

# Wrapped Forages for Horses

**Cecilia E. Müller**

*Faculty of Veterinary Medicine and Animal Science  
Department of Animal Nutrition and Management  
Uppsala*

**Doctoral thesis  
Swedish University of Agricultural Sciences  
Uppsala 2007**

**Acta Universitatis Agriculturae Sueciae**

2007: 44

ISSN 1652-6880  
ISBN 978-91-576-7343-5  
© 2007 Cecilia Müller, Uppsala  
Tryck: SLU Service/Repro, Uppsala 2007

## Abstract

Müller, C.E. 2007. *Wrapped forages for horses*. Doctoral dissertation. ISSN 1652-6880, ISBN 978-91-576-7343-5.

Wrapped forages, in the form of silage and haylage, have become more common in horse diets during recent years. Silage and haylage is commonly produced in big bales. However, for use in stables with few animals, these bales often contain too much forage to be consumed before onset of aerobic deterioration. Smaller bales are therefore of interest, but knowledge of the chemical composition (including vitamin content), fermentation pattern and changes in those variables during storage of small bales is limited, and was therefore investigated. Small bale forage contained higher pH, higher ethanol and lower lactic acid content, compared to general levels in chopped silo silage, but low levels of ammonia-N and butyric acid. There were no general effects of dry matter or extent of fermentation on  $\alpha$ -tocopherol and  $\beta$ -carotene contents in the preserved forages, but linear positive correlations between the vitamins and lactic acid existed. In general, long-term storage (14 months) of small bales influenced fermentation variables, yeasts and pH, but silage was affected by storage to a larger extent than haylage. Although changes occurred during storage, values in two-month old bales correlated well with values obtained after 14 months.

The influence of forage conservation methods on horse preference was also investigated. Hay, haylage and silage were produced from the same grass crops and the forages were offered simultaneously to horses. Silage was the first chosen forage, had the highest rate of consumption and the longest eating time, while hay had the lowest consumption rate and the shortest eating time. Haylage was intermediate between hay and silage in both eating time and rate of consumption.

The influence of forage conservation methods on equine hindgut fermentation was studied using fistulated horses. Hay, haylage and silage were produced from the same grass crop and fed in a changeover study. Horses were sampled after being fed the forage for 21 days, and a kinetic study of colon fermentation was performed in each period. Forage conservation method had no effect on microbial or chemical composition in the right ventral colon or faeces on Day 21. All forages showed similar fermentation kinetics in the right ventral colon before (0h) and at 2, 4, 8 and 12 h after feeding.

*Keywords:* silage, haylage, hay, horse, preference, hindgut fermentation, storage, bales

*Author's address:* Cecilia Müller, Kungsängen Research Centre, Department of Animal Nutrition and Management, SLU, 753 23 UPPSALA, Sweden.

E-mail: Cecilia.Muller@huv.slu.se

# Zusammenfassung

Müller, C.E. 2007. Folienverpacktes Weidefutter für Pferde. Doktorabhandlung. ISSN 1652-6880, ISBN 978-91-576-7343-5.

Folienverpacktes Weidefutter, in Form von Silage und Heusilage, hat in den letzten Jahren im zunehmenden Maß das Heu in den Pferderationen ersetzt. Silage und Heusilage wird allgemein in den grossen Ballen produziert. Für den Gebrauch in kleineren Pferdeställen, enthalten diese Ballen jedoch häufig zu viel Futter das ein Verfaulungsprozess startet bevor die Ballen verbraucht sind. Aus diesem Grund wurde die Möglichkeit untersucht Kleiballen zu produzieren. Jedoch sind Kenntnisse über die chemischen Zusammensetzung (einschließlich Vitamininhalt), des Gärusters und der Veränderungen während der Lagerung in solchen Ballen begrenzt, und diese Variablen wurden folglich untersucht. Kleine Ballen enthalten einen höheren pH und höhere Äthanolkonzentration aber weniger Milchsäuregehalt, verglichen mit allgemeinen Niveaus in gehäckselten Silage, aber niedrigen Niveaus des Ammoniak-N und Buttersäure. Es gab keine allgemeinen Veränderungen der Trockensubstanz und/oder Gärumfang auf  $\alpha$ -tocopherol- und  $\beta$ -carotininhalt in konserviertem Futter. Im Allgemeinen wurden langfristige Lagerung (14 Monate) der kleine Ballen beeinflussten Gärungsvariablen, Hefen und pH aber Silage Veränderungen in mehr Variablen als Heusilage unterworfen. Obgleich Veränderungen während der Lagerungen eintraten, stimmten Werte in den 2 Monate alten Ballen gut mit den Werten den 14 Monate alten Ballen überein.

Der Einfluß der Konservierungsmethode des Grünfutters auf die Präferenz von Pferden wurde auch untersucht. Heu, Heusilage und Silage wurde aus dem gleichen Gras produziert und angeboten Pferden. Die Konservierungsmethode wirkte auf die Pferd Präferenz zur Bevorzugung von Silage. Silage war die erste Wahl, hatte den höchsten Verzehr und die längste Freßzeit, während Heu den niedrigsten Verzehr und die kürzeste hatte. Heusilage war zwischen Heu und Silage in der Verzehr und in der Freßzeit zu finden.

Der Einfluß der Konservierungsmethode des Grünfutters auf die Fermentation im Dickdarm (rechter ventraler Kolon) wurde an fistulierten Pferde untersucht. Heu, Heusilage und Silage wurden aus dem gleichen Gras produziert und in einem change-over Experiment vollzogen. Die Konservierungsmethode hatte keinen Effekt auf die mikrobielle oder chemische Zusammensetzung im Dickdarm gemessen am 21 Tag jeder Fütterungsperiode. Eine kinetische Studie der Kolon wurde auch durchgeführt. Resultate zeigten, daß alle Futtertypen im gleichen Bereich nach 0, 2, 4, 8 und 12 Stunden Werte in der selben Größenordnung aufwiesen.

*Schlüsselwörter:* Silage, Heusilage, Heu, Pferd, Präferenz, Dickdarm, Konservierungsmethode, Fermentation

*Adresse des Autors:* Cecilia Müller, Kungsängen Research Centre, Department of Animal Nutrition and Management, SLU, 753 23 UPPSALA, Schweden.  
E-mail: Cecilia.Muller@huv.slu.se

*"Skulle det någon gång undantagsvis förekomma, att man nödgas lägga för sina hästar ett fördärfoat hö, måste man, sedan man valt ut det bästa, sorgfälligt skaka och lufta det, samt minst 12 timmar före fodringen bespränga det med saltvatten. Sedan höet därefter torkat i friska luften, må det i all världens dar läggas för de stackars krakarna."*

*Greve och Hippolog Carl Gustaf Wrangel, 1839-1908.*

*"Should it ever occur, that one must lay before one's horses a spoiled hay, one must, after selecting the best, carefully shake and aerate it, and at least 12 hours before feeding, sprinkle it with salted water. Thence, after the hay has dried in fresh air, may it in pity be laid before the poor rips."*

*Count and Hippologist Carl Gustaf Wrangel, 1839-1908*

*(free translation by the author of the thesis).*

***Till Mamma och Pappa***

# Contents

## **Introduction, 9**

The role of forage in equine nutrition, 9

Forage conservation methods, 10

*Hay, 10*

*Silage and haylage, 12*

Tocopherols and carotenes in forage in relation to equine nutrition, 16

Factors influencing forage intake and preference in horses, 17

Factors influencing the ecosystem in the equine hindgut, 18

## **Aims of the thesis, 19**

### **Material and methods, 19**

Grass crops (I-V), 20

Production of small bale silage and haylage (I-IV), 20

*Aerobic storage stability (I), 21*

Preference study (IV), 21

Studying the equine hindgut (V), 21

Statistical analysis (I-V), 22

### **Results, 22**

Production of small bale silage and haylage (I-IV), 22

Fermentation pattern and microbial counts in small bale silage and haylage (I-V), 22

*Aerobic storage stability (I), 23*

Storage of small bale silage and haylage (II), 23

Tocopherols and carotenes in small bale silage and haylage (III), 24

Horse preference for hay, haylage and silage (IV), 24

Composition of hindgut content in horses fed hay, haylage and silage (V), 24

### **Discussion, 25**

Production of small bale silage and haylage, 25

Influence of conservation method on forage composition, 26

*Influence of epiphytic microflora on fermentation, 29*

*Aerobic storage stability, 30*

Storage of small bale silage and haylage, 31

Tocopherols and carotenes in preserved forage, 32

Influence of forage conservation method on preference of horses, 32

Influence of forage conservation method on hindgut fermentation in horses, 34

*Digestion of water-soluble carbohydrates in forages, 36*

### **Main conclusions, 38**

### **Populärvetenskaplig sammanfattning, 38**

### **References, 41**

### **Acknowledgements, 55**

# Appendix

## Papers I-V

The present thesis is based on the following papers, which are referred to in the text by their Roman numerals:

- I. Müller, C.E. 2005. Fermentation patterns of small-bale silage and haylage produced as a feed for horses. *Grass and Forage Science* **60**, 109-118.
- II. Müller, C.E., Pauly, T.M. & Udén, P. 2007. Storage of small bale silage and haylage—influence of storage period on fermentation variables and microbiological composition. *Grass and Forage Science* **62** (3), xx-xx (*In Press*).
- III. Müller, C.E., Möller, J., Krogh Jensen, S. & Udén, P. 2007. Tocopherol and carotenoid levels in baled silage and haylage in relation to horse requirements. *Animal Feed Science and Technology* (*In press*).
- IV. Müller, C.E. & Udén, P. 2007. Preference of horses for grass conserved as hay, haylage or silage. *Animal Feed Science and Technology* **132**, 66-78.
- V. Müller, C.E. & Udén, P. 2007. Effect of forage conservation method on microbial and chemical composition in the hindgut of horses fed hay, haylage and silage (*Submitted*).

Reprints are published with kind permission of the publishers concerned.

The experiment in Paper **IV** was approved by the Ethical Committee in Uppsala, Sweden. The experiment in Paper **V** was approved by the Department of Health and Animal Care of the French Veterinary Authority, France.

## List of abbreviations and definitions

Ammonia-N	ammonia-nitrogen
ATP	adenosine-triphosphate
$a_w$	water activity
CFU	colony forming units
Co-EDTA	cobalt(III)ethylenediamine-tetraacetate
CP	crude protein
DM	dry matter
DNA	deoxyribonucleic acid
FM	fresh matter
ha	hectare
Haylage	airtight stored grass containing $\geq 500$ g DM/kg
ip	intraperitoneal
LAB	lactic acid bacteria
LD <sub>50</sub>	lethal dose 50 %
NADH	nicotinamide adenine dinucleotide
NDF	neutral detergent fibre
$r^2$	square of the correlation coefficient
RAO	recurrent airway obstruction
SCFOS	short-chain fructo-oligosaccharides
SD	standard deviation
SEM	standard error of mean
Silage	ensiled grass containing less than 500 g DM/kg
SLU	Swedish University of Agricultural Sciences
VDMI	voluntary dry matter intake
VFA	volatile fatty acids
WSC	water-soluble carbohydrates



# Introduction

## The role of forage in equine nutrition

Horses (*Equus caballus*) have evolved as grass eaters, capable of surviving on plant tissues due to microbial fermentation of the feed in the hindgut (Janis, 1976). Hindgut fermentation was adopted as a digestive strategy early in evolution, and enabled equids to survive on grassland characterized by plants with high fibre contents (Sneddon & Argenzio, 1998). Some 25 million years ago, the ancestor of the modern horse, known from fossil records as *Meryhippus*, was adapted to life as a fast-moving grazer of the plains (Sneddon, 1993).

The modern horse is still a grass eater, designed for hindgut fermentation of fibrous feeds into volatile fatty acids (VFA) (Hintz *et al.*, 1971; Argenzio, Southworth & Stevens, 1974). The equine digestive system is adapted to grass, and as in other herbivores, the digestive system works best with the feed it is adapted to (Hummel *et al.*, 2006). Feeding horses small amounts of fibrous feeds is connected with digestive upsets (Clarke, Roberts & Argenzio, 1990; Archer & Proudman, 2006) and development of stereotypic behaviour (McGreevy *et al.*, 1995; Goodwin, Davidson & Harris, 2002). Gastric ulcers have also been reported in ponies fed only concentrates, in contrast to ponies fed hay (Coenen, 1990), and the risk for colic incidents has been found to increase if more than 2.7 kg oats is fed daily (Hudson *et al.*, 2001). Recent studies also show that the mandibular motion is different when horses are eating a pelleted concentrate feed compared to hay, and that diets high in pelleted feed may have an impact on normal dental wear (Bonin *et al.*, 2007). Forage should thus constitute the major proportion in diets to horses, and pasture grass would be the simplest way of ensuring this. However, in northern countries, pasture cannot be provided all year round due to the cold winter climate. The summer grass therefore has to be conserved, and traditionally, haymaking has been the most common way to conserve forage for horses (Thompson, 1983).

The need for conserved forages in a transportable form has a long history in equine management. At the end of the 19<sup>th</sup> century, the UK cavalry had a large requirement of hay in bales, and at the 1878 Agricultural Exhibition in Paris, the British military bought four of the first stationary hay balers available to bale hay for horses in battle (Thompson, 1983). Thompson (1983) also described a similar requirement for forage in a moveable form by the civil society; at the end of 1800 and beginning of 1900, about 200 000 horses existed in London which had to be provided hay and other feedstuffs from farms outside the city. The present structure of the Swedish horse population is somewhat similar. At least 285 000 horses exist in Sweden (Bratt, 2001), and 75% of these are housed in areas close to cities (Persson, 2005). In recent years, hay in horse diets has been partly replaced by baled silage (<500 g dry matter (DM)/kg) and haylage (≥500 g

DM/kg) (Billysson, 2002; Holmquist & Müller, 2002; Schwarz *et al.*, 2005). The idea of feeding silage to horses is however, not new, as Nourse reported in 1897 that maize silage was a suitable feed for horses and mules if the animals were successively adapted to it.

A common way to produce silage and haylage is in the form of big bales. Today, bales account for approximately 60% of the total amount of silage conserved in Sweden (Wilkinson & Toivonen, 2003). Big bales are, however, not ideal for use in most horse stables, as more than 75% of the Swedish stables house only one to four horses (Persson, 2005). Thus, the big bales contain too much forage, if the bale is to be consumed before onset of aerobic deterioration. Handling of big bales has also been mentioned as a problem at small un-mechanized horse farms (Müller, 2002). Therefore, smaller bales are of interest for horse feeding purposes, but machinery equipment for production of small bale silage and haylage is not readily available at present. Also, scientific knowledge of wrapped forages as substitutes for hay in horse diets is limited and needs further study.

## **Forage conservation methods**

### *Hay*

Hay has been the traditional type of conserved forage used for horses, but if not produced and stored properly (i.e. dry and airy), it is readily subjected to mould growth (Lacey, 1989). The preservative effect of hay-making is due to a low water activity ( $a_w$ ), and the dried hay must be kept as such in order to retain its nutritive value and hygienic quality, and for minimizing DM losses (Gregory *et al.*, 1963; Sullivan, 1973; Clevström *et al.*, 1981; Clevström and Ljunggren, 1984; Hlödversson, 1985; Lacey, 1989). The dried hay should have a DM level above 840 g/kg to keep mould counts low and to keep the hay from heating (Gregory *et al.*, 1963). Spontaneous heating of damp hay renders it brown and/or black due to the formation of indigestible Maillard products (Sullivan, 1973).

Barn-drying hay at harvest was found to be superior to field-drying and chemical preservation in restricting DM losses and mould growth under Swedish conditions (Clevström *et al.* 1981; Clevström & Ljunggren, 1984; Hlödversson, 1985). Clevström & Ljunggren (1984) investigated the mycological flora in field- and barn-dried hay, harvested from the same crop at different DM levels (600, 700 and 800 g DM/kg), and stored for six months. There was a heavy growth of *Aspergillus*, *Penicillium* and occasionally *Rhizopus* in the field-dried hay. The barn-dried hay also showed growth of *Aspergillus* and *Penicillium*, but the number of CFU (colony forming units)/g was much less than in the field-dried hay. The lowest mould spore load was found in the barn-dried hay harvested at 600 g DM/kg. Clevström *et al.* (1981) studied chemical preservation of hay (550 and 730 g DM/kg) by addition of various amounts of sodium chloride, propionic acid or formic acid, and found that mould counts were particularly high in hay bales treated with propionic or formic acid. Treatment with formic acid also resulted in an almost pure culture of *A. flavus*, and both aflatoxins B<sub>1</sub> and G<sub>1</sub> were found (635-1000 µg/l) in the hay.

Dry matter losses in hay during storage are primarily caused by fungal growth (Hlödversson, 1985). The extent of mould growth in stored hay can be different at the surface compared to within the stack or the bales, as direct contact with moist air can create conditions favourable to mould growth. In a preliminary report, Sundberg & Lindahl (2006) showed fluctuations in relative moisture,  $a_w$  and the presence and quantity of different mould species at the surface, at 25-cm depth and at 50-cm depth in hay bales stored in stacks from August to May. Registrations or samples were taken once every second month, and at three different farms. Results showed that the relative moisture at the surface, but not at 25- or 50-cm depth, followed the relative moisture in the air to a great extent. Water activity was higher at the surface than at 25- or 50-cm depth, and was  $\geq 0.70$  at the surface from October until May. At 25- and 50-cm depth,  $a_w$  generally fluctuated around 0.60 during the same period. Active growth of most filamentous fungi is restricted at  $a_w$  0.80 (Hocking, Miscamble & Pitt, 1994; Adams & Moss, 1995), although xerophilic fungi may be able to grow at  $a_w$  0.61 (Adams & Moss, 1995). The mould growth in the study of Sundberg & Lindahl (2006) was larger (in CFU/g) at the surface than at 25- and 50-cm depth, and a greater diversity of mould species was also found at the surface.

#### Respirable particles in forage and respiratory disorders

Mould growth in forage for horses should be avoided as moulds produce both spores and mycotoxins. Mould spores, together with actinomycetes such as *Micropolyspora faeni*, play a large role in the aetiology of recurrent airway obstruction (RAO), also known as “heaves” or “broken wind”, which is a chronic respiratory disease in horses (Falk-Rønne *et al.*, 1984; Clarke, 1987; Robinson *et al.*, 1996; Vandeput *et al.*, 1998). Airborne endotoxins can also contribute to pulmonary dysfunction like RAO (Künzle *et al.*, 2007). Gohlke (1957) argued that the condition was recognized as a manmade disease already by Aristoteles (384–322 BC), and has since been attributed to the use of mouldy feeds and bedding materials. A genetic background in the susceptibility of acquiring the disease is also suspected (Ewart & Robinson, 2007). Respiratory problems were found to be the second largest reason (8 to 9%) for culling of Swedish Warmblood horses born during 1965–1982, and 93% of the respiratory problems were defined as RAO (Wallin *et al.*, 2000). In the UK, the estimated true prevalence of RAO in the horse population was 14 % (Hotchkiss, Reid & Christley, 2007). In a recent Swedish study based on insurance company registrations, Penell *et al.* (2005) found that respiratory diseases accounted for 5% of the proportional morbidity in the insured equine population in Sweden. If these studies are comparable, respiratory diseases seems to have decreased slightly as a cause of culling of horses in Sweden. However, as the disease results in laboured breathing (Robinson *et al.*, 1996), it is still a serious welfare issue for affected horses. Also, impaired performance, the chronic state of the condition and veterinary costs associated with care of RAO-horses make preventive measures important.

As RAO is associated with respirable particles (0.5–3.0  $\mu\text{m}$ ) such as mould spores, forages with less mould growth or fewer counts of respirable particles are

of interest for horse feeding purposes (Clarke, 1987; Raymond *et al.*, 1997; Vandenput *et al.*, 1997). Vandenput *et al.* (1997) showed that even “good quality” (meaning not visibly dusty when shaken) grass hay contained higher levels of respirable particles than haylage (780 g DM/kg) or silage (480–490 g DM/kg), and silage contained less respirable particles and less thermophilic actinomycetes than haylage. Vandenput *et al.* (1998) also reported that to maintain a RAO-horse in clinical remission, it was more important to eliminate hay from the surrounding environment than to change the type of bedding material. Horses with RAO fed grass silage showed no difference in pulmonary function (mechanics of breathing and arterial blood analysis) compared to healthy control horses fed hay, or compared to themselves after being two months on pasture. However, if RAO-horses were fed hay, they developed clinical signs of RAO and had significant changes in pulmonary function parameters within eight days (s.d.± 3 days) (Vandenput *et al.*, 1998). A pasture-like environment can be achieved by using silage and wooden shavings instead of hay and straw, as showed by McGorum, Ellison & Cullen (1998), who measured total and respirable dust and airborne endotoxins in different equine management systems.

One method to reduce the amount of respirable particles in hay is to soak it in water before feeding. Moore-Colyer (1996) found that soaking hay for 30 minutes reduced the amount of respirable particles present in dry hay by about 90%, and Blackman & Moore-Colyer (1998) showed that soaking hay for 10 minutes, or steaming hay for 80 minutes, reduced the amount of respirable particles in dry hay by 93%. However, the procedure of soaking hay is labour-intensive and leads to nutritional losses (in particular P, K, Mg, Na and Cu), even if they are small when the hay is not “oversoaked” (*e.g.* soaked for longer periods than 30 minutes) (Moore-Colyer, 1996; Blackman & Moore-Colyer 1998). The steaming procedure was found to be very expensive in relation to soaking (Blackman & Moore-Colyer, 1998). As it is not known what actually happens with the mould spores during soaking or steaming, the safety of these procedures can be questioned. Soaking of hay also provides possibilities for other microorganisms to start growing, and hay that has been “oversoaked” may be questionable from this perspective as well. Also, if mycotoxins are present in mouldy dry hay, it is not known whether they remain in the hay or in the water after soaking. Mycotoxins produced by *Fusarium spp.* are well known health hazards for horses (Asquith, 1991), but intoxication of horses by *e.g. Aspergillus spp.*, *Penicillium spp.* and *Stachybotrys spp.* toxins has also been reported (Aller, Edds & Asquith, 1981; Asquith, 1991; Vesonder *et al.*, 1991; Ochoi *et al.*, 1992; Barnett *et al.*, 1995; Le Bars & Le Bars, 1996).

### *Silage and haylage*

Finner (1966) classified silages in three categories: direct-cut silage, wilted silage and low-moisture silage, the latter also named haylage. A typical haylage would, according to Finner (1966), contain 400 to 600 g DM/kg. In this thesis, haylage has been defined as containing  $\geq 500$  g DM/kg. Silage is preserved by the fermentative activities of lactic acid bacteria (LAB) in anaerobic environments, resulting in a lowered pH. Production of lactic acid decreases with increasing DM

content (Jackson & Forbes, 1970). Haylage is preserved through a combination of drying, airtight storage and, depending on the possibility for LAB to grow, also presence of lactic acid and decreased pH. Wilting of crops for haylage increases the concentration of solubles in the liquid fraction, which decreases  $a_w$  of the crop (Greenhill, 1964). The effect of a lowered  $a_w$  is inhibitory on overall microbial activity (Adams & Moss, 1995). Greenhill (1964), however, proposed that in low moisture crops, the limited availability of the plant juice was probably the reason for low lactic acid production, and not the lowered  $a_w$  in itself. The differences in the conservation methods generally result in higher content of water-soluble carbohydrates (WSC), higher pH and lower concentration of VFA, lactic acid, ammonia-N and alcohols (ethanol and 2,3-butanediol) in haylage compared to silage (Gordon *et al.*, 1961; Greenhill, 1964; Finner, 1966; Nicholson *et al.*, 1991; Pahlow & Weissbach, 1996; Dawson *et al.*, 1999; Driehuis & van Wijkelaar, 2000; Han *et al.*, 2006).

LAB are present on the grass crop in the field, and the most frequently found species are heterofermentative *Leuconostoc mesenteroides* and *Lactobacillus fermentum*, as well as homofermentative *Lactobacillus plantarum* and *Pediococcus spp.* Pahlow & Dinter (1987) reported that the relative proportion of *Pediococci spp.* increased with plant maturity, but the ratio of hetero- to homofermentative LAB remained quite stable at 1:1.2. Wilting has been demonstrated to reduce total LAB counts on the fresh crop (Pahlow & Dinter, 1987). Glucose and fructose present in the crop are considered as the primary energy sources of homofermentative LAB during ensiling. LAB metabolism by the Embden-Meyerhof-Parnas glycolytic pathway yields lactate, adenosinetriphosphate (ATP) and water from pyruvate (Gibbs *et al.*, 1950). Grass fructans (mainly  $\beta$ -2,6-linked levans, also known as phleins according to Suzuki (1993)) can be indirectly involved as fructose sources during ensiling as shown by Merry *et al.* (1995), who found fructans to be degraded to fructose during ensiling. Fructans were degraded even in the absence of bacteria, but degradation was more rapid if bacteria were present (Merry *et al.*, 1995). However, isolation of 712 strains of LAB from grass forage showed that only 16 strains were able to ferment phlein-type fructans (Müller & Lier, 1994). Müller & Steller (1995) also demonstrated that different strains of LAB had different abilities to enzymatically degrade fructans of  $\beta$ -2,6-linked levan type and  $\beta$ -2,1-linked inulin type.

*Streptococci spp.*, *Pediococci spp.* and certain *Lactobacilli spp.* are all classified as LAB, but have the ability to produce other fermentation end-products than lactic acid from pyruvate by the glycolytic pathway. Products such as acetate, CO<sub>2</sub>, ethanol and 2,3-butanediol, among others, can be detected when glucose availability is low (Thomas, Ellwood & Longyear, 1979). In a heterofermentative process, the end-products formed depend on the type of hexose substrate utilized and on the bacterial species. Lactate, ethanol and acetate are, however, normally predominating (McDonald, Henderson & Heron, 1991).

### Badly fermented silage

Badly fermented silage is often characterized by high concentrations of butyric acid and/or ammonia, and can have a pH between 5 and 7 (or higher). Presence of butyric acid and ammonia in silage is normally a result of the activity of clostridia or enterobacteria (McDonald, Henderson & Heron, 1991). The clostridial species found in silages usually belong to one of three phenotypically different groups, described by Pahlow *et al.* (2003) as; 1) proteolytic clostridia producing ammonia, amines, acetic acid and butyric acid from peptides and amino acids. This group includes *Clostridium sporogenes* and *C. bifermentans*, which have a limited ability to ferment carbohydrates; 2) the *C. butyricum*-group, which ferments monosaccharides to mainly butyric acid and acetic acid; and 3) the *C. tyrobutyricum* group, which can use a limited number of carbohydrates, but mainly ferments lactic acid to butyric acid even when pH is low. The latter group is the most studied clostridia in silage due to its economic impact in the dairy industry, causing late blowing of hard cheese. *C. tyrobutyricum* has also been found to be the most common clostridial species in Swedish big bale silage (Jonsson, 1990). The studies of clostridial species isolated from silage have been based on classic microbiology rather than modern DNA-based techniques. Therefore, the species reported should be looked upon as phenotypic groups, rather than strict taxonomic species (Pahlow *et al.*, 2003).

The growth of clostridia in silage has been shown to depend on  $a_w$  and DM content (Hengeveld, 1983), the relation between  $a_w$  and DM content in different crops, the amount of WSC in relation to DM level, buffering capacity of the crop, degree of laceration,  $\text{NO}_3$ -concentration of the crop, epiphytic micro-flora, ensiling technology and the use of additives (Wieringa, 1958; Weissbach, Schmidt & Hein, 1974; Spoelstra, 1990; Pauly, 1999). The problem with prediction of clostridial growth in silage is that silage is a heterogenous material. Spoelstra (1990) showed that the magnitude of variation in pH and DM content in a clamp silo was much smaller than the magnitude of variation in counts of clostridial spores in the same samples. Thus, clostridial growth in micro-niches might not have a very large influence on the chemical quality of silage (microbiological niches can exist at distances of about 10  $\mu\text{m}$ ), and may not be very well correlated to the counts of clostridial spores, but the effect of clostridial growth (or of other microbial species), may seriously affect the hygienic quality of the forage (Spoelstra, 1990; Pahlow *et al.*, 2003). Long-stemmed bale silage is even more heterogenous than chopped clamp silage, and niches of clostridial growth with high levels of spores, butyric acid and ammonia have been found in bales where the general DM level would indicate restricted clostridial growth (Pauly, 1999).

The main concern of feeding badly fermented silage to horses is health disturbances. Strictly proteolytic clostridia such as *C. botulinum* are rarely found in silage (Notermans, Kozaki & van Schothorst, 1979; Spoelstra, 1981), but can produce the lethal neurotoxin botulin under certain conditions (Roberts, 1988; Hatheway, 1989). Notermans, Kozaki & van Schothorst (1979) showed that toxin production by *C. botulinum* in grass took place only at  $a_w \geq 0.94$  at pH 6.5 and 5.8.

At pH 5.3, toxin production (with grass as a substrate) was demonstrated only at  $a_w \geq 0.985$ . Case reports of feed-related botulism in horses often fail to show the presence of botulinum neurotoxin in affected animals or in suspected feed or water, due to difficulties with sampling and analytical methods, as very small amounts ( $LD_{50}/kg$  body weight 0.5–2.0 ng i.p. in mice and guinea pigs) of the toxin is lethal (Gill, 1982). Also, the clinical signs usually appear three to seven days after ingestion of the toxin, at which time there may be no detectable toxin in the serum or gut content of the horse (Blood *et al.*, 1979) and most often no feed left to sample. A number of published case reports have therefore based the botulism diagnosis on clinical symptoms. A clear connection between feeding silage and incidents of botulism does not seem to be evident in the literature. Rather, inclusion of cadavers or a general poor feed hygiene in feedstuffs of different types seems to be involved. In cases of botulism where silage was fed, the forage was reported as being badly fermented with a high pH (Haagsma *et al.*, 1990), with inclusion of cadavers (Gudmundsson, 1997) or with a high pH and a strong smell of ammonia (Ricketts *et al.*, 1984). In cases where silage was not fed, inclusion of cadavers in oat chaff (Kelly *et al.*, 1984) or alfalfa hay cubes (Kinde *et al.*, 1991), grass clippings subjected to heating (Switzer *et al.*, 1984), feed-through dirt from a rack where only green oat hay had been fed (Heath *et al.*, 1990), feed and water contaminated with carcasses *via* birds as vectors (Schoenbaum *et al.*, 2000) and round bale hay stored outdoors resulting in rotten and mouldy material in the bale centre (Hunter *et al.*, 2002), were all reported as sources of the neurotoxin.

The other group of bacteria associated with high levels of ammonia in silage is enterobacteria. Enterobacteria found in silages are Gram-negative bacteria which are facultatively anaerobic and have catalase activity as well as  $NO_3^-$ -reducing ability (Spoelstra, 1987; Heron, Wilkinson & Duffus, 1993). The fermentation products from enterobacteria, using glucose as a substrate in anaerobic environments, are acetic and formic acid as well as ethanol and butanediol, depending on the bacterial species (Pahlow *et al.*, 2003). Ethanol and butanediol are not desirable as they do not contribute to a decrease in pH. The number of enterobacteria can increase during wilting of grass crops (Spoelstra, 1987) and during the first days of ensiling (Heron, Wilkinson & Duffus, 1993), but after four (Östling & Lindgren, 1995) to nine (Heron, Wilkinson & Duffus, 1993) days, they are usually not present in the silage. Also, Byrne *et al.* (2002) did not find any detectable levels of enteric bacteria in grass after 19 days of ensiling, and *Escherichia coli* O157:H7 added at harvest did not survive the ensiling process. Although the species of enterobacteria most frequently found in silages are considered to be non-pathogenic, they can contain endotoxins in the outer cell membrane, which may be associated with health problems in dairy cows (Lindgren, 1991). Also, van Duijkeren, van Aasten & Gaastra (2000) found F17-fimbriae positive *E. coli* genes and haemolytic *E. coli* strains only in faeces from diarrhoeic horses, but not in faeces from healthy horses. However, if F17-positive or haemolytic *E. coli* were responsible for the diarrhoea in the horses was not known.

## **Tocopherols and carotenes in forage in relation to equine nutrition**

Specific factors influencing the content of tocopherols (vitamin E) and carotenes (provitamin A) in wrapped forages have not been investigated to any large extent. Vitamin data in equine feed tables is largely based on vitamin contents in hay and, to a lesser extent, silo silage. As wrapped forages are gaining popularity as feeds for horses (*e.g.* Holmquist & Müller, 2002), knowledge of the influence of forage conservation methods on vitamin content is important. Blakley & Bell (1994) reported that a low vitamin A and E status in horses was associated with winter feeding of harvested feeds. It has also previously been reported that a diet consisting of dried hay and oats does not contain sufficient amounts of vitamin A and E to maintain adequate serum levels of these vitamins in weanlings and pregnant mares during the winter period (Mäenpää, Koskinen & Koskinen, 1988). It is known that airtight storage of moist barley (280 g water/kg) can lower the total amount of vitamin E, compared to airtight storage of grain at a lower water content (200 g water/kg) and compared to heat-dried grain (Hakkarainen, Työppönen & Bengtsson, 1983 a, b). Haylage is also preserved largely due to airtight storage, but it is not known if a reduction in total vitamin E, similar to the reduction in grain, takes place in haylage. Ballet, Robert & Williams (2000) showed in a review that the levels of different vitamins in forages were highly variable, and that the average (and range) of  $\alpha$ -tocopherol and  $\beta$ -carotene content was generally lower in hay than in silage. As both  $\alpha$ -tocopherol and  $\beta$ -carotene are destroyed by oxidation, ultraviolet radiation and heat (Seshan & Sen, 1942; Sullivan, 1973), different forage conservation methods may influence the content of these vitamins differently. Bauernfeind (1980) suggested that  $\alpha$ -tocopherol was more stable in an acid than in an alkali environment, and that it was relatively stable to heat and light in the absence of oxygen.

There are eight forms of vitamin E found naturally in plants, of which  $\alpha$ -tocopherol has the widest distribution and the highest vitamin E activity in animal tissue (Aitken & Hankin, 1970). Pagan, Kane & Nash (2005) showed that vitamin E from a natural source was more effective than synthetic vitamin E at elevating plasma  $\alpha$ -tocopherol levels in horses. This should be both a physiological and economical incentive to preserve vitamin E levels in conserved forages close to the initial levels of the fresh crops. Saastamoinen & Juusela (1993) found a seasonal variation in serum vitamin E content in horses. Serum levels of vitamin E was highest during grazing, and decreased during a subsequent indoor period when horses were fed timothy hay, oats and a supplement containing 1 mg vitamin E/kg bodyweight. Vitamin E is an important anti-oxidant in the blood (Moffarts *et al.*, 2005), and is vital for the humoral immune response in horses (Baalsrud & Øvernes, 1986). Diseases responsive to vitamin E and selenium supplementation, such as muscular dystrophy (Wilson *et al.*, 1976; Ronéus, 1982) and a form of degenerative disease of the spinal cord and brainstem (equine degenerative myeloencephalopathy) (Mayhew *et al.*, 1987), exist in foals and young horses. In horses 2 years of age or older, a neurodegenerative disorder



of the somatic lower motor neurons (equine motor neuron disease) can be prevented by supplementing the stable diet with vitamin E, or by providing access to green forage (Divers *et al.*, 2006).

$\beta$ -carotene in green forage is the main source of vitamin A (retinol) to grazing animals, although other carotenes may also be nutritionally important (Britton & Goodwin, 1973).  $\beta$ -carotene is mainly associated with reproductive function in mares (Peltier *et al.*, 1997), function of mucous membranes, cell differentiation at tissue growth and normal function of retinal cells (Lewis, 1996). Saastamoinen & Juusela (1992) found no seasonal variation of serum retinol in horses that were grazing or subsequently kept indoors, where they were fed timothy hay, oats and different amounts of a supplement containing vitamin A. On the other hand, Blakley & Bell (1994) found higher plasma levels of retinol in horses from May to August, compared to other times of the year, and Mäenpää, Koskinen & Koskinen (1988) also found higher serum retinol levels in mares and foals when they were on pasture compared to when kept and fed indoors. Excess intake of carotene from pasture has not been reported to result in toxicity in horses (NRC, 1989), but excess feeding of vitamin A (retinol) supplements have been reported to result in *e.g.* bone fragility, teratogenesis, loss of hair and epidermis, ataxia and depression (Donoghue *et al.*, 1981; NRC, 1989). Naturally occurring  $\beta$ -carotene in forage is therefore a safer (and less expensive) source of vitamin A than supplements, but information on the content of  $\beta$ -carotene in wrapped forages is scarce. The influence of different forage conservation methods on carotene content in grass has been reported to vary from an 80% reduction after slow field-drying of hay (Sullivan, 1973) to less than 30% in well-preserved and rapidly acidified silage (Watson & Nash, 1960; Shahane & Mungikar, 1991).

### **Factors influencing forage intake and preference in horses**

Ensiling has been reported to reduce the voluntary intake of forages in horses (McLean *et al.*, 1995; Moore-Colyer & Longland, 2000), and in sheep and cows (Buchanan-Smith, 1990; Erdman, 1993). However, in comparative studies of voluntary intake in horses, forages conserved as hay, haylage and silage had very different neutral detergent fibre (NDF) contents and different nutritive values (McLean *et al.*, 1995; Moore-Colyer & Longland, 2000), making it difficult to know if the nutrient content (plant maturity and botanical composition) or the conservation method caused the differences in intake. Voluntary dry matter intake (VDMI) of forage is probably partly related to the NDF content of the forage, but other factors are clearly involved, as prediction equations based on NDF content have been reported to have  $r^2$ -values of 0.11 (Dulphy *et al.*, 1997) to 0.50 (Lawrence, Lawrence & Coleman, 2001). Other factors influencing voluntary intake of silage may be acetic acid concentration, as foals have been reported to reject drinking water containing more than 0.16 ml acetic acid/100 ml water (Randall, Schurg & Church, 1978). Austbø (1990) also reported that horses rejected clamp silage when it had a noticeable smell of butyric acid.

Voluntary intake and preference do not measure the same thing (Forbes, 1986; Lawrence *et al.*, 1987; LaCasha *et al.*, 1999), and the relation between voluntary intake and preference is unknown. Opinions about horse preferences for different forages exist, but the scientific knowledge in the area is scarce. Reports have mainly dealt with voluntary intake and digestibility of forage (Austbø, 1990; Smolders, Steg & Hindle, 1990; Istasse *et al.*, 1996; Moore-Colyer & Longland, 2000; Bergero, Peiretti & Cola 2002), effects of forage on behaviour (Goodwin, Davidson & Harris, 2002) or effect of plant species on voluntary intake and digestibility (Darlington & Hershberger, 1968; LaCasha *et al.*, 1999). Studies concerning horse preference for forages have, to the authors' knowledge, mainly covered pasture (Archer 1971, 1973, 1978; Naujeck, Hill & Gibb, 2004). The digestibility and nutritive value of a forage are important feed attributes, but only if the forage is not rejected by the horse. When voluntary intake of single forages is studied, the results give no information of the preferences when horses are given multiple choices. One example of this is the experiment of Lawrence *et al.* (1987), who studied intake of dried or acid treated (0.80 propionic acid and 0.20 acetic acid) lucerne (*Medicago sativa*) hay in horses. When horses were given each forage separately, there was no intake difference among treatments, but when given both forages simultaneously, horses consumed more of the dried hay. These "Cafeteria" type tests with more than one feed at a time may therefore be a method to gain better knowledge of active feed choices made by the horse, than studies of one feed at a time.

### **Factors influencing the ecosystem in the equine hindgut**

The equine hindgut is inhabited by a number of different microorganisms using different substrates and producing mainly microbial cells, gases, VFA, ammonia and some lactate (Hintz, Argenzio & Schryver, 1971; Argenzio, Southworth & Stevens, 1974; Udén & van Soest, 1982; de Fombelle *et al.*, 2003). de Fombelle *et al.* (2003) demonstrated the presence of microorganisms all along the digestive tract of horses, but the number of cellulolytic bacteria and VFA production was highest in the hindgut. Changes in the microbial community and biochemistry in the hindgut (including faeces) of horses have been associated with different types of colic (Reeves, Salman & Smith, 1996), laminitis (Garner *et al.*, 1978; Goodson *et al.*, 1988; Rowe, Lees & Pethick, 1994; Milinovich *et al.*, 2006; van Eps & Pollitt, 2006), treatment with antibiotics (Kropp, 1991), equine intestinal clostridiosis (Wierup, 1977) and change of faeces' appearance, bad coat and lowered performance (Ronéus *et al.*, 1993). The microbial and fermentation profiles of both caecum and right ventral colon have been found to be directly related to the composition of the diet in studies examining different ratios of hay to concentrates (Kern *et al.*, 1973; Moore & Dehority, 1992; de Fombelle *et al.*, 2001; Julliand *et al.*, 2001; Medina *et al.*, 2002). The microbial profiles of faeces in horses on early or late summer pasture were also found to differ (Darby *et al.*, 1995). Corresponding knowledge of the influence of hay, haylage and silage on the hindgut fermentation in horses is lacking. The changes taking place in the forage during conservation, such as the increase in VFA and lactic acid content, decrease in pH and change in WSC fraction in silage, and sometimes also haylage,

affects not only the forage itself, but may also have an influence on digestion and hindgut fermentation in the horse.

## **Aims of the thesis**

The general aim of the thesis was to investigate production and storage of small bale grass silage and haylage, and to study the influence of forage conservation method on chemical and microbiological variables in small bale silage and haylage, on horse preference and on equine hindgut fermentation. The specific objectives were:

- To investigate the possibility to use high-density hay balers for production of small bale silage and haylage, and to study the resulting fermentation pattern and microbial composition in the small bales
- To investigate stability of chemical and microbiological variables during long-term storage of small bale silage and haylage
- To investigate the influence of extent of fermentation on tocopherol and carotene content in small bale silage and haylage
- To investigate horse preferences for silage, haylage and hay
- To investigate the influence of silage, haylage and hay on hindgut fermentation and microbiology in horses

## **Materials and methods**

This thesis is based on five experiments. Three of the experiments (**I–III**) focused primarily on production and storage of small bale silage and haylage, and influence of DM on fermentation and chemical composition (including tocopherols and carotenes in Paper **III**) of the forage. In Paper **IV**, horse preference for grass conserved as hay, haylage and silage was investigated. In Paper **V**, hindgut fermentation in horses fed hay, haylage and silage was studied. The experiments were mainly conducted at Kungsängen Research Centre, Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, Uppsala. Analyses of tocopherols and carotenes (**III**) were performed at Department of Animal Health, Welfare and Nutrition, Danish Institute of Agricultural Sciences, Research Centre Foulum, Tjele, Denmark. Experiments with horses in Paper **V** were done at Établissement National d'Enseignement Supérieur Agronomique de Dijon, France. Details of experimental, analytical and statistical procedures are given in the respective papers, but a general description is given below.

## Grass crops

The grass crops used in the experiments consisted mainly of timothy (*Phleum pratense*) and meadow fescue (*Festuca pratensis*). The crop in Experiment II in Paper I also contained 0.05–0.1 of perennial ryegrass (*Lolium perenne*) and 0.05 red clover (*Trifolium pratense*). Couch grass (*Agropyron repens*) was present in the crop in Papers II–V in approximate amounts of 0.1. In Paper V, dandelions (*Taraxacum vulgare*) compromised about 0.05 of the sward. The crops used in Paper II–V were first cuts of a permanent grass ley, which was fertilized with NPK 21-3-10 (430 kg/ha) before the first cut. The same crop was used for the different treatments within experiments in each paper (I–V), as the main interest was the influence of different conservation methods, and not plant maturity, botanical composition or other factors related to different crops.

## Production of small bale silage and haylage

The production of silage and haylage of different DM levels was done by varying wilting times of the cut crop before baling. In Papers I–IV, small square bales were produced directly at harvest using high-density balers intended for hay production. The high-density baler used in Paper III and V was modified by replacing the original knotters with knotters intended for a large square baler. This made it possible to use a stronger twine for binding the bales. In Paper III, a mini-round baler was also used. Wrapping of small square bales was done using a mini-wrapper with a rotating table (I–V), and small round bales were wrapped using a conventional round bale wrapper (III), also with a rotating table. In Paper V, a rebaling method was employed. The forage was harvested in big round bales, ensiled for approximately seven months and then opened and rebaled into small square bales, using the machinery equipment mentioned previously. All wrapped bales in Papers I–V were stored in a fenced bale-yard during ensiling, with care taken to protect the bales from birds and rodents. Bales were inspected regularly, and if damages of the stretch film were detected, they were immediately repaired with special plastic tape. Bales with more serious damages in the stretch film were excluded for further use in the studies.

Use of additives (chemical and biological) and number of stretch film layers (6, 8 or 10) were studied in Paper I, which consisted of two repeated experiments, with variations in crops and additive inoculants. The same chemical additive was used in Experiments 1 and 2, but the type of bacterial inoculant varied. In paper V, a LAB inoculant was added to both silage and haylage.

A method to study tightness in wrapped bales was used in Paper I and III. The method was based on the gas entry rate of bales. A small negative pressure was created inside the bale, using a one-way valve inserted through the plastic and a hand-operated vacuum pump. An injection needle connected to a manometer was then inserted through the valve membrane, and the gas entry rate was recorded as the time it took for the negative pressure to rise from -20 to -15 mm water column.

### *Aerobic storage stability*

The aerobic storage stability of silage and haylage upon exposure to air was investigated in Experiment 1 (**I**), by measuring increase in CO<sub>2</sub> in forage samples placed in a standardized aerobic environment (constant temperature 24°C). Forage samples (0.187 kg DM) were placed in identical vessels, and about 500 l of CO<sub>2</sub>-free air/kg DM was passed through the vessels. CO<sub>2</sub> was measured in the outflow air at 1 h intervals. Production of CO<sub>2</sub> indicated microbial activity in the sample (Honig, 1990), and when the level reached 10 ml CO<sub>2</sub>/l, the recording was terminated. The microbial growth during aerobic storage was also studied by quantitative determination of fungi and WSC in bales sampled at Day 6 after opening (Experiment 1, Paper **I**).

### **Preference study**

Wrapped forages used in Paper **IV** were produced as described previously for small bale silage and haylage, and hay was harvested in small square bales and barn-dried. All forages originated from the same crop and harvest. The horses used in the preference study were privately owned, of different breeds and with different previous experiences of eating silage and haylage. The horses were kept at an early autumn pasture during the experiment. The experimental model was most easily described as a Cafeteria model, where each horse had access to equal amounts (on dry matter basis) of all forages during two hours daily. The first choice, forage consumption, eating time and eating behaviour of individual horses were registered daily for five consecutive days in four periods.

### **Studying the equine hindgut**

Haylage and silage in Paper **V** were harvested as described previously, and hay was barn-dried (small square bales). The experimental horses used in Paper **V** were fistulated both in the caecum and the right ventral colon, but only the colon fistula was used for sampling. The experimental model was a crossover design where all horses were subjected to all treatments (Table 1) after a common pre-period. Horses were fed the experimental forage, salt and minerals, and had free access to water. Colon and faecal samples were taken at Day 21 in each period, and analyzed for microbial and chemical composition. Fermentation kinetics of the right ventral colon was followed during two subsequent days in each period, with sampling before (0 h) and 2, 4, 8 and 12 h after the morning meal. Samples from the kinetic study were analyzed for chemical composition, but not for microbial variables.

Table 1. *Experimental design in Paper V*

<b>Horse</b>	<b>Pre-period</b>	<b>Period I</b>	<b>Period II</b>	<b>Period III</b>
Horse G	hay	haylage	hay	silage
Horse K	hay	silage	hay	haylage
Horse P	hay	haylage	hay	silage

## Statistical analysis

The statistical methods and models used are described in detail in each paper. Variables were normally distributed except for pH in Paper **IV**. pH was therefore transformed to  $[H^+]$  before statistical analysis. Analysis of variance in Papers **I–V** was performed using SAS general linear models procedure (SAS Institute, SAS Inc. Cary, NC, USA). Correlation analysis in Papers **II–III** was performed using SAS correlation procedure, and Pearson correlation coefficients ( $\rho$ ) were calculated according to Milton (1992) as:

$$\rho = \text{Cov}(X, Y) / \sqrt{(\text{Var}X)(\text{Var} Y)}$$

SAS mixed models procedure was used in Paper **IV**, where the variables “eating time” and “forage consumption” were analysed using a multivariate repeated measurements model. In all experiments, differences were regarded as statistically significant when  $P < 0.05$ .

## Results

### Production of small bale silage and haylage

Production of small square bale silage and haylage suffered from problems with the twine at baling (**I, II, IV**), but these were minimized with the replacement of the knotters (**III** and **V**). Small square bales generally had a higher bale density than small round bales, but interactions between bale type and DM content were also present (**III**). Bale weight was found to be important for the possibility to wrap the bales correctly, as bales weighing less than 30–35 kg were pulled off the wrapper table as by the plastic film. Increasing the number of stretch film layers from six to ten resulted in higher levels of  $CO_2$  inside the bale, but gas entry rate or fermentation variables were not affected by the amount of stretch film applied (**I**).

### Fermentation pattern and microbial counts in small bale silage and haylage

Use of inoculant additives at DM levels between 350 and 600 g/kg improved lactic acid fermentation and decreased pH compared to controls in haylage in Experiment 1 and in silage in experiment 2 (**I**). Haylage generally showed less signs of lactic acid fermentation and had higher pH than silage, but levels of butyric acid and ammonia were low (**I–V**). Ethanol content was higher than lactic

acid content in control haylage in Experiment 1 (I), in both types of haylage in Paper II–IV and in haylage in Paper V (Table 2). Concentration of ethanol and lactic acid was similar in silage in Paper III (Table 2). However, ethanol concentration was higher in silage than in haylage in Paper II–V, but not in Paper I (Experiment 2). Water-soluble carbohydrate contents were generally higher in haylages than in silages (I–V). The WSC fraction differed between forages in Paper V, where the fresh crop for hay contained half the concentration of fructans found in the fresh crops for silage and haylage. This difference was the opposite in the preserved forages, where hay had the highest fructan content (V). Mould counts and enterobacteria were generally higher in hay than in silage or haylage (IV, V). Clostridial spore counts were higher in silage than in haylage only in Paper III.

### *Aerobic storage stability*

Additives prolonged aerobic storage stability in Experiment 1 (I), as control forage reached 10 ml CO<sub>2</sub>/l in outflow air earlier than forages treated with inoculants or chemicals. Additive-treated bales also expressed lower mould counts than control bales after being open for six days. Bales treated with the chemical additive had the highest WSC content both at Day 0 (opening day) and at Day 6 after opening in Experiment 1 (I), and the WSC content at Day 6 did not diverge to any large extent from WSC content at Day 0.

Table 2. Content of dry matter, ethanol, lactic acid and pH in silage and haylage in Papers I–V

Paper	Forage type/treatment	Dry matter g/kg	Ethanol g/kg DM	Lactic acid g/kg DM	pH
I <sup>1</sup>	Haylage (control, Exp.1)	544	19.3	4.9	6.07
	Silage (control, Exp.2)	339	4.5	30.4	5.19
	Haylage (control, Exp.2)	496	4.6	12.5	5.46
II <sup>2</sup>	Silage	306	26.9	38.3	4.65
	Haylage (HLL)	565	13.8	2.0	5.55
	Haylage (HLH)	672	8.6	0.4	5.72
III <sup>3</sup>	Silage	285	32.8	32.9	5.16
	Haylage (HL1)	505	21.8	3.1	5.51
	Haylage (HL2)	564	17.3	1.4	5.51
IV <sup>4</sup>	Silage	309	22.8	31.8	4.94
	Haylage (HLL)	577	7.9	2.6	5.63
	Haylage (HLH)	684	4.3	0.3	5.81
V <sup>5</sup>	Silage	343	9.7	43.0	4.44
	Haylage	548	5.9	1.3	5.60

<sup>1</sup>Data from Table 2 (Experiment 1), Table 3 and Table 4 (Experiment 2) in Paper I, <sup>2</sup>From Table 2 in Paper II, <sup>3</sup>From Table 1 in Paper III, <sup>4</sup>From Table 3 in Paper IV, <sup>5</sup>From Table 2 in Paper V.

### **Storage of small bale silage and haylage**

Long-term storage of small square bales generally resulted in lower pH-value, 2,3-butanediol and WSC concentration, and higher content of lactic and acetic acid, ethanol, ammonia-N and yeast counts, compared to short-term storage (II).

Interactions between forage type and storage time were present, as silage responded differently to long-term storage than either of the two haylage types. pH, succinic acid and 2,3-butanediol were lower and lactic and acetic acid higher after long- than after short-term storage in silage. The same variables of both haylage types were unaffected by long-term storage. It was observed that silage bales deviated from a square shape, while haylage bales were rectangular (II). A reduction in WSC content during storage was similar (about 20 g WSC/kg DM) for all forage types (II).

### **Tocopherols and carotenes in small bale silage and haylage**

The DM level of grass conserved as silage or haylage did not have any general linear effect on the content of  $\alpha$ -tocopherol or  $\beta$ -carotene in the preserved forages (III). However, haylage with lower DM level (500 g DM/kg) contained less  $\alpha$ -tocopherol and  $\beta$ -carotene than silage (300 g DM/kg) and haylage with higher DM level (600 g DM/kg). Interactions between DM treatment and bale type (round or square) were found for several variables, and  $\alpha$ -tocopherol and  $\beta$ -carotene content were highest in silage in square bales. Proportions of  $\alpha$ -tocopherol and  $\beta$ -carotene in preserved forages in relation to initial content in the fresh crop were highest in silage in square bales and high DM haylage in square and round bales, and were about 0.6 for  $\alpha$ -tocopherol and 0.85 for  $\beta$ -carotene. Weak linear correlations were found between fermentation variables and  $\alpha$ -tocopherol or  $\beta$ -carotene, of which lactic acid had the highest Pearson correlation coefficients (positive for both  $\alpha$ -tocopherol and  $\beta$ -carotene).

### **Horse preference for hay, haylage and silage**

In Paper IV, silage, haylages (two different DM levels) and hay from the same crop had different chemical and microbial compositions with the exception of ash and crude protein content and counts of yeast and clostridial spores. Horses preferred silage when given the opportunity to choose simultaneously between hay, haylages and silage (IV). Eating time was longest and consumption highest for silage, and silage was never left in favour of any other forage. In 72 out of 84 times, silage was the first chosen forage. Hay had the shortest eating time and lowest consumption, and was never completely consumed. The haylages were intermediate between hay and silage in eating time and amount consumed. At the end of each observation period, horses alternated between the forages more frequently than at the beginning.

### **Composition of hindgut content in horses fed hay, haylage and silage**

Hay, haylage and silage produced from the same crop had different chemical composition except content of acid detergent fibre, *in vitro* digestible organic matter, estimated metabolizable energy and counts of yeasts and clostridial spores (V). The different forages did not produce different responses in microbial or



chemical composition in colon or faeces of fistulated horses sampled at Day 21 (V). Colon samples were generally lower than faecal samples in DM content, counts of lactobacilli and lactate utilizing bacteria, and higher in pH, acetic, propionic and butyric acids as well as in total organic acids. When haylage was fed, fermentation kinetics in colon was different for acetic acid, valeric acids and total organic acids, which concentrations were lower compared to when hay and silage was fed. Also, feeding hay resulted in higher DM, *i*-butyric and *i*-valeric acid concentration in the kinetic study, but differences were small and the proportions of acetate:propionate:butyrate:valerate to total organic acids in colon were similar among forage treatments. The content of WSC in colon samples from the kinetic study was close to zero. There were general differences, although very small, between sampling times in the kinetic study, as butyric and valeric acids were highest at 0 h, but no interaction between forage type and sampling time, meaning that all forages behaved similarly in the colon at the different sampling times. Small differences among horses were observed in chemical variables in colon both at Day 21 and in the kinetic study, indicating some individual variation in colon fermentation in horses.

## Discussion

### Production of small bale silage and haylage

The modification of the knotters in the high-density baler was a simple and low-cost way to improve the possibility to use this type of machine for baling crops with a lower DM content than hay (III, V). The number of stretch film layers only influenced the amount of CO<sub>2</sub> inside the bales, and a difference existed only between 6 and 10 layers (I). As CO<sub>2</sub> in silage is produced during plant respiration and the following fermentation, the higher CO<sub>2</sub>-concentration in bales wrapped with 10 layers of stretch film may indicate a tighter seal, as suggested by Paillat & Gaillard (2001). In Finland, Heikkilä *et al.* (2002) found lower yeast and mould counts in bales with six layers of white stretch film compared to four layers, but there was no difference between six and eight layers. O’Kiely, Forristal & Lenehan (2000 a) compared 2, 4 and 6 layers of stretch film on big round bale silage. An improvement in digestibility and less surface rotting and mould development were found when the number of layers was increased from 2 to 4, but only a smaller improvement from 4 to 6 layers. No differences in yeast or mould counts were found between 6, 8 and 10 layers of stretch film in Experiment 1 (I). Black stretch film was used by O’Kiely, Forristal & Lenehan (2000 a), and as black stretch film has been shown to have a higher CO<sub>2</sub>-permeability than white stretch film, due to different absorption of radiation (Möller, Klaesson & Lingvall, 1999), studies using different film colours may not be directly comparable. Paillat & Gaillard (2001) also stressed the fact that the stretch film could react quite differently in different climates.

The bale density was generally higher in square than in round bales (III), but interactions between bale type and DM content were also present. Bosma (1981) found a large impact of crude fibre content in the crop on porosity or density (kg DM/m<sup>3</sup>) under laboratory conditions. The higher the crude fibre content, the lower the density. Plant maturity stage, chop length and moisture content have also been found to influence silage density (Bosma, 1981; Honig, 1987). Honig (1987) found that the higher the DM and the coarser the grass, the higher consolidation was needed to keep the gas flow at a constant level. At 200 g DM/kg, a density of 160 kg DM/m<sup>3</sup> and at 400 g DM/kg, a density of 225 kg DM/m<sup>3</sup> was needed to keep the gas flow in the grass at the same level. A high gas flow may facilitate oxygen penetration (Williams, 1994) and subsequent microbial growth. In haylages in Papers I–V, the DM level was higher than 400 g/kg and density was seldom larger than 170 kg DM/m<sup>3</sup> in small bales. Still, mould growth was low in all haylages (I–V). This supports the findings of O’Kiely, Forristal & Lenehan (2000b), who reported that neither bale density nor degree of wilting were important for avoidance of mould growth when a sufficient number of stretch film layers was used to exclude oxygen-containing air from the forage. The same conclusion about wilting was made by Undersander, Wood & Foster (2005), who showed that a DM range from 370 to 770 g/kg in lucerne or lucerne-grass mixtures was acceptable for preserving the crop as silage or haylage, if sufficiently thick plastic (more than 13µm in total) was used, and bales were wrapped within 24 hours after baling.

### **Influence of conservation method on forage composition**

The fermentation pattern differed between silage and haylage treatments in all experiments in this thesis. The fermentation was more extensive in silage than in haylage, as indicated by a higher concentration of lactic acid and a lower pH in silage (I–V). The haylages generally had higher concentration of ethanol than lactic acid (I–V) except in Experiment 2 (I), and silage in Paper III contained equal amounts of ethanol and lactic acid. Ethanol fermentation in silage or haylage is not desirable as it does not contribute to a decrease in pH, and as CO<sub>2</sub> is produced during ethanol formation, DM losses occur (McDonald, Henderson & Heron, 1991). Ethanol in silage can be produced during heterolactic fermentation by LAB (Buyze, Vanden Hamer & De Haan, 1957), by yeasts (Jonsson & Pahlow, 1984), by enterobacteria and by proteolytic clostridia through Stickland reactions (Pahlow *et al.*, 2003). The production of ethanol in high-dry matter grass silage (400–500 g/kg) was discussed by Driehuis & van Wikselaar (1996, 2000), who suspected yeast to be the responsible organism. Driehuis & van Wikselaar (2000) also found a positive correlation between ethanol and WSC content in silages, which indicated that the ethanol producing microorganism was more osmotolerant than epiphytic LAB, and that a substrate other than WSC was used for ethanol formation. O’Brien *et al.* (2007 b) reported the yeast species *Pichia anomala* and mucoraceous moulds to be positively correlated with ethanol content in baled silage, and also that a higher DM level favoured *P. anomala* and mucoraceous moulds. Yeast counts were found to increase in wilted material (up to 500 g DM/kg), but ethanol content did not follow the increase in DM or yeast counts,

but rather tended to change in opposite direction (Jonsson *et al.*, 1990). In Papers **II–III**, yeast counts seemed to be higher in silage and haylage containing more ethanol than lactic acid. However, mixed fermentations with both LAB and yeasts have also been reported to occur, typically when crops rich in WSC are used for ensiling purposes (Ashbell *et al.*, 1987).

Driehuis *et al.* (1997) found that high DM silages (i.e. haylage) contained much lower amounts of ethanol when inoculated with LAB than when not. Adding osmotolerant inoculants to high-DM crops could therefore be a way to enhance the rate of lactic acid production and pH decline, and to avoid ethanol formation. In Paper **I**, where two different inoculants were used, fermentation in haylage in Experiment 1 was similar to the results of Driehuis *et al.* (1997), but in Experiment 2, content of lactic acid, ethanol and pH were not different between control and inoculated haylage.

Production of ethanol in silage and haylage can also occur as a result of plant enzyme activity in the initial stages of anaerobiosis. In absence of oxygen, glycolysis produces ethanol, lactic acid and alanine from pyruvate as a consequence of inhibition of the respiratory chain. Pyruvate decarboxylase is the initial enzyme in the reaction and has an acid pH optimum. If accumulation of organic acids occurs during initial stages, as proposed by McDonald, Henderson & Heron (1991), pH is reduced and pyruvate decarboxylase activated. However, haylages in Papers **I–V** were heavily wilted, and as plant respiration generally decreases and ceases with increasing DM content (Wylam, 1953; Sullivan, 1973), plant enzymes seem less likely as the main cause of ethanol formation.

Ethanol can be rapidly absorbed from the gastrointestinal tract, but the horse appears to have an unusually high concentration of alcohol dehydrogenase in the liver (Chapman & Rudram, 1978). These authors administered 0.57 and 1.14 g ethanol/kg body weight using a nasogastric tube and found the highest ethanol content in the blood 1 h after administration, irrespective of the amount infused into the stomach. The mean rate of ethanol elimination from the blood was 6.3 mg/100 ml/h. Kristensen *et al.* (2007) concluded that alcohol concentrations typical in corn silage (4 to 14 g ethanol and 1 to 6 g propanol per kg DM), did not show any indication of saturation of hepatic alcohol metabolism in dairy cows fed the silages, but that ruminal metabolism and post-ingestion blood levels of alcohols could be affected. Silage in Papers **II** and **III** also contained 2,3-butanediol in amounts close to the ethanol content. Studies on utilization of 2,3-butanediol by horses have, to the author's knowledge, not been published, but Mathison, Fenton & Milligan (1981) concluded that feed intake, growth rate and diet digestibility in sheep were not affected by a 5% inclusion of 2,3-butanediol to a hay diet.

Content of WSC were generally higher in haylages than in silages (**I–V**), which was in agreement with the results of Gordon *et al.* (1961), Finner (1966) and Nicholson *et al.* (1991). The higher WSC content in haylage could be explained by a generally lower microbial activity compared to silage. The differences in the WSC fraction among the preserved forages in Paper **V** could be explained by

respiration and hydrolysis during wilting and ensiling (Wylam, 1953; Sullivan, 1973). Content of sucrose and fructans was lower and glucose and fructose higher or similar in preserved forages compared to their corresponding fresh crops, which indicated a degradation of sucrose and fructans to hexoses during storage. This degradation was larger in silage and haylage than in hay (**V**). Lundén Pettersson & Lindgren (1990) found values of glucose, fructose, sucrose and fructans in wilted fresh grass similar to the values in the fresh crop in Paper **V**, and also that the main changes in WSC, both during wilting and ensiling, was seen in the sucrose and fructan fractions.

The baled forages in Papers **I–V** contained low levels of lactic acid and had higher pH-values than usually found in chopped silages stored in silos, which is in accordance with previous studies (Haigh, 1990; Field & Wilman, 1996; Slotner & Bertilsson, 2006). However, the wrapped forages could not be considered as badly fermented, as levels of butyric acid and ammonia-N were low, or badly preserved, as mould counts were low. pH has previously been reported to differ between bale silage and silo silage. Field & Wilman (1996) used data from 5650 clamp silages and 820 baled silages, and found that pH was generally higher in bales than in clamps at similar DM levels. Also, Haigh (1990) made an on-farm survey of big bale silages, and showed that total lactic and acetic acid concentrations were lower and pH 0.8 units higher than in comparable bunker silages. This indicates that intervals for quality variables used for conventional silo silage cannot be applied directly to baled forages. Also, Wieringa (1958) found the correlations established between pH, butyric acid, ammonia and lactic acid in wet silage not to be applicable to heavily wilted silages. The equation of Weissbach (1968), relating pH to DM for avoidance of clostridial (butyric acid) fermentation, was valid for silages containing 150 to 500 g DM/kg. Thus, these equations can not be directly applied to haylage. This is in accordance with Muck (1988), who stated that fermentation was not as important in silages with higher DM levels (>550 g/kg, i.e. haylage) to prevent general growth of clostridia, as clostridial activity was inhibited at these DM contents (Wieringa, 1958; Leibensperger & Pitt, 1987).

Hengeveld (1983) found a decrease in clostridial spore counts in silage when tedding frequency was increased from once to twice per day during wilting. This is in accordance with the results of Paper **III**, where clostridial spore counts decreased with increased number of mechanical field treatments and increasing dry matter level. In Papers **I**, **II**, **IV** and **V**, there were no differences in clostridial spore counts between DM treatments. Enterobacterial counts were higher in hay than in wrapped forages (**IV**, **V**), but were present also in silages and haylages (**II–V**). The number of enterobacteria usually decreases during the first days of ensiling (Heron, Wilkinson & Duffus, 1993; Östling & Lindgren, 1995), but can increase during wilting of grass crops (Spoelstra, 1987).

Mould counts were generally higher in hay than in silage or haylage (**IV**, **V**) but no differences in mould counts between silage and haylage were detected (**I–V**). Due to the detrimental effects of mould spores on equine airways (e.g. Robinson *et al.*, 1996), and the presence of mycotoxins in mouldy forages (O'Brien *et al.*, 2006), care should be taken to inhibit mould growth in feeds. Roquefortin C,

mycophenolic acid and andrastin A were found in amounts of up to 20 mg/kg, and roquefortines A, B and D, festuclavine, marcfortine A and agroclavine in amounts of 0.1–5 mg/kg, in visibly mouldy bale silage (O'Brien *et al.*, 2006). Although effects of the different mycotoxins have not been studied to any large extent in animals, detrimental influence on health cannot be ruled out (Scudamore & Livesey, 1998; Wilkinson, 1999). Intoxication of horses by both *Aspergillus spp.* and *Penicillium spp.* has been reported (Aller, Edds & Asquith, 1981; Asquith, 1991; Vesonder *et al.*, 1991; Ocholi *et al.*, 1992; Barnett *et al.*, 1995).

### *Influence of epiphytic microflora on fermentation*

As the majority of horses in Sweden are used for “leisure riding” and have comparably low requirements for energy and protein (NRC, 1989), silage and haylage crops are cut rather late in the season, compared to silage harvested for lactating dairy cows. Cutting the crop at a late maturity stage also results in the standing crop being inhabited by a different epiphytic microflora than at an early cut (Behrendt, Müller & Seyfarth, 1997). This might influence the direction of fermentation and the hygienic quality of the preserved forage. The epiphytic microflora on fresh crops consists mainly of LAB, enterobacteria and fungi (Lacey, 1989; Pahlow, 1991; Behrendt, Müller & Seyfarth, 1997). Pahlow (1991) presented data showing the influence of cutting date on the composition of the epiphytic microflora on grass, and found late cuts to result in increased counts of LAB, enterobacteria, clostridia and moulds but not yeasts. Lacey (1989) also found filamentous fungi to develop during plant growth, especially during senescence and ripening. Behrendt, Müller & Seyfarth (1997) found higher numbers of heterotrophic bacteria and filamentous fungi on a late first cut crop than on a regrowth crop cut at the same time (mid-July). The microbial populations were generally characterized by a low diversity, and the largest numbers of taxa were found on late cut material (mid-July). Enterobacteria were found particularly on overmature grass, did not show up until the end of May and were higher in numbers on the late first cut crop than on the regrowth crop. The dominating species among enterobacteria was *Pantoea agglomerans* (teleomorph *Erwinia herbicola*) (Behrendt, Müller & Seyfarth, 1997). In Papers **I**, **II**, **IV** and **V**, microbial counts of LAB on the fresh crop were within normal ranges (Pahlow & Dinter, 1987) or lower, and enterobacteria counts were higher than any other microbial variable in the fresh crop (**II**, **IV**, **V**), probably reflecting the epiphytic flora.

The type of sward used for silage or haylage production may also influence the fermentation outcome. Swards commonly used for forage production at small horse farms in Sweden are extensively managed, old, permanent grass leys also used for grazing. Keating & O’Kiely (2000) found differences in conservation characteristics between *Lolium spp.* swards managed for intense silage production and old, permanent grass swards. Silage from permanent grass swards was higher in pH and had lower ratios of lactic acid to acetic acid + ethanol, compared to silage from *Lolium spp.* swards. The sward used in Papers **II–V** was a permanent grass ley, and this might have contributed to the higher ethanol and lower lactic acid content in the forages.

Wilting and pre-treatment of the crop may also influence the epiphytic microflora. Pahlow & Dinter (1987) found that the number of LAB/g fresh matter of the harvested crop ranged from log 2.85–6.78 for both grass and maize, that chopping increased and wilting (in most cases) reduced the number of LAB (wilted to 330 g/kg within 24 h). Muck (1987) found that the major factors correlated to the number of LAB (on chopped lucerne) were wilting time, average wilting temperature and the rate of drying. A slow drying rate resulted in higher LAB numbers at longer wilting times. The same author also found that the greatest rate of LAB growth occurred during the first day of wilting. Wilting for extended periods during wet weather conditions may also increase the number of enterobacteria (Beck, 1965). In this thesis, only Paper V allowed a proper comparison of fresh crops within an experiment, but no effect of wilting on numbers of LAB or enterobacteria was found. However, enterobacteria compete with LAB and can produce the undesired compounds ethanol and butanediol during fermentation (Pahlow *et al.*, 2003). Also, enterobacteria usually die off in silage due to presence of lactic and acetic acid and a lowering of the pH (Heron, Wilkinson & Duffus, 1993; Östling & Lindgren, 1993, 1995). In baled haylage, lactic acid contents were very low and pH rarely below 5.40 (I–V). The influence of high counts of enterobacteria on the fresh crop on direction of fermentation in baled haylage may therefore be an area requiring further study.

#### *Aerobic storage stability*

Haylage may heat very rapidly after opening, due to a high content of residual WSC (Savoie & Jofriet, 2003). Therefore, it may be important to inhibit microbial activity in haylage during aerobic storage, especially if big bales are used in small stables where the number of horses is low. When oxygen diffuses into the silage, lactate-assimilating yeasts grow and a gradient of pH, lactic acid as well as other acids, and oxygen is created from the silage surface, producing heat and niches with micro-environments suitable for different micro-organisms (Jonsson, 1991). Using additives prolonged aerobic storage stability in opened bales (I). The reason for this was probably the presence of sodium benzoate and sodium propionate in the additives. In order for sodium benzoate to be effective against fungi, pH should be lower than 6 (Woolford, 1975), which was the case in Paper I. Some LAB inoculants may also have the ability to reduce aerobic deterioration. Ström (2005) used *Lactobacillus plantarum* MiLAB 393 as an inoculant in silage, and found it to inhibit spoilage yeast. The precise mode of inhibition was not found, but synergy between lactic acid and other antimicrobial substances produced by MiLAB 393 probably explained the anti-fungal activity. The antifungal properties of the inoculants used in Papers I and V were not known.

Compaction of the silage has been regarded as a factor having a major influence on the aerobic stability. Less dense silage has higher oxygen diffusion rates (Rees, Audsley & Neale, 1983), as oxygen penetration depends mainly on diffusion and volumetric flow (McGechan & Williams, 1994). Oxygen entering the forage allows growth of aerobic microorganisms (Jonsson, 1991). However, Hoxey & Billington (1987) found no differences in aerobic deterioration for low dry matter

silage with different compaction levels. For high dry matter silages, the aerobic deterioration was limited to a region close to the silage face, but compaction reduced oxygen penetration and aerobic deterioration (Hoxey & Billington, 1987). Thus, to minimize aerobic deterioration in baled haylage after opening, a high bale density and use of additives at ensiling (I) may be helpful.

### **Storage of small bale silage and haylage**

There was a risk of poor bale density (I, III) or deviations from a square bale shape (II) in small bale silage and haylage. As long as the stretch film layers provide an airtight seal, bale density is less important for storage stability (O’Kiely, Forristal & Lenehan, 2000 b). However, bales with an uneven shape may be difficult to wrap with a proper 50% overlap of the layers which in turn could provide a way for air to enter. Also, bales with a tendency to lose shape, such as low DM and/or low density bales may be more subjected to damage during handling (Randby & Fyhri, 2005). Disruption of the stretch film layers could impair storage stability, and in such cases, bale density may be important for storage stability. In Paper II, silage bales deviated from a rectangular form, and silage was the only treatment where mould counts were above detection level. However, mould counts in Paper II were on average very low. If the deviating bale shape seriously impaired tightness of the wrapping, mould counts would probably have been much larger, especially in long-term stored bales.

There is little information on losses in baled silage during storage (Savoie & Jofriet, 2003). The model of aerobic fungal growth in silage developed by Muck, Pitt & Leibensperger (1991) and Pitt, Muck & Pickering (1991), and the model of air infiltration losses by McGechan & Williams (1994), were predominantly based on information from silage stored in conventional silos, which differs from baled silage in both fermentation variables (Haigh, 1990; Field & Wilman, 1996; Fychan & Jones, 1996) and density (Ruxton & Gibson, 1995). Bales also have a much higher surface to volume ratio than silos (Forristal & O’Kiely, 2005), resulting in larger volumes of forage being close to the surface in bales. The sensitivity of baled silage during storage was demonstrated by McNamara *et al.* (2002), who investigated bird damages on wrapped silage bales in Ireland. McNamara *et al.* (2002) found that even small stretch film damage could result in quantitative DM losses due to mould growth, especially in high DM silages. The extent of mould growth in baled silage in Ireland was studied by O’Brien *et al.* (2005, 2007 a), who confirmed that unsatisfactory storage conditions and insufficient amounts of stretch film layers caused mould growth in bales. O’Brien *et al.* (2005) found 40% of all bales examined (on 35 farms) to have visible damages in the stretch film, and 90% of the bales had visible mould growth. It was also found that well-managed bales contained lower counts of mould propagules than normal on-farm managed bales. In well-managed bales, *P. roqueforti* spores and visible mould growth was absent, in contrast to on-farm managed bales (O’Brien *et al.*, 2007 b). In Paper II, storage of small square bales for 14 months generally resulted in changes in fermentation parameters, WSC content and yeast counts, and silage was more prone to change than haylage. No increase in mould growth was

detected during storage in any of the forages, but yeast counts were generally higher in long-term stored bales. O'Brien *et al.* (2007 b) and Jonsson *et al.* (1990) both found *Candida lambica* (teleomorph *Pichia fermentans*) and *Hansenula anomala* (teleomorph *Pichia anomala*) to be the most common yeast species in baled silage. These genera were also the dominating yeast species in silage where air was allowed to penetrate during fermentation (Jonsson & Pahlow, 1984).

### **Tocopherols and carotenes in preserved forage**

The reduction in tocopherol and carotene contents in silage and haylage in Paper **III** was not clearly associated with the extent of fermentation, but positive linear correlations were strongest between lactic acid and  $\alpha$ -tocopherol, and between lactic acid and  $\beta$ -carotene. This may indicate that lactic acid fermentation was more important than the extent of fermentation (or DM level) itself for preservation of  $\alpha$ -tocopherol and  $\beta$ -carotene in grass. Thus, the larger reduction in total vitamin E content associated with decreasing DM contents in airtight stored grain (Hakkarainen, Työppönen & Bengtsson (1983 a, b) was not evident for grass silage and haylage (**III**).

The results of Paper **III**, as well as the variation reported in the reviews of Ballet, Robert & Williams (2000) and of Nozière *et al.* (2006), indicated that the conservation method was not the primary determinant of  $\alpha$ -tocopherol and  $\beta$ -carotene content in forage. The content of  $\alpha$ -tocopherol probably depends to a larger extent on plant maturity, since mature plants have lower leaf:stem ratio than young plants (Miller, 1958), and leaves contain higher levels of  $\alpha$ -tocopherol than stems (Brown, 1953; Horvath *et al.*, 2006). Also, the botanical composition of the crop influences the amount of carotenes in forages. Grasses have been shown to have lower losses of  $\beta$ -carotene than lucerne and red clover during wilting, probably due to lower levels of an unidentified aerobic oxygenase enzyme in grasses (Kalač & Kyzlink, 1980), as well as lower leaf losses during harvest. For determination of factors affecting both  $\alpha$ -tocopherol and  $\beta$ -carotene levels in forage, the influence of harvest methods and wilting need further attention.

Some, but not all, of the forages in Paper **III** contained sufficient amounts of  $\alpha$ -tocopherol to theoretically satisfy the vitamin E requirements of horses at maintenance. Exercise, growth, pregnancy and lactation require additional vitamin E (NRC, 1989), and the content of  $\alpha$ -tocopherol in the conserved forages might be too low if forage constitutes the entire ration. All treatments contained sufficient amounts of  $\beta$ -carotene to, in theory, cover the requirement of provitamin A for all categories of healthy horses (NRC, 1989).

### **Influence of forage conservation method on preference of horses**

The horses' preference for silage in Paper **IV** was clear, although the reason for this preference was unknown. One cause may be the lower DM level, making silage resemble pasture grass in its physical appearance. Waring (1974) observed horses that had developed a "hay wetting procedure", in which the horses soaked



mouthfuls of hay in their water supply before chewing. The reason for this behaviour was not explained, but Waring (1974) also mentioned the resemblance of wet hay to fresh grass as a possible reason. The silage in Paper **IV** had a slightly lower NDF content than the other forages and this might have played a role in the preference of the horses. Although preference and VDMI do not measure the same thing, it should be noted that correlations between NDF content and VDMI in horses are poor (Dulphy *et al.*, 1997; Cuddeford, 2005). Also, there was no difference in NDF content between hay and any of the haylages, but eating time and consumption differed between the same forages (**IV**). Silage and hay contained equal levels of *in vitro* digestible organic matter and estimated metabolizable energy, whereas both of the haylages contained approximately 97 % of the same variables in silage and hay. Therefore, NDF concentration, *in vitro* digestible organic matter or metabolizable energy content was not considered to have had any major impact on preference.

Comparisons of studies investigating horse preference are problematic, as the number of published studies is few. The area is also difficult to interpret and understand, partly because preference is dependent on the available alternatives. Studies of feral horses in Wyoming (Crane, Smith & Reynolds, 1997), Camargue (Duncan, 1992) and Alberta (Salter & Hudson, 1979) reported that abundance and succulence (measured as percent moisture) of graminoid vegetation in proximity to preferred habitats seemed to be primary factors influencing habitat selection. Also, the diet composition followed seasonal availability of different plant species, but proportional intake of different species was fairly constant between seasons (Salter & Hudson, 1979; Duncan, 1992; Crane, Smith & Reynolds, 1997) with a few exceptions reported by Duncan (1992). Horses are selective but do not consume only the preferred feed but survey their environment and move back and forth between different feed alternatives both at pasture (Archer, 1971, 1973, 1978; Naujeck, Hill & Gibb, 2004) and when fed different preserved feeds indoors (Goodwin, Davidson & Harris, 2002). This behaviour was also seen in Paper **IV**, as horses did not ignore the less preferred forages, but were observed to alternate between the forages, even though a clear preference for silage existed.

A lower voluntary intake of clamp silage compared to big bale silage, haylage and hay, was reported by Moore-Colyer & Longland (2000), who found ponies to consume only about half the amount of clamp silage compared to the other forages (on a DM basis). However, the forages used in the study had substantial differences in nutritive value (CP content was 154, 111, 70 and 44 g/kg DM in clamp silage, big bale silage, haylage and hay respectively), and were therefore not suitable for comparing the effects of different conservation methods. McLean *et al.* (1995) compared hay and clamp silage fed to horses and also found lower voluntary intakes of the silage, but the nutritive content of the forages used differed greatly, as hay contained 728 and silage 540 g NDF/kg DM. Austbø (1990) used the same crop for production of clamp silage, big bale haylage and hay, and found a lower intake and larger feed refusals of silage, but only when it had a noticeable smell of butyric acid. When the silage had a pleasant smell, intake was not different from hay or haylage. Rejection of butyric acid-containing silage has also been reported in ruminants (Waldo, Keys & Gordon, 1972; Demarquilly

& Dulphy, 1977). The silage in Paper **IV** had very low concentration of butyric acid, and there was no general rejection of silage by the horses. The results of Paper **IV** showed that different forage conservation methods can influence horse preference for forage. This finding can perhaps contribute to future studies of factors that regulate forage intake in horses, which at present is limited (Cuddeford, 2005).

Individual preferences existed as there were interactions between horses and forages in Paper **IV**. Individual variation in selectivity has previously also been reported among horses on pasture (Marinier & Alexander, 1991). This individual variation, together with the influence of previously fed diets, constitutes additional difficulties in studying preference. Confounding effects from previous feeds were reported by LaCasha *et al.* (1999), who measured VDMI and digestibility of three different forages in a changeover study. At the end of the experiment, horses were given simultaneous access to the three forages fed previously, and were found to consume less of the forage they had been fed immediately before. How long this memory of a feed lasts is uncertain, making it difficult to take even a known feeding history of an animal into account.

### **Influence of forage conservation method on hindgut fermentation in horses**

There were no differences in chemical or microbial composition of colon content or faeces in horses fed hay, haylage and silage. This showed that the forage conservation methods employed did not affect hindgut fermentation differently (**V**). Hindgut fermentation values were comparable to a previous study where horses were fed hay with a similar CP content (Hintz, Argenzio & Schryver, 1971). In comparison with horses fed timothy hay (Kern *et al.*, 1974) or pasture grass and hay (Mackie & Wilkins, 1988), acetate was lower and propionate higher in colon samples in Paper **V**. It should however be noted that the crop used in Paper **V** was harvested early in the season and contained twice as much CP compared to the hay used by Kern *et al.* (1974). Crops harvested early contain less indigestible and more readily digestible components, which may explain the somewhat higher propionate percentage in the colon fluid in Paper **V**. Propionate and valerate concentrations were found to increase in caecal fluid of horses when grain was added to forage diets (Hintz, Argenzio & Schryver, 1971; Kern *et al.*, 1973). This indicated that some of the grain reached the caecum and provided a substrate for microbial fermentation (Crawford *et al.*, 1968).

Small differences in hindgut fermentation variables between individual horses were noted in Paper **V**. However, these differences were not considered to have had any large influence on the results. The differences were probably a sign of individual variation in hindgut fermentation among horses. Individual variations have previously been reported to be larger in horses compared to ruminants and rabbits, when measuring digesta particle size (Udén, 1982), digestibility (Udén & Van Soest, 1982) and retention time of liquid (CoEDTA) and solid (Cr) digesta markers (Udén *et al.*, 1982). It should also be noted that caecal cannulation has

been reported to increase total tract mean retention time of both hay and concentrates compared to before cannulation (Pulse, Baker & Potter, 1973; Austbø & Volden, 2006). The horses in Paper V were cannulated in both the caecum and right ventral colon, which have previously been reported to lower the total tract mean retention time (Drogoul, Poncet & Tisserand, 2000). The mean retention time could affect the extent of digestion and fermentation of the feed (Van Weyenberg, Sales & Janssens, 2006), and therefore studies using fistulated horses can produce different results than studies using euthanized animals.

In the kinetic study of the colon in Paper V, no evident pattern of cyclic changes in VFA or pH was found, but minor irregular differences in pH, total organic acids and individual acids were present between sampling times. As there were no interactions between forage type and sampling time, all forages had similar fermentation kinetics in the right ventral colon. The amount of total and individual VFA in the colon at a given time is the result of production and absorption of VFA. Argenzio, Southworth & Stevens (1974) studied *in vitro* transport of VFAs in isolated mucosa from the large intestine of horses, and found that although individual acids were present in equal 30 mM concentrations in the lumen, they were transported to the blood side in the order of acetate > propionate > butyrate. Acetate was transported to the blood side at a greater rate than propionate or butyrate, but the absorption ( $\mu\text{mol}/\text{cm}^2$ ) of acetate from the lumen bath was not different from absorption of propionate or butyrate. Mucosa of caecum and colon absorbed VFA at approximately equivalent rates (Argenzio, Southworth & Stevens, 1974). The net appearance and disappearance of VFA in the equine large intestine has also been correlated with cyclic changes of  $\text{Na}^+$  in the same compartment (Argenzio and Stevens, 1975).

Lactate levels in colon and faeces were low in Paper V. Alexander, MacPherson & Oxford (1952) and Alexander & Davies (1963) reported that lactic acid was not commonly found in colon liquid. However, the anaerobic Gram-negative cocci *Veillonella gazogenes* was frequently isolated from intestinal sites, and this cocci was found to produce gas and VFA only with lactate as substrate (Alexander, MacPherson & Oxford, 1952). The reason for the low lactate levels in equine colon in Paper V could thus have been due to conversion of lactate to propionate, as has been reported in the rumen of sheep (Mackie, Gilchrist & Heath, 1984). However, these authors found only 0.025 of the total VFA to come from lactate on a high-forage diet low in readily fermentable carbohydrates. Also, the propionate concentration and propionate proportion of total organic acids were not exceptionally high in Paper V. Alexander & Davies (1963) found the highest amount of lactic acid in the stomach of horses, but could not show that lactic acid was absorbed in the small intestine as the lactate concentration was sometimes higher in the caudal ileum compared to in the cranial ileum. Argenzio, Southworth & Stevens (1974) and de Fombelle *et al.* (2003), on the other hand, found decreasing concentrations of lactic acid from the stomach and along the small intestine in horses fed different diets, and in the large intestine, lactic acid had almost completely disappeared. Both lactate-producing and lactate-utilizing organisms were present in different compartments of the alimentary canal. The dominance of lactic acid in the stomach, and of VFA in the hindgut, may represent

different environments or substrate availability favouring different microorganisms (Alexander & Davies, 1963; de Fombelle *et al.*, 2003). In Paper V, the number of lactate utilizing bacteria in the colon was close to log 7 CFU/ml, and the number of lactate producing bacteria was about log 6 CFU/ml, which were both considered as normal levels (Julliand, 2005). Chamberlain, Thomas & Anderson (1981) found that protozoa played a central role in metabolism of lactic acid in the rumen of sheep fed grass silage diets. The role of protozoa in the equine hindgut is, however, not very well understood (Ike *et al.*, 1983; Moore & Dehority, 1992).

Cellulolytic bacteria were a minor component of total anaerobic bacteria in colon (V), which was in accordance with Kern *et al.* (1973) and Julliand (2005). *Ruminococcus flavefaciens* has been identified as the predominating cellulolytic species in pony and donkey caecum (Julliand *et al.*, 1999), together with *R. albus* and *Fibrobacter succinogenes* (Julliand, 2005). Major end-products from cellobiose fermentation by isolated strains of *R. flavefaciens* from equines were comparable (with some deviations) to corresponding rumen strains, except for ethanol production, which was larger in the equine strain, and malate and fumarate, which were not detected from isolated equine species of *R. flavefaciens* (Julliand *et al.*, 1999). *In vitro* fermentation end-products of the equine fungi *Piromyces citronii*, found in caecum of ponies and donkeys, were formate, acetate and ethanol from fermentation of glucose and cellobiose, in contrast to rumen strains of the fungi, which also produced lactate (Julliand *et al.*, 1998).

Using faecal samples as indicators of the status in the colon of forage-fed horses were applicable for lactic acid, *i*-butyric acid, valeric acids, total anaerobic bacteria, cellulolytic bacteria and streptococci (V). However, as concluded by da Veiga, Chaucheyras-Durand & Julliand (2005), more information of the influence of different diets on caecal, colonic and faecal ecosystems is needed before faeces can be regarded as a substitute for caecal and colon samples in horses. However, if variables in faeces diverge strongly from normal, they may serve as indicators of upsets in the alimentary canal. Milinovich *et al.* (2006) and van Eps & Pollitt (2006) induced laminitis in horses using an overload of fructo-oligosaccharides administered via nasogastric tubing, which resulted in a faecal pH of 4 to 5 in conjunction with watery diarrhoea. In these cases, faecal pH and consistency can clearly be regarded as a sign of a disturbed gut function.

#### *Digestion of water-soluble carbohydrates in forages*

Although there were differences in the water-soluble carbohydrate fraction between hay, haylage and silage, these differences were not manifested in colon samples (V). The very low concentrations of glucose, fructose and sucrose in colon (V), can be explained by the presence of brush border disaccharidases in the small intestine and the transport of glucose and fructose across the small intestinal membrane (Roberts, 1975; Dyer *et al.*, 2002). Varloud *et al.* (2004) also reported that “sugar and starch” in different diets were digested to about 0.90 in samples taken from the caecum of horses. Fructan content was also very low in colon samples in Paper V. The fructans were probably degraded prior to right ventral

colon, as the glycosidic linkage between C2 and C6 ( $\beta$ -2,6-linkage) in the furanosyl units is regarded as very unstable in acid environments (Smith, 1973), such as in the pyloric part of the equine stomach (Frape, 2004). Temperate grasses such as Timothy and Meadow fescue contain fructans mainly of the levan type, which consists of  $\beta$ -2,6-linked fructose units and a sucrose residue (Chatterton *et al.*, 1990; Suzuki, 1993; Cairns *et al.*, 1999). Very small amounts of cereal fructans ( $\beta$ -2,1-linked) have been found to be absorbed in the small intestine of rats (Nilsson *et al.*, 1988), but in fistulated pigs fed  $\beta$ -2,1-linked fructan (inulin), more than half of the oral dose had disappeared prior to the caecum (Böhmer, Branner & Roth-Maier, 2005). In horses, Coenen, Mößeler & Vervuert (2006) argued evidence for microbial digestion of inulin (from Jerusalem artichoke) pre-caecally through measurement of fermentative gases in the horses' breath. Also, Respondek *et al.* (2005) were unable to recover any short-chain fructo-oligosaccharides (SCFOS,  $\beta$ -2,1-linked) from the digesta in the stomach and small intestine of horses fed a diet of straw, concentrates and added SCFOS when horses were sampled two hours after feeding. Pre-caecal microbial digestion of SCFOS and pre-colonic digestion of grass fructans in horses thus seems likely to exist, and could be supported by the findings of de Fombelle *et al.* (2003), who demonstrated the presence and activity of microorganisms along the entire gastrointestinal tract of horses, and of Yuki *et al.* (2000) who reported the presence of *Lactobacillus spp.* in the non-secreting area of the horse stomach.

It has recently been reported that equine laminitis can be experimentally induced by an orally administered overload of  $\beta$ -2,1-linked inulin from chicory roots (van Eps & Pollitt, 2006; Milinovich *et al.*, 2006). Berg *et al.* (2005) demonstrated lower pH and higher lactate and VFA content in faeces from horses receiving increasing amounts of a  $\beta$ -2,1-linked SCFOS in the feed. However, a connection between an overload of  $\beta$ -2,1-linked SCFOS and presence of  $\beta$ -2,6-linked levans in grass in their abilities to induce laminitis or disturbances in the gastrointestinal tract has not yet been established. Bailey, Marr & Elliot (2004) also argued that the induction of laminitis by different models may not precisely mirror naturally occurring laminitis. If a connection between the fructan types were to be proven, the reduced fructan levels in silage or haylage (V) would render these forages more suitable than hay to horses prone to develop laminitis. On the other hand, no difference in fructan content in the colon was found between horses fed hay, haylage or silage (V). It should also be noted, that the lowest dose of fructans used to induce laminitis was 7.5 g/kg body weight (van Eps & Pollitt, 2006). The highest fructan content found in Paper V was 34 g/kg DM in the fresh crop used for silage. To reach the dose 7.5 g fructans/kg body weight by eating this forage, a horse weighing 500 kg would have to consume 110 kg DM. Thus, the levels of fructans present in the fresh crops or preserved forages in Paper V do not seem to be sufficient to be able to induce laminitis in horses.

## Main conclusions

- Small bale silage and haylage production is possible using high-density hay balers, but modification of the knotting mechanism may be necessary. Ethanol can be a common fermentation product in small bale silage and haylage, and can exceed the content of lactic acid.
- Long-term storage (14 months) of small silage bales resulted in changes in fermentation variables compared to short-term storage (2 months), but values at 14 months were well correlated with values at 2 months. Small haylage bales were not affected by the same storage time.
- Content of  $\alpha$ -tocopherol and  $\beta$ -carotene in small bale silage and haylage was not clearly associated to the extent of fermentation, but correlated to lactic acid content.
- Different forage conservation methods had an impact on horse preference in favour of silage. Silage had the longest eating times and highest consumption, followed by haylage and hay in descending order.
- Conservation of forage as hay, haylage or silage did not produce different responses in microbial or chemical composition in the hindgut of fistulated horses fed the forages.

## Populärvetenskaplig sammanfattning

Det har länge varit tradition att utfodra hästar med vallfoder i form av hö. På senare år har dock ensilage och hösilage ( $\geq 50$  % torrsbstanshalt (ts)) i inplastade balar i viss utsträckning ersatt hö i hästfoderstaterna. Det kan finnas flera anledningar till detta, men en orsak är sannolikt svårigheten att producera och lagra hö torrt. Torr lagring av hö är en förutsättning för att undvika tillväxt av mögel. Förekomst av mögelsporer i foder och strömedel är förknippat med sjukdomen Recurrent Airway Obstruction (RAO), eller ”kwickdrag” som den också kallas. RAO är en kronisk luftvägssjukdom som innebär att hästens lungor inte har samma kapacitet som lungorna hos en helt frisk häst, vilket påverkar hästens prestationsförmåga negativt. Ensilage och hösilage innehåller i allmänhet färre mögelsporer än hö, och både skörd och lagring är mindre väderberoende än skörd och lagring av hö.

Inplastat vallfoder produceras vanligen i stora runda eller fyrkantiga balar (ca 400-700 kg). Problemet med stora balar i hästsammanhang är att de flesta hästar finns på gårdar eller i stall med ett fåtal djur. För ett sådant stall innehåller stora ensilage- och hösilagebalar i allmänhet för mycket foder för att de skall hinna utfodras innan fodret blivit skämt, särskilt under de varmare årstiderna. Små hästgårdar utan mekaniserad balhantering kan också ha problem med att hantera

storbalar. Ensilage och hösilage i mindre balar kan därför vara ett intressant alternativ, men det finns i dagsläget inga kommersiellt tillgängliga balpressar som är anpassade för en sådan småbalsproduktion. Kunskapen om hösilage och ensilage med avseende på fodrets användbarhet och inverkan på digestionskanalen hos hästar är också begränsad. Syftet med de studier som avhandlingen grundar sig på var därför att närmare studera både produktion av småbalar med tillgängliga metoder, och användning av inplastat vallfoder i hästfoderstater.

I den första studien (**I**) undersöktes om det var möjligt att använda en konventionell höpress (glidkolvspress) för att producera ensilage (35 % ts) och hösilage (50-60 % ts) i småbalar. I försöket studerades också om det var nödvändigt att använda ensileringsmedel (biologiska och kemiska) för att få en bra hygienisk kvalitet på fodret, om balarnas hygieniska kvalitet var stabil efter öppning och hur många lager plast (6, 8 eller 10) som behövdes för att balarna skulle vara täta. Resultaten visade att ensileringsmedel inte var nödvändigt för att få en acceptabel hygienisk kvalitet, men att det förlängde hållbarheten på fodret efter öppning av balarna. Antalet plastlager hade främst effekt på mängden koldioxid inne i oöppnade balar, vilken var högst med 10 lager plast. Det fanns en del praktiska problem med att få tillräckligt hårt pressade balar utan att äventyra pressens knytmekanism, och utan att riskera att balbanden gick sönder direkt vid pressning. I studie **III** och **V** löstes detta problem genom att byta ut pressens originalknytare till sådana knytare som normalt sitter i stora fyrkantsbalpressar. På så sätt gick det att använda starkare och grövre pressgarn, och balarna kunde pressas hårdare och få högre baldensitet ( $\text{kg ts/m}^3$ ) utan att balbanden gick sönder.

För att undersöka om småbalarna var hållbara över en längre lagringsperiod, och om den kemiska och mikrobiologiska sammansättningen i fodret förändrades över tiden, gjordes en lagringsstudie med provtagning av balarna 2 och 14 månader efter inplastning (**II**). Denna studie gjordes på ensilage (35 % ts), hösilage med låg ts-halt (55 %) och hösilage med hög ts-halt (70 %), som producerats från samma vall och samma skörd. Resultaten visade att 14 månaders lagring av balarna påverkade alla kemiska fermentationsvariabler utom smörsyra- och etanolhalt i ensilaget, men de båda hösilagen uppvisade inte några förändringar. Av de mikrobiologiska variablerna var det bara jästantalet som ökade med lagringstiden. Även om förändringar i de kemiska fermentationsvariablerna ägde rum under lagringen överensstämde analysresultaten efter 2 och 14 månaders lagring mycket väl med varandra. Det innebar att prov som togs 2 månader efter inplastning var godtagbara för att åskådliggöra sammansättningen i fodret även efter 14 månaders lagring. Detta gäller under förutsättning att plasten bibehållits intakt, att balarna inte skadats på något sätt och att ingen smörsyrajäsnings skett.

I samband med att framför allt hösilage börjat användas i större utsträckning till hästar, har också funderingar kring vitamininnehållet i inplastat vallfoder uppkommit. Det är sedan tidigare känt att E-vitamininnehållet i lufttätt lagrat korn kan minska kraftigt under lagringen, särskilt då ts-halten är runt 72 %. Eftersom hösilage också till stor del konserveras genom lufttät lagring, uppstod frågan om innehållet av vitamin E i gräs påverkades av lagring på samma sätt som korn. Eftersom naturligt vitamin E tas upp bättre i hästens kropp än syntetiskt framställt

vitamin E, är det en god idé att försöka bevara så mycket som möjligt av det naturliga E-vitaminet i fodret. Likaså uppkom frågeställningar kring hur betakaroten, ett förstadie till vitamin A, påverkades av lufttät lagring eftersom tidigare studier visat att mängden betakaroten kan minska med upp till 80 % vid långsam torkning av gräs till hö, men att förlusterna i allmänhet är mindre än 30 % i välfermenterat ensilage. En studie av innehållet av vitamin E och betakaroten i ensilage (30 % ts), hösilage 1 (50 % ts) och hösilage 2 (60 % ts) genomfördes därför (III). Gräs ensilerades i små fyrkantiga och små runda balar som lagrades i 11 månader innan de öppnades och provtogs. Resultaten visade att ts-haltens inverkan var svår att tolka. Ensilage i fyrkantsbalar och hösilage 2 i rund- och fyrkantsbalar innehöll 60 % av den initiala mängden E-vitamin i grönmassan, och motsvarande siffra för betakaroten var 86 %. Ensilage i rundbalar och hösilage 1 i rund- och fyrkantsbalar innehöll 39 % av den initiala E-vitaminhalten, motsvarande siffra för betakaroten var 33 %. Det fanns alltså ingen tydlig generell effekt av ts-halt och fermentationsgrad (eller balform) på vitamininnehållet. Det fanns dock ett samband mellan innehållet av mjölksyra och båda vitaminerna, vilket indikerade att en bra konserveringsprocess (mjölksyrarjäsning) kan vara fördelaktig för att bevara innehållet av vitamin E och betakaroten i inplastat vallfoder. Andra faktorer än konserveringsmetoden verkar också spela större roll för innehållet av vitamin E och betakaroten i vallfoder, som t ex växternas botaniska utvecklingsstadium vid skörd, den botaniska sammansättningen av vallen och förtorkningen vid skörd.

Att använda ensilage och hösilage som hästfoder har ibland diskuterats, eftersom enstaka studier påvisat en nedgång i hästarnas konsumtion när de utfodrats med ensilage jämfört med hö. Att hästen äter sitt vallfoder är naturligtvis viktigt, eftersom fodervägran för just grovfoder kan ge mycket stora problem med störningar i mag-tarmkanalen (som tex kolik). För att undersöka om hästar hade någon preferens för vallfoder konserverade på olika sätt, genomfördes därför en studie där försökshästar fick välja fritt vilket vallfoder de ville äta (IV). För att kunna göra en sådan jämförelse var det viktigt att de foder som ingick i studien inte hade olika näringsinnehåll eller olika botanisk sammansättning. I de tidigare studierna användes foder från olika vallar och skördar, vilket gjorde det svårt att tolka resultaten. Ensilage (30 % ts), hösilage med låg ts (55 %), hösilage med hög ts (70 %) och hö (88 % ts) producerades därför från samma gräsvall och samma skörd i små fyrkantsbalar. Hästarna erbjöds sedan 1 kg ts av varje foder samtidigt, under 2 timmar per dag i 20 dagar. Hästarnas förstahandsval, hur länge och hur mycket de åt av varje foder registrerades. Hästarna åt mest av ensilaget, därefter kom hösilaget med låg ts, sedan hösilaget med hög ts och sist höet. Samma ordning gällde för ättiden och för hästarnas förstahandsval. Höet åts aldrig upp helt och hållet, och hästarna lämnade aldrig ensilaget för att äta av ett annat foder efter att ha luktat och/eller smakat på det förstnämnda. Studien påvisade alltså att konserveringsmetoden kunde inverka på hästarnas preferens för vallfoder, och att det var till ensilagens fördel, även om orsaken till denna preferens inte kunde förklaras. Detta resultat gäller för välfermenterade ensilage. Feljästa ensilage med höga halter av smörsyra och ammoniak kan inte förväntas ge samma resultat.



Torrsubstanshalten i gräs som pressas och plastas in spelar en avgörande roll för hur fodret kommer att konserveras. Grönmassa som har en låg ts-halt kommer att ensileras i större utsträckning än grönmassa med en hög ts-halt. Det beror på att mjölksyrabakterierna, som sköter om själva ensileringen, behöver vatten för att kunna bilda mjölksyra från socker i gräset. Det innebär också att ensilage innehåller mer mjölksyra och andra organiska syror, samt mindre socker och har ett lägre pH-värde jämfört med hösilage och hö. Skillnaden mellan hö och hösilage kan variera beroende på hösilagens ts-halt, men generellt sett innehåller hö mer socker och har ett något högre pH-värde än hö. Mängden mjölksyra och andra organiska syror i hösilage är vanligtvis mycket låg och skiljer sig inte nämnvärt från hö, men kan variera beroende på ts-halten och mjölksyrabakteriernas aktivitet. Hur dessa olikheter i den kemiska sammansättningen i hö, hösilage och ensilage påverkar hästens grovtarmsmiljö har inte undersökts tidigare, och därför genomfördes ett utfodringsförsök (V). Hö, hösilage (55 % ts) och ensilage (35 % ts) producerades från samma vall och samma skörd. För att kunna studera aktiviteten i hästens tarm vid utfodring med de olika vallfodren användes fistulerade försökshästar. Fisteln var placerad i högra ventralkolon och möjliggjorde provtagning av hästens grovtarmsinnehåll. Provtagning av tarminnehållet och av träck gjordes efter 21 dagars utfodring med respektive vallfoder. Dessutom utfördes en kinetikstudie, som innebar att tarminnehållet provtogs före utfodring, och sedan efter 2, 4, 8 och 12 h, för att följa fermentationsförloppet i kolon. Utfodring med de olika vallfodren gav inte upphov till några skillnader i mikrobiologiska eller kemiska variabler i varken tarminnehåll eller träck. Det var alltså inte möjligt att från proverna på tarminnehåll eller träck avgöra om hästen hade ätit hö, hösilage eller ensilage. Kinetikstudien påvisade också att de olika fodren uppförde sig likadant i kolon vid de olika provtagningstidpunkterna.

## References

- Adams, M.R. & Moss, M.O. 1995. *Food Microbiology*. The Royal Society of Chemistry, Cambridge, UK. pp. 18-54.
- Aitken, F.C. & Hankin, R.G. 1970. *Vitamins in Feeds for Livestock*. Technical Communication. No. 25, Commonwealth Bureau of Animal Nutrition. C.A.B., Farnham Royal, Bucks, England, UK.
- Alexander, F. & Davies, M.E. 1963. Production and fermentation of lactate by bacteria in the alimentary canal of the horse and pig. *Journal of Comparative Pathology* 7, 1-8.
- Alexander, F., MacPherson, M.J.D. and Oxford, A.E. 1952. Fermentative activities of some members of the normal coccal flora of the horse's large intestine. *Journal of Comparative Pathology* 62, 252-259.
- Aller, W.W., Edds, G.T. & Asquith, R.L. 1981. Effects of aflatoxins in young ponies. *American Journal of Veterinary Research* 42, 2162-2164.
- Archer, D.C. & Proudman, C.J. 2006. Epidemiological clues to preventing colic. *The Veterinary Journal* 172, 29-39.
- Archer, M. 1971. Preliminary studies on the palatability of grasses, legumes and herbs to horses. *Veterinary Record* 89, 236-240.
- Archer, M. 1973. The species preferences of grazing horses. *Journal of British Grassland Society* 28, 123-128.

- Archer, M. 1978. Further studies on palatability of grasses to horses. *Journal of the British Grassland Society* 33, 239-243.
- Argenzio, R.A., Southworth, M. & Stevens, C.E., 1974. Sites of organic acid production and absorption in the equine gastrointestinal tract. *American Journal of Physiology* 226, 1043-1050.
- Argenzio, R.A. & Stevens, C.E. 1975. Cyclic changes in ionic composition of digesta in the equine intestinal tract. *American Journal of Physiology* 228, 1224-1230.
- Ashbell, G., Pahlow, G., Dinter, B. & Weinberg, Z.G. 1987. Dynamics of orange peel fermentation during ensilage. *Journal of Applied Bacteriology* 63, 275-279.
- Asquith, R.L. 1991. Mycotoxicoses in horses. In: J.E. Smith & R.S. Henderson (Eds.). *Mycotoxins and Animal Foods*. CRC Press Inc., Boca Raton, USA. pp. 679-688.
- Austbø, D. 1990. Høy, rundballesurfor og surfor fra plansilo til hest. Fordøyelseforsøg, vannopptak og test på rullende matte (In *Norweigan*). In: *Husdyrsforsøgsmøtet Norges landbrukshøgskole*. Statens fagteneste for landbruket, nr 4. pp. 174-178.
- Austbø, D. & Volden, H. 2006. Influence of passage model and caecal cannulation on estimated passage kinetics of roughage and concentrate in the gastrointestinal tract of horses. *Livestock Science* 100, 33-43.
- Baalsrud, K.J. & Øvernes, G. 1986. Influence of vitamin E and selenium supplement on antibody production in horses. *Equine Veterinary Journal* 18, 472-474.
- Bailey, S.R., Marr, C.M. & Elliott, J. 2004. Current research and theories on the pathogenesis of acute laminitis in the horse. *The Veterinary Journal* 167, 129-142.
- Ballet, N., Robert, J.C. & Williams, P.E.V. 2000. Vitamins in forages. In: Givens, D.I., Owen, E., Axford, R.F.E. & Omed, H.M. (Eds). *Forage Evaluation in Ruminant Nutrition*. CABI Publishing, Oxon, UK. pp. 399-431.
- Barnett, D.T., Mowrey, R.A., Hagler, W.M. Jr, Bristol, D.G. & Mansmann, R.A. 1995. *The correlation of selected mycotoxins to the incidence of colic in horses*. In: Proceedings of the Fourteenth Equine Nutrition and Physiology Symposium, Ontario, California. pp. 242-247.
- Bauernfeind, J.C., 1980. Tocopherols in foods. In: Machlin, L.J. (ed.) *Vitamin E: A Comprehensive Treatise*. Marcel Dekker, New York and Basel, pp. 99-167.
- Beck, T. 1965. Investigations on the ecology and physiology of the silage microflora. *Landwirtschaftliche Forschung* 18, 243-250.
- Behrendt, U., Müller, T. & Seyfarth, W. 1997. The influence of extensification in grassland management on the populations of microorganisms in the phyllosphere of grasses. *Microbiological Research* 152, 75-85.
- Berg, E.L., Fu, C.J., Porter, J.H. & Kerley, M.S. 2005. Fructooligosaccharide supplementation in the yearling horse: Effects on fecal pH, microbial content, and volatile fatty acid concentrations. *Journal of Animal Science* 83, 1549-1553.
- Bergero, D., Peiretti, P.G. & Cola, E. 2002. Intake and apparent digestibility of perennial ryegrass haylages fed to ponies either at maintenance or at work. *Livestock Production Science* 77, 325-329.
- Billysson, F. 2002. *A survey of the feeding of horses at riding schools in southern Sweden (In Swedish)*. BSc thesis P 00/02:12. Swedish University of Agricultural Sciences, Alnarp, Sweden.
- Blackman, M. & Moore-Colyer, M.J.S. 1998. Hay for horses: the effects of three different wetting treatments on dust and nutrient content. *Animal Science* 66, 745-750.
- Blakley, B.R. & Bell, R.J. 1994. The vitamin A and vitamin E status of horses raised in Alberta and Saskatchewan. *Canadian Veterinary Journal* 35, 297-300.
- Blood, D.C., Henderson, J.A., Radostits, O.M., Arundel, J.H. & Gay, C.C. 1979. *Veterinary Medicine*. 5<sup>th</sup> ed. Baillière Tindall, Cassell Ltd., London, UK. pp. 441-442.
- Böhmer, B.M., Branner, G.R. & Roth-Maier, D.A. 2005. Precaecal and faecal digestibility of inulin (DP 10-12) or an inulin/*Enterococcus faecium* mix and effects on nutrient digestibility and microbial gut flora. *Journal of Animal Physiology and Animal Nutrition* 89, 388-396.
- Bonin, S.J., Clayton, H.M., Lanovaz, J.L. & Johnston, T. 2007. Comparison of mandibular motion in horses chewing hay and pellets. *Equine Veterinary Journal*, In press. doi: 10.2746/042516407X157792.

- Bosma, A.H. 1981. *Factors affecting the density of ensiled pre-wilted grass*. Paper No. 25. In: Sixth silage conference – silage production and utilization. (Eds. R.D. Harkess & M.E. Castle). Queen Margaret College, Edinburgh, UK. pp. 49-50.
- Bratt, M. 2001. *Möjligheterna att uppskatta hästpopulationens storlek och struktur*. Förstudie på uppdrag av Jordbruksverket (*In Swedish*). Statistiska centralbyrån, Örebro, Sweden.
- Britton, G. & Goodwin, T.W. 1973. Chlorophyll, carotenoid pigments and sterols. In: Butler, G.W. & Bailey, R.W. (Eds.), *Chemistry and biochemistry of herbage*, Vol 1. Academic Press Inc. Ltd, London, UK. pp. 502.
- Brown, F. 1953. The tocopherol content of farm feeding-stuffs. *Journal of the Science of Food and Agriculture* 4, 161-165.
- Buchanan-Smith, J.G. 1990. An investigation into palatability as a factor responsible for reduced intake of silage by sheep. *Animal Production* 50, 253-260.
- Buyze, G., Vandem Hamer, J.A. & De Haan, P.G. 1957. Correlation between hexose-monophosphate shunt, glycolytic system and fermentation type in lactobacilli. *Antonie van Leeuwenhoek* 23, 345-350.
- Byrne, C.M., O'Kiely, P., Bolton, D.J., Sheridan, J.J., McDowell, D.A. & Blair, I.S. 2002. Fate of *Escherichia coli* O157:H7 during silage fermentation. *Journal of Food Protection* 65 (12), 1854-1860.
- Cairns, A.J., Nash, R., de Carvalho, M-A. M. & Sims, I.M. 1999. Characterization of the enzymatic polymerization of 2,6-linked fructans by leaf extracts from timothy grass (*Phleum pratense*). *New Phytologist* 142, 79-91.
- Chamberlain, D.G., Thomas, P.C. & Anderson, F.J. 1981. *Lactic acid metabolism in the rumen of animals given silage diets*. Paper No. 13. In: Sixth silage conference – silage production and utilization. (Eds. R.D. Harkess & M.E. Castle). Queen Margaret College, Edinburgh, UK. pp. 25-26.
- Chapman, D.I. & Rudram, D.A. 1978. The disposition and excretion of ethanol by the horse. *Journal of Veterinary Pharmacology and Therapy* 1, 293-298.
- Chatterton, N.J., Harrison, P.A., Thornley, W.R. & Draper, E.A. 1990. Oligosaccharides in foliage of *Agropyron*, *Bromus*, *Dactylis*, *Festuca* and *Phleum*. *New Phytologist* 114, 167-171.
- Clarke, A.F. 1987. A review of environmental and host factors in relation to equine respiratory disease. *Equine Veterinary Journal* 19 (5), 435-441.
- Clarke, L.L., Roberts, M.C & Argenzio, R.A. 1990. Feeding and digestive problems in horses. *Veterinary Clinics of North America: Equine Practice* 6 (2), 433-450.
- Clevström, G. & Ljunggren, H. 1984. Occurrence of storage fungi, especially aflatoxin-forming *Aspergillus flavus* in soil, greenstuff and prepared hay. *Journal of Stored Products Research* 20, 71-82.
- Clevström, G., Göransson, B., Hlödversson, R. & Pettersson, H. 1981. Aflatoxin formation in hay treated with formic acid and in isolated strains of *Aspergillus flavus*. *Journal of Stored Products Research* 17 (4), 151-161.
- Coenen, M. 1990. Beobachtungen zum Vorkommen fütterungsbedingter Magenulcera beim Pferd (*In German*). *Schweizer Archiv für Tierheilkunde* 132, 121-126.
- Coenen, M., Mößler, A. & Vervuert, I. 2006. Fermentative gases in breath indicate that inulin and starch start to be degraded by microbial fermentation in the stomach and small intestine of the horse in contrast to pectin and cellulose. *Journal of Nutrition* 136, 2108S-2110S.
- Crane, K.K., Smith, M.A. & Reynolds, D. 1997. Habitat selection patterns of feral horses in southcentral Wyoming. *Journal of Range Management* 50 (4), 374-380.
- Crawford, B.H., Baker, J.P., Lieb, S. & Mitchell, G.E. 1968. Starch and cellulose digestion by equine intestinal microflora (Abstract). *Journal of Animal Science* 27, 1151.
- Cuddeford, D. 2005. *Voluntary food intake by equids*. In: Proceedings Equine Nutrition Conference, Hannover, 1-2 October 2005, Germany. *Pferdeheilkunde* 21, 7-8.
- Darby, N.W., Mosley, J.C., Davitt, B.B. & Bohach, G.A. 1995. Effects of diet on ungulate excretion of *Enterococcus* spp., *Streptococcus bovis* and *Streptococcus equinus* in feces. *Journal of Environmental Quality* 24, 719-724.
- Darlington, J.M. & Hershberger, T.V. 1968. Effect of forage maturity on digestibility,

- intake and nutritive value of alfalfa, timothy and orchardgrass by equine. *Journal of Animal Science* 27, 1572-1576.
- Dawson, L.E.R., Ferris, C.P., Steen, R.W.J., Gordon, F.J. & Kilpatrick, D.J. 1999. The effects of wilting grass before ensiling on silage intake. *Grass and Forage Science* 54, 237-247.
- Demarquilly, C. & Dulphy, J.P. 1977. Effect of ensiling on feed intake and animal performance. *Proceedings International Meeting on Animal Production from Temperate Grasslands*. Dublin, Ireland, pp. 53-61.
- Divers, T.J., Mohammed, H.O., Hintz, H.F. & de Lahunta, A. 2006. Equine Motor Neuron Disease: A review of clinical and experimental studies. *Clinical Techniques in Equine Practice* 5, 24-29.
- Donoghue, S., Kronfeld, D.S., Berkowitz, S.J. & Copp, R.L. 1981. Vitamin A nutrition of the equine: Growth, serum biochemistry, and haematology. *Journal of Nutrition* 111, 365-374.
- Driehuis F. & van Wikselaar P.G. 1996. *The occurrence of alcoholic fermentation in high dry matter grass silages*. In: Proceedings of the XI<sup>th</sup> International Silage Conference, Aberystwyth, Wales, UK, 1996, pp. 254-255.
- Driehuis, F. & van Wikselaar, P.G. 2000. The occurrence and prevention of ethanol fermentation in high-dry-matter grass silage. *Journal of the Science of Food and Agriculture* 80, 711-718.
- Driehuis F., van Wikselaar P.G., van Vuuren, A.M. & Spoelstra, S.F. 1997. Effect of a bacterial inoculant on rate of fermentation and chemical composition of high dry matter grass silages. *Journal of Agricultural Science Cambridge* 128, 323-329.
- Drogoul, C., Poncet, C. & Tisserand, J.L. 2000. Feeding ground and pelleted hay rather than chopped hay to ponies: 1. Consequences for *in vivo* digestibility and rate of passage of digesta. *Animal Feed Science and Technology* 87, 117-130.
- van Duijkeren, E., van Aasten, A.J.A.M & Gaastra, W. 2000. Characterization of *Escherichia coli* isolated from adult horses with and without enteritis. *Veterinary Quarterly* 22, 162-166.
- Dulphy, J.P., Martin-Rosset, W., Dubroeuq, H. & Jailler, M. 1997. Evaluation of voluntary intake of forage trough-fed light horses. Comparison with sheep. Factors of variation and prediction. *Livestock Production Science* 52, 97-104.
- Duncan, P. 1992. *Horses and Grasses*. The nutritional ecology of equids and their impact on the Camargue. *Ecological Studies*, 87. Springer Verlag, New York, USA. pp. 74-125.
- Dyer, J., Fernandez-Castaño Merediz, E., Salmon, K.S.H., Proudman, C.J., Edwards, G.B. & Shirazi-Beechey, S.P. 2002. Molecular characterisation of carbohydrate digestion and absorption in equine small intestine. *Equine Veterinary Journal* 34, 349-358.
- Erdman, R. 1993. *Silage fermentation characteristics affecting feed intake*. In: Silage Production from Seed to Animal. Proceedings National Silage Production Conference, 23-25 February 1993, Syracuse, New York, USA. NRAES 67, pp. 210-219.
- Ewart, S.L. & Robinson, N.E. 2007. Genes and respiratory disease: a first step on a long journey. Review Article. *Equine Veterinary Journal* 39 (1). In Press. doi: 10.2746/042516407X194296.
- Falk-Rønne, J., Gravesen, S., Larsen, L. & Svenningsen, J. 1984. Mikroorganismer i luften i en hestestald (In Danish). *Dansk Veterinær Tidsskrift* 67 (21), 1079-1083.
- Field, M. & Wilman, D. 1996. *pH in relation to dry matter content in clamped and baled grass silages harvested in England and Wales*. In: Proceedings of the XI<sup>th</sup> International Silage Conference, Aberystwyth, Wales, UK, 1996, pp. 126-127.
- Finner, M.F. 1966. Harvesting and handling low-moisture silage. *Transactions of the ASAE* 9, 377-381.
- de Fombelle, A., Julliard, V., Drogoul, C. & Jacotot, E. 2001. Feeding and microbial disorders in horses: 1- effect of an abrupt incorporation of two levels of barley in a hay diet on microbial profile and activities. *Journal of Equine Veterinary Science* 21 (9), 439-445.
- de Fombelle, A., Varloud, M., Goachet, A-G., Jacotot, E., Philippeau, C., Drogoul, C. &

- Julliand, V. 2003. Characterization of the microbial and biochemical profile of the different segments of the digestive tract in horses given two distinct diets. *Animal Science* 77, 293-304.
- Forbes, J.M. 1986. *The Voluntary Food Intake of Farm Animals*. Butterworths & Co. Ltd. London, UK. pp. 130-143.
- Forristal, P.D. & O'Kiely, P. 2005. *Update on technologies for producing and feeding silage*. In: Silage production and utilisation. Proceedings of the XIVth International Silage Conference, July 2005, Belfast, Northern Ireland. pp. 83-96.
- Frape, D. 2004. *Equine Nutrition and Feeding*. 3<sup>rd</sup> ed. Blackwell Publishing Ltd. UK. pp. 8.
- Fychan, R. & Jones, R. 1996. *The effect of harvesting technology on silage quality and effluent production*. In: Proceedings of the XI<sup>th</sup> International Silage Conference, Aberystwyth, Wales, UK, 1996, pp. 218-219.
- Garner, H.E., Moore, J.N., Johnson, J.H., Clarke, L., Amend, J.F., Tritschler, L.G., Coffman, J.R., Sprouse, R.F., Hutcheson, D.P. & Salem, C.A. 1978. Changes in the caecal flora associated with the onset of laminitis. *Equine Veterinary Journal* 10, 249-252.
- Gibbs, M., Dunrose, R., Bennett, F.A. & Bubeck, M.R. 1950. On the mechanism of bacterial fermentation of glucose to lactic acid studied with C<sup>14</sup>-glucose. *Journal of Biological Chemistry* 184, 545-549.
- Gill, D.M. 1982. Bacterial toxins: a table of lethal amounts. *Microbiological Reviews* 46, 86-94.
- Gohlke, P. 1957. *Aristoteles: Tierheilkunde*. 2<sup>nd</sup> ed. Paderborn, Germany. pp. 366.
- Goodson, J., Tyznik, W. J., Cline, J.H. & Dehority, B.A. 1988. Effects of an abrupt diet change from hay to concentrate on microbial numbers and physical environment in the cecum of the pony. *Applied and Environmental Microbiology* 54 (8), 1946-1950.
- Goodwin, D., Davidson, H.P.B & Harris, P. 2002. Foraging enrichment for stabled horses: effects on behaviour and selection. *Equine Veterinary Journal* 34, 686-691.
- Gordon, C.H., Derbyshire, J.C., Wiseman, H.G., Kane, E.A. & Melin, C.G. 1961. Preservation and feeding value of alfalfa stored as hay, haylage and direct-cut silage. *Journal of Dairy Science* 44, 1299-1311.
- Greenhill, W.L. 1964. Plant juices in relation to silage fermentation. III. Effect of water activity of juice. *Journal of the British Grassland Society* 19, 336-339.
- Gregory P.H., Lacey M.E., Festenstein G.N. & Siner F.A. 1963. Microbial and biochemical changes during the moulding of hay. *Journal of General Microbiology* 33, 147-174.
- Gudmundsson, S.H. 1997. Tutorial article: Type B botulinum intoxication in horses: case report and literature review. *Equine Veterinary Education* 9 (3), 156-159.
- Haagsma, J., Haesebrouck, F., Devriese, L. & Bertels, G. 1990. An outbreak of botulism type B in horses. *Veterinary Record* 127, 206.
- Haigh, P.M. 1990. The effect of dry matter content on the preservation of big bale grass silage made during the autumn on commercial farms in South Wales 1983-87. *Grass and Forage Science* 45, 29-34.
- Hakkarainen, R.V., Työppönen, J.T. & Bengtsson, G.S. 1983 a. Relative and quantitative changes in total vitamin E and isomer content of barley during conventional and air-tight storage with special reference to annual variations. *Acta Agriculturae Scandinavica* 33, 395-400.
- Hakkarainen, R.V., Työppönen, J.T. & Bengtsson, G.S. 1983 b. Changes in the content and composition of vitamin E in damp barley stored in air-tight bins. *Journal of the Science of Food and Agriculture* 34, 1029-1038.
- Han, K.J., Collins, M., Vanzant, E.S. & Dougherty, C.T. 2006. Characteristics of baled silage made from first and second harvests of wilted and severely wilted forages. *Grass and Forage Science* 61, 22-31.
- Hatheway, C.L. 1989. Bacterial sources of clostridial neurotoxins. In: Simpson, L.L. (Ed.). *Botulinum neurotoxin and tetanus toxin*. Academic Press, Inc. San Diego, California, USA. pp. 3-24.
- Heath, S.E., Bell, J.R., Chirino-Trejo, M., Schuh, J-A.C.L. & Harland, R.J. 1990. Feedthrough dirt as a source of *Clostridium botulinum* type C intoxication in a group of farm horses. *Canadian Veterinary Journal* 31, 13-19.

- Heikkilä, T., Jaakola, S., Saaritalo, E., Suokannas, A. & Helminen, J. 2002. *Effects of wilting time, silage additive, and plastic layers on the quality of round bale silage*. In: Proceedings of the XIIIth International Silage Conference, 11-13 September 2002, Auchincruive, Scotland, UK. pp. 158-160.
- Hengeveld, A.G. 1983. *Sporen van boterzuurbacteriën in kuilvoer*. Rep. 88. Proefstation voor de Runveehouderij, schapenhouderij en paardenhouderij, Lelystad, The Netherlands. In: Pahlow, G., Muck, R.E., Driehuis, F., Oude Elferink, S.J.W.H. & Spoelstra, S.F. 2003. Microbiology of ensiling. In: *Silage Science and Technology*. Eds. D.R. Buxton, R.E. Muck & J.H. Harrison. ASA, CSSA, SSSA Agronomy no. 42, Madison, Wisconsin, USA.
- Heron, S.J.E., Wilkinson, J.F. & Duffus, C.M. 1993. Enterobacteria associated with grass and silages. *Journal of Applied Bacteriology* 75, 13-17.
- Hintz, H.F., Argenzio R.A. & Schryver H.F. 1971. Digestion coefficients, blood glucose levels and molar percentage of volatile fatty acids in intestinal fluid of ponies fed varying forage-grain ratios. *Journal of Animal Science* 33, 992-995.
- Hintz, H.F., Hogue, D.E., Walker, E.F., Lowe, J.E. & Schryver, H.F. 1971. Apparent digestion in various segments of the digestive tract of ponies fed diets with varying roughage-grain ratios. *Journal of Animal Science* 32 (2), 245-248.
- Hlödversson, R. 1985. *Methods for estimating and preventing storage losses in moist hay*. Dissertation. Report 144. Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Hocking, A.D., Miscamble, B.F. & Pitt, J.I. 1994. Water relations of *Alternaria alternata*, *Cladosporium cladosporioides*, *Cladosporium sphaerospermum*, *Curvularia lunata* and *Curvularia pallescens*. *Mycological Research* 98 (1), 91-94.
- Holmquist, S. & Müller, C.E. 2002. *Problems related to feeding forages to horses*. In: Proceedings of the XIIIth International Silage Conference, 11-13 September 2002, Auchincruive, Scotland, pp. 152-153.
- Honig, H. 1987. *Influence of forage type and consolidation on gas exchange and losses in silo*. Theatre Paper 26. In: Eight Silage Conference. The AFRC Institute for Grassland and Animal Production, Hurley, Berks, UK. pp. 51-52.
- Horvath, G., Wessjohann, L., Bigirimana, J., Jansen, M., Guisez, Y., Caybergs, R. & Horemans, N. 2006. Differential distribution of tocopherols and tocotrienols in photosynthetic and non-photosynthetic tissues. *Phytochemistry* 67, 1185-1195.
- Hotchkiss, J.W., Reid, S.W.J. & Christley, R.M. 2007. A survey of horse owners in Great Britain regarding the horses in their care. Part 2: Risk factors for recurrent airway obstruction. *Equine Veterinary Journal*, In press. doi: 10.2746/042516407X180129.
- Hoxey, R.P. & Billington, R.S. 1987. *Measurements of temperature, carbon dioxide and oxygen in two 20 tonne silage bunkers: from ensiling to post opening*. Poster 22. In: Eight Silage Conference. The AFRC Institute for Grassland and Animal Production, Hurley, Berks, UK. pp. 131-132.
- Hummel, J., Südekum, K.-H., Streich, W.J. & Clauss, M. 2006. Forage fermentation patterns and their implications for herbivore ingesta retention times. *Functional Ecology* 20, 989-1002.
- Hudson, J.M., Cohen, N.D., Gibbs, P.G. & Thompson, J.A. 2001. Feeding practices associated with colic in horses. *Journal of American Veterinary Medicine Association* 219, 1419-1425.
- Hunter, J.M., Rohrbach, B.W., Andrews, F.M. & Whitlock, R.H. 2002. Round bale grass hay: a risk factor for botulism in horses. *Compendium on Continuing Education for the Practicing Veterinarian* 24 (2), 166-168.
- Ike, K., Nuruki, R., Imai, S. & Ishii, T. 1983. Composition of intestinal ciliates and bacteria excreted in feces of the racehorse. *Japanese Journal of Veterinary Science* 45 (2), 157-163.
- Istasse, L., Van Eenaeme, C., Hornick, J.L., Van Calster, P. & Huet, D. 1996. *Composition, intakes and apparent digestibility of 3 grass silages offered to horses*. In: Proceedings of the British Society of Animal Science 1996, Scarborough, UK, p. 647.
- Jackson, N. & Forbes, T.J. 1970. The voluntary intake by cattle of four silages differing in dry matter content. *Animal Production* 12, 591-599.

- Janis, C. 1976. The evolutionary strategy of the equidae and the origins of rumen and cecal digestion. *Evolution* 30, 757-774.
- Jonsson, A. 1990. Enumeration and confirmation of *Clostridium tyrobutyricum* in silages using neutral red, D-cycloserine, and lactate dehydrogenase activity. *Journal of Dairy Science* 73, 719-725.
- Jonsson, A. 1991. Growth of *Clostridium tyrobutyricum* during fermentation and aerobic deterioration of grass silage. *Journal of the Science of Food and Agriculture* 54 (4), 557-568.
- Jonsson, A., Lindberg, H., Sundås, S., Lingvall, P. & Lindgren, S. 1990. Effect of additives on the quality of big-bale silage. *Animal Feed Science and Technology* 31, 139-155.
- Jonsson, A. & Pahlow, G. 1984. Systematic classification and biochemical characterization of yeasts growing in grass silage inoculated with *Lactobacillus* cultures. *Animal Research and Development* 20, 7-22.
- Julliand, V. 2005. *Impact of nutrition on the microflora of the gastrointestinal tract in horses*. In: Lindner, A. (Ed.). Applied Equine Nutrition. 1<sup>st</sup> Equine Nutrition Conference, Hannover, 2005. Wageningen Academic Publishers, The Netherlands. pp. 85-103.
- Julliand, V., de Fombelle, A., Drogoul, C. & Jacotot, E. 2001. Feeding and microbial disorders in horses: part 3 – Effects of three hay:grain ratios on microbial profile and activities. *Journal of Equine Veterinary Science* 21 (11), 543-546.
- Julliand, V., Riondet, C., de Vaux, A., Alcaraz, G. & Fonty, G. 1998. Comparison of metabolic activities between *Piromyces citronii*, an equine fungal species, and *Piromyces communis*, a ruminal species. *Animal Feed Science and Technology* 70, 161-168.
- Julliand, V., de Vaux, A., Millet, L. & Fonty, G. 1999. Identification of *Ruminococcus flavefaciens* as the predominant cellulolytic bacterial species of the equine cecum. *Applied and Environmental Microbiology* 65 (8), 3738-3741.
- Kalač, P. & Kyzlink, V. 1980. The enzymic nature of the degradation of beta-carotene in red clover and in other forage crops during silage-making with acid additives. *Animal Feed Science and Technology* 5, 59-68.
- Keating, T. & O'Kiely, P. 2000. Comparison of old permanent grassland, *Lolium perenne* and *Lolium multiflorum* swards grown for silage: 2. Effects on conservation characteristics in laboratory silos. *Irish Journal of Agricultural and Food Research* 39 (1), 25-33.
- Kelly, A. P., Jones, R.T., Gillick, J.C. & Sims, L.D. 1984. Outbreak of botulism in horses. *Equine Veterinary Journal* 16 (6), 519-521.
- Kern, D.L., Slyter, L.L., Weaver, J.M., Leffel, E.C. & Samuelson, G. 1973. Pony cecum vs. steer rumen: the effect of oats and hay on the microbial ecosystem. *Journal of Animal Science* 37, 463-469.
- Kern, D.L., Slyter, L.L., Leffel, E.C., Weaver, J.M. & Oltjen, R.R. 1974. Ponies vs. steers: microbial and chemical characteristics of intestinal digesta. *Journal of Animal Science* 38 (3), 559-564.
- Kinde, H., Bettley, R.L., Ardans, A., Galey, F.D., Daft, B.M., Walker, R.L., Eklund, M.W. & Byrd, J.W. 1991. *Clostridium botulinum* type-C intoxication associated with consumption of processed alfalfa hay cubes in horses. *Journal of the American Veterinary Medical Association* 199 (6), 742-746.
- Kristensen, N.B., Storm, A., Raun, B.M.L., Røjen, B.A. & Harmon, D.L. 2007. Metabolism of silage alcohols in lactating dairy cows. *Journal of Dairy Science* 90, 1364-1377.
- Kropp, S. 1991. *Bakteriologische Untersuchungen zur Zusammensetzung der Darmflora des Pferdes und deren Beeinflussung durch Chemotherapeutika (In German)*. Dissertation. Tierärztliche Hochschule Hannover, Germany. pp. 11-27.
- Künzle, F., Gerber, V., Van Der Haegen, A., Wampfler, B., Straub, R. & Marti, E. 2007. IgE-bearing cells in bronchoalveolar lavage fluid and allergen-specific IgE levels in sera from RAO-affected horses. *Journal of Veterinary Medicine A* 54, 40-47.
- LaCasha, P.A., Brady, H.A., Allen, V.G., Richardson, C.R. & Pond, K.R. 1999. Voluntary intake, digestibility and subsequent selection of Matua Bromegrass, Coastal Bermudagrass and Alfalfa hays by yearling horses. *Journal of Animal Science* 77, 2766-2773.

- Lacey, J. 1989. Pre- and post-harvest ecology of fungi causing spoilage of foods and other stored products. *Journal of Applied Bacteriology* 67 (supplement), 11S-25S.
- Lawrence, A.C. St., Lawrence, L.M. & Coleman, R.J. 2001. *Using an empirical equation to predict voluntary intake of grass hays by mature equids*. Proceedings of the 17<sup>th</sup> Equine Nutrition and Physiology Symposium, Kentucky, USA. pp. 99-100.
- Lawrence, L.M., Moore, K.J., Hintz, H.F., Jaster, E.H. & Wischover, L. 1987. Acceptability of alfalfa hay treated with an organic acid preservative for horses. *Canadian Journal of Animal Science* 67, 217-220.
- Le Bars, J. & Le Bars, P. 1996. Recent acute and subacute mycotoxicoses recognized in France. *Veterinary Record* 27, 383-394.
- Leibensperger, R.Y & Pitt, R.E. 1987. A model of clostridial dominance in ensilage. *Grass and Forage Science* 42, 297-317.
- Lewis, L.D. 1996. *Feeding and care of the horse*. 2<sup>nd</sup> ed. Lippincott Williams and Wilkins, Pennsylvania, USA. pp. 42-48.
- Lindgren, S. 1991. Hygienic problems in conserved forage. In: Pahlow, G. & Honig, H. (Eds.). *Forage conservation towards 2000*. Landbauforschung Völkenrode, Sonderheft 123, Germany. pp. 177-190.
- Lundén Pettersson, K. & Lindgren, S. 1990. The influence of the carbohydrate fraction and additives on silage quality. *Grass and Forage Science* 45, 223-233.
- Mackie, R.I., Gilchrist, F.M. & Heath, S. 1984. An *in vivo* study of ruminal microorganisms influencing lactate turnover and its contribution to volatile fatty acid production. *Journal of Agricultural Science* 103, 37-51.
- Mackie, R.I & Wilkins, C. 1988. Enumeration of anaerobic bacterial microflora of the equine intestinal tract. *Applied and Environmental Microbiology* 54 (9), 2155-2160.
- Mäenpää, P.H., Koskinen, T. & Koskinen, E. 1988. Serum profiles of vitamins A, E and D in mares and foals during different seasons. *Journal of Animal Science* 66, 1418-1423.
- Marinier, S.L. & Alexander, A.J. 1991. Selective grazing behaviour in horses: Development of methodology and preliminary use of tests to measure individual grazing ability. *Applied Animal Behaviour Science* 30, 203-221.
- Mathison, G.W., Fenton, M. & Milligan, L.P. 1981. Utilization of 2,3-butanediol by sheep. *Canadian Journal of Animal Science* 61, 649-656.
- Mayhew, I.G., Brown, C.M., Stowe, H.D., Trapp, A.L., Derksen, F.J. & Clement, S.F. 1987. Equine degenerative myeloencephalopathy: a vitamin E deficiency that may be familial. *Journal of Veterinary Internal Medicine* 1, 45-50.
- McDonald, P., Henderson, A.R. & Heron, S.J.E. 1991. *The biochemistry of silage*. 2<sup>nd</sup> ed. Chalcombe Publications, Marlow, UK. pp. 52, 91-94, 272.
- McGechan, M.B. & Williams, A.G. 1994. A model of air infiltration losses during silage storage. *Journal of Agricultural Engineering Research* 57, 237-249.
- McGorum, B.C., Ellison, J. & Cullen, R.T. 1998. Total and respirable airborne dust endotoxin concentrations in three equine management systems. *Equine Veterinary Journal* 30 (5), 430-434.
- McGreevy, P.D., Cripps, P.J., French, N.P., Green, L.E. & Nicol, C.J. 1995. Management factors associated with stereotypic and redirected behaviour in the Thoroughbred horse. *Equine Veterinary Journal* 27 (2), 86-91.
- McLean, B., Afzalzadeh, A., Bates, L., Mayes, R.W. & Hovell, F.D. DeB. 1995. Voluntary intake, digestibility and rate of passage of a hay and a silage fed to horses and to cattle. *Animal Science* 60, 555.
- McNamara K., O'Kiely P., Whelan J., Forristal P.D. & Lenehan J.J. 2002. Simulated bird damage to the plastic stretch-film surrounding baled silage and its effects on conservation characteristics. *Irish Journal of Agricultural and Food Research* 41, 29-41.
- Medina, B., Girard, I.D., Jacotot, E. & Jullian, V. 2002. Effect of a preparation of *Saccharomyces cerevisiae* on microbial profile and fermentation patterns in the large intestine of horses fed a high fibre or a high starch diet. *Journal of Animal Science* 80, 2600-2609.
- Merry, R.J., Winters, A.L., Thomas, P.I., Müller, M. & Müller, T. 1995. Degradation of fructans by epiphytic and inoculated lactic acid bacteria and by plant enzymes during



- ensilage of normal and sterile hybrid ryegrass. *Journal of Applied Bacteriology* 79, 583-591.
- Milutinovic, G.J., Trott, D.J., Burrell, P.C., van Eps, A.W., Thoenner, M.B., Blackall, L.L., Al Jassim, R.A.M., Morton, J.M & Pollitt, C.C. 2006. Changes in equine hindgut bacterial populations during oligofructose-induced laminitis. *Environmental Microbiology* 8 (5), 885-898.
- Miller, D.F. 1958. Composition of cereal grains and forages. *Natl. Acad. Sci., Natl. Res. Council Pub.* 585. Washington, USA. In: Butler, G.W. & Bailey, R.W. (Eds.), *Chemistry and biochemistry of herbage*, Vol 3. Academic Press Inc. Ltd, London, UK.
- Milton, J.S. 1992. Statistical methods in the biological and health sciences. 2<sup>nd</sup> Ed. McGraw-Hill Inc., New York, USA. pp. 346-390.
- Moffarts, de B., Kirschvink, N., Art, T., Pincemail, J. & Lekeux, P. 2005. Effect of oral antioxidant supplementation on blood antioxidant status in trained thoroughbred horses. *Veterinary Journal* 169, 65-74.
- Moore, B.E. & Dehority, B.E. 1992. Effects of diet and protozoa on total and cellulolytic bacterial and fungal concentrations in the cecum and colon of the equine. *Journal of Animal Science* 70 (Supplement 1), 240.
- Moore-Colyer, M.J.S. 1996. Effects of soaking hay fodder for horses on dust and mineral content. *Animal Science* 63, 337-342.
- Moore-Colyer, M.J.S. & Longland, A.C. 2000. Intakes and *in vivo* apparent digestibilities of four types of conserved grass forage by ponies. *Animal Science* 71, 527-534.
- Muck, R.E. 1987. *Factors affecting numbers of lactic acid bacteria on lucerne prior to ensiling*. Theatre Paper 2. In: Eight Silage Conference. The AFRC Institute for Grassland and Animal Production, Hurley, Berks, UK. pp. 3-4
- Muck, R.E. 1988. Factors influencing silage quality and their implications for management. *Journal of Dairy Science* 7, 2992-3002.
- Muck R.E., Pitt R.E. & Leibensperger, R.Y. 1991. A model of aerobic fungal growth in silage. 1. Microbial characteristics. *Grass and Forage Science* 46, 283-299.
- Müller, C.E. 2002. *Small square bale silage for horses and horse-owners*. In: Proceedings of the XIIIth International Silage Conference, September 11-13, 2002, Auchincruive, Scotland, UK. pp. 330-331.
- Müller, M. & Lier, D. 1994. Fermentation of fructans by epiphytic lactic acid bacteria. *Journal of Applied Bacteriology* 76, 406-411.
- Müller, M. & Steller, J. 1995. Comparative studies of the degradation of grass fructan and inulin by strains of *Lactobacillus paracasei* subsp. *paracasei* and *Lactobacillus plantarum*. *Journal of Applied Bacteriology* 78, 229-236.
- Möller, K., Klaesson, T. & Lingvall, P. 1999. *Correlation between colour and temperature of LDPE stretch film used in silage bales*. In: The XIIth International Silage Conference: Silage production in relation to animal performance, animal health, meat and milk quality. Swedish University of Agricultural Sciences, Uppsala, Sweden. pp. 251-252.
- Naujeck, A., Hill, J. & Gibb M.J. 2004. Influence of sward height on diet selection by horses. *Applied Animal Behaviour Science* 90, 49-63.
- Nicholson, J.W.G., McQueen, R.E., Charmley, E. & Bush, R.S. 1991. Forage conservation in round bales or silage bags: effect on ensiling characteristics and animal performance. *Canadian Journal of Animal Science* 71, 1167-1180.
- Nilsson, U., Öste, R., Jägerstad, M. & Birkhed, D. 1988. Cereal fructans: *In vitro* and *in vivo* studies on availability in rats and humans. *Journal of Nutrition* 118, 1325-1330.
- Notermans, S., Kozaki, S. & van Schothorst, M. 1979. Toxin production by *Clostridium botulinum* in grass. *Applied and Environmental Microbiology* 38 (5), 767-771.
- Nourse, D.O. 1897. Silage for horses. *Bulletin no. 80. New series. Volume VI. No. 9. Virginia Agricultural Experiment Station*. Blacksburg, Montgomery Co., Virginia, USA.
- Nozière, P., Graulet, B., Lucas, A., Martin, B., Grolier, P. & Doreau, M. 2006. Carotenoids for ruminants: From forages to dairy products. *Animal Feed Science and Technology* 131, 418-450.
- NRC, 1989. *Nutrient requirements of horses*. 5<sup>th</sup> revised edition. National Research Council. National Academy Press, Washington D.C., USA. pp. 19-24.

- O'Brien, M., O'Kiely, P., Forristal, P.D. & Fuller, H.T. 2005. Fungi isolated from contaminated baled grass silage on farms in the Irish Midlands. *FEMS Microbiology Letters* 247, 131-135.
- O'Brien, M., O'Kiely, P., Forristal, P.D. & Fuller, H.T. 2007 a. Quantification and identification of fungal propagules in well-managed baled grass silage and in normal on-farm produced bales. *Animal Feed Science and Technology* 132, 283-297.
- O'Brien, M., O'Kiely, P., Forristal, P.D. & Fuller, H.T. 2007 b. Visible fungal growth on baled grass silage during the winter feeding season in Ireland and silage characteristics associated with the occurrence of fungi. *Animal Feed Science and Technology*, In press. doi: 10.1016/j.anifeedsci.2007.01.010.
- O'Brien, M., Nielsen, K.F., O'Kiely, P., Forristal, P.D., Fuller, H.T. & Frisvad, J.C. 2006. Mycotoxins and other secondary metabolites produced *in vitro* by *Penicillium paneum* Frisvad and *Penicillium roqueforti* Thom isolated from baled grass silage in Ireland. *Journal of Agricultural and Food Chemistry* 54, 9268-9276.
- Ocholi, R.A., Chima, J.C., Chukwu, C.O. & Irokanulo, E. 1992. Mycotoxicosis associated with *Penicillium purpurogenum* in horses in Nigeria. *Veterinary Record* 130 (22), 495.
- O'Kiely, P., Forristal, P.D. & Lenehan, J.J. 2000 a. Baled silage conservation characteristics as influenced by forage dry matter concentration, bale density and the number of wraps of plastic film used. *Irish Journal of Food and Agriculture Research* 39 (3), 468.
- O'Kiely, P., Forristal, P.D. & Lenehan, J.J. 2000 b. *Forage dry matter concentration, bale density and the quantity of plastic wrap used as factors affecting baled silage conservation*. In: Pullar, D. (Ed.) Beef from Grass and Forage. Occasional Symposium No. 35 British Grassland Society, 2000. Stafford, UK.
- Östling, C. & Lindgren, S. 1993. Inhibition of enterobacteria and *Listeria* growth by lactic, acetic and formic acids. *Journal of Applied Bacteriology* 75, 18-24.
- Östling, C. & Lindgren, S. 1995. Influences of enterobacteria on the fermentation and aerobic stability of grass silages. *Grass and Forage Science* 50, 41-47.
- Pagan, J.D., Kane, E. & Nash, D. 2005. *Form and source of tocopherol affects vitamin E status in Thoroughbred horses*. Proceedings Equine Nutrition Conference, Hannover, 1-2 October 2005, Germany. *Pferdeheilkunde* 21, 101-102.
- Pahlow, G. 1991. *Role of microflora in forage conservation*. In: Forage Conservation towards 2000. Landbauforschung Völkenrode, Braunschweig –Völkenrode (FAL). (Eds. Pahlow, G. & Honig, H). Sonderheft 123. pp. 26-36.
- Pahlow, G. & Dinter, B. 1987. *Epiphytic lactic acid bacteria of forages – methods of evaluation and first results*. Theatre Paper 1. In: Eight Silage Conference. The AFRC Institute for Grassland and Animal Production, Hurley, Berks, UK. pp. 1-2.
- Pahlow, G., Muck, R.E., Driehuis, F., Oude Elferink, S.J.W.H. & Spoelstra, S.F. 2003. Microbiology of ensiling. In: D.R. Buxton, R.E. Muck & J.H. Harrison (Eds.). *Silage Science and Technology*. ASA, CSSA, SSSA Agronomy no. 42, Madison, Wisconsin, USA. pp. 43, 50-51, 54.
- Pahlow, G. & Weissbach, F. 1996. *Effect of the numbers of epiphytic lactic acid bacteria (LAB) and of inoculation on the rate of pH-decline in direct cut and wilted grass silages*. In: Proceedings of the XIth International Silage Conference, Aberystwyth, UK. pp. 104-105
- Paillat, J-M. & Gaillard, F. 2001. Air-tightness of wrapped bales and resistance of polythene stretch film under tropical and temperate conditions. *Journal of Agricultural Engineering Research* 79 (1), 15-22.
- Pauly, T.M. 1999. Heterogeneity and hygienic quality of grass silage. Dissertation. Agraria 157. *Acta Universitatis Agriculturae Sueciae*, Uppsala, Sweden.
- Peltier, M.M., Peltier, M.R., Sharp, D.C. & Ott, E.A. 1997. Effect of  $\beta$ -carotene administration on reproductive function of horse and pony mares. *Theriogenology* 48, 893-906.
- Penell, J.C., Egenvall, A., Bonnett, B.N., Olson, P. & Pringle, J. 2005. Specific causes of morbidity among Swedish horses insured for veterinary care between 1997-2000. *Veterinary Record* 157 (15), 670-477.

- Persson, P. 2005. *Kartläggning och analys av hästverksamheten i Sverige (In Swedish)*. Rapport, samordningsenheten. Jordbruksverket, Jönköping. ISSN 1102-3007.
- Pitt, R.E., Muck, R.E. & Pickering, N.B. 1991. A model of aerobic fungal growth in silage. 2. Aerobic stability. *Grass and Forage Science* 46, 301-312.
- Pulse, R.E., Baker, J.P. & Potter, G.D. 1973. Effects of caecal fistulation upon nutrient digestion and indicator retention in horses. *Journal of Animal Science* 37, 488-492.
- Randall, R.P., Schurg, W.A. & Church, D.C. 1978. Response of horses to sweet, salty, sour and bitter solutions. *Journal of Animal Science* 47, 51-55.
- Randby, Å.T. & Fyhri, T. 2005. *Transport of wrapped silage bales*. Silage production and utilisation. In: Proceedings of the XIVth International Silage Conference, Belfast, Northern Ireland. pp. 246.
- Raymond, S.L., Curtis, E.F., Winfield, L.M. & Clarke, A.F. 1997. A comparison of respirable particles associated with various forage products for horses. *Equine Practice* 19 (2), 23-26.
- Rees, D.V.H., Audsley, E. & Neale, M.A. 1983. Some physical properties that affect the rate of diffusion of oxygen into silage. *Journal of Agricultural Science Cambridge* 100, 601-605.
- Reeves, M.J., Salman, M.D. & Smith, G., 1996. Risk factors for equine acute abdominal disease (colic): Results from a multi-center case-control study. *Preventive Veterinary Medicine* 26, 285-301.
- Respondek, F., Goachet, A-G., Rudeaux, F. & Julliand, V. 2005. *Effects of short-chain fructo-oligosaccharides on the microbial and biochemical profile of different segments of the gastrointestinal tract in horses*. Proceedings Equine Nutrition Conference, Hannover, Germany. *Pferdeheilkunde* 21, 69-70.
- Ricketts, S.W., Greet, T.R.C., Glyn, P.J., Ginnett, C.D.R., McAllister, E.P., McCaig, J., Skinner, P.H., Webbon, P.M., Frappe, D.L., Smith, G.R. & Murray, L.G. 1984. Thirteen cases of botulism in horses fed big bale silage. *Equine Veterinary Journal* 16 (6), 515-518.
- Roberts, M.C. 1975. Carbohydrate digestion and absorption studies in the horse. *Research in Veterinary Science* 18, 64-69.
- Roberts, T.A. 1988. Botulism. In: Stark, B.A. & Wilkinson, J.M. (Eds.). *Silage and health*. Chalcombe Publications, UK. pp. 35-43.
- Robinson, N.E., Derksen, F.J., Olszewski, M.A. & Buechner-Maxwell, V.A. 1996. The pathogenesis of chronic obstructive pulmonary disease of horses. *British Veterinary Journal* 152, 283-306.
- Ronéus, B. 1982. Glutathione peroxidase and selenium in the blood of healthy horses and foals affected by muscular dystrophy. *Scandinavian Journal of Veterinary Science* 34, 350-353.
- Ronéus, B., Ronéus, N. Franklin, A. & Jonsson, P. 1993. Behandling med standardiserad kolikultur vid tarmflorerubbning hos hästar (Treatment of horses with disturbed intestinal microflora using standardised coli culture) (In Swedish). *Svensk Veterinärtidning* 45, 201-204.
- Rowe, J.B., Lees, M.J. & Pethick, D.W. 1994. Prevention of acidosis and laminitis associated with grain feeding in horses. *Journal of Nutrition* 124, 2742S-2744S.
- Ruxton, G.D. & Gibson, G.J. 1995. A mathematical model of the aerobic deterioration of big bale silage and its implications for the growth of *Listeria monocytogenes*. *Grass and Forage Science* 50, 331-344.
- Saastamoinen, M.T. & Juusela, J. 1992. Influence of dietary supplementation on serum vitamin A and D concentrations and their seasonal variation in horses. *Agricultural Science in Finland* 1 (5), 477-482.
- Saastamoinen, M. T. & Juusela, J. 1993. Serum vitamin E concentration of horses on different vitamin E supplementation levels. *Acta Agriculturae Scandinavica Section A Animal Science* 43, 52-57.
- Salter, R.E. & Hudson, R.J. 1979. Feeding ecology of feral horses in Western Alberta. *Journal of Range Management* 32 (3), 221-225.

- Savoie, P. & Jofriet, J.C. 2003. Silage storage. In: D.R. Buxton, R.E. Muck & J.H. Harrison (Eds.). *Silage Science and Technology*. ASA, CSSA, SSSA Agronomy no. 42, Madison, Wisconsin, USA. pp. 440-445.
- Schoenbaum, M.A., Hall, M.S., Glock, R.D., Grant, K., Allen, J.L., Schiefer, T.J., Scigliabaglio, P. & Whitlock, R.H. 2000. An outbreak of type C botulism in 12 horses and a mule. *Journal of the American Veterinary Medical Association* 217 (3), 365-368.
- Schwarz, F.J., Sliwinski, H., Schuster, M. & Rosenberger, E., 2005. *Variation in the nutrient composition of different feedstuffs for horses*. Proceedings Equine Nutrition Conference 1-2 October 2005, Hannover, Germany. *Pferdeheilkunde* 21, 9-10.
- Scudamore, K.A. & Livesey, C.T. 1998. Occurrence and significance of mycotoxins in forage crops and silage: a review. *Journal of the Science of Food and Agriculture* 77, 1-17.
- Seshan, P.A. & Sen, K.C., 1942. Studies on carotene in relation to animal nutrition. Part III. Stability of carotene in plant material with special reference to hay making and storage. *Journal of Agricultural Science* 32, 275-285.
- Shahane, J. & Mungikar, A.M. 1991. Stability of  $\beta$ -carotene during silage preparation from pressed lucerne. *Comparative Physiology and Ecology* 16, 27-31.
- Slottner, D. & Bertilsson, J. 2006. Effect of ensiling technology on protein degradation during ensilage. *Animal Feed Science and Technology* 127, 101-111.
- Smith, D. 1973. The non-structural carbohydrates. Chapter 3. In: Butler, G.W. & Bailey, R.W. *Chemistry and biochemistry of herbage*. Vol 1. Academic Press Inc., London, Ltd. pp. 109-110.
- Smolders, E.A.A., Steg, A. & Hindle, V.A. 1990. Organic matter digestibility in horses and its prediction. *Netherlands Journal of Agricultural Science* 38, 435-447.
- Sneddon, J. C. 1993. Physiological effects of hypertonic dehydration on body fluid pools in arid-adapted mammals. How do Arab-based horses compare? *Comparative Biochemistry and Physiology* 104 A, (2), 201-213.
- Sneddon, J.C. & Argenzio, R.A. 1998. Feeding strategy and water homeostasis in equids: the role of the hindgut. *Journal of Arid Environments* 38, 493-509.
- Spoelstra, S.F. 1981. *Spores of lactate-fermenting clostridia in grass silage*. Paper No. 5. In: Sixth silage conference – silage production and utilization. (Eds. R.D. Harkess & M.E. Castle). Queen Margaret College, Edinburgh, UK. pp. 9-10
- Spoelstra, S.F. 1987. Degradation of nitrate by enterobacteria during silage fermentation of grass. *Netherlands Journal of Agricultural Science* 35, 43-54.
- Spoelstra, S.F. 1990. Comparison of the content of clostridial spores in wilted grass silage ensiled either in laboratory, pilot-scale or farm silos. *Netherlands Journal of Agricultural Science* 38, 423-434.
- Ström, K. 2005. *Fungal inhibitory lactic acid bacteria*. Characterization and application of *Lactobacillus plantarum* MiLAB 393. Doctoral thesis. Department of Microbiology, Swedish University of Agricultural Sciences, Uppsala, Sweden.
- Sullivan, J.T. 1973. Drying and storing herbage as hay. In: G.W. Butler & R.W. Bailey (Eds.). *Chemistry and Biochemistry of Herbage*. Vol.3. Academic Press Inc. Ltd. London, UK. Chapter 27. pp. 1-31.
- Sundberg, M. & Lindahl, C. 2006. Uppfuktning och mögelbildning vid lagring av hö till hästar – försök 2005-2006. Preliminary report (*In Swedish*), November 2006. *Swedish Institute of Agricultural and Environmental Engineering*, Uppsala, Sweden.
- Suzuki, M. 1993. History of fructan research: Rose to Edelman. In: M. Suzuki & N.J. Chatterton (Eds.) *Science and Technology of fructans*. CRC Press Inc., Boca raton, Florida, USA. pp. 21-39.
- Switzer, J.W., Jensen, M., Riemann, H.P and Airola, W.A. 1984. An outbreak of suspected type D botulism in horses in California. *California Veterinarian* 7, 14-17.
- Thomas, T.D., Ellwood, D.C. & Longyear, V.M.C. 1979. Change from homo- to heterolactic fermentation by *Streptococcus lactis* resulting from glucose limitation in anaerobic chemostat cultures. *Journal of Bacteriology* 138 (1), 109-117.
- Thompson, F.M.L. 1983. Horses and hay in Britain 1830-1918. In: F.M.L. Thompson (Ed.). *Horses in European economic history. A preliminary canter*. The British Agricultural History Society, Reading, UK. pp. 50-72.

- Udén, P. 1982. The determination of digesta particle size in some herbivores. *Animal Feed Science and Technology* 7, 35-44.
- Udén, P., Rounsaville, T.R., Wiggans, G.R. & Van Soest, P.J. 1982. The measurement of liquid and solid digesta retention in ruminants, equines and rabbits given timothy (*Phleum pratense*) hay. *British Journal of Nutrition* 48, 329-339.
- Udén, P. & van Soest, P.J. 1982. Comparative digestion of timothy (*Phleum pratense*) fibre by ruminants, equines and rabbits. *British Journal of Nutrition* 47, 267-272.
- Undersander, D., Wood, T. & Foster, W. 2005. *Wrapping rectangular bales with plastic to preserve wet hay or make haylage*. In: Silage production and utilisation. Proceedings of the XIVth International Silage Conference, Belfast, Northern Ireland. pp. 247.
- Van Eps, A.W. & Pollitt, C.C. 2006. Equine laminitis induced with oligofructose. *Equine Veterinary Journal* 38 (3), 203-208.
- Van Weyenberg, S., Sales, J. & Janssens, G.P.J. 2006. Passage rate of digesta through the equine gastrointestinal tract: A review. *Livestock Science* 99, 3-12.
- Vandenput, S., Duvivier, D.H., Votion, D., Art, T. & Lekeux, P. 1998. Environmental control to maintain stabled COPD horses in clinical remission: effects on pulmonary function. *Equine Veterinary Journal* 30 (2), 93-96.
- Vandenput, S., Istasse, L., Nicks, B. & Lekeux, P. 1997. Airborne dust and aeroallergen concentrations in different sources of feed and bedding for horses. *Veterinary Quarterly* 19, 154-158.
- Varlout, M., de Fombelle, A., Goachet, A.G., Drogoul, C. & Julliand, V. 2004. Partial and total apparent digestibility of dietary carbohydrates in horses as affected by the diet. *Animal Science* 79, 61-72.
- da Veiga, L., Chaucheyras-Durand, F. & Julliand, V. 2005. *Comparative study of colon and faeces microbial communities and activities in horses fed a high starch diet*. In: Proceedings Equine Nutrition Conference, Hannover, 1-2 October 2005, Germany. *Pferdeheilkunde* 21, 45-46.
- Vesonder, R., Haliburton, J., Stubblefield, R., Gilmore, W. & Peterson, S. 1991. *Aspergillus flavus* and aflatoxins B<sub>1</sub>, B<sub>2</sub> and M<sub>1</sub> in corn associated with equine death. *Archives of Environmental and Contamination Toxicology* 20, 151-153.
- Waldo, D.R., Keys, J.E. Jr. & Gordon, C.H. 1972. Preservation efficiency and dairy heifer response from unwilted formic and wilted untreated silages. *Journal of Dairy Science* 56, 129-136.
- Wallin, L., Strandberg, E., Philipsson, J. & Dalin, G. 2000. Estimates of longevity and causes of culling and death in Swedish warmblood and coldblood horses. *Livestock Production Science* 63, 275-289.
- Waring, G.H. 1974. Behavioral adaptation of feeding in horses. *Journal of Animal Science* 39, 137.
- Watson, S.J. & Nash, M.J. 1960. *Conservation of grass and forage crops*. Oliver and Boyd, Edinburgh, Scotland, UK. In: Butler, G.W. & Bailey, R.W. (Eds.). *Chemistry and biochemistry of herbage*. Vol. 3. Academic Press Inc. Ltd., London, UK.
- Weissbach, F. 1968. Beziehungen zwischen Ausgangsmaterial und Gärungsverlauf bei der Grünfuttersilierung (Relationships between herbage and the course of fermentation in ensiling green fodder) (*In German*). Habilitation, *University of Rostock*, Germany.
- Weissbach, F., Schmidt, L. & Hein, E. 1974. *Method of anticipation of the run of fermentation in silage making based on the chemical composition of the green fodder*. p. 663-673. In: Proceedings of the 12<sup>th</sup> International Grassland Congress, (Eds. V.G. Iglovikov & A.P. Movsisyants). Vol. 3, Part 2. Moscow. 11-20 June 1974. Russian Academy of Agricultural Sciences, Lgovaya.
- Wieringa, G.W. 1958. The effect of wilting on butyric acid fermentation in silage. *Netherlands Journal of Agricultural Sciences* 6, 204-210.
- Wierup, M. 1977. *Equine Intestinal Clostridiosis*. Acta Veterinaria Scandinavica Supplementum 62. PhD-thesis. Royal Veterinary College, Uppsala, Sweden.
- Wilkinson, J.M. 1999. Silage and animal health. *Natural Toxins* 7, 221-232.
- Wilkinson, J.M. & Toivonen, M.I. 2003. *World Silage – A survey of forage conservation around the world*. Appendix 38. Chalcombe Publications, Lincoln, UK.

- Williams, A.G. 1994. The permeability and porosity of grass silage as affected by dry matter. *Journal of Agricultural Engineering Research* 59, 133-140.
- Wilson, T.M., Morrison, H.A., Palmer, N.C., Finley, G.G., & van Dreumel, A.A. 1976. Myodegeneration and suspected selenium/vitamin E deficiency in horses. *Journal of the American Veterinary Medical Association* 169, 213-217.
- Woolford, M.K. 1975. Microbiological screening of food preservatives, cold sterilants and specific antimicrobial agents as potential silage additives. *Journal of the Science of Food and Agriculture* 26, 229-237.
- Wylam, C. 1953. Analytical studies on the carbohydrates of grasses and clovers. III. – Carbohydrate breakdown during wilting and ensilage. *Journal of the Science of Food and Agriculture* 4, 527-531.
- Yuki, N., Shimazaki, T., Kushiro, A., Watanabe, K., Uchida, K., Yuyama, T. & Morotomi, M. 2000. Colonization of the stratified squamous epithelium of the nonsecreting area of horse stomach by Lactobacilli. *Applied and Environmental Microbiology* 66 (11), 5030-5034.

## Acknowledgements

The experiments in this thesis were mainly performed at Kungsängen Research Centre, Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, Uppsala. The horses in Paper V were located at ENESAD (Établissement National d'Enseignement Supérieur Agronomique de Dijon), Dijon, France.

Financial support was provided by The Swedish Farmers Foundation for Agricultural Research–Horse Research Committee; Trioplast Inc., Smålandsstenar, Sweden; Agria Sweden and Krafft Hästfoder, Sweden. Additives were supplied by Medipharm, Kågeröd, Sweden; Hanson & Möhring, Sweden and Addcon Agrar GmbH, Germany. Machinery equipment was supplied by Tellefsdal/Möre maskiner, Sweden; Lely Welger Maschinenfabrik, Germany; New Holland/Söderberg & Haak, Sweden; Kverneland/Taarup, Sweden and Trejon, Sweden. All are greatly acknowledged for their contributions.

The Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, Sweden is greatly acknowledged for providing research facilities, computers, travel grants etc.

There are a number of people who have, in different ways, directly or indirectly or both, helped out during the work with this thesis, and to whom I would like to express my sincere gratitude:

First of all, my very special THANKS to my main supervisor, Docent Peter Udén. I think it would take up too much space to mention all the things that I have learned from you. I am very grateful that you have given me free reins throughout the work with this thesis, but also for always having time for my questions and manuscripts, and for pointing me in the right direction when it was needed. You have a rare ability to make me teach myself, which I appreciate very much. I think that I have been very, very fortunate to have you as my supervisor.

Mrs. Margie Knipe, thank you for excellent and rapid linguistic revision and for your interest in my work. All faults still residing in the text are entirely my own... Also, Thank You both Peter and Margie for your warm hospitality and for all the invitations and dinners at your home. I have enjoyed them very much!

Agr.D. Thomas Pauly, my co-supervisor, thank you for taking of your time to help me with all sorts of things, such as reading manuscripts, answering questions about silage microbiology, checking my German translations and lifting small bales here and there. I know that you have a very tight schedule, and therefore extra thanks for also taking the time to put in a joke here and now!

DVM Merike Ronéus, my second co-supervisor. Thank you for the discussions about horses and forages and everything related and unrelated! I thought it was

very nice to have your support, especially during the beginning of the work with my thesis.

The laboratory staff at Kungsängen; Börje Ericsson, Håkan Wallin, Barbro Näslund, Camilla Andersson, thank you for excellent analytical work and advice on sample handling and preparation. A special thanks to Börje, I don't know how you manage to keep a good mood all the time?! Thank you also for letting me invade the microbiology room (among other spaces) from time after another.

Professor Dietrich von Rosen, Department of Biometry and Engineering, SLU, thanks for statistical consultation and for your patience and understanding of our experimental designs. I know better now...hopefully.

Thanks to all of my colleagues at the "Forage Conservation Group": Agr.D. Rolf Spörndly (special thanks for reading my thesis), Agr.D. Martin Knický and Rainer Nylund, for all help with the small bale experiments. A special thanks to Johan Andersson, for fixing every little practical problem during the experiments, almost before they even occurred. Thanks also to Kerstin Burstedt for being my translator to the impossible administration system of SLU.

Thanks to all the master-students that have helped out with the experiments in different ways, especially MSc. Jenny Möller (Paper **III**), and MSc. Jenny Redgård (Paper **IV**), thank you! Also, thanks to former colleague Agr.D. David Slotner, who carried around a lot of small bales during the experiments and thought it was "good exercise"... Also, special thanks to PhD-student Sara Muhonen, my collaborator in the experiments at ENESAD in France, and her supervisor Agr.D. Anna Jansson.

Thanks to all former and present colleagues at Kungsängen Research Centre, for all the discussions and laughters during coffee breaks and lunches. I am sure that the discussions going on around the tables in the kitchen are neither more diverse nor more detailed at any other workplace! Agr.D. Torsten Eriksson, thank you for showing interest in my results and for helpful discussions about them, and Agr.D. Sigrid Agenäs, thanks for showing interest in my work and for reading the thesis in advance to printing.

Thanks to my fellow PhD-students at Kungsängen; Linda Forsbäck, Maria Eriksson, Lotta Jönsson, Sofie Fröberg and Anna Werner; especially thanks to Linda and Maria for improving my lunch habits by at least 1000 %. I am afraid that I will fail to keep up the improvement by myself ☺.

Tack också till Staffan Söderberg, Hölö; Urban och Ronae Brunsberg, Enköping, Henrik Lillje, Arboga och Johan Bertilsson, Köping, för ert intresse för mitt forskningsarbete, och för er vilja att dela med er av era egna erfarenheter av produktion av vallfoder till hästar i större kvantiteter.

Agronom Claes Nilsson, min mentor i PUMA-kursen. Tack för att du visat mig hur många olika saker en agronom kan duga till!

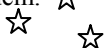


Tack till Lektor Pia Haxner, som ställde upp med hästinhysning och -passning när det var körigt, och för att du är du ☺.

Mina grannar Emma & Johan Thorén-Hellsten och Anna Jansson & Per Gustafsson för att ni ställt upp med hästpassning när det har behövts. Tack till Anna också för alla pratstunder på hemmaplan om jobbet och hästforskning.

Suzanne Cewé, vilken tur jag hade som träffade på just dig och Rauen för sisådär 6-7 år sedan! Tack för att du inte bara är en superb ridlärare, utan också en god vän som dessutom förstår och har intresse av mitt forskningsarbete. Om alla tränare var lika intresserade av hästutfodring som du, skulle nog många hästar må lite bättre ;-)

Min bror Martin, min svägerska Anne, brorsbarnen Lina och Sanna Lyrup-Müller, tack för att ni ser till att jag inte försvinner in i mina böcker och papper, och för att ni alltid finns där. Likaså Emma Jönsson med familj och Sara N. Rasmussen med familj, mina vänner sedan barnsben, för att ni alltid finns där även när tiden egentligen inte räcker till, och även om ni ibland befinner er på andra sidan jordklotet (eller i alla fall 65 mil bort...). ”Riktiga vänner är som stjärnor, de finns alltid där även om man inte alltid ser dem.” ☆ ☆



Meine Oma, Du bist mein Idol.

Dennah<sup>ox</sup>, och alla andra kära hästvänner – tack för att ni är min fristad där det bara går att vara ”här och nu”.

Min mamma Kerstin och min pappa Heribert – jag har så oerhört mycket att tacka er för, men mest av allt vill jag säga tack för er ousinliga uppmuntran för alla mina påhitt, och för all hjälp jag alltid fått när något har varit problematiskt eller svårt. Ni har alltid gett mig alla möjligheter, och jag har lärt mig mycket mer av er än vad ni nog egentligen själva tror.