

Interaction of misfolded proteins and mitochondria in neurodegenerative disorders

Andrey Y. Abramov^a, Alexey V. Berezhnov^b, Evgeniya I. Fedotova^b, Valery P. Zinchenko^b,
Ludmila P. Dolgacheva^b

^aDepartment of Molecular Neuroscience, UCL Institute of Neurology, London WC1N 3BG,
United Kingdom

^bInstitute of Cell Biophysics Russian Academy of Sciences, Pushchino 142290, Russia

Corresponding author: Andrey Y Abramov, Department of Molecular Neuroscience,
UCL Institute of Neurology, London WC1N 3BG, United Kingdom e-mail:
a.abramov@ucl.ac.uk

Keywords: neurodegeneration, mitochondria, alpha-synuclein, tau, huntingtin, beta-amyloid

Abstract

The number of the people affected by neurodegenerative disorders is growing dramatically due to the aging of population. The major neurodegenerative diseases share some common pathological features including involvement of mitochondria in the mechanism of pathology and misfolding and accumulation of abnormally aggregated proteins. Neurotoxicity of aggregated beta-amyloid, tau, alpha-synuclein and huntingtin is linked to effects of these proteins on mitochondria. All these misfolded aggregates affect mitochondrial energy metabolism by inhibiting diverse mitochondrial complexes and limits ATP availability in neurons. Beta-amyloid, tau, alpha-synuclein and huntingtin are shown to be involved in increased production of reactive oxygen species which can be generated in mitochondria or can target this organelle. Most of these aggregated proteins are able to deregulate mitochondrial calcium handling that in combination with oxidative stress lead to opening of the mitochondrial permeability transition pore. Despite some of the common features, aggregated beta-amyloid, tau, alpha-synuclein and huntingtin have diverse targets in mitochondria that can partially explain neurotoxic effect of these proteins in different brain regions.

Introduction

Aging of the population in the majority of countries leads to increase in the number of people with age-related disorders, including neurodegenerative diseases. Despite the differences in aetiology and different mechanisms of cell loss, most of neurodegenerative disorders share some of common features including the involvement of mitochondrial dysfunction and oxidative stress in development of pathology and, specific for each disease misfolded proteins which aggregates in the brain of patients. Thus, for Alzheimer's disease there are two misfolded proteins – β -amyloid (main component of the senile plaques) and tau protein (aggregates of which form an intracellular tangles) (1). It should be noted that tau aggregates are not specific for Alzheimer's only and appeared in the number of other neurodegenerative disorders. Tau mutations are shown to be linked to familial form of frontotemporal dementia (2, 3). Second most common neurodegenerative disorder – Parkinson's disease is characterised by intracellular occlusions called Lewy bodies, which are formed by aggregated α -synuclein (4). One of the main histopathological features of Huntington disease is an aggregate of huntingtin protein (5). All these aggregates in the brain consist mostly of protein fibrils and for long time are believed to be

the trigger of cellular pathology and neurodegeneration in these diseases. Only recently a number of studies showed that small oligomeric forms of these proteins are more toxic than monomeric or fibril forms.

Mitochondrion is an organelle which is strongly implicated in the mechanisms of neurodegeneration. Being the major energy producer in the cell, mitochondria play important role in the mechanism of cell death, calcium and redox signalling (6). Ability of mitochondria to produce reactive oxygen species (ROS) in the electron transport chain, TCA cycle and some other enzymes has a functional implication in cell signalling, however overproduction of ROS in mitochondria links this organelle to the age-related pathology and neurodegenerative disorders (7, 8). However, some of the mitochondrial enzymes could be specifically linked to functions involved in maintenance of neuronal homeostasis. Thus, the enzyme monoamine oxidase, which is located on the outer membrane of mitochondria, is involved in the homeostasis of neurotransmitters – dopamine, serotonin and norepinephrine (9, 10). Neurons are also dependent on mitochondrial function due to several reasons: a) Neurons predominantly produce ATP via oxidative phosphorylation in the mitochondria, with almost no contribution from glycolysis; b) The brain consumes 10 times more oxygen and glucose than any other organ or tissue that may results in higher probability of ROS production. c) Neurons are long-lived differentiated cells that are therefore more dependent on the processes of mitochondrial dynamics and removal of unwanted mitochondria (mitophagy) compare to cells from other tissues. d) Neurons are excitable cells with high calcium fluxes. Mitochondria play a role of buffering Ca^{2+} that shapes calcium signals and protects cells from calcium excitotoxicity. e) Neuronal processes, that is, axons and dendrites, may be very long and therefore depend on mitochondrial transport for transfer of energy molecules to different parts of the cell. Because of the importance of all mitochondrial functions to neuronal health, it is perhaps understandable why neurons are vulnerable to dysfunction in any mitochondrial pathway. Mitochondrial pathology has been associated with a wide range of neurodegenerative diseases. In primary mitochondrial diseases, that is, diseases caused by mutations in mitochondrial DNA or nuclear DNA encoding mitochondrial proteins, it is clear that perturbation in mitochondrial function alone is sufficient and necessary to trigger neuronal death. It is less clear whether the mitochondrial dysfunction seen in the sporadic late onset neurodegenerative diseases is necessary for pathogenesis or a bystander effect of disease, which is mostly associated with misfolded aggregates.

For long time, the role of misfolded proteins in mitochondrial function was disputable due to lack of evidence of location of misfolded aggregates in mitochondria. Recent studies demonstrated that aggregated peptides could be delivered to mitochondria (11).

In this review we discuss the different mechanisms by which misfolded proteins affect mitochondrial function and ROS production and how mitochondrial dysfunction and protein aggregation could be related to progressive neuronal death in different forms of neurodegeneration.

Effects of β -Amyloid on mitochondria

The amyloid precursor protein (APP) is cleaved by β and γ -secretase generating a range of β -amyloid (β A) peptides between 39 and 43 amino acid residues long, where the hydrophobic nature of β A 1-40 and β A 1-42 promotes self-aggregation and neurotoxicity. A series of conformational changes of β A via dimers, oligomers, protofibrils and fibrils leads ultimately to a deposition of amyloid plaques.

β A shown to have a strong effect on mitochondrial enzymes which contains iron-sulphur centre – the most of these enzymes are the complexes of electron transport chain (Figure 1) and TCA cycle - α -ketoglutarate dehydrogenase and aconitase (12-15). In intact cortical and hippocampal neurons, β A reduce ATP levels through inhibition of the complexes I and IV (16) and induces profound mitochondrial depolarisation of two types – slow mitochondrial depolarisation and sharp and transient loss of $\Delta\psi_m$ (17, 18). This mitochondrial depolarisation was dependent on the β A-induced calcium signal (17, 19) and induction of ROS overproduction from NADPH oxidase (20, 21). Deregulation of calcium homeostasis has been demonstrated in Alzheimer's disease (AD), with β A causing increased cytoplasmic calcium levels and mitochondrial calcium overload, resulting in increase in ROS production and opening of the PTP (19, 20). β A is able to induce opening of PTP in isolated mitochondria (22, 23) and primary astrocytes (17, 24, 25). Furthermore β A may directly interact with cyclophilin D (a PTP component) forming a complex in the mitochondria that has reduced threshold for opening in the presence of mPTP inducers. Prevention of PTP opening by inducing cyclophilin D deficiency (molecular inhibition of PTP opening) is also able to improve mitochondrial function and learning/memory in an aging AD mouse model (26).

A reduction in complex IV activity has been demonstrated in mitochondria from the hippocampus and platelets of AD patients, as well as in AD animal models and AD hybrid cells (27). Aggregation of β A leads to oxidative stress, mitochondrial dysfunction and energy failure prior to the development of plaque pathology (28). Activation of the DNA repairing enzyme PARP in AD due to overproduction of ROS in NADPH oxidase leads to consumption of NAD and restriction of substrates (Figure 1) for mitochondrial complex I, resulting in collapse in bioenergetics and cell death (29, 30). Provision of mitochondrial substrates can prevent amyloid induced cell death (17, 21). A perturbation in mitochondrial dynamics has also been described in AD human brain and cell models. Fragmented mitochondria are seen in AD hippocampus in association with a downregulation of mitochondrial fusion proteins (MFN-1, MFN-2, OPA-1), with an increase in expression of the mitochondrial fission protein Fis-1 (31).

Mitochondria also can regulate aggregation of β A, tau or alpha-synuclein. Inorganic polyphosphate plays important signalling role in brain (32, 33) and in mammalian cells produced in mitochondria (34). Inorganic polyphosphate accelerates aggregation of β A, tau and alpha-synuclein forming non-toxic fibrils playing role cytoprotective modifier (35).

Role of tau in mitochondrial physiology and pathology

Tau protein (tubulin-associated unit) refers to microtubule-associated proteins. It is a soluble, natively unfolded, and phosphorylated protein, ubiquitously expressed in most tissues and organs. This protein exists as six alternatively spliced isoforms and is encoded by a single gene, *mapt*, that is located on chromosome 17 in humans (36). Tau is found in all cellular and subcellular compartments but is most prominent in the axons of neurons of the central nervous system (37, 38). Tau protein plays an important role in neuronal physiology, in microtubule assembly and dynamics (39), in promoting axonal out growth (40), axonal transport and in signal transduction (41). Physiological and pathological activity of tau is dependent on the phosphorylation (tau is phosphoprotein) and alternative splicing and on the level of aggregation. The soluble prefibrillar aggregates of tau proteins cause the most damage to neurons. In disease, tau dissociates from microtubules and forms large, primarily intracellular, β -sheet rich fibrils (42). Tau protein is involved in the pathogenesis of many neurodegenerative diseases, specifically in Alzheimer's disease and frontotemporal dementia. Pathologies and dementias of the nervous system are associated with tau proteins that have become defective and no longer stabilize

microtubules properly. The abnormal tau function leads to the deficits in fast axonal transport, dystrophic neurites, and abnormal mitochondrial distribution (43-45). This abnormal distribution of mitochondria is more likely to be induced by impairment the fission and fusion of mitochondria by tau (46). It also have been shown that in human tau transgenic mice and flies, F-actin is increased, which disrupts the physical association of mitochondria and the fission protein DRP1 (Figure 1), leading to mitochondrial elongation (46). The resulting neurotoxicity can be rescued either by reducing mitochondrial fusion, or by enhancing fission, or by reversing actin stabilization. The possible effect of tau on mitochondrial complex I have been shown triple knockout Alzheimer's disease mouse mitochondria (47). The 10+16 intronic mutation in MAPT gene, encoding tau increase in the production of 4R tau isoforms, which are more prone to aggregation. Human iPSC derived neurons with this mutation are associated with partially suppressed complex I-driven respiration that lead to F1Fo-ATPase to be switched in reverse mode. This combination increased mitochondrial membrane potential that trigger ROS production in electron transport chain which causes oxidative stress and cell death (48).

Role of α -synuclein in mitochondrial physiology and pathology

α -Synuclein is strongly implicated in pathology of Parkinson's disease as a main component of Lewy body – neuronal aggregated inclusions. Lewy body is one of the major pathological hallmarks of this neurodegenerative disorder. One of autosomal-dominant familial Parkinson's disease can be attributed solely to mutations in the SNCA gene (which encoded α -synuclein) or by genetic duplication or triplication of the wild-type SCNA locus (49). Native monomeric form of α -synuclein is soluble protein which aggregates to form insoluble fibrils via a series of conformational changes including most toxic intermediates – oligomeric.

Monomeric α -synuclein plays important physiological roles in synaptic signal transduction (50, 51) and as a regulator ATP production (Figure 1). Thus, monomeric α -synuclein binds F0-F1-ATPsynthase and increase efficiency of this enzyme to produce ATP (52).

Monomeric, oligomeric and fibrillary α -synucleins are able to penetrate through plasma and intracellular membranes (53, 54). The high degree of curvature of mitochondrial membranes and the presence of cardiolipin contribute to the interaction of α -synuclein with this organelle. It is known that α -synuclein preferentially binds to negatively charged lipids

(55-58). Previously, it has been found that α -synuclein specifically binds to mitochondria but no other cell organelles (53, 59).

Parkinson's disease is linked to mitochondrial abnormalities more than any other neurodegenerative disorder. It has been proven by toxins (rotenone and MPTP) and by the fact that most of the familial forms of Parkinson's disease are associated with mitochondria. Pathogenesis of Parkinson's disease is characterized by decreased activity of mitochondrial respiratory chain complex I in the nigrostriatal system by 25-30%. Importantly, oligomeric α -synuclein is able to inhibit complex I (60, 61) or even damage this component of electron transport chain (62). In agreement to this oligomeric α -synuclein had no effect on the neurons with complex I mutation (63). In transgenic mice overexpressing a wild-type α -synuclein occur not only breach morphology, loss of mitochondrial membrane potential ($\Delta\psi$), and fragmentation of the mitochondria, predisposing to neurodegeneration (64-66).

Mitochondria play important role in maintenance of calcium homeostasis in physiology and pathology (67). For monomeric and oligomeric α -synuclein shown ability to stimulate calcium signal (Figure 1) by incorporation into plasma membrane and forming a pore (68, 69). Mitochondrial calcium overload and reactive oxygen species are the major triggers for mitochondrial permeability transition pore (mPTP) (70). Although activation of production of ROS in mitochondria by α -synuclein is disputable it was shown recently that β -sheet-rich α -synuclein oligomers, and to a lesser extent α -synuclein fibrils are initiated of increase ROS production and lipid peroxidation (71, 72) Ability of oligomeric (but not monomeric) α -synuclein to induce both calcium rise and ROS production increases the probability of mPTP opening. However, using of purified recombinant human α -synuclein on isolated mitochondria it has been shown that the addition of oligomeric forms of α -synuclein reduced retention time exogenously added Ca^{2+} , promoted of Ca^{2+} induced swelling and mitochondrial depolarization and accelerated secretion of cytochrome C. Inhibition of mPTP prevented these oligomer-induced changes of mitochondrial parameters (61).

In all eukaryotic cells, endoplasmic reticulum (ER) and mitochondria interact closely using a specific sub-domains of ER and MITO membrane, forming MAM structures (mitochondria-associated membranes, see Figure 1) (73). MAM site of ER has a unique lipid composition, enriched of cholesterol and the anionic phospholipids, with the characteristics of the lipid raft (74). It has been shown that α -synuclein affects a key MAM function - calcium transport between the ER and mitochondria and the wild type α -

synuclein from cell lines and brain tissue of human and mouse is present in MAM structures (73, 75). It was found that pathogenic point mutations in human α -synuclein result in its reduced association with MAM, coincident with a lower degree of apposition of ER with mitochondria, a decrease in MAM function, and an increase in mitochondrial fragmentation compared with wild-type (76).

α -Synuclein can interfere with number of mitochondrial transport proteins including TOM machinery and VDAC (57). Thus, it was shown that certain types of post-translationally modified α -synuclein binds with high affinity to the receptor TOM20, related to mitochondrial protein importing machinery TOM. This binding prevented the interaction of TOM20 with its co-receptor, TOM22, and impaired mitochondrial protein import (77).

Huntingtin protein and mitochondria

Huntington's disease is neurodegenerative disorder caused by mutation of a CAG repeat located in exon 1 of Huntingtin gene. Vulnerability and degeneration of striatal neurons are the first obvious signs of early-grade Huntington's disease. Importantly, a mutation of huntingtin protein (Htt) changes the location of this protein from the nucleus and cytoplasm for wild type Htt to mitochondria for mutant Htt (78, 79). For wild type Htt the role in vesicle trafficking, secretion pathways and apoptosis was demonstrated (80, 81).

The Htt function should affect a multitude of signalling pathways that could be confirmed by lethality of *htt* knockout mouse model (82).

The co-localisation of mutant Htt to the mitochondria leads to inhibition of electron transport chain and reduction of energy levels in Huntington's disease which initiates striatal cell death (Figure 1). Expression of mutant Htt in mice resulted in a reduced mitochondrial membrane potential, suggesting that one or more of the electron transport chain complexes are not working correctly. Thus, in mice with mutated Htt activity of the complex II and complex IV is reduced and expression of mitochondrial complex II is also significantly decreased (83-85). Lower mitochondrial membrane potential in neurons with mutated Htt leads to dramatically reduced calcium buffering capacity (86). Lower mitochondrial buffering capacity in cells with mutated Htt could be the reason for induction of mitochondrial PTP and cell death (87).

Conclusions and future directions

Despite the difference in toxicity between β A, α -synuclein, tau and Htt and their original location (extracellular, intracellular or membranes) all these misfolded proteins and peptides have similarities in the mechanisms of neurodegeneration induced by their aggregates. One of the most important steps is an induction of oligomerisation of these peptides. Considering possible role of monomers (α -synuclein, tau and huntingtin) in cell physiology aggregation of these peptides is the initial step in the mechanism of their toxicity. Although all these misfolded proteins initiate ROS production and inhibit mitochondrial respiration via different mechanisms, it results in induction of cell death through opening of mPTP.

Funding: This work was supported by the Federal Targeted Programme for Research and Development in Priority Areas of Development of the Russian Scientific and Technological Complex for 2014-2020 [grant number 14.616.21.0054, project identifier RFMEFI61615X0054].

References

1. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO molecular medicine*. 2016;8(6):595-608.
2. Liu F, Gong CX. Tau exon 10 alternative splicing and tauopathies. *Molecular neurodegeneration*. 2008;3:8.
3. Spillantini MG, Goedert M. Tau pathology and neurodegeneration. *The Lancet Neurology*. 2013;12(6):609-22.
4. Gandhi S, Wood NW. Molecular pathogenesis of Parkinson's disease. *Human molecular genetics*. 2005;14 Spec No. 2:2749-55.
5. Saudou F, Humbert S. The Biology of Huntingtin. *Neuron*. 2016;89(5):910-26.
6. Burchell VS, Gandhi S, Deas E, Wood NW, Abramov AY, Plun-Favreau H. Targeting mitochondrial dysfunction in neurodegenerative disease: Part I. Expert opinion on therapeutic targets. 2010;14(4):369-85.
7. Angelova PR, Abramov AY. Functional role of mitochondrial reactive oxygen species in physiology. *Free radical biology & medicine*. 2016;100:81-5.
8. Balaban RS, Nemoto S, Finkel T. Mitochondria, oxidants, and aging. *Cell*. 2005;120(4):483-95.
9. Finberg JP. Update on the pharmacology of selective inhibitors of MAO-A and MAO-B: focus on modulation of CNS monoamine neurotransmitter release. *Pharmacology & therapeutics*. 2014;143(2):133-52.
10. Vaarmann A, Gandhi S, Abramov AY. Dopamine induces Ca²⁺ signaling in astrocytes through reactive oxygen species generated by monoamine oxidase. *The Journal of biological chemistry*. 2010;285(32):25018-23.
11. Ruan L, Zhou C, Jin E, Kucharavy A, Zhang Y, Wen Z, et al. Cytosolic proteostasis through importing of misfolded proteins into mitochondria. *Nature*. 2017;543(7645):443-6.
12. Blass JP, Gibson GE. The role of oxidative abnormalities in the pathophysiology of Alzheimer's disease. *Revue neurologique*. 1991;147(6-7):513-25.
13. Casley CS, Canevari L, Land JM, Clark JB, Sharpe MA. Beta-amyloid inhibits integrated mitochondrial respiration and key enzyme activities. *Journal of neurochemistry*. 2002;80(1):91-100.
14. Longo VD, Viola KL, Klein WL, Finch CE. Reversible inactivation of superoxide-sensitive aconitase in A β 1-42-treated neuronal cell lines. *Journal of neurochemistry*. 2000;75(5):1977-85.

15. Canevari L, Clark JB, Bates TE. beta-Amyloid fragment 25-35 selectively decreases complex IV activity in isolated mitochondria. *FEBS letters*. 1999;457(1):131-4.
16. Casley CS, Land JM, Sharpe MA, Clark JB, Duchen MR, Canevari L. Beta-amyloid fragment 25-35 causes mitochondrial dysfunction in primary cortical neurons. *Neurobiology of disease*. 2002;10(3):258-67.
17. Abramov AY, Canevari L, Duchen MR. Beta-amyloid peptides induce mitochondrial dysfunction and oxidative stress in astrocytes and death of neurons through activation of NADPH oxidase. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2004;24(2):565-75.
18. Ionov M, Burchell V, Klajnert B, Bryszewska M, Abramov AY. Mechanism of neuroprotection of melatonin against beta-amyloid neurotoxicity. *Neuroscience*. 2011;180:229-37.
19. Abramov AY, Canevari L, Duchen MR. Changes in intracellular calcium and glutathione in astrocytes as the primary mechanism of amyloid neurotoxicity. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2003;23(12):5088-95.
20. Abramov AY, Canevari L, Duchen MR. Calcium signals induced by amyloid beta peptide and their consequences in neurons and astrocytes in culture. *Biochimica et biophysica acta*. 2004;1742(1-3):81-7.
21. Abramov AY, Duchen MR. The role of an astrocytic NADPH oxidase in the neurotoxicity of amyloid beta peptides. *Philosophical transactions of the Royal Society of London Series B, Biological sciences*. 2005;360(1464):2309-14.
22. Parks JK, Smith TS, Trimmer PA, Bennett JP, Jr., Parker WD, Jr. Neurotoxic Abeta peptides increase oxidative stress in vivo through NMDA-receptor and nitric-oxide-synthase mechanisms, and inhibit complex IV activity and induce a mitochondrial permeability transition in vitro. *Journal of neurochemistry*. 2001;76(4):1050-6.
23. Shevtzova EF, Kireeva EG, Bachurin SO. Effect of beta-amyloid peptide fragment 25-35 on nonselective permeability of mitochondria. *Bulletin of experimental biology and medicine*. 2001;132(6):1173-6.
24. Abramov AY, Jacobson J, Wientjes F, Hothersall J, Canevari L, Duchen MR. Expression and modulation of an NADPH oxidase in mammalian astrocytes. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2005;25(40):9176-84.
25. Abramov AY, Scorziello A, Duchen MR. Three distinct mechanisms generate oxygen free radicals in neurons and contribute to cell death during anoxia and reoxygenation. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2007;27(5):1129-38.
26. Du H, Guo L, Fang F, Chen D, Sosunov AA, McKhann GM, et al. Cyclophilin D deficiency attenuates mitochondrial and neuronal perturbation and ameliorates learning and memory in Alzheimer's disease. *Nature medicine*. 2008;14(10):1097-105.
27. Du H, Guo L, Yan S, Sosunov AA, McKhann GM, Yan SS. Early deficits in synaptic mitochondria in an Alzheimer's disease mouse model. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;107(43):18670-5.
28. Caspersen C, Wang N, Yao J, Sosunov A, Chen X, Lustbader JW, et al. Mitochondrial Abeta: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 2005;19(14):2040-1.
29. Abeti R, Abramov AY, Duchen MR. Beta-amyloid activates PARP causing astrocytic metabolic failure and neuronal death. *Brain : a journal of neurology*. 2011;134(Pt 6):1658-72.
30. Angelova PR, Abramov AY. Interaction of neurons and astrocytes underlies the mechanism of Abeta-induced neurotoxicity. *Biochemical Society transactions*. 2014;42(5):1286-90.
31. Wang X, Su B, Siedlak SL, Moreira PI, Fujioka H, Wang Y, et al. Amyloid-beta overproduction causes abnormal mitochondrial dynamics via differential modulation of mitochondrial fission/fusion proteins. *Proceedings of the National Academy of Sciences of the United States of America*. 2008;105(49):19318-23.
32. Holmstrom KM, Marina N, Baev AY, Wood NW, Gourine AV, Abramov AY. Signalling properties of inorganic polyphosphate in the mammalian brain. *Nature communications*. 2013;4:1362.
33. Angelova PR, Baev AY, Berezhnov AV, Abramov AY. Role of inorganic polyphosphate in mammalian cells: from signal transduction and mitochondrial metabolism to cell death. *Biochemical Society transactions*. 2016;44(1):40-5.
34. Pavlov E, Aschar-Sobbi R, Campanella M, Turner RJ, Gomez-Garcia MR, Abramov AY. Inorganic polyphosphate and energy metabolism in mammalian cells. *The Journal of biological chemistry*. 2010;285(13):9420-8.
35. Cremers CM, Knoefler D, Gates S, Martin N, Dahl JU, Lempart J, et al. Polyphosphate: A Conserved Modifier of Amyloidogenic Processes. *Molecular cell*. 2016;63(5):768-80.
36. Neve RL, Harris P, Kosik KS, Kurnit DM, Donlon TA. Identification of cDNA clones for the human microtubule-associated protein tau and chromosomal localization of the genes for tau and microtubule-associated protein 2. *Brain research*. 1986;387(3):271-80.
37. Gorath M, Stahnke T, Mronga T, Goldbaum O, Richter-Landsberg C. Developmental changes of tau protein and mRNA in cultured rat brain oligodendrocytes. *Glia*. 2001;36(1):89-101.
38. Kumar P, Jha NK, Jha SK, Ramani K, Ambasta RK. Tau phosphorylation, molecular chaperones, and ubiquitin E3 ligase: clinical relevance in Alzheimer's disease. *Journal of Alzheimer's disease : JAD*. 2015;43(2):341-61.

39. Drubin DG, Kirschner MW. Tau protein function in living cells. *The Journal of cell biology*. 1986;103(6 Pt 2):2739-46.
40. Wang Y, Mandelkow E. Tau in physiology and pathology. *Nature reviews Neuroscience*. 2016;17(1):5-21.
41. Mazzaro N, Barini E, Spillantini MG, Goedert M, Medini P, Gasparini L. Tau-Driven Neuronal and Neurotrophic Dysfunction in a Mouse Model of Early Tauopathy. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2016;36(7):2086-100.
42. Spillantini MG, Goedert M. Tau protein pathology in neurodegenerative diseases. *Trends in neurosciences*. 1998;21(10):428-33.
43. Stokin GB, Lillo C, Falzone TL, Brusch RG, Rockenstein E, Mount SL, et al. Axonopathy and transport deficits early in the pathogenesis of Alzheimer's disease. *Science*. 2005;307(5713):1282-8.
44. Praprotnik D, Smith MA, Richey PL, Vinters HV, Perry G. Filament heterogeneity within the dystrophic neurites of senile plaques suggests blockage of fast axonal transport in Alzheimer's disease. *Acta neuropathologica*. 1996;91(3):226-35.
45. Wang X, Su B, Lee HG, Li X, Perry G, Smith MA, et al. Impaired balance of mitochondrial fission and fusion in Alzheimer's disease. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2009;29(28):9090-103.
46. DuBoff B, Gotz J, Feany MB. Tau promotes neurodegeneration via DRP1 mislocalization in vivo. *Neuron*. 2012;75(4):618-32.
47. Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, Meier F, et al. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;106(47):20057-62.
48. Esteras N, Rohrer JD, Hardy J, Wray S, Abramov AY. Mitochondrial hyperpolarization in iPSC-derived neurons from patients of FTDP-17 with 10+16 MAPT mutation leads to oxidative stress and neurodegeneration. *Redox biology*. 2017;12:410-22.
49. Singleton AB, Farrer M, Johnson J, Singleton A, Hague S, Kachergus J, et al. alpha-Synuclein locus triplication causes Parkinson's disease. *Science*. 2003;302(5646):841.
50. Burre J, Sharma M, Tsetsenis T, Buchman V, Etherton MR, Sudhof TC. Alpha-synuclein promotes SNARE-complex assembly in vivo and in vitro. *Science*. 2010;329(5999):1663-7.
51. Vargas KJ, Makani S, Davis T, Westphal CH, Castillo PE, Chandra SS. Synucleins regulate the kinetics of synaptic vesicle endocytosis. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2014;34(28):9364-76.
52. Ludtmann MH, Angelova PR, Ninkina NN, Gandhi S, Buchman VL, Abramov AY. Monomeric Alpha-Synuclein Exerts a Physiological Role on Brain ATP Synthase. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2016;36(41):10510-21.
53. Nakamura K, Nemani VM, Wallender EK, Kaehlcke K, Ott M, Edwards RH. Optical reporters for the conformation of alpha-synuclein reveal a specific interaction with mitochondria. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2008;28(47):12305-17.
54. Cremades N, Cohen SI, Deas E, Abramov AY, Chen AY, Orte A, et al. Direct observation of the interconversion of normal and toxic forms of alpha-synuclein. *Cell*. 2012;149(5):1048-59.
55. Davidson WS, Jonas A, Clayton DF, George JM. Stabilization of alpha-synuclein secondary structure upon binding to synthetic membranes. *The Journal of biological chemistry*. 1998;273(16):9443-9.
56. Pfefferkorn CM, Jiang Z, Lee JC. Biophysics of alpha-synuclein membrane interactions. *Biochimica et biophysica acta*. 2012;1818(2):162-71.
57. Rostovtseva TK, Gurnev PA, Protchenko O, Hoogerheide DP, Yap TL, Philpott CC, et al. alpha-Synuclein Shows High Affinity Interaction with Voltage-dependent Anion Channel, Suggesting Mechanisms of Mitochondrial Regulation and Toxicity in Parkinson Disease. *The Journal of biological chemistry*. 2015;290(30):18467-77.
58. Shvadchak VV, Yushchenko DA, Pievo R, Jovin TM. The mode of alpha-synuclein binding to membranes depends on lipid composition and lipid to protein ratio. *FEBS letters*. 2011;585(22):3513-9.
59. Martin LJ, Pan Y, Price AC, Sterling W, Copeland NG, Jenkins NA, et al. Parkinson's disease alpha-synuclein transgenic mice develop neuronal mitochondrial degeneration and cell death. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2006;26(1):41-50.
60. Loeb V, Yakunin E, Saada A, Sharon R. The transgenic overexpression of alpha-synuclein and not its related pathology associates with complex I inhibition. *The Journal of biological chemistry*. 2010;285(10):7334-43.
61. Luth ES, Stavrovskaya IG, Bartels T, Kristal BS, Selkoe DJ. Soluble, prefibrillar alpha-synuclein oligomers promote complex I-dependent, Ca²⁺-induced mitochondrial dysfunction. *The Journal of biological chemistry*. 2014;289(31):21490-507.
62. Devi L, Raghavendran V, Prabhu BM, Avadhani NG, Anandatheerthavarada HK. Mitochondrial import and accumulation of alpha-synuclein impair complex I in human dopaminergic neuronal cultures and Parkinson disease brain. *The Journal of biological chemistry*. 2008;283(14):9089-100.

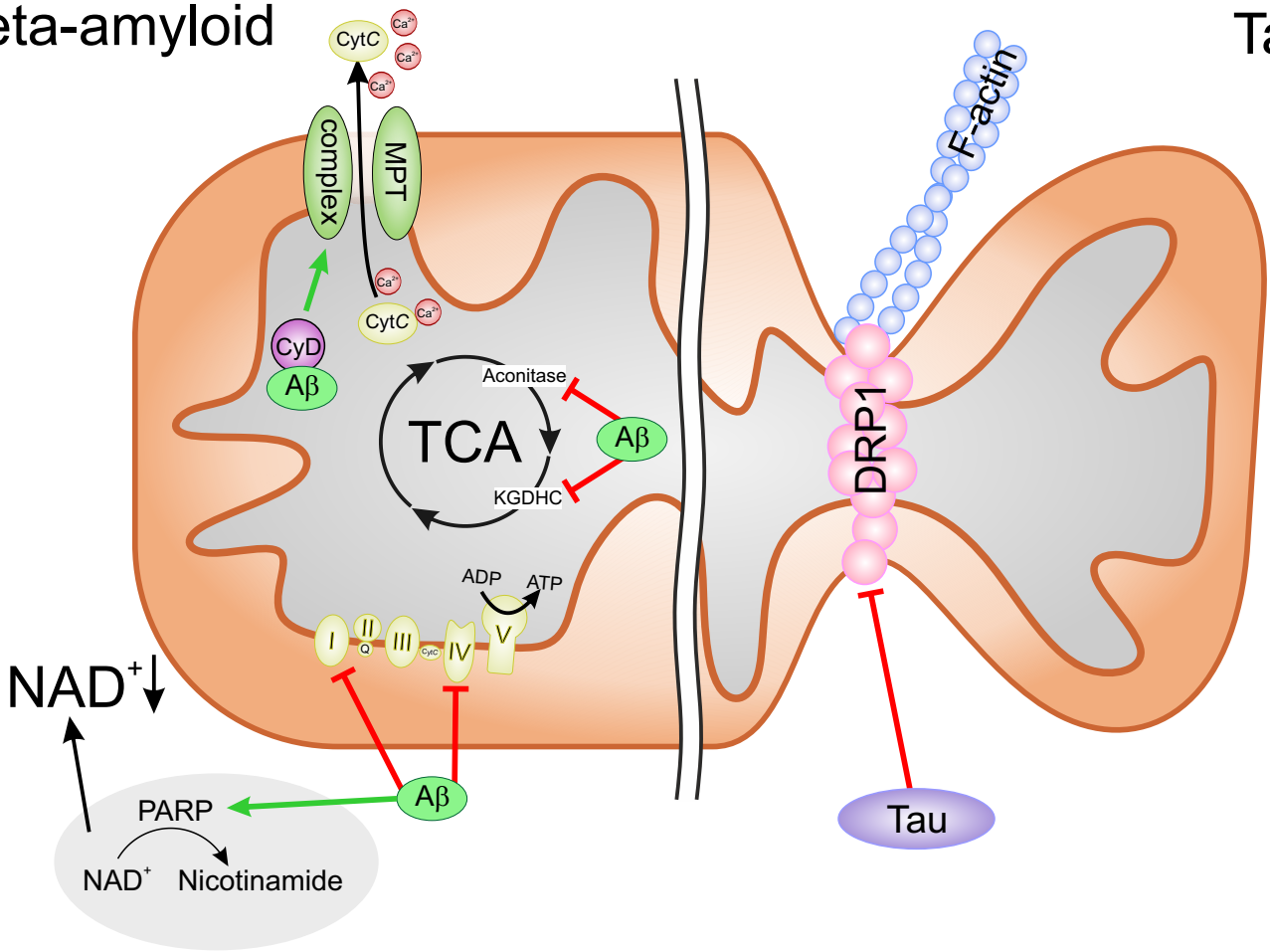
63. Reeve AK, Ludtmann MH, Angelova PR, Simcox EM, Horrocks MH, Klenerman D, et al. Aggregated alpha-synuclein and complex I deficiency: exploration of their relationship in differentiated neurons. *Cell death & disease*. 2015;6:e1820.
64. Zhu Y, Duan C, Lu L, Gao H, Zhao C, Yu S, et al. alpha-Synuclein overexpression impairs mitochondrial function by associating with adenylate translocator. *The international journal of biochemistry & cell biology*. 2011;43(5):732-41.
65. Nakamura K, Nemani VM, Azarbal F, Skibinski G, Levy JM, Egami K, et al. Direct membrane association drives mitochondrial fission by the Parkinson disease-associated protein alpha-synuclein. *The Journal of biological chemistry*. 2011;286(23):20710-26.
66. Mullin S, Schapira A. alpha-Synuclein and mitochondrial dysfunction in Parkinson's disease. *Molecular neurobiology*. 2013;47(2):587-97.
67. Abeti R, Abramov AY. Mitochondrial Ca(2+) in neurodegenerative disorders. *Pharmacological research*. 2015;99:377-81.
68. Angelova PR, Ludtmann MH, Horrocks MH, Negoda A, Cremades N, Klenerman D, et al. Ca²⁺ is a key factor in alpha-synuclein-induced neurotoxicity. *Journal of cell science*. 2016;129(9):1792-801.
69. Zakharov SD, Hulleman JD, Dutseva EA, Antonenko YN, Rochet JC, Cramer WA. Helical alpha-synuclein forms highly conductive ion channels. *Biochemistry*. 2007;46(50):14369-79.
70. Bernardi P, Rasola A, Forte M, Lippe G. The Mitochondrial Permeability Transition Pore: Channel Formation by F-ATP Synthase, Integration in Signal Transduction, and Role in Pathophysiology. *Physiological reviews*. 2015;95(4):1111-55.
71. Deas E, Cremades N, Angelova PR, Ludtmann MH, Yao Z, Chen S, et al. Alpha-Synuclein Oligomers Interact with Metal Ions to Induce Oxidative Stress and Neuronal Death in Parkinson's Disease. *Antioxidants & redox signaling*. 2016;24(7):376-91.
72. Angelova PR, Horrocks MH, Klenerman D, Gandhi S, Abramov AY, Shchepinov MS. Lipid peroxidation is essential for alpha-synuclein-induced cell death. *Journal of neurochemistry*. 2015;133(4):582-9.
73. Hayashi T, Rizzuto R, Hajnoczky G, Su TP. MAM: more than just a housekeeper. *Trends in cell biology*. 2009;19(2):81-8.
74. Hayashi T, Fujimoto M. Detergent-resistant microdomains determine the localization of sigma-1 receptors to the endoplasmic reticulum-mitochondria junction. *Molecular pharmacology*. 2010;77(4):517-28.
75. Cali T, Ottolini D, Negro A, Brini M. alpha-Synuclein controls mitochondrial calcium homeostasis by enhancing endoplasmic reticulum-mitochondria interactions. *The Journal of biological chemistry*. 2012;287(22):17914-29.
76. Area-Gomez E. Assessing the function of mitochondria-associated ER membranes. *Methods in enzymology*. 2014;547:181-97.
77. Di Maio R, Barrett PJ, Hoffman EK, Barrett CW, Zharikov A, Borah A, et al. alpha-Synuclein binds to TOM20 and inhibits mitochondrial protein import in Parkinson's disease. *Science translational medicine*. 2016;8(342):342ra78.
78. Hoogeveen AT, Willemsen R, Meyer N, de Rooij KE, Roos RA, van Ommen GJ, et al. Characterization and localization of the Huntington disease gene product. *Human molecular genetics*. 1993;2(12):2069-73.
79. Rockabrand E, Slepko N, Pantalone A, Nukala VN, Kazantsev A, Marsh JL, et al. The first 17 amino acids of Huntingtin modulate its sub-cellular localization, aggregation and effects on calcium homeostasis. *Human molecular genetics*. 2007;16(1):61-77.
80. Brandstaetter H, Kruppa AJ, Buss F. Huntingtin is required for ER-to-Golgi transport and for secretory vesicle fusion at the plasma membrane. *Disease models & mechanisms*. 2014;7(12):1335-40.
81. Rigamonti D, Bauer JH, De-Fraja C, Conti L, Sipione S, Sciorati C, et al. Wild-type huntingtin protects from apoptosis upstream of caspase-3. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2000;20(10):3705-13.
82. Nasir J, Floresco SB, O'Kusky JR, Diewert VM, Richman JM, Zeisler J, et al. Targeted disruption of the Huntington's disease gene results in embryonic lethality and behavioral and morphological changes in heterozygotes. *Cell*. 1995;81(5):811-23.
83. Bae BI, Xu H, Igarashi S, Fujimuro M, Agrawal N, Taya Y, et al. p53 mediates cellular dysfunction and behavioral abnormalities in Huntington's disease. *Neuron*. 2005;47(1):29-41.
84. Benchoua A, Trioulier Y, Zala D, Gaillard MC, Lefort N, Dufour N, et al. Involvement of mitochondrial complex II defects in neuronal death produced by N-terminus fragment of mutated huntingtin. *Molecular biology of the cell*. 2006;17(4):1652-63.
85. Benchoua A, Trioulier Y, Diguët E, Malgorn C, Gaillard MC, Dufour N, et al. Dopamine determines the vulnerability of striatal neurons to the N-terminal fragment of mutant huntingtin through the regulation of mitochondrial complex II. *Human molecular genetics*. 2008;17(10):1446-56.
86. Panov AV, Gutekunst CA, Leavitt BR, Hayden MR, Burke JR, Strittmatter WJ, et al. Early mitochondrial calcium defects in Huntington's disease are a direct effect of polyglutamines. *Nature neuroscience*. 2002;5(8):731-6.
87. Lim D, Fedrizzi L, Tartari M, Zuccato C, Cattaneo E, Brini M, et al. Calcium homeostasis and mitochondrial dysfunction in striatal neurons of Huntington disease. *The Journal of biological chemistry*. 2008;283(9):5780-9.

Figure legend

Figure 1. Effects of the major misfolded proteins on mitochondria. Beta-amyloid, tau, alpha-synuclein and huntingtin protein have a direct effect on mitochondria. Oligomeric beta-amyloid, alpha-synuclein and huntingtin protein inhibit complexes of electron transport chain, tau play important role in mitochondrial dynamics. Mitochondrial dysfunction induced by aggregated proteins lead to neuronal cell death

Beta-amyloid

Tau



Alpha-synuclein

Huntingtin protein

