

Adaptation of ion regulatory capacities in fish gills under hypercapnic acidosis



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Introduction

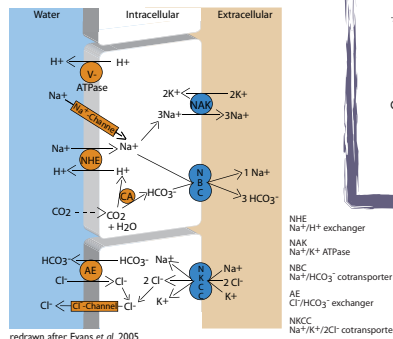
The currently rising CO₂ levels in the atmosphere and in marine surface waters combine with trends of increasing temperatures.

Simultaneous shifts in those factors will likely enhance sensitivity especially in stenothermal animals (reviewed in Pörtner *et al.* 2004).

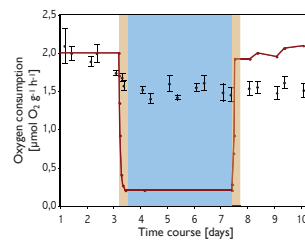
The preservation of ion balance and the regulation of pH despite environmental fluctuations play a special role in this context, and support the maintenance of vital cellular functions. Therefore, it comprises a significant fraction of the energy budget.

Here, we investigated the time course of acclimation to hypercapnia (10,000 ppm) in the benthic eelpout *Zoarces viviparus* at optimum temperature.

The contribution of different transporters to ion and pH regulation in the gills were studied on functional, translational and transcriptional levels.



Whole Animal Regulation Capacity



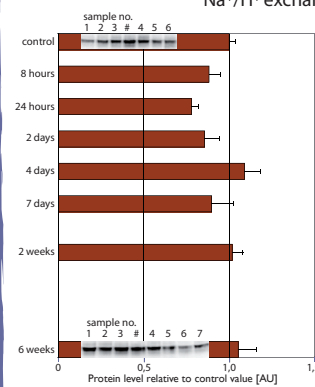
Zoarces viviparus shows a high capacity for pH and ion regulation, comparable to other studies on fish (reviewed in Ishimatsu *et al.* 2005).

Whole animal oxygen consumption remains constant during and after acute exposure to 10,000 ppm hypercapnic incubation and recovery (time course 3.5 - 4.0 - 3.0 days).



Ion Transporter Protein Levels

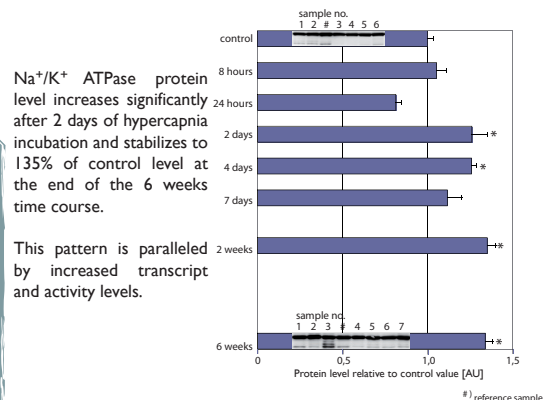
Na⁺/H⁺ exchanger



Na⁺/H⁺ exchanger protein level shows a slight decrease within the first 2 days of hypercapnia incubation, followed by recovery after 4 days and stabilization at control level until the end of the 6 weeks trial.

The protein levels correspond to the respective transcript levels.

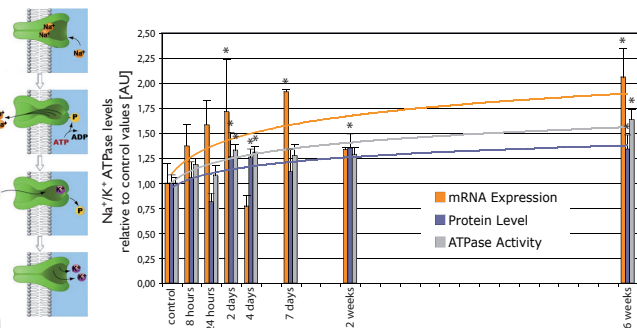
Na⁺/K⁺ ATPase



Na⁺/K⁺ ATPase protein level increases significantly after 2 days of hypercapnia incubation and stabilizes to 135% of control level at the end of the 6 weeks time course.

This pattern is paralleled by increased transcript and activity levels.

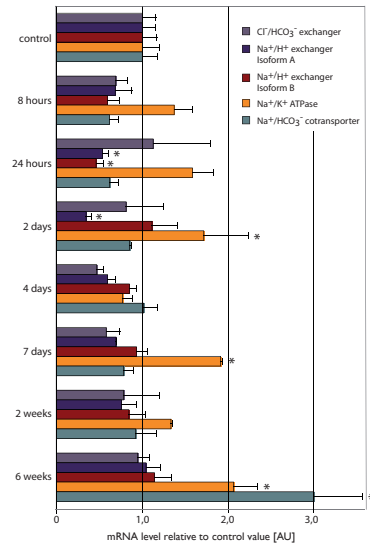
Gill Na⁺/K⁺ ATPase Capacity



Na⁺/K⁺ ATPase regulation follows a similar pattern on different regulation levels.

Higher functional capacities are established by higher protein and mRNA levels.

Ion Transporter mRNA Expression



Overall, the expression of the „passive“ transporters is reduced during the early phase of acclimation, and is restored at the end.

The expression of the active Na⁺ pump remains high during the early phase and doubles until the end of acclimation.

During early phase the Na⁺/HCO₃⁻ cotransporter behaves like the other passive transporters, but expression is about threefold at the end of acclimation.

Experimental Procedure

Long term exposure to hypercapnia

Individuals of *Zoarces viviparus* were kept at optimum temperature (10°C) before and during exposure to 10,000 ppm hypercapnic incubation (corresponding pH of 6.9).

Groups of 8 - 10 fishes were sampled for each point of the time course.

Gill tissue was dissected and frozen immediately in liquid nitrogen.

RNA was isolated and used for mRNA quantification of single ion transporters by Real-time PCR.

Tissue homogenates were prepared to determine Na⁺/K⁺ ATPase activity as well as Na⁺/H⁺ exchanger and Na⁺/K⁺ ATPase protein quantities by Western blotting.

Conclusion & Prospects

Even though whole animal energy consumption is not affected by acute hypercapnic incubation, large rearrangements in all investigated gill transporters at the molecular level could be shown.

Since the protein levels of Na⁺/H⁺ exchanger and Na⁺/K⁺ ATPase follow their respective mRNA patterns, similar increments of Na⁺/HCO₃⁻ cotransporter may be predicted according to the observed higher mRNA levels.

Elevated Na⁺/K⁺ ATPase capacities might influence the energy budget of the eurythermal species.

Literature

Evans, D.H., Piermarini, P.M. & Choe, K.P. (2005). "The multifunctional fish gill: dominant site of gas exchange, osmoregulation, acid-base regulation, and excretion of nitrogenous waste." *Physiological Reviews* **85**(1): 97-177.

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