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The abundance, taxa richness and diversity of periphytic algae in the Szigetköz region 1991-1995.

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Abstract

Quality and quantity of the periphytic algae were investigated before and after the diversion of the main arm of Danube at two big branch-system of Szigetköz. Before the diversion 198 taxa, after it 108 taxa were recorded at Cikolasziget, and 138 taxa before and 96 after the diversion were observed at Ásványráró respectively. While the number of taxa decreased the abundance increased, the diversity decreased at all the sampling points. Floristic changes also were detected.

Introduction

The periphytic algae are ideal organisms for biomonitoring (Whitton 1991). The algae have a short lifetime so they usually react fast to changes in their environment. The changes in the quantity of communities indicate the changes in their habitat. Most taxa of attached or periphytic algae are fixed to the substrata so their presence or absence and changes in their quantity are characteristic for the habitat.

Study area:

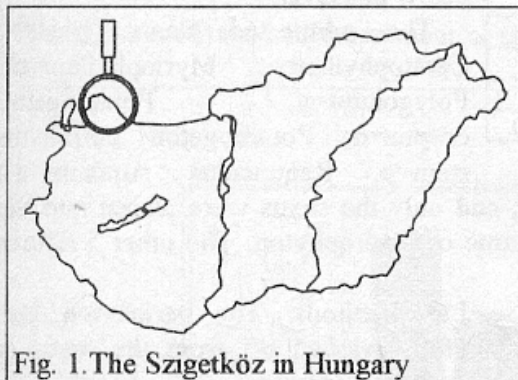


Fig. 1. The Szigetköz in Hungary

The Danube is the only Hungarian river which has an extended branch system. So Szigetköz stands alone for its value as an unique habitat.

The Hungarian and Czechoslovakian governments decided to build a water power plant to satisfy a growing energy demand in 70s. This region is endangered by the Bős-Nagymaros Barrage System.

The Szigetköz is situated at the northwestern part of Hungary, where the Danube enters the Carpathian Basin (Fig.1.). Here, the large quick flowing river slows down, and the main arm of the river is divided many smaller arms. The Szigetköz is an island bounded by the main arm of the river (Old-Danube) and the Mosoni Danube. This island is 52 km long, on average 7-8 km wide its area is 375 sq. km.

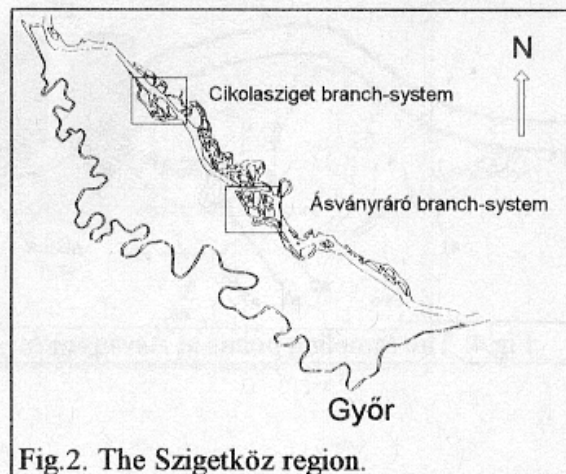


Fig.2. The Szigetköz region.

Many experts drew attention to the series of very complex environmental processes resulting in predominantly negative degradative changes in the area. (Expert group... 1994). Owing to these risks shown by analyses and environmental impact assessments, the Hungarian party suspended construction in May of 1989. The Slovak party continued the construction, and a modified version - so called variant C - began to operate on 25th October 1992. The main arm (Old-Danube) was diverted, only a small portion of the total discharge (10-20 %) flow in the bed of Old-Danube. Consequently the Szigetköz branch-system lost most of their water supply and large areas became dry and the water level decreased in every branche. After the diversion 1992 a bit more water has been brought back to these branches, however the natural water level fluctuations are missing nevertheless.

Our investigations on the periphytic algae join to the Hungarian biomonitoring research in Szigetköz.

The references of previous algological work at Szigetköz (mainly on fitoplankton) available in Ács & Buczkó article in this volume.

Material and Methods

The samples were collected at 8 sampling points of the two big branch systems (Fig.2.) at Cikolasziget branch system = CBS (Fig 3.) and Ásványráró branch-system ÁBS (Fig.4.) before and after the diversion of Danube. The first two figure of the 8 places code of samples refer to the sampling points.

Sampling methods: The stem of macrophyta (*Phragmites*=n and *Typha*=g) were cut off at water level, and about 10 cm below this level.

Table 1.	Cikolasziget	Ásványráró
1991	13-15.07. and 07.09.	
1992		26.06.
1995	23.08 and 7.10.	23.08 and 7.10.

were carefully taken off, the leaves were cut off, and only the stems were placed into the sampling jars. (The 3d figure of code gives the name of macrophyton. The other 5 figures give the date of the collection see Table.1)

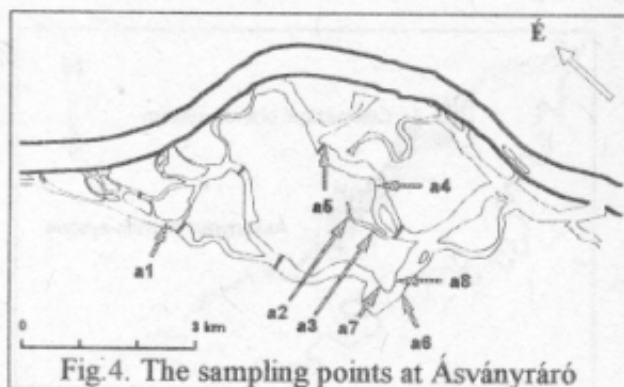


Fig. 4. The sampling points at Ásványráró

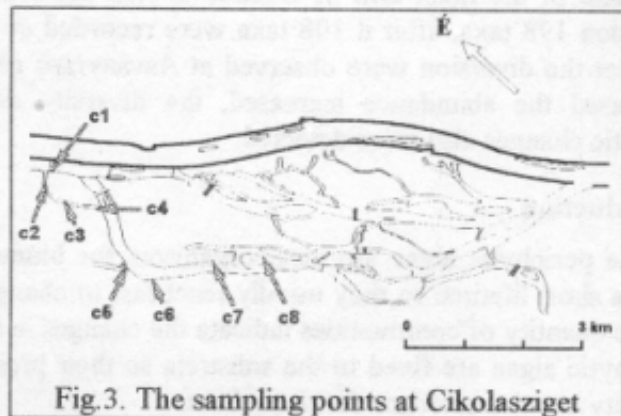


Fig. 3. The sampling points at Cikolasziget

The submersed plants (*Najas*=j, *Ceratophyllum*=c, *Myriophyllum*=m, *Polygonum*=y, *Potamogeton crispus*=o, *Potamogeton perfoliatus* stem=p, *Ranunculus Aquatilis*=q.)

Lab methods: The periphyton was carefully washed off from the stems in lab, these samples were handled as plankton samples. The abundance refers to the surface unit of stems. Diatoms were identified under light and scanning electron microscope, after the cleaning of the valves by H_2O_2 . For statistical analysis SYN-TAX III. program was used (Podani 1988).

Results and discussion

The quantity of periphyton

After the diversion the abundance of attached algae increased at both branch-system, in every sampling point (Fig. 5.). The average of abundance increased as well as the standard deviation, so the increase of maximum even more.

At CBS about four times more attached algae lived in every macrophyton than in 1991. At ÁBS the abundance of attached algae increased about ten times. Moreover the density of macrophyta also increased, so the quantity of benthic flora multiplied here. This means, the benthic eutrophication became definitely obvious.

The increase of abundance was observable in summer and also in autumn. For example 2. Table show the changes of abundance according to seasons at CBS.

	average of abundance *10 ⁶	standard deviation	maximum *10 ⁶	minimum *10 ⁶	number of samples
1991 summer Cikolasziget	1.09	0.822	2.16	0.16	10
1991 autumn Cikolasziget	1.15	0.900	2.50	0.01	10
1995 summer Cikolasziget	3.82	3.219	10.74	0.14	10
1995 autumn Cikolasziget	5.21	3.894	10.48	0.50	8

The number of taxa and the number of taxa/samples decreased in both branch system.

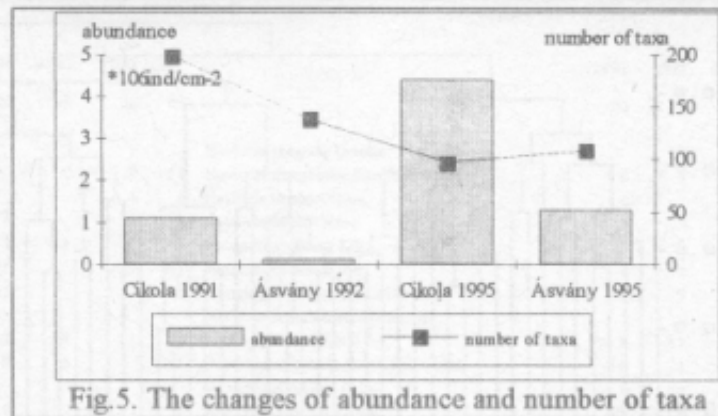


Fig. 5. The changes of abundance and number of taxa

Richness of taxa

A totale of 63 samples were analyzed during this study. Table 3. contains the taxa which occurred in more than 5 p.c. of samples.

Achnanthes minutissima was the most abundant and the most frequent species.

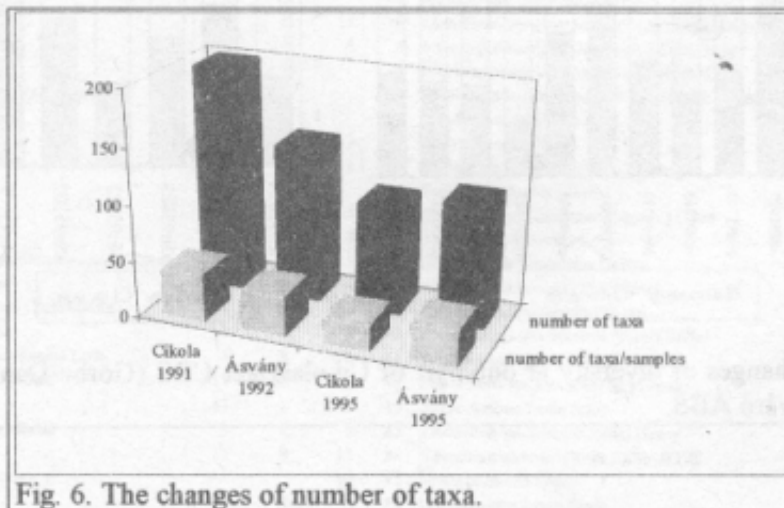


Fig. 6. The changes of number of taxa.

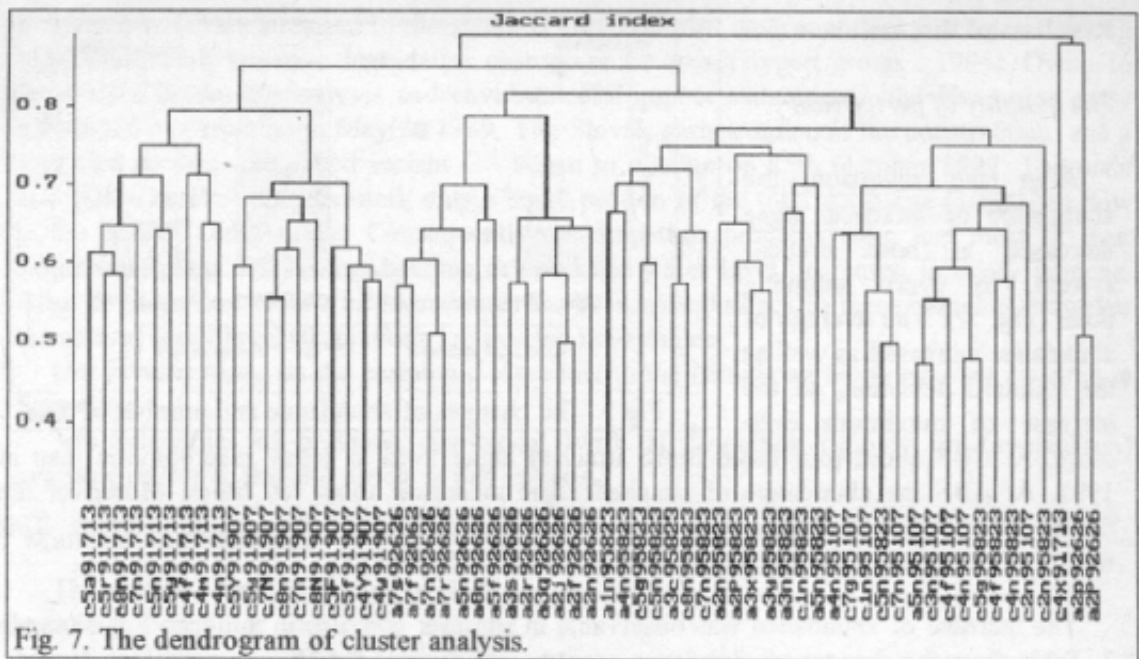


Fig. 7. The dendrogram of cluster analysis.

With the exceptions 3 samples the other 60 samples form clearly three groups (Fig. 7.). The first group contains the samples collected in 1991 at CBS. The second one consists of the samples of ÁBS in 1992. The third group contains the samples of 1995, here, the cluster analysis did not show separation according to the origine of the samples. The arrangements of groups refer to seasons instead of the sampling points.

It seems that, the formerly different floristic composition found at CBS and ÁBS disappeared until 1995 and the floristic composition of periphytic algae became uniform.

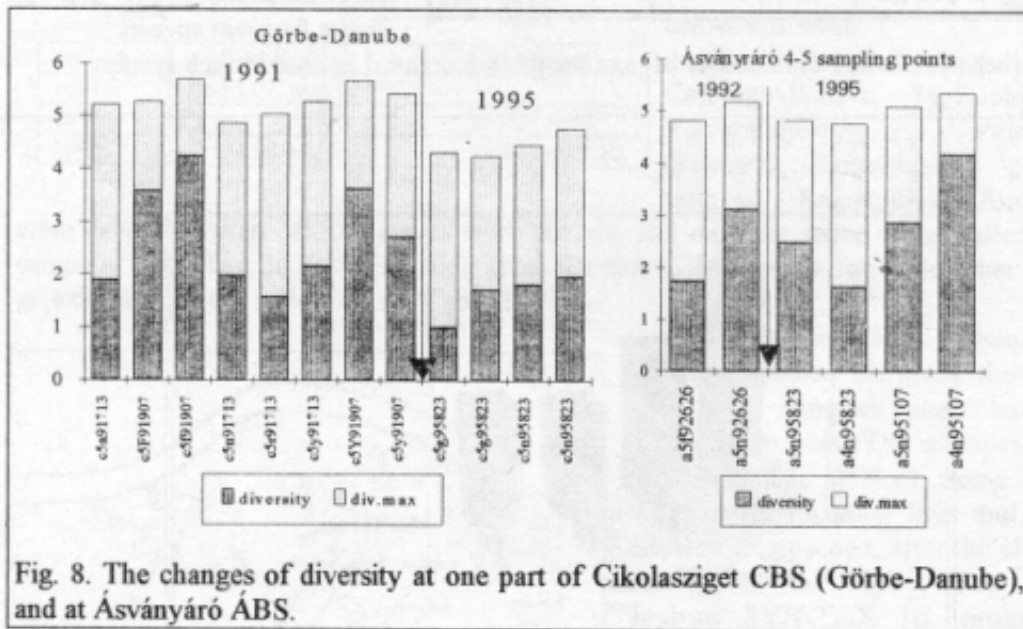


Fig. 8. The changes of diversity at one part of Cikolasziget CBS (Görbe-Danube), and at Ásványáró ÁBS.

Table 3. Taxa occurring in more than 5 p. c. of the samples

	1991	1992	1995 sum			1991	1992	1995 sum	
number of samples	20	15	28	63		20	15	28	63
Cyanobacteria					<i>Navicula gregaria</i> Donkin		4		4
<i>Anabaena catenulata</i> (Kütz.) Born. & Flah.		2	9	11	<i>Navicula margalithii</i> Lange-Bertalot	9	2	19	30
<i>Coelastrum kuetzingianum</i> Näg.			4	4	<i>Navicula minima</i> Grun.	4			4
<i>Merisnopeia glauca</i> (Ehr.) Näg.	6		2	8	<i>Navicula papula</i> Kütz.	2	2	1	5
<i>Merisnopeia warmingiana</i> Lagerheim	3	1	1	5	<i>Navicula pygmaea</i> Kütz.	11			11
<i>Planktolyngbya subtilis</i> (W. West) Anagnostidis & K.	18		25	43	<i>Navicula radiosa</i> Kütz.	8	7	1	16
<i>Pseudanabaena costata</i> Laut. erb.	15			15	<i>Navicula rhynchocephala</i> Kütz.	16	7	11	34
Cryptophyta					<i>Navicula subminutula</i> Manguin				4
<i>Cryptomonas erosa</i> Ehr.	2	4		6	<i>Navicula veneta</i> Kütz.	7	11	24	42
<i>Cryptomonas ovata</i> Ehrenberg	8	5		13	<i>Nitzschia acicularis</i> (Kütz.) W. Smith	10	1	2	13
Chrysophyta, Chrysophyceae					<i>Nitzschia amphibia</i> Grun.	5	4	6	15
<i>Chrysoococcus rufescens</i> Klebs	9			9	<i>Nitzschia angustata</i> Grun.	3	4	6	13
<i>Dinobryon divergens</i> Imhof	3	2		5	<i>Nitzschia capitellata</i> Hust.				4
<i>Dinobryon sertularia</i> Ehr.	4	4		8	<i>Nitzschia dissipata</i> (Kütz.) Grun.	7	5	17	29
Chrysophyta, Bacillariophyceae					<i>Nitzschia fonticola</i> Grun.	8			1
<i>Achnanthes delicatula</i> (Kütz.) Grun.		4		4	<i>Nitzschia frustulata</i> (Kütz.) Grun.		6	8	14
<i>Achnanthes lanceolata</i> Bréb.	7	5	1	13	<i>Nitzschia fruticosa</i> Hust.	14	9		23
<i>Achnanthes lanceolata</i> var. <i>rostrata</i> Hust.	4			4	<i>Nitzschia inconspicua</i> Grun.		5		5
<i>Achnanthes minutissima</i> Kütz.	20	13	27	60	<i>Nitzschia linearis</i> (Agardh) W. Smith	14	3	10	27
<i>Achnanthes plonensis</i> Hust. ed.	4	1	8	13	<i>Nitzschia palca</i> (Kütz.) W. Smith	7		9	16
<i>Amphora coffeaeformis</i> (Agardh) Kützing			4	4	<i>Nitzschia recta</i> Hantzsch	5	4	8	17
<i>Amphora ovalis</i> (Kütz.) Kütz.	5	4	10	19	<i>Nitzschia signondea</i> (Nitzsch) W. Smith	6		2	8
<i>Amphora pediculus</i> (Kütz.) Grun.	14	5	19	38	<i>Nitzschia sublinearis</i> Hust.				4
<i>Asterionella formosa</i> Hass.	1	1	3	5	<i>Rhizocosphaenia abbreviata</i> (Agardh) Lange	9	7	19	35
<i>Aulacoseira distans</i> (Ehr.) Kütz.	1	3	1	5	<i>Skeletonema potamos</i> (Weber) Hasle	4		3	7
<i>Aulacoseira granulata</i> var. <i>angustissima</i> Müll.	1	1	3	5	<i>Stephanodiscus hantzschii</i> f. <i>tenuis</i> (Hust.) Hak et S.	8			8
<i>Aulacoseira italica</i> (Ehr.) Sim.		3	1	4	<i>Stephanodiscus hantzschii</i> Grun. f. <i>hantzschii</i>	4			4
<i>Aulacoseira italica</i> var. <i>tenuissima</i> (Grun.) O. Müll.	7		1	8	<i>Stephanodiscus invisitatus</i> Hohn et Heller	6			6
<i>Cocconeis pediculus</i> Ehr.	16	6	17	39	<i>Surirella ovalis</i> Bréb.			5	5
<i>Cocconeis placentula</i> Ehr.	5	10	25	40	<i>Thalassiorisica pseudonana</i> Hasle et Heimd	14			14
<i>Cyclotephanos dubius</i> (Fricke) Round	4			4	Chlorophyta				
<i>Cyclotella meneghiniana</i> Kütz.	14			14	<i>Characium ensiforme</i> Herm.	16	8	3	27
<i>Cyclotella pseudostelligera</i> Hust.	4			4	<i>Characium ornithocephalum</i> A. Br.	3	2	1	6
<i>Cymatopleura solea</i> (Bréb.) W. Smith	3		1	4	<i>Chlamydomonas reinhardtii</i> Dang.	9	13	9	31
<i>Cymbella affinis</i> Kütz.	11	7	27	45	<i>Cladophora fracta</i> (Dillw.) Kütz.	5			5
<i>Cymbella aspera</i> (Ehr.) Cleve	5	6		11	<i>Cladophora glomerata</i> (L.) Kütz.	1		3	4
<i>Cymbella caespitosa</i> (Kütz.) Brun.	1	1	4	6	<i>Coelastrum microporum</i> Näg.	2	9	1	12
<i>Cymbella cistula</i> (Ehr.) Kütz.	1	2	2	5	<i>Coelastrum sphaericum</i> Näg.	3	1	4	8
<i>Cymbella cymbiformis</i> Agardh	5		4	9	<i>Cosmarium granatum</i> Bréb.	5		2	7
<i>Cymbella microcephala</i> Grun.	5		11	16	<i>Crucigenia quadrata</i> Motr.	2	5	2	9
<i>Cymbella minuta</i> Hilse ex Rabenhorst	16		2	18	<i>Crucigenia tetrapedia</i> (Kütz.) W. & G. S. West	2	5	2	9
<i>Cymbella proxima</i> Rütmer			8	8	<i>Dictyosphaerium ehrenbergianum</i> Näg.	1	4		5
<i>Cymbella silesiaca</i> Bleisch		12	11	23	<i>Dictyosphaerium pulchellum</i> Wood.	5	5	2	12
<i>Diatoma tenuis</i> Agardh	4	5		9	<i>Didymocystis planctonica</i> Kors.	7			7
<i>Diatoma vulgare</i> Bory	9	6	11	26	<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	9	9	12	30
<i>Epithema adnata</i> (Kütz.) Bréb.	4	1	1	6	<i>Monoraphidium griffithii</i> (Berk.) Kom.-Legn.	3	7	3	13
<i>Epithema sorex</i> Kütz.	3	1		4	<i>Monoraphidium minutum</i> (Näg.) Kom.-Legn.	3	3	4	10
<i>Eunotia exigua</i> (Bréb. ex Kütz.) Rabenhorst	4			4	<i>Monoraphidium mirabile</i> (W. & G. S. West) Kom.-Legn.	3	3	7	13
<i>Fragilaria brevistriata</i> Grun.	8	9		17	<i>Neodesmus danubialis</i> Hind.	1	3		4
<i>Fragilaria capucina</i> Desm.			7	7	<i>Oocystis borgeri</i> Snow	5		1	6
<i>Fragilaria capucina</i> var. <i>vaucheriae</i> (Kütz.) Lange-Bert.	3	1	1	5	<i>Pediastrum boryanum</i> (Turp.) Menegh.	1	2	1	4
<i>Fragilaria crotonensis</i> Kütton		5		5	<i>Pediastrum duplex</i> Meyen		3	1	4
<i>Fragilaria pinnata</i> Ehr.	4	11	5	20	<i>Scenedesmus acuminatus</i> (Lagerh.) Chod.	1	11	12	24
<i>Fragilaria ulna</i> (Nitzsch) Ehr.	20	9	9	38	<i>Scenedesmus acutus</i> Meyen	2	1	5	8
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bert. var. <i>acus</i> (Kütz.)	13	9	5	27	<i>Scenedesmus bicaudatus</i> Dedus.				4
<i>Gomphonema acuminatum</i> Ehr.	6	3	4	13	<i>Scenedesmus ecomis</i> (Ehr.) Chod.	4	10	12	26
<i>Gomphonema angustatum</i> (Kütz.) Rabenhorst	1		11	12	<i>Scenedesmus opoliensis</i> P. Richt.				10
<i>Gomphonema minutum</i> Agardh			8	8	<i>Scenedesmus quadricauda</i> (Furp.) Bréb.	7	10	17	34
<i>Gomphonema olivaceum</i> (Homemann) Bréb.	4	3	6	13	<i>Scenedesmus spinosus</i> Chod.	2	1	6	9
<i>Gomphonema parvulum</i> (Kütz.) Kütz.	17	11	13	41	<i>Schroederia setigera</i> (Schröd.) I. cumm.	3	6		9
<i>Gomphonema truncatum</i> Ehr.	11	1	7	19	<i>Stigeoclonium tenue</i> Kütz.	18	7	7	32
<i>Gyrosigma acuminatum</i> (Kütz.) Rabb.	3	1	9	13	<i>Tetraedron caudatum</i> (Corda) Hansg.	1	4	2	7
<i>Melosira varians</i> Ag.	13	8	13	34	<i>Tetrastrum glabrum</i> (Roll) Ahlstr. & Tiff.	4	1	3	8
<i>Navicula capitatoradiata</i> Germain			12	12	<i>Ulothrix zonata</i> Kütz.	5			5
<i>Navicula cryptocephala</i> Kütz.	16	9	5	30	<i>Uronema elongatum</i> Hodg.	6			6

Diversity

The diversity and its maximum follow the decrease of number of taxa at CBS. Fig. 8. shows the changes of diversity at sampling point c5. Both values are decreased here. This phenomenon can be observed at every sampling points at CBS which is the most disturbed area.

By contrast, Fig. 8. presents the same values at the almost undisturbed part of ÁBS. Though the diversity in undisturbed area no decrease, we suppose, that the changes at CBS caused by the diversion of Danube, and connected work impact and no other influences in the concerned part of Szigetköz caused the decrease of diversity instead of other changes (e.g. the influence of weather or amount of discharge).

Summary

To follow the long term changes in Szigetköz, the samples collected in 1991 and in 1992 constitute a status survey, and they were compared to samples collected in 1995. 8 sampling points chosen in the branch systems called Cíkolasziget and Ásványráró, respectively. The quantity of periphyton increased in every sampling points (bentonic eutrophication!) while the number of taxa and the diversity decreased. In the more disturbed branch system (Cíkolasziget) more changes were observed than in the moderately disturbed Ásványráró branch system.

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