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Reversal of density dependence of juvenile *Littorina littorea* (Gastropoda) growth in response to periphyton nutrient status

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Abstract

Experimental periphyton communities were grown in aquaria receiving media of differently enriched seawater (fully enriched, without Si enrichment, without N and P enrichment) and supplied differently with medium (batch and weekly replacement). Periphyton was subject to grazing by 1–6 individuals of juvenile *Littorina littorea*. Periphyton biomass was higher in the replacement aquaria than in the batch aquaria and higher in the full and the –Si medium than in the –NP medium. The N:C ratio of the periphyton increased with *Littorina* number in the batch aquaria and was unaffected by *Littorina* number in the replacement aquaria. Diatoms were most dominant in the –NP treatments and rarest in the –Si treatments. Chlorophytes were dominant in the –Si and the fully enriched treatments, but also Cyanobacteria contributed significantly to periphyton biomass in those treatments under nutrient replacement. Somatic growth of *Littorina* was negatively correlated to *Littorina* density in the replacement aquaria and positively density dependent in the batch aquaria. The latter is explained by improved food quality under stronger grazing pressure. © 2001 Elsevier Science B.V. All rights reserved.

KeyWords: Herbivory; *Littorina*; Periphyton; Food quality; Benthos

1. Introduction

During the last decade, it has become increasingly clear that the growth and reproduction of herbivorous animals might not only be limited by the total amount or energy content of their food but also by its chemical composition (Muller-Navarra and Lampert, 1996). Essential organic constituents, particularly highly unsaturated fatty acids (Brett and Muller-Navarra, 1997; Muller-Navarra et al., 2000) and mineral nutrients, particularly nitrogen and phosphorous (Hessen, 1992; Urabe and Watanabe, 1992; Sterner, 1993) have received attention. In some cases, correlations between the content of mineral nutrients and essential organic constituents make the distinction between the

two types of 'food quality limitation' difficult (Muller-Navarra, 1995; Sterner and Schulz, 1998). In the case of unicellular algae, mineral nutrient content of biomass is closely related to the extent of algal nutrient limitation (Droop, 1973, 1985). Microalgal nutrient status can be co-determined by grazing because of animal nutrient excretion and because less algae can share the same supply of mineral nutrients if grazing reduces biomass (Sterner, 1989, 1990). The resulting negative correlation between grazing and nutrient limitation could lead to an improved food quality at higher grazing pressure. If food quality limitation limits herbivore growth, the traditional view of a negative relationship between population densities and growth rates would be turned upside down. More grazers would reduce the amount of food but improve its nutritional quality. This

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reversal of density dependence has been shown by continuous culture experiments with the freshwater zooplankton *ap nia ealeata* and the P-limited planktonic green alga *cenedes'1'ts acttts* (Sommer, 1992) as food organism. A reversal of density-dependent regulation would require substantial changes in traditional models of population interactions, which frequently rest on the assumption of negative density dependence.

Here, I present laboratory experiments with juveniles of the common periwinkle (*Littorina littorea*) and Baltic Sea periphyton which show a reversal of density dependence induced by nutrient limitation of periphyton. *Littorina littorea* is an important herbivore on Northern European and American coasts, feeding on a wide array of benthic food items. If given a choice, they prefer periphytic microalgae, filamentous macroalgae or microscopic germlings of macroalgae over leathery macroalgae (Lubchenco, 1978; Steneck and Watling, 1982; Worm et al., 1999; Lotze et al., 2000). There is no pronounced selectivity in periphyton feeding, except for some relative protection of algae attached tightly to the substratum and gelatinous algae (Sommer, 1999a,b).

2. Material and methods

The experiments were performed in 36 aquaria with a bottom area of 1.5 dm² and a volume of 2.3 dm³. The bottom of each aquarium was covered by 28 sandstone tiles (of 5.29 cm² each), which could be removed individually for subsampling. The aquaria were filled with 1.5 dm³ growth medium. Periphyton for the experiments was obtained by incubating rough, transparent plastic sheets at ca. 1 m depth in Kiel Fjord (western Baltic Sea) from 3 May to 11 June 1999. After incubation, periphyton was scratched from the plastic sheets, suspended in 250 cm³ sterile seawater, mixed well and distributed equally to the 36 aquaria. The algae were permitted to settle and start growth on the tiles until 22 June. Then juvenile *Littorina* individuals (1.8–2.3 mm shell height) were placed in the aquaria. Those snails had been kept under abundant food supply in an aquarium for two months before the start of the experiment. Every four weeks, one randomly selected tile was removed for

sampling and replaced by a fresh one. The experiment was terminated after 16 weeks. The day-light cycle was 14:10 hrs; photon flux density was ca. 40 J.E cm⁻² s⁻¹; the experimental temperature was 15°C.

The experimental treatments differed in the number of snails (6 levels from 1 to 6 individuals per aquarium), the composition of the medium (3 levels; full, -Si, -NP) and the supply mode of the medium (replacement of 2/3 each week; no replacement). There was a fully factorial combination of all treatments. The full medium consisted of filtered (0.2 J.m) Baltic Sea water enriched with 20 J.M Si, 15 J.M N (supplied as NH₄NO₃), and 2 J.M P. The -Si medium contained no Si enrichment and the -NP medium contained no enrichment with N and P. The background concentrations in the filtered seawater were variable, but always <10% of the enrichments. The two supply modes were intended to contrast scenarios where nutrient supply to the algae was dominated by external sources (replacement) and by excretion by snails (no replacement = batch).

Periphyton was scratched from the sample tiles and suspended in filtered seawater in order to be split into subsamples for the analysis of particulate C, N, P, and cell counts. Chemical analysis was performed on periphyton filtered onto precombusted glass fiber filters. Particulate C and N were analysed in a CN analyser (Fisons NA 1500 N), particulate P was measured according to oceanographic standard methods (Grasshoff, 1976) in an autoanalyser after muffling the filters at 545°C and dissolving the ash in H₂SO₄. Subsamples for cell counts were treated like plankton samples and counted under the inverted microscope (Utermohl, 1958). At least 400 individuals were counted, which gives 95% confidence limit of ±10% for the estimated cell number (Lund et al., 1958). Taxonomic identification was performed according to Pankow (1990). Cell volumes were calculated according to the geometric formulae suggested by Hillebrand et al. (1999) after measurements of 20 individuals per taxon.

Littorina growth was measured as per capita gain of soft-body organic substance (ash-free dry mass = AFDM) during the experiment (16 weeks). AFDM for the end point of the experiments

Table 1

ANOVAs of response variables with medium composition (M) and supply mode (S) as categorical factors and *Littorina* number as quantitative factor. Response variables: periphyton biomass [ln (J.g C cm⁻²)], periphyton N:C ratio [ln (mol N/mol C)], relative biomass of higher taxa [arcsine(B_i/B_{tot})^{-1/2}], *Littorina* growth [mg AFDM/ind];significance levels: * p < 0.05, ** p < 0.01, *** p < 0.001, ****p < 0.0001

Dependent variable	Significant main effects (F-ratio, significance)	Significant interactions (F-ratio, significance)
Biomass	M (12.75****) S (84.4****) L (238.5****)	M X S (6.5**)
N:C	M (35.1****) S (159.7****) L (22.2****)	M X S (4.6*) S X L (25.6****)
Diatoms	M (1226****) S (9.7**)	M X S (9.6**)
Green algae	M (476****) S (64.0****)	M X S (19.3****)
Cyanobacteria	M (6.1**) S (6.3*) L (5.6*)	M X S (7.1**)
<i>Littorina</i> growth	M (9.3**) S (66.4****)	M X S (5.9**) S X L (17.9****)

was measured directly while start values were estimated from a shell height (mm)-mass (mg) regression:

$$AFDM = 0.05525H^{2.96};$$

$$r^2 = 0.72; \text{ d.f.} = 35; \text{ p} < 0.00001$$

This regression had been obtained with *Littorina* individuals from the same source as the experiments.

The experimental results were first analysed by a multifactor ANOVA (general linear models, Stagraphics) with three independent variables: medium composition and supply mode as categorical factors and *Littorina* number as quantitative factor (Table 1). The number of cases entered in each ANOVA was 36. Mean values over the entire duration of the experiment were chosen for the periphyton parameters, because the calculation of *Littorina* growth refers to the same period. The response of dependent variables (periphyton C, periphyton N:C ratio, *Littorina* growth) to *Littorina* number was then analysed by individual

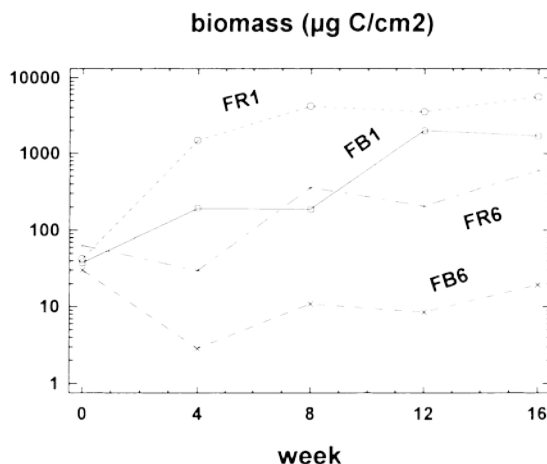


Fig. 1. Time course of periphyton biomass (POC) in the full medium;FB1: full medium, batch, 1 *Littorina*;FB6: full medium, batch, 6 *Littorina*;FR1: full medium, replacement, 1 *Littorina*;FR6: full medium, replacement, 6 *Littorina*.

regression analysis for each combination of medium composition and supply mode.

3. Results

3 Perip Yton nio'1'ass and stoic io'1'etrY

Already after four weeks a clear response of periphyton biomass measured as POC to the different treatments could be observed. Those differences were maintained throughout the entire duration of the experiment (Figs 1-3). In the ANOVA, biomass showed a significant response to medium composition, supply mode and *Littorina* densities (Table 1). Interaction effects between medium composition and supply mode were also significant. Biomass levels were lower in the batch than in the replacement treatments and lower in the -NP treatments than in the other two nutrient treatments which formed a homogeneous group according to Scheffe's multiple range test. Log biomass declined linearly with *Littorina* density (Table 2).

The relationship between particulate P and particulate N was examined by a regression analysis according to the model

$$N = aP^n$$

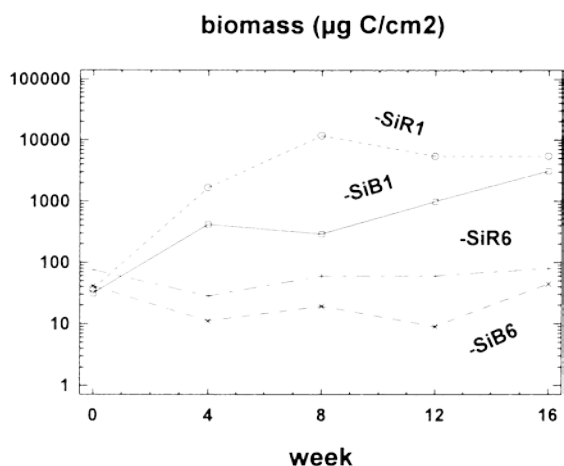


Fig. 2. Time course of periphyton biomass (POC) in the medium without Si-enrichment; -SiB1: without Si, batch, 1 *Littorina*; -SiB6: without Si, batch, 6 *Littorina*; -SiR1: without Si, replacement, 1 *Littorina*; -SiR6: without Si, replacement, 6 *Littorina*.

which showed an almost linear relationship (Table 3). At exponents close to 1, parameter a is an estimate of the N:P ratio in algal biomass. This was clearly below the Redfield-ratio (16:1), indicating that N rather than P was the potentially limiting nutrient (Hillebrand and Sommer, 1999). Therefore, and because of the close and linear relationship between N and P, only the N:C ratios will be analysed as indicators of algal nutrient status. They were significantly influenced by all experimental factors (Table 4, Fig. 4). Biomass N content was higher under medium replacement than under batch conditions and lower in the -NP medium than in the other two media (Scheffe's multiple range test). In the batch experiments N:C increased with *Littorina* density but it was not significantly influenced

Table 2

Linear regressions of periphyton biomass ($\ln \text{J.g C cm}^{-2}$) on *Littorina* number per vessel, for treatment codes see text and legend of Fig. 4

Treatment	$a \pm \text{SE}$	$n \pm \text{SE}$	r^2	p
FR	8.29 ± 0.59	-0.53 ± 0.15	0.75	0.0260
-SiR	9.04 ± 0.40	-0.82 ± 0.10	0.94	0.0013
-NPR	7.70 ± 0.40	-0.82 ± 0.10	0.94	0.0013
FB	7.06 ± 0.58	-0.83 ± 0.15	0.88	0.0052
-SiB	7.12 ± 0.51	-0.73 ± 0.13	0.89	0.0051
-NPB	7.16 ± 0.23	-0.89 ± 0.06	0.98	0.0001

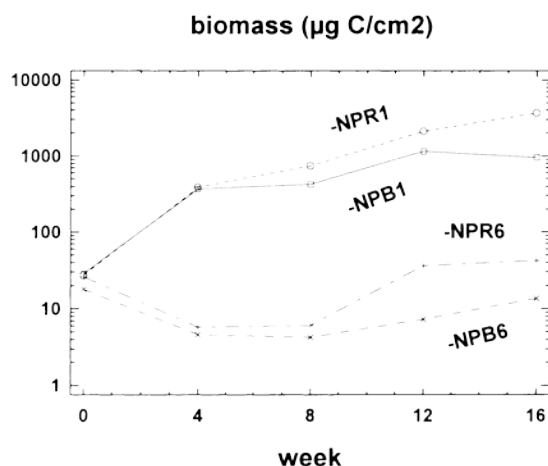


Fig. 3. Time course of periphyton biomass (POC) in the medium without N- and P enrichment; -NPB1: without N and P, batch, 1 *Littorina*; -NPB6: without N and P, batch, 6 *Littorina*; -NPR1: without N and P, replacement, 1 *Littorina*; -NPR6: without N and P, replacement, 6 *Littorina*.

by *Littorina* density under nutrient replacement (Table 4).

3 Periphyton taxonomic composition

Throughout the experiment, the bulk of algal biomass was composed of three higher taxa, Bacillariophyceae (diatoms), Chlorophyta (green algae) and Cyanobacteria (blue-green algae), while other groups were negligible. Medium composition exerted the greatest influence, while supply mode had a significant influence only in some cases (Table 1). *Littorina* density was not an important factor in determining higher level taxonomic composition. Diatoms were dominant in the -NP treatments, contributed less to total biomass in the F treatments, and still less in the -Si treatments (Fig. 5). Green algae (mainly *Enteromorpha intestinalis*, *Cladophora pycnoides*,

Table 3

Regression of particulate N on particulate P (both molar) according to the model $N = aP^n$

Time (weeks)	$a \pm \text{SE}$	$n \pm \text{SE}$	r^2	p
4	8.33 ± 1.95	0.95 ± 0.05	0.90	< 0.00001
8	9.49 ± 2.10	1.00 ± 0.05	0.92	< 0.00001
12	7.97 ± 1.52	0.94 ± 0.05	0.91	< 0.00001
16	7.88 ± 1.06	0.92 ± 0.04	0.94	< 0.00001

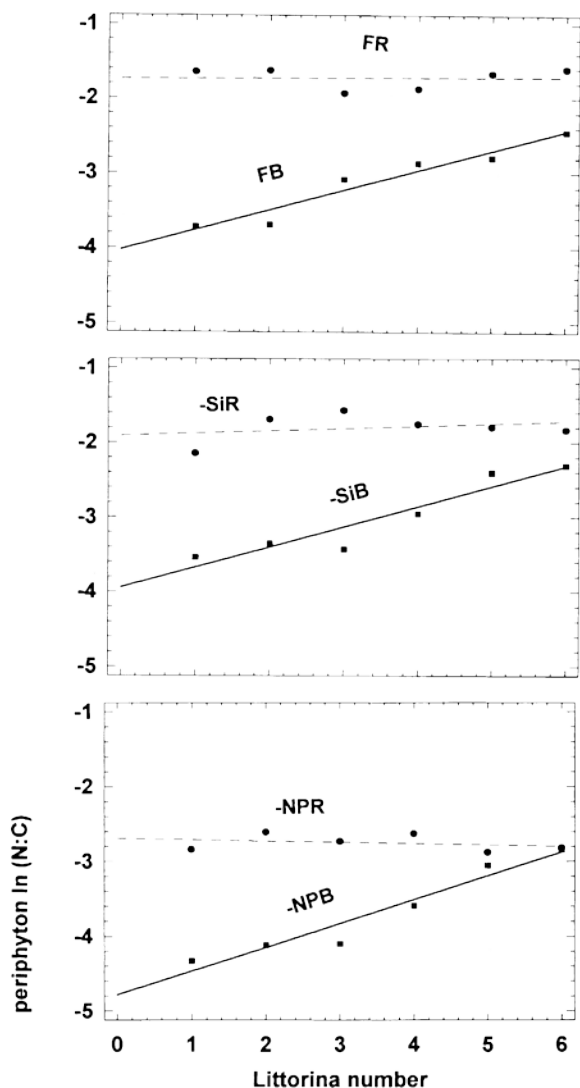


Fig. 4. Periphyton N:C ratio (log transformed) in response to *Littorina* density. F: full enrichment; -Si: enrichment without Si; -NP: enrichment without N and P; B: batch; R: replacement.

Table 4

Linear regressions of \log^e periphyton N:C ratio on *Littorina* number per vessel

Treatment	a ± SE	n ± SE	r ²	p
FR	-1.74 ± 0.16	-0.005 ± 0.038	0.005	0.896
-SiR	-1.92 ± 0.19	0.03 ± 0.5	0.09	0.564
-NPR	-2.60 ± 0.11	0.02 ± 0.03	0.09	0.559
FB	-4.03 ± 0.13	0.27 ± 0.03	0.94	0.0014
-SiB	-3.95 ± 0.18	0.27 ± 0.05	0.89	0.0045
-NPB	-4.78 ± 0.16	0.32 ± 0.04	0.94	0.0015

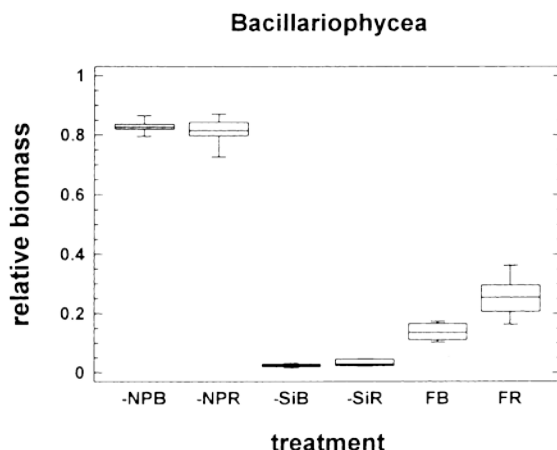


Fig. 5. Relative biomass (taxon biomass/total biomass) of Bacillariophyceae in response to nutrient treatments, box-and-whisker plots showing medians, quartiles and full range of data; F: full enrichment; -Si: enrichment without Si; -NP: enrichment without N and P; B: batch; R: replacement.

lotrix, unidentified flagellates/zoo-spores) and Cyanobacteria (mainly *Anaena naltica*) performed best in the -Si medium, second best in the full medium and worst in the -NP medium (Figs 6 and 7). Medium replacement favoured Cyanobacteria while the batch supply mode favoured the green algae. Individual species responded to the different media in the same way as the higher taxa to which they belonged. However, there were some differences in the response of diatom species/genera to the supply mode. A positive response to medium replacement was found in *Fraeilaria tantlata*, *Acantodes loneipes*, *A'p ora co aei or'1'is*, and *tatroneis constricta*. A negative response to medium replacement was found in unidentified, small centric diatoms, *BerleleYa* (= *A'p ipetra*) *rtilans*, and *Nitzsc ia spp.* *elosira '1'onili or'1'is* responded neutrally to the supply mode.

3.3 *Littorina* growth

The gain of ash-free dry mass responded significantly to the medium composition and the supply mode (Table 1). *Littorina* density did not show up as a significant main effect, but there was a significant interaction between *Littorina* density and supply mode. Although individual regressions of *Littorina* growth on *Littorina* number were insignificant in

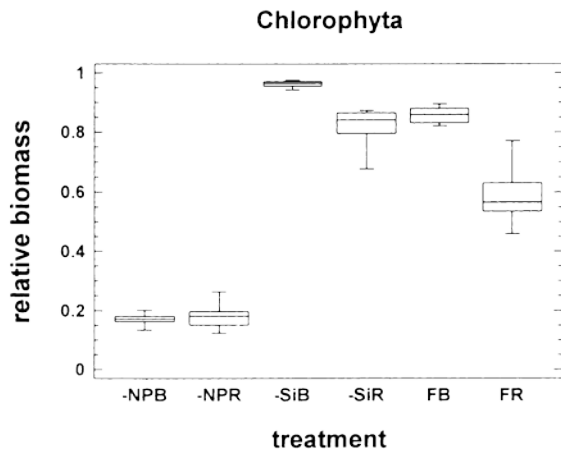


Fig. 6. Relative biomass (taxon biomass/total biomass) of Chlorophyta in response to nutrient treatments, box-and-whisker plots showing median, quartiles and full range of data; F: full enrichment; -Si: enrichment without Si; -NP: enrichment without N and P; B: batch; R: replacement.

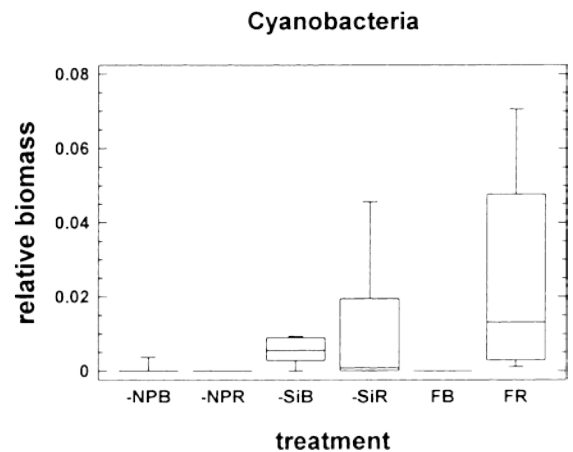


Fig. 7. Relative biomass (taxon biomass/total biomass) of Cyanobacteria in response to nutrient treatments, box-and-whisker plots showing median, quartiles, and full range of data; F: full enrichment; -Si: enrichment without Si; -NP: enrichment without N and P; B: batch; R: replacement.

most cases a consistent contrast in the sign of the regression coefficient between the different nutrient supply modes could be found (Table 5). The regression lines for the replacement and the batch mode were significantly different from each other. In the batch experiments, *Littorina* growth responded positively to density while it responded negatively under nutrient replacement (Fig. 8).

4. Discussion

The N:C stoichiometry of periphyton biomass spans one order of magnitude from 0.01 to 0.21. The lowest margin is ca. one half of the minimum value (0.022) reported by Hillebrand and Sommer

(1999) from a growth experiment in the western Baltic Sea and indicates severe nitrogen limitation of growth. In their study, a C:N:P ratio of 119:17:1 was found to indicate optimal growth, which is remarkably similar to the analogous Redfield-ratio for phytoplankton (106:16:1; Goldman et al., 1979). Due to the scatter of data, Hillebrand and Sommer (1999) have suggested N:C < 0.1 as 'safe' indication of N limitation, provided that N:P < 13. Using this criterion, periphyton growth has to be considered N saturated in the full and the -Si media under the replacement supply mode and N limited in the other treatments, including the possibility of Si limitation of diatoms. The absence of a response of algal N:C to *Littorina* density under medium replacement indicates, that external nutrient supply was sufficient to

Table 5

Linear regressions of *Littorina* growth (in mg AFDM ind⁻¹) on *Littorina* number; probabilities of error: p_{ind} : for individual regression; p_a : for comparison of intercepts; p_b : for comparison of slopes

Treatment	$a \pm SE$	$n \pm SE$	r^2	p_{ind}	p_a	p_b
FR	2.41 ± 0.46	-0.16 ± 0.12	0.31	0.896	0.0059	0.0041
FB	0.06 ± 0.17	0.09 ± 0.04	0.52	0.107		
SiR	2.95 ± 0.33	-0.33 ± 0.175	0.48	0.126	0.0046	0.0476
-SiB	0.36 ± 0.17	0.08 ± 0.04	0.47	0.131		
-NPR	1.34 ± 0.26	-0.15 ± 0.07	0.56	0.087	< 0.0001	0.0825
-NPB	-0.08 ± 0.09	0.13 ± 0.02	0.88	0.0085		

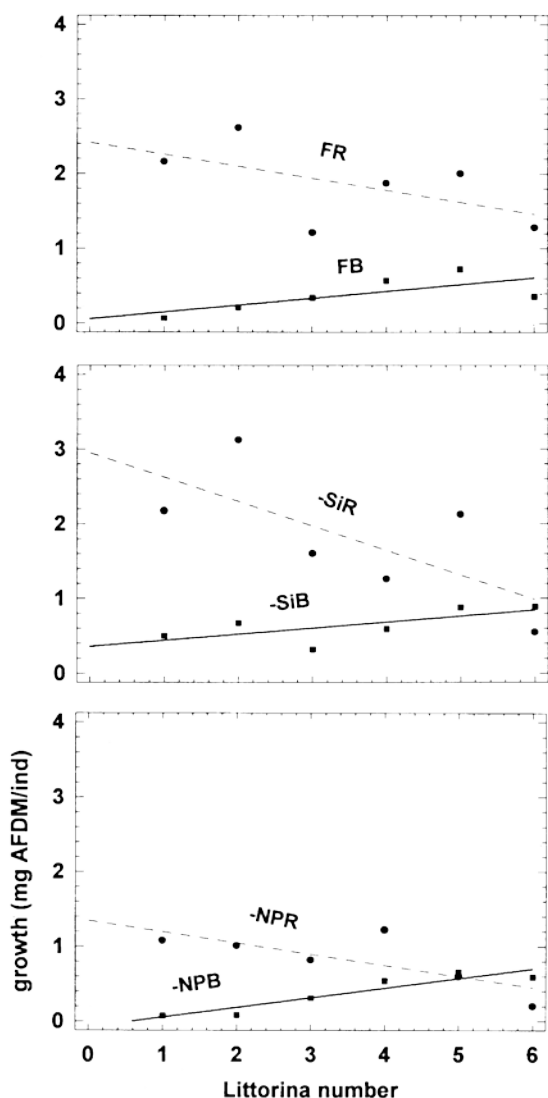


Fig. 8. *Littorina* growth (mg AFDW gained per individual) in response to *Littorina* density. F: full enrichment; -Si: enrichment without Si; -NP: enrichment without N and P; B: batch; R: replacement.

observed any effect on algal N:C by the herbivores. Conversely, in the batch treatments biomass reduction by *Littorina* and nutrient supply by excretion significantly alleviated N limitation; however, this did not lead to N-replete conditions (Fig. 7).

The performance of diatoms in the different media conforms to the resource ratio hypothesis developed in the context of Tilman's (1982) competition theory

and confirmed for periphyton communities by Sommer (1996) and Hillebrand and Sommer (1997). In both studies, an increasing importance of diatoms with increasing Si:N ratios had been found. In the present study, the largest relative biomass of diatoms was found in the -NP medium (highest Si:N ratio) and lowest in the -Si medium (lowest Si:N ratio). However, there is one surprise that cannot be explained at present. The potentially N₂-fixing Cyanobacterium *Anaena naltica* was favoured by medium exchange (no N limitation), while it would have been expected to be most strongly favoured under the most severe N limitation. However, *Anaena* growth might have been restricted by P shortage in the -NP treatments without medium exchange.

The shift from a negative to a positive density dependence of *Littorina* growth indicates poor food quality under nutrient-limited conditions. Here, I use the biomass specific cell quota of nitrogen (N:C) as an index of chemical food quality. The choice does not necessarily imply that I assume nitrogen limitation of *Littorina* growth. Other substances correlated with cellular N might also have been limiting for *Littorina* growth, as has been suggested by Muller-Navarra (1995) for a case of apparent P limitation of the freshwater zooplankton *Daphnia*. In their review, Sterner and Schulz, 1998 propose two possible ways in which limitation by food quality and food quantity could interact. Poor food quality can reduce growth rates at any level of food quantity, or poor food quality could be compensated by enhanced ingestion at high food concentrations. This would permit maximal growth rates even with suboptimal food with a later transition from food limitation to saturation for the low quality food. Compensatory feeding cannot be tested with the data of the present study, because the highest periphyton biomass levels were attained under experimental condition without N limitation (medium replacement, full or -Si medium). Therefore, the combined influence of food quantity and food quality was tested by a multiple regression analysis. A multiplicative effect of food quantity and of food quality would show up as an additive effect if both measures are transformed logarithmically. Therefore, log¹⁰ of periphyton C (mg/cm²) and of biomass N:C (mol/mol) were used as independent variables. Additionally, the arcsine-square root transformed values of diatom, green algal, and cyanobacterial relative

biomass were entered as candidate variables to account for taxonomy effects of food quality. Both the forward and the backward stepwise variable selection (F -to-enter = 4.0) removed all candidate independent variables except for $\log^{10}C$ and $\log^{10}(N:C)$:

$$G = 1.92(\pm 0.34) + 0.39(\pm 0.09)\log^{10}C$$

$$+ 1.54(\pm 0.19)\log^{10}(N:C);$$

$$r^2 = 0.74; \quad p < 0.0001$$

where G = individual growth.

It should be noted that the experimental system is an highly artificial model system which demonstrates the reversal of density dependence of herbivore growth between open (medium exchange) and closed (batch) systems. Especially the batch systems do not mimic the behaviour of natural periphyton-grazer systems, because those are almost always open systems. A large portion of the nutrients excreted by herbivores will usually be washed away by waves and fertilise phytoplankton rather than periphyton. The other possible beneficial effect (reduction of algae which have to share the same nutrient flux) should be similar in open and closed systems.

The reversal of density dependence of herbivore growth because of food quality limitation has previously been reported from zooplankton (Sommer, 1992). The two studies differ from each other in the response variable of the herbivore. In the zooplankton study *in situ* reproduction rates were measured as indicator of food limitation, while here only somatic growth of juveniles was measured. It remains to be seen whether limitation of somatic growth would eventually translate into limitation of population growth of *Littorina*. Traditional predator-prey models frequently include some modification of the logistic growth equation, which assumes negative density dependence of growth rates because of intraspecific competition. For systems where nutrient dynamics are dominated by herbivore recycling, such models should be modified to include intraspecific facilitation (positive density dependence) as well.

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