International Journal of Systematic and Evolutionary Microbiology (2008), 58, 408-413

Actinomycetospora chiangmaiensis gen. nov., sp. nov., a new member of the family *Pseudonocardiaceae*

Yi Jiang,^{1,2} Jutta Wiese,¹ Shu-Kun Tang,² Li-Hua Xu,² Johannes F. Imhoff¹ and Cheng-Lin Jiang²

¹Leibniz-Institut für Meereswissenschaften, IFM-GEOMAR, Düsternbrooker Weg 20, D-24105 Kiel, Germany

²Yunnan Institute of Microbiology, Yunnan University, Kunming 650091, China

A novel actinomycete strain, YIM 0006^{T} , was isolated from soil of a tropical rainforest in northern Thailand. The isolate displayed the following characteristics: aerial mycelium is absent, short spore chains are formed directly on the substrate mycelium, contains *meso*-diaminopimelic acid, arabinose and galactose (cell-wall chemotype IV), the diagnostic phospholipid is phosphatidylcholine, MK-9(H₄) is the predominant menaquinone and the G+C content of the genomic DNA is 69.0 mol%. Phylogenetic analysis and phenotypic characteristics showed that strain YIM 0006^T belongs to the family *Pseudonocardiaceae* but can be distinguished from representatives of all genera classified in the family. The novel genus and species *Actinomycetospora chiangmaiensis* gen. nov., sp. nov. are proposed, with strain YIM 0006^T (=CCTCC AA 205017^T =DSM 45062^T) as the type strain of *Actinomycetospora chiangmaiensis*.

The first description of the family Pseudonocardiaceae was given by Embley et al. (1988), and the description was emended by Stackebrandt et al. (1997) on the basis of 16S rRNA gene sequence analysis. The family currently consists of 14 genera with validly published names: Actinoalloteichus (Tamura et al., 2000), Actinopolyspora (Gochnauer et al., 1975), Amycolatopsis (Lechevalier et al., 1986), Crossiella (Labeda, 2001), Goodfellowia (Labeda & Kroppenstedt, 2006), Kibdelosporangium (Shearer et al., 1986), Kutzneria (Stackebrandt et al., 1994), Prauserella (Kim & Goodfellow, 1999), Pseudonocardia (Henssen, 1957), Saccharomonospora (Nonomura & Ohara, 1971), Saccharopolyspora (Lacey & Goodfellow, 1975), Streptoalloteichus (Tomita et al., 1987), Thermobispora (Wang et al., 1996) and Thermocrispum (Korn-Wendisch et al., 1995). Strain YIM 0006^T was isolated during an investigation of actinomycete diversity in soil from a tropical rainforest in Chiang Mai, in northern Thailand. Here, we report on the classification and characterization of strain YIM 0006^T and propose a novel genus and species of the family Pseudonocardiaceae to accommodate the strain.

Strain YIM 0006^{T} was isolated from a soil sample after 2 weeks incubation at 28 °C on starch-glycerol medium as described by Jiang *et al.* (2006). Cultural characteristics of

the strain were determined after growth at 28 °C for 2 weeks by methods used in the International Streptomyces Project (ISP; Shirling & Gottlieb, 1966) as well as by using Czapek's medium and nutrient agar (Dong & Cai, 2001). Colour determination was performed with colour chips from the ISCC-NBS Color Charts Standard Samples no. 2106 (Kelly, 1964). Morphological observations of spore chains and mycelia were made by light microscopy (Olympus microscope BH-2) and scanning electron microscopy (Philips XL30 ESEM-TMP) after 20–50 days incubation. Gram staining (Hucker's modification; Society for American Bacteriologists, 1957) and Ziehl–Neelsen preparations (Gordon, 1967) were evaluated by light microscopy.

Growth of strain YIM 0006^{T} was poor on most media tested, although the strain grew well but slowly on yeast extractmalt extract agar (ISP 2). Soluble pigments were not produced under any conditions tested in this study. Aerial mycelium was not observed on any of the tested media. The vegetative mycelium fragmented into rod-shaped elements (Fig. 1a) and was pale to brilliant orange–yellow in colour. Short spore chains were formed directly on the vegetative mycelium (Fig. 1a, b). The strain displayed bud-like structures of the spore chains, as has also been described for members of the genus *Pseudonocardia* (Huang *et al.*, 2002). Spores were short and rod-shaped, $0.3-0.6 \times 0.8 1.2 \ \mu$ m. The spore surface was smooth.

Correspondence Johannes F. Imhoff

jimhoff@ifm-geomar.de

Li-Hua Xu lihxu@ynu.edu.cn

Downloaded from www.microbiologyresearch.

The GenBank/EMBL/DDBJ accession number for the 16S rRNA gene sequence of strain YIM 0006^{T} is AM398646.

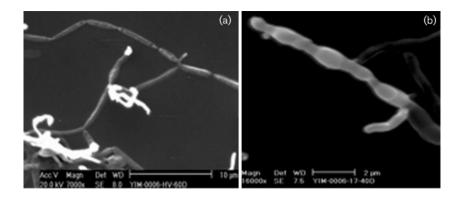


Fig. 1. (a) Scanning electron micrograph of fragments of vegetative mycelium and short spore chains of strain YIM 0006^{T} on HV medium (Hayakawa & Nonomura, 1987) after incubation for 60 days. Bar, 10 μ m. (b) Scanning electron micrograph of short spore chains of strain YIM 0006^{T} on glycerol-asparagine medium (ISP 5) after incubation for 40 days. Bar, 2 μ m.

All tests of physiological and biochemical characteristics of strain YIM 0006^T were performed at 28 °C and recorded after 7, 14, 20 and 30 days, except for the nitrate reduction test, which was recorded after 1, 3 and 5 days. Carbon- and nitrogen-source utilization as well as acid production from sugars under aerobic conditions were examined according to the method of Kämpfer *et al.* (1991). The isolate used a range of carbon sources (see species description). Galactose, arabinose, mannose, raffinose, inositol, mannitol and sodium citrate were not utilized. Tests of gelatin liquefaction, milk coagulation, milk peptonization, starch hydrolysis, nitrate reduction, growth on cellulose, H₂S and melanin production were negative.

Cell material for the extraction of chromosomal DNA and chemotaxonomic studies was obtained after cultivation at 28 °C for 7-10 days in ISP 2 broth (Shirling & Gottlieb, 1966) supplemented with the vitamin mixture of HV medium (Hayakawa & Nonomura, 1987) as a shaking culture. Procedures for analysis of diagnostic cell-wall amino acids and sugars followed those described by Staneck & Roberts (1974). Polar lipids were extracted, examined by two-dimensional TLC and identified using the procedures of Minnikin et al. (1984). Menaquinones were extracted according to Minnikin et al. (1984) and separated by HPLC (Kroppenstedt, 1982). Cellular fatty acid composition was determined as described by Sasser (1990) using the Microbial Identification System (MIDI, Inc.). Chromosomal DNA for genomic DNA G+C content analysis was extracted as described by Marmur (1961). The DNA G+C content was determined by the HPLC method (Tamaoka & Komagata, 1984) with an Agilent 1100 LC system (IRIS Technologies).

The cell wall of strain YIM 0006^{T} contained *meso*diaminopimelic acid as the diagnostic peptidoglycan diamino acid. Whole-cell hydrolysates contained arabinose and galactose as diagnostic sugars (cell-wall chemotype IV; Lechevalier & Lechevalier, 1970). Analysis of phospholipids revealed phosphatidylcholine, phosphatidylinositol and phosphatidylglycerol, indicating phospholipid type PIII (Lechevalier *et al.*, 1977). The predominant menaquinone was MK-9(H₄). The fatty acid profile consisted mainly of iso-branched saturated hexadecanoic acid. The predominant components, as proportions of the total fatty acid composition, were iso- $C_{14:0}$ (1.1 %), iso- $C_{15:0}$ (3.2 %), iso- $C_{16:1}$ H (2.6 %), iso- $C_{16:0}$ (29.8 %), $C_{16:1}\omega7c/iso-C_{15:0}$ 2-OH (17.6 %), $C_{16:0}$ (10.8 %), $C_{16:0}$ 10-methyl (7.2 %), iso- $C_{17:0}$ (2.5 %), anteiso- $C_{17:0}$ (5.5 %), $C_{17:1}\omega8c$ (4.4 %), $C_{17:0}$ (1.6 %), $C_{17:0}$ 10-methyl (1.3 %), $C_{18:1}\omega9c$ (2.3 %) and $C_{18:0}$ (3.7 %). The G+C content of genomic DNA of the strain was 69.0 mol%.

Genomic DNA extraction and PCR amplification of the 16S rRNA gene of strain YIM 0006^T were carried out using procedures described by Xu et al. (2003). The 16S rRNA gene sequence (1456 nucleotides) was compared with corresponding sequences of the family Pseudonocardiaceae from the GenBank/EMBL/DDBJ database by using BLAST (Altschul et al., 1997), BLAST 2 sequences (Tatusova & Madden, 1999) and FASTA (Pearson, 1990). The alignment was performed using CLUSTAL_X (Thompson et al., 1997) and corrected manually. Phylogenetic analysis was conducted using MEGA version 3.1 (Kumar et al., 2004) and the PhyML online web server (Guindon et al., 2005). A distance matrix was generated according to Kimura's twoparameter model (Kimura, 1980, 1983) and a phylogenetic tree was constructed using the neighbour-joining method (Saitou & Nei, 1987). The topology of the tree was evaluated by performing a bootstrap analysis (Felsenstein, 1985) using 1000 resamplings. A maximum-likelihood tree was calculated using the GTR (general time-reversible) substitution model and bootstrap values from 500 resamplings.

A database search demonstrated that strain YIM 0006^{T} belongs to the family *Pseudonocardiaceae* (Stackebrandt *et al.*, 1997). Phylogenetic study was performed with 16S rRNA gene sequences of type strains of *Pseudonocardia* species with validly published names, as far as available, including [*Actinobispora*] *xinjiangensis*, [*Actinobispora*] *aurantiaca*, [*Actinobispora*] *alaniniphila* and [*Actinobispora*] *yunnanensis*, which were combined into the genus *Pseudonocardia* by Huang *et al.* (2002) as well as with sequences of representative type strains of the other 13 genera of the *Pseudonocardiaceae*. The closest relatives of strain YIM 0006^T were *Pseudonocardia halophobica* DSM 43089^T, with 95.24 % sequence identity, *Pseudonocardia benzenivorans* DSM 44703^T and *Pseudonocardia alni* IMSNU

 20049^{T} (both 95.10%). The sequence identity of YIM 0006^{T} to *Kibdelosporangium aridum* DSM 43828^{T} was 94.28%, and the identity to type strains belonging to other genera of the family *Pseudonocardiaceae* was below 94.20%.

According to the phylogenetic tree (Fig. 2), strain YIM 0006^{T} formed a distinct subclade between the genera *Pseudonocardia* and *Kibdelosporangium*. Although the 16S rRNA gene sequence similarity between strain YIM 0006^{T} and members of the genus *Pseudonocardia* fell into the range between the *Pseudonocardia* species (99.6–93.6%) given by Huang *et al.* (2002), the separate branching of the isolate is clearly supported by high bootstrap values of 96% (percentage of 1000 resamplings) and 97% (percentage of 500 resamplings) after calculation of the neighbour-joining tree (Fig. 2) and the maximum-likelihood tree (not shown), respectively.

Strain YIM 0006^{T} and representatives of the next most closely related genus *Pseudonocardia* have the same cell-wall chemotype (chemotype IV; *meso*-diaminopimelic acid, arabinose and galactose), fatty acid type and DNA G+C

content, while the menaquinone pattern clearly distinguishes the new isolate from members of the genus *Pseudonocardia* (Table 1). Several chemotaxonomic characteristics and the absence of sporangium-like structures clearly differentiate strain YIM 0006^{T} from representatives of the phylogenetically close genus *Kibdelosporangium* (Table 1).

On the basis of a combination of phylogenetic distinctness and differences in chemotaxonomic and morphological characteristics, we consider that strain YIM 0006^T represents a novel genus and species, for which the name *Actinomycetospora chiangmaiensis* gen. nov., sp. nov. is proposed.

Description of Actinomycetospora gen. nov.

Actinomycetospora (Ac.ti'no.my.ce.to.spo'ra. N.Gr. n. actinomyces -etos an actinomycete; Gr. fem. n. spora a seed and, in bacteriology, a spore; N.L. fem. n. Actinomycetospora referring to an actinomycete with spore chains).

Aerobic, Gram-positive, non-acid-fast, non-motile actinomycetes. Substrate mycelium fragments into rod-shaped

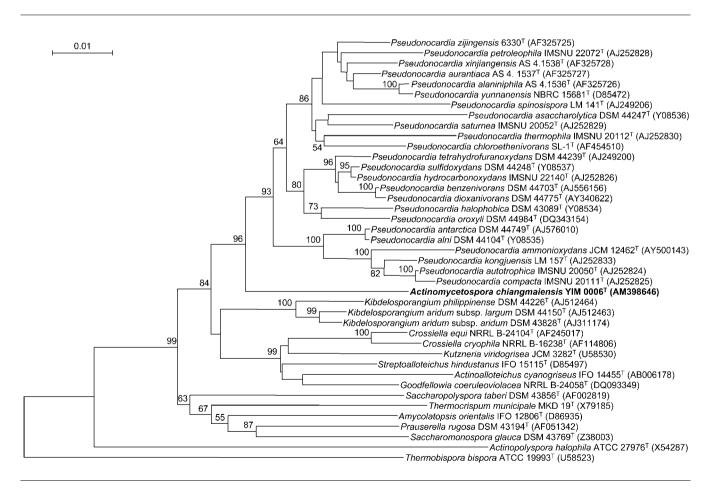


Fig. 2. Neighbour-joining tree derived from 16S rRNA gene sequences showing the relationship of YIM 0006^T and representative species of the 14 genera of the family *Pseudonocardiaceae*. Numbers at branch nodes are bootstrap percentages (1000 resamplings; only values over 50% are given). Bar, 1% sequence divergence.

Genus	Some key morphological characteristics	Sugar pattern	Phospholipids*	Major menaquinone(s)	Fatty acid type†	Fatty acid G+C content type† (mol%)
Kibdelosporangium	Long chains of spores, sporangium-like structure of the aerial	4	PE, PI, PME, PG, DPG, MK-9(H ₂ , H ₄ , H ₆)	MK-9(H ₂ , H ₄ , H ₆)	3с	66
Pseudonocardia	Long or short chains of spores, fragmentation of aerial and substrate mycelia (v), production of spores by budding or	Gic (v), Mia (v) Ara, Gal	PE or PC, PME, PI, PG, MK-8(H ₄) DPG, GlcNu	MK-8(H ₄)	2b	68–79
Actinomycetospora gen. nov. (YIM 0006	septation (V), swollen hyphal segments (V) <i>tctinomycetospora</i> Short chains of spores, bud-like structures, fragmentation of gen. nov. (YIM 0006 ^T) substrate mycelium, absence of aerial mycelium	Ara, Gal	PC, PI, PG	MK-9(H ₄)	2b	69

elements. No aerial mycelium is produced on any medium tested. The cell wall contains *meso*-diaminopimelic acid. Whole-cell hydrolysates contain arabinose and galactose (cell-wall chemotype IV). Phosphatidylcholine is the diagnostic phospholipid, with phosphatidylinositol and phosphatidylglycerol. The predominant menaquinone is MK-9(H₄). The type species is *Actinomycetospora chiangmaiensis*.

Description of Actinomycetospora chiangmaiensis sp. nov.

Actinomycetospora chiangmaiensis (chiang.mai.en'sis. N.L. fem. adj. chiangmaiensis pertaining to Chiang Mai, a city in the north of Thailand in the vicinity of which the type strain was found).

In addition to the characteristics given in the genus description, this species has the following properties. Vegetative mycelium is pale to brilliant orange-yellow in colour. Short spore chains are formed directly from vegetative mycelium. Spores are short and rod-shaped. Spore surfaces are smooth. No soluble pigment is produced. Glucose, fructose, xylose, ribose, rhamnose, sucrose, lactose, sorbitol, glycerol, sodium acetate, asparagine, glycine, histidine and methionine are utilized as sole carbon sources. Acid is not produced from these carbon sources. Gelatin liquefaction, milk coagulation and peptonization, starch hydrolysis, nitrate reduction, growth on cellulose, H₂S and melanin production are negative. The major cellular fatty acids are iso-C_{16:0} (29.8%), C_{16:1}w7c/iso-C_{15:0} 2-OH (17.6%), $C_{16:0}$ (10.8%) and $C_{16:0}$ 10-methyl (7.2%). The G+C content of the DNA of the type strain is 69 mol%.

The type strain, YIM 0006^{T} (=CCTCC AA 205017^T =DSM 45062^T), was isolated from soil collected from a tropical rainforest located at Chiang Mai in the north of Thailand.

Acknowledgements

This research was supported by the National Basic Research Program of China (no. 2004CB719601), the National Natural Science Foundation of China (no. 30560001), the Yunnan Provincial International Cooperative Program (no. 2005GH21), the Yunnan Provincial Natural Science Foundation (no. 2004 C0002Q), the Program for New Century Excellent Talents in University and the Centre of Marine Natural Products, which is funded by the Ministry of Science, Economic Affairs and Transport of the state of Schleswig-Holstein (Germany). We thank Dr Jörg Süling, Miss Xiang-Feng Cai and Mr Yun Chen for technical assistance and discussions.

References

Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W. & Lipman, D. J. (1997). Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25, 3389–3402.

Dong, X.-Z. & Cai, M.-Y. (editors) (2001). Determination of biochemical properties. In *Manual for the Systematic Identification of General Bacteria*, pp. 370–398. Beijing: Science Press (in Chinese).

Data for reference genera were derived from Shearer

Huang et al. (2002), Reichert et al. (1998) and Kämpfer et al. (2006).

et al. (1993), Henssen (1957),

Table 1. Morphological and chemotaxonomic characteristics of strain YIM 0006^T and related genera of the family *Pseudonocardiaceae*

et al. (1986), Mertz & Yao (1988), Tomita

†According to the classification of Kroppenstedt (1985)

Embley, M. T., Smida, J. & Stackebrandt, E. (1988). The phylogeny of mycolate-less wall chemotype IV actinomycetes and description of *Pseudonocardiaceae* fam. nov. *Syst Appl Microbiol* 11, 16–19.

Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39, 783–791.

Gochnauer, M. B., Leppard, G. G., Komaratat, P., Kates, M., Novitsky, T. & Kushner, D. J. (1975). Isolation and characterization of *Actinopolyspora halophila*, gen. et sp. nov., an extremely halophilic actinomycete. *Can J Microbiol* **21**, 1500–1511.

Gordon, R. E. (1967). The taxonomy of soil bacteria. In *The Ecology of Soil Bacteria*, pp. 293–321. Edited by T. R. G. Gray & D. Parkinson. Liverpool: Liverpool University Press.

Guindon, S., Lethiec, F., Duroux, P. & Gascuel, O. (2005). PHYML Online – a web server for fast maximum likelihood-based phylogenetic inference. *Nucleic Acids Res* **33**, W557–W559.

Hayakawa, M. & Nonomura, H. (1987). Humic acid-vitamin agar, a new medium for the selective isolation of soil actinomycetes. *J Ferment Technol* 65, 501–509.

Henssen, A. (1957). Beiträge zur Morphologie und Systematik der thermophilen Actinomyceten. Arch Mikrobiol 26, 373–414 (in German).

Huang, Y., Wang, L. M., Lu, Z. T., Hong, L., Liu, Z. H., Tan, G. Y. A. & Goodfellow, M. (2002). Proposal to combine the genera *Actinobispora* and *Pseudonocardia* in an emended genus *Pseudonocardia*, and description of *Pseudonocardia zijingensis* sp. nov. *Int J Syst Evol Microbiol* **52**, 977–982.

Jiang, Y., Duan, S. R., Tang, S. K., Cheng, H. H., Li, W. J. & Xu, L. H. (2006). Isolation methods of rare actinomycetes. *Microbiology* (*Beijing*) 33, 181–183.

Kämpfer, P., Kroppenstedt, R. M. & Dott, W. (1991). A numerical classification of the genera *Streptomyces* and *Streptoverticillium* using miniaturized physiological tests. *J Gen Microbiol* 137, 1831–1891.

Kämpfer, P., Kohlmeyer, U., Thiemer, B. & Andreesen, J. R. (2006). *Pseudonocardia tetrahydrofuranoxydans* sp. nov. *Int J Syst Evol Microbiol* 56, 1535–1538.

Kelly, K. L. (1964). Inter-Society Color Council – National Bureau of Standards Color Name Charts Illustrated with Centroid Colors. Washington, DC: US Government Printing Office.

Kim, S. B. & Goodfellow, M. (1999). Reclassification of *Amycolatopsis* rugosa Lechevalier *et al.* 1986 as *Prauserella rugosa* gen. nov., comb. nov. *Int J Syst Bacteriol* **49**, 507–512.

Kimura, **M.** (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 16, 111–120.

Kimura, M. (1983). The Neutral Theory of Molecular Evolution. Cambridge: Cambridge University Press.

Korn-Wendisch, F., Rainey, F., Kroppenstedt, R. M., Kempf, A., Majazza, A., Kutzner, H. J. & Stackebrandt, E. (1995). *Thermocrispum* gen. nov., a new genus of the order *Actinomycetales*, and description of *Thermocrispum municipale* sp. nov. and *Thermocrispum agreste* sp. nov. *Int J Syst Bacteriol* **45**, 67–77.

Kroppenstedt, R. M. (1982). Separation of bacterial menaquinones by HPLC using reverse phase (RP18) and a silver loaded ion exchanger as stationary phases. *J Liq Chromatogr* **5**, 2359–2367.

Kroppenstedt, R. M. (1985). Fatty acid and menaquinone analysis of actinomycetes and related organisms. In *Chemical Methods in Bacterial Systematics* (Society for Applied Bacteriology Technical Series vol. 20), pp. 173–199. Edited by M. Goodfellow & D. E. Minnikin. New York: Academic Press.

Kumar, S., Tamura, K. & Nei, M. (2004). MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. *Brief Bioinform* 5, 150–163.

Labeda, D. P. (2001). Crossiella gen. nov., a new genus related to Streptoalloteichus. Int J Syst Evol Microbiol 51, 1575–1579.

Labeda, D. P. & Kroppenstedt, R. M. (2006). *Goodfellowia* gen. nov., a new genus of the *Pseudonocardineae* related to *Actinoalloteichus*, containing *Goodfellowia coeruleoviolacea* gen. nov., comb. nov. *Int J Syst Evol Microbiol* 56, 1203–1207.

Lacey, J. & Goodfellow, M. (1975). A novel actinomycete from sugarcane bagasse, *Saccharopolyspora hirsuta* gen. et sp. nov. *J Gen Microbiol* 88, 75–85.

Lechevalier, M. P. & Lechevalier, H. A. (1970). Chemical composition as a criterion in the classification of aerobic actinomycetes. *Int J Syst Bacteriol* **20**, 435–443.

Lechevalier, M. P., De Bièvre, C. & Lechevalier, H. A. (1977). Chemotaxonomy of aerobic actinomycetes: phospholipid composition. *Biochem Syst Ecol* 5, 249–260.

Lechevalier, M. P., Prauser, H., Labeda, D. P. & Ruan, J. S. (1986). Two new genera of nocardioform actinomycetes: *Amycolata* gen. nov. and *Amycolatopsis* gen. nov. *Int J Syst Bacteriol* **36**, 29–37.

Marmur, J. (1961). A procedure for the isolation of deoxyribonucleic acid from microorganisms. *J Mol Biol* **3**, 208–218.

Mertz, F. P. & Yao, R. C. (1988). Kibdelosporangium philippinense sp. nov. isolated from soil. Int J Syst Bacteriol 38, 282–286.

Minnikin, D. E., O'Donnell, A. G., Goodfellow, M., Alderson, G., Athalye, M., Schaal, A. & Parlett, J. H. (1984). An integrated procedure for the extraction of isoprenoid quinones and polar lipids. *J Microbiol Methods* **2**, 233–241.

Nonomura, H. & Ohara, Y. (1971). Distribution of actinomycetes in soil. X. New genus and species of monosporic actinomycetes. *J Ferment Technol* 49, 895–903.

Pearson, W. R. (1990). Rapid and sensitive sequence comparison with FASTP and FASTA. *Methods Enzymol* **183**, 63–98.

Reichert, K., Lipski, A., Pradella, S., Stackebrandt, E. & Altendorf, K. (1998). *Pseudonocardia asaccharolytica* sp. nov. and *Pseudonocardia sulfidoxydans* sp. nov., two new dimethyl disulfide-degrading actinomycetes and emended description of the genus *Pseudonocardia*. Int J Syst Bacteriol 48, 441–449.

Saitou, N. & Nei, M. (1987). The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4, 406–425.

Sasser, M. (1990). Identification of bacteria by gas chromatography of cellular fatty acids. USFCC Newsl 20, 16.

Shearer, M. C., Colman, P. M., Ferrin, R. M., Nisbet, L. J. & Nash, C. H. (1986). New genus of the *Actinomycetales: Kibdelosporangium aridum* gen. nov., sp. nov. *Int J Syst Bacteriol* **36**, 47–54.

Shirling, E. B. & Gottlieb, D. (1966). Methods for characterization of *Streptomyces* species. *Int J Syst Bacteriol* 16, 313–340.

Society for American Bacteriologists (1957). *Manual of Microbiological Methods.* New York: McGraw-Hill.

Stackebrandt, E., Kroppenstedt, R. M., Jahnke, K. D., Kemmerling, C. & Gürtler, H. (1994). Transfer of *Streptosporangium viridogriseum* (Okuda et al. 1966), *Streptosporangium viridogriseum* subsp. *kofuense* (Nonomura and Ohara 1969), and *Streptosporangium albidum* (Furumai et al. 1968) to *Kutzneria* gen. nov. as *Kutzneria viridogrisea* comb. nov., *Kutzneria kofuensis* comb. nov., and *Kutzneria albida* comb. nov., respectively, and emendation of the genus *Streptosporangium*. Int J Syst Bacteriol 44, 265–269.

Stackebrandt, E., Rainey, F. A. & Ward-Rainey, N. L. (1997). Proposal for a new hierarchic classification system, *Actinobacteria* classis nov. *Int J Syst Bacteriol* **47**, 479–491.

Staneck, J. L. & Roberts, G. D. (1974). Simplified approach to identification of aerobic actinomycetes by thin-layer chromatography. *Appl Microbiol* **28**, 226–231.

International Journal of Systematic and Evolutionary Microbiology 58

Tamaoka, J. & Komagata, K. (1984). Determination of DNA base composition by reversed-phase high-performance liquid chromatography. *FEMS Microbiol Lett* **25**, 125–128.

Tamura, T., Liu, Z. H., Zhang, Y. M. & Hatano, K. (2000). Actinoalloteichus cyanogriseus gen. nov., sp. nov. Int J Syst Evol Microbiol 50, 1435–1440.

Tatusova, T. A. & Madden, T. L. (1999). BLAST 2 sequences – a new tool for comparing protein and nucleotide sequences. *FEMS Microbiol Lett* 174, 247–250.

Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G. (1997). The CLUSTAL_X Windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res* **25**, 4876–4882.

Tomita, K., Nakakita, Y., Hoshino, Y., Numata, K. & Kawaguchi, H. (1987). New genus of the *Actinomycetales: Streptoalloteichus hindustanus* gen. nov., nom. rev.; sp. nov., nom. rev. *Int J Syst Bacteriol* **37**, 211–213.

Tomita, K., Hoshino, Y. & Miyaki, T. (1993). *Kibdelosporangium albatum* sp. nov., producer of the antiviral antibiotics cycloviravins. *Int J Syst Bacteriol* 43, 297–301.

Wang, Y., Zhang, Z. & Ruan, J. S. (1996). A proposal to transfer *Microbispora bispora* (Lechevalier 1965) to a new genus, *Thermobispora* gen. nov., as *Thermobispora bispora* comb. nov. *Int J Syst Bacteriol* 46, 933–938.

Xu, P., Li, W. J., Xu, L. H. & Jiang, C. L. (2003). A microwave based method for genomic DNA extraction from actinomycetes. *Microbiology (Beijing)* 30, 82–84.