# Photosynthetic Light-Harvesting Systems Organization and Function

Proceedings of an International Workshop October 12–16, 1987 Freising, Fed. Rep. of Germany

Editors Hugo Scheer · Siegfried Schneider



Walter de Gruyter · Berlin · New York 1988

# CONTENTS

List of Participants	XIII
SECTION I. ORGANIZATION: BIOCHEMICAL METHODS	
Introduction: The Biochemistry of Light-Harvesting Complexes by R.J. Cogdell	1
Phycobilisome-Thylakoid Interaction: The Nature of High Molecular Weight Polypeptides by E. Gantt C.A. Lipschultz and F.X. Cunningham Jr	11
On the Structure of Photosystem II-Phycobilisome Complexes of Cyanobacteria by E. Mörschel and GH. Schatz	21
Structure of Cryptophyte Photosynthetic Membranes by W. Wehrmeyer	35
Structural and Phylogenetic Relationships of Phycoerythrins from Cyanobacteria, Red Algae and Cryptophyceae by W. Sidler and H. Zuber	49
Isolation and Characterization of the Components of the Phycobilisome from <u>Mastigocladus</u> <u>laminosus</u> and Cross- linking Experiments by R. Rümbeli and H. Zuber	61
C-Phycocyanin from Mastigocladus laminosus: Chromophore Assignment in Higher Aggregates by Cystein Modification by R. Fischer, S. Siebzehnrübl and H. Scheer	71
Photochromic Properties of C-Phycocyanin by G. Schmidt, S. Siebzehnrübl, R. Fischer and H. Scheer	77
Concerning the Relationship of Light Harvesting Bili- proteins to Phycochromes in Cyanobacteria by W. Kufer	89
Subunit Structure and Reassembly of the Light-Harvesting Complex from Rhodospirillum rubrum G9+ by R. Ghosh, Th. Rosatzin and R. Bachofen	93
Primary Structure Analyses of Bacterial Antenna Polypeptides - Correlation of Aromatic Amino Acids with Spectral Properties - Structural Similarities with Reaction Center Polymentides	
by R.A. Brunisholz and H. Zuber	103

The Structure of the "Core" of the Purple Bacterial Photo- synthetic Unit by D.J. Dawkins, L.A. Ferguson and R.J. Cogdell	115
A Comparison of the Bacteriochlorophyll CBinding Proteins of Chlorobium and Chloroflexus by P.D. Gerola, P. Højrup and J.M. Olson	129
Interactions between Bacteriochlorophyll c Molecules in Oligomers and in Chlorosomes of Green Photosynthetic Bacteria by D.C. Brune, G.H. King and R.F. Blankenship	141
Light-Harvesting Complexes of Chlorophyll c-Containing Algae by A.W.D. Larkum and R.G. Hiller	153
Isolation and Characterization of a Chlorophyll a/c-Hetero- xanthin/Diadinoxanthin Light-Harvesting Complex from Pleurochloris meiringensis (Xanthophyceae)	167
The Antenna Components of Photosystem II with Emphasis on the Major Pigment-Protein, LHC IIb by G.F. Peter and P. Thornber	175
SECTION II: ORGANIZATION: MOLECULAR GENETICS AND CRYSTALLOGRAPHY	
Molecular Biology of Antennas by G. Drews	187
High-Resolution Crystal Structure of C-Phycocyanin and Polarized Optical Spectra of Single Crystals by T. Schirmer, W. Bode and R. Huber	195
Crystallization and Spectroscopic Investigation of Purple Bacterial B800-850 and RC-B875 Complexes by W. Welte, T. Wacker and A. Becker	201
Structure of the Light-Harvesting Chlorophyll a/b-Protein Complex from Chloroplast Membranes by W. Kühlbrandt	211
Phycobilisomes of Synchechococcus Sp. PCC 7002, Pseudanabaena Sp. PCC 7409, and Cyanophora paradoxa: An Analysis by Molecular Genetics by D.A. Bryant	217
Organization and Assembly of Bacterial Antenna Complexes by G. Drews	233

The Use of Mutants to Investigate the Organization of the Photosynthetic Apparatus of <u>Rhodobacter</u> sphaeroides by C.N. Hunter and R. van Grondelle
Mechanisms of Plastid and Nuclear Gene Expression During Thylakoid Membrane Biogenesis in Higher Plants by P. Westhoff, H. Grüne, H. Schrubar, A. Oswald, M. Streubel, U. Ljungberg and R.G. Herrmann
SECTION III: ORGANIZATION: SPECIAL SPECTROSCOPY TECHNIQUES AND MODELS
Assigment of Spectral Forms in the Photosynthetic Antennas to Chemically Defined Chromophores by A. Scherz 277
Linear Dichroism and Orientation of Pigments in Phycobilisomes and their Subunits by L. Juszcak, N.E. Geacintov, B.A. Zilinskas and J. Breton 281
Low Temperature Spectroscopy of Cyanobacterial Antenna Pigments by W. Köhler, J. Friedrich, R. Fischer and H. Scheer
Chromophore Conformations in Phycocyanin and Allophycocyanin as Studied by Resonance Raman Spectroscopy by B. Szalontai, V. Csizmadia, Z. Gombos, K. Csatorday and M. Lutz
Coherent Anti-Stokes Raman Spectroscopy of Phycobilisomes, Phycocyanin and Allophycocyanin from <u>Mastigocladus</u> laminosus
by S. Schneider, F. Baumann, W. Steiner, R. Fischer, S. Siebzehnrübl and H. Scheer
Optical Absorption and Circular Dichroism of Bacteriochlorophyll Oligomers in Triton X-100 and in the Light-Harvesting-Complex B850; A Comparative Study by V. Rozenbach-Belkin, P. Braun, P. Kovatch and A.Scherz 323
Absorption Detected Magnetic Resonance in Zero Magnetic Field on Antenna Complexes from <u>Rps. acidophila</u> 7050 - The Temperature Dependence of the Carotenoid TripTet State Properties by J. Ullrich, J.U. Y. Schütz and H.C. Wolf
Effect of Lithium Dodecyl Sulfate on B 800-850 Antenna Complexes from <u>Rhodopseudomonas</u> acidophila: A Resonance Raman Study by B. Robert and H. Frank

Bacteriochlorophyll a/b in Antenna Complexes of Purple Bacteria by B. Robert, A. Vermeglio, R. Steiner, H. Scheer and M. Lutz	355
Bacteriochlorophyll c Aggregates in Carbon Tetrachloride as Models for Chlorophyll Organization in Green Photo- synthetic Bacteria by J.M. Olson and J.P. Pedersen	365
Orientation of the Pigments in the Reaction Center and the Core Antenna of Photosystem II by J. Breton, J. Duranton and K. Satoh	375
Non-Linear Absorption Spectroscopy of Antenna Chlorophyll a in Higher Plants by D. Leupold, H. Stiel and P. Hoffmann	387
SECTION IV: FUNCTION: ELECTRONIC EXCITATION AND ENERGY TRANSFER	
Excitation Energy Transfer in Photosynthesis by R. van Grondelle and V. Sundström	403
Fluorescence Spectroscopy of Allophycocyanin Complexes from Synechococcus 6301 Strain AN112 by P.Maxson, K. Sauer and A.N. Glazer	439
Picosecond Energy Transfer Kinetics in Allophycocyanin Aggregates from <u>Mastigocladus</u> laminosus by E. Bittersmann, W. Reuter, W. Wehrmeyer and A.R. Holzwarth	451
Picosecond Time-Resolved Energy Transfer Kinetics within C-Phycocyanin and Allophycocyanin Aggregates by T. Gillbro, A. Sandström, V. Sundström, R. Fischer and H. Scheer	457
Energy Transfer in "Native" and Chemically Modified C-Phyco- cyanin Trimers and the Constituent Subunits by S. Schneider, P. Geiselhart, F. Baumann, S. Siebzehnrübl, R. Fischer and H. Scheer	469
Effect of Protein Environment and Excitonic Coupling on the Excited-State Properties of the Bilinchromophores in C-Phycocyanin by S. Schneider, Ch. Scharnagl, M. Dürring, T. Schirmer and W. Bode	483
Excitation Energy Migration in C-Phycocyanin Aggregates Isolated from Phormidium luridum: Predictions from the Förster's Inductive Resonance Theory by J. Grabowski and G.S. Björn	491

Energy Transfer Calculations for two C-Phycocyanins Based on Refined X-Ray Crystal Structure Coordinates of Chromophores by K. Sauer and H. Scheer	507
Energy Transfer in Light-Harvesting Antenna of Purple Bacteria Studied by Picosecond Spectroscopy by V. Sundström, H. Bergström, T. Gillbro, R. van Grondelle, W. Westerhuis, R.A. Niederman and R.J. Cogdell	513
Excitation Energy Transfer in the Light-Harvesting Antenna of Photosynthetic Purple Bacteria: The Role of the Long-Wave- Length Absorbing Pigment B896 by R. van Grondelle, H. Bergström, V. Sundström, R.J. van Dorssen, M. Vos and C.N. Hunter	519
The Function of Chlorosomes in Energy Transfer in Green Photo- synthetic Bacteria by R.J. van Dorssen, M. Vos and J. Amesz	531
Energy Transfer in <u>Chloroflexus</u> aurantiacus: Effects of Temperature and <u>Anaerobic Conditions</u> by B.P. Wittmershaus, D.C. Brune and R.E. Blankenship	543
Interpretation of Optical Spectra of Bacteriochlorophyll Antenna Complexes by R.M. Pearlstein	555
Time Resolution and Kinetics of "F680" at Low Temperatures in Spinach Chloroplasts by R. Knox and S. Lin	567
Picosecond Studies of Fluorescence and Absorbance Changes in Photosystem II Particles from <u>Synechococcus</u> <u>Sp.</u> by A.R. Holzwarth, G.H. Schatz and H. Brock	579
Analysis of Excitation Energy Transfer in Thylakoid Membranes by the Time-Resolved Fluorescence Spectra by M. Mimuro	. 589

V. CONCLUDING REMARKS

Future Problems on Antenna Systems and Summary Remarks by E. Gantt	601
Author Index	605
Subject Index	609

# LOW TEMPERATURE SPECTROSCOPY OF CYANOBACTERIAL ANTENNA PIGMENTS

#### W. Köhler, J. Friedrich

Physikalisches Institut und Bayreuther Institut für Makromolekülforschung (BIMF), Universität Bayreuth, D-8580 Bayreuth, F.R.G.

R. Fischer, H. Scheer Botanisches Institut, Universität München D-8000 München 19, F.R.G.

## Abstract

Disorder on a microscopic scale leads to inhomogeneous line broadening of the optical spectra of chromophores, which prevents high resolution spectroscopy in a straightforward manner. In this paper emphasis is put on hole burning experiments on C-phycocyanin (PC) and phycobilisomes (PBS) of <u>Masticogladus laminosus</u>. This technique is capable of resolving the zero phonon fine structure in spite of disorder. From the measured hole profiles energy transfer times within a broad frequency range of the phycobilisome absorption could be estimated. From fluorescence line narrowing experiments combined with hole burning, details about the level structure and the loss of correlation in energy transfer processes could be elucidated. From temperature dependent hole filling experiments the distribution of conformational barriers of the chromophore attached proteins could be measured.

### Introduction

In the case of antenna pigments the knowledge of the details of the electronic-vibrational level structure is a prerequisite for understanding the

Abbreviations: PE phycoerythrin, PC phycocyanin, APC allophycocyanin, PBS phycobilisome

energy transfer mechanisms and, hence, the functioning of these pigments in the photosynthetic process (1,2). Spectroscopy at liquid Helium temperatures can usually provide this information in organic molecular systems. This results from two facts: First, the linewidths may narrow dramatically as the temperature is lowered, and second, most of the intensity may be confined to the so-called 'zero phonon transitions'. Hence, low temperatures principally allow for highly resolved spectra with very good signal to noise ratio (3).

Unfortunately, disorder on a microscopic scale prevents straightforward high resolution spectroscopy in the pigments studied, e.g. PC and PBS from <u>M. laminosus</u>, because of the concommitant inhomogeneous line broadening. Inhomogeneous line broadening not only obscures all the information contained in the homogeneous line shape function but also wipes out the vibrational pattern to a high degree. Here we wish to report how some of these drawbacks can be overcome by spectral hole burning and fluore-scence line narrowing techniques.

How disorder on a microscopic scale obscures spectroscopic information

To understand the kind of information one can obtain from a homogeneous line shape function of guest molecules in a host matrix we assume that the host-guest system is perfectly ordered and the concentration is low enough so that each guest molecule has the same microscopic environment. In this case the absorption line shape has the typical form shown in Fig. 1. It consists of a narrow so-called zero phonon line and a broad so-called phonon side band. The relative intensity in the narrow zero phonon line determines the Debye-Waller-factor, a characteristic parameter of the considered host-guest system, which is, for many systems on the order of 0.4-0.8 for temperatures around 4K. Above 40 K, or so, it becomes extremely small so that the zero phonon structure tends to vanish. The phonon side band is an outcome of the Franck-Condon-principle applied to the vibrations of the host material: The excited guest molecule has a different charge distribution. Hence, the matrix is polarized and tends to assume a new equilibrium configuration while the guest molecule is in the

294



Fig. 1. Zero phonon line with homogeneous width  $\gamma$  and phonon sideband.  $\Delta$  is half the Stokes-shift

excited state. This is equivalent to say that the matrix vibrates as a consequence of the guest excitation. Matrix vibrations are usually called phonons. If the matrix or, strictly speaking, the immediate environment of the excited chromophore is very rigid, the coupling of the electronic excitation to the matrix environment (to the lattice) is small and, hence, the phonon side band carries little oscillator strength and the Debye-Waller-factor will be close to 1. The spectrum is then dominated by the narrow and intense zero phonon lines.

The zero phonon line is a purely electronic transition, i.e. the vibrational state of the lattice does not change during this kind of excitation. Since the lifetime of an electronic level is orders of magnitude longer than that of a vibrational level, its width  $\gamma$  is, according to the uncertainty relation, much narrower, and, hence, its peak intensity may be orders of magnitude higher than the phonon side band. Hence, we have a typical absorption line shape as depicted in Fig. 1. Along these lines of reasoning it seems to be clear that one can determine the lifetime of an electronic

level in case one succeeds in measuring its homogeneous zero phonon line shape. This, however, is not an easy task because of several reasons: First, only at extremely low temperatures on the order of 1 K is the width determined by the true lifetime of the electronic level excited. With increasing temperatures the dynamics of the host matrix broaden the line without changing its lifetime. Second, many dye molecules have lifetimes on the order of 1 nsec, hence the width will be on the order of 10 to 100 MHz  $(0.0003 - 0.003 \text{ cm}^{-1})$  and the required resolution of the spectrometer has to be on the order of  $10^7$ . Third, and most important, the presence of microscopic disorder on a molecular level obscures the homogeneous line shape function. Hence, a straightforward spectroscopy is not possible (for a review, see (1)). How disorder changes the spectral properties of a chromophore in a host matrix is discussed in the following paragraph.

Fig. 2 symbolizes a disordered matrix. The solvent cage of molecule 1 is different from that of molecule 2 and 3. Hence, as schematically shown



#### Fig. 2. Schematic representation of inhomogeneous line broadening (b) as a result of microscopic disorder (a)

in Fig. 2b, these molecules absorb at different frequencies. As a consequence, spatial disorder leads to a spread of absorption frequencies, which is called inhomogeneous line broadening. At low temperatures, the inhomogeneous width may be larger than the homogeneous width by more than 4 orders of magnitudes. Hence, all details of the homogeneous molecular line shape are wiped out due to disorder.

Fluorescence line narrowing (FLN) and hole burning (HB) spectroscopy

Both FLN and HB are high resolution techniques designed to overcome inhomogeneous line broadening. The basic idea of FLN is sketched in Fig. 3a. A narrow bandwidth laser with frequency  $\omega_{\rm L}$  excites only those centers in the inhomogeneous band which are accidentally tuned with their absorption freugency to  $\omega_{\rm L}$ . Then, only molecules absorbing in a frequency range on the order of the homogeneous width around  $\omega_{\rm L}$  can emit fluorescence. If the fluorescence is detected with a high resolution spectrometer, the lines will show a structure as shown in Fig. 3a. A drawback of FLN is the fact that it is very difficult to observe resonance fluorescence and, hence, the lines are broadened by vibrational relaxation and other processes.

Hole burning is shown in Fig. 3b. It always works in case the guest molecules are photoreactive, and the zero phonon line carries enough oscillator strength. Then, those molecules excited within a frequency range



Fig. 3. Schematic representation of fluorescence line narrowing (a) and spectral hole burning (b). of the homogeneous linewidth  $\gamma$  around  $\omega_{L}$  are transformed to a product, hence, the number of molecular absorbers at  $\omega_{L}$  is diminished and a hole appears in the spectrum. Contrary to FLN, HB allows for a resonant detection at the laser frequency; therefore, this technique has the capability of measuring the homogeneous linewidth. One has, however, to be careful to rule out all slow relaxation processes such as spectral diffusion, because the time scale of HB-experiments is slow and a lot of processes may occur which broaden the hole.

Hole burning experiments on phycobilisomes and spectrally resolved energy transfer times

Fig. 4 shows an absorption spectrum of phycobilisomes of <u>M. laminosus</u> at a temperature of 4 K in a saccharose/phosphate buffer solution. The



Hole burning in PBS of M. lamino-Fig. 4. sus. Burning positions are indicated by arrows. Typical hole shapes obtained in the PC and APC-peaks are shown on an enlarged scale. PBS preparation, modified from Nies and Wehrmeyer (4), was made in 0.9 M phosphate buffer (pH 6). Centrifugation was repeated twice. Isolated PBS solution was saturated with saccharose, to ensure coupling in a low temperature glass.

spectrum is characterized by a clearly resolved structure which originates from the various pigments which build the highly organized phycobilisome assembly APC, PC, PE. In spite of the highly organized structure, there is a remarkable amount of disorder as documented by the inhomogeneous linewidths. It is a question of great interest whether the observed disorder is an intrinsic property of the chromoprotein or of the proteinaceous environment. In case of the PBS, most of the chromophores are known to interact strongly with the protein. Consequently, we believe that disorder is intrinsic and reflects the different conformational states of the latter (5). The arrows in Fig. 4 indicate the positions where hole burning was performed. Typical holes are shown in the same figure as insets. Their width is on the order of  $0.4 \text{ cm}^{-1}$ . It is interesting to compare this width with that measured for isolated phycocyanine (6). This is done in Fig. 5 for phycobilisome and PC in saccharose/buffer solution. The hole width in



Fig. 5. Comparison of spectral holes in isolated PC and phycobilisomes (PC-peak).

the phycobilisome system is much larger than in the isolated PC. We interprete this finding in the following way: Hole burning on the PC-trimer occurs effectively only on the red edge of the visible band. Hence, it seems that it is only the fluorescing chromophore(s) (or chromophore states) which are effectively burnt. In these chromophores, the fluorescence lifetime is mainly governed by intramolecular decay processes, which occur on the order of nanoseconds. At sufficiently low temperatures, where dephasing is small, these decay processes determine the width of the hole. In the phycobilisomes there is, apart from these decay processes, a fast energy transfer to acceptor chromoproteins, like APC. In case these energy transfer processes dominate the dynamics of the excited level, the transfer times can be determined from the measured hole width. From our results we would estimate transfer times on the order of 30 ps. It is interesting to note that these results do not depend very much on the position of excitation (see Fig. 4).

We conclude this section with a few remarks on the advantages and disadvantages of the hole burning method in determining lifetimes. Unlike fluorescence detection methods, which suffer from overlapping contributions from the various pigments, hole burning is a resonant method which directly yields information on the burnt state. No kinetic model is necessary in evaluating this information. However, the width of the hole is not only determined by the lifetime of the state considered, but depends also on the pure dephasing time. It is difficult to determine the latter one separately and, hence, the hole burning results have to be considered as lower bounds for the transfer times.

# Spectral distribution of the photoproduct

The nature of the hole burning reaction is, as of yet, not clear. Basically one discerns between photochemical – and photophysical reactions. In the first case it is assumed that photochemical changes of the dye molecule itself (i.e. of the chromophore) occur (e.g. bond breaking, proton transfer reactions, etc.). In the second case it is assumed that the dye molecule itself is unchanged whereas the surrounding changes (e.g. a change in the conformational state of the protein). Photochemical reactions usually lead to large spectral shifts, while photophysical reactions lead to small spectral shifts (7). In many cases (including chromoproteins) it may be difficult to distinguish both types of reactions in a clear cut way. Fig. 6 shows (together with the absorption spectrum) the difference in optical



Fig. 6. Difference in optical density (△OD) before and after burning the phycobilisome spectrum around 6370 A.

density before and after burning. Note, that most of the product occurs in a rather narrow range around the educt which favours a photophysical hole burning reaction. An interesting observation is the phototransformation which occurs in the APC-range around 6560 A. Note, that laser irradiation is performed at 6370 A in the PC-range. As obvious, the spectral range burnt in the APC-spectrum is almost as broad as the inhomogeneous band. Hence, frequency selection is lost in this case. Interestingly, only the long wavelength edge of the APC- band is burnt. It seems, that only this long wavelength range of the band is photoreactive.

## Conformational barriers

In case the reaction product is determined by the interaction between probe molecule (i.e. chromophore) and environment, we expect that, due to the disorder of the system, the reaction barriers are not well defined, but are rather characterized by a broad distribution. Hole burning offers a possibility for directly measuring this distribution of barriers (5,7). To this end we make use of temperature cycling experiments: A hole is burnt at a low temperature  $T_b$  ('burn'). Then, the temperature of the sample is raised to some value T (the 'excursion temperature') and cycled back to  $T_b$ , where the hole is measured again.

In our case burning was performed at the APC-peak, near 6550 A (see Fig. 4).  $T_b$  was 4 K. The changes of the hole area during such a cycle is a measure for the number of product molecules having returned to their educt state. The experiment is performed as a function of the excursion temperature T. For a fixed excursion temperature T, all those centers return to the educt state characterized by barriers

$$V_{T} = kT \ln R_{0} \tau$$
 (1)

 $R_0$  is the attempt frequency and  $\tau$  the experimental time needed to drive the system through one cycle. In  $R_0 \tau$  is on the order of 30. Centers with barriers higher than  $V_T$  stay unaffected. The variation of the excursion temperature enables a sampling of the distribution of barrier heights. Fig. 7a shows the result for PBS. For comparison, the results obtained for a series of organic glasses are also shown.



Fig. 7. Annealing of a spectral hole burnt into the PBS absorption (a). For the same comparison for organic behavior glasses is also shown. The distribution of reaction barriers can be described by a well superposition of glasslike states and a discrete feature distributed in a Gaussian fashion (b).

The experiments on the protein sample can be perfectly fitted by assuming that the distribution of barrier heights consists of a superposition of a glass-like distribution and a Gaussian distribution which is centered around 650 cm<sup>-1</sup> and which has a width of 210 cm<sup>-1</sup>. We interprete these findings in the following way: We assume that hole burning is due to a conformational change of the protein environment around the irradiated chromophore (photophysical HB- mechanism). On one hand, a protein is very much like a glass, i.e. it is disordered on a microscopic scale and can exist in many conformational states with different barriers. We have recently shown that in a glass the probability of finding a barrier with height V is proportional to  $1/\sqrt{V}$  (5,7). On the other hand a protein has also very specific conformational states, i.e. 'functionally important states', which are related to well defined motions, say of a large part of the peptid chain (8). It is plausible that the barrier height of such a specific conformational state depends on the actual substate of the glass-like states which the protein occupies, and, hence the barrier is spread around some mean value, which in our case is  $650 \text{ cm}^{-1}$ . The decomposition of the experimental results in a glass-like distribution and a symmetrically broadened discrete feature is shown in Fig. 7b.

Fluorescence line narrowing experiments and level structure in C-phycocyanin

The hole burning experiments document well the zero phonon nature of the optical transitions in chromoproteins (9-11) in contrast to the results obtained for reaction centers of <u>Rhodopseudomonas viridis</u> and <u>Rhodopseudomonas sphaeroides</u> (12-14). However, the fluorescence emission is broad irrespective of the mode of excitation and detection. A simple explanation of this phenomenon is the assumption of a loss of energy correlation due to transfer processes, that is, a specific level with a sharp frequency in a sensitizing state populates the whole inhomogeneous band of the fluorescing state. The idea was to narrow the fluorescence by resonant excitation of the fluorescing state. Fig. 8 shows that this idea did not work out. There is no sharply structured fluorescence even in case excitation is carried out far in the red edge of the PC-absorption. This phenomenon can be under-

stood on the basis of the special level structure involved (15) and the loss of energy correlation. As to the special level structure we have to assume that the fluorescing state consists in reality of (at least) two states with strongly overlapping inhomogeneous bands. The loss of energy corre-



Fig. 8. Selectively excited fluorescence of PC-trimer as a function of excitation wavelength.

lation means that, though the inhomogeneous bands are strongly overlapping, the energy spacing between the two states in an individual molecule can have any value between zero and (roughly) the inhomogeneous width. Since these states are coupled via energy transfer processes the maximum of the fluorescence does not shift with excitation frequency and, as the frequency is scanned into the red, the fluorescence shows a characteristic cut off at the laser frequency. Another consequence of this special level structure is a wavelength dependent hole burning efficiency: As long as the excitation frequency is far enough to the blue of the states involved, there is a high probability that the second level is lower in energy. Hence, the probability for energy transfer is high and, correspondingly, the probability for hole burning or resonance fluorescence is low. As the laser frequency is turned to the red, the probability of finding an acceptor state with lower energy decreases, hence, the efficiency for hole burning and resonance emission increases. Both phenomena can indeed be observed.

#### Acknowledgement

The authors acknowledge grants from the Deutsche Forschungsgemeinschaft, SFB 213 (Bayreuth) and SFB 143-A1 (Munich).

#### References

- 1. Scheer, H. 1981. Ang.Chemie 93, 230; Int.Ed.Engl. 20, 241
- 2. Cogdell, R.J. and H. Scheer. 1985. Photochem. Photobiol. 42, 669
- Friedrich, J. and D. Haarer. 1984. Ang.Chemie <u>96</u>, 96; Int. Ed. Engl. <u>23</u>, 113
- 4. Nies, M. and W. Wehrmeyer. 1980. Planta 150, 330
- Köhler, W., J. Friedrich and H. Scheer. 1987. Phys.Rev.A, submitted
- Füglistaller, P., H. Widmer, G. Frank and H. Zuber. 1981. Arch. Microbiol. 129, 268
- Köhler, W. and J. Friedrich. 1987. Ber. Bunsenges. Phys. Chemie <u>91</u>, 858
- 8. Fraunfelder, H. 1984. Helvet. Phys. Acta 57, 165
- Friedrich, J., H. Scheer, B Zickendraht-Wendelstadt and D. Haarer. 1981. J.Am.Chem.Soc. 103, 1030
- Friedrich, J., H. Scheer, B. Zickendraht-Wendelstadt and D. Haarer. 1980. J.Chem.Phys. <u>74</u>, 2260
- 11. Köhler, W., J. Friedrich and H. Scheer. 1987. Chem.Phys. Lett., submitted
- 12. Meech, S.R., A.J. Hoff and D.A. Wiersma. 1985. Chem.Phys. Lett. 21, 287

- 13. Boxer, S.G., D.J. Lockart and T.R. Middeldorf. 1986. Chem.Phys. Lett. <u>123</u>, 476
- 14. Gillie, J.K., B.L. Fearey, J.M. Hayes, G.J. Small and J.H. Golbeck. 1986. Chem.Phys.Lett. <u>134</u>, 316
- 15. Zickendraht-Wendelstadt, B., J. Friedrich and W. Rüdiger. 1979. Photochem.Photobiol. 31, 367