

**Genetic analysis of host and phosphite
mediated resistance to *Phytophthora cinnamomi*
in *Arabidopsis thaliana***

by

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Declaration

The work described in this thesis was undertaken while I was an enrolled student for the degree of Doctor of Philosophy at Murdoch University, Western Australia. I declare that this thesis is my own account of my research and contains as its main content work which has not previously been submitted for a degree at any tertiary education institution.

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Abstract

Phosphite (Phi), an analogue of phosphate (Pi) is highly effective for the control of *Phytophthora cinnamomi*, a devastating necrotrophic pathogen worldwide. This study describes the effect of phosphite (Phi) on the induction of defence responses in *Phytophthora cinnamomi*-infected *Arabidopsis thaliana* accessions Ler and Col-0, and mutants defective in salicylic acid (SA), jasmonic acid (JA), ethylene (ET), abscisic acid (ABA), phosphate starvation response (PSR) and auxin response signalling pathways. The inoculation of the resistant Col-0 with *P. cinnamomi* induced a rapid increase in callose deposition (by 6 h after inoculation) and hydrogen peroxide (H₂O₂) production (by 24 h after inoculation) whereas inoculation of susceptible Ler showed a delayed and reduced response. Treatment of Ler with Phi produced a response to *P. cinnamomi* inoculation similar to that observed in Col-0 in terms of timing and magnitude suggesting Phi primes the plant for a rapid and intense response to infection involving heightened activation of a range of defence responses.

A reliable method for measuring disease progression is important when evaluating susceptibility in host–pathogen interactions. A sensitive quantitative polymerase chain reaction (QPCR) assay was developed for the quantitative measurement of *P. cinnamomi* DNA (biomass) *in planta* that avoids problems caused by variation in DNA extraction efficiency and degradation of host DNA during host tissue necrosis. Purified plasmid DNA, containing the pScFvB1 mouse gene, was added during DNA extraction and the pathogen's biomass was normalized based on plasmid DNA rather than host DNA or sample fresh weight. It was demonstrated that normalization of pathogen DNA to sample fresh weight or host DNA in samples with varying degrees of necrosis led to an overestimation of the pathogen's biomass.

Inoculation of mutants in the SA, JA, and ET defence signalling pathways did not affect the resistance of Col-0 suggesting alternative pathways are involved. A high level

susceptibility was observed in the *aba2-4* mutant suggesting a role for ABA signalling in the induction of resistance to *P. cinnamomi*. Phi treatment of *aba2-4* increased resistance but not to the wild type levels indicating a possible role for ABA-dependent and ABA independent signalling in Phi mediated resistance. Application of Phi to non-inoculated *A. thaliana* seedlings elevated transcription of defence genes in the SA (*PR1* and *PR5*) and JA/ET (*THI2.1* and *PDF1.2*) pathways. Furthermore, analysis of gene expression in Col-0 revealed that either Phi or *P. cinnamomi* caused the down-regulation of the transcriptional level of *AtMYC2* (a positive regulator of ABA signalling which also negatively regulates JA-related genes) and increased the transcriptional abundance of *PDF1.2*. Together these results suggest that the resistance response of Col-0 and Phi treatment both act partially through an ABA dependent mechanism which is independent of the antagonism between ABA and elements of the JA/ET pathway such as *PDF1.2*.

Phosphite has been suggested to interfere with various plant processes including Pi homeostasis therefore the potential involvement of the Pi and auxin signalling pathways in resistance to *P. cinnamomi* was investigated using several PSR and auxin response pathway mutants. The mutants *tir1-1*, an auxin response mutant deficient in the auxin-stimulated SCF (Skp1–Cullin–F-Box) ubiquitination pathway and *phr1-1*, a mutant defective in response to Pi starvation were highly susceptible to *P. cinnamomi* compared to their parental background Col-0. Complementation restored resistance to the level observed in Col-0. Moreover, inhibition of auxin transporters by TIBA (2,3,5-triiodobenzoic acid) led to a significant increase in susceptibility of *Lupinus angustifolius* seedlings to *P. cinnamomi* supporting the importance of the auxin signalling pathway in *P. cinnamomi* resistance. The 26S proteasome subunits mutants; *rpn10-1* (Defective in ubiquitin/26S proteasome-mediated proteolysis) and *pbe1-1* (proteasome subunit beta type-5-A) were also susceptible to *P. cinnamomi*. The *rpn10-1*

mutant has also been associated with the auxin signalling pathway and the susceptibility of *rpn10-1* and *pbe1-1* indicates that the 26S proteasome and auxin signalling could play a role in resistance to *P. cinnamomi*. Given the apparent involvement of auxin and PSR signalling in the resistance to *P. cinnamomi*, the possible involvement of these pathways in Phi mediated resistance was also investigated. Application of Phi at both low and high concentrations attenuated some of the Pi starvation inducible genes such as *At4*, *AtACP5* and *AtPT2*. However, in phosphate sufficient plants, Phi treatment mimicked Pi starvation responses in terms of enhanced expression of *PHR1*, *AUX1*, *AXR1*, *AXR2* and *SGT1B*; suppression of primary root elongation, and increased root hair formation. Together, these results suggest that the auxin response pathway, particularly auxin sensitivity and transport, plays a role in the plant's resistance to *P. cinnamomi* and suggest that phosphite-mediated resistance may in some part be through its effect on stimulation of the auxin response pathway.

Statement of the contributions of jointly authored papers

The following manuscripts have either been published or have been prepared/submitted to scientific journals.

Chapter 2: Eshraghi, L., Anderson, J., Aryamanesh, N., Shearer, B., McComb, J., Hardy, G. E. S. and O'Brien, P. A. (2011) Phosphite primed defence responses and enhanced expression of defence genes in *Arabidopsis thaliana* infected with *Phytophthora cinnamomi*. *Plant Pathology*, 60, 1086-1095.

The contribution of work for this paper is 90% by the candidate of this thesis, Leila Eshraghi including the design, performance and analysis of experiments and writing of the manuscript, and 10% for all other authors in terms of advice on the experimental design, approach and revising the manuscript.

Chapter 3: Eshraghi, L., Aryamanesh, N., Anderson, J. P., McComb, J., Hardy, G. E. S., Shearer, B. and O'Brien, P. A. (2011) A quantitative PCR assay for accurate in planta quantification of the necrotrophic pathogen *Phytophthora cinnamomi*. *European Journal of Plant Pathology*, 131, 419-430.

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Chapter 4: Eshraghi, L., Anderson, J., Aryamanesh, N., Shearer, B., McComb, J. and Hardy, G. E. S. (2012) Defence signalling pathways involved in plant resistance and phosphite-mediated control of *Phytophthora cinnamomi*. Submitted to *Planta* (March 2012).

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Chapter 5: Eshraghi, L., Aryamanesh, N., Anderson, J. P., McComb, J., Shearer, B. and Hardy, G. E. S. (2012) Suppression of auxin response pathway enhances susceptibility to *Phytophthora cinnamomi* and phosphite stimulates *Arabidopsis* auxin signalling pathway. Prepared for *BMC Biology*.

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Coordinating Supervisor, Professor Giles Hardy

Signature:

Conference publications pertaining to this thesis

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Eshraghi L, Aryamanesh N, Anderson A J, McComb J, Hardy G E S, Shearer B & O'Brien P A (2011). Evaluating the role of defence pathways of *Arabidopsis thaliana* in resistance to *Phytophthora cinnamomi*. In 'Asian Association of Societies for Plant Pathology (AASPP) and the Australasian Plant Pathology Society (APPS). 26- 29 April 2011 Darwin, Australia.

Eshraghi L, McComb J, Hardy G E S & O'Brien P A (2008). The role of Phosphite in inducing resistance to *Phytophthora cinnamomi* in *Arabidopsis thaliana*. In '9th International Congress of Plant Pathology', 24–29 August 2008; Turin, Italy.

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and discipline.

List of abbreviations

| | |
|-------------|--|
| Δ ct | Change in threshold cycle |
| ABA | Abscisic acid |
| ANOVA | Analysis of variance |
| AGI | <i>Arabidopsis</i> Genome Initiative |
| ARG | Auxin responsive genes |
| ARF | Auxin response factor |
| ASK | <i>Arabidopsis</i> SKP1-like |
| AUX/IAA | Auxin/indole-3-acetic-acid |
| AXR | Auxin resistant |
| COI | Coronitine insensitive |
| COP | Constitutive photomorphogenic |
| CTR | Constitutive triple response |
| Ct | Threshold cycle |
| CUL | Cullin |
| CV | Coefficient of variation |
| E1 | Enzyme 1 (same as UBA, ubiquitin activating enzyme) |
| E2 | Enzyme 2 (same as UBC, ubiquitin conjugating enzyme) |
| E3 | Enzyme 3 (same as ubiquitin protein ligase) |
| EIN | Ethylene insensitive |
| ET | Ethylene |
| ETI | Effector-triggered immunity |
| ETR | Ethylene receptor |
| ERF | Ethylene response factor |
| FBX2 | F-box protein 2 |
| GFP | Green fluorescent protein |

| | |
|-------|---|
| GUS | Beta-glucuronidase |
| GMO | Genetically Modified Organisms |
| HR | Hypersensitive response |
| IAA | Indole-3-acetic acid |
| ISI | Induces systemic resistance |
| JA | Jasmonic acid |
| LRR | Leucine-rich repeat |
| MAPK | Mitogen-activated protein kinase |
| MES | 2-morpholinoethanesulfonic acid |
| Myc | Epitope tag from c-Myc protein |
| NIM | Non-inducible immunity |
| NO | Nitric oxide |
| NPR | Non-expressor of pathogenesis-related genes |
| PAMP | Pathogen associated molecular pattern |
| Pc | <i>Phytophthora cinnamomi</i> |
| PCR | Polymerase chain reaction |
| PCD | Programmed cell death |
| PDF | Plant defensin |
| Pi | Phosphate |
| Phi | Phosphite |
| PHR1 | Phosphate starvation response 1 |
| PIN | Pin-formed |
| PPCK1 | Phosphoenolpyruvate carboxylase kinase 1 |
| PR | Pathogenesis-related |
| PSR | Phosphate starvation response |
| PTI | PAMP-triggered immunity |

| | |
|----------------|--|
| QPCR | Quantitative polymerase change reaction |
| R gene/protein | Resistance gene/protein |
| RAR | Required for MIA12 resistance |
| RBX | RING-box protein, same as ROC1 and Hrt1p |
| RING | Really interesting new gene protein domain |
| RNA | Ribonucleic acid |
| ROS | Reaction oxygen species |
| ROC | Regulator of cullins |
| RT | Reverse transcription |
| RNAase | Ribonuclease |
| SA | Salicylic acid |
| SAR | Systemic acquired resistance |
| SE | Standard error |
| SCF | Skp1-Cullin1-F-box |
| SGT | Suppressor of G2 allele of skp1 |
| SKP | S phase kinase-associated protein |
| SON | Suppressor of nim1-1 |
| T-DNA | Transfer DNA |
| THI | thionin |
| TIBA | 2,3,5-triiodobenzoic acid |
| TIR1 | Transport inhibitor response1 |
| UBA | Ubiquitin activating enzyme |
| UBC | Ubiquitin conjugating enzyme |
| U-box | UFD2-homology domain |
| UPP | Ubiquitin proteasome pathway |