

THE CHANGE OF BODY COMPOSITION OF INDIGENOUS RAMS ON DIFFERENT BODY WEIGHT AND FEEDING LEVEL

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ABSTRACT

The aim of this research was to investigate the influence of body weight and feeding level on the change of body composition of indigenous ram. This research was expected to establish the efficient feeding level to produce high growth rate and meat quality of ram in tropical environment. Sixteen rams with different body weight (BW) were divided into 2 groups, i.e. light BW (10 + 1.47 kg) and heavy BW (18 + 1.18 kg). The experimental design was Split Plot design with two factors. The main plot was body weight (light and heavy) and the sub plot was level of feeding (FL, 1x maintenance and 1.5 x maintenance). The parameters measured in this experiment were live weight gain and body composition. The results of this study showed that body water, body protein and body fat were significantly higher ($P < 0.01$) in heavy body weight (HBW) than those in light body weight (LBW), and was higher in 1.5xM than those 1xM. Ram of HW and LW had similar change rate of physical composition. Ram given 1.5xM had higher rate of change of body fat and body water than those given feed at maintenance level. It can be concluded that there was no interaction between BW and FL on the change of body water, body protein and body fat. Ram of HBW and LBW had similar rate of change of physical composition. Level of feeding affected body water content, the higher level of feeding resulted in fatter ram, leading to less body water content.

Keywords: Body Weight, Feeding Level, Maintenance, Body Composition

INTRODUCTION

The success of ruminant management is indicated by high body weight gain. To increase the livestock productivity, daily management must be done in a good way. Productivity of ruminant is affected by feed, genetic and environmental factors. Production of meat, the primarily goal of ruminant production was depend on the growth process. The growth phenomenon is the essential process in the livestock and meat industry, both from the standpoint of animal growth and in production of most feedstuffs that ruminants convert to meat. Basic understanding of animal growth has potential for solving problems of efficiency in ruminant production.

Tissues grow and develop chronologically in specific "growth waves." Certain tissues grow and mature before others; growth starts with neural tissue

and proceeds to bone, muscle tissue, and finally adipose tissue. Within each of these tissues, development can be early, medium, or late depend on its location in the body. For example, deposition of fat near the kidney precedes deposition at intermuscular, subcutaneous, and intramuscular sites. Hence, body shape and composition change as an animal matures. Supply of dietary nutrients must be coordinated with this progression to maintain optimum growth rates.

Animal growth involves hyperplasia and hypertrophy of the body cells. The growth pattern follows a sigmoid curve (S curve) (Edey, 1983), consisting of a prepubertal accelerating followed by a linear phase and finally a self-decelerating phase. Each component of the body (muscle, fat and bone) has its own growth curve and live weight change is an integration of all phases. Some researchers reported that muscle and fat productions have a

correlation with age and body weight (Ensminger, 1992; Burton and Reid, 1968). In animals containing less than 31% of fat, the amounts of body components increased linearly with the increasing of body weight; above this concentration of fat, the weights of water and protein increased at decreasing rates and the amounts of fat and energy increased at increasing rates, as body weight increased.

Feed is needed by an animal to fulfill the nutrient requirement for maintenance and production. The first function of feed is to maintain the body condition without developing body tissues and organ (Pond *et al.*, 1995). After the maintenance requirement has been fulfilled, feed is used for growth and other production such as muscle

was expected to establish the efficient feeding level to produce high growth rate of rams in tropical environment.

MATERIALS AND METHODS

Materials

Sixteen thin tailed rams with different body weight (BW) were divided into 2 groups, i.e. light BW (weighed 10 ± 1.47 kg; aged 6 to 7 months) and heavy BW (weighed 18 ± 1.18 kg; aged 9 to 12 months). The rams were placed in individual pens and fed a complete feed containing $\pm 12.6\%$ crude protein of dry matter and 14.66 kJ/g of energy (Table 1).

Table 1. Diet Formulation and its Chemical Composition.

Feedstuff	%	DM	CP	EE	Cfi	NFE	Energy
					----- % DM -----		--kJ/g--
Napier grass	20	89.96	8.31	2.41	43.54	23.78	13.28
Coffee hulls	10	89.35	9.24	2.03	29.22	42.58	16.00
Rice bran	35	92.07	8.15	1.61	23.33	40.03	15.49
Cassava pulp	18	88.54	2.87	1.35	12.74	60.99	13.69
SBM	15	89.88	46.56	2.42	2.61	31.72	16.77
Mineral mix	1	-	-	-	-	-	-
Salt	1	-	-	-	-	-	-
Complete Feed	100	92.11	12.94	1.86	22.48	42.13	14.66

SBM = Soybean Meal; DM = Dry Matter; CP = Crude Protein; EE = Ether Extract; Cfi = Crude fiber; NFE = Nitrogen Free Extract.

growth and fat deposition. Waters cited by Burton and Reid (1968) stated that cattle fed at maintenance or slightly submaintenance level after they had been reared on high nutrition to an age of 9 to 17 months, continued to grow in height and length of skeleton for at least 1 year, during which time body fat was depleted. This observation indicated that certain tissues (bone) have a higher priority claim for nutrients than other tissues on certain stages of growth. Feeding level influences growth rate, fat deposition and carcass composition in every stage of age; animals with different feeding level have different body composition, although they are in the same body weight and breed (Hammond cited by Lawrie, 1995).

The aim of this research was to investigate the influence of body weight and feeding level on the change of body composition in rams. This research

Methods

Experimental Design

The experimental design was "Split Plot Design" with two factors. The main plot was 2 groups of BW (light BW and heavy BW) and the sub plot were two levels of feeding, it was allowed to fulfill 1x maintenance (equal to 2.4% BW) and 1.5 x maintenance (equal to 3.6% BW) requirements.

Experimental Procedures

The study was carried out in 14 weeks with 4 periods, i.e; adaptation period (2 weeks), preliminary period (2 weeks) and feeding treatment period (10 weeks). In order to determine dry matter requirement, every ram were weighed at the beginning of adaptation periode. The feeding level of 1xM was determined by maintaining the animals at zero gain, while the

1.5xM was obtained by multiplying 1xM level feeding with 1.5. The diets from this calculation were given to rams based on DM requirement.

Parameters

The parameters measured in this experiment were live weight gain and body composition. The live weight gain (LWG) measured by subtracting final weight by initial weight of the experimental period. Body composition was measured using Urea Space (US) technique as recommended by Astuti and Sastradipradja (1998). The changes of body composition (water, protein, and fat) were measured in the 2nd, 6th and 8th weeks of treatment period. Urea dilution trials were performed on all 16 rams. Urea space technique procedures were as follow:

A day before the trial, the sheep were weighed to measure their body weight and calculate the metabolic body weight (BW^{0.75}). The dose was 0.65 ml urea solution for every kilogram metabolic body weight. A hundred grams of crystal urea was diluted into 500 ml NaCl to make urea solution.

Before urea dilution being administered into the rams blood, 10 ml blood sample was collected from the jugular vein of each sheep to determine plasma urea concentration. Then, a urea solution was administered using catheter, inserted into a jugular vein. After infusion, the catheter was flushed with 10 ml of NaCl and then removed. The actual quantity of urea injected was determined gravimetrically by weighing syringes before and after injection.

Ten milliliters blood sample was collected from the jugular vein 12 min after the mean of infusion time. The blood sample was than stored in push-up tubes, placed immediately in an ice bath and then centrifuged at 3000 rpm for 10 min. An amount of blood plasma subsample was taken from the sample and frozen at -5 °C for the subsequent determination of plasma urea concentration.

Urea concentration in the blood plasma was measured using Berthelot method with a commercial kit (Bavaria diagnostica, Hamburg, Germany) containing 4 solutions. Before blood plasma being analyzed, preparations of standard solution need to be done, there was R1A made by mixed R2 to R1. Urea standard were made to have 0 mg/dl (100% aquabides), 12.5 mg/dl (aquabides : STD = 3:1), 25 mg/dl (aquabidest : STD = 1:1) and 50 mg/dl (100%

STD). Ten microliters of blood plasma and urea standard was taken by micropipette into the tube. Then, 1000 µl R1A and aquabides were add into each tube and incubate in the water-bath at 37°C for 5 min. Next step were adding 100 µl of R3 into each tube and incubate in the water-bath at 37°C for 5 min. Each sample was then measured for urea determination using spectrophotometer.

Urea space was calculated from the dose of urea N injection, urea space was calculated on both a live weight (LW) and empty body weight (EBW) basis using the following equation (Bartle *et al.*, 1983):

$$US (\%) = \frac{(\text{Dose of urea - N (mg) injected})}{\text{Change in plasma urea - N (mg/100 ml)} \times 10 \times LW}$$

Change in BUN was expressed in mg of urea-N/ 100 ml plasma and body weight as either LW or EBW. Change in BUN was calculated by difference between BUN at 0 and 12 minute after urea injection. Empty body weight was defined as body weight minus the content of alimentary tract. The equation for determining empty body water (EBH₂O) following Rule *et al.* (1986), while for protein (EBPro) and fat (EBFat) were calculated as Panaretto and Till (1963). All parameters, except EBPro (in kg) were expressed s percentage of empty body weight, as follow:

$$EB-H_2O (\%) = 59.1 + 0,22 \times US (\%) - 0.44 LW \dots\dots (1)$$

$$EB-H_2O (kg) = [EB-H_2O (\%) \times EBW (kg)]/100\% \dots\dots (2)$$

$$EB-Pro (kg) = 0.265 \times EB-H_2O (kg) - 0.47 \dots\dots (3)$$

$$EB-Pro (\%) = 100 \times [EB-Pro (kg)/EBW \dots\dots (4)$$

$$EB-Fat (\%) = 98.0 - 1,32 \times EB-H_2O (\%) \dots\dots (5)$$

$$EB-Fat (kg) = [EB-Fat (\%) \times EBW (kg)]/100\% \dots\dots (6)$$

Data Analysis

Data were analyzed using analysis of variance by SAS Institute Inc. (1998). Split plot was used to analyze main effects of body weight, feeding levels and their interactions. Least square means were computed and tested for treatment differences. If there were no differences in two-way interactions (P>0.05), then data were then analyzed for the main effects of weight and feed in one-way anova.

RESULTS AND DISCUSSION

Water Intake and Live Weight Gain

The effect of the treatments on the water intake and live weight gain (LWG) are presented in Table 2.

There was an interaction between body weight and feeding level on LWG ($P < 0.05$).

Heavy sheep with maintenance level of feeding have greater LWG than the light one, and light sheep

showed that CP and energy balance for LW and HW was the same, it was 8.6 g for every 1 MJ of energy intake.

Table 2. Effects of Body Weight and Feeding Level and Their Interactions on Water Intake, CP Intake, Energy Intake and Live Weight Gain of Indigenous Rams.

Parameters	Light body weight		Heavy body weight		Significance		
	1xM	1.5xM	1xM	1.5xM	BW	FL	INT
LWG (g/d)	-0.01	0.00	-0.03	0.05	ns	**	**
CP intake (g/d)	26.86	49.91	57.36	100.53	**	**	**
Energy intake (MJ/d)	3.12	5.79	6.66	11.67	**	**	**
TDN (g/d)	171.91	319.39	367.07	643.37	**	**	**
Water intake (ml/d)	1299	1412	1797	2647	**	*	ns

LWG = Live weight gain; M = maintenance; BW = body weight; FL = feeding levels; INT = interaction between body weight and level of feeding; ** = highly significant ($P < 0.01$); * = significant ($P < 0.05$); ns = non significant ($P > 0.050$).

on the 1.5xM had less LWG than did the heavy one. The LWG were -0.01 and 0.00 g/d for light sheep on the 1xM and 1.5xM, respectively. The heavy sheep had -0.03 and 0.05 g/d LWG at maintenance and 1.5xM, respectively. Interaction between body weight and feeding levels on LWG were influenced by DMI. Tillman *et al.* (1991) informed that LWG was influenced by DMI.

The data of LWG of light and heavy body weight indicated that the feed given to the sheep did not fulfill the nutrient requirement for sheep. Present study was in contrast with feeding level requirement according to Kearn (1982), that sheep with 10 kg of BW would have 25 g/d of LWG if fed at 3.6 % of live weight and sheep with 20 kg BW on the 3% of live weight. In present study, sheep with LBW and HBW needed 3.6% of BW to fulfill maintenance requirement. Kearn (1982) mentioned that nutrient requirement of sheep on 10 kg of body weight on the maintenance level was 26 g and 140 g of CP and TDN, respectively; while nutrient requirement of sheep with 20 kg were 44 g CP and 240 g TDN. This experiment (Table 2) showed that CP intake and TDN of that sheep on 10 and 18 kg of body weight were 26.86 g/d; 171.91 g/d and 57.36 g/d; 367.07 g/d. for maintenance requirement, respectively. This experiment showed that CP and TDN for maintenance requirement of indigenous sheep were higher than those recommended by Kearn (1982). This experiment also

There was no interaction between body weight and feeding level on water intake ($P > 0.05$). Water intake was higher in heavy body weight than those in light body weight ($P > 0.05$), and those in the maintenance feeding level was lower than those in 1.5xM of feeding level ($P > 0.05$). Water intake increased with the increasing of DMI and those in the heavy sheep have higher water intake because of the larger size will make higher body water evaporation than those in the light body weight.

Body Composition

Body water

There was no interaction ($P > 0.05$) between body weight and feeding level on both body water weight and percentage. The body water weight in the second week was higher ($P < 0.01$) in the heavy sheep compared to those in the light sheep, and those in the 1xM was lower ($P < 0.01$) than those in the 1.5xM (Table 3). The same cases occurs in the 6th and 8th week of parameters measured, heavy sheep had higher body water content compared to those in the light sheep ($P < 0.05$). However, it was in contrast with the percentage of body water content from EBW. Increasing in animal age will followed by increasing the percentage of body fat and decreasing the proportion of body water, and slightly decreasing the body protein.

Light sheep have greater ($P<0.01$) body water percentage than did in the heavy sheep; and those in 1.5xM was lower ($P<0.01$) than in 1xM. It means

Body protein

There was no interaction between body weight and feeding level on body protein content in body

Table 3. Effects of Body Weight and Feeding Level and Their Interactions on Body Water, Weekly Change of Body Water and Water Intake of Indigenous Rams.

Parameters	Light weight		Heavy weight		Significance		
	1xM	1.5xM	1xM	1.5xM	BW	FL	INT
EB-H2O							
Week 2 (kg)	3.56	4.13	7.05	8.20	**	**	ns
(%EBW)	58.75	58.70	58.41	58.30	**	*	ns
Week 6 (kg)	3.35	4.01	6.91	8.46	**	*	ns
(%EBW)	58.77	58.71	58.42	58.27	**	**	ns
Week 8 (kg)	3.21	4.15	6.69	8.36	**	**	ns
(%EBW)	58.79	58.70	58.45	58.28	**	**	ns
The change of EB-H2O							
Week 2-6 (kg/d)	-0.22	-0.16	-0.14	0.26	ns	ns	ns
(%/d)	0.01	0.01	0.01	-0.03	ns	ns	ns
Week 6-8 (kg/d)	-0.14	0.19	-0.22	-0.10	ns	ns	ns
(%/d)	0.02	-0.01	0.02	0.01	ns	*	ns
Week 2-8 (kg/d)	-0.35	0.03	-0.36	0.16	ns	*	ns
(%/d)	0.04	0.00	0.04	-0.02	ns	**	ns

M = maintenance; BW = body weight; FL = feeding levels; INT = interaction between body weight and level of feeding; **= highly significant ($P<0.01$); * = significant ($P<0.05$); ns = non significant ($P>0.050$).

that as animal age increased, body water was decreased, confirmed with some study previously reported (Berg and Butterfield, 1976); Tillman *et al.*, 1991).

There were significant differences ($P<0.05$) between feeding level in both body water weight and the percentage. It was considered to be caused by water intake being significantly different ($P<0.05$) between body weight and between feeding level (Table 3). Body water composition was affected by age, nutrition, animal type and decrease with the increasing of body weight (Jesse *et al.*, 1976).

There were no significant differences between body weights and feeding levels in the change of body water weight between 2-6 weeks ($P>0.05$) (Figure 2), ranging from -0.22 to 0.26 kg/d and -0.03 to 0.02% in percentage, respectively. On the other hand, the percentage of the change of body water on 6-8, and 2-8 weeks were higher ($P<0.05$) in 1xM than those in 1.5xM. The percentage of the change of body water was greater ($P<0.01$) in 1.5xM of feeding level of the weight of body water.

composition. The body protein in 2nd, 6th and 8th weeks in the heavy sheep were significantly higher ($P<0.01$) compared to those in the light sheep, both in weight and percentage of body protein (See Table 5). It can be attributed to the fact that CP intake in this experiment were significantly different ($P<0.05$) between body weight. Protein is exist in the muscle and bone, which arrange the body; so heavy body weight would have high body protein weight than those on the light body weight.

The percentage of body protein were higher ($P<0.05$) in those in the heavy sheep compared to those in the light sheep in every week of parameters measured, either between body weight or feeding level. The result of this experiment was in agreement with Soeparno (1994) who stated that most of body composition was affected by body weight and to some extent was affected by age of the animal. The percentage of body protein was in contrast with those cited by Judge *et al.* (1989), that body protein was 16 to 22% of empty body weight.

Table 4. Effects of Body Weight and Feeding Level and Their Interactions on Body Protein, The Change Of Body Protein and CP Intake of Indigenous Rams.

Parameters	Light weight		Heavy weight		Effects		
	1xM	1.5xM	1xM	1.5xM	BW	FL	INT
EB-Pro							
Week 2 (kg)	0.47	0.62	1.39	1.70	**	*	ns
(% EBW)	7.76	8.76	11.56	12.09	**	**	ns
Week 6 (kg)	0.41	0.59	1.36	1.77	**	*	ns
(% EBW)	7.36	8.39	11.47	12.19	**	ns	ns
Week 8 (kg)	0.38	0.63	1.31	1.71	**	**	ns
(% EBW)	6.88	8.79	11.37	12.15	**	**	ns
The change of EB-Pro							
Week 2-6 (kg/d)	0.06	0.04	0.04	-0.07	ns	ns	ns
(%/d)	0.40	0.50	0.09	-0.10	ns	ns	ns
Week 6-8 (kg/d)	-0.03	0.05	-0.06	-0.03	ns	ns	ns
(%/d)	-0.48	0.54	-0.11	-0.04	ns	ns	ns
Week 2-8 (kg/d)	0.18	0.01	-0.10	0.04	ns	*	ns
(%/d)	-0.16	0.04	-0.20	0.06	ns	ns	ns
CP intake (g/d)							
Week 2-6	26.86	49.54	57.36	99.72	**	**	**
Week 6-8	26.86	49.68	57.36	102.10	**	**	**
Week 2-8	26.86	49.91	57.30	100.53	**	**	**

CP = crude protein; M = maintenance; BW = body weight; FL = feeding levels; INT = interaction between body weight and level of feeding; ** = highly significant ($P < 0.01$); * = significant ($P < 0.05$); ns = non significant ($P > 0.050$).

The percentage of body protein in 6th week was not affected by feeding levels. This result was in agreement with Tillman *et al.* (1991) and Soeparno (1994) who have reported that body protein percentage is not affected by age and feed on the stage of maturity.

From the point of view of body protein change, the differences of body protein in every week parameters measured were not followed by the change of body protein and its percentage in the 2-6, 6-8 and 2-8 weeks measurement ($P > 0.05$), ranged from -0.10 to 0.18 kg/d. This was in agreement with Tillman *et al.* (1991) that body protein was tended to be constant on the stage of maturity and was not affected by feed. It was proved by CP intake in 2-6, 6-8 and 2-8 week were significantly different ($P < 0.05$) in body weight and feeding level. However, body protein is affected by genetic, because protein exists in the muscle and bone, and those were arranged the body. The research conducted by Garrett *et al.* (1959) reported that sheep fed on maintenance level had 16.2% body

protein and increased with the increasing of animal size (and gain).

Body fat

Observations on the body fat obtained that the change of body fat and energy intake as shown in Table 5.

There was no interaction ($P > 0.05$) between body weight and feeding level on body fat in every week of measurement. The percentage of body fat was higher ($P < 0.01$) in the heavy sheep than those in light sheep. The body fat was higher ($P < 0.01$) in 1.5xM of feeding levels than that in 1xM. The average of body fat in this experiment was higher than the research reported by Garrett *et al.* (1959) that sheep body fat at maintenance level of feeding was 18.7% of body weight, and increased with body weight.

The changes of body fat were significantly different ($P < 0.01$) in every week of measurement, except from 2nd to 6th weeks of data collection period ($P < 0.05$). Energy intake were higher

Table 5. Effects of Body Weight and Feeding Level and Their Interactions on Body Fat, Weekly Change of Body Fat and Energy Intake of Indigenous Ram.

Parameters	Light weight		Heavy weight		Effects		
	1xM	1.5xM	1xM	1.5xM	BW	FL	INT
Body fat							
Week 2 (kg)	1.24	1.45	2.53	2.96	**	*	ns
(%)	20.44	20.52	20.90	21.05	**	**	ns
Week 6 (kg)	1.19	1.42	2.47	3.07	**	**	ns
(%)	20.43	20.51	20.88	21.08	**	**	ns
Week 8 (kg)	1.12	1.34	2.39	3.03	**	**	ns
(%)	20.40	20.52	20.85	21.07	**	**	ns
The change of Body fat (kg/d)							
Week 2-6 (kg/d)	0.05	0.03	0.05	-0.10	ns	ns	ns
(% EBW)	0.02	0.01	0.02	-0.03	ns	ns	ns
Week 6-8 (kg/d)	-0.08	0.04	-0.08	-0.08	ns	*	ns
(% EBW)	-0.03	0.01	-0.03	-0.01	ns	*	ns
Week 2-8 (kg/d)	-0.10	0.01	-0.14	0.06	ns	*	ns
(% EBW)	-0.04	0.00	-0.05	0.02	ns	*	ns
Energy intake (MJ/d)							
Week 2-6	3.12	5.75	6.66	11.57	**	**	**
Week 6-8	3.12	5.77	6.66	11.85	**	**	**
Week 2-8	3.12	5.79	6.66	11.68	**	**	**

BW = main effect of body weight; FL = effect level of feeding; INT = interaction between body weight and level of feeding; M = maintenance; **= significant at $P < 0.01$; * = significant at $P < 0.05$; ns = non significant at $P > 0.05$).

($P < 0.01$) in heavy sheep compared to those in light sheep, and was higher in 1.5xM ($P < 0.01$) of feeding levels. High energy intake resulted in reduction of body protein and increase of body fat (Crouse *et al.* cited by Soeparno, 1994). These results showed that sheep, with heavy weight and light weight given 1.5xM feeding level, needed 6 weeks to increase body fat, from week 2 to week 8 of feeding treatment period.

CONCLUSION

Based on the results in this study, it can be concluded that there was no interaction between BW and FL on the change of body water, body protein and body fat. Rams given 1.5xM feeding level had higher rate of change of body fat and body water than those given feed at maintenance level. Both light and heavy rams lose body weight when they were fed 2.4% body weight. It is recommended that rams should be offered diet 3.6% of body weight, with CP

and energy balance being 8.6 g for every 1 MJ of gross energy intake to meet the maintenance requirement.

REFERENCES

- Anggorodi, R. 1994. Ilmu Makanan Ternak Umum. 5th Ed, PT Gramedia, Jakarta.
- Astuti, D.A. and D. Sastradipradja. 1998. Evaluation of body composition using urea dilution and slaughter technique of growing priangan sheep. *Media Veteriner*. 6(3):5-9
- Bartle, S. J., J. R. Males and R. L. Preston. 1983. Evaluation of urea dilution as an estimator of body composition in mature cows. *J. Anim. Sci.* 60: 410-418
- Berg, R. T. and R. M. Butterfield. 1976. *New Concepts of Cattle Growth*. 1st Ed, Sydney University Press, Sydney.
- Burton, J. H. and J.T. Reid. 1968. Interrelationship among energy input, body size, age and body

- composition of sheep. *J. Nutrition*. 97: 517-524
- Edey, T.N. 1983. *Tropical Sheep and Goat Production* Freeman and Co., San Fransisco.
- Ensminger, M.C. 1992. *Beef Cattle Science*. Animal Agriculture Series. 7th Ed, Interstate Printer and Publisher Inc, Danville.
- Garrett, W. N., J. H. Mayer, and G. P. Lofgreen. 1959. The comparative energy requirements of sheep and cattle for maintenance and gain. *J. Anim. Sci.* 18: 528-547
- Jesse, G.W., G.B. Thomson, J.L. Clark, H.B. Hendrick and K.G. Weimer. 1976. Effects of ration energy and slaughter weight on composition of empty body and carcass gain of beef. *J. Anim. Sci.* 53 : 418
- Judge, M.D., E.D. Aberle, J.C. Forrest, H.B. Hedrick and R.A. Merkel. 1989. *Principles of Meat Science*. Kendall/Hunt Publishing Company, Dubuque, Iowa.
- Kearl, L. C. 1982. *Nutrition Requirements of Ruminants in Developing Countries*. International Feedstuff Utah Agriculture Experiment Station. 1st Ed, Utah State University, Logan.
- Lawrie, R.A. 1995. *Meat Science*. Low Temperature Research Station, Nottingham University. Cambridge.
- Panaretto, B.A. and A.R. Till. 1963. Body Composition In vivo (II). *Aust. J. Agric. Res.* 14: 926-943.
- Pond, W.G, D.C. Church and K.R. Pond. 1995. *Basic Animal Nutrition and Feeding*. 4th, John Wiley and Sons, Inc., New York.
- Rule, D. C., R. N. Arnold, E. J. Hentges and D. C. Beitz. 1986. Evaluation of urea dilution as a technique for estimating body composition of beef steers in vivo” validation of published equations and comparison with chemical composition. *J. Anim. Sci.* 64:1935-1948
- Soeparno. 1994. *Ilmu dan Teknologi Daging*. 1st Ed, Gadjah Mada University Press, Yogyakarta.
- Tillman, A. D., H. Hartadi, S. Reksohadiprojo, S. Prawirokusumo and S. Lebdoesoekojo. 1991. *Ilmu Makanan Ternak Dasar*. 5th Ed, Gadjah Mada University Press, Yogyakarta.