These results also indicate that baclofen's reductions of excitatory synaptic transmission are resistant to blockade by bicuculline and indeed baclofen's reductions of EPSP amplitudes are not antagonized by this GABA<sub>A</sub> receptor antagonist [56].

The advent of *in vitro* slice preparations of the mammalian CNS has made it possible to evaluate the action of known concentrations of drugs on neurons in these preparations under steady state conditions. The EC<sub>50</sub> for baclofen's depressions of excitatory synaptic transmission is approximately 1 µM and significant depressions are observed at concentrations between 10 and 100 nM [5-7, 24, 47, 56, 70, 82]. These latter concentrations are approximately equal to those obtained in the cerebrospinal fluid after systemic administration of therapeutic doses in man [66, 103]. In our study of neocortical neurons, there was no apparent difference in the concentration dependence of baclofen's reductions of EPSPs and its reductions of either type of IPSP [56].

In summary, the majority of the studies on the action of baclofen on synaptic transmission seem to favor the conclusion that baclofen reduces synaptic transmission by a presynaptic action. It should be noted, however, that none of these studies provide direct evidence for such a mechanism, with the exceptions of the studies on the frog motoneuron and the crayfish neuromuscular junction. In most studies, it was concluded that the action was presynaptic either because no evidence of a postsynaptic action was found or the postsynaptic changes that were observed were considered to be insufficient to account for the effects on postsynaptic potentials. As we already mentioned, there is, however, direct evidence for effects of baclofen on neurotransmitter release and also recent evidence that baclofen has a direct effect on postsynaptic membrane conductance. In addition, there are several electrophysiological studies which have directly addressed the effects of baclofen on presynaptic afferent terminals or experimental models thereof. These results are presented in the following sections.

#### **Presynaptic Actions of Baclofen**

The most direct evidence for a presynaptic action of baclofen is its demonstrated inhibition of neurotransmitter release. Baclofen has been shown to decrease the evoked release of several putative neurotransmitters, including monoamines [15, 40, 94] and excitatory amino acids [24, 61, 82, 89, 90]. Baclofen's action to reduce the release of excitatory amino acids is consistent with baclofen's selective inhibition of synaptic transmission that is thought to be mediated by these excitatory amino acids.

Although baclofen depresses GABA<sub>A</sub> receptor-mediated IPSPs by an action which is not postsynaptic [56, 96] and dendritically located GABA<sub>B</sub> receptors have been demonstrated on central GABAergic neurons [19, 106], baclofen does not reduce the directly evoked release of GABA from brain slices [24, 61, 89]. Collins et al. [24] demonstrated, however, that baclofen significantly reduces GABA release that is evoked by electrical stimulation of excitatory afferents. They proposed that baclofen reduces stimulation-evoked GABA release and GABAergic inhibition via its direct action to reduce the release of excitatory amino acid neurotransmitters and consequently, the excitatory drive of GABA releasing interneurons [cf. 56, 96].

Several careful electrophysiological studies have addressed the mechanism of baclofen's putative presynaptic inhibition of synaptic transmission. Unfortunately these studies have not provided any consistent and positive evidence in this regard. The first suggestion that baclofen may activate presynaptic inhibitory mechanisms in the spinal cord by enhancing primary afferent depolarization as described for benzodiazepines [see: 49, 95] was abandoned after more experimental data were obtained. It was shown that baclofen depresses rather than enhances the excitability of primary afferents [22, 29, 42] and presynaptic inhibition was unchanged or reduced by baclofen [1, 71, 72].

Davidoff and Sears proposed that baclofen's depressions of afferent excitability were secondary to its action of hyperpolarizing presynaptic terminals and suggested that this action might account for baclofen's depressions of synaptic transmission [29]. The magnitude of baclofen's reductions of terminal excitability were considered to be insufficient, however, to account for its depressions of synaptically evoked responses [42]. The action potential invasion of the terminal region of afferent fibers does not seem to be impaired by baclofen, because its depressions of spinal monosynaptic reflex responses can be temporarily overcome by post-tetanic potentiation [22]. Shapovalov and Shiriaev also concluded that baclofen did not impair presynaptic terminal invasion [97].

The terminal region of primary afferent fibers carry GABA<sub>B</sub> binding sites [91] and GABA<sub>A</sub> and GABA<sub>B</sub> receptors coexist on the perikarya of small caliber primary afferent fibers [33]. However, due to their small size, primary afferent terminals have resisted analysis with intracellular recording techniques. An often used substitute for the analysis of ionic mechanisms that are supposed to occur in the terminal region are recordings from dorsal root ganglion cells in vivo and in vitro. In dorsal root ganglion neurons in culture, baclofen reduces the duration of the calcium component of action potentials [35]. Similar results were obtained in neurons of the myenteric plexus [23]. These results provide suggestive evidence that baclofen reduces neurotransmitter release by blocking inward calcium currents in presynaptic terminals [see also: 97].

Baclofen had no effect, however, on inward calcium currents recorded from cultured hippocampal neurons under voltageclamp conditions [45], nor on the duration of the calcium component of action potentials recorded from rat neocortical neurons [56]. Barry [8] concluded that baclofen's presynaptic inhibition of synaptic transmission at the crayfish neuromuscular junction was unlikely to be the result of an effect on presynaptic calcium influx. In an in vitro preparation of the hippocampus, baclofen reduced the extracellular calcium concentration measured with ion-sensitive microelectrodes ([50] decreases in extracellular calcium concentration reflect the movement of calcium ions into pre- and postsynaptic elements as a consequence of neuronal activity). In these experiments. however, consistent reductions in the presynaptic component of stimulation-evoked calcium entry were only observed when baclofen was applied at a concentration of 50 µM. This is approximately 50-fold greater than the  $EC_{50}$  for baclofen's depressions of synaptic transmission (see above). Thus the findings that baclofen reduces somatic calcium currents in dorsal root ganglion or myenteric plexus neurons do not necessarily extrapolate to CNS neurons, and at present there is little evidence to support the claim that baclofen inhibits transmitter release and, thereby, synaptic transmission by reducing presynaptic calcium influx.

### Postsynaptic Actions of Baclofen

Baclofen has a dose- and concentration-dependent hyperpolarizing action on some mammalian central neurons [65, 73, 79] which is associated with an increase of postsynaptic conductance [20, 44, 47, 56, 59, 76, 78, 87, 99]. This action is unaffected by blockade of synaptic transmission and is therefore indeed a direct postsynaptic action and not a disfacilitation secondary to removal of tonic excitatory input [20, 76, 78, 87, 99]. This action of baclofen results in decreases in the direct excitability of central neurons [56, 60]. The baclofen concentrations at which these effects become apparent are approximately equal

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to the concentrations at which baclofen produces effects on synaptic transmission  $(EC_{50} < 2 \mu M)$  [56, 59, 78, 99].

The mean reversal potential of baclofeninduced changes in  $E_m$  and the dependence of this reversal potential on the extracellular potassium concentration indicate that these changes are secondary to an increase in the conductance of the postsynaptic membrane to potassium ions [56, 59, 78, 99]. In contrast to GABA<sub>A</sub> receptormediated responses, the amplitude and reversal potential of baclofen-induced changes in  $E_m$  are unaffected by reductions of the extracellular chloride concentration [59, 78] or by intracellular injections of chloride ions [14, 76, 78].

It was suggested on the basis of current recordings from hippocampal clamp neurons that baclofen-induced conductance changes were voltage-dependent [59, 78]. This was verified directly in voltage clamp experiments on cultured hippocampal neurons in which baclofen was shown to activate a potassium conductance which is voltage-dependent and inward rectifying [45]. Baclofen-induced currents were also recorded under voltage clamp conditions from rat neocortical neurons [56]. The conductance activated by baclofen is sensitive to blockade by the potassium channel blockers 4-aminopyridine and barium ions; however, the present results indicate that it is different from any of the previously identified potassium conductances [45, 59, 78, 99; but see: 14].

Baclofen-induced increases in postsynaptic potassium conductance are insensitive to blockade by concentrations of GABA<sub>A</sub> receptor antagonists that completely antagonize GABA-mediated increases in chloride conductance [20, 44, 45, 56, 59, 76, 78, 87; but see: 60]. Pentobarbitone, which increases the effect of GABA on chloride-conductance, does not alter the action of baclofen [78]. GABA can, however, mimic the action of baclofen to increase postsynaptic potassium conductance when GABA<sub>A</sub> receptors are blocked with appropriate antagonists [44, 45, 78]. These results are consistent with the conclusion that the postsynaptic effects of baclofen are mediated by GABA<sub>B</sub> receptors.

The extent to which baclofen's action to increase postsynaptic potassium conductance contributes to its depressions of synaptic transmission in various structures or to the in vivo effects of baclofen is debatable. As noted, there are many studies in which baclofen produced profound effects on synaptic transmission and yet no evidence for a postsynaptic action was found. That an increase in postsynaptic potassium conductance is inhibitory is obvious, however, and in some neurons the hyperpolarizations observed are as great as 20 mV. It is unfortunate that in many of these studies the effects on synaptic responses were not investigated. Gähwiler and Brown [45] proposed that if baclofen increased the potassium conductance of presynaptic terminals, this action might indirectly lead to a decrease in presynaptic calcium influx by decreasing the duration of the action potential. In neocortical neurons, however, baclofen's depressions of postsynaptic potentials and its action to increase somatic potassium conductance could be temporally dissociated.

# Effect of Baclofen on Exogeneously Applied Neurotransmitters

Saito et al. [93] reported that, in low concentrations, baclofen selectively reduced the depolarizing actions of substance P in spinal motoneurons and suggested that it was a substance P antagonist. This suggestion was not supported by subsequent electrophysiological studies however [31, 37, 41, 42, 51, 84], and baclofen does not interfer with the binding of <sup>3</sup>H substance P [see: 17].

It was shown in several extracellularly conducted studies that baclofen can inhibit responses to exogenously applied excitatory neurotransmitters [27, 32, 41, 80, 83, 93]. In further such investigations, however, it was demonstrated that inhibitions of responses to these chemical excitants required doses or concentrations of baclofen significantly greater than those at which baclofen inhibited synaptically evoked responses [30, 42, 52, 82]. In our intracellular study of neocortical neurons, we found that applications of baclofen that produced virtually complete depressions of stimulation-evoked EPSPs did not significantly reduce depolarizations produced by L-glutamate, L-aspartate, or N-methyl-D-aspartate [56]. The observed occasional and modest reductions of depolarizations produced by these excitatory amino acids were similar in magnitude and duration to baclofen-induced decreases in direct excitability, thus suggesting that they were the result of baclofen's action to increase the postsynaptic potassium conductance of rat neocortical neurons. That baclofen does not postsynaptically block conductance increases produced by these substances is directly supported by our findings that baclofen had no effect on L-glutamate-evoked inward currents recorded in neocortical neurons under voltage clamp conditions.

Baclofen applications that reduce the conductance increases associated with stimulation-evoked GABAergic IPSPs have no effect on conductance increases produced by the direct application of GABA or the  $GABA_A$  agonist muscimol [56, 96]. Even at high concentrations, baclofen had no effect on currents evoked in neocortical

neurons by iontophoretically applied GABA [56].

# GABA<sub>B</sub> Receptor Mediated Synaptic Processes

It was proposed by Newberry and Nicoll [77, 78] that slow (long-latency) IPSPs evoked in hippocampal CA1 neurons and hyperpolarizations produced by baclofen may each be secondary to activation of GABA<sub>B</sub> receptors. According to this assumption, baclofen and the endogenous transmitter responsible for the long-latency IPSPs should act on the same population of postsynaptic receptors to increase the same postsynaptic potassium conductance. Due to the lack of established antagonists of either baclofen-induced hyperpolarizations or long-latency IPSPs, this possibility cannot be tested directly at present. Interestingly, however, baclofen reduces slow IPSPs in hippocampal neurons [14, 60] and similar IPSPs evoked in neocortical neurons [56, 57]. The mechanism of this effect of baclofen is unresolved.

### Conclusions

The presently available data indicates that baclofen has both presynaptic and postsynaptic effects in the mammalian CNS. Baclofen's action to increase postsynaptic potassium conductance directly depresses neuronal excitability, however, several lines of evidence indicate that this effect is not alone responsible for baclofen's marked depression of postsynaptic potentials. Although most of the recordings probably have been obtained by somatic impalements, the data suggest that the primary mechanism by which baclofen depresses synaptic transmission is a presynaptic reduction of transmitter release. This conclusion is supported directly by the established effect of baclofen to reduce the evoked release of several putative neurotransmitters.

At present, the mechanism by which baclofen reduces presynaptic transmitter release is unclear. Presynaptic inhibition secondary to depolarization of afferent terminals can be excluded, however, and the data indicate that baclofen does not reduce synaptic transmission by hyperpolarizing presynaptic fibers sufficiently to impair action potential invasion of the terminal region. There is some evidence which suggests that baclofen may reduce transmitter release by reducing inward calcium currents in presynaptic terminals. There is no direct evidence for such a mechanism, however, and at least some calcium currents do not appear to be affected by baclofen.

### References

- 1. Ashby P, White DG (1973) "Presynaptic" inhibition in spasticity and the effect of  $\beta$ (4-chlorophenyl)-GABA. J Neurol Sci 20: 329–338
- 2. Auden N, Wachtel H (1977) Biochemical effects of baclofen ( $\beta$ -parachlorophenyl-GABA) on the dopamine and the noradrenaline in the rat brain. Acta Pharmacol Toxicol (Copenh) 40:310-320
- 3. Ault B, Evans RH (1978) Central depressant action of baclofen. J Physiol (Lond) 284:131P
- 4. Ault B, Evans RH (1981) The depressant action of baclofen on the isolated spinal cord of the neonatal rat. Eur J Pharmacol 71:357-364
- 5. Ault B, Nadler JV (1982) Baclofen selectively inhibits transmission at synapses made by axons of CA3 pyramidal cells in the hippocampal slice. J Pharmacol Exp Ther 223:291-297
- Ault B, Nadler JV (1983) Anticonvulsantlike actions of baclofen in the rat hippocampal slice. Br J Pharmacol 78:701 -708
- Ault B, Nadler JV (1983) Effect of baclofen on synaptically-induced cell firing in the rat hippocampal slice. Br J Pharmacol 80:211 -219
- Barry SR (1984) Baclofen has a presynaptic action at the crayfish neuromuscular junction. Brain Res 311:152-156
- Bein HJ (1972) Pharmakologische Differenzierung von Muskelrelaxantien. In: Birkmayer W (ed) Aspekte der Muskelspastik. Huber, Wien, pp 76–82
- Ben-Ari V, Krnjevic K, Reiffenstein RJ, Reinhardt W (1981) Inhibition conductance changes and action of γ-aminobutyrate in rat hippocampus. Neuroscience 6:2445 -2463
- 11. Benecke R, Meyer-Lohmann J (1974) Effects of an antispastic drug  $\beta$ -(4-chlorophenyl)-gamma-aminobutyric acid on Renshaw cell activity. Neuropharma-cology 13: 1067–1075
- Bianchine JR (1985) Drugs for Parkinson's disease, spasticity, and acute muscle spasms. In: Goodman, Gilman (eds) The Pharmacological Basis of Therapeutics. Macmillan, London Basingstoke, pp 473 -490
- Birkmayer RW (1972) Spasticity a topical survey. Huber, Wien
- Blaxter TJ, Carlen PL (1985) Pre- and postsynaptic effects of baclofen in the rat hippocampal slice. Brain Res 341:195-199

- Bowery NG, Hill DR, Hudson AL, Doble AL, Middlemiss A, Shaw J, Turnbull M (1980) (-)Baclofen decreases neurotransmitter release in the mammalian CNS by an action at a novel GABA receptor. Nature 283:92-94
- Bowery NG, Doble A, Hill DR, Hudson AL, Shaw JS, Turnbull MJ, Warrington R (1981) Bicuculline-insensitive GABA receptors on peripheral autonomic nerve terminals. Eur J Pharmacol 71:53-70
- Bowery NG (1982) Baclofen: 10 years on. Trends in Pharmacological Science 3:400 -403
- Bowery NG, Hill DR, Hudson AL (1983) Characteristics of GABA<sub>B</sub> receptor binding sites on rat whole brain synaptic membranes. Br J Pharmacol 73:191–206
- Bowery NG, Prive GW, Turnbull MJ, Wilkin GP (1983) Evidence for the presence of GABA<sub>B</sub> receptors on cerebellar Purkinje dendrites. Br J Pharmacol 79:9P
- Brady RJ, Swann JW (1984) Postsynaptic actions of baclofen associated with its antagonism of bicuculline-induced epileptogenesis in hippocampus. Cell Mol Neurobiol 4:403–408
- Cain CR, Simmonds MA (1982) Effects of baclofen on the olfactory cortex slice preparation. Neuropharmacology 21:371-373
- 22. Capek R, Esplin B (1982) Baclofen-induced decrease of excitability of primary afferents and depression of monosynaptic transmission in cat spinal cord. Can J Physiol Pharmacol 60:160–166
- Cherubini E, North RA (1984) Inhibition of calcium spikes and transmitter release by gamma-aminobutyric acid in the guineapig myenteric plexus. Br J Pharmacol 82:101-105
- 24. Collins GGS, Anson J, Kelly EP (1982) Baclofen: Effects on evoked field potentials and amino acid neurotransmitter release in the rat olfactory cortex slice. Brain Res 238:371-383
- Connors BW, Gutnick MJ, Prince DA (1982) Electrophysiological properties of neocortical neurons in vitro. J Neurophysiol 48:1302–1320
- Curtis DR, Duggan AW, Felix D, Johnston GAR, McLennan H (1971) Antagonism between bicuculline and GABA in the cat brain. Brain Res 33:57-73
- 27. Curtis DR, Game CJA, Johnston GAR, McCulloch RM (1974) Central effects of  $\beta$ -(p-chlorophenyl)-gamma-aminobutyric acid. Brain Res 70:158–170

- Curtis DR, Lodge D, Bornstein JC, Peet MJ (1981) Selective effects of (-)-baclofen on spinal synaptic transmission in the cat. Exp Brain Res 42: 158-170
- Davidoff RA, Sears ES (1974) The effects of Lioresal on synaptic activity in the isolated spinal cord. Neurology 24:957–963
- Davies J (1981) Selective depression of synaptic excitation in cat spinal neurons by baclofen: an iontophoretic study. Br J Pharmacol 72:373-384
- 31. Davies J, Dray A (1976) Substance P in the substantia nigra. Brain Res 107:623-627
- Davies J, Watkins JC (1974) The action of β-phenyl-GABA derivatives on neurones of the cerebral cortex. Brain Res 70:501-505
- 33. Desarmenien M, Feltz P, Occhiopinti G, Santangelo F, Schlichter R (1984) Coexistence of GABA<sub>A</sub> and GABA<sub>B</sub> receptors on A delta and C primary afferents. Br J Pharmacol 81:327-333
- Dreifuss JJ, Kelly JS, Krnjevic K (1969) Cortical inhibition and gamma-aminobutyric acid. Exp Brain Res 9:137-154
- Dunlap K (1981) Two types of gammaaminobutyric acid receptor on embryonic sensory neurons. Br J Pharmacol 74:579 -585
- Enna SJ, Gallagher JP (1983) Biochemical and electrophysiological characteristics of mammalian GABA receptors. Int Rev Neurobiol 2:181-212
- 37. Evans RH, Francis AA, Watkins JC (1976) The effects of substance P like peptides on spinal motoneurones in vitro and antagonism by Lioresal. Proc Br Pharmacol Soc C18
- Evans RH, Watkins JC (1978) Specific antagonism of excitant amino acids in the isolated spinal cord of the neonatal rat. Eur J Pharmacol 50: 123-129
- 39. Fagg GE, Foster AC (1983) Amino acid neurotransmitters and their pathways in the mammalian central nervous system. Neuroscience 9:701-719
- Fillenz M, Fung SC (1983) Effect of GABA on <sup>3</sup>(H)noradrenaline release from rat hippocampal synaptosomes. J Physiol (Lond) 339:39-40P
- Fotherby KJ, Morrish NJ, Ryall RW (1976) Is lioresal (baclofen) an antagonist of substance P? Brain Res 113:210-213
- 42. Fox S, Krnjevic K, Morris ME, Puil E, Werman R (1978) Action of baclofen on mammalian synaptic transmission. Neuroscience 3:495-515
- 43. Fukuda H. Kudo Y, Ono H (1977) Effects of  $\beta$ -(p-chlorophenyl)-GABA (baclofen) on spinal synaptic activity. Eur J Pharmacol 44:17-24

- 44. Gähwiler BH, Maurer R, Wüthrich HJ (1984) Pitrazepin, a novel GABA A antagonist. Neurosci Lett 45:311–316
- 45. Gähwiler BH, Brown DA (1985) GABA<sub>B</sub> receptor-activated K<sup>+</sup> current in voltageclamped CA<sub>3</sub> pyramidal cells in hippocampal cultures. Proc Natl Acad Sci USA 82:1558–1562
- 46. Gehlert DR, Yamamura HI, Wamsley JK (1985) Gamma-aminobutyric acid-B-receptors in the rat brain: Quantitative autoradiographic localization using μ<sup>3</sup>H°(-)-baclofen. Neurosci Lett 56: 183-188
- 47. Haas HL, Greene RW, Olpe HR (1985) Stereoselectivity of L-baclofen in hippocampal slices of the rat. Neurosci Lett 55:1-4
- Haefely W (1983) Tranquillizers. In: Grahame-Smith DG, Cowen PJ (eds) Psychopharmacology I, part 1: Preclinical psychopharmacology. Excerpta Medica, Amsterdam Oxford Princeton, Elsevier, Amsterdam, pp 107–151
- Haefely W, Polc P (1983) Electrophysiological studies on the interaction of anxiolytic drugs with GABAergic mechanisms. In: Malick JB, Enna SJ, Yamamura HI (eds) Anxiolytics: Neurochemical, Behavioral and Clinical Perspecitives. Raven Press, New York, pp 113-145
- Heinemann U, Hamon B, Konnerth A (1984) GABA and baclofen reduce changes in extracellular free calcium in area CA1 of rat hippocampal slices. Neurosci Lett 47:295-300
- 51. Henry JL, Ben-Ari Y (1976) Actions of the p-chlorophenyl derivative of GABA, lioresal, on nociceptive and non-nociceptive units in the spinal cord of the cat. Brain Res 117:540-544
- 52. Henry JL (1982) Pharmacological studies on the prolonged depressant effects of baclofen on lumbar dorsal horn units in the cat. Neuropharmacology 21:1085-1093
- 53. Henry JL (1982) Effects of intravenously administered enantiomers of baclofen on functionally identified units in lumbar dorsal horn of the spinal cat. Neuropharmacology 21:1073-1083
- 54. Hill DR, Bowery NG (1981) <sup>3</sup>H-baclofen and <sup>3</sup>H-GABA bind to bicuculline-insensitive GABA<sub>B</sub> sites in rat brain. Nature 290:149–152
- 55. Howe JR, Sutor B, Zieglgänsberger W (1985) The hyperpolarization by baclofen of rat neocortical neurons is probably unrelated to its depression of postsynaptic potentials. Neurosci Lett [Suppl] 22: S386
- 56. Howe JR, Sutor B, Zieglgänsberger W

- Physiol (submitted for publication)
  57. Howe JR, Sutor B, Zieglgänsberger W (1986) Characteristics of a long-duration inhibitory postsynaptic potential in rat frontal neocortical neurons *in vitro*. Cell Mol Neurobiol (submitted for publication)
- 58. Howe JR, Zieglgänsberger W (1986) D-Baclofen does not antagonize the actions of L-baclofen on rat neocortical neurons. Neurosci Lett (submitted for publication)
- 59. Inoue M, Matsuo T, Ogata N (1985) Baclofen activates voltage-dependent and 4-aminopyridine sensitive K<sup>+</sup> conductance in guinea-pig hippocampal pyramidal cells maintained *in vitro*. Br J Pharmacol 84:833-841
- 60. Inoue M, Matsuo T, Ogata N (1985) Characterization of pre- and postsynaptic actions of (-)-baclofen in the guinea-pig hippocampus in vitro. Br J Pharmacol 84:843-851
- Johnston GAR, Hailstone MH, Freeman CG (1980) Baclofen: stereoselective inhibition of excitant amino acid release. J Pharm Pharmacol 32:230
- 62. Karbon EW, Duman R, Enna SJ (1983) Biochemical identification of multiple GABA<sub>B</sub> binding sites: association with noradrenergic terminals in rat forebrain. Brain Res 274: 393–396
- Kato M, Waldmann U, Murakami S (1978) Effects of baclofen on spinal neurones of cats. Neuropharmacology 17:827-833
- 64. Keberle H, Faigle JW (1972) Synthese sowie Beziehung zwischen Struktur und Wirkung der Gamma-aminobuttersäurederivate. In: Birkmayer W (ed) Aspekte der Muskelspastik. Huber, Wien, pp 90–93
- 65. Klee MR, Misgeld U, Zeise ML (1981) Pharmacological differences between CA3 and dentate granule cells in hippocampal slices. In: Feher O, Joo F (eds) Advances in Physiological Science, vol 36. Akademiai Kiado, Budapest, pp 155–164
- 66. Knutsson E, Lindblom U, Martensson A (1974) Plasma and cerebrospinal fluid levels of baclofen (Lioresal) at optimal therapeutic responses in spastic paresis. J Neurol Sci 23:473-484
- Koketsu K, Shoji T, Yamamoto K (1974) Effects of GABA on presynaptic nerve terminals in bullfrog sympathetic ganglia. Experientia 30:382–383
- Krogsgaard-Larsen P, Falch E, Jacobsen P (1984) GABA agonists: Structural requirements for interaction with the GABA-benzodiazepine receptor complex. In: Bowery

The Neuropharmacology of Baclofen

NG (ed) Actions and Interactions of GABA and Benzodiazepines. Raven Press, New York, pp 109–132

- Krnjevič K (1974) Chemical nature of synaptic transmission in vertebrates. Physiol Rev 54:418-540
- Lanthorn TH, Cotman CW (1981) Baclofen selectively inhibits excitatory synaptic transmission in the hippocampus. Brain Res 225:171-178
- Laskey W (1974) Effects of a GABA-derivative on excitability of Ia afferent terminals. Proc Can Fed Biol Soc 17:55
- 72. Laskey W, Esplin B, Capek R (1975) Effects of the antispastic agent,  $\beta$ -(4-chlorophenyl)gamma-aminobutyric acid (CPG), on spinal reflexes. Proc Can Fed Biol Soc 18:30
- 73. Misgeld U, Klee MR, Zeise ML (1982) Differences in burst characteristics and drug sensitivity between CA3 neurons and granule cells. In: Klee MR, Lux HD, Speckmann EJ (eds), Physiology and Pharmacology of Epileptogenic Phenomena. Raven Press, New York, pp 131–139
- Möhler H, Okada T (1977) GABA receptor binding with <sup>3</sup>(H) + bicuculline-methiodide in the rat CNS. Nature 267:65-67
- 75. Mugnaini E, Oertel W (1985) An atlas of the distribution of gabaergic neurons and terminals as revealed by GAD immunohistochemistry. In: Björklund A, Hökfelt T (eds) Handbook of Chemical Neuroanatomy. GABA and Neuropeptides in the CNS, part I. Elsevier, Amsterdam, pp 436-608
- Newberry NR, Nicoll RA (1984) Direct hyperpolarizing action of baclofen on hippocampal pyramidal cells. Nature 308:450
   –452
- 77. Newberry NR, Nicoll RA (1984) A bicuculline-resistant inhibitory post-synaptic potential in rat hippocampal pyramidal cells *in vitro*. J Physiol (Lond) 348:239–254
- Newberry NR, Nicoll RA (1985) Comparison of the action of baclofen with gammaaminobutyric acid on rat hippocampal pyramidal cells *in vitro*. J Physiol (Lond) 360:161-185
- 79. Ogata N, Abe H (1982) Neuropharmacology in the brain slice: Effects of substance P on neurons in the guinea pig hypothalamus. Comp Biochem Physiol 72:171-178
- 80. Olpe HR, Koella WP, Wolf P, Haas HL (1977) The action of baclofen on neurones of the substantia nigra and of the ventral tegmental area. Brain Res 134:577-580
- Olpe HR, Demieville H, Baltzer V, Bencze WL, Koella WP, Wolf P, Haas HL (1978) The biological activity of D- and L-baclofen (Lioresal<sup>®</sup>). Eur J Pharmacol 52:133-136
- 82. Olpe HR, Baudry M, Fagni L, Lynch G

(1982) The blocking action of baclofen on excitatory transmission in the rat hippocampal slice. J Neurosci 2:698-703

- Ono H, Fukuda H, Kudo Y (1979) Mechanisms of depressant action of baclofen on the spinal reflex in the rat. Neuropharmacology 18:647-653
- 84. Phillis JW (1976) Is β-(4-chlorophenyl)-GABA a specific antagonist of substance P on cerebral cortical neurons? Experientia 32:593-594
- Pierau FK, Zimmermann P (1973) Action of a GABA-derivative on postsynaptic potentials and membrane properties of cats' spinal motoneurons. Brain Res 54:376-380
- Pierau FK, Matheson GK, Wurster RD (1975) Presynaptic action of β-(4-chlorophenyl)-GABA. Exp Neurol 48:343–351
- Pinnock RD (1984) Hyperpolarizing action of baclofen on neurons in the rat substantia nigra slice. Brain Res 322:337-340
- Polc P, Haefely W (1976) Effects of two benzodiazepines, phenobarbitone and baclofen on synaptic transmission in the cat cuneate nucleus. Naunyn Schmiedebergs Arch Pharmacol 294:121-131
- Potashner SJ (1979) Baclofen: Effects on amino acid release and metabolism in slices of guinea pig cerebral cortex. J Neurochem 32:103-109
- Potashner SJ, Gerard D (1983) Kainate-enhanced release of D-<sup>3</sup>(H)aspartate from cerebral cortex and striatum: reversal by baclofen and pentobarbital. J Neurochem 40:1548–1557
- Price GW, Wilkin GP, Turnbull MJ, Bowery NG (1984) Are baclofen-sensitive GABA<sub>B</sub> receptors present on primary afferent terminals of the spinal cord? Nature 301:71-73
- 92. Roberts E, Chase TN, Tower DB (1976) GABA in Nervous System Function. Raven Press, New York
- 93. Saito KS, Konishi S, Otsuka M (1975) Antagonism between lioresal and substance P in rat spinal cord. Brain Res 97:177-180
- 94. Schlicker E, Classen K, Göthert M (1984) GABA<sub>B</sub> receptor-mediated inhibition of serotonin release in the rat brain. Naunyn Schmiedebergs Arch Pharmacol 326:99-105
- Schmidt RF (1971) Presynaptic inhibition in the vertebrate central nervous system. Erg Physiol Biol Exp Pharmacol 63:20-101
- 96. Scholfield CN (1983) Baclofen blocks postsynaptic inhibition but not the effect of muscimol in the olfactory cortex. Br J Pharmacol 78:79–84
- 97. Shapovalov AI, Shiriaev BI (1982) Selective modulation of chemical transmission at a

dual-action synapse (with special reference to baclofen). General Physiology and Biophysics 1:423-433

- 98. Simmonds MA (1984) Physiological and pharmacological characterization of the actions of GABA. In: Bowery NG (ed) Actions and interactions of GABA and benzodiazepines. Raven Press, New York, pp 27-41
- 99. Stevens D, Gallagher JP, Shinnick-Gallagher P (1985) Further studies on the action of baclofen on neurons of the dorsolateral septal nucleus of the rat, in vitro. Brain Res 358:360-363
- 100. Sutor B, Zieglgänsberger W (1984) A GABA-mediated, chloride-dependent depolarizing IPSP in neocortical neurons of the rat in vitro. Pflügers Arch [Suppl] 400:R37
- 101. Sutor B (1985) Nachweis eines GABA-vermittelten, inhibitorischen postsynaptischen Potentials in Neuronen des Neokortex der Ratte. Ph. D. Thesis, University of Erlangen-Nürnberg
- 102. Sutor B, Howe J, Zieglgänsberger W (1985) Baclofen depresses stimulation-evoked postsynaptic potentials of rat neocortical neurons *in vitro*. Naunyn Schmiedebergs Arch Pharmacol [Suppl] 329:381
- 103. Swahn CG, Beving H, Sedvall G (1979) Mass fragmentographic determination of 4-amino-3-p-chlorophenylbutyric acid (baclofen) in cerebrospinal fluid and serum. J Chromatogr 162:433-438
- 104. Swaynok J, Dickson C (1984) D-Baclofen is an agonist/antagonist at baclofen receptors mediating antinociception in the spinal cord. Soc Neurosci Abstr 10:32.13
- 105. Terrence CF, Sax M, Fromm GH, Chang CH, Yoo CS (1983) Effect of baclofen enantiomorphs on the spinal trigeminal nucleus and steric similarities with carbamazepine. Pharmacology 27:85–94
- 106. Wilkin GP, Hudson AL, Hill DR, Bowery NG (1981) Autoradiographic localization of GABA<sub>B</sub>-receptors in rat cerebellum. Nature 294:584–587
- 107. Wilson PR, Yaksh TL (1978) Baclofen is antinociceptive in the spinal intrathecal space of animals. Eur J Pharmacol 51:323 -330
- 108. Wojcik WJ, Neff NH (1984)  $\gamma$ -Aminobutyric acid B receptors are negatively coupled to adenylate cyclase in brain and in the cerebellum these receptors may be associated with granule cells. Mol Pharmacol 25:24–28
- 109. Young RR, Delwaide PJ (1981) Drug therapy: spasticity. N Engl J Med 304:28-33
- 110. Zieglgänsberger W (1986) Central control

of nociception. In: Handbook of Physiology. The Nervous System IV. pp 100–210 111. Žukin SR, Young AB, Snyder SH (1974)

Gamma-aminobutyric acid binding to receptor sites in the rat central nervous system. Proc Natl Acad Sci USA 71:4802-4807