

NANOFILTRATION TREATMENT FOR PESTICIDES REMOVAL: A CASE STUDY FOR ATRAZINE AND DIMETHOATE

TAN LIAN SEE

UNIVERSITI SAINS MALAYSIA

NANOFILTRATION TREATMENT FOR PESTICIDES REMOVAL: A CASE STUDY FOR ATRAZINE AND DIMETHOATE

by

TAN LIAN SEE

Thesis submitted in fulfillment of the requirements for the degree of Master of Science

NOVEMBER 2007

ACKNOWLEDGEMENTS

First of all, I would like to express my deepest gratitude to my family for their endless love, support and blessing for me to pursue my Master's degree. My deepest appreciation also goes to both my dedicated supervisor and co-supervisor, Prof. Abdul Latif Ahmad and Dr. Syamsul Rizal Abd. Shukor for their patient guidance, support and help throughout the course of my research.

I would like to thank the membrane research group; KK Lau, Mei Fong, Choi Yee, Siew Chun, Pei Ching, Choe Peng, Hairul, Wahidah, Kak Z, Sunarti, Derek and Kak Ina for their knowledge sharing and guidance throughout my research. To Ivy, Sam, Chai, Jia Huey, Cheng Teng, Yin Fong, Kelly, Fadhil, Bee Hong, Lip Han, Wei Wen, Kak Nida, Mun Kong and other friends in USM; Thank you for your help and support as well as making my life meaningful and interesting. Many thanks to Ooi BS, Yii Seng, Khai Mun, Raymond, Wee Kang and Oscar for their advices and helps.

Sincere thanks to all the technicians and staff of School of Chemical Engineering for their cooperation, kindness and warmest helping hand. Special thank goes to En. Shamsul Hidayat and En. Faiza for their enormous help and support throughout my experimental works. I would also like to express my deepest gratitude to Kuok Foundation for providing me with scholarship for the past two years as well as to USM for funding this research with USM Short Term Grant. Last but not least, I would like to thank my special friend, Lee Chang Quan, for his encouragement and unconditional support all this while. To all the people who have helped me throughout my research, directly or indirectly; your contribution shall not be forgotten. Thank you.

Tan Lian See November 2007

ii

TABLE OF CONTENTS

		Page	
AKN	OWLEDGEMENTS	ii	
TABLE OF CONTENTS			
LIST	LIST OF TABLES		
LIST	OF FIGURES	viii	
LIST	OF PLATES	xii	
LIST	OF SYMBOL	xiii	
LIST	OF ABBREVIATION	XV	
ABS	TRAK	xvi	
ABS	TRACT	xviii	
СНА	PTER ONE : INTRODUCTION	1	
1.0	Research Overview	1	
1.1	Pesticides in Agriculture Sector	1	
1.2	Pesticides in Source of Drinking Water	3	
1.3	Problem Statement	5	
1.4	Objectives of Research	7	
1.5	Scope of Study	7	
1.6	Organization of the Thesis	9	
СНА	PTER TWO : LITERATURE REVIEW	11	
2.0	Introduction	11	
2.1	Pesticide	11	
	2.1.1 Atrazine	12	
	2.1.1(a) Environmental Impact	16	
	2.1.1(b) Toxicology Effect	16	
	2.1.2 Dimethoate	17	
	2.1.2(a) Environmental Impact	18	
	2.1.2(b) Toxicology Effect	18	
2.2	Pesticides Pollution in Water	19	
2.3	Treatment Technologies for Pesticides Removal	22	
2.4	Nanofiltration	27	
	2.4.1 Principle of Nanofiltration	27	
	2.4.2 Applications of Nanofiltration in Pesticides Removal	29	

	2.4.3 Transport Models for Nanofiltration	30
	2.4.3(a) Transport Equations	32
	2.4.3(b) Model Development	33
2.5	Statistical Tools	40
2.6	Summary	41
CHAP	PTER THREE : MATERIALS AND METHODS	42
3.0	Introduction	42
3.1	Materials	42
	3.1.1 Membrane	42
	3.1.2 Pesticides	
	3.1.2(a) Atrazine	43
	3.1.2(b) Dimethoate	43
	3.1.3 Chemicals	43
	3.1.3(a) Organic Solvent	45
	3.1.3(b) pH Adjustment	45
3.2	Experimental Set-Up	45
	3.2.1 Dead End Filtration Rig	45
3.3	Analysis Method	48
	3.3.1 High Performance Liquid Chromatography (HPLC)	48
	3.3.2 Water Composition Analysis	50
	3.3.2(a) Chemical Oxygen Demand (COD) Measurement	50
	3.3.2(b) DR 2800 Spectrophotometer	51
	3.3.3 pH Measurement	51
3.4	Experimental Procedure	52
	3.4.1 Flowchart of the Overall Experimental Works	52
	3.4.2 Filtration Experiments	53
	3.4.3 Filtration Studies at Various Conditions	54
	3.4.3(a) Comparison between NF90, NF200, NF270 and DK	55
	3.4.3(b) Effect of Water Quality	55
	3.4.3(c) Effect of Operating Pressure	56
	3.4.3(d) Effect of Pesticides Concentration	56
	3.4.3(e) Effect of Stirring Rate	56
	3.4.3 (f) Effect of Solution's pH	56
	3.4.3(g) Filtration of Binary Solutes	56
	3.4.4 Percentage of Rejection and Permeate Flux Calculation	57

3.5	Statistical Analysis	57
	3.5.1 Experimental Design	58
3.6	Spiegler-Kedem Model	58
CHA	PTER FOUR : RESULTS AND DISCUSSION	60
4.0	Introduction	60
4.1	Comparison between Membranes	60
	4.1.1 Pure Water Permeability	61
	4.1.2 Pesticide Solution Permeability	63
	4.1.3 Effect of Water Quality	68
	4.1.3(a) Water Composition	69
	4.1.3(b) Effect of Water Quality on Pesticide Rejection	70
	4.1.3(c) Effect of Water Quality on Permeate Flux	72
4.2	Parameter Study for Nanofiltration	74
	4.2.1 Effect of Operating Pressure	74
	4.2.1(a) Effect of Operating Pressure on Pesticide	75
	Rejection	
	4.2.1(b) Effect of Operating Pressure on Permeate Flux	76
	4.2.2 Effect of Feed Pesticide Concentration	78
	4.2.2(a) Effect of Feed Pesticide Concentration on	78
	Pesticide Rejection	
	4.2.2(b) Effect of Feed Pesticide Concentration on	80
	Permeate Flux	
	4.2.3 Effect of Stirring Rate	82
	4.2.3(a) Effect of Stirring Rate on Pesticide Rejection	82
	4.2.3(b) Effect of Stirring Rate on Permeate Flux	84
	4.2.4 Effect of Initial pH of solution	85
	4.2.4(a) Effect of Initial pH of solution on Pesticide	86
	Rejection	
	4.2.4(b) Effect of Initial pH of solution on Permeate Flux	88
	4.2.5 Effect of Binary Solute Mixture	90
	4.2.5(a) Effect of Binary Solute Mixture on Pesticide	90
	Rejection	
	4.2.5(b) Effect of Binary Solute Mixture on Permeate Flux	92
4.3	Statistical Analysis	92
	4.3.1 General Factorial Design	93

	4.3.2 ANOVA Analysis	93
	4.4.3 Confirmation Test	108
4.4	Modeling and Simulation	108
	4.4.1 Parameter Estimation	110
	4.4.2 Comparison between Experimental and Modeling Data	110
	4.4.3 Concentration Polarization Profile	113
CHA	PTER FIVE : CONCLUSIONS AND RECOMMENDATIONS	114
5.1	Conclusions	114
5.2	Recommendations	116
REFERENCES		118
APP	ENDICES	135
Appe	ndix A Chromatogram for Pesticides	135
Appe	ndix B Calibration Curve for COD Measurement	136
LIST	OF PUBLICATIONS, SEMINARS AND AWARDS	137

LIST OF TABLES

Table 2.1	Chemical classification system of pesticide	13
Table 2.2	Summary of cases of pesticides contamination in water	20
Table 2.3	Treatment methods available for pesticides removal and findings by researchers	24
Table 3.1	Specification of membrane used	43
Table 3.2	Purity and manufacturer of pesticides used	43
Table 3.3	Physical properties of atrazine (Kamrin, 1997)	44
Table 3.4	Physical properties of dimethoate (Kamrin, 1997)	45
Table 3.5	Structural specifications of SterlitechTM HP4750 (Sterlitech, 2005)	48
Table 3.6	Specification for DR 2800 Spectrophotometer	53
Table 3.7	Experimental design for atrazine and dimethoate	60
Table 4.1	Content summary for chapter results and discussions	62
Table 4.2	Composition of feed water	70
Table 4.3	Experimental results based on the general factorial experimental design	95
Table 4.4	ANOVA for the significance of model and model terms for atrazine	96
Table 4.5	ANOVA for the significance of model and model terms for dimethoate	96
Table 4.6	Statistical parameters obtained from the ANOVA for the reduced models	97
Table 4.7	Empirical models in terms of coded values	98
Table 4.8	Confirmation experiments for atrazine	110
Table 4.9	Confirmation experiments for dimethoate	110
Table 4.10	Parameters estimated using Lavenberg Marquardt Method on the experimental results	111

LIST OF FIGURES

		Page
Figure 1.1	Interactions between pesticides and ecosystems (Beitz et al., 1994)	5
Figure 1.2	Pesticide fate and transport (Schnoor, 1992)	6
Figure 2.1	Structure of atrazine	16
Figure 2.2	Structure of dimethoate	17
Figure 2.3	Schematic of dead-end nanofiltration process	28
Figure 2.4	Convective and diffusive flows perpendicular to the membrane surface (Field, 1996)	28
Figure 2.5	Physical interpretation of the Spiegler-Kedem model for solute transport (Williams, 2005)	32
Figure 3.1	Diagram of experimental set-up	46
Figure 3.2	Diagram of the Sterlitech HP4750 Stirred Cell (Sterlitech, 2005)	46
Figure 3.3	Calibration curve for atrazine	49
Figure 3.4	Calibration curve for dimethoate	50
Figure 3.5	Flowchart of the overall experimental works	53
Figure 4.1	Pure water permeability	63
Figure 4.2	Comparison between membranes on atrazine rejection with time at operating pressure of 6 x 10 ⁵ Pa and stirring rate of 1000 rpm. Feed pesticide concentration was 10 mg/L	63
Figure 4.3	Comparison between membranes on dimethoate rejection with time at operating pressure of 6 x 10 ⁵ Pa and stirring rate of 1000 rpm. Feed pesticide concentration was 10 mg/L	64
Figure 4.4	Comparison between membranes on atrazine and dimethoate	65
Figure 4.5	Comparison between membranes on permeate flux during atrazine rejection with time at operating pressure of 6 x 10 ⁵ Pa and stirring rate of 1000 rpm. Feed pesticide concentration was 10 mg/L	67

Figure 4.6	Comparison between membranes on permeate flux during dimethoate rejection with time at operating pressure of 6×10^5 Pa and stirring rate of 1000 rpm. Feed concentration was 10 mg/L	67
Figure 4.7	Effect of water quality on rejection of atrazine at operating pressure of 6x10 ⁵ Pa and stirring rate of 1000 rpm. Feed concentration was 10mg/L	70
Figure 4.8	Effect of water quality on rejection of dimethoate at operating pressure of 6×10^5 Pa and stirring rate of 1000 rpm. Feed concentration was 10 mg/L	71
Figure 4.9	Effect of water quality on flux performance during rejection of atrazine at operating pressure of 6 x 10 ⁵ Pa and stirring rate of 1000 rpm. Feed concentration was 10 mg/L	73
Figure 4.10	Effect of water quality on flux performance during rejection of dimethoate at operating pressure of 6 x 10 ⁵ Pa and stirring rate of 1000 rpm. Feed concentration was 10 mg/L	73
Figure 4.11	Effect of operating pressure on rejection of atrazine at feed concentration of 10 mg/L and stirring rate 1000 rpm	75
Figure 4.12	Effect of operating pressure on rejection of dimethoate at feed concentration of 10 mg/L and stirring rate 1000 rpm	76
Figure 4.13	Effect of operating pressure on permeate flux during rejection of atrazine at feed concentration of 10 mg/L and stirring rate 1000 rpm	77
Figure 4.14	Effect of operating pressure on permeate flux during rejection of dimethoate at feed concentration of 10 mg/L and stirring rate 1000 rpm	77
Figure 4.15	Effect of pesticide concentration on rejection of atrazine at operating pressure of 6 x 10 ⁵ Pa and stirring rate 1000 rpm	79
Figure 4.16	Effect of pesticide concentration on rejection of dimethoate at operating pressure of 6 x 10^5 Pa and stirring rate 1000 rpm	79
Figure 4.17	Effect of pesticide concentration on permeate flux during rejection of atrazine at operating pressure of 6 x 10^5 Pa and stirring rate 1000 rpm	81

Figure 4.18	Effect of pesticide concentration on permeate flux during rejection of dimethoate at operating pressure of 6 x 10 ⁵ Pa and stirring rate 1000 rpm	81
Figure 4.19	Effect of stirring rate on rejection of atrazine at operating pressure of 6 x10 ⁵ Pa and feed concentration of 10 mg/L	83
Figure 4.20	Effect of stirring rate on rejection of dimethoate at operating pressure of 6 x10 ⁵ Pa and feed concentration of 10 mg/L	83
Figure 4.21	Effect of stirring rate on permeate flux during rejection of atrazine at operating pressure of 6 x 10 ⁵ Pa and feed concentration of 10 mg/L	84
Figure 4.22	Effect of stirring rate on permeate flux during rejection of dimethoate at operating pressure of 6 x 10 ⁵ Pa and feed concentration of 10 mg/L	85
Figure 4.23	Effect of initial pH of solution on rejection of atrazine at operating pressure of 6 x 10 ⁵ Pa, feed concentration of 10 mg/L and 1000 rpm	86
Figure 4.24	Effect of initial pH of solution on rejection of dimethoate at operating pressure of 6 x 10 ⁵ Pa, feed concentration of 10 mg/L and stirring rate 1000 rpm	87
Figure 4.25	Effect of initial pH of solution on permeate flux during rejection of atrazine at operating pressure of 6 x 10^5 Pa, feed concentration of 10 mg/L and stirring rate 1000 rpm	89
Figure 4.26	Effect of initial pH of solution on permeate flux during rejection of dimethoate at operating pressure of 6 x 10 ⁵ Pa, feed concentration of 10 mg/L and stirring rate 1000 rpm	89
Figure 4.27	Effect of binary solute mixture on rejection of atrazine and dimethoate on (a) NF90, (b) NF200, (c) NF270 and (d) DK at operating pressure of 6 x 10^5 Pa, total pesticides concentration of 10 mg/L and stirring rate 1000 rpm	91
Figure 4.28	Effect of binary solute mixture on permeate flux during rejection of mixture of atrazine and dimethoate at operating pressure of 6×10^5 Pa, total pesticides concentration of 10 mg/L and stirring rate 1000 rpm	92
Figure 4.29	DESIGN-EXPERT plots for atrazine and dimethoate. Predicted versus actual values plot for (a) % rejection; (b) flux for atrazine and (c) %rejection; (d) flux for dimethoate	99

Figure 4.30	DESIGN-EXPERT plots. Normal probability of residual for (a) % rejection; (b) flux for atrazine and (c)% rejection; (b) flux for dimethoate	100
Figure 4.31	DESIGN-EXPERT plots. Studentized residuals versus predicted values plot for (a) % rejection; (b) flux for atrazine and (c) %rejection; (d) flux for dimethoate	101
Figure 4.32	DESIGN-EXPERT plots. Interaction graph between (a) membranes and pressure; (b) membranes and pesticide concentration	102
Figure 4.33	DESIGN-EXPERT plots. Interaction graph between pressure and stirring rate for (a) NF90; (b) NF200; (c) NF270; (d) DK for rejection of atrazine	103
Figure 4.34	DESIGN-EXPERT plots. Interaction graph between membrane and pressure for flux during rejection of atrazine	104
Figure 4.35	DESIGN-EXPERT plots. Interaction graph between membrane and pressure for rejection of dimethoate	105
Figure 4.36	DESIGN-EXPERT plots. Interaction between (a) membranes and pressure; (b) membranes and stirring rate for flux during rejection of dimethoate	106
Figure 4.37	DESIGN-EXPERT plots. Interaction between pressure and stirring rate for (a) NF90; (b) NF200; (c) NF270; (d) DK for flux during rejection of dimethoate	107
Figure 4.38	Solute rejection against permeate flux curve from experimental data and the predicted results from Spiegler-Kedem model	111
Figure 4.39	Solute rejection plotted against pressure using the experimental data and predicted results from Spiegler-Kedem model	112
Figure 4.40	Permeate flux plotted against pressure using the experimental data and predicted results from Spiegler-Kedem model	112
Figure 4.41	Concentration polarization profile plotted against pressure	113

LIST OF PLATES

Page

Plate 3.1 Dead end filtration rig

LIST OF SYMBOLS

		Unit
a	Osmotic constant	(m³Pa/g)
A	Membrane area	(m²)
C_{bs}	Solute bulk concentration	(mg/L)
C _f	Pesticide concentration of feed	(mg/L)
C_p	Pesticide concentration of permeate	(mg/L)
C_s	Solute concentration in solution	(mol/m ³)
C_{ms}	Solute concentration at the wall of feed side	(mg/L)
\overline{C}_{s}	Average solute concentration in solution	(mol/m ³)
dC_s	Solute concentration different in solution	(mol/m ³)
dP	Hydrostatic pressure driven different	(N/m²)
dx	Axial direction difference	(m)
D_{sw}	Diffusion coefficient of solute in water	(m²/s)
J_{s}	Solute flux	(mol/m ² .s)
J_{v}	Solvent fluxes	(m/s)
k _s	Mass transfer coefficient	(m/s)
K _{o/w}	Octanol/water partition coefficient	(dimensionless)
L_p	Membrane permeability	(m³/(m².s.Pa)
т	Solute molar mass	(g/mol)
P_s	Solute permeability	(mol/N.s)
$P_{\overline{s}}$	Local solute permeability	(m²/s)
P_{w}	Hydrodinamic permeability	(m³/N.s)
$\overline{P_{_{W}}}$	Specific hydraulic permeability	(m ⁴ /N.s)

r_{sc}	Radius of stirred cell	(m)
R	Percentage of pesticide rejection	(%)
R ²	Coefficient of determination	(dimensionless)
R_{os}	Observed rejection	(dimensionless)
R_s	True rejection	(dimensionless)
R_{g}	Ideal gas constant	(m ³ Pa/mol.K)
Re	Reynold number	(dimensionless)
Sc	Schmidt number	(dimensionless)
Sh	Sherwood number	(dimensionless)
t	Time	(S)
Т	Operating temperature	(Kelvin)
v_w	Permeate flux	(m ³ /m ² .s)
V	Cumulative volume	(m ³)
x	Vertical distance from the membrane surface	(m)

Greek letters		
σ_{s}	Reflection coefficient	(dimensionless)
$d\pi$	Osmotic pressure different	(N/m ²)
ΔP	Hydrostatic pressure driven difference	(N/m ²)
δ	Concentration polarization layer thickness	(m)
ρ	Density of solution	(kg/m ³)
ω	Stirring rate	(s ⁻¹)
$\Delta\pi$	Osmotic pressure difference	(N/m ²)
Δt	Time difference	(s)
μ	Viscosity of solution	(kg/m.s)
ΔV	Cumulative volume difference	(m ³)

LIST OF ABBREVIATION

AOP's	Advanced oxidation processes
AChE	Enzyme acetyl cholinesterase
ANOVA	Analysis of variance
CFR	Case fatality rate
CFSK	Combined Film Theory-Spiegler-Kedem
COD	Chemical Oxygen Demand
CV	Coefficient of variance
HPLC	High Performance Liquid Chromatography
IARC	International Agency for Research on Cancer
MCPA	Malaysian CropLife and Public Health Association
MF	Microfiltration
NF	Nanofiltration
OZ	Organo-zeolite
PAC	Powdered activated carbon
rpm	Rotation per minute
RC	Regenerated Cellulose
RO	Reverse osmosis
SK	Spiegler-Kedem
UF	Ultrafiltration
UK	United Kingdom
USA	United States of America
UV/VIS	Ultraviolet-visible
WHO	World Health Organization

RAWATAN PENURASAN NANO UNTUK PENYINGKIRAN RACUN SERANGGA: KAJIAN KES BAGI ATRAZIN DAN DIMETOAT

ABSTRAK

Tesis ini memfokuskan kepada penyingkiran racun serangga dari larutan akues menggunakan membran penurasan nano. Dua jenis racun serangga, atrazin dan dimetoat, telah dipilih untuk diuji. Empat jenis membran penurasan nano (NF90, NF200, NF270 and DK) telah diuji dengan menjalankan penurasan hujung mati teraduk menggunakan larutan racun serangga. Melalui kajian ini, ia didapati bahawa NF90 menunjukkan prestasi penolakan yang terbaik, diikuti dengan NF200 dan DK. Sementara itu, walaupun NF270 menunjukkan prestasi hasil telapan yang tertinggi di antara empat membran yang diuji, ia menunjukkan prestasi penolakan yang atrazin dengan lebih baik daripada dimethoate secara konsisten.

Prestasi penolakan dapat ditingkatkan sebanyak lebih kurang 10% apabila penurasan dijalankan pada racun serangga yang dilarutkan dalam air paip atau air sungai. Pemantauan ini menunjukkan bahawa dalam kes sebenar di loji rawatan air, penolakan racun serangga yang lebih baik dapat dijangkakan. Akan tetapi, ini diperoleh dengan pengorbanan prestasi hasil telapan kerana hasil telapan yang lebih rendah dapat diperhatikan bagi penurasan air paip dan air sungai.

Prestasi penolakan dan hasil telapan didapati menunjukkan kesan positif apabila tekanan operasi dan kadar adukan ditingkatkan. Akan tetapi, ia dikenalpasti bahawa peningkatan kepekatan suapan racun serangga telah mengurangkan prestasi penolakan dan hasil telapan. Sementara itu, meningkatkan pH larutan telah menaikkan prestasi penolakan bagi NF200, NF270 dan DK. Walau bagaimanapun, prestasi hasil telapan menurun. Kes yang berlainan dapat dipantau bagi NF90 yang mana ia

xvi

menunjukkan prestasi yang konsisten tanpa mengira keadaan pH larutan. Pengaruh bahan larut perduaan atrazin-dimetoat terhadap proses penolakan juga dikaji dan ia dikenalpasti mengurangkan sedikit prestasi penolakan membran penurasan nano. Sungguhpun begitu, prestasi telapan tidak terpengaruh oleh sistem perduaan tersebut.

Keputusan ANOVA (analisis varians) daripada rekaan faktorial am menunjukkan bahawa secara amnya, kepekatan suapan racun serangga tidak memainkan peranan yang penting dalam prestasi penolakan dan hasil telapan racun serangga. Ia hanya didapati menunjukkan kesan yang ketara bagi prestasi penolakan atrazin. Faktor lain seperti jenis membran, tekanan operasi dan kadar adukan didapati memainkan peranan yang penting dalam prestasi penolakan dan hasil telapan racun serangga.

Pengesahan data eksperimen untuk membran dengan prestasi yang terbaik, NF90, menggunakan model Spiegler-Kedem telah menunjukkan penganggaran data eksperimen yang baik. Penentuan pekali (R²) yang diperolehi untuk pemadanan data adalah 0.9871 bagi atrazin dan 0.9692 bagi dimetoat.

NANOFILTRATION TREATMENT FOR PESTICIDES REMOVAL: A CASE STUDY FOR ATRAZINE AND DIMETHOATE

ABSTRACT

This thesis focuses on the removal of pesticide from aqueous solution using nanofiltration membrane. Two pesticides, atrazine and dimethoate, were selected for study in this research. Four nanofiltration membranes (NF90, NF200, NF270 and DK) were subjected to a stirred dead-end filtration of the pesticide solution. It was found that NF90 showed the best rejection performance, followed by NF200 and DK. Meanwhile, although NF270 showed the highest permeate flux out of the four membranes tested, it showed the poorest rejection. In overall, for the four membranes tested, atrazine was consistently better rejected than dimethoate.

The rejection performance was further enhanced by approximately 10% when filtration was done with the pesticide being dissolved in tap water or river water. This observation showed that in actual case of filtration in water treatment plant, better pesticide rejection performance could be expected. However, this was obtained at the expense of flux performance since lower permeate flux was observed for filtration of tap water and river water.

Increasing operating pressure and stirring rate posed positive effects on both rejection and flux performance of nanofiltration membranes. However, increasing the feed pesticide concentration reduced the rejection and flux performance. Increasing initial pH of solution increased the rejection performance of NF200, NF270 and DK. Nevertheless, this was accompanied with reduced permeate flux. Exceptional case was observed for NF90 whereby it showed somewhat consistent performance regardless of the initial pH of solution. The effect of atrazine-dimethoate binary solutes on the rejection was also studied and it was found that the presence of binary solutes

slightly reduced the rejection performance of the nanofiltration membranes. Nevertheless, the flux performance was observed not to be affected by the binary system.

ANOVA (analysis of variance) results from general factorial design showed that generally, feed pesticides concentration did not play significant roles in rejection and flux performance of pesticides in the system. It was found to be significant only in the case of rejection performance of atrazine. Other factors such as type of membrane, operating pressure and stirring rate were found to play significant role in the rejection and flux performance of pesticides.

Verification of experimental data for the best-performed membrane, NF90, using Spiegler-Kedem model showed that the model provided a good estimation of experimental data. The coefficient of determination (\mathbb{R}^2) obtained for the fitted data was 0.9871 and 0.9692 for atrazine and dimethoate, respectively.

CHAPTER 1

INTRODUCTION

1.0 Research Overview

This chapter provides an overview of the research background. The scope covers from current scenario of pesticides usage in agriculture and how the pesticides found their way into the sources of drinking water. Then, the problem statement of this research is presented, followed by the objectives of this research as well as the scope of study and the organization of this thesis.

1.1 Pesticides in Agriculture Sector

Malaysia is actively involved in agriculture practice, planting oil palm, paddy, fruit, vegetables and others for both local consumption and export purposes. In order to achieve the objectives such as to maintain the quantity and quality of agriculture productions, pesticides are used in agriculture sector as a mean of pest control for sustainability of the industry.

In Malaysia, the annual pesticides sales figure exceeds RM 300 million. It is estimated that annual crop losses in our country could exceed 30% without pesticides (MCPA, 2005). On the other hand, insects, weeds, fungi, viruses, parasites, birds and rodents consume or destroy approximately 48% of the world's annual food production (Yedla and Dikshit, 2005).

Crop losses could be translated into less food supply and subsequently resulted in surging of food price. It is inevitable that more pesticides will be utilized to cope with the demands by the increasing population as well as to fulfill the vision of our Prime

Minister, Datuk Seri Abdullah Ahmad Badawi, to transform agriculture sector into the main industry in Malaysia.

Malaysian CropLife and Public Health Association (MCPA) (2005) defined pesticide as chemical or a biological product developed and used for pests control. In agriculture, "pests" include insects, diseases, weeds and other organisms like nematodes which can have a devastating effect on the quality and quantity of crops harvested for food, grain and fibre.

Pesticides are also used in the public health sector to control vector borne diseases like malaria, dengue fever (both diseases spread by mosquitoes), river blindness disease (spread by snails), houseflies, cockroaches and termites that destroy building structures (MCPA, 2005). Ballantyne and Marrs (2004) stated that the word 'pesticides' is used to cover substances that control organisms (insects, fungi, plants, slugs, snails, weeds, micro-organism, nematodes, etc) which destroy plant life and interfere with food chain, and which act as vectors to disease organism to man and animals.

There are a variety of pesticides to be chosen from, be it fungicides, insecticides, herbicides, rodenticides, molluscides and nematicides. In Malaysia, there are over a thousand of pesticide products registered to Department of Agriculture (Department of Agriculture, 2005). It is estimated that about 70% of pesticides sold goes to the agriculture sector (MCPA, 2005).

1.2 Pesticides in Source of Drinking Water

In Malaysia, little attention has been given to the presence of pesticides in the source of drinking water and its adverse effects on human health. These huge amount of pesticides used are the emerging contaminants in drinking water supplies.

Overseas, public attention on the potential long-term consequences of pesticides on human health and environment has started since 1962 when Carson (1962) highlighted the matter in her book 'Silent Spring'. In year 2003, test done by a well-known non-government organization on Coca-Cola and Pepsi in India revealed the presence of unacceptably high levels of pesticide residue. In one sample of Coca-Cola, the presence of lindane, a carcinogen, was 140 times higher than the allowable limit (Kapoor, 2006). As for our country, the presence of pesticides was acknowledged by Tan *et al.* (1995) to be present in almost all the river systems in Peninsular Malaysia. In June 2005, it was reported in The Star newspaper (Jessy, 2005) that uncontrolled agricultural activities at Cameron Highlands had contaminated the rivers so acute that even drinking water from the treatment plants was polluted.

The effect of pesticides on the environment is very complex as undesirable transfers occur continually among different environmental sections. Pesticides that are sprayed in the air may eventually end up in soils or water. The atmosphere is an effective medium which can move airborne pesticides away from their application sites and redeposit them in far away locations (Majewski, 1991). On the other hand, pesticides applied directly to the soil may be washed off by rain into nearby bodies of surface water or percolate through the soil to lower soil layers and groundwater (Kamrin, 1997). Furthermore, it has been reported that pesticides metabolites have high potential of leaching in soil (Fava *et al.*, 2005). However, it was noted that the

movement of pesticide in and through the soil is primary a function of water solubility of the pesticides and of the adsorption capacities of the soil type (Lichtenstein, 1972).

Pesticides uses and transfers have already extended to urbanized catchments (Blanchoud *et al.*, 2004). As a matter of fact, contamination of drinking water by pesticides can occur through carelessness and nonagriculture uses as well such as accidental major spill, application in lawns and golf courses and back-siphoning (Gustafson, 1993). The following are potential sources of pesticides that can lead to drinking water contamination listed by Gustafson (1993):

- i) Application near surface water bodies
- ii) Spray drift during major application
- iii) Small intentional spills
- iv) Lawns and golf courses
- v) Agriculture drainage wells
- vi) Point sources at storage facilities
- vii) Poorly constructed wells
- viii) Glasshouses and nurseries
- ix) Abandoned wells
- x) Back-siphoning
- xi) Application near sinkholes

Beitz *et al.* (1994) has mapped out the multiple interaction between pesticides and ecosystem as shown in Figure 1.1. Meanwhile, Schnoor (1992) summarized the fate and transport of pesticides in the environment in Figure 1.2. The figures clearly show that pesticides would eventually end up becoming a possible threat to human's health via atmosphere and water. Low-level residues of pesticides in water generally may not present acute toxicity problems, but chronic effects will likely be of concern (Carsel and Smith, 1987). This is because pesticides could have chronic effects such as cancer (Davies and Doon, 1987; Doull, 1989; Younes and Galal-Gorchev, 2000), Parkinson's disease (Cone, 2005), reproductive effects, fetal damage, delayed neurologic manifestations and possible immunologic disorders (Doull, 1989).



Figure 1.1: Interactions between pesticides and ecosystems (Beitz et al., 1994).

1.3 Problem Statement

In 1987, World Health Organization estimated that up to 500,000 illnesses and as many as 20,000 deaths can be attributed annually to pesticidal chemicals worldwide (Wilkinson, 1987). Presently, on a worldwide basis, intoxications attributed to pesticides have been estimated to be as high as 3 million cases of acute and severe poisoning annually, with many unreported cases and with some 220 000 deaths (Kumazawa and Suzuki, 2000). This situation creates urgency to produce solution to remove pesticides from the source of drinking water. This is because pesticides will continue to be effective pest controls, but it is up to us to find ways to avoid many of the pesticides poisonings and contaminations that exist today.



Figure 1.2: Pesticide fate and transport (Schnoor, 1992).

Traditionally, removal of pesticides for the production of drinking water is done by adsorption using activated carbon which is effective but requires frequent regeneration (Van der Bruggen *et al.*, 1998). Besides that, chlorination, air stripping, coagulation, ozonation and advanced oxidation are also being used for treatment of pesticides from water (WHO, 2005). These methods however suffer from their own limitations and have limited successful applications in this area, leaving the vacuum to be filled by membrane technology. Although there could be fouling in membranes over the time, membrane processes are becoming increasingly widespread in water treatment applications due to its ability to achieve high removals of constituents such as dissolved solids, organic carbons, inorganic ions and organic compounds.

1.4 Objectives of Research

The objective of this research is to examine the performance of nanofiltration membranes in rejection of pesticides in aqueous solution. In order to achieve the main objective, the measurable objectives of this study are stated as follows:

- To evaluate the separation performance for different pesticides, different water quality and different commercial nanofiltration membranes in lab scale operation.
- To study the effect of experimental parameters namely operating pressure, pesticide concentration, stirring rate, pH of solution and binary solute mixture on rejection of pesticides and permeate flux.
- To investigate the significance of experimental variables such as membrane, operating pressure, pesticides concentration and stirring rate and on rejection of pesticides and permeate flux by applying general factorial design.
- To model the separation process and validate it using experimental data.

1.5 Scope of Study

This study focused on removal of atrazine and dimethoate from aqueous solution. Atrazine was selected as subject of study because this herbicide is commonly

used in the plantations around the world as well as in Malaysia (Chooi, 2005). Extensive amount of its usage has ranked it among the most common pesticides found in surface water and groundwater (Plakas *et al.*, 2006). On the other hand, dimethoate is also widely used in Malaysia and it is being regulated in guidelines for drinking water by World Health Organization. Nevertheless, data on effectiveness of dimethoate rejection using membranes has not been found so far (WHO, 2005).

Pesticides leaching would normally cause minute concentration of pesticides to be present in water and its chronic effect to the livings has been of more concern. Therefore, four polyamide nanofiltration membranes, NF90, NF200, NF270 and DK were examined for its performance in the pesticides rejection. Although the range of usual run-off case is in μ g/L, the range of concentration used in this study is in mg/L as to consider in case of accidental spill of pesticides in water source.

A comparative study of atrazine and dimethoate rejection and permeate flux with NF90, NF200, NF270 and DK nanofiltration membranes was first conducted to gain initial insight of the performance of each membrane. Deionized water was used as solvent throughout the experimental works in order to get an accurate measurement of performance for pure pesticide concentration in water. However, a sub-section of this research was dedicated to investigating the membrane performance in different water quality. This would provide clearer understanding on nanofiltration performance in the real case of water filtration.

After that, a comprehensive study on the effect of experimental parameters towards the rejection and flux performance of atrazine and dimethoate with the nanofiltration membranes was carried out. Statistical tool was later applied to determine the significance of experimental variables. Finally, verification of

experimental data for the best-performed membrane was done by using Spiegler-Kedem model. This model was selected because unlike other models which are either only valid for reverse osmosis or inorganic molecules, this model has been acknowledged to be valid for retention of organic molecules in nanofiltration (Van der Bruggen and Vandecasteele, 2002).

1.6 Organization of the Thesis

There are five chapters in this thesis. An overview on pesticides in agriculture sector and how pesticides could end up in source of drinking water are outlined in Chapter One.

Chapter Two presents a review of the literature. It is divided into five major sections. The first section gives a review about the current scenario of pesticides pollution in water. This is followed by explanation on pesticides which include their environmental fate and toxicology effect in section two. Detailed information on the pesticides chosen as subject of study, namely atrazine and dimethoate, is also presented in the section. Description on treatment technologies available for pesticides removal is provided in section three. Then, explanation on membrane filtration and nanofiltration process is given in section four. Section five focuses on the statistical approach used for factorial experimental design. A short summary on the literature review is presented in section six.

Chapter Three covers the methodology for the experimental work done in this research. This chapter is divided into six sections. The first section presents the materials such as membranes and chemicals used in the experiments. The second section gives a general description experimental set-up while the third section provides brief explanation on the analysis equipment used in this study. On the other hand, the

fourth section is a general description of the experimental procedures. The fifth section provides brief description on the statistical analysis approach chosen for this research while the final section gives explanation on how the modeling work was done.

Chapter Four presents all the acquired results and discusses on the findings. It is grouped into four main sections. Section one presents the results and discussion on the comparison of performances between the membranes tested by monitoring their performances when using pure water, pesticide solution in pure water and pesticide solution in different water quality. Section two examines the effect of operating conditions to the performances of the nanofiltration membranes while section three presents the statistical analysis done based on the general factorial design in order to investigate the significance of several main operating conditions to the performances of the nanofiltration membranes tested and interaction between the factors as well as the final regression model obtained from ANOVA. In section four, the performance of the best-performed membrane is validated using Spiegler-Kedem model.

Finally, Chapter Five gives the conclusion and some recommendations for future research. The conclusions are written according to the finding found in Chapter Four. Based on the conclusion, recommendations for future work are suggested.

CHAPTER 2

LITERATURE REVIEW

2.0 Introduction

This chapter provides the literature review of the pesticides pollution in water in section one. Section two provides an overview of history and chemical classification of pesticides as well as information regarding atrazine and dimethoate. After that, review of treatment technology available for pesticides removal is presented, followed by review on nanofiltration process. Literature review on statistical tools used to analyzed data is presented in section five and finally, a short summary on this chapter is provided in section six.

2.1 Pesticide

Pesticide is a general term originated in the early days of its development. Initially, they were classified as contact poisons and stomach poisons. Later on, the classification was made based on their chemical composition and their application. Based on their application, pesticides can be classified into insecticides, acaricides, fungicides, bactericides, nematicides, rodenticides, molluscicides, weedicides, herbicides and soil fumigants. Until 1950's, insecticides were the major contributor of pesticides production. However, since 1954 when phenoxyacetic acid, a highly valuable herbicide, was discovered, the importance of weed control in protecting agricultural products was noticed. Since then, herbicides have been dominating the production as well as the market over insecticides (Yedla and Dikshit, 2005).

It was until 1962 when Rachel Carson's *Silent Spring* appeared that people were made aware of the hidden costs of pesticides and their potential in causing

adverse effects on human health and the environment (Wilkinson, 1987). Human can be exposed to pesticides either through skin, oral consumption or respiration (Ballantyne and Marrs, 2004). In order to provide a better picture of the types of pesticides available, their impact to environments as well as toxicology effect, Table 2.1 presents a general overview of several common groups of pesticides which are classified based on their chemical composition.

2.1.1 Atrazine

Atrazine (2-chloro-4-(ethylamino)-6-isopropylamino-s-triazine) is a type of triazine herbicide (Kamrin, 1997). It was discovered in 1952 by scientists from J.R. Geigy Ltd. in Switzerland. Its first application was done in 1954 and it was patented in 1955 (Castelo-Grande *et al.*, 2005). Since then, atrazine has become the most widely used herbicide in agricultural and forestry applications, with 70,000–90,000 tonnes applied annually in the world. Atrazine has high effectiveness in inhibiting the growth of target weeds including variety of plants and some species of algae by interfering with the normal function of photosynthesis (Graymore *et al.*, 2001).

Herbicide contamination of water environment is of considerable concern because of the potential health hazard towards humans and animals, necessitating stringent legislation on the purity of drinking water. Coincidentally, being one of the most frequently applied herbicides in the agricultural sector, atrazine has frequently been detected in fresh water at levels exceeding the permissible limits (Chingombe *et al.*, 2006). This has led World Health Organization (WHO) to set guideline for drinking water and it allows a maximum of 0.002 mg/L of atrazine in drinking water (WHO, 2005). Figure 2.1 shows the molecular structure of atrazine.

Table 2.1: Chemical classification system of pesticide.

Category	General information	Environmental Impact	Toxicology Effect
Carbamates	-Members of this family are effective as	-Generally are non persistent in the	-Chronic exposure may cause adverse effects on
(Kamrin, 1997)	insecticides, herbicides and fungicides,	environment.	organs or acetylcholinesterase levels.
	although they are most commonly used	-Degrade through chemical hydrolysis	
	as insecticides.	and microbial processes and are unlikely	
	-Work on both target and non-target	to bioaccumulate in aquatic systems.	
	species through inhibition of enzyme		
	acetyl cholinesterase, a substance that		
	transmit a nerve impulse from a nerve		
	cell to a specific receptor such as		
	another nerve cell or a muscle cell.		
Organochlorine	-Insecticides composed primarily of	-According to Beitz et al. (1994), a	-May adversely affect fertility and reproduction at
pesticides	carbon, hydrogen, and chlorine	pesticide is considered as high persistent	high doses and have carcinogenic effects at
	(Coming Clean, 2006).	in soil and water if its half-life is more than	chronic exposure (Kamrin, 1997).
	-Powerful pesticides obtained at	1 month. Kamrin (1997) generalized that	
	relatively low cost (Mitchell, 1966).	organochlorine pesticides possesses half-	
	-The compound's resistance to	life between 2 to 10 years after it is	
	biochemical degradation, coupled with	applied to soil.	
	its solubility in fats (lipids), leads to		
	bioaccumulation in living organisms.		
	This has led to its banning or restriction		
	for agriculture uses in many countries		
	(Kamrin, 1997).		

13

Table 2.1 Continue	30 (a)		
Organophosphorus	-Inhibit the enzyme acetyl	-Usually degrade within 2-4 weeks of	-The commercial products are categorized from
pesticides	cholinesterase (AChE) and thus over-	application in soil (Ibrahim et al., 1998).	extremely toxic to moderately toxic. They are
	stimulate neurological activity in	-Lakes and streams may be susceptible	efficiently absorbed by inhalation, ingestion and
	organism (Pope, 1999).	to pesticide runoff if application occurs	skin penetration (Yusof et al., 1995).
	-Have higher toxicity than	prior to rainfall (Kamrin, 1997)	-Although the degradation process usually leads
	organochlorine pesticides, but are		to the formation of less harmful breakdown
	widely used in the world to replace the		products, in some instances, it can produce
	organochlorine pesticides due to its		more toxic products (Kamrin, 1997). Thus, the
	short half-life (Derache, 1977).		widespread uses of OPPs, coupled with the
	-Due to the magnitude of their		relative lack of reliable data on the risks of long-
	application, they are frequently		term exposure to human health have raised
	associated with toxicity to animals and		concern (Yusof et al., 1995).
	and humans (Evgenidou <i>et al.</i> , 2005)		
Phenoxy and	-One of the most controversial phenoxy	-Most of the phenoxy and benzoic acid	-Most phenoxy herbicides are moderately toxic
benzoic acid	compounds is 2,4,5-T. This compound	herbicides are of low persistence in soil,	while most benzoic acid herbicides are slightly
herbicides	was used extensively, together with	lasting up to 2 weeks.	toxic.
	2,4-D, in the formulation of Agent	-However, despite the relatively rapid	-Phenoxy herbicides could cause liver and
	Orange during the Vietnam War	breakdown, they tend to be mobile in soil	kidney damage as well as reproductive effects.
	(Sahabat Alam Malaysia, 1984).	and thus, have the ability to move from	Benzoic acid herbicides could have teratogenic
	- 2,4,5-T has been banned in Malaysia,	soil into surface water or groundwater	effects at very high doses (Kamrin, 1997).
	but 2,4-D is still being sold (Department	(Kamrin, 1997).	
	of Agriculture, 2005).		

Table 2.1 Continued (a)

Table 2.1 Continue	ed (b)		
Phyrethroids	-Have biological origins.	-Phyrethroids concentration decrease	-Categorized to be from slightly to moderately
(Kamrin, 1997)		rapidly in pond waters and in laboratories	toxic.
		degradation studies due to sorption,	-Exposure to high doses can be fatal
		suspended particles and plants. Microbial	-Can cause liver effects.
		and photodegradation can also occur.	
Triazine	-Herbicides which consist of a single	-Triazines do not adsorb to soil particles	-Triazine compounds are categorized from
(Kamrin, 1997)	ring structure with three nitrogen atoms.	and may leach through soil.	moderately toxic to slightly toxic.
	-Used against a wide variety of weed	-They are relatively stable in water (i.e.	-It could cause disruption of metabolism of
	species by interfering with	not easily hydrolyzed).	vitamins, adverse reproductive effects and liver
	photosynthesis.		damage with high doses.



Figure 2.1: Structure of atrazine.

2.1.1(a) Environmental Impact

Atrazine is a common soil and water pollutant as atrazine applied to cropland can be transported to groundwater by infiltration or to surface waters by water runoff (Nélieu *et al.*, 2000; Barreiro *et al.*, 2007). This is because it has the properties of high leaching potential, persistence in soil, slow hydrolysis and low vapor pressure (Boyd, 2000). Atrazine also persists under cool, dry conditions, in a stable pH environment (Ta *et al.*, 2006).

Atrazine is soluble in water (28 mg/L, 20°C) and is moderately toxic to fish and highly toxic to aquatic invertebrates. It has half-life in soil and water of over 41 and 55 days, respectively (Perez *et al.*, 2006).

2.1.1(b)Toxicology Effect

Atrazine is a stable and low-biodegradable aromatic compound that can cause adverse effects on human health (Chu *et al.*, 2007). It can exhibit phenomena called biomagnification, which means it has the ability to concentrate in animal tissues as it moves up in the food chain (Castelo-Grande *et al.*, 2005).

Atrazine is classified as possibly carcinogenic by the IARC (International Agency for Research on Cancer) and is related to ovary cancer. The simultaneous intake of nitrate and atrazine holds the risk of formation of the strongly genotoxic *N*-

nitrosoatrazine. Chromosomal damage in human lymphocyst and an enhanced occurrence of non-Hogdkin's lymphoma were reported even at very low concentration of 0.1 μ g/L *N*-nitrosoatrazine (Zhang *et al.*, 2004). Atrazine also has the power to increase the toxicity of arsenic in human cells (Gonzalez-Barreiro *et al.*, 2006).

2.1.2 Dimethoate

Dimethoate (O, O – dimethyl S – methylcarbamoylmethyl phosphorodithioate) is an organophosphorus insecticide with a contact and systemic action. It was introduced in 1956 (Fischer *et al.*, 1997). It is widely used against a broad range of insects and mites and is also used for indoor control of houseflies (Sharma *et al.*, 2005). Dimethoate exerts its neurotoxicity by phosphorylation of the enzyme acetyl cholinesterase (AChE) in the central and peripheral nervous systems (Sivapiriya *et al.*, 2006).

Dimethoate is a kind of high-effective pesticide that is extensively applied in agriculture. However, its residues have a negative effect on the environment and on the health of people because of its toxicity and stability (Zhang *et al.*, 2007). Therefore, dimethoate is listed as one of the chemicals from agriculture activities in which guideline value has been established. Its value in drinking water must be ensured to be below 0.006 mg/L (WHO, 2005). Figure 2.2 shows the chemical structure of dimethoate. It is an aliphatic derivative of organophosphorus pesticides.



Figure 2.2: Structure of dimethoate.

2.1.2(a) Environmental Impact

The acute and lethal effects of dimethoate have been determined in several aquatic invertebrate species. Acetyl cholinesterase (AChE) inhibition causes an accumulation of acetylcholine at nerve synapses and disruption of nerve function (Lundebyel *et al.*, 1997).

Dimethoate is highly soluble in water (25 g/L, 21°C) and it adsorbs poorly to any of the soils (Kamrin, 1997; Hernandez-Soriano *et al.*, 2006). Its high solubility in water has subjected it to considerable leaching as it can move rapidly through soils and end up in surface water or groundwater. It has half-life in soil and water of over 20 and 8 days, respectively (Kamrin, 1997).

2.1.2(b)Toxicology Effect

Dimethoate is identified as moderately toxic compound in U.S. Environmental Protection Agency (Wilczek, 2005). Dimethoate poisoning is usually associated with neuromuscular transmission block in both animals and humans (Sivapiriya *et al.*, 2006). Doull (1989) reported that dimethoate could cause mutagenicity, fetotoxicity and reproductive effects. Thyroid impairment can also occur due to its toxicity (Mahjoubi-Samet *et al.*, 2005).

Shahar *et al.* (2005) reported that a patient developed overt parkinsonism presenting with a resting tremor, expressionless face and lack of blinking along with marked cogwheel rigidity after an accidental ingestion of a raw eggplant sprayed with dimethoate. According to a survey on acute poisoning with pesticides in the state of Mato Grosso do Sul, Brazil, dimethoate was also associated with the highest case fatality rate (CFR) compared to other insecticides and herbicides (Recena *et al.*, 2006). In another study done on 802 patients with chlorpyrifos, dimethoate, or fenthion

poisoning admitted to three hospitals in Sri Lanka, the proportion of dying was significantly higher with dimethoate and dimethoate-poisoned patients died sooner than those ingesting other pesticides (Eddleston *et al.*, 2005).

More alarmingly, omethoate, the degradation product of dimethoate has been proven to be 10 times more toxic than its parent compound and it is more of a potential inhibitor to cholinesterase activity than dimethoate (Evgenidou, 2006). Moreover, the synergy effect of dimethoate-atrazine is more lethal than the effect of the individual pesticide since the toxicity of dimethoate was enhanced significantly when they are in binary combination (Anderson and Zhu, 2004).

2.2 Pesticides Pollution in Water

Pesticide pollution in water may arise from runoff and leaching. Only a part of the applied amount of a pesticide is bioactive while the rest is distributed in the environment. Certain remaining amounts attached to the soil are leached out, migrate into groundwater or are distributed by surface runoff. A certain quantity reaches the air and can diffuse over long distances. The diagrams of pesticides cycle can be referred in Figure 1.1 and Figure 1.2.

Contamination of surface and ground waters by pesticides has been documented by several researchers in recent years. The findings are summarized in Table 2.2. In fact, accidents of pesticides' spills had been recorded in United States in 1969 and 1986. These two accidents were the cause for the development of chemical monitoring and acceleration of the sanitation programme and their implementation on the control of water quality (Van-Urk *et al.*, 1993).

Reference	Findings/Conclusions
Gotz <i>et al.</i> (1998)	Elevated amounts of some triazines pesticides and
	dimethoate were recorded in the River Elbe near Hamburg.
El-Kabbany et al.	The water monitoring study confirmed the presence of
(2000)	organophosphorus pesticides and carbamate pesticides in
	drainage water and canal water in El-Haram, Giza.
Zhang et al. (2002)	Total concentration for organophosphorus pesticides ranged
	from 134.8 to 354.6 ng/L and total concentration for
	organochlorine pesticides ranged from 115.4 to 414.7 ng/L
	was detected at Jiulong River Estuary, China.
Gomez-Gutierrez et al.	Concentrations ranged from 0.4 to 19.5 ng/L for the
(2006)	organochlorine compounds and up to 170 ng/L for the more
	polar pesticides were found from the samples from the Ebro
	River, Spain.
Guzzella et al. (2006)	The results of the two-year monitoring campaign allowed to
	conclude that atrazine, even though it was banned in Italy in
	1986, was still the main groundwater contaminant. It was
	present in 100% of the considered samples and 30% of
	them exceeded the allowable limit.
Konstantinou et al.	Pesticides which were mostly detected were herbicides,
(2006)	organophosphorus insecticides as well as the banned
	organochlorines insecticides due to their persistence in the
	aquatic environment. Rivers were found to be more polluted
	than lakes. The detected concentrations of most pesticides
	followed a seasonal variation, with maximum values
	occurring during the late spring and summer period followed
	by a decrease during winter. Nationwide, in many cases, the
	reported concentrations ranged in low ppb levels. However,
	elevated concentrations were recorded in areas of high
	pesticide use and intense agricultural practices.
Lapworth and Gooddy	Diuron was observed in 90% of groundwaters sampled
(2006)	between year 2003 and 2004. Longer-term (1989-2005)
	monitoring showed that pollution of the aquifer by atrazine,
	simazine, and more recently diuron, shows a positive
	correlation with periods of high groundwater levels.

Table 2.2: Summary of cases of pesticides contamination in water.

Table 2.2 Continued.

Morvan <i>et al.</i> (2006)	Five years after the ending of atrazine application which was
	used since the sixties, atrazine and its degradation product,
	deethylatrazine were still systematically quantified at the
	outlet of the watershed with concentrations from 0.07 to 0.43
	$\mu g/L$ for atrazine, and between 0.14 and 1.16 $\mu g/L$ for
	deethylatrazine.
Blanchoud et al. (2007)	The obtained data showed a similar contribution by urban
	pesticides in the Marne River, France due to runoff over
	impervious surfaces as compared to agricultural pesticides
	used on cultivated soils (about 11 tons/year in both cases).
Moore et al. (2007)	Both surface water and sediment phases of the three
	Mississippi Delta oxbow lakes examined had greater levels
	of current-use pesticide contamination than other previously
	studied watersheds due to more intensive cultivation and the
	concomitantly greater amounts of pesticide run-off from
	adjacent agricultural fields.

The implementation on the control of water quality is important because different type of pesticides have different decaying period. Pesticides such as organophosphorus pesticides have half-life between several weeks to a few months while organochlorine pesticides can accumulate in the environment and in livings for years. The persistence and fate of pesticides is also dependent to a multitude of environmental factors such as soil types, temperature, light, moisture, microorganism, etc. It is for these reasons that no absolute half-life can be attributed to any pesticides (Lichtenstein, 1972). A study by Halimah *et al.* (2005) showed that a double increment from recommended pesticides dosage increases the day of detection after treatment from one day to five days. In a study carried out in Mississippi soybean fields, it was found that the downstream of the Yazoo River was contaminated with methaxychlor and endosulfan even after 3 weeks and 3.5 km far from the application site (Yedla and Dikshit, 2005).

During runoff events, residues of pesticides find way into coastal waters, rivers, estuaries and mangroves and were reported to be responsible for many massive fish kills (Yedla and Dikshit, 2005). Even after degradation, the breakdown products of pesticides can be less, more or similar in toxicity when compared to the parent chemical. This scenario inadvertently affects the aquatic livings by enhancing or decreasing populations, inhibiting or stimulating respiration or inhibiting the growth of the aquatic livings (Sumasundaram and Coats, 1990). If this situation is left unattended, it will eventually lead to ecological chaos. Besides, more humans are falling victim to the man-made poisons (Sahabat Alam Malaysia, 1984). In view of this unhealthy scenario, more attention has been given by regulatory bodies to monitor the water quality from pesticides contamination.

In Malaysia, no specific local water regulation on pesticides content has been implemented by government so far. However, there is a provision for the regulation of import, manufacture and sales of pesticides which exists under the Pesticides Act 1974 (amended in 2004). Its objective is to ensure that pesticides imported, manufactured and sold in the country are of good quality and that they will not cause adverse effects on man, food crops and the environment (Department of Agriculture, 2006). This outline of objective is still very wide, general and vague. Nevertheless, according to the Drinking Water Quality Standards in Malaysia, the content of certain pesticides in drinking water is monitored up to four times a year by referring to the limit set by World Health Organization (WHO) (Perbadanan Bekalan Air Pulau Pinang, 2007).

2.3 Treatment Technologies for Pesticides Removal

Unlike heavy metals and other pollutants, pesticides are lethal to the environment even at micro level of concentrations (Yedla and Dikshit, 2005). The conventional water treatment processes such as alum coagulation, clarification and chlorination had been concluded to produce insignificant pesticides removal (Sisodia *et*

al., 1996). This situation was also demonstrated at the A.H. Weeks drinking water treatment plant at Ontario, Canada, whereby sand filtration and flocculation by aluminum sulfate were proven ineffective in removing trace levels of atrazine (Hua *et al.*, 2006). Hence, in order to protect the environment and to meet the stringent enforcement regulations, many researchers are competing to produce effective, reliable and economical way for pesticide-containing water treatment system.

Current treatment technologies for pesticides treatment can be divided into three categories. They are chemical, physical and biological methods. Chemical treatment using advanced oxidation processes (AOP's) has gained interest of researchers lately due to its ability to destruct organic compounds. Biological method is still in its infancy in this area and it is still facing many challenges that needed to be overcome. On the other hand, physical treatment such as adsorption and membrane filtration has produced satisfactory results in removing pesticides from water. Table 2.3 summarizes the available methods for pesticides removal and findings by researchers.

2.4 Membrane Filtration

Membrane is a barrier which separates two phases and restricts the transport of various chemical species (Strathmann, 1990). Microfiltration, ultrafiltration and reverse osmosis are among the well established membrane separation processes. Ultrafiltration and microfiltration are basically similar in their mode of separation. They separate by molecular sieving through increasingly fine pores. However, in reverse osmosis membranes, the membrane pores are so small that solutes permeate the membrane by dissolving in the membrane material and diffusing down on a concentration gradient. Separation occurs because of the differences in solubilities and mobilities of different solutes in the membrane (Baker, 2004). Nanofiltration is the most recently developed pressure-driven membrane separation process and has properties that lie between those of ultrafiltration and reverse osmosis (Oatleya *et al.*, 2005).

Table 2.3: Treatment methods available for pesticides removal and findings by researchers.

Treatment methods	Findings/Conclusions	References
Advanced oxidation processes (AOP's)	-Based on the production of hydroxyl radicals as oxidizing agents to mineralize synthetic organic chemicals. -These radicals are the second strongest oxidative species after fluorine (reduction potential $E_0 = 2.8$ V) and they attack unselectively most of the organic molecules, resulting in a partial or total decomposition. - The commonly applied AOP's are ozonation, Fenton's process and photocatalytic oxidation.	Evgenidou <i>et al.</i> (2005)
(a) Ozonation	-Decrease of 66–96% of the atrazine concentrations was observed when ozone treatment was integrated in the A.H. Weeks drinking water treatment plant which serves the City of Windsor, Ontario Canada.	Hua <i>et al.</i> (2006)
	-Complete degradation of a mixture of several pesticides (alachlor, atrazine, chlorfenvinphos, diuron and isoproturon) in aqueous solutions at pilot-plant scale was found to be slow. -It was also hard to accomplish as large amounts of the oxidant were required to lower the organic content of the solutions.	Maldonado <i>et al.</i> (2006)
(b) Fenton's process	-Studied on the oxidation of atrazine in the presence of hydrogen peroxide (H ₂ O ₂) and ferrihydrite at different concentrations and pHs. -The decomposition product of atrazine was detected only after 1 day of experiment and a decrease of atrazine by 21% was observed over a period of 8 days.	Barreiro <i>et al.</i> (2007)
(c) Photocatalytic oxidation	-Studied on photocatalytic oxidation of azynphos-methyl and dimethoate in water. -It was found that TiO ₂ absorbed light only in the narrow UV range of the solar spectrum, therefore, FeCl ₃ , another photocatalyst and also, probably, photosensitizer for TiO ₂ , must be used. -Concluded that solar light can be used together with small amount of FeCl ₃ and TiO ₂ for a tertiary treatment to eliminate pollutant as pesticides in a short time as the decomposition took place at higher rates compared to photocatalytic oxidation by using only either of them.	Dominguez <i>et al.</i> (1998)
	-TiO ₂ was a more efficient photocatalyst compared to ZnO since the oxidation and decomposition of dimethoate proceeded at higher reaction rates. -However, complete mineralization was not achieved in both systems and the authors proposed longer irradiation time or higher quantities of the catalysts for better performance.	Evgenidou <i>et al.</i> (2005)