REVIEW

Review: Infectious Diseases and Coagulation Disorders

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Infection, both bacterial and nonbacterial, may be associated with coagulation disorders, resulting in disseminated intravascular coagulation and multiorgan failure. In the last few decades a series of in vivo and in vitro studies has provided more insight into the pathogenetic mechanisms and the role of cytokines in these processes. Because of the growing interest in this field, the complexity of the subject, and the fact that many physicians must deal with a variety of infections, current data are reviewed on the association between infectious diseases and the coagulation system. Novel therapeutic intervention strategies that will probably become available in the near future are mentioned, along with those of special interest for infectious disorders for which only supportive care can be given.

Systemic infections may be complicated by activation of the coagulation cascade, varying from subclinical activation, which is indicated by a rise in laboratory markers for thrombin and fibrin generation, to fulminant disseminated intravascular coagulation (DIC) with the formation of microvascular thrombi in various organs [1]. Bleeding, thrombosis, or both may be the presenting clinical features. DIC may contribute to multiorgan failure (MOF) and is associated with a high mortality in both bacterial and nonbacterial disease [2, 3].

Studies of gram-negative sepsis in humans and experimental animals have shown that cytokines play a pivotal role in the activation and regulation of the coagulation cascade, although the interactions are complicated, and the effects are time-dependent and transient [4]. Activation of the coagulation system has also been documented for nonbacterial pathogens (i.e., viruses causing hemorrhagic fevers [HFs] [5, 6], protozoa [malaria] [7, 8], and fungi [9]).

Since no specific effective treatment is yet available for DIC, therapy focuses on treatment of the underlying disorder (e.g., antibiotics for bacterial infection). For many infectious diseases, such as viral HF, causal therapy is not available, and only supportive care can be provided. To improve therapy and

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supportive care, a better understanding of the pathogenesis of bleeding and thrombotic complications is needed. Novel therapeutic agents that intervene in the coagulation and cytokine cascades will become available in the near future and may have a positive clinical effect, especially for infections for which only supportive care can be given.

Because of the growing interest in this field, the complexity of the subject, and the fact that many physicians deal with a variety of infections, we will review current data on the association between infectious diseases and the coagulation system.

Clinical Aspects of Hemostasis in Bacterial and Viral Infections

Viral and bacterial infections may influence hemostasis and can lead to thrombohemorrhagic complications or syndromes such as DIC, hemolytic uremic syndrome (HUS), thrombotic thrombocytopenic purpura (TTP), or vasculitis. Symptoms and signs may be dominated by bleeding, thrombosis, or both.

DIC is an acquired disorder in which the hemostatic system is activated, resulting in activation of platelets and the conversion of fibrinogen to fibrin [3, 10]. This may lead to generalized microvascular thrombosis and MOF and to life-threatening hemorrhage due to consumption of coagulation factors and activation of the fibrinolytic system. DIC as a consumptive coagulopathy, with consumption of both platelets and clotting factors, must be distinguished from the nonconsumptive coagulopathies, such as HUS and TTP, characterized by the consumption of platelets but not clotting factors. HUS and TTP are regarded as variants of a single syndrome characterized by thrombocytopenia, hemolytic anemia, fever, renal abnormalities, and neurologic disturbances. Endothelial injury is consid-

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ered the primary event, which, as shown in recent studies, may lead to excessive release of extremely large polymers of von Willebrand factor that cannot be processed to smaller forms because of deficiency of von Willebrand factor-cleaving protease [11–14].

Vasculitis, which may be triggered by infection, is characterized by local or more generalized vascular changes, resulting from infarction secondary to occlusion by thrombi of the lumen of small blood vessels in the upper part of the dermis. Vascular occlusion may lead to ischemic tissue injury due to local vascular occlusion or bleeding due to local tissue damage [15, 16].

None of these syndromes (DIC, MOF, HUS, TTP, or vasculitis) are specific for a certain pathogen, and their occurrence depends on factors such as virulence of the pathogen, the patient's prior condition, source and size of the inoculum, and availability of antimicrobial treatment. For example, these coagulopathic syndromes are encountered more frequently in gram-negative infection than in gram-positive and other nonbacterial infections. Vasculitis (i.e., cytomegalovirus [CMV] vasculitis) occurs in human immunodeficiency virus (HIV)– immunocompromised persons but is rare in persons not immunodepressed, underlining the role of the host's status. There is a relation between the size of the inoculum and the occurrence of the sepsis syndrome, as has been shown in an experimental gram-negative sepsis model in baboons [17].

In some cases of MOF, bleeding and (microvascular) thrombosis may be coexisting features, making it impossible to determine the principal defect (thrombosis or bleeding). Taking these considerations into account, the classification of clinical entities according to bleeding or thrombosis will be the basis for discussion, although the complexity and overlap in clinical presentation must be kept in mind.

Hemorrhage. Hemorrhage may occur as a single clinical phenomenon or may be part of a more complex derangement of the coagulation cascade due to DIC or septic vasculitis, as in gram-negative bacterial sepsis (i.e., meningococcemia [1, 3, 16]). Severity may vary from local defects in hemostasis with oozing from arterial or venous puncture sites to more general complications, such as petechiae, purpura, ecchymosis, gut bleeding, hemoptysis, or even MOF due to adrenal bleeding (as in Waterhouse-Friderichsen syndrome). The clinical signs are not pathogen-specific and in general depend on the severity of infection. In specific infections, such as viral HF, bleeding

complications are prominent [18–20]. Among the viral HFs, Dengue [18–20], Marburg [21, 22], and Ebola [23, 24] are the most important. Dengue is the most prevalent (table 1).

Bacterial and viral infections may result in a vasculitis-like syndrome with either bleeding manifestations or ischemic injury [15, 16, 25]. Vasculitis is well documented in CMV infection [26, 27], occurring predominantly in the vasculature of the gastrointestinal tract, where it causes colitis [28, 29]; the central nervous system, where it causes cerebral infarction [30, 31]; and the skin, where it results in petechiae, purpura papules, localized ulcers, or a diffuse maculopapular eruption [32].

HIV infection may be accompanied by vasculitis syndromes, such as polyarteritis nodosa, Henoch-Schönlein purpura, and leukocytoclastic vasculitis [33–36]. Hepatitis B and C infection may cause polyarteritis-like vasculitis [37–39]. Parvovirus B19 has been suggested to be associated with vasculitis-like syndromes, including Kawasaki disease, polyarteritis nodosa, and Wegener's granulomatosis [40–42].

Leptospirosis, especially Weil's syndrome, may present with hemoptysis, epistaxis, intestinal bleeding, adrenal bleeding, hematuria, and even subarachnoid hemorrhage [43]. The pathogenesis may be either primary activation of coagulation or diffuse vasculitis, resulting in bleeding or ischemia of the vascularized tissue. TTP, with bleeding as the presenting symptom, may also occur in the course of Weil's syndrome [44]. In other viral and bacterial infections associated with TTP or HUS, bleeding also is often the prominent and presenting symptom [45–47].

Thrombosis and hemorrhage: DIC. Viral and bacterial infections may theoretically result in local thromboembolic disease, that is, deep venous thrombosis or pulmonary embolism. In a thromboembolic prevention study of low-dose subcutaneous standard heparin for hospitalized patients with infectious diseases, morbidity due to thromboembolic disease was significantly reduced in the heparin group, compared with the group receiving no prophylaxis. There was, however, no beneficial effect of prophylaxis on mortality due to thromboembolic complications [48]. In chronic viral diseases, such as CMV or HIV infection, the risk of thromboembolic complications is relatively low [49, 50].

MOF is a clinical entity characterized by generalized microvascular thrombosis that may develop as part of the DIC syndrome, especially in bacterial gram-negative sepsis but also in

 Table 1.
 Viral infections hallmarked by hemorrhage or viral hemorrhagic fevers (HFs).

Virus	Geographic distribution	Source of infection
Dengue HF	Southeast Asia, Caribbean, Central/South America, China	Mosquito
Chikungunya	Southeast Asia	Mosquito
Ebola	Zaire, Sudan	Unknown
Marburg HF	Zimbabwe, Kenya, Uganda	Unknown
Lassa fever	West Africa	Rodent
Yellow fever	South America, Africa	Mosquito
Omsk HF	Former Soviet Union	Tick
Hantaan	Central Europe, former Soviet Union, Korea, Japan, eastern China	Rodent

severe viral infections. Viral HF (table 1) is complicated by DIC in the most severe cases [18–24]. DIC is not frequently encountered in other viral infections [51, 52] but has been found in cases of rotavirus [53, 54], varicella, rubella, rubeola, and influenza infections [55–61].

TTP and HUS, triggered by a viral or bacterial infection [45–47], frequently lead to bleeding symptoms, as has been discussed, but platelet and fibrin thrombi may also be generated in various organs, leading to prominent symptoms with organ dysfunction. The variety and complexity of the clinical presentation of the coagulopathy syndromes confront the clinician with the difficult questions of whether, when, and how to pro-

vide optimal supportive care to combat the coagulation disorders in addition to giving antimicrobial therapy. In making this decision, he or she will probably be guided by the clinically most pronounced presenting symptom.

Principles of Hemostasis, Coagulation, and Fibrinolysis: General Aspects

The hemostatic mechanism consists of primary hemostasis, coagulation, and fibrinolysis (figure 1). Primary hemostasis is maintained by the adhesion and aggregation of platelets to form a hemostatic plug that is stabilized by fibrin strands [62, 63].

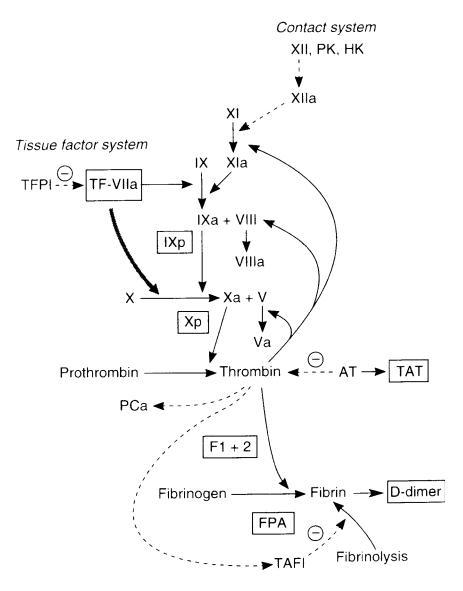


Figure 1. Model of hypothetical coagulation cascade. Products of coagulation activation are shown in squares: IXp, factor IX activation peptide; Xp, factor X activation peptide; F1 + 2, prothrombin fragment 1 + 2; FPA, fibrinopeptide A; D-dimer, fibrin split product; TAT, thrombin-antithrombin complex. Natural inhibitors shown: TFPI, tissue factor pathway inhibitor; AT, antithrombin; PCa, activated protein C (which cleaves activated factors V and VIII). Coagulation proteins are given in roman numerals: PK, prekallikrein; HK, high–molecular weight kininogen; TAFI, recently identified "thrombin activatable fibrinolytic inhibitor."

Coagulation, the second hemostatic defense mechanism, is generated by a series of linked coagulation protease-zymogen reactions, ultimately resulting in the formation of fibrin. In the current concept of coagulation, thrombin generation is induced by the assembly of tissue factor (TF) VIIa complex, the socalled extrinsic route of coagulation.

Coagulation is counteracted by different inhibitory mechanisms. A first mechanism is made up of the circulating inhibitors of blood coagulation, that is, antithrombin, proteins C and S, and TF pathway inhibitor (TFPI). A second inhibitory mechanism consists of the endothelium-bound modulators heparin sulfate and thrombomodulin, which facilitate the inhibitory activity of antithrombin and protein C, respectively. The third mechanism, the fibrinolytic system, is activated by tissue plasminogen activator (tPA) and urokinase after their synthesis and release from the endothelial cell system. These activators initiate the conversion of plasminogen to plasmin, which hydrolyzes polymerized fibrin strands into soluble fibrin degradation products. Infection may lead to an imbalance between platelet function and the regulatory mechanisms of the coagulation cascade and fibrinolysis, resulting in bleeding, thrombosis, or both.

Pathophysiologic Mechanisms of Hemostasis in Bacterial and Viral Infection

On the basis of data from experimental studies that have revealed the complexity of the interactions among infectious pathogens, cytokines, effector cells, and the coagulation system, we will discuss current insights into the pathogenesis of abnormal hemostasis in infectious diseases. Cytokines are thought to play an essential role in activation of the coagulation system during viral and bacterial infections [64, 65].

Coagulation Activation

TF pathway. The TF pathway (extrinsic route) is the main route for activation of the coagulation cascade in sepsis. Endotoxins, lipopolysaccharide constituents of the outer membrane of gram-negative microorganisms, play a pivotal role in the development of the gram-negative sepsis syndrome [66]. Levels of circulating endotoxin are prognostic markers for the clinical outcome of the septic syndrome [67, 68]. Injection of endotoxin or tumor necrosis factor (TNF)- α into healthy volunteers results in activation of the coagulation system via activation of the TF pathway [64, 69–71]. TNF- α injected into volunteers initiates the release of interleukin (IL)-6 and IL-8, which is followed by an increase in the levels of circulating markers of thrombin and fibrin generation (prothrombin fragments [F1 + F2], thrombin-antithrombin complexes [TAT], fibrinopeptide A) [17, 64, 72]. This activation is preceded by a primary but transient activation of fibrinolysis; in contrast, after activation of the coagulation cascade, activation of fibrinolysis is secondary [52]. The net effect of an injection of endotoxin or TNF- α is a procoagulant state. In this experimental model, coagulation and fibrinolysis seem to be activated independently, which may lead to an imbalance between activation of coagulation and regulation (activation/inhibition) of fibrinolysis [73]. The same imbalance has been observed in baboons after injection of IL-1 [74]. Administration of IL-6 into patients with renal cell carcinoma also results in activation of coagulation, as reflected by a rise in the concentration of F1 + F2 and TAT complexes, but here fibrinolysis is not affected [75].

TNF- α induces the expression of TF on monocytes [76] and endothelial cells [77–79]. In chimpanzees and baboons, antibodies directed against TF or factor VIIa or treatment with TFPI prevents activation of the common pathway [71, 80–84], whereas antibodies directed against factor XII do not prevent activation [85, 86], emphasizing the role of the TF pathway. IL-10 can inhibit the coagulant response during systemic infection by influencing the expression of TF on monocytes [87, 88].

As mentioned above, TNF is not an essential factor in the mediation of activation of coagulation in sepsis. This has become evident from studies of volunteers treated with anti-TNF in which endotoxin-induced activation of coagulation did not change [89, 90]. Surprisingly, administration of IL-1ra, a major natural inhibitor of IL-1, results in inhibition of both the coagulation cascade and fibrinolysis in baboons with lethal bacteremia and patients with the sepsis syndrome [74, 91]. Treatment of patients with IL1-ra is associated with decreases in the concentrations of TAT and plasmin-a2 antiplasmin (PAP) complexes and tPA and plasminogen activator inhibitor (PAI)-1 [91]. Treating chimpanzees with anti-IL-6 after the administration of low-dose endotoxin prevents activation of the coagulation cascade but does not affect fibrinolysis [92]. In conclusion, activation of the TF pathway after endotoxin release is largely, although not exclusively, mediated and regulated by cytokines.

Contact activation. Although the factor XII pathway does not seem to play an essential role in activation of the coagulation cascade in experimental sepsis in baboons [85, 86], blocking of the contact activation system by the administration of monoclonal antibodies against factor XIIa could prevent lethal hypotension. This effect is most probably mediated by the generation of kinins, such as bradykinin [85]. Moreover, the contact system seems to play an independent role in the activation of the fibrinolytic system [93]. However, the importance of this route in sepsis has not yet been completely clarified (table 2).

Endothelial Cell and TF Expression

The role of endothelial cells seems to be crucial in the development of shock and activation of coagulation [94–96]. Endothelial cell injury is a common feature of viral infection and can alter hemostasis in a direct or indirect manner. The en-

Viruses		
Dengue virus	Herpes simplex virus	
Ebola virus	Poliovirus	
Marburg virus	Adenovirus	
Hantaan virus	Parainfluenza virus	
Cytomegalovirus	Echovirus	
Measles	Human immunodeficiency virus	
Human T cell leukemia virus type 1	Mumps	

Table 2.Viruses infecting endothelial cells.

dothelial cell can be directly infected by different viruses [97–99], for example, herpes simplex, adenovirus, parainfluenzavirus, poliovirus, echovirus, measles, mumps [100], CMV [101], human T cell leukemia virus type 1 [102], and HIV [103]. The ability to infect endothelial cells has also been demonstrated in HFs caused by Dengue, Marburg, Ebola, Hantaan, and Lassa viruses [104]. Such infections may result in a procoagulant state, mainly by inducing TF expression on the endothelial surface [105–107], probably mediated by cytokines such as IL-1 [105–112], TNF- α , and IL-6 [113–115].

An additional cause of enhanced coagulability may be prothrombinase complex assembly on the CMV surface [116]. This observation suggests that the CMV surface contains the necessary procoagulant phospholipids for assembly of the coagulation enzyme complex leading to thrombin generation. A similar observation has been made for HIV [117]. However not all viral infections affecting endothelial cells result in activation of coagulation, which may indicate that activation of endothelium is one factor in a multifactorial process (figure 2).

Fibrinolysis

The process of fibrinolysis involves the General aspects. enzymatic cascade, which helps to break down cross-linked fibrin molecules. Fibrinolysis may be activated primarily-and thus independently of activation of the coagulation cascade-or secondarily in response to fibrin formation. If fibrinolysis is not balanced in time and scope, either bleeding or thrombosis may ensue. After the injection of TNF into healthy volunteers, activation of the coagulation cascade is preceded by a transient activation of fibrinolysis, which is reflected by increased circulating levels of tPA and urokinase plasminogen activator, followed by an increase in PAI-I, suppressing fibrinolysis [64, 72, 118, 119]. Coagulation activation triggers a secondary activation of fibrinolysis [52], which is rapidly shut off by the release of relatively large amounts of PAI-I, such that the net effect of the injection of a bolus of endotoxin is a procoagulant state.

Infusion of IL-1 α into baboons also elicits an early and tran-

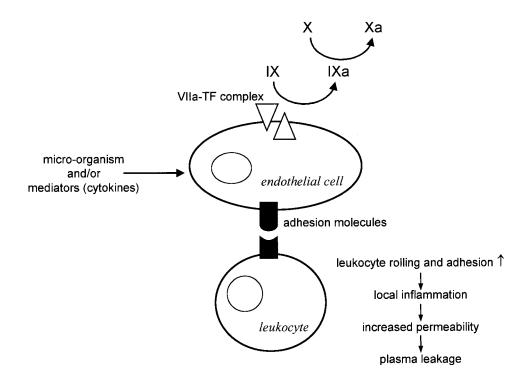


Figure 2. Endothelial cell in infection. During infection, endothelial cell may change to procoagulant state by expressing tissue factor on membrane. Tissue factor is main inducer of extrinsic pathway of blood coagulation. Increased endothelial permeability resulting in plasma leakage may be the consequence of local or more generalized inflammatory response, with increased leukocyte adherence to endothelial cell. This mechanism is triggered by up-regulation of endothelial cell adhesion molecules (i.e., intercellular adhesion molecule 1, vascular cell adhesion molecule 1, Eselectin, P-selectin).

sient activation of fibrinolysis [74] followed by the production of PAI-1, leading to a balance. Fibrinolytic activation is followed by activation of coagulation, thus showing the same imbalance between coagulation and fibrinolysis observed after the administration of TNF- α . This procoagulant imbalance is also present in DIC, where the net effect will be a tendency toward diffuse bleeding and microvascular thrombosis.

Treatment with anti-TNF in the human and chimpanzee sepsis models inhibits the fibrinolytic system, as reflected by the absence of a rise in tPA, PAI-1, and PAP complexes [89, 90, 120], whereas coagulation is not affected, showing that TNF plays an important role in regulation of the fibrinolytic response. Administration of IL-1ra into baboons with lethal bacteremia or patients with sepsis inhibits both coagulation and fibrinolysis [74, 91], as indicated by decreased concentrations of TAT and PAP complexes and tPA and PAI-1 [91]. Treating chimpanzees with anti–IL-6 after the administration of lowdose endotoxin prevents activation of the coagulation cascade and does not affect fibrinolysis [91].

Protein C/S system. During sepsis, the protein C/S system is down-regulated [121]. After the release of TNF- α , thrombomodulin is also down-regulated, resulting in a further decrease in protein C activity, thereby enhancing the procoagulant state. Some viruses can induce specific changes in the coagulation inhibitory system. During the course of HIV infection, the protein C/S system may be impaired as a result of an acquired protein S deficiency, the pathogenesis of which is not yet clarified [122-125]. In children, protein S deficiency seems to correlate with the duration of HIV infection [122], but such a relationship has not been found in adults [123]. Increased plasma concentrations of the C4b-binding protein, an acutephase protein that binds protein S, may result in decreased levels of free protein S. Antiphospholipid antibodies, which may be present in persons infected with HIV, might interfere with the protein C-protein S complex and diminish its activity [123, 126-129]. In patients with Dengue HF, we also found decreased levels of both protein C and protein S activity (current authors, unpublished data). As in Dengue infection, HIV can affect the endothelial cell; hypothetically, this could lead to decreased production of protein S.

Thrombocytopenia

Thrombocytopenia is seen in the course of many viral infections but is only occasionally serious enough to lead to hemostatic impairment and bleeding complications [130]. It is assumed that thrombocytopenia is mainly immune mediated [131]. The mechanism is decreased thrombopoiesis [132, 133], increased platelet consumption [134], or a combination of both. Direct interaction of the virus with platelets [5, 135, 136] may lead to thrombocytopenia or thrombocytopathy. Endothelial injury by the virus [137] may lead to increased adherence and consumption of platelets [138]. Viral infections that have been associated with thrombocytopenia are mumps [139, 140], rubella [141–143], rubeola [144], varicella [145–147], disseminated herpes simplex [148], CMV infection [149, 150], infectious mononucleosis [151–155], Hantaan virus infection [156, 157], Dengue HF [5, 135, 158], Crimean-Congo HF [159], and Marburg HF [160]. Dengue fever is associated with thrombocytopenia even in mild and uncomplicated cases [5]; therefore, thrombocytopenia cannot be the only explanation for the occurrence of bleeding.

Soluble Adhesion Molecules

The change of the endothelial cell from a resting to a procoagulant state may be associated with expression of endothelial surface adhesion molecules [112, 161, 162]. These molecules, that is, the intercellular adhesion molecule, the vascular cell adhesion molecule, E-selectin, and the von Willebrand factor, play an essential role in the binding of leukocytes, resulting in a local inflammatory response, endothelial cell damage, and subsequent plasma leakage and shock [163-166]. Vasculitis also occurs and has been documented in association with such viruses as CMV and HIV [26, 167, 168]. The finding of increased plasma concentrations of these endothelial surface adhesion molecules is thought to reflect the level of activation and perhaps damage of the endothelial cell. Measuring circulating endothelial cells by immunofluorescence or immunomagnetic separation may provide additional information about the activation of vascular endothelium [169-171].

Conclusions

Infectious diseases are often accompanied by activation of coagulation. Although direct interactions between the infectious agent and the coagulation system occur, cytokines are believed to be important mediators in this process. During systemic gram-negative and gram-positive bacterial infections, activation of coagulation is mediated via the extrinsic TF pathway. Experimental studies suggest that, as a rule, coagulation and fibrinolysis occur independently of one another, and the overall result is usually a procoagulant tendency. The latter may result in DIC with microvascular thrombosis and organ failure. Bleeding may result from consumption of platelets and clotting proteins in traumatized tissues.

Although this scenario has been derived largely from studies with purified endotoxin or gram-negative bacteria, we expect similar alterations to occur in gram-positive and in specific viral infections, because some of the key intermediate cytokines (TNF, IL-1, IL-6) are involved. It should be noted, however, that the mechanisms by which viruses presumably induce coagulation in vivo are still speculative, since only in vitro data are available. The critical cellular elements involved in these reactions may not be the same for bacterial and viral infections, but we speculate that the vascular endothelium is a main target. Endothelial cells may turn into a procoagulant state either by stimulation of cytokines in concert with circulating blood cells, such as lymphocytes or platelets, or by direct infection (viruses) of endothelial cells.

Bleeding in infectious disease is most likely a multifactorial process resulting from a combination of thrombocytopenia, consumption of clotting factors, (local) hyperfibrinolysis, and vascular damage or leakage. In addition, immunologically mediated vasculitis may contribute to bleeding in specific infections.

The clinical consequences of chronic viral infections (e.g., CMV and HIV) for the development of thrombotic complications, vasculitis, and atherosclerosis are of great interest. One could hypothesize that since CMV infection may lead to transformation of the endothelial cell to a procoagulant state but may also induce vasculitis and atherosclerosis [172], there could well be a common pathway in the pathophysiologic mechanism of these physiologic and anatomic entities. The role of other pathogens, such as *Chlamydia pneumoniae* that have the potency to affect the endothelium and contribute to the pathogenesis of atherosclerosis is of interest [173–175].

Knowledge of the underlying mechanisms leading to thrombosis or bleeding is fundamental for the development of therapeutic strategies. In gram-negative infections, insight into the important roles of endotoxin and specific cytokines has already led to clinical therapeutic trials with selective inhibitors of the TF pathway (monoclonal antibodies, Fab fragments, modified factor VIIa, nematode anticoagulant protein, TFPI) [176, 177]. Studies with anti-TF antibodies and TFPI in primates have shown that, in addition to inhibition of coagulation activity, these agents may have significant anti-inflammatory properties and may markedly reduce mortality in otherwise lethal infections [178, 179].

Given the potential role that endothelial TF may play in some of the thrombohemorrhagic complications of viral disease, we expect that therapeutic intervention at the TF level or aimed at one of the critical cytokines that mediate its cellular expression may potentially favorably alter the clinical course of these infections. Many issues remain to be answered, and thus there is an urgent need for more clinical and experimental studies, particularly with respect to the relationship between viral infections and the mechanisms leading to bleeding/DIC. Needless to say, treatment of the causal infectious agent remains the cornerstone of therapy.

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References

- Levi M, ten Cate H, van der Poll T, van Deventer SJH. Pathogenesis of disseminated intravascular coagulation in sepsis. JAMA 1993; 270:975–9.
- Bauer KA, Weitz JI. Laboratory markers of coagulation and fibrinolysis. In: Colman RW, Hirsh J, Marder VJ, Salzman EW, eds. Hemostasis and thrombosis: basic principles and clinical practice, 3rd ed. Philadelphia: JB Lippincott 1994:1197–210.
- ten Cate H, Brandjes DPM, Wolters HJ, van Deventer SJH. Disseminated intravascular coagulation: pathophysiology, diagnosis and treatment. New Horiz 1993; 1:312–23.
- ten Cate JW, van der Poll T, Levi M, ten Cate H, van Deventer SJH. Cytokines: triggers of clinical thrombotic disease. Thromb Haemost 1997;78:415–9.
- Bhamarapravati N. Hemostatic defects in Dengue hemorrhagic fever. Rev Infect Dis 1989; 11(suppl):S826–9.
- Heller MV, Marta RF, Sturk A, et al. Early markers of blood coagulation and fibrinolysis activation in Argentine hemorrhagic fever. Thromb Haemost 1995; 73:368–73.
- Clemens R, Pramoolsinsap C, Lorenz R, et al. Activation of the coagulation cascade in severe falciparum malaria through the intrinsic pathway. Br J Haematol 1994;87:100–5.
- Mohanty D, Ghosh K, Nandwani SK, et al. Fibrinolysis, inhibitors of blood coagulation and monocyte derived coagulant activity in acute malaria. Am J Hematol 1997; 54:23–9.
- Fera G, Semararo N, De Mitrio V, Schiraldi O. Disseminated intravascular coagulation associated with disseminated cryptococcosis in a patient with acquired immunodeficiency syndrome. Infection 1993; 21:171–3.
- ten Cate H, Levi M, Biemond BJ, et al. Markers of hemostatic activation in experimental endotoxemia: implications for therapy. In: Müller-Berghaus G, Madlener K, Blombäck M, ten Cate JW, eds. DIC. Pathogenesis, diagnosis and therapy of disseminated intravascular fibrin formation. Amsterdam: Excerpta Medica, **1993**:81–9.
- Furlan M, Robles R, Galbusera M, et al. Von Willebrand factor cleaving protease in thrombotic thrombocytopenic purpura and the hemolytic uremic syndrome. N Engl J Med 1998; 339:1578–84.
- Tsai HM, Chun-Yet Lian E. Antibodies to von Willebrand factor cleaving protease in acute thrombotic thrombocytopenic purpura. N Engl J med 1998; 339:1585–94.
- Moake JL. Moschcowitz, multimers, and metalloprotease [editorial]. N Engl J Med 1998; 339:1629–31.
- Moake JL, Eisenstaedt RS. Thrombotic thhrombocytopenic purpura and the hemolytic uremic syndrome. In: Colman RW, Hirsh J, Marder VJ, Salzman EW, eds. Hemostasis and thrombosis: basic principles and clinical practice. 3rd ed. Philadelphia: JB Lippincott 1994:1064–75.
- Lie JT. Vasculitis associated with infectious agents. Curr Opin Rheumatol 1996;8:26–9.
- Ackerman AB, Chongchitnant N, Sanchez J, Guo Y, et al. Inflammatory diseases. In: Histologic diagnosis of inflammatory skin diseases. 2nd ed. Baltimore: Williams & Wilkins 1997:170–786.
- De Boer JP, Creasy AA, Chang A, et al. Activation patterns of coagulation and fibrinolysis in baboons following infusion with lethal or sublethal dose of *Escherichia coli*. Circ Shock **1993**; 39:59–67.
- Hayes EB, Gubler DJ. Dengue and dengue hemorrhagic fever. Pediatr Infect Dis J 1992; 11:311–7.
- Sumarmo, Wulur H, Jahja E, Gubler DJ, Suharyono W, Sorensen K. Clinical observations on virologically confirmed fatal dengue infections in Jakarta, Indonesia. Bull World Health Organ 1983;61:693–701.
- Kuberski T, Rosen L, Reed D, Mataika J. Clinical and laboratory observations on patients with primary and secondary dengue type I infections with hemorrhagic manifestations in Fiji. Am J Trop Med Hyg 1977;26: 775–83.
- 21. Egbring R, Slenczka W, Baltzer G. Clinical manifestations and mechanism of the hemorrhagic diathesis in Marburg viral disease. In: Martini GA,

Siegert R, eds. Marburg virus disease. New York: Springer Verlag, **1971**: 41–8.

- Gear JSS, Cassel GA, Gear AJ, et al. Outbreak of Marburg virus disease in Johannesburg. Br Med J 1975;4:489–93.
- Report of a WHO international study team. Ebola hemorrhagic fever in Sudan, 1976. Bull World Health Organ 1978; 56:247–70.
- Report of an international commission. Ebola hemorrhagic fever in Zaire, 1976. Bull World Health Organ 1978; 56:271–93.
- Guillevin L, Lhote F, Ghérardi R. The spectrum and treatment of virus associated vasculitides. Curr Opin Rheumatol 1997;9:31–6.
- Golden MP, Hammer SM, Wanke CA, Albrecht MA. Cytomegalovirus vasculitis. Case reports and review of the literature. Medicine 1994;73: 246–55.
- Ho DD, Rota TR, Andrews CA, Hirsch MS. Replication of human cytomegalovirus in endothelial cells. J Infect Dis 1984;150:956–7.
- Goodman MD, Porter DD. Cytomegalovirus vasculitis with fatal colonic hemorrhage. Arch Pathol 1973;96:281–4.
- Foucar E, Mukai K, Foucar K, Sutherland DER, van Buren CT. Colon ulceration in lethal cytomegalovirus infection. Am J Clin Pathol 1981; 76:788–801.
- Booss J, Dann PR, Winkler SR, Griffith BP, Kim JH. Mechanisms of injury to the central nervous system following experimental cytomegalovirus infection. Am J Otolaryngol 1990;11:313–7.
- Koeppen AH, Lansing LS, Peng SK, Smith RS. Central nervous system vasculitis in cytomegalovirus infection. J Neurol Sci 1981;51:395–410.
- Lin CS, Penha PD, Krishman MN, Zak FG. Cytomegalic inclusion disease of the skin. Arch Dermatol 1981;117:282–4.
- Libman BS, Quinsmorio FP, Stimmler MM. Polyarteritis nodosa like vasculitis in human immunodeficiency virus infection. J Rheumatol 1995; 22:351–5.
- Calabrese LH. Vasculitis and infection with the human immunodeficiency virus. Rheum Dis Clin North Am 1991;17:131–47.
- Gherardi R, Belec L, Mhiri C, et al. The spectrum of vasculitis in human immunodeficiency virus infected patients. Arthritis Rheum 1993;36: 1164–74.
- 36. Oehler R. vaskulitis bei HIV infizierten patienten. Med Klin 1993; 15:327-9.
- Sergent JS, Lockshin MD, Christian CL, Gocke DJ. Vasculitis with hepatitis B antigenemia. Medicine (Baltimore) 1976; 55:1–18.
- Carson CW, Conn DL, Czaja AJ, Wright TL, Brecher ME. Frequency and significance of antibodies to hepatitis C virus in polyarteritis nodosa. J Rheumatol 1993;20:304–9.
- Quint L, Deny P, Guillevin L, et al. Hepatitis C virus in patients with polyarteritis nodosa: prevalence in 38 patients. Clin Exp Rheumatol 1991;9:253–7.
- Leruez-Ville M, Laugé A, Morinet F, Guillevin L, Deny P. Polyarteritis nodosa and parvovirus B19. Lancet 1994;344:263–4.
- Nikkari S, Mertsola J, Korvenranta H, Vainionpää R, Toivanen P. Wegener's granulomatosis and parvovirus B19 infection Arthritis Rheum 1994; 37:1707–8.
- Yoto Y, Kudoh T, Haseyama K, Suzuki N, Chiba S, Matsunaga Y. Human parvovirus B19 infection in Kawasaki disease. Lancet 1994; 344:58–9.
- Lecour H, Miranda M, Magro C, et al. Human leptospirosis—a review of 50 cases. Infection 1989;17:8–12.
- Laing RW, Teh C, Toh CH. Thrombotic thrombocytopenic purpura (TTP) complicating leptospirosis: a previously undescribed association. J Clin Pathol 1990;43:961–2.
- Badesha PS, Saklayen MG. Hemolytic uremic syndrome as a presenting form of HIV infection. Nephron 1996;72:472–5.
- Chu QD, Medeiros LJ, Fisher AE, Chaquette RF, Crowley JP. Thrombotic thrombocytopenic purpura and HIV infection. South Med J 1995; 88: 82–6.
- Qadri SM, Kayali S. Enterohemorrhagic *Escherichia coli*. A dangerous foodborne pathogen. Postgrad Med **1998**;103:179–80.
- 48. Gardlund B. Randomised controlled trial of low dose heparin for prevention

of fatal pulmonary embolism in patients with infectious diseases. The heparin prophylaxis study group. Lancet **1996**;347:1357–61.

- Laing RBS, Brettle RP, Leen CLS. Venous thrombosis in HIV infection. Int J STD AIDS 1996;7:82–5.
- Jenkins RE, Peters BS, Pinchin AJ. Thromboembolic disease in AIDS is associated with cytomegalovirus disease. AIDS 1991; 5:1540–2.
- Marder VJ. Consumptive thrombohemorrhagic disorders. In: Williams WJ, Beutler E, Erslev AJ, Lichtman MA, eds. Hematology. 3rd ed. New York: McGraw-Hill, 1983;1433–61.
- Marder VJ, Feinstein DI, Francis CW, Colman RW. Consumptive thrombohemorrhagic disorders. In: Colman RW, Hirsh J, Marder VJ, Salzman EW, eds. Hemostasis and thrombosis: basic principles and clinical practice. 3rd ed. Philadelphia: JB Lippincott, 1994:1023–63.
- Limbos MA, Lieberman JM. Disseminated intravascular coagulation associated with rotavirus gastroenteritis: report of two cases. Clin Infect Dis 1996;22:834–6.
- WHO scientific working group. Rotavirus and other viral diarrheas. Bull World Health Organ 1980; 58:183–98.
- McKay DG, Margaretten W. Disseminated intravascular coagulation in virus diseases. Arch Intern Med 1967;120:129–52.
- Linder M, Müller-Berghaus G, Lasch HG, Gagel. Virus infection and blood coagulation. Thromb Diath Haemorrh 1970;23:1–11.
- Talley NA, Assumpcao CAR. Disseminated intravascular clotting complicating viral pneumonia due to influenza. Med J Aust 1971;2:763–6.
- Davison AM, Thomson D, Robson JS. Intravascular coagulation complicating Influenza A virus infection. Br Med J 1973; 1:654–5.
- Whitaker AN, Bunce I, Graeme ER. Disseminated intravascular coagulation and acute renal failure in Influenza A2 infection. Med J Aust 1974;2: 196–201.
- Settle H, Glueck HI. Disseminated intravascular coagulation associated with influenza. Ohio State Med J 1975;71:541–3.
- Anderson DR, Schwartz J, Hunter NJ, Cottrill C, Bisaccia E, Klainer AS. Varicella hepatitis: a fatal case in a previously healthy, immunocompetent adult. Arch Intern Med 1994;154:2101–5.
- Davie EW, Fujikawa K, Kisiel W. The coagulation cascade: initiation, maintenance, and regulation. Biochemistry 1991; 30:10363–70.
- 63. Van Gorp ECM, ten Cate H. Coagulation monitoring. In: Webb AR, Shapiro MJ, Singer M, Suter PM, eds. Oxford textbook of critical care medicine. Oxford, UK: Oxford University Press, **1999**:1170–3.
- 64. Van Deventer SJH, Buller HR, ten Cate JW, Aarden LA, Hack CE, Sturk A. Experimental endotoxinaemia in humans: analysis of cytokine release and coagulation, fibrinolytic, and complement pathways. Blood 1990; 76:2520–6.
- Michie HR, Manogue KR, Spriggs DR, et al. Detection of circulating tumor necrosis factor after endotoxin administration. N Engl J Med 1988;318: 1481–6.
- Morrison DC, Ryan JC. Endotoxin and disease mechanisms. Annu Rev Med 1987; 38:417–32.
- Brandtzaeg P, Kieruif P, Gaustad P, et al. Plasma endotoxin as a predictor of multiple organ failure and death in systemic meningococcal disease. J Infect Dis 1989;159:195–204.
- Van Deventer SJH, Büller HR, ten Cate JW, Sturk A, Pauw W. Endotoxinaemia: an early predictor of septicemia in febrile patients. Lancet 1988; 1:605–9.
- Bauer KA, ten Cate H, Barzegar S, Spriggs DR, Sherman ML, Rosenberg RD. Tumor necrosis factor infusions have a procoagulant effect on the hemostatic mechanism of humans. Blood 1989;74:165–72.
- van der Poll T, Buller HR, ten Cate H, et al. Activation of coagulation after administration of tumor necrosis factor to normal subjects. N Engl J Med 1990; 322:1622–7.
- Levi M, ten Cate H, Bauer KA, Buller HR, ten Cate JW, Rosenberg RD. Dose dependent endotoxin induced cytokine release and coagulation activation in chimpanzees. Thromb Haemost 1991;65:793.
- 72. Suffredini AF, Harpel PC, Parillo JE. Promotion and subsequent inhibition

of plasminogen activator after administration of intravenous endotoxin to normal subjects. N Engl J Med **1989**;320:1165–72.

- van der Poll T, Levi M, van Deventer SJH. TNF and the dysbalance between coagulant and anticoagulant mechanisms in septicemia. Update. Intensive Care Emerg Med 1990; 14:269–73.
- 74. Jansen PM, Boermeester MA, Fischer E, et al. Contribution of interleukin-1 to activation of coagulation and fibrinolysis, to neutrophil degranulation and the release of sPLA2 in sepsis. Studies in non-human primates following interleukin 1α administration and during lethal bacteremia. Blood **1995**;86:1027–34.
- Stouthard JML, Levi M, Hack CE, et al. Interleukin-6 stimulates coagulation, not fibrinolysis, in humans. Thromb Haemost 1996;76:738–42.
- Conkling PR, Greenberg CS, Weinberg JB. Tumor necrosis factor induces tissue factor-like activity in human leukemia cell line U937 and peripheral blood monocytes. Blood 1988;72:128–33.
- Conway EM, Bach R, Rosenberg RD, Konigsberg WH. Tumor necrosis factor enhances expression of tissue factor mRNA in endothelial cells. Thromb Res 1989; 53:231–41.
- Bevilacqua MP, Pober JS, Majeau GR, Fiers W, Cotran RS, Gimbrone MA. Recombinant tumor necrosis factor produces pro-coagulant activity in cultured human vascular endothelium. Proc Natl Acad Sci USA 1986; 83:4533–7.
- Osterud B, Flaegstad T. Increased tissue thromboplastin activity in monocytes of patients with meningococcal infections related to unfavourable prognosis. Thromb Haemost 1983;49:5–7.
- Levi M, ten Cate H, Bauer KA, et al. Inhibition of endotoxin induced activation of coagulation and fibrinolysis by pentoxifylline or by a monoclonal anti-tissue factor antibody in chimpanzees. J Clin Invest 1994;93: 114–20.
- Taylor FB Jr, Chang A, Ruf W, et al. Lethal *E. coli* septic shock is prevented by blocking tissue factor with monoclonal antibody. Circ Shock **1991**; 33:127–34.
- Creasy AA, Chang ACK, Feigen L, Wün TC, Taylor FB Jr, Hinshaw LB. Tissue factor pathway inhibitor reduces mortality from *Escherichia coli* septic shock. J Clin Invest **1993**;91:2850–60.
- Biemond BJ, ten Cate H, Levi M, et al. Complete inhibition of endotoxininduced coagulation activation in chimpanzees with a monoclonal Fab fragment against factor VII/VIIa. Thromb Haemost 1995;73:223–30.
- Biemond BJ, ten Cate H, Levi M, et al. Complete inhibition of endotoxin induced coagulation activation in chimpanzees with a monoclonal antibody to factor VII/VIIa. Circulation 1992; 86:679.
- Pixley RA, de la Cadena R, Page JD, et al. The contact system contributes to hypotension but not to disseminated intravascular coagulation in lethal bacteremia. J Clin Invest **1993**;91:61–8.
- Pixley RA, de la Cadena, Page JD, et al. Activation of the contact system in lethal hypotensive bacteremia in a baboon model. Am J Pathol 1992;140:897–906.
- Pajkrt D, van der Poll T, Levi M, et al. Interleukin 10 inhibits activation of coagulation and fibrinolysis during human endotoxemia. Blood 1997;89:2701–5.
- Pradier O, Gérard C, Delvaux A, et al. Interleukin-10 inhibits the induction of monocyte procoagulant activity by bacterial lipopolysaccharide. Eur J Immunol 1993;23:2700–4.
- van der Poll T, Levi M, van Deventer SJH, et al. Differential effects of antitumor necrosis factor monoclonal antibodies on systemic inflammatory responses in experimental endotoxemia in chimpanzees. Blood 1994;83: 446–51.
- van der Poll T, Coyle SM, Levi M, et al. Effect of recombinant dimeric tumor necrosis factor receptor on inflammatory responses to intravenous endotoxin in normal humans. Blood **1997**;89:3727–34.
- Boermeester MA, van Leeuwen PAM, Coyle SM, et al. Interleukin-1 receptor blockade in patients with sepsis syndrome: evidence that interleukin-1 contributes to the release of interleukin-6, elastase and phos-

pholipase A2, and to the activation of the complement, coagulation and fibrinolytic systems. Arch Surg **1995**; 130:739–48.

- van der Poll T, Levi M, Hack CE, et al. Elimination of interleukin 6 attenuates coagulation activation in experimental endotoxemia in chimpanzees. J Exp Med 1994;179:1253–9.
- 93. Levi M, Hack CE, de Boer JP, Brandjes DPM, Büller HR, ten Cate JW. Reduction of contact activation related fibrinolytic activity in factor XII deficient patients: further evidence for the role of the contact system in fibrinolysis in vivo. J Clin Invest 1991;88:1155–60.
- Cotran RS. The endothelium and inflammation: new insights. Monogr Pathol 1982;23:18–37.
- Stemerman MB, Colton C, Morell E. Perturbations of the endothelium. Prog Hemost Thromb 1984; 7:289–324.
- Kaiser L, Sparks HV Jr. Endothelial cells: not just a cellophane wrapper. Arch Intern Med 1987;147:569–73.
- Friedman H, Macarak E, MacGregor R, Wolfe J, Kefalides N. Virus infection in endothelial cells. J Infect Dis 1981;143:266–73.
- Friedman H, Wolfe J, Kefalides N, Macarak E. Susceptibility of endothelial cells derived from different blood vessels to common viruses. In Vitro Cell Dev Biol 1986; 22:397–401.
- Friedman H. Infection of endothelial cells by common human viruses. Rev Infect Dis 1989;11(suppl):S700–4.
- Friedman HM, Macarak EJ, MacGregor RR, Wolfe J, Kefalides NA. Virus infection of endothelial cells. J Infect Dis 1981;143:266–73.
- Ho DD, Rota TR, Andrews CA, Hirsch MS. Replication of human cytomegalovirus in endothelial cells. J Infect Dis 1984; 150:956–7.
- Hoxie JA, Matthews DM, Cines DB. Infection of human endothelial cells by human T-cell leukemia virus type I. Proc Natl Acad Sci USA 1984;81: 7591–5.
- 103. Wiley CA, Schrier RD, Nelson JA, Lampert PW, Oldstone MBA. Cellular localization of human immunodeficiency virus infection within the brains of acquired immune deficiency syndrome patients. Proc Natl Acad Sci USA **1986**; 83:7089–93.
- 104. Butthep P, Bunyaratvej A, Bhamarapravati N. Dengue virus and endothelial cell: a related phenomenon to thrombocytopenia and granulocytopenia in Dengue hemorrhagic fever. Southeast Asian J Trop Med Public Health 1993; 24:246–9.
- Bevilacqua M, Pober J, Wheeler M, Cotran R, Gimbrone M. Interleukin-1 activation of vascular endothelium: effects on procoagulant activity and leucocyte adhesion. Am J Pathol 1985;121:394–403.
- Adams DH, Wyner LR, Karnovsky MJ. Experimental graft arteriosclerosis.
 Immunocytochemical analysis of lesion development. Transplantation 1993; 56:794–9.
- 107. Schorer A, Kaplan M, Rao G, Moldow C. Interleukin-1 stimulates endothelial cell tissue factor production and expression by a prostaglandinindependent mechanism. Thromb Haemost **1986**; 56:256–9.
- van Dam-Mieras MCE, Muller AD, van Hinsbergh VWM, Mullers WJHA, Bomans PHH, Bruggeman CA. The procoagulant response of cytomegalovirus infected endothelial cells. Thromb Haemost 1992; 68: 364–70.
- 109. van Dam-Mieras MCE, Bruggeman CA, Muller AD, Debie WHM, Zwaal RFA. Induction of endothelial cell procoagulant activity by cytomegalovirus infection. Thromb Res 1987;47:69.
- Etingin OR, Silverstein RL, Friedman HM, Hajjar DP. Viral activation of the coagulation cascade: molecular interactions at the surface of infected endothelial cells. Cell **1990**;61:657–62.
- 111. Visser MR, Tracy PB, Vercelotti GM, Goodman JL, White JG, Jacob HS. Enhanced thrombin generation and platelet binding on herpes simplex virus-infected endothelium. Proc Natl Acad Sci USA 1988;85:8227–30.
- Etingin OR, Silverstein RL, Hajjar DP. Identification of a monocyte receptor on herpes virus-infected cells. Proc Natl Acad Sci USA 1991;88:7200–3.
- Dudding L, Haskel S, Clark BD, Auron PE, Sporn S, Huang ES. Cytomegalovirus infection stimulates expression of monocyte associated mediator genes. J Immunol 1989;143:3343–52.

- 114. Smith PD, Saini SS, Raffeld M, Manischewitz JF, Wahl SM. Cytomegalovirus induction of tumor necrosis factor-α by human monocytes and mucosal macrophages. J Clin Invest 1992; 90:1642–8.
- Almeida GD, Porada CD, St Jeor S, Ascensao J. Human cytomegalovirus alters interleukin 6 production by endothelial cells. Blood 1994;83:370–6.
- Pryzdial ELG, Wright JF. Prothrombinase assembly on an enveloped virus: evidence that the cytomegalovirus surface contains procoagulant phospholipid. Blood 1994; 84:3749–57.
- 117. Aloia RC, Tian H, Jensen FC. Lipid composition and fluidity of the human immunodeficiency virus envelope and host cell plasma membranes. Proc Natl Acad Sci USA 1993;90:5181–5.
- 118. Voss R, Matthias FR, Borkowski G, Reitz D. Activation and inhibition of fibrinolysis in septic patients in an intensive care unit. Br J Haematol 1990; 75:99–102.
- van der Poll T, Levi M, Buller HR, et al. Fibrinolytic response to tumor necrosis factor in healthy subjects. J Exp Med 1991;174:729–73.
- 120. Biemond BJ, Levi M, ten Cate H, et al. Plasminogen activator and PAI-I release during experimental endotoxemia in chimpanzees: effects of various interventions in the cytokine and coagulation cascades. Clin Sci 1995; 88:587–94.
- Esmon CT. The regulation of natural anticoagulant pathways. Science 1987;235:1348–52.
- Sugerman RW, Church JA, Goldsmith JC, Ens GE. Acquired protein S deficiency in children infected with human immunodeficiency virus. Pediatr Infect Dis J 1996;15:106–11.
- 123. Stahl CP, Wideman CS, Spira TJ, Haff EC, Hixon GJ, Evatt BL. Protein S deficiency in men with long term human immunodeficiency virus infection. Blood 1993;81:1801–7.
- Lafeuillade A, Alessi MC, Poizot-Martin I, et al. Endothelial cell dysfunction in HIV infection. J Acquir Immune Defic Syndr 1992;5:127–31.
- 125. Bissuel F, Verruyer M, Causse X, Dechavane M, Trepo C. Acquired protein S deficiency: correlation with advanced disease in HIV-infected patients. J Acquir Immune Defic Syndr 1992; 5:484–9.
- 126. Malia RG, Kitchen S, Greaves M, Preston FE. Inhibition of activated protein C and its cofactor protein S by antiphospholipid antibodies. Br J Haematol 1990; 76:101–7.
- 127. Lo SCL, Salem HH, Howard MA, Oldmeadow MJ, Firkin BG. Studies of natural anticoagulant proteins and anticardiolipin antibodies in patients with the lupus anticoagulant. Br J Haematol **1990**; 76:380–6.
- Hassell KL, Kressin DC, Neumann A, Ellison R, Marlar RA. Correlation of antiphospholipid antibodies and protein S deficiency with thrombosis in HIV-infected men. Blood Coagul Fibrinolysis 1994; 5:455–62.
- Llorente MR, Carton JA, Carcaba V, et al. Antiphospholipid antibodies in human immunodeficiency virus infection. Med Clin (Barc) 1994;103: 10–3.
- Levin J. Bleeding with infectious diseases. In: Ratnoff OD, Forbes CD, eds. Disorders of hemostasis. New York: Grune & Stratton, 1984:367–78.
- Kelton J, Neame P. Hemorrhagic complications of infection. In: Hemostasis and thrombosis. Philadelphia, JB Lippincott 1987:965–74.
- Bierman HR, Nelson ER. Hematodepressive virus diseases of Thailand. Ann Intern Med 1965; 62:867–84.
- Nakao S, Lai CJ, Young NS. Dengue virus, a flavivirus, propagates in human bone marrow progenitors and hematopoietic cell lines. Blood 1989;74:1235–40.
- Na-Nakorn S, Suingdumrong A, Pootrakul S, Bhamarapravati N. Bone marrow studies in Thai hemorrhagic fever. Bull WHO 1966; 35:54–5.
- Halstead SB. Antibody, macrophages, Dengue virus infection, shock and hemorrhage: a pathogenetic cascade. Rev Infect Dis 1989;11(suppl): S830–9.
- Terada H, Baldini M, Ebbe S, Madoff MA. Interaction of Influenza virus with blood platelets. Blood 1966; 28:213–28.
- Stemerman MB. Vascular intimal components: precursors of thrombosis. Prog Hemost Thromb 1974;2:1–47.
- 138. Curwen KD, Gimbrone MA Jr, Handin RI. In vitro studies of thrombores-

istance: the role of prostacyclin (PGI2) in platelet adhesion to cultured normal and virally transformed human vascular endothelial cells. Lab Invest **1980**;42:366–74.

- Kolars CP, Spink WW. Thrombopenic purpura as a complication of mumps. JAMA 1958; 168:2213–5.
- Graham DY, Brown CH III, Benrey J, Butel JS. Thrombocytopenia: a complication of mumps. JAMA 1974;227:1162–4.
- 141. Myllylä G, Vaheri A, Vesikari T, Penttinen K. Interaction between human blood platelets, viruses and antibodies. Post rubella thrombocytopenic purpura and platelet aggregation by rubella antigen-antibody interaction. Clin Exp Immunol **1969**;4:323–32.
- 142. Hirsch EO, Gardner FH. The transfusion of human blood platelets, with a note on the transfusion of granulocytes. J Lab Clin Med 1952; 39: 556–69
- Morse EE, Zinkham WH, Jackson DP. Thrombocytopenic purpura following rubella infection in children and adults. Arch Intern Med 1966; 117: 573–9.
- Hudson JB, Weinstein L, Chang TW. Thrombocytopenic purpura in measles. J Pediatr 1956;48:48–56.
- Charkes ND. Purpuric chickenpox: report of a case, review of the literature, and classification by clinical features. Ann Intern Med 1961;54:745–59
- 146. Tobin JD Jr, ten Bensel RW. Varicella with thrombocytopenia causing fatal intracerebral hemorrhage. Am J Dis Child 1972; 124:577–8.
- Brook I. Disseminated varicella with pneumonia, meningoencephalitis, thrombocytopenia,, and fatal intracranial hemorrhage. South Med J 1979; 72:756–7.
- Whitaker JA 3d, Hardison JE. Severe thrombocytopenia after generalized herpes simplex virus-2 (HSV-2) infection. South Med J 1978;71:864–5.
- Chanarin I, Walford DM. Thrombocytopenic purpura in cytomegalovirus mononucleosis. Lancet 1973;2:238–9.
- Sahud MA, Bachelor MM. Cytomegalovirus-induced thrombocytopenia: an unusual case report. Arch Intern Med 1978;138:1573–5.
- Carter RL. Platelet levels in infectious mononucleosis. Blood 1965;25: 817–21.
- Sharp AA. Platelets, bleeding and haemostasis in infectious mononucleosis. In: Carter RL, Penman HG, eds. Infectious mononucleosis. Oxford, UK: Blackwell Scientific, 1969:99–110.
- Casey TP, Matthews JRD. Thrombocytopenic purpura in infectious mononucleosis. N Z Med J 1973;77:318–20.
- Ellman L, Carvalho A, Jacobson BM, Colman RW. Platelet autoantibody in a case of infectious mononucleosis presenting as thrombocytopenic purpura. Am J Med 1973; 55:723–6.
- Mazza J, Magnin GE. Severe thrombocytopenia in infectious mononucleosis: report of two cases and review of the literature. Wis Med J 1975; 74:124–7.
- Lee M. Korean hemorrhagic fever (hemorrhagic fever with renal syndrome).
 2nd ed. Seoul: Seoul National University Press, 1986:83–106.
- 157. Lo TT, Wang SH, Hwang HF, Pan NC, Qing WO, Wei WY. Mechanisms of bleeding and the search for treatment in epidemic hemorrhagic fever. Chin J Infect Dis 1986;4:716–9.
- Suvatte V. Dengue hemorrhagic fever: hematologic abnormalities and pathogenesis. J Med Assoc Thai 1978;61(suppl):53–8.
- 159. Swanepoel R, Shepherd AJ, Leman PA, et al. Epidemiologic and clinical features of Crimean-Congo hemorrhagic fever in Southern Africa. Am J Trop Med Hyg **1987**; 36:120–32.
- 160. Martini GA. Marburg virus disease: clinical syndrome. In: Martini GA, Siegert R, eds. Marburg virus disease. New York: Springer Verlag, 1971: 1–9.
- McEver R. GMP-140: a receptor for neutrophils and monocytes on activated platelets and endothelium. J Cell Biochem 1991;45:156–61.
- Etingin O, Silverstein R, Hajjar D. Von Willebrand factor mediates platelet adhesion to virally infected endothelial cells. Proc Natl Acad Sci USA 1993;90:5153–6.
- 163. van Gorp ECM, Suharti C, ten Cate H, et al. Endothelial cell activation

in (Dengue) hemorrhagic fever [abstract]. In: Program and abstracts of the 38th Interscience Conference on Antimicrobial Agents and Chemotherapy (San Diego). Washington, DC: American Society for Microbiology, **1998**.

- 164. Anderson R, Wang S, Osiowy C, Issekutz AC. Activation of endothelial cells via antibody-enhanced dengue virus infection of peripheral blood monocytes. J Virol 1997;71:4226–32.
- 165. Takahashi M, Ikeda U, Masuyama J, et al. Monocyte-endothelial cell interaction induces expression of adhesion molecules on human umbilical cord endothelial cells. Cardiovasc Res 1996; 32:422–9.
- 166. Seigneur M, Constans J, Blann A, et al. Soluble adhesion molecules and endothelial cell damage in HIV-infected patients. Thromb Haemost 1997; 77:646–9.
- Calabrese LH. Vasculitis and infection with the human immunodeficiency virus. Rheum Dis Clin North Am 1991; 17:131–47.
- Gherardi R, Belec L, Mhiri C, et al. The spectrum of vasculitis in human immunodeficiency virus infected patients. Arthritis Rheum 1993;36: 1164–74.
- 169. Drancourt M, George F, Brouqui P, Sampol J, Raoult D. Diagnosis of Mediterranean spotted fever by indirect immunofluorescence of *Rickettsia conorii* in circulating endothelial cells isolated with monoclonal antibody–coated immunomagnetic beads. J Infect Dis **1992**;166:660–3.
- 170. George F, Brouqui P, Boffa MC, et al. Demonstration of *Rickettsia conorii*induced endothelial injury in vivo by measuring circulating endothelial cells, thrombomodulin, and von Willebrand factor in patients with Mediterranean spotted fever. Blood **1993**;82:2109–16.

- Solovey A, Lin Y, Browne P, Choong S, Wayner E, Hebbel RP. Circulating activated endothelial cells in sickle cell anemia. N Engl J Med 1997; 337: 1584–90.
- Nicholson AC, Hajjar DP. Herpesviruses in atherosclerosis and thrombosis. Etiologic agents or ubiquitous bystanders? Arterioscler Thromb Vasc Biol 1998; 18:339–48.
- 173. Danesh J, Collins R, Peto R. Chronic infections and coronary heart disease: is there a link? [review]. Lancet 1997; 350:430–6.
- 174. Gurfinkel E, Bozovich G, Daroca A, et al. Randomised trial of roxithromycin in non Q wave coronary syndromes: Roxis pilot study. Lancet 1997; 350:404–7.
- 175. Gupta S, Leatham EW, Carrington D, et al. Elevated *Chlamydia pneumoniae* antibodies, cardiovascular events, and azithromycine in male survivors of myocardial infarction. Circulation **1997**;96:404–7.
- 176. Cappello M, Vlasuk GP, Bergum PW, et al. Ancylostoma caninum anticoagulant peptide: a hookworm-derived inhibitor of human coagulation factor Xa. Proc Natl Acad Sci USA 1995;92:6152–6.
- 177. Stanssens P, Bergum PW, Gansemans Y, et al. Anticoagulant repertoire of the hookworm *Ancylostoma caninum*. Proc Natl Acad Sci USA **1996**; 93:2149–54.
- 178. Taylor FB Jr, Chang A, Ruf W, et al. Lethal *E. coli* septic shock is prevented by blocking tissue factor with monoclonal antibodies. Circ Shock 1991; 33:127–37.
- Creasey AA, Chang ACK, Feigen L, Wün TC, Taylor FB Jr, Hinshaw LB. Tissue factor pathway inhibitor reduces mortality from *Escherichia coli* septic shock. J Clin Invest **1993**;91:2850.