



# BIOGENESIS AND FUNCTION OF PLANT LIPIDS

---

Proceedings of the Symposium on Recent Advances in the Biogenesis and Function of Plant Lipids held in Paris, June 4-7, 1980.

*Editors*

P. Mazliak  
P. Benveniste  
C. Costes  
*and*  
R. Douce



1980

ELSEVIER/NORTH-HOLLAND BIOMEDICAL PRESS  
AMSTERDAM · NEW YORK · OXFORD

6 155 909 #1

© 1980 Elsevier/North-Holland Biomedical Press

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission of the copyright owner.

ISBN for this volume: 0-444-80273-8

ISBN for the series: 0-444-80081-6

Published by:

Elsevier/North-Holland Biomedical Press  
335 Jan van Galenstraat, P.O. Box 211  
Amsterdam, The Netherlands

Sole distributors for the USA and Canada:

Elsevier North Holland Inc.  
52 Vanderbilt Avenue  
New York, N.Y. 10017

**Library of Congress Cataloging in Publication Data**

**Symposium on Recent Advances in the Biogenesis and  
Function of Plant Lipids, Paris, 1980.  
Biogenesis and function of plant lipids.**

(Developments in plant biology ; v. 6)

Bibliography: p.

Includes index.

1. Plant lipids--Congresses. I. Mazliak, P.

II. Title. III. Series.

QK898.L56S96 1980

581.19'247

80-20135

ISBN 0-444-80273-8

9570342

Printed in The Netherlands



## CONTENTS

Dedication	V
Preface	VII
LIPID METABOLISM IN CHLOROPLASTS AND LEAVES	
Oleic acid, the central substrate P.K. Stumpf, D.N. Kuhn, D.J. Murphy, M.R. Pollard, T. McKeon and J. MacCarthy	3
The role of chloroplasts in leaf lipid metabolism and poly- unsaturated fatty acid synthesis G. Roughan and R. Slack	11
Use of mesophyll protoplasts to study lipid metabolism in cell organelles R. Haas and E. Heinz	19
The structural organization of chloroplast lipids <i>in vivo</i> and in model systems: Some aspects C. Liljenberg	29
Lysogalactolipids as intermediates in galactolipid synthesis in chloroplasts K.-P. Heise and A. Sauer	39
Galactolipid formation in chloroplast envelopes. A discussion on differences between experiments <i>in vivo</i> and <i>in vitro</i> J.F.G.M. Wintermans, A. van Besouw, G. Bögemann and J. Aerts	49
Studies on the biosynthesis of sulfoquinovosyl diacylglycerol in higher plants J.B. Mudd, R. Dezacks and J. Smith	57
Long chain acyl-coenzyme A thioesters as substrates in glycerolipid biosynthesis of chloroplasts M. Bertrams and E. Heinz	67
Synthesis of long-chain acyl-CoA in chloroplast envelope membranes J. Joyard and P.K. Stumpf	73
Cooperation between chloroplasts and the extra-plastidial compartment for the biosynthesis of leaf lipids A. Trémolières, J.-P. Dubacq, D. Drapier and P. Mazliak	77
Phospholipid exchange proteins from photosynthetic tissues M. Julienne, D. Douady, J.-P. Dubacq, A. Trémolières, D. Drapier, M. Grosbois, P. Mazliak and J.-C. Kader	81
Changes in lipid composition and synthesis and in chloroplast structure observed in greening barley leaves A.O. Davies, A.T. James, R. Jeffcoat and J.L. Harwood	85

A hypothetic role for phosphatidylglycerol and 3- <i>trans</i> -hexadecenoic acid in the light reactions of the photosynthetic process J.-C. Duval, J.-P. Dubacq and A. Trémolières	91
Trans-3-hexadecenoic acid and grana stacking T. Guillot-Salomon, A. Trémolières, C. Tuquet and J.-P. Dubacq	95
Lipid changes in plastids isolated from alfalfa seedlings grown under salt-stress F. Harzallah-Skhiri, T. Guillot-Salomon and M. Signol	99
Influence of growth temperature on acyl lipids of leaves D.J. Chapman and J. Barber	103
Fatty acid synthesis in isolated chromoplasts and chromoplast homogenates. ACP stimulation, substrate utilisation, and cerulenin inhibition B. Liedvogel and H. Kleinig	107
Partial purification and properties of a soluble fatty acid synthetase from olive leaves L.M. Daza, M. Garrido and J.P. Donaire	111
Function and distribution of phospholipids in spinach thylakoid membranes as revealed by phospholipase A <sub>2</sub> treatment A. Rawyler and P.-A. Siegenthaler	117
Characterization and meaning of chloroplast lipoxygenase activities R. Douillard	121
Galactolipid biosynthesis in <i>Euglena gracilis</i> E. Blee and R. Schantz	127
Phosphatidylglycerol biosynthesis in <i>Euglena gracilis</i> A. Chammai and R. Schantz	131
Discovery of a new glyceroglycolipid in blue-green algae and its role in galactolipid biosynthesis G.B. Feige, E. Heinz, K. Wrage, N. Cochems and E. Ponzelar	135
 LIPID METABOLISM IN NON-PHOTOSYNTHETIC TISSUES	
Fatty acid synthesis J.L. Harwood	143
Phospholipid localization in biological membranes J.A.F. Op den Kamp, T. Kauerz and G. van Meer	153
Phospholipid exchange protein from higher plants M. Yamada, T. Tanaka and J.-I. Ohnishi	161
Lipid metabolism as a factor in environmental adaptation P.J.C. Kuiper	169
Biogenesis of lipids in oilseed plants L.-Å. Appelqvist	177

The biochemistry of lipids in cereal crops T. Galliard and P.J. Barnes	191
Lipid composition of sycamore cells cultivated at various temperatures M. Gawer, F. Trapy, J. Guern and P. Mazliak	199
Oxygen and temperature effects on the fatty acid composition of sycamore cells ( <i>Acer pseudoplatanus</i> L.) F. Rebeille, R. Bligny and R. Douce	203
Desaturation of fatty acids in lipids in response to the growth temperature in the blue-green alga, <i>Anabaena variabilis</i> N. Sato and N. Murata	207
Subcellular localisation of fatty acid synthetases in cell cultures of higher plants F. Spener	211
Biosynthesis of parinaric acid (9,11,13,15-octadecatetraenoic acid) M. Noda, K. Ohga, Y. Nakagawa and K. Ichihara	215
A comparison of the polypeptide and phospholipid composition of oil body and microsomal preparations from safflower and linseed cotyledons R. Slack and G. Roughan	219
Lipid metabolism in developing wheat seeds D.N. Stokes, T. Galliard and J.L. Harwood	223
$\alpha$ -ketodicarboxylic acids in lipogenic substrates in Flacourtiaceae I. Tober and F. Spener	227
Root phospholipid composition as a factor of the differential Ca-sensitivity of plants M. Rossignol and C. Grignon	231
Calcium binding, phosphatidic acid formation and fatty acid breakdown in plant mitochondria J. Dupont and C. Lance	235
The availability of palmitoyl-CoA influences the activity of palmitoyl-CoA hydrolase in carrot homogenate P. Baardseth and E. Slinde	239
Effect of sodium chloride and calcium sulfate on the lipid composition of sunflower leaves ( <i>Helianthus annuus</i> L.) L. Bettaieb, M. Gharsalli and A. Cherif	243
Relationships between temperature, microviscosity and desaturases activities in the microsomal membranes of two fungi with different behaviour L. Chavant, C. Montant and C. Wolf	249
<i>Scopulariopsis brevicaulis</i> (Bainier) : Study of the lipid content in relation to growth J.-L. Fonvieille and M. Sancholle	253
Fatty acid biosynthesis in cell free preparations of <i>Anabaena cylindrica</i> Z.T. Al'Araji and T.J. Walton	259

The effect of biotrophic fungal infection on the lipid metabolism of green plants D.M. Lösel	263
WAX METABOLISM	
Light promotes synthesis of the very long fatty acid acyl chains in maize wax P. von Wettstein-Knowles, P. Avato and J. D. Mikkelsen	271
Alkanes and alkenes in the epicuticular waxes from Cistus plants P.-G. Gülz	275
Stearoyl-CoA metabolism in the microsomes from leek epidermal cells: Thioesterase, acyltransferase and elongase activities T. Abdul-Karim, R. Lessire and C. Cassagne	281
Synthesis of lipids by epidermal and mesophyll protoplasts isolated from barley leaf sheaths J.D. Mikkelsen	285
Long chain fatty acid activation: Relations with very long chain fatty acids biosynthesis R. Lessire and C. Cassagne	291
PIGMENTS	
Prenylquinones in plant leaves H.K. Lichtenthaler	299
Current concept of organization of chlorophyll biosynthesis A.A. Shlyk	311
Carotenoid biosynthesis in plants J.W. Porter, S.L. Spurgeon and D. Pan	321
The role of carotenoids in chloroplasts of higher plants D. Siefertmann-Harms	331
Biosynthesis of $\alpha$ -tocopherol and plastoquinone-9 in spinach chloroplasts J. Soll and G. Schultz	341
Carotenoid biosynthesis in <i>Scenedesmus obliquus</i> (Chlorophyta): Experiments with deuterium oxide G. Britton and A.P. Mundy	345
The influence of kinetin on the chlorophyll biosynthesis in radish cotyledons C. Buschmann	349
Properties of membrane-bound 3-hydroxy-3-methylglutaryl-coenzyme A reductase (EC 1.1.1.34) from radish seedlings and some aspects of its regulation T.J. Bach, H.K. Lichtenthaler and J. Rétey	355

Carotenoid biosynthesis: Biogenesis of capsanthin and capsorubin in pepper fruits ( <i>Capsicum annuum</i> ) B. Camara and R. Monéger	363
Level of chlorophyll b and the light harvesting chlorophyll-protein complex in <i>Raphanus</i> seedlings grown at different light quanta fluence rates U. Prenzel, H.K. Lichtenthaler and D. Meier	369
Circular dichroism studies of the spontaneous organization of xanthophyll compounds in water-alcohol solutions. Correlations with the role played by the polyene substances in the structural organization of biological systems J. Lematre, B. Maudinas and C. Ernst	373
Light influence on zeaxanthin epoxidation inhibition of S-ethyl dipropylthiocarbamate R.E. Wilkinson	375
Lipids, pigments, light-harvesting chlorophyll protein complex and structure of a virescent mutant of maize E. Selstam	379
STEROLS AND TERPENES	
Function as an evolutionary determinant of biosynthesis W.R. Nes	387
The use of mutants and azasterols in studies of yeast sterol biosynthesis A.C. Oehlschlager, H.D. Pierce, Jr., A.M. Pierce, R.H. Angus, E. Quantin-Martenot, A.M. Unrau and R. Srinivasan	395
Biosynthesis of sterol conjugates in plants Z.A. Wojciechowski	405
A monolayer study of lipid: Protein interactions in the chloroplast membrane D.G. Bishop and J.R. Kenrick	415
Incorporation of <sup>14</sup> C-labeled CO <sub>2</sub> , phosphoglycerate, phosphoenol- pyruvate, pyruvate, acetate and mevalonate into terpenoids and acyllipids of isolated intact spinach chloroplasts K.H. Grumbach	421
Site of synthesis of geranylgeraniol derivatives in intact spinach chloroplasts M.A. Block, J. Joyard and R. Douce	427
Latency of uridine diphosphate glucose-sterol-β-D-glucosyl-transferase and uridine diphosphatase in purified membrane fractions from maize coleoptiles M. M'Voula-Tsiéri, P. Benveniste, E. Martenot and M.-A. Hartmann-Bouillon	431

Biosynthesis and cellular localisation of terpene hydrocarbons in maritime pine C. Bernard-Dagan, M. Gleizes, G. Pauly, J.P. Carde and A. Marpeau	437
Membrane systems involved in synthesis and transport of monoterpene hydrocarbons in pine leaves J.-P. Carde, C. Bernard-Dagan and M. Gleizes	441
Presence of steryl glycosides and amyirin glycosides in a blue-green alga: <i>Nostoc commune</i> and a red alga: <i>Porphyridium</i> sp. R. Duperon, P. Doireau, A. Verger and P. Duperon	445
Author index	449



## BIOSYNTHESIS OF $\alpha$ -TOCOPHEROL AND PLASTOQUINONE-9 IN SPINACH CHLOROPLASTS

JÖRGEN SOLL AND GERNOT SCHULTZ

Institut für Tierernährung, Arbeitsgruppe für Phytochemie und Futtermittelkunde, Tierärztliche Hochschule, D - 3000 Hannover 1 (FRG)

### ABSTRACT

Prenylation and methylation reaction in  $\alpha$ T biosynthesis is localized in the envelope membranes of the chloroplasts, while PQ-9 biosynthesis takes place in the envelope membranes and also in the thylakoid membranes. The sequence in  $\alpha$ -T biosynthesis in spinach is (see also Figure 1): Homogentisate + Phytyl-PP  $\longrightarrow$  Me-6-PQH<sub>2</sub>  $\longrightarrow$  2,3-Me<sub>2</sub>PQH<sub>2</sub>  $\longrightarrow$   $\gamma$ T  $\longrightarrow$   $\alpha$ T; for the PQ-9 biosynthesis it is: Homogentisate + Solanesyl-PP  $\longrightarrow$  Me-6-SQH<sub>2</sub>  $\longrightarrow$  PQH<sub>2</sub>.

### RESULTS

The two major prenylquinones of the chloroplast are  $\alpha$ T and PQ-9. The aromatic moiety of both derives from homogentisate<sup>1</sup>; the prenyl sidechain originates from a polyprenyl-PP (C<sub>20</sub> in the case of  $\alpha$ T, C<sub>45</sub> in that of PQ-9). Homogentisate is formed by an intraplastidic occurring shikimate pathway<sup>2,3</sup>.

### $\alpha$ -Tocopherol biosynthesis

The only site of  $\alpha$ T biosynthesis in spinach chloroplasts are the envelope membranes<sup>4,5</sup>. Phytyl-PP derives by reduction from GGPP which is synthesized by a recombined system of chloroplast envelope or thylakoid membranes with stroma (soluble fraction)<sup>6</sup>. Homogentisate is solely prenylated with phytyl-PP to form Me-6-PQH<sub>2</sub><sup>5</sup>. There is no stimulation by other chloroplast fractions like stroma or thylakoid membranes<sup>5</sup>. The prenyltransferase in spinach shows a strong specificity for phytyl-PP (26 pmol/h. mg protein); GGPP is inactive in this system<sup>5</sup>! From the possible position isomers only Me-6-PQH<sub>2</sub> but not Me-5- or Me-3-PQH<sub>2</sub> are formed<sup>5</sup>. A kinase which forms phytyl-PP from phytol + ATP is localized in the stroma<sup>5</sup>.

The following two methylation steps with SAM as methyl-group donor to form

---

Abbreviations: GGPP - geranylgeranyl-PP; Me-6-GGQH<sub>2</sub> - 2-methyl-6-geranylgeranylquinol; Me-6-PQH<sub>2</sub> and isomers - 2-methyl-6-phytylquinol and 5- and 3-phytyl-isomers; 2,3-Me<sub>2</sub>PQH<sub>2</sub> - 2,3-dimethyl-5-phytylquinol; Me<sub>3</sub>PQH<sub>2</sub> - trimethylphytylquinol; Me-6-SQH<sub>2</sub> - 2-methyl-6-solanesylquinol; PQ-9 -<sup>3</sup>plastoquinone-9; PQH<sub>2</sub> - plastoquinol-9; SAM - S-adenosylmethionine;  $\alpha$ T,  $\gamma$ T,  $\delta$ T -  $\alpha$ ,  $\gamma$ ,  $\delta$ -tocopherol;  $\gamma$ T<sub>3</sub> -  $\gamma$ -tocotrienol.

$\alpha$ T from Me-6-PQH<sub>2</sub> are also only due to the envelope membranes<sup>4</sup>: (that is: Me-6-PQH<sub>2</sub>  $\xrightarrow{\text{SAM}}$  2,3-Me<sub>2</sub>PQH<sub>2</sub>  $\longrightarrow$   $\gamma$ T  $\xrightarrow{\text{SAM}}$   $\alpha$ T). Again no stimulation occurs by combining envelope membranes with stroma protein. In marked contrast to the prenylation enzyme, the methyl transferase exhibits a preference for Me-6-GGQH<sub>2</sub> (2 nmol/h. mg protein) in comparison to Me-6-PQH<sub>2</sub> (0,7 nmol/h. mg protein)<sup>4,8</sup>. In spinach, however only products of the pathway with phytyl-PP as substrate are found.

The cyclization of the prenylquinol to the corresponding chromanol (in this case: 2,3-Me<sub>2</sub>PQH<sub>2</sub>  $\longrightarrow$   $\gamma$ T) which is a prerequisite for the second methylation step takes place only in intact chloroplasts<sup>7,8</sup> but not in isolated envelope membranes. This might be due to lack of cofactors in the medium used. It is also probable that the procedure to prepare envelope membranes<sup>4,9</sup> markedly effects the enzymes involved in the cyclization reaction.

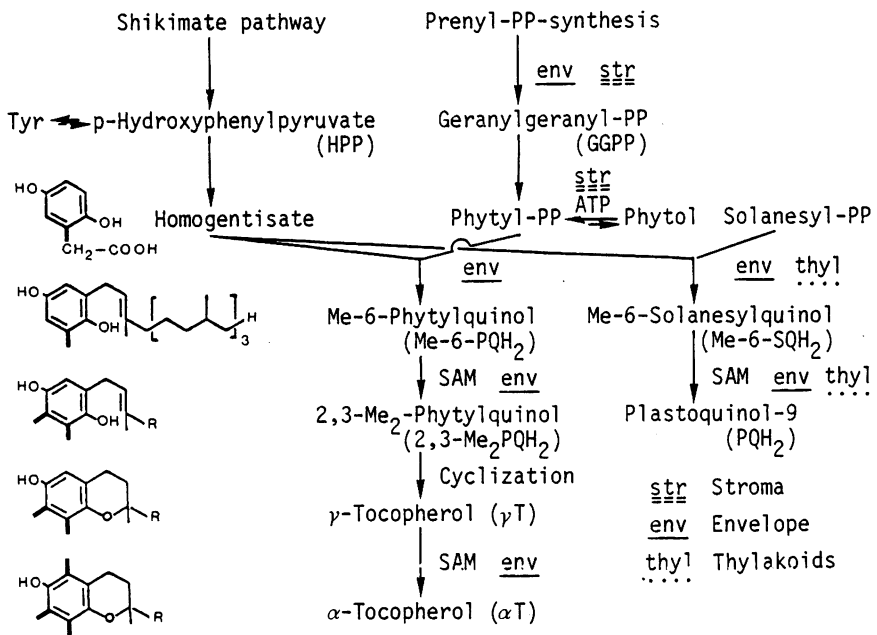


Fig. 1. Biosynthesis of  $\alpha$ T and PQ-9 in spinach chloroplasts.

### Plastoquinone-9 biosynthesis

PQ-9 biosynthesis, both prenylation and methylation is not only performed by the envelope membranes (1,2 pmol/h. mg protein and 10 pmol/h. mg protein) but also at low rates by the thylakoid membranes (0,013 pmol/h. mg protein and 0,35 pmol/h. mg protein)<sup>5</sup>. However, if one takes into account the rate of thylakoid protein to that of envelope protein per mg chlorophyll the yields in total are not as different as they are calculated on the basis of protein itself. The sequence of reactions involved in the PQ-9 biosynthesis is similar to  $\alpha$ T. Solanesyl-PP ( $C_{45}$ ) serves as prenyl compound in the prenylation reaction to form Me-6-SQH<sub>2</sub> with homogentisate. In the following steps Me-6-SQH<sub>2</sub> is methylated with SAM to yield PQ-9. In either case,  $\alpha$ T and PQ-9 biosynthesis, the quinol stage of the precursor is involved in the methylation reaction and not quinone stage<sup>7</sup>.

### CONCLUSIONS

Prenylquinones synthesized in chloroplasts operate in different ways in the photosynthetic mechanism. Whereas PQ-9 acts in the photosynthetic electron transport,  $\alpha$ T inactivates energized oxygen species formed by light (by scavenging radicals and/or quenching of  $^1O_2$ )<sup>11,12</sup>.

Both  $\alpha$ T and PQ-9, are formed by enzyme systems of lipophilic membranes of the chloroplast being in direct contact to more hydrophilic areas of the stroma. The steps of the synthesis, both prenylation and methylation of the quinols, are conceivable as electrophilic substitution of aromatics, the mechanism is not yet clear.

In fewer cases, tocopherols and tocotrienols in plants occur also in laticifers<sup>13</sup> and oil seeds but compartmentation of synthesis in comparison to chloroplast remains of future interest.

### ACKNOWLEDGEMENTS

Financial support by the Deutsche Forschungsgemeinschaft and the Centre National de la Recherche Scientifique (ERA 847 to Prof. Dr. R. Douce, Biologie Végétale, CENG and USM-G, Grenoble, France) are gratefully acknowledged. We thank Dr. D.R. Threlfall (University of Hull, U.K.) for making his manuscript available to us prior to publication.

## REFERENCES

1. Threlfall, D.R. (1971) *Vitam. Horm.*, 29, 153-200.
2. Bickel, H., Palme, L. and Schultz, G. (1978) *Phytochemistry*, 17, 119-124.
3. Löffelhardt, W. and Kindl, H. (1979) *FEBS Letters*, 104, 332-334.
4. Soll, J., Douce, R. and Schultz, G. (1980) *FEBS Letters*, 112, 243-246.
5. Soll, J., Kemmerling, M. and Schultz, G. (1980) *Arch. Biochem. Biophys.*, submitted for publication.
6. Block, M. and Douce, R. (1980) *Biochim Biophys. Acta*, in press.
7. Soll, J. and Schultz, G. (1980) *Phytochemistry*, 19, 215-218.
8. Soll, J. and Schultz, G. (1979) *Biochem. Biophys. Res. Commun.*, 91, 715-720.
9. Douce, R. and Joyard, J. (1979) *Adv. Bot. Res.*, 7, 1-116.
10. Hutson, K.G. and Threlfall, D.R. (1980) *Biochim. Biophys. Acta*, submitted for publication.
11. Foote, C.S., Clough, R.L. and Yee, B.G. (1978) *Tocopherol, Oxygen and Biomembranes* (de Duve, C., and Hayaishi, O., eds.), Elsevier/North-Holland, Amsterdam, pp. 13-21.
12. Yamauchi, R. and Matsushita, S. (1979) *Agric. Biol. Chem.*, 43, 2157-2161.
13. Peake, I.R., Audley, B.G. and Pennock, J.F. (1970) *Biochem. J.*, 119, 58p.