

**EFFECT OF RIFAMPICIN AND OMEPRAZOLE IN
COMBINATION WITH CHLOROQUINE ON
CHLOROQUINE RESISTANT PLASMODIA
IN VITRO AND IN VIVO**

By

LOW JEN HOU

**Thesis submitted in fulfillment of the requirements for the degree of
Master of Science**

January 2007

**EFFECT OF RIFAMPICIN AND OMEPRAZOLE IN
COMBINATION WITH CHLOROQUINE ON
CHLOROQUINE RESISTANT PLASMODIA
IN VITRO AND IN VIVO**

By

LOW JEN HOU

Thesis submitted in fulfillment of the requirements for the degree of

Master of Science

January 2007

ACKNOWLEDGEMENTS

For their invaluable contribution in the successful completion of this thesis, I would like to express my deepest gratitude and heartiest thanks to the following:

First and foremost to my supervisor Assoc. Prof. Dr S. Sivachandra Raju whom I cannot thank enough for his continuous assistance and invaluable advice with many aspects of the study, throughout the duration of the project (including its initial design, technical aid in the in vivo study) and training in critical thinking. His great patience and tolerance with the many difficulties I faced initially during experimentation is very much appreciated.

To my co-supervisor Assoc. Prof. Dr M. Ravichandran for providing invaluable technical aid in the in vitro study, resolving many knotty issues with culture and for always being a constant source of good advice and an example of diligence.

To my co-supervisor Prof. Norazmi Mohd Nor for helpful and scholarly advice on direction of project and providing the needed material for the in vivo study.

The Department of Pharmacology and Microbiology for providing much need support and research facilities and to Universiti Sains Malaysia for the short term research grants.

My senior colleague Noor A'shikin Azahri, who was in the early stages of the project, a source of good advice, encouragement and enthusiasm.

My colleague Khairul Mohd Fadzli Mustaffa for providing massive technical assistance during the study, for his unflagging spirit and diligence and organization as well as general assistance in literature review.

My colleagues Tan Hooi Min and Nasriyyah for providing much needed technical assistance during the study.

My senior colleague Syed Atif Ali for scholarly advice on various aspects of tissue culture and to my senior colleague Robaiza for showing me the ropes for in vivo studies and a source of scholarly advice on various aspects of maintaining parasites in vitro.

I would like to extend my thanks also to Microbiology staff En. Mohd Shukri Abdullah and En. Abdullah Bujang for guidance on slide interpretation and also to En. Chan Guan Tiong for meticulous maintenance of the cultures in liquid nitrogen. I would like to thank Dr Ayub for statistical advice and a 2nd opinion on result interpretation. My thanks goes out to Dr Mohd Suhaimi and Dr Wan Nazirah for their help in checking the manuscript.

Last but definitely not least, a very special thank you to my parents for their unconditional support and understanding.

TABLE OF CONTENTS

	Page
Acknowledgements	i
Table of Contents	iii
List of Tables	vii
List of Figures	ix
List of Abbreviations	xiii
Abstrak	xiv
Abstract	xvii
	Page
CHAPTER 1: INTRODUCTION	
1.1 Malaria overview	1
1.2 Malaria lifecycle and clinical manifestations	3
1.3 Parasite genetics and biochemistry	6
1.4 Methods of parasite detection and enumeration	12
1.5 In vivo model: murine malaria	16
1.6 Chloroquine	21
1.7 Current therapy for chloroquine resistant malaria	23
1.8 Drug transport	26
1.8.1 General	26
1.8.2 P-glycoproteins (P-gp)	27
1.8.3 P-gp substrates and inhibitors	30

1.9 Mechanism of action of chloroquine and its transport	34
1.10 Chloroquine resistance and postulated mechanisms	37
1.11 Previous chemosensitizer studies	47
1.12 Chemosensitizers assayed in current study	48
1.12.1 Rifampicin	48
1.12.2 Omeprazole	50
1.13 Objective of study	53

CHAPTER 2: MATERIALS AND METHODS

2.1 In vitro study	54
2.1.1 Preparation of components for <i>P.falciparum</i> culture	54
2.1.1.1 Parasite material	54
2.1.1.2 Preparation of human serum	54
2.1.1.3 Preparation of human blood	55
2.1.1.4 Preparation of RPMI 1640 culture media	56
2.1.2 Retrieval of <i>P. falciparum</i> from liquid nitrogen	58
2.1.3 Continuous culture of <i>P. falciparum</i> in vitro	60
2.1.4 Determination of parasitemia through thin smear light microscopy	61
2.1.5 Cryopreservation of <i>P. falciparum</i>	62
2.1.6 Synchronization of <i>P. falciparum</i> cultures	63
2.1.7 Drug testing for <i>P. falciparum</i>	64
2.1.7.1 Drug preparation	64

2.1.7.2	Conducting in vitro tests	65
2.1.7.3	Determination of parasite growth through radioactive tritiated hypoxanthine	66
2.1.7.4	Statistical analysis	68
2.2	In vivo study	69
2.2.1	Malarial parasite, animal host and general setup	69
2.2.2	Initiation of infection and passaging to subsequent hosts	69
2.2.3	Drug testing for <i>P. berghei</i>	70
2.3	Determination of genotype of chloroquine resistance for TM90C2B	72
2.3.1	Genomic DNA extraction of <i>P. falciparum</i>	72
2.3.2	Primer design	73
2.3.3	PCR	75
2.3.4	Electrophoresis and DNA purification	76
CHAPTER 3:	RESULTS	79
3.1	Genetics of chloroquine resistance	79
3.2	In vitro drug combinations	84
3.2.1	Chloroquine and Rifampicin on HB3 (CQS strain)	84
3.2.2	Chloroquine and Rifampicin on TM90C2B (CQR strain)	89
3.2.3	Chloroquine and Rifampicin on Dd2 (CQR strain)	94

3.2.4 Chloroquine and Omeprazole on HB3 (CQS strain)	99
3.2.5 Chloroquine and Omeprazole on TM90C2B (CQR strain)	104
3.2.6 Chloroquine and Verapamil on Dd2 (CQR strain)	109
3.3 In vivo drug combinations	
3.3.1 Chloroquine and Rifampicin on NK65	114
3.3.2 Chloroquine and Omeprazole on NK65 and ANKA	122
CHAPTER 4: DISCUSSION AND CONCLUSION	
4.1 Genetics for chloroquine resistance in TM90C2B	140
4.2 Response to Chloroquine	144
4.3 Response to Verapamil and combination therapy with Chloroquine	145
4.4 Response to Rifampicin and combination therapy with Chloroquine	152
4.5 Response to Omeprazole and combination therapy with Chloroquine	158
4.6 Conclusion	163
REFERENCES	164
APPENDICES	186
Appendix I Sequencing results and comparison with known genes	
Appendix II Content of solutions/reagents	
Appendix III Drug structures, molecular weights and unit conversion	
Appendix IV Miscellaneous pictures of study	
Appendix V Papers/Presentations arising from this thesis	

LIST OF TABLES

		Page
Table 1.1	Mortality due to malaria (1,000s) in year 2000	2
Table 1.2	Comparison of various assays for detection and enumeration of Plasmodia in vitro	15
Table 1.3	Different characteristics of <i>P. berghei</i> and human parasites	20
Table 3.1	Summary of sequencing results	83
Table 3.2	IC ₅₀ and FIC values for various combinations of chloroquine and rifampicin against <i>P. falciparum</i> strain HB3	88
Table 3.3	IC ₅₀ and FIC values for various combinations of chloroquine and rifampicin against <i>P. falciparum</i> strain TM90C2B	93
Table 3.4	IC ₅₀ and FIC values for various combinations of chloroquine and rifampicin against <i>P. falciparum</i> strain Dd2	98
Table 3.5	IC ₅₀ and FIC values for various combinations of chloroquine and omeprazole against <i>P. falciparum</i> strain HB3	103
Table 3.6	IC ₅₀ and FIC values for various combinations of chloroquine and omeprazole against <i>P. falciparum</i> strain TM90C2B	108
Table 3.7	IC ₅₀ and FIC values for various combinations of chloroquine and verapamil against <i>P. falciparum</i> strain Dd2	113
Table 3.8	Effect of chloroquine and rifampicin on day 7 survival of mice injected with <i>P. berghei</i> strain NK65	120
Table 3.9	Effect of chloroquine and rifampicin on recrudescence of <i>P. berghei</i> strain NK65 in mice	121
Table 3.10	Effect of various doses of omeprazole in combination with chloroquine 5 mg/kg/d on clearing <i>P. berghei</i> strain NK65	130

Table 3.11	Effect of various doses of omeprazole in combination with chloroquine 5 mg/kg/d on day of recrudescence of <i>P. berghei</i> strain ANKA	131
Table 3.12	Effect of various doses of omeprazole in combination with chloroquine 5 mg/kg/d on 7 day survival and 30 day survival of mice infected with <i>P. berghei</i> strain ANKA	132
Table 3.13	Effect of various doses of omeprazole in combination with chloroquine 10 mg/kg/d on clearing <i>P. berghei</i> strain ANKA	135
Table 3.14	Effect of various doses of omeprazole in combination with chloroquine 10 mg/kg/d on day of recrudescence of <i>P. berghei</i> strain ANKA in mice	136
Table 3.15	Effect of various doses of omeprazole in combination with chloroquine 10 mg/kg/d on 7 day survival and 30 day survival of <i>P. berghei</i> strain ANKA in mice	137

LIST OF FIGURES

		Page
Figure 1.1	Lifecycle of Plasmodia	4
Figure 1.2	Acute, chronic, and pregnancy-related manifestations of malaria	5
Figure 1.3	Various postulations on mechanism of increased cell membrane permeability	8
Figure 1.4	Purine metabolism in the parasite	10
Figure 1.5	Haemoglobin digestion and haeme polymerization	11
Figure 1.6	Molecular structure of chloroquine	22
Figure 1.7	Cross sectional view of p-glycoprotein	33
Figure 1.8	Areas with reduced susceptibility of <i>P. falciparum</i> to chloroquine and sulfadoxine-pyrimethamine (SP) and areas designated as multidrug resistant according to WHO.	38
Figure 1.9	Postulated mechanisms of chloroquine transport in <i>P.falciparum</i>	45
Figure 1.10	Molecular structure of rifampicin	52
Figure 1.11	Molecular structure of omeprazole	52
Figure 3.1	A photograph showing typical DNA bands for unpurified pfcr1 76, pfmdr 86 and pfcr1 219 products	80
Figure 3.2	A photograph showing typical DNA bands for unpurified pfmdr 86 and pfmdr 1034 products.	81
Figure 3.3	Photographs showing typical DNA bands for all purified PCR products.	82
Figure 3.4	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose rifampicin (RIF) against <i>P. falciparum</i> strain HB3	85
Figure 3.5	Dose-Response curves for effect of chloroquine (CQ) combined with higher dose rifampicin (RIF) against <i>P.</i>	86

	<i>falciparum</i> strain HB3	
Figure 3.6	Isobologram of chloroquine (CQ) with rifampicin (RIF) for <i>P. falciparum</i> strain HB3	87
Figure 3.7	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose rifampicin (RIF) against <i>P. falciparum</i> strain TM90C2B	90
Figure 3.8	Dose-Response curves for effect of chloroquine (CQ) combined with higher dose rifampicin (RIF) against <i>P. falciparum</i> strain TM90C2B	91
Figure 3.9	Isobologram of chloroquine (CQ) with rifampicin (RIF) for <i>P. falciparum</i> strain TM90C2B	92
Figure 3.10	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose rifampicin (RIF) against <i>P. falciparum</i> strain Dd2	95
Figure 3.11	Dose-Response curves for effect of chloroquine (CQ) combined with higher dose rifampicin (RIF) against <i>P. falciparum</i> strain Dd2	96
Figure 3.12	Isobologram of chloroquine (CQ) with rifampicin (RIF) for <i>P. falciparum</i> strain Dd2	97
Figure 3.13	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose omeprazole (OMP) against <i>P. falciparum</i> strain HB3	100
Figure 3.14	Dose-Response curves for effect of chloroquine (CQ) combined with higher dose omeprazole (OMP) against <i>P. falciparum</i> strain HB3	101
Figure 3.15	Isobologram of chloroquine (CQ) with omeprazole (OMP) for <i>P. falciparum</i> strain HB3	102
Figure 3.16	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose omeprazole (OMP) against <i>P. falciparum</i> strain TM90C2B	105
Figure 3.17	Dose-Response curves for effect of chloroquine (CQ) combined with higher dose omeprazole (OMP) against <i>P. falciparum</i> strain TM90C2B	106

Figure 3.18	Isobologram of chloroquine (CQ) with omeprazole (OMP) for <i>P. falciparum</i> strain TM90C2B	107
Figure 3.19	Dose-Response curves for effect of chloroquine (CQ) combined with lower dose verapamil (VPL) against <i>P. falciparum</i> strain Dd2	110
Figure 3.20	Dose-Response curves for effect of Chloroquine (CQ) combined with higher dose Verapamil (VPL) against <i>P. falciparum</i> strain Dd2	111
Figure 3.21	Isobologram of chloroquine (CQ) with Verapamil (VPL) for <i>P. falciparum</i> strain Dd2	112
Figure 3.22	Rate of parasitemia decline for various chloroquine doses for mice infected with <i>P. berghei</i> (NK65)	115
Figure 3.23	Effect of rifampicin on parasitemia in mice infected with <i>P. berghei</i> (NK65) on chloroquine 5 mg/kg/day	117
Figure 3.24	Effect of rifampicin on parasitemia in mice infected with <i>P. berghei</i> (NK65) on chloroquine 10mg/kg/day	118
Figure 3.25	Effect of rifampicin on parasitemia in mice infected with <i>P. berghei</i> (NK65) on chloroquine 20mg/kg/day	119
Figure 3.26	Rate of decline of parasitemia (NK65 and ANKA) for various chloroquine doses	123
Figure 3.27	Response of NK65, ANKA and ANKA pressure group to chloroquine 5 mg/kg/day	124
Figure 3.28	Response of NK65, ANKA and ANKA pressure group to chloroquine 10 mg/kg/day	125
Figure 3.29	Effect of omeprazole on parasitemia in mice infected with <i>P. berghei</i> (ANKA) as compared to controls	127
Figure 3.30	Effect of omeprazole on parasitemia in mice infected with <i>P. berghei</i> (ANKA) on chloroquine 5 mg/kg/day	129
Figure 3.31	Effect of omeprazole on parasitemia in mice infected with <i>P. berghei</i> (ANKA) on chloroquine 10 mg/kg/day	134
Figure 3.32	Effect of omeprazole on parasitemia in mice in ANKA pressure group on chloroquine 5 mg/kg/day	139

Figure 4.1	Hypothetical model for chloroquine resistance based on chloride ion transport	150
Figure 4.2	Hypothetical model for chloroquine resistance based on direct chloroquine transport	151

LIST OF ABBREVIATIONS

ABC	ATP-binding cassette transporter superfamily
ATP	adenosine triphosphate
CDC	Center for Disease Control
CFTR	cystic fibrosis transmembrane conductance regulator
CQ	chloroquine
CQR	chloroquine resistant
CQS	chloroquine sensitive
CYP	cytochrome P-450 mixed function oxidases
<i>dhfr</i>	dihydrofolate reductase gene
<i>dhps</i>	dihydropteroate synthase gene
DMSO	dimethyl sulfoxide
DNA	deoxyribonucleic acid
dNTP	deoxyribonucleoside triphosphate
dpm	disintegrations per minute
dsDNA	double-stranded deoxyribonucleic acid
DV	digestive vacuole
FIC	fractional inhibitory concentration
h	hours
IC ₅₀	median inhibitory concentration
IRBC	infected red blood cells
MDR	multi drug resistance
min	minutes
MRP	multi resistant protein
NAD	nicotinamide adenine dinucleotide
OMP	omeprazole
PCR	polymerase chain reaction
PCV	packed cell volume
Pfcr1	plasmodium falciparum chloroquine resistance transporter
Pfmdr1	plasmodium falciparum multidrug resistance gene
Pgh1	p-glycoprotein homologue 1
P-gp	p-glycoprotein
pLDH	parasite lactate dehydrogenase
RBC	red blood cells
RIF	rifampicin
RNA	ribonucleic acid
RPMI	Roswell Park Memorial Institute
s	seconds
SNP	single-nucleotide polymorphism
TCM	tissue culture medium
VPL	verapamil
WHO	World Health Organization

ABSTRAK

Kekebalan terhadap chloroquine telah mengakibatkan peningkatan kematian dan morbiditi daripada penyakit malaria *P. falciparum*. Ketiadaan vaksin yang berkesan mengakibatkan terapi malaria bergantung pada kombinasi ubat alternatif seperti Mefloquine dan Artesunate dan tahap kekebalan terhadap ubat-ubat alternatif tersebut kerap meningkat. Adalah diketahui bahawa agen perencat glycoprotein P berupaya menghalang kekebalan sel barah terhadap pelbagai ubat melalui pencegahan effluks ubat anti-barah oleh glycoprotein P. *P. falciparum* juga didapati membentuk homolog glycoprotein P (pgh1) pada membrannya dan Verapamil dan pengujakimia lain (contohnya: antihistamin) boleh mengekang kekebalan Chloroquine. Oleh demikian, adalah dicadangkan bahawa agen perencat glycoprotein P boleh berfungsi sebagai pengujakimia yang berkesan kerana pengangkut dalam kelas "ABC superfamili" kerap mempunyai substrat dan agen perencat yang serupa.

Dua agen perencat p-gp (rifampicin dan omeprazole) diuji dalam kajian ini secara in vitro dan in vivo dengan matlamat menghalang kekebalan terhadap Chloroquine.

P. falciparum yang sensitif terhadap chloroquine and yang kebal terhadap chloroquine (strain HB3, TM90C2B, Dd2) dicairkan daripada keadaan sejukbeku dan dikultur dalam media RPMI 1640, sel darah merah 'O' dan serum pada 37°C berdasarkan protocol Trager-Jansen yang diubahsuai. Chloroquine dicairkan secara berperingkat dan diuji bersama Rifampicin/Omeprazole dalam nisbah

kepekatan yang berbeza-beza dalam plat micro ELISA steril yang mengandungi kultur parasit. Selepas tempoh inkubasi yang tertentu, plat ujian dikeluarkan and keputusan dicapai melalui kaedah kematangan schizont atau melalui pergabungan hipozantina radioaktif dengan DNA parasit.

P. berghei (strain NK65, ANKA) juga dicairkan dan disuntik ke dalam ruang peritoneal tikus Swiss Albino yang berumur 6-7 minggu. Bila virulen menjadi stabil, pelbagai amaun Chloroquine diberikan bersama Rifampicin atau Omeprazole mengikut regim Peter's selama 4 hari berturut-turut. Peratusan parasitemia diketahui melalui calitan nipis darah tikus pada sisip kaca. Hari parasit muncul kembali dalam darah dan hari kematian tikus juga dicatat.

Kajian in vitro menunjukkan bahawa Rifampicin bersifat amat antagonis terhadap chloroquine untuk strain HB3 dan TM90C2B (FIC purata >2). Takat antagonis berkurangan apabila tempoh inkubasi drug bersama parasit dipanjangkan ke 72 jam. Keputusan ini disokong dalam kajian in vivo di mana tikus yang diberi rifampicin serta chloroquine menunjukkan parasit muncul kembali dengan lebih awal dan kematian tikus meningkat.

Kajian in vitro Omeprazole menunjukkan ia juga bersifat antagonis terhadap Chloroquine tetapi tidak seteruk berbanding interaksi Rifampicin-Chloroquine. Pada dos harian 5-20 mg/kg, kajian in vivo mendedahkan bahawa Omeprazole mengurangkan kematian tikus dalam tempoh pemerhatian sebulan dan melambatkan kemunculan semula parasit dalam darah, tetapi keputusan tersebut tidak mencapai tahap signifikan bila diuji dengan statistik. Tiada kesan yang nyata pada kadar penurunan parasitemia.

Rifampicin bersifat amat antagonis terhadap chloroquine secara in vitro and in vivo. Maka, kombinasi Rifampicin-Chloroquine patut dielakkan dalam amalan klinikal. Kemungkinan bahawa penggunaan Rifampicin dalam pesakit TB yang juga menghidap malaria boleh menyebarkan kekebalan Chloroquine kerana pemanjangan tempoh untuk membasmi parasit.

Kedua-dua inhibitor p-glycoprotein tidak merupakan pengujakimia yang berkesan buat gejala kekebalan chloroquine. Maka mekanisme kekebalan parasit malaria terhadap Chloroquine perlu diselidik semula untuk menjumpai agen yang lebih sesuai untuk mengekang kekebalan Chloroquine.

ABSTRACT

Chloroquine resistance has resulted in a resurgence of morbidity and mortality from *P. falciparum* malaria. The lack of an effective vaccine forces dependence on alternative drug combination therapies based on mefloquine or artesunate, of which the incidence of multidrug resistance is steadily rising. P-glycoprotein inhibitors reverse multidrug resistance in tumour cells by preventing efflux of anti-neoplastic drugs by p-glycoprotein, an ATP-binding cassette transporter (ABC). *P. falciparum* was also found to possess a p-gp homologue (pgh1) and that verapamil and other chemosensitizers (antihistamines, antidepressants) are able to reverse chloroquine resistance. It was postulated that potent p-gp inhibitors would be effective chemosensitizers as transporters of the ABC superfamily frequently share similar substrates and inhibitors. 2 known p-gp inhibitors (rifampicin and omeprazole) are assayed in this study, *in vitro* and *in vivo*, aiming to achieve chemoreversal of chloroquine resistance.

Chloroquine-sensitive and chloroquine-resistant *P. falciparum* (HB3, TM90C2B, Dd2 strains) were thawed and maintained in continuous culture using RPMI 1640 media, 'O' RBCs and pooled serum at 37°C using a modified Trager-Jansen candle jar protocol. Serial two fold dilutions of chloroquine and assayed chemosensitizers were performed on sterile ELISA microplates in checkerboard pattern to achieve different drug ratios in respective culture wells.

After a fixed incubation period, test plates were harvested and results obtained by assessing schizont maturation or incorporation of radioactive hypoxanthine.

P. berghei (NK65, ANKA strains) were thawed and injected intraperitoneally into naïve Swiss Albino mice (6-7 weeks old). Upon stabilization of virulence, various chloroquine doses combined with rifampicin or omeprazole were administered intraperitoneally following a standard Peters' consecutive 4-day treatment regime. Percentage parasitemia was assessed by thin smears of tail blood. Day of recrudescence and survival within a month were also noted.

In vitro rifampicin showed marked antagonism when combined with chloroquine against *P. falciparum* strains HB3 and TM90C2B (average FIC >2). Antagonism was less marked with a longer incubation period (72 hours). These results were supported in the *in vivo* study where mice receiving rifampicin and chloroquine showed earlier recrudescence and a drop in overall survival.

In vitro tests showed that omeprazole is mildly antagonistic when combined with chloroquine. The *in vivo* study revealed that for ANKA and NK65 strains, omeprazole at doses of 5, 10 and 20 mg/kg/day, slightly improves day of recrudescence and overall survival figures at the end of a month without reaching significance. Effect on rate of decline of parasitemia is very minimal.

As rifampicin is strongly antagonistic to chloroquine both *in vitro* and *in vivo*, such combinations should be avoided in clinical practice. It is possible that concomitant use of rifampicin in TB patients who also contracted malaria, may have helped propagate chloroquine resistance due to the lengthier clearance times of the parasite. As neither p-glycoprotein inhibitor is a suitable chemosensitizer for chloroquine resistance perhaps a reevaluation of resistance mechanisms would expedite the search for a suitable chemoreversal agent.

CHAPTER 1

INTRODUCTION

1.1 Malaria overview

Malaria, a parasitic infection transmitted by certain anopheline mosquitoes, is one of the most prevalent and pernicious diseases of humans, estimated to kill between 700,000 – 2.7 million people (mainly children) worldwide each year as shown in Table 1.1. It affects about 5% of the world's population. (Breman, 2001)

While Malaysia has one of the oldest malaria control programs in the world and large scale eradication programs and vector control measures that have resulted in a steep drop in incidence in the nineties, from 2.99/1000 population in 1994 to 0.56/1000 population in 2000. However despite the general success in controlling malaria, it remains an important health issue in remote areas. A good majority of cases occur in Sabah where emerging chloroquine resistance has been noted.

The widespread availability of cheap and effective anti-malarials, particularly chloroquine and sulphadoxine-pyrimethamine has curbed the extent of mortality and morbidity, but it has also encouraged the development and spread of resistance. While many strategies have been offered to counter chloroquine resistance, the lack of a truly effective vaccine has spurred investigators to intensify the search for cheap and potent new drugs/drug combinations to address this need.

Table 1.1 Deaths and malaria-related deaths (1,000s)

Area	Population	All deaths (%)	Malaria deaths (%)	Malaria/total (%)
World	6,122,210	56,554	1,124	2.0
Africa	655,476	10,681	963 (85.7)	9.0
America	837,967	(18.9)	1 (-)	0.02
Eastern	493,091	5,911 (10.5)	55 (4.9)	1.3
Mediterranean	874,178	4,156 (7.3)	0	-
Europe	1,559,810	9,703 (17.2)	95 (8.5)	0.7
Southeast Asia	1,701,689	14,467	10 (0.9)	0
Western Pacific		(25.6)		
		11,636		
		(20.6)		

Adapted from the World Health Organization World Health Report, 2002

1.2 Malaria lifecycle and clinical manifestations

There are four species of Plasmodia which infect humans, but *Plasmodium falciparum* accounts for the majority of instances of morbidity and mortality. Its virulence and adaptability are important factors which make it a deadly threat. While drug resistance for *P. vivax* has also been noted, this phenomenon is nowhere near as widespread as that of *P. falciparum*.

The lifecycle of a malarial parasite is a complex one as shown in Figure 1.1. An infected mosquito injects sporozoites into the blood of the host. Sporozoites enter liver cells, multiply and release merozoites into the blood stream. Merozoites invade red blood cells and start a phase of multiplication where they mature from ring forms to early trophozoites, late trophozoites, immature schizonts and mature schizonts which burst to release another batch of merozoites; a cycle which can be repeated indefinitely causing malarial symptoms. A small portion of merozoites develop sexually into gametocytes which are then taken up by mosquitoes when they feed, thus completing the cycle. A summary of various clinical manifestations of malaria is given in Figure 1.2.

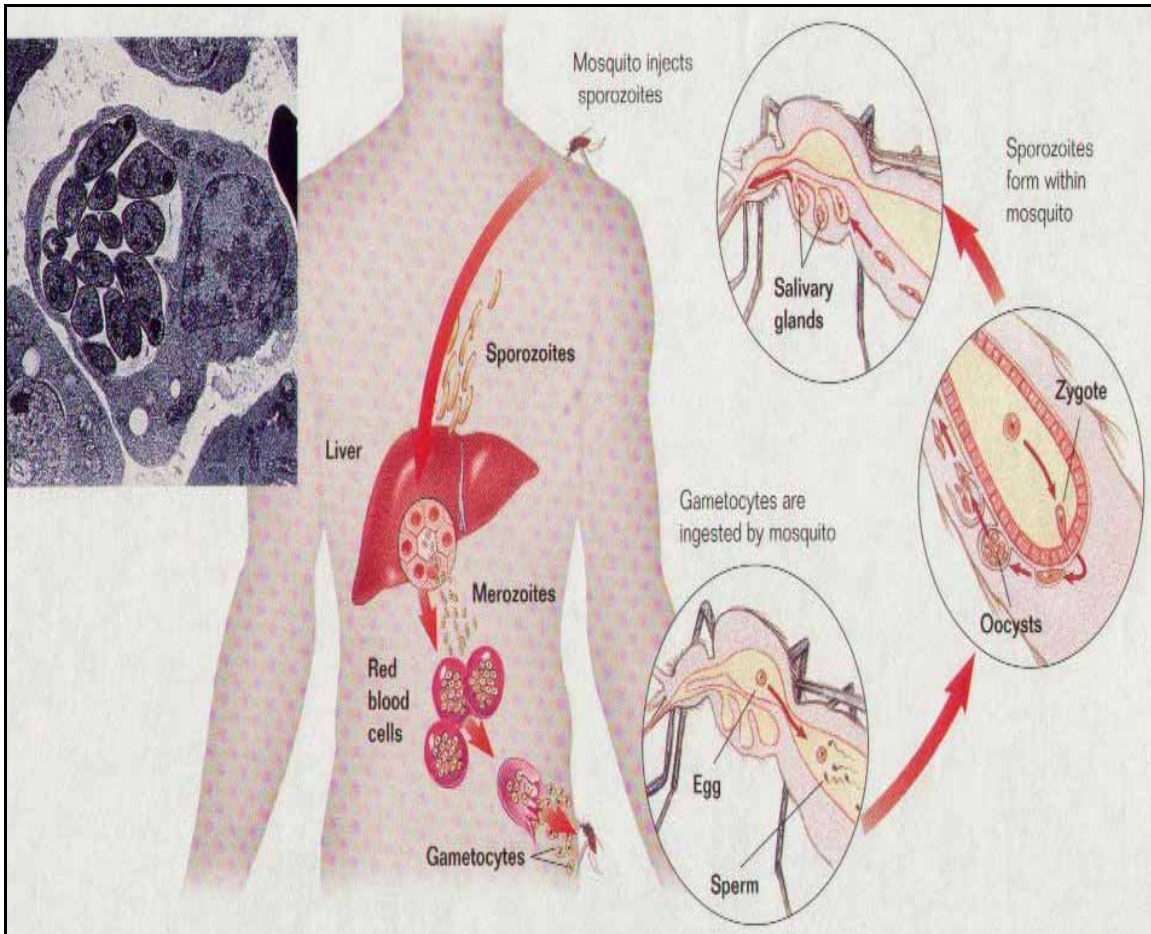


Figure 1.1 Lifecycle of Plasmodia