

## Research



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# The marine nitrogen cycle: recent discoveries, uncertainties and the potential relevance of climate change

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The ocean's nitrogen cycle is driven by complex microbial transformations, including nitrogen fixation, assimilation, nitrification, anammox and denitrification. Dinitrogen is the most abundant form of nitrogen in sea water but only accessible by nitrogen-fixing microbes. Denitrification and nitrification are both regulated by oxygen concentrations and potentially produce nitrous oxide (N<sub>2</sub>O), a climate-relevant atmospheric trace gas. The world's oceans, including the coastal areas and upwelling areas, contribute about 30 per cent to the atmospheric N<sub>2</sub>O budget and are, therefore, a major source of this gas to the atmosphere. Human activities now add more nitrogen to the environment than is naturally fixed. More than half of the nitrogen reaches the coastal ocean via river input and atmospheric deposition, of which the latter affects even remote oceanic regions. A nitrogen budget for the coastal and open ocean, where inputs and outputs match rather well, is presented. Furthermore, predicted climate change will impact the expansion of the oceans' oxygen minimum zones, the productivity of surface waters and presumably other microbial processes, with unpredictable consequences for the cycling of nitrogen. Nitrogen cycling is closely intertwined with that of carbon, phosphorous and other biologically important elements via biological stoichiometric requirements. This linkage implies that human alterations of nitrogen cycling are likely to have major consequences for other biogeochemical processes and ecosystem functions and services.

## 1. Introduction

The nitrogen cycle on the Earth has evolved over three billion years through biogeochemical and microbial processes coupled via natural feedbacks that keep the nitrogen cycle of the oceans in approximate balance [1]. However, recent findings strongly suggest a significant imbalance of the oceans' nitrogen budget towards higher losses than inputs [2,3]. Moreover, new processes have been identified, such as the anaerobic ammonium oxidation (anammox) process [4], the conversion of ammonia and nitrite to dinitrogen gas or the denitrification by eukaryotic species such as foraminifera [5]. Previously, canonical denitrification by prokaryotes was thought to be the only removal process of reactive nitrogen. Likewise, the single major N source to the ocean was considered to be the nitrogen-fixing photoautotrophic bacteria of the genus *Trichodesmium*, a species that occurs globally in tropical and subtropical waters [6]. However, today, numerous other nitrogen-fixing bacteria have been described which may raise the estimate of the global nitrogen input to the oceans significantly [7]. Moreover, nitrogen fixation seems to be a significant process not only in tropical surface waters and some benthic systems [8], but also in anoxic waters, at the sea floor [9], in estuaries and in river plumes where diatom-diazotroph associations prevail [10,11]. Nitrification, the microbially mediated,

two-step oxidation of ammonium to nitrite and nitrate, was traditionally attributed to bacteria, but ammonia-oxidizing archaea now seem even more important [12–14].

On top of these recent findings and uncertainties are the perturbation of the nitrogen cycle by fossil fuel combustion and the production of artificial fertilizers, which together exceed the amount produced by natural nitrogen fixation [15,16]. The extra nitrogen disrupts the N balance in marine systems, fuels eutrophication of coastal oceans and supports extension of hypoxic zones [17]. These sites act as an additional, poorly quantified, source of nitrous oxide ( $\text{N}_2\text{O}$ ). Climate change imposes numerous threats to the functioning of the Earth's elemental cycles. This overview paper will give a brief insight into the functioning of the marine nitrogen cycle, how the major processes are interlinked, what is known and where the critical limits of our understanding are.

## 2. Surface water processes: assimilation of dissolved inorganic nitrogen, dissolved organic nitrogen and nitrogen fixation

### (a) Global patterns in dissolved inorganic nitrogen concentrations

Nitrogen is an essential element for all life forms but, in sea water, nitrogen mostly occurs as inert dissolved  $\text{N}_2$  gas (more than 95%) that is inaccessible to most species. The rest is reactive nitrogen ( $\text{N}_r$ ), such as nitrate, ammonia and dissolved organic compounds. Nitrate concentrations in surface waters vary from almost zero in the tropics and subtropics up to several tens of micromoles per litre in the temperate and Arctic and Antarctic oceans (figure 1a).

The increase in deep-water nutrient concentration from the northern Atlantic towards the Indian and Pacific oceans (figure 1b) is a result of a global circulation pattern, called the oceanic 'conveyor belt' [19]. The circulation is driven by the excess salt in the northern Atlantic Ocean where surface waters sink to the bottom and travel southwards. This southward current partly supplies water to the Antarctic Circumpolar Current, and continues to the Indian and the Pacific Oceans [19] from whence the flow returns to the surface to travel back through the Indonesian Islands the Tasman Sea [20], and further into the Indian and Atlantic Oceans. The residence time of the Atlantic water is around 180 years, whereas the complete overturning of ocean deep waters takes approximately 1000 years [19].

Four major processes deliver nutrient-rich waters to the surface: upwelling, winter mixing, eddies and diffusion from below a permanent or seasonal thermocline (table 1). Coastal upwelling is especially strongly developed in eastern boundary currents along the west coasts off North and South America and Africa, where nutrient-rich water from approximately 200 m is brought to the surface (figure 1a). The upwelled water stimulates primary production and the export of organic material, with the consequence of oxygen deficiency in subsurface waters owing to remineralization. The recycling of organic matter at depth leads to accumulation of nutrients with nitrate concentrations between 20 and  $50 \mu\text{mol l}^{-1}$  (figure 1b). At high latitudes, deep winter-time convection mixes the waters down to a depth of several hundred metres so that seasonal interfaces are destroyed.

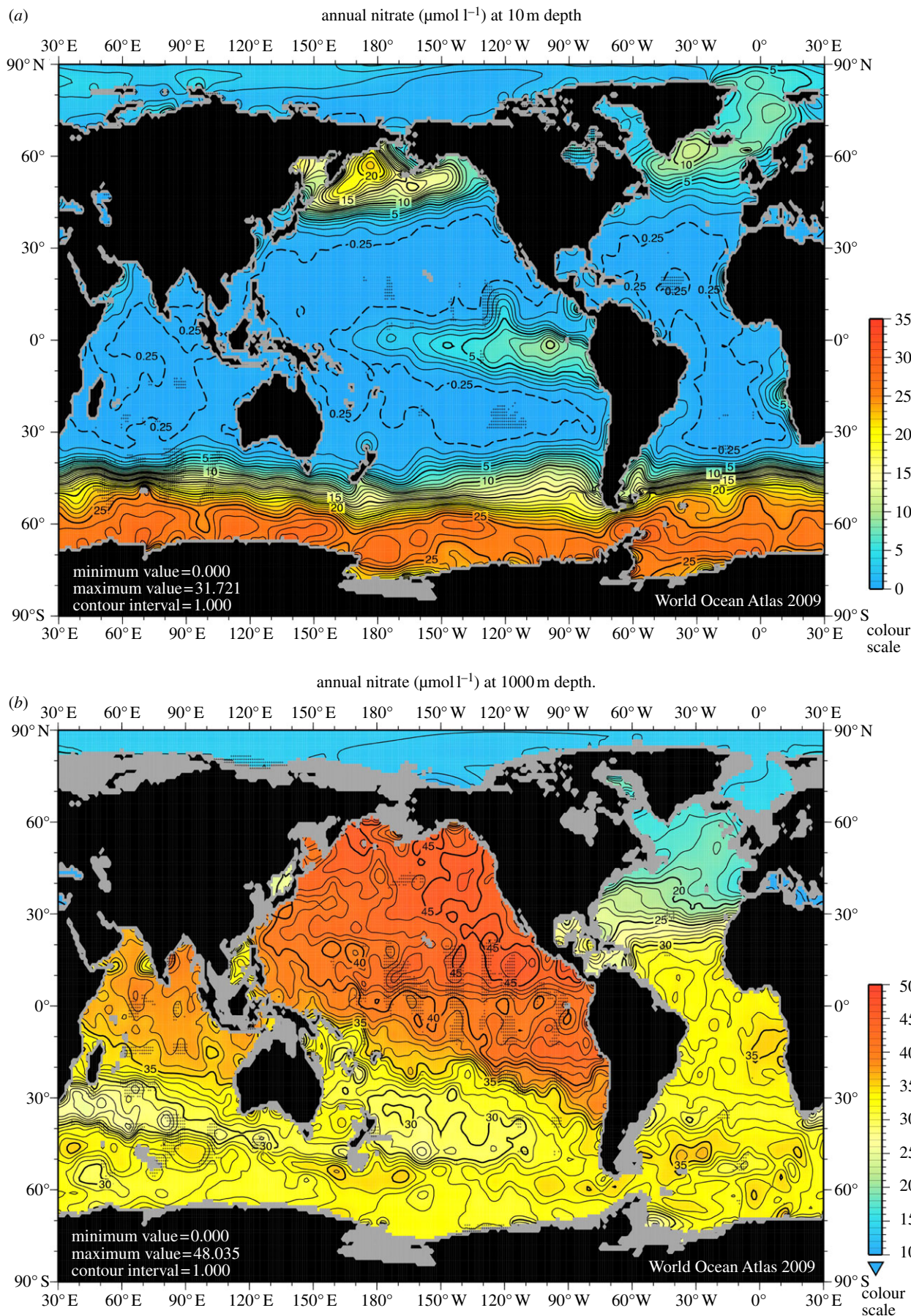
Thus, nutrients from greater depth are redistributed into the surface layer to provide the nutrient reservoir for the spring bloom in the following year. In the open ocean, eddy fields are generated by baroclinic or barotropic instabilities and large-scale gyres by the divergence of the horizontal wind field. Within these, structure nutrients are supplied to the surface by Ekman pumping, stimulating blooms of phytoplankton [21,22]. To what extent these sites contribute to global primary production is difficult to estimate, because the occurrence of eddies is transient and varies greatly in time and location. For the Atlantic Ocean, roughly one-third of the total N flux may be delivered by eddy transport [23]. Phytoplankton living in surface waters drives the N cycle and often consumes all inorganic nutrients down to a depth where ambient light level is 1–0.1% of the surface irradiation. At these depths, turbulent diffusion is another important source for primary production [24].

Our understanding of ocean productivity regulation was significantly advanced when the concept of new and regenerated production was introduced by Dugdale & Goering [25]. They defined new production as that based on the input of nitrate from outside the euphotic zone (including nitrogen fixation), whereas regenerated production (based on ammonium) fuels a 'microbial loop' first introduced by Azam *et al.* [26] within a defined surface layer of the ocean. Based on Dugdale and Goering's concept, only the amount of newly produced biomass can be quantitatively exported to the deep ocean without running down the primary production system [27]. However, this concept assumes that sources of new nitrogen are small, and that the ocean is in steady state, which often seems not to be the case [3,6,28]. Other potential problems with the Dugdale and Goering concept are bacterial uptake of dissolved inorganic nitrogen (DIN), assimilation of dissolved organic nitrogen (DON) and nitrification in the euphotic zone [29,30]. Of the global net primary production in the oceans of  $1900 \text{ Tg C yr}^{-1}$  [31],  $320 \text{ Tg yr}^{-1}$  should be N, using a C : N ratio of 6. To these, about  $140 \text{ Tg N yr}^{-1}$  are added by nitrogen fixation [32], whereas global losses may be around  $400 \text{ Tg N yr}^{-1}$  [33]. A global nitrogen budget considering all natural and human sources and incorporating recent discoveries has not yet been put together, but qualitative attempts have been made [3,6,32,34].

Nitrogen in atmospheric deposition develops into a significant N source in the open ocean, where usually very little combined nitrogen occurs in surface waters [15]. To what extent these atmospheric nutrients support oceanic productivity, and decrease oxygen concentrations, remains unclear, but Dentener *et al.* [35] estimate a total nitrogen deposition of  $46.2 \text{ Tg N yr}^{-1}$ , whereas Duce *et al.* [15] derive a global estimate of  $67 \text{ Tg N yr}^{-1}$ .

### (b) Nitrogen fixation

Nitrogen-fixing organisms are independent of combined N sources and able to use the dissolved  $\text{N}_2$  gas, which has concentrations of over  $400 \mu\text{mol l}^{-1}$  in sea water. In order to be used as a source of nitrogen, dinitrogen has to be reduced to ammonium for which specific enzymes are required [36]. Moreover, nitrogen-fixing species rely on phosphate and require iron in much larger quantities than other phytoplankton [37]. Therefore, co-limitation of Fe and P occurs [38] and may even regulate the global occurrence of nitrogen-fixing species. Because the Fe input from land is lower in the Pacific than the Atlantic Ocean



**Figure 1.** Nitrate concentrations in (a) surface waters of the ocean and (b) at 1000 m depth. Adapted from Boyer *et al.* [18].

[39], the tropical Atlantic Ocean may experience higher overall N fixation than the Pacific [40]. But recent evidence suggests higher rates in the Pacific Ocean than previously assumed, possibly higher than in the Atlantic Ocean [28,41].

Nitrogen fixation in the ocean has long been focused on the large bloom-forming genus *Trichodesmium* [6], until smaller unicellular species were discovered and their potential fixation activity demonstrated [7,42,43]. These are heterotrophic

**Table 1.** Summary of major nitrogen nutrient sources to the ocean.

source of nutrient	major sites of occurrence	example of sites where process is of significance	type of nutrient	timing
convective overturning	high latitudes	North Atlantic, Greenland and Norwegian Seas	$\text{NO}_3^-$	autumn, winter
wind induced and Ekman upwelling	eastern boundary upwelling systems	off Oregon/California/Mexico, Peru/Chile, Iberian Peninsula/Mauretania, Namibia, NW Indian Ocean (Arabian Sea)	$\text{NO}_3^-$ , DON	during upwelling season (depending on the direction of the wind fields)
eddy activity/gyres	everywhere	frontal systems; Gulf stream, Kuroshio, Polar front	$\text{NO}_3^-$	sporadically/permanent
diffusion	thermoclines	oligotrophic ocean	$\text{NO}_3^-$ , $\text{NH}_4^+$	permanent, depending on the gradient
rivers	continental shelves	Amazon, Mississippi, Yangtze, Mekong	$\text{NO}_3^-$ , DON	permanent
atmospheric deposition	coastal, but globally relevant	oligotrophic tropical ocean	$\text{NH}_4^+$ , $\text{NO}_3^-$ , DON	sporadically
shelf processes	on continental shelves	European shelf, Patagonian shelf	$\text{NO}_3^-$ , DON	permanent
regeneration of particles	everywhere in the water column and in sediments	high latitude surface waters in summer, low latitude surface waters, benthic boundary layers	$\text{NH}_4^+$ , $\text{NO}_3^-$ , $\text{NO}_2^-$ , DON	permanent

nitrogen fixers, including the photoheterotroph group-A (UCYN-A), which lacks the photosystem II [44], and *Richelia intracellularis*, a symbiont that fixes nitrogen in diatoms cells and has been found in tropical river plumes [45,46].

Nitrogen fixation may also be an important source of nitrogen to support biological production in deep-sea environments. Mehta *et al.* [47] described a diverse group of organisms possessing *nifH* genes, and therefore potentially capable of fixing  $\text{N}_2$  from hydrothermal vent sites of the Juan de Fuca Ridge system. Other archaeal *nifH* sequences, many of uncertain phylogeny, have been linked to subsurface circulation through hydrothermal vent systems [48] and to methane cold seeps [9,49–51] and a mud volcano [52]. Further work at a cold seep field has revealed that methanotrophic archaea form syntrophic associations with sulfur-reducing bacteria, and that these consortia actively fix  $\text{N}_2$  [9,50,51]. The overall contribution of these benthic diazotrophs to the oceanic nitrogen budget remains poorly known, but may be significant.

Nitrogen fixation rates based on direct measurements are sparse, and the variability is extremely high [6]. Therefore, global estimates of total marine nitrogen fixation are often based on geochemical factors such as unbalanced N:P ratios ( $\text{N}^*$  method) which are assumed to support nitrogen fixation. Recently, Groszkopf *et al.* [53] reported a global marine N-fixation rate of  $177 \text{ Tg N yr}^{-1}$  based on extrapolation of direct measurement, and suggest that  $\text{N}_2$  fixers other than *Trichodesmium* are quite important outside the tropics. From global nutrient distributions and an ocean circulation model, Deutsch *et al.* [32] calculated a nitrogen fixation rate of  $140 \text{ Tg N yr}^{-1}$ , which agrees well with other estimates in the range of  $100\text{--}200 \text{ Tg N yr}^{-1}$  [6]. It seems that lower estimates mainly result from model exercises

where additional factors limiting N fixation such as iron have been implemented [54,55]. The global N-fixation rate of  $140 \text{ Tg N yr}^{-1}$  has been largely unquestioned and is widely used [3].

### (c) The role of dissolved organic nitrogen in nitrogen cycling

Dissolved organic nitrogen is rather uniformly distributed in the water column of the open ocean, with slightly higher concentrations in the surface than at depth, but with DON increasing considerably towards coastal areas and in estuaries [56]. With a mean concentration of  $5.8 \pm 2 \mu\text{mol l}^{-1}$ , DON may potentially be more important than inorganic forms, because DON concentrations comprise between 18 per cent and 85 per cent of the total nitrogen pool in coastal and open ocean surface water, respectively, with particulate nitrogen being negligible [56]. The DON pool is not as inert as suggested by the relatively high and constant concentrations found in the oceans, but a small part of it is rather dynamic and consumed by phytoplankton and bacteria [57]. Furthermore, DON is mainly of autochthonous origin because it stems from direct release by phytoplankton and bacteria [58,59], egestion and excretion from micro- and meso-zooplankton [60], or viral lysis of bacterioplankton [61].

In coastal waters, rivers are a major source of allochthonous DON, and its composition, bioavailability and quantities may vary with land use [62]. Another allochthonous source relevant also for the open ocean is the DON in atmospheric deposition [63,64]. DON is actively channelled into cells via membrane transport systems [65] and seems to be a quite active component of coastal nitrogen cycling [66,67]. Better understanding

of the dynamic of DON is essential to quantify its role in the nitrogen and carbon cycles and how these will respond to anthropogenic perturbations and global change.

#### (d) Stoichiometry of C : N : P in the ocean

The close similarity between nitrogen and phosphorous ratios in plankton and in deep-water nutrients was first noted by Redfield [68], who suggested that life in the ocean adjusts the nutrients according to its requirements. Today, the perception is, rather, vice versa, that life has adjusted to the oceanic ratios. The C : N : P ratio of 106 : 16 : 1 on a molar basis is still a fundamental concept in marine sciences and mirrors the metabolic demands of an average living cell [69]. This 'Redfield ratio' allows linking elemental cycles and has been widely used in ecosystem and element flux models [32]. The cellular stoichiometry is important for the regulation of organic C and N cycling and also for the linkage between single-cell activity and ecosystem function. Heterotrophic organisms maintain low nitrate concentrations in the water when organic C : N ratios match the stoichiometric demands, but as soon as organic carbon becomes limiting, denitrification decreases, and nitrification is enhanced [70]. It may be quite important to focus nutrient management scenarios to maintain balanced elemental ratios because nitrogen supply in excess of the demand leads to significant enrichment of  $N_r$  [71].

The ultimately limiting element for ocean productivity had been debated over decades, and a consensus seems to be that nitrogen is limiting on short time-scales, whereas on geological time scales it seems to be phosphorus [72,73]. Both these views are based on input and loss rates and the turnover of the elements, which all may vary over time [74]. However, human perturbations affecting the input or loss of substances ultimately impact the global elemental cycles and the productivity.

### 3. Biogeochemistry of oxygen deficiency zones

Oxygen controls the distributions of N-cycle processes based on the fact that some microbial reactions require oxygen and others are inhibited by it. Both the microbial pathways that lead to net loss of fixed nitrogen, denitrification and anammox, occur only in the near or total absence of oxygen. These conditions occur in coastal and shelf sediments around the world and in a few locations in the water column where relatively high oxygen utilization rates and low ventilation rates lead to oxygen depleted zones of several hundred metres thickness (e.g. Arabian Sea). Regions where the oxygen concentration is low enough to induce anaerobic metabolism are known as oxygen minimum zones or more precisely oxygen-deficient zones (ODZs) experiencing oxygen concentrations less than  $10 \mu\text{mol l}^{-1}$ , an expression we use throughout this text. Nitrogen cycling in ODZs is fundamentally different from the rest of the open ocean, because only in ODZs can net loss of fixed N occur in the water column.

Denitrification and anammox both lead to the formation of gaseous dinitrogen (and  $N_2O$  in the case of the former) and both consume DIN, but have potentially different effects on the carbon cycle. Denitrifying bacteria are mostly heterotrophs, and respire DIN at  $O_2$  concentrations below  $60 \mu\text{mol l}^{-1}$  while oxidizing organic matter. Some denitrifiers, however, are autotrophs, and fix  $CO_2$  while oxidizing reduced sulfur

compounds with nitrate. Anammox bacteria are also autotrophic, fixing  $CO_2$  while oxidizing ammonium plus nitrite to dinitrogen, and oxidizing nitrite to nitrate, all anaerobically. In the open ocean ODZs, denitrification and anammox are both constrained by the stoichiometry of the organic matter supply, and anammox must depend on denitrification for the continuous supply of DIN. The complete utilization of organic matter of average composition (see stoichiometry above) under anaerobic conditions should lead to the loss of fixed N from both denitrification and anammox in the ratio of 71 : 29, i.e. 29 per cent of the fixed N loss is attributed to anammox.

In sediments and coastal systems, however, there are multiple sources of organic matter and DIN, such that the average composition rule does not apply. With an allochthonous supply of ammonium in a stratified basin or sediment, the proportion of anammox can be much higher. It was surprising, therefore, that the first reported direct measurements of anammox and denitrification in the ODZs (where the average rule should apply) failed to detect denitrification. This suggested that all of the fixed N loss was due to anammox. This is apparently a robust result, although the number of measurements is still rather small [75–77]. Only in the Arabian Sea has denitrification been detected at appreciable levels [77,78], whereas another study nearby failed to detect either anammox or denitrification [79]. There are several alternative explanations for the variation in the proportion of anammox and denitrification in the ODZs, but this is an active area of research with no current consensus.

From stoichiometric model studies, Koeve & Kähler [80] conclude that the proportion of anammox and denitrification does not vary widely in the ocean, and direct measurements of excess  $N_2$  and DIN concentrations verify that organic matter loss in the ODZs does indeed produce net Redfield stoichiometry [81]. Thus, resolving the question of the relative contribution of anammox and denitrification has small ramifications for the magnitude of the overall fixed N loss. One fundamental difference between the two processes is that denitrification involves  $N_2O$  as an intermediate, whereas anammox does not. This issue is explored in §3a.

#### (a) Nitrous oxide production, consumption and release from the ocean

The marine pathways of  $N_2O$  and the quantification of its oceanic emissions have received increased attention during recent decades [82,83].  $N_2O$  acts as a strong greenhouse gas and in the stratosphere it is the precursor of ozone-depleting nitric oxide (NO) radicals [84,85]. Because of the ongoing decline of chlorofluorocarbons and the continuous increase of  $N_2O$  in the atmosphere, the contributions of  $N_2O$  to both the greenhouse effect and ozone depletion will be even more pronounced in the twenty-first century [84,86]. The oceans, including coastal areas such as continental shelves, estuaries and upwelling areas are a major source of  $N_2O$  and contribute about 30 per cent ( $5.5 \text{ Tg N yr}^{-1}$ ) to the atmospheric  $N_2O$  budget [84]. Oceanic  $N_2O$  is produced as a by-product during bacterial as well as archaeal nitrification; however, nitrifying archaea dominate  $N_2O$  production [87,88].  $N_2O$  occurs also as an intermediate during denitrification. It is produced by the reduction of  $NO_3^-$  and, in turn, can be further reduced to  $N_2$ . Nitrification is the dominating  $N_2O$  production process, whereas denitrification contributes only 7–35% to the overall  $N_2O$  water column budget in the

ocean [89,90]. The importance of other microbial processes, such as dissimilatory nitrate reduction to ammonia and anammox, for oceanic,  $\text{N}_2\text{O}$  production is largely unknown [83].

The amount of  $\text{N}_2\text{O}$  produced during both nitrification and denitrification strongly depends on the prevailing dissolved oxygen ( $\text{O}_2$ ) concentrations and is significantly enhanced under low (i.e. suboxic)  $\text{O}_2$  conditions [2,91].  $\text{N}_2\text{O}$  is usually not detectable in anoxic waters because of its reduction to  $\text{N}_2$  during denitrification. Thus, significantly enhanced  $\text{N}_2\text{O}$  concentrations are generally found at oxic/suboxic boundaries in the oceans [2,83]. The strong  $\text{O}_2$  sensitivity of  $\text{N}_2\text{O}$  production is also observed in coastal systems that are characterized by seasonal shifts in the  $\text{O}_2$  regime [92]. A biological source of  $\text{N}_2\text{O}$  in the well-oxygenated, mixed layer/euphotic zone seems to be unlikely (see discussion in Freing *et al.* [90]). A recent study of both the  $\text{N}_2\text{O}$  air/sea fluxes and  $\text{N}_2\text{O}$  diapycnal fluxes into the mixed layer in the coastal upwelling off Mauritania, northwest Africa indicated that surfactants may have a dampening effect on air–sea exchanges of  $\text{N}_2\text{O}$  [93]. However, for a reassessment of the global coastal  $\text{N}_2\text{O}$  emissions, further laboratory and field studies of the effect of surfactants on the air/sea flux of  $\text{N}_2\text{O}$  are needed.

Global maps of  $\text{N}_2\text{O}$  in the surface ocean [94,95] show supersaturation of  $\text{N}_2\text{O}$  in coastal and equatorial upwelling regions as well as  $\text{N}_2\text{O}$  anomalies close to zero (i.e. near equilibrium) in large parts of the open ocean. Since the studies of Nevison *et al.* [94] and Suntharalingam & Sarmiento [95], the amount of available  $\text{N}_2\text{O}$  data has been steadily increasing. Therefore, the project ‘Marine Methane and Nitrous Oxide’ database has been launched with the aim to collect and archive  $\text{N}_2\text{O}$  datasets and to provide actual fields of surface  $\text{N}_2\text{O}$  for emission estimates [96].

The observed ongoing deoxygenation of the open ocean [97] can be translated into an additional  $\text{N}_2\text{O}$  accumulation of less than 6 per cent [83]. In contrast to the open ocean, Naqvi *et al.* [92] cautioned that global  $\text{N}_2\text{O}$  emissions from shallow hypoxic/anoxic coastal systems might increase significantly in the future. It seems realistic to expect that the  $\text{N}_2\text{O}$  emissions from shallow hypoxic/anoxic coastal systems will increase in the near future due to increasing nutrient inputs (caused by the ongoing industrialization and intensification of agricultural activities), whereas future  $\text{N}_2\text{O}$  emissions from the open ocean owing to decreasing oxygen in mid water depths seem to be of minor significance.

## 4. Shelf processes

The coastal ocean is conveniently partitioned into a proximal, i.e. spatially heterogeneous part, including estuaries, lagoons, etc., and a distal compartment, i.e. the continental shelf. The coastal ocean is the eventual sink of most anthropogenic DIN delivered by rivers, groundwater and atmospheric deposition.

### (a) Proximal coastal systems

Each year, rivers transport about 40–66  $\text{Tg N yr}^{-1}$  to coastal ecosystems, 40 per cent in the form of DIN (mainly nitrate), 40 per cent in the form of particulate nitrogen and the remaining 20 per cent as dissolved organic nitrogen [62]. The variability in the export is impacted by anthropogenic nitrogen input, whereas DON export varies between 10 and 25 per cent of total N [62]. Submarine groundwater discharge delivers about 4  $\text{Tg N yr}^{-1}$  to near-shore ecosystems. Further inputs

of nitrogen to the proximate coastal ocean include nitrogen fixation in the water column, sediments and vegetated ecosystems totalling about 15  $\text{Tg N yr}^{-1}$  and atmospheric deposition (about 1  $\text{Tg N yr}^{-1}$ ). Losses of fixed nitrogen include export to the continental shelf (38  $\text{Tg N yr}^{-1}$ ), emission of  $\text{N}_2\text{O}$  (0.5  $\text{Tg N yr}^{-1}$ ), denitrification (4–8  $\text{Tg N yr}^{-1}$ ), fish landings (3.7  $\text{Tg N yr}^{-1}$ ) and burial (22  $\text{Tg N yr}^{-1}$ ; figure 2). This burial occurs mainly in vegetated systems such as seagrass meadows and mangroves [98], because of trapping of particulate nitrogen and assimilation of dissolved inorganic and organic nitrogen [67]. The present-day and future nitrogen cycling in proximal coastal systems differs substantially from that before the Anthropocene. Eutrophication has resulted in community shifts such as a loss of submerged vegetation and proliferation of macroalgae and phytoplankton. This will eventually lead to lower rates of nitrogen burial. Denitrification may be higher or lower depending on conditions [99], whereas  $\text{N}_2\text{O}$  emission is higher because of extensive oxic–anoxic interfaces with high rates of nitrification and denitrification.

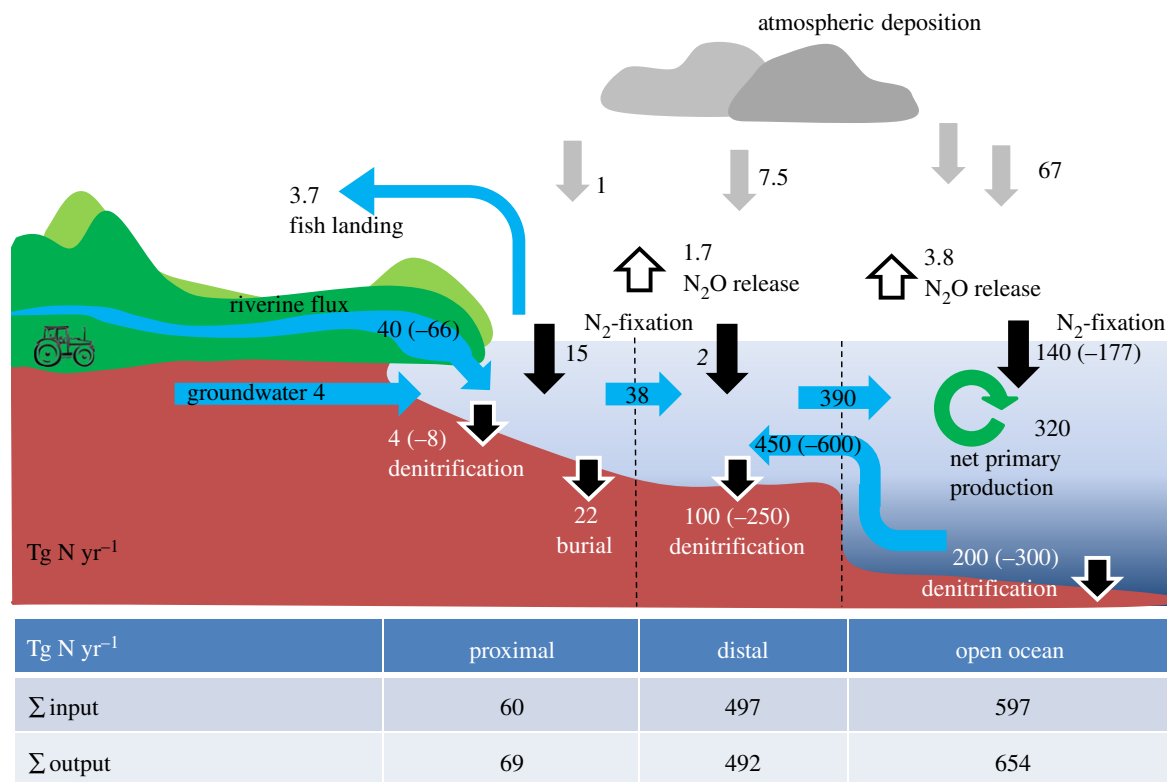
### (b) Distal coastal systems

The fixed nitrogen budget of continental shelf systems is primarily governed by exchanges with the open ocean (figure 2). Globally, continental shelves receive about 38  $\text{Tg N yr}^{-1}$  exported from the proximal coastal zone, 7.5  $\text{Tg N yr}^{-1}$  from atmospheric deposition (almost all anthropogenic) and a small contribution by nitrogen-fixing organisms (about 2  $\text{Tg N yr}^{-1}$ ). Continental shelf systems lose nitrogen via denitrification (100–250  $\text{Tg N yr}^{-1}$ ); the uncertainty, however, is high [100]. Because the global estimate of 400  $\text{Tg N yr}^{-1}$  removal per year exists, about 200–300  $\text{Tg N yr}^{-1}$  remain for the open ocean (figure 2). Recent estimates suggested five times higher denitrification rates in permeable sediments [101], so the number may need an upward revision in the future. Most nitrogen entering the continental shelf comes from nitrate exchanges with the open ocean (450–600  $\text{Tg N yr}^{-1}$ ). The majority of the nitrogen imported from subsurface oceanic waters is returned in the form of DON and particulate organic nitrogen (PON) to open ocean surface waters (about 390  $\text{Tg N yr}^{-1}$ ). This organic nitrogen exported from coastal systems supports the reported heterotrophy of oceanic gyre ecosystems and constitutes a source of fixed nitrogen for micro-organisms. A budget of these numbers for the three systems (figure 2) reveals a relatively low uncertainty in the numbers around 10 per cent that supports the reliability of the estimates.

## 5. Model achievements and global budgets considering nitrogen

Ocean biogeochemical and ecosystem models often have N, as model currency, i.e. nitrogen is modelled as a state variable. The models in use differ in many aspects, in particular regarding complexity in model formulation and spatial coverage. Detailed process models such as nitrogen-fixing models, e.g. Fennel *et al.* [102] and Hood *et al.* [103], and denitrification models, for example [104,105], are designed to investigate single processes (see review by Hood [106]).

Ocean biogeochemical circulation models are often based on the nitrogen–phytoplankton–zooplankton–detritus model of Fasham *et al.* [107], which was the first model to consider DON and bacteria as state variables. This model was coupled



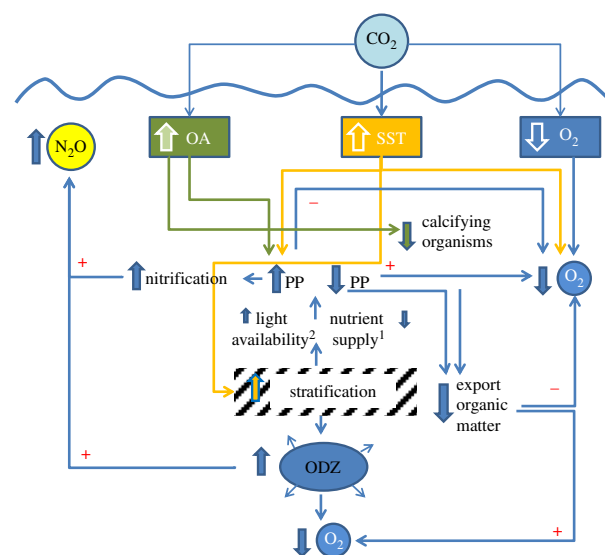
**Figure 2.** Nitrogen budget of the ocean. All numbers are given in the text. The table below considers the smaller numbers, therefore the larger numbers are given in parentheses; net primary production is no input/loss term.

to a circulation model and estimated basin-wide nitrate and chlorophyll-*a* and the relative magnitude of different new N sources [108]. Since then, the complexity of the models has increased by including multiple nutrients, or plankton functional groups. The development of the concept of excess nitrogen  $N^*$  [109] or excess phosphorous  $P^*$  [32] was a milestone because it allows coupling of the nitrogen to phosphorous and carbon cycles [3,32]. The imbalance between input and losses and the findings of higher estimates of both terms would result in higher turnover rates of the nitrogen cycle and the question of how other cycles are still linked.

A major uncertainty in global modelling remains meteorological forcing, especially in the tropics. Large differences exist between monthly mean climatologic datasets and reanalysis data of NCEP/NCAR or ECMWF in wind stress forcing [110], as well as in atmospheric temperature of up to 0.8 K in the late 1980s and early 1990s [111]. A further uncertainty is the lack of consensus concerning the question of whether or not it is necessary to include explicit formulations of microbes as state variables [106].

## 6. Impact of climate change

A recent summary explored climate change effects on the future ocean [112] caused by the increase of  $CO_2$  in the atmosphere [84,113]. There are direct (warming and acidification) and indirect (oxygen depletion) effects. These factors produce further changes that contribute more or less clearly to the many possible changes in ocean biogeochemistry (figure 3). Warming of the sea water, a direct consequence of the increasing air temperature, enhances stratification of the ocean and microbial activity. This may not develop equally everywhere in the ocean. In the Arctic and in low latitudes (from 20° N to



**Figure 3.** Schematic of the major effects of climate change on the ocean biogeochemistry. The graphic does not consider the differences between effects in Arctic and tropical latitudes except for the different effects on primary production (PP); nutrient supply (1) from below the thermocline is restricted due to stronger stratification in low latitudes, but PP is enhanced (2) at high latitudes because of higher light availability. Plus and minus symbols emphasize the positive or negative feedback of a process. OA, ocean acidification; SST, sea surface temperature; PP, primary production; ODZ, oxygen deficiency zone;  $O_2$ , oxygen concentrations,  $N_2O$ , nitrous oxide.

20° S) stronger stratification of the water column is expected than in the rest of the ocean, which would, in turn, affect nutrient supply and light availability. Another important impact is the declining solubility of gases, especially of  $O_2$  itself, potentially increasing the size of ODZ [97,114]. Ocean acidification

may stimulate N<sub>2</sub>-fixing cyanobacteria [115]. In areas where primary production increases, more oxygen is consumed at depth during remineralization and potentially more N<sub>2</sub>O can escape to the atmosphere when the water masses reach the surface (see section §3*a*). However, increasing temperatures may change the balance between production and respiration so that less export of organic matter may occur counteracting oxygen losses in the deep ocean.

Ocean acidification may directly result in significantly reduced nitrification rates accompanied reduced N<sub>2</sub>O production [116]. However, this scenario might be globally counteracted by the generation of more N<sub>2</sub>O when the nitrification process itself shows an overall positive response to lower pH values such as some studies suggest [87,117]. Therefore, it seems reasonable to assume only a minor effect of decreasing

pH on N<sub>2</sub>O production during nitrification, but mesocosm and field experiments to verify the effect of ocean acidification on N<sub>2</sub>O production in the ocean are missing. Recently published model results by Suntharalingam *et al.* [118] show that atmospheric deposition of anthropogenic N to the ocean might lead to an enhanced global oceanic N<sub>2</sub>O production (0.06–0.34 Tg N y<sup>-1</sup>) especially in regions which are sensitive to changes in dissolved oxygen such as coastal zones. At this point, it is difficult to predict which process may dominate, because the interaction of processes is too complex to evaluate the net effect on ocean carbon and nitrogen cycles.

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## References

- Canfield DE, Glazer AN, Falkowski PG. 2010 The evolution and future of earth's nitrogen cycle. *Science* **330**, 192–196. (doi:10.1126/science.1186120)
- Codispoti LA, Brandes JA, Christensen JP, Devol AH, Naqvi SWA, Paerl HW, Yoshinari T. 2001 The oceanic fixed nitrogen and nitrous oxide budgets: moving targets as we enter the anthropocene? *Sci. Marina* **65**, 85–105. (doi:10.3989/scimar.2001.65s285)
- Gruber N, Galloway JN. 2008 An Earth-system perspective of the global nitrogen cycle. *Nature* **451**, 293–296. (doi:10.1038/nature06592)
- van de Graaf AA, Mulder A, Bruijn Pd, Jetten MSM, Robertson LA, Kuenen JG. 1995 Anaerobic oxidation of ammonium is a biologically mediated process. *Appl. Environ. Microbiol.* **61**, 1246–1251.
- Risgaard-Petersen N. 2006 Evidence for complete denitrification in a benthic foraminifer. *Nature* **443**, 94–96. (doi:10.1038/nature05070)
- Capone DG, Burns JA, Montoya JP, Subramaniam A, Mahaffey C, Gundersoen T, Michaels AF, Carpenter EJ. 2005 Nitrogen fixation by *Trichodesmium* spp.: an important source of new nitrogen to the tropical and subtropical North Atlantic Ocean. *Glob. Biogeochem. Cycles* **19**, 1–17. (doi:10.1029/2004GB002331)
- Zehr JP, Carpenter EJ, Villareal TA. 1998 New perspectives on nitrogen-fixing microorganisms in tropical and subtropical oceans. *Trends Microbiol.* **8**, 68–73. (doi:10.1016/S0966-842X(99)01670-4)
- Capone DG. 1988 Benthic nitrogen fixation. In *Nitrogen cycling in coastal marine environments* (eds TH Blackburn, J Sorensen), pp. 85–123. Scientific Committee on Problems of the Environment (SCOPE). London, UK: John Wiley and Sons.
- Dekas AE, Poretsky RS, Orphan VJ. 2009 Deep-sea archaea fix and share nitrogen in methane-consuming microbial consortia. *Science* **326**, 422–426. (doi:10.1126/science.1178223)
- Carpenter EJ, Montoya JP, Burns J, Mulholland MR, Subramaniam A, Capone DG. 1999 Extensive bloom of a N<sub>2</sub>-fixing diatom/cyanobacterial association in the tropical Atlantic Ocean. *Mar. Ecol. Prog. Ser.* **185**, 273–283. (doi:10.3354/meps185273)
- Grosse J, Bombar D, Doan HN, Lam NN, Voss M. 2010 Mekong River influence on nitrogen fixation rates and phytoplankton species composition. *Limnol. Oceanogr.* **55**, 1668–1680. (doi:10.4319/lo.2010.55.4.1668)
- Könneke M, Bernhard AE, Torre JRdl, Walker CB, Waterbury JB, Stahl DA. 2005 Isolation of an autotrophic ammonia-oxidizing marine archaeon. *Nature* **437**, 543–546. (doi:10.1038/nature03911)
- Martens-Habbena W, Berube PM, Urakawa H, Torre JRdl, Stahl DA. 2009 Ammonia oxidation kinetics determine niche separation of nitrifying Archaea and Bacteria. *Nature* **461**, 976–981. (doi:10.1038/nature08465)
- Wuchter C *et al.* 2006 Archaeal nitrification in the ocean. *Proc. Natl Acad. Sci. USA* **103**, 12 317–12 322. (doi:10.1073/pnas.0600756103)
- Duce RA *et al.* 2008 Impacts of atmospheric anthropogenic nitrogen on the open ocean. *Science* **320**, 893–897. (doi:10.1126/science.1150369)
- Vitousek PM, Mooney HA, Lubchenko J, Melillo JM. 1997 Human domination of earth's ecosystems. *Science* **277**, 494–499. (doi:10.1126/science.277.5325.494)
- Diaz RJ, Rosenberg R. 2008 Spreading dead zones and consequences for marine ecosystems. *Science* **321**, 926–929. (doi:10.1126/science.1156401)
- Boyer TP, Antonov JI, Garcia HE, Johnson DR, Locarnini RA, Mishonov AV, Pitcher MT, Baranova OK, Smolyar IV. 2006 *NOAA atlas NESDIS 60. World ocean database 2005* (ed. S Levitus). Washington, DC: U.S. Government Printing Office.
- Broecker WS. 1991 The great ocean conveyor. *Oceanography* **4**, 79–89. (doi:10.5670/oceanog.1991.07)
- Speich S, Blanke B, Vries P.d., Drijfhout S, Döös K, Ganachaud A, Marsh R. 2002 Tasman leakage: a new route in the global ocean conveyor belt. *Geophys. Res. Lett.* **29**, 55-1–55-4. (doi:10.1029/2001GL014586)
- Fong AA, Karl DM, Lukas R, Letelier RM, Zehr JP, Church MJ. 2008 Nitrogen fixation in an anticyclonic eddy in the oligotrophic north pacific ocean. *ISME J.* **2**, 663–676. (doi:10.1038/ismej.2008.22)
- McGillicuddy DJ *et al.* 2007 Eddy/wind interactions stimulate extraordinary mid-ocean plankton blooms. *Science* **316**, 1021–1026. (doi:10.1126/science.1136256)
- Oschlies A, Garçon V. 1998 Eddy-induced enhancement of primary production in a model of the North Atlantic Ocean. *Nature* **394**, 266–269. (doi:10.1038/28373)
- Schafstall J, Dengler M, Brandt P, Bange H. 2010 Tidal-induced mixing and diapycnal nutrient fluxes in the Mauritanian upwelling region. *J. Geophys. Res.* **115**, C10014. (doi:10.1029/2009JC005940)
- Dugdale RE, Goering JJ. 1967 Uptake of new and regenerated forms of nitrogen in primary productivity. *Limnol. Oceanogr.* **12**, 196–206. (doi:10.4319/lo.1967.12.2.0196)
- Azam F, Fenchel T, Field JG, Gray JS, Meyer-Reil LA, Thingstad F. 1983 The ecological role of water-column microbes in the sea. *Mar. Ecol. Prog. Ser.* **10**, 257–263. (doi:10.3354/meps010257)
- Eppley RW, Peterson BJ. 1979 Particulate organic matter flux and planktonic new production in the deep ocean. *Nature* **282**, 677–680. (doi:10.1038/282677a0)
- Karl D *et al.* 2002 Dinitrogen fixation in the world's oceans. *Biogeochemistry* **57**, 47–98. (doi:10.1023/A:1015798105851)
- Bronk DA, Glibert PM, Ward BB. 1994 Nitrogen uptake, dissolved organic nitrogen release, and new production. *Science* **265**, 1843–1846. (doi:10.1126/science.265.5180.1843)
- Yool A, Martin AP, Fernández C, Clark DR. 2007 The significance of nitrification for oceanic new production. *Nature* **447**, 999–1002. (doi:10.1038/nature05885)
- Behrenfeld MJ *et al.* 2006 Climate-driven trends in contemporary ocean productivity. *Nature* **444**, 752–755. (doi:10.1038/nature05317)



32. Deutsch C, Sarmiento JL, Sigman DM, Gruber N, Dunne JP. 2007 Spatial coupling of nitrogen inputs and losses in the ocean. *Nature* **445**, 163–167. (doi:10.1038/nature05392)
33. Codispoti LA. 2007 An oceanic fixed nitrogen sink exceeding 400 TgN a<sup>-1</sup> vs the concept of homeostasis in the fixed-nitrogen inventory. *Biogeosciences* **4**, 233–253. (doi:10.5194/bg-4-233-2007)
34. Galloway JN, Townsend AR, Erismann JW, Bekunda M, Cai Z, Freney JR, Martinelli LA, Seitzinger SP, Sutton MA. 2008 Transformation of the nitrogen cycle: recent trends, questions, and potential solutions. *Science* **320**, 889–892. (doi:10.1126/science.1136674)
35. Dentener F *et al.* 2006 Nitrogen and sulfur deposition on regional and global scales: a multimodel evaluation. *Glob. Biogeochem. Cycles* **20**, GB4003. (doi:10.1029/2005GB002672)
36. Mulholland MR, Capone DG. 2000 The nitrogen physiology of the marine N<sub>2</sub>-fixing cyanobacteria *Trichodesmium* spp. *Trends Plant Sci.* **4**, 148–153. (doi:10.1016/S1360-1385(00)01576-4)
37. Berman-Frank I, Quigg A, Finkel ZV, Irwin AJ, Haramaty L. 2007 Nitrogen-fixation strategies and Fe requirements in cyanobacteria. *Limnol. Oceanogr.* **52**, 2260–2269. (doi:10.4319/lo.2007.52.5.2260)
38. Mills MM, Ridame C, Davey M, Roche JL, Geider RJ. 2004 Iron and phosphorus co-limit nitrogen fixation in the eastern tropical North Atlantic. *Nature* **429**, 292–294. (doi:10.1038/nature02550)
39. Jickells TD *et al.* 2005 Global iron connections between desert dust, ocean biogeochemistry, and climate. *Science* **308**, 67–71. (doi:10.1126/science.1105959)
40. Moore CM *et al.* 2009 Large-scale distribution of Atlantic nitrogen fixation controlled by iron availability. *Nat. Geosci.* **2**, 867–871. (doi:10.1038/NGEO1667)
41. Deutsch C, Gruber N, Key RM, Sarmiento JL. 2001 Denitrification and N<sub>2</sub> fixation in the Pacific Ocean. *Glob. Biogeochem. Cycles* **15**, 483–506. (doi:10.1029/2000GB001291)
42. Montoya JP, Holl CM, Zehr JP, Hansen A, Villareal TA, Capone DG. 2004 High rates of N<sub>2</sub> fixation by unicellular diazotrophs in the oligotrophic Pacific Ocean. *Nature* **430**, 1027–1031. (doi:10.1038/nature02824)
43. Zehr JP, Waterbury JB, Turner PJ, Montoya JP, Omoregie E, Steward GF, Hansen A, Karl DM. 2001 Unicellular cyanobacteria fix N<sub>2</sub> in the subtropical North Pacific Ocean. *Nature* **412**, 635–639. (doi:10.1038/35088063)
44. Zehr JP, Bench SR, Carter BJ, Hewson I, Niazi F, Shi T, Tripp HJ, Affourtit JP. 2008 Globally distributed uncultivated oceanic N<sub>2</sub>-fixing cyanobacteria lack oxygenic photosystem II. *Science* **322**, 1110–1112. (doi:10.1126/science.1165340)
45. Foster RA, Kuypers MM, Vagner T, Paerl RW, Musat N, Zehr JP. 2011 Nitrogen fixation and transfer in open ocean diatom–cyanobacterial symbioses. *ISME J.* **5**, 1484–1493. (doi:10.1038/ismej.2011.26)
46. Bombar D, Moisaner PH, Dippner JW, Foster RA, Voss M, Karfeld B, Zehr JP. 2011 Distribution of diazotrophic microorganisms and nifH gene expression in the Mekong River plume during intermonsoon. *Mar. Ecol. Prog. Ser.* **424**, 39–52. (doi:10.3354/meps08976)
47. Mehta MP, Butterfield DA, Baross JA. 2003 Phylogenetic diversity of nitrogenase (nifH) genes in deep-sea and hydrothermal vent environments of the Juan de Fuca Ridge. *Appl. Environ. Microbiol.* **69**, 960–970. (doi:10.1128/AEM.69.2.960-970.2003)
48. Mehta MP, Huber JA, Baross JA. 2005 Incidence of novel and potentially archaeal nitrogenase genes in the deep Northeast Pacific Ocean. *Environ. Microbiol.* **7**, 1525–1534. (doi:10.1111/j.1462-2920.2005.00836.x)
49. Dang H, Luan X, Zhao J, Li J. 2009 Diverse and novel nifH and nifH-like gene sequences in the deep-sea methane seep sediments of the Okhotsk Sea. *Appl. Environ. Microbiol.* **75**, 2238–2245. (doi:10.1128/AEM.02556-08)
50. Orphan VJ, Turk KA, Green AM, House CH. 2009 Patterns of 15N assimilation and growth of methanotrophic ANME-2 archaea and sulfate-reducing bacteria within structured syntrophic consortia revealed by FISH-SIMS. *Environ. Microbiol.* **11**, 1777–1791. (doi:10.1111/j.1462-2920.2009.01903.x)
51. Pernthaler A, Dekas AE, Brown CT, Goffredi SK, Embaye T, Orphan VJ. 2008 Diverse syntrophic partnerships from deep-sea methane vents revealed by direct cell capture and metagenomics. *Proc. Natl Acad. Sci. USA* **105**, 7052–7057. (doi:10.1073/pnas.0711303105)
52. Miyazaki J, Higa R, Toki T, Ashi J, Tsunogai U, Nunoura T, Imachi H, Takai K. 2009 Molecular characterization of potential nitrogen fixation by anaerobic methane-oxidizing archaea in the methane seep sediments at the number 8 Kumano Knoll in the Kumano Basin, offshore of Japan. *Appl. Environ. Microbiol.* **75**, 7153–7162. (doi:10.1128/aem.01184-09)
53. Groszkopf T *et al.* 2012 Doubling of marine dinitrogen-fixation rates based on direct measurements. *Nature* **488**, 361–364. (doi:10.1038/nature11338)
54. Monteiro FM, Dutkiewicz S, Follows MJ. 2011 Biogeographical controls on the marine nitrogen fixers. *Glob. Biogeochem. Cycles* **25**, GB2003. (doi:10.1029/2010GB003902)
55. Moore JK, Doney SC, Lindsay K. 2004 Upper ocean ecosystem dynamics and iron cycling in a global three-dimensional model. *Glob. Biogeochem. Cycles* **18**, GB4028. (doi:10.1029/2004GB002220)
56. Bronk DA. 2002 Dynamics of DON. In *Biogeochemistry of marine dissolved organic matter* (eds DA Hansell, CA Carlson), pp. 153–200. Amsterdam, The Netherlands: Academic Press.
57. Kirchmann D. 2000 Uptake and regeneration of inorganic nutrients by marine heterotrophic bacteria. In *Microbial ecology of the oceans* (ed. DL Kirchmann), pp. 261–288, Wiley Series in Ecological and Applied Microbiology. John Wiley & Sons, Inc.
58. Bronk DA, Ward BB. 1999 Gross and net nitrogen uptake and DON release in the euphotic zone of Monterey Bay, California. *Limnol. Oceanogr.* **44**, 573–585. (doi:10.4319/lo.1999.44.3.0573)
59. Ogawa H, Amagai Y, Koike I, Kaiser K, Benner R. 2001 Production of refractory dissolved organic matter by bacteria. *Science* **292**, 917–920. (doi:10.1126/science.1057627)
60. Nagata T, Kirchman DL. 1992 Release of dissolved organic matter by heterotrophic protozoa: implications for microbial food webs. *Arch. Hydrobiol. Beih. Ergebn. Limnol.* **35**, 99–109.
61. Fuhrman JA, Noble RT. 1995 Viruses and protists cause similar bacterial mortality in coastal seawater. *Limnol. Oceanogr.* **40**, 1236–1242. (doi:10.4319/lo.1995.40.7.1236)
62. Seitzinger SP, Harrison JA, Dumont E, Beusen AHW, Bouwman AF. 2005 Sources and delivery of carbon, nitrogen, and phosphorus to the coastal zone: an overview of global nutrient export from watersheds (NEWS) models and their application. *Glob. Biogeochem. Cycles* **19**, 1–11. (doi:10.1029/2005GB002606)
63. Cornell S, Rendell A, Jickells T. 1995 Atmospheric inputs of dissolved organic nitrogen to the oceans. *Nature* **376**, 243–246. (doi:10.1038/376243a0)
64. Jickells T, Baker AR, Cape JN, Cornell SE, Nemitz E. 2013 The cycling of organic nitrogen through the atmosphere. *Phil. Trans. R. Soc. B* **368**, 20130115. (doi:10.1098/rstb.2013.0115)
65. Mulholland MR, Lomas MW. 2008 Nitrogen uptake and assimilation. In *Nitrogen in the marine environment*, 2nd edn. (eds D Capone, D Bronk, M Mulholland, E Carpenter), pp. 303–384. New York, NY: Academic Press.
66. Korth F, Deutsch B, Liskow I, Voss M. 2011 Uptake of dissolved organic nitrogen by size-fractionated plankton along a salinity gradient from the North Sea to the Baltic Sea. *Biogeochemistry* **111**, 347–360. (doi:10.1007/s10533-011-9656-1)
67. Van Engeland T, Bouma TJ, Morris EP, Brun FG, Peralta G, Lara M, Hendriks IE, Soetaert K, Middelburg JJ. 2011 Potential uptake of dissolved organic matter by seagrasses and macroalgae. *Mar. Ecol. Prog. Ser.* **427**, 71–81. (doi:10.3354/meps09054)
68. Redfield AC. 1934 On the proportions of organic derivatives in seawater and their relation to the composition of plankton. *James Johnstone Mem. Vol. University of Liverpool*, 176–192.
69. Klausmeier CA, Litchman E, Daufresne T, Levin SA. 2004 Optimal nitrogen-to-phosphorus stoichiometry of phytoplankton. *Nature* **429**, 171–174. (doi:10.1038/nature02454)
70. Taylor PG, Townsend AR. 2010 Stoichiometric control of organic carbon–nitrate relationships from soils to the sea. *Nature* **464**, 1178–1181. (doi:10.1038/nature08985)

71. Lunau M, Voss M, Erickson M, Dziallas C, Casciotti K, Ducklow H. 2012 Excess nitrate loads to coastal waters reduces nitrate removal efficiency: mechanism and implications for coastal eutrophication. *Environ. Microbiol.* (doi:10.1111/j.1462-2920.2012.02773.x)
72. Falkowski PG. 1997 Evolution of the nitrogen cycle and its influence on the biological sequestration of CO<sub>2</sub> in the ocean. *Nature* **387**, 272–275. (doi:10.1038/387272a0)
73. Tyrrell T. 1999 The relative influences of nitrogen and phosphorus on oceanic primary production. *Nature* **400**, 525–531. (doi:10.1038/22941)
74. Falkowski PG, Katz ME, Knoll AH, Quigg A, Raven JA, Schofield O, Taylor FJR. 2004 The evolution of modern eukaryotic phytoplankton. *Science* **305**, 354–360. (doi:10.1126/science.1095964)
75. Hamersley MR *et al.* 2007 Anaerobic ammonium oxidation in the Peruvian oxygen minimum zone. *Limnol. Oceanogr.* **52**, 923–933. (doi:10.4319/lo.2007.52.3.0923)
76. Thamdrup B, Dalsgaard T, Jensen MM, Ulloa O, Fariás L, Escribano R. 2006 Anaerobic ammonium oxidation in the oxygen-deficient waters off northern Chile. *Limnol. Oceanogr.* **51**, 2145–2156. (doi:10.4319/lo.2006.51.5.2145)
77. Ward BB, Devol AH, Rich JJ, Chang BX, Bulow SE, Naik H, Pratihary A, Jayakumar A. 2009 Denitrification as the dominant nitrogen loss process in the Arabian Sea. *Nature* **461**, 78–82. (doi:10.1038/nature08276)
78. Bulow SE, Rich JJ, Naik HS, Pratihary AK, Ward BB. 2010 Denitrification exceeds anammox as a nitrogen loss pathway in the Arabian Sea oxygen minimum zone. *Deep Sea Res. I: Oceanogr. Res. Papers* **57**, 384–393. (doi:10.1016/j.dsr.2009.10.014)
79. Jensen MM, Lam P, Revsbech NP, Nagel B, Gaye B, Jetten MS, Kuypers MM. 2011 Intensive nitrogen loss over the Omani Shelf due to anammox coupled with dissimilatory nitrite reduction to ammonium. *ISME J.* **5**, 1660–1670. (doi:10.1038/ismej.2011.44)
80. Koeve W, Kähler P. 2010 Heterotrophic denitrification vs. autotrophic anammox: quantifying collateral effects on the oceanic carbon cycle. *Biogeosciences* **7**, 2327–2337. (doi:10.5194/bg-7-2327-2010)
81. Chang BX, Devol AH, Emerson SR. 2010 Denitrification and the nitrogen gas excess in the eastern tropical South Pacific oxygen deficient zone. *Deep Sea Res. I: Oceanogr. Res. Papers* **57**, 1092–1101. (doi:10.1016/j.dsr.2010.05.009)
82. Bange H. 2008 Gaseous nitrogen compounds (NO, N<sub>2</sub>O, N<sub>2</sub>, NH<sub>3</sub>) in the ocean. In *Nitrogen in the marine environment* (eds DG Capone, DA Bronk, MR Mulholland, EJ Carpenter), pp. 51–94. New York, NY: Elsevier.
83. Bange H, Freing A, Kock A, Löscher C. 2010 Marine pathways to nitrous oxide. In *Nitrous oxide and climate change* (ed. K Smith), pp. 36–62. London, UK: Earthscan.
84. IPCC. 2007 Climate change 2007: the physical science basis. In *Contribution of working group I to the fourth assessment report of the Intergovernmental Panel on Climate Change* (eds S Solomon, D Qin, M Manning, Z Chen, M Marquis, KB Averyt, M Tignor, HL Miller), pp. 996. Cambridge, UK: Cambridge University Press.
85. WMO. 2010 Scientific assessment of ozone depletion: 2010 Global Ozone Research and Monitoring Project-Report No. 52. Geneva, Switzerland: WMO.
86. Ravishankara A, Daniel JS, Portmann RW. 2009 Nitrous oxide (N<sub>2</sub>O): the dominant ozone-depleting substance emitted in the 21st century. *Science* **326**, 123–125. (doi:10.1126/science.1176985)
87. Löscher CR, Kock A, Koenneke M, LaRoche J, Bange HW, Schmitz RA. 2012 Production of oceanic nitrous oxide by ammonia-oxidizing archaea. *Biogeosciences* **9**, 2095–2122. (doi:10.5194/bg-9-2419-2012)
88. Alyson ES, Carolyn B, Matthew RM, Casciotti KL. 2011 Isotopic signature of N<sub>2</sub>O produced by marine ammonia-oxidizing archaea. *Science* **333**, 1282–1285. (doi:10.1126/science.1208239)
89. Bange H, Andreae MO. 1999 Nitrous oxide in the deep waters of the world's oceans. *Glob. Biogeochem. Cycles* **13**, 1127–1135. (doi:10.1029/1999GB900082)
90. Freing A, Wallace DWR, Bange HW. 2012 Global oceanic production of nitrous oxide. *Phil. Trans. R. Soc. B* **367**, 1245–1255. (doi:10.1098/rstb.2011.0360)
91. Codispoti L, Elkins J, Yoshinari T, Friederich G, Sakamoto C, Packard T. 1992 On the nitrous oxide flux from productive regions that contain low oxygen waters. In *Oceanography of the Indian Ocean* (ed. BN Desai), pp. 271–284. Rotterdam, The Netherlands: Balkema.
92. Naqvi SWA, Bange HW, Fariás L, Monteiro PMS, Scranton MI, Zhang J. 2010 Marine hypoxia/anoxia as a source of CH<sub>4</sub> and N<sub>2</sub>O. *Biogeosciences* **7**, 2159–2190. (doi:10.5194/bg-7-2159-2010)
93. Kock A, Schafstall J, Dengler M, Brandt P, Bange HW. 2012 Sea-to-air and diapycnal nitrous oxide fluxes in the eastern tropical North Atlantic Ocean. *Biogeosciences* **9**, 957–964. (doi:10.5194/bg-9-957-2012)
94. Nevison CD, Weiss RF, Erickson III DJ. 1995 Global oceanic emissions of nitrous oxide. *J. Geophys. Res.* **100**, 15 809–15 820. (doi:10.1029/95JC00684)
95. Suntharalingam P, Sarmiento JL. 2000 Factors governing the oceanic nitrous oxide distribution: simulations with an ocean general circulation model. *Glob. Biogeochem. Cycles* **14**, 429–454. (doi:10.1029/1999GB900032)
96. Bange HW, Bell TG, Cornejo M, Freing A, Uher G, Upstill-Goddard RC, Zhang G. 2009 MEMENTO: a proposal to develop a database of marine nitrous oxide and methane measurements. *Environ. Chem.* **6**, 195–197. (doi:10.1071/EN09033)
97. Stramma L, Johnson GC, Sprintall J, Mohrholz V. 2008 Expanding oxygen-minimum zones in the tropical oceans. *Science* **320**, 655–658. (doi:10.1126/science.1153847)
98. Bouillon S *et al.* 2008 Mangrove production and carbon sinks: a revision of global budget estimates. *Glob. Biogeochem. Cycles* **22**, GB2013. (doi:10.1029/2007GB003052)
99. Middelburg JJ, Levin LA. 2009 Coastal hypoxia and sediment biogeochemistry. *Biogeosciences* **6**, 1273–1293. (doi:10.5194/bg-6-1273-2009)
100. Voss M *et al.* 2011 Nitrogen processes in coastal and marine ecosystems. In *The European nitrogen assessment* (eds MA Sutton, CM Howard, JW Erisman, G Billen, A Bleeker, P Grennfelt, Hv Grinsven, B Grizzetti), pp. 147–176. Cambridge, UK: Cambridge University Press.
101. Gao H *et al.* 2012 Intensive and extensive nitrogen loss from intertidal permeable sediments of the Wadden Sea. *Limnol. Oceanogr.* **57**, 185–198. (doi:10.4319/lo.2012.57.1.0185)
102. Fennel K, Spitz YH, Letelier RM, Abbott MR, Karl DM. 2002 A deterministic model for N<sub>2</sub> fixation at stn. ALOHA in the subtropical North Pacific Ocean. *Deep Sea Res.* **49**, 1–3.
103. Hood RR, Coles VJ, Capone DG. 2004 Modeling the distribution of *Trichodesmium* and nitrogen fixation in the Atlantic Ocean. *J. Geophys. Res.* **109**, C06006. (doi:10.1029/2002JC001753)
104. Oguz T, Ducklow HW, Malanotte-Rizzoli P. 2000 Modeling distinct vertical biogeochemical structure of the Black Sea: dynamical coupling of the oxic, suboxic, and anoxic layers. *Glob. Biogeochem. Cycles* **14**, 1331–1352. (doi:10.1029/1999GB001253)
105. Voss M, Dippner JW, Montoya JP. 2001 Nitrogen isotope patterns in the oxygen deficient waters of the Eastern Tropical Pacific Ocean (ETNP). *Deep Sea Res. I* **48**, 1905–1921. (doi:10.1016/S0967-0637(00)00110-2)
106. Hood RR, Christian JR. 2008 Ocean nitrogen cycle modeling. In *Nitrogen in the marine environment* (eds DG Capone, DA Bronk, MR Mulholland, EJ Carpenter), pp. 1445–1495. Amsterdam, The Netherlands: Elsevier.
107. Fasham MJR, Ducklow HW, McKelvie SM. 1990 A nitrogen-based model of plankton dynamics in the oceanic mixed layer. *J. Mar. Res.* **48**, 591–639.
108. Sarmiento JL, Slater RD, Fasham MJR, Ducklow HW, Toggweiler JR, Evans GT. 1993 A seasonal 3-dimensional ecosystem model of nitrogen cycling in the North Atlantic euphotic zone. *Glob. Biogeochem. Cycles* **7**, 417–450. (doi:10.1029/93GB00375)
109. Gruber N, Sarmiento J. 1997 Global patterns of marine nitrogen fixation and denitrification. *Glob. Biogeochem. Cycles* **11**, 235–266. (doi:10.1029/97GB00077)
110. Josey SA, Kent EC, Taylor PK. 2002 Wind stress forcing of the ocean in the SOC climatology: comparisons with the NCEP-NCAR, ECMWF, UWM/COADS, and Hellerman and Rosenstein datasets. *J. Phys. Oceanogr.* **32**, 1993–2019. (doi:10.1175/1520-0485(2002)032<1993:WSFOTO>2.0.CO;2)
111. Trenberth KE, Stepaniak DP, Hurrell JW, Fiorino M. 2001 Quality of reanalyses in the tropics. *J. Clim.* **14**, 1499–1510. (doi:10.1175/1520-0442(2001)014<1499:QORITT>2.0.CO;2)

112. Gruber N. 2011 Warming up, turning sour, losing breath: ocean biogeochemistry under global change. *Phil. Trans. R. Soc. A* **369**, 1980–1996. (doi:10.1098/rsta.2011.0003)
113. Feely RA, Doney SC, Cooley SR. 2009 Ocean acidification: present conditions and future changes in a high-CO<sub>2</sub> world. *Oceanography* **22**, 36–47. (doi:10.5670/oceanog.2009.95)
114. Keeling RF, Körtzinger A, Gruber N. 2010 Ocean deoxygenation in a warming world. *Annu. Rev. Mar. Sci.* **2**, 199–229. (doi:10.1146/annurev.marine.010908.163855)
115. Hutchins DA, Fu F-X, Zhang Y, Warner ME, Feng Y, Portune K, Bernhardt PW, Mulholland MR. 2007 CO<sub>2</sub> control of *Trichodesmium* N<sub>2</sub> fixation, photosynthesis, growth rates, and elemental ratios: implications for past, present, and future ocean biogeochemistry. *Limnol. Oceanogr.* **52**, 1293–1304. (doi:10.4319/lo.2007.52.4.1293)
116. Beman JM, Chow C-E, King AL, Feng Y, Fuhrman JA, Andersson A, Bates NR, Popp BN, Hutchins DA. 2011 Global declines in oceanic nitrification rates as consequence of ocean acidification. *Proc. Natl Acad. Sci. USA* **108**, 208–213. (doi:10.1073/pnas.1011053108)
117. de Bie MJ, Middelburg JJ, Starink M, Laanbroek HJ. 2002 Factors controlling nitrous oxide at the microbial community and estuarine scale. *Mar. Ecol. Prog. Ser.* **240**, 1–9. (doi:10.3354/meps240001)
118. Suntharalingam P, Buitenhuis E, Le Quéré C, Dentener F, Nevison C, Butler JH, Bange HW, Forster G. 2012 Quantifying the impact of anthropogenic nitrogen deposition on oceanic nitrous oxide. *Geophys. Res. Lett.* **39**, L07605. (doi:10.1029/2011GL050778)