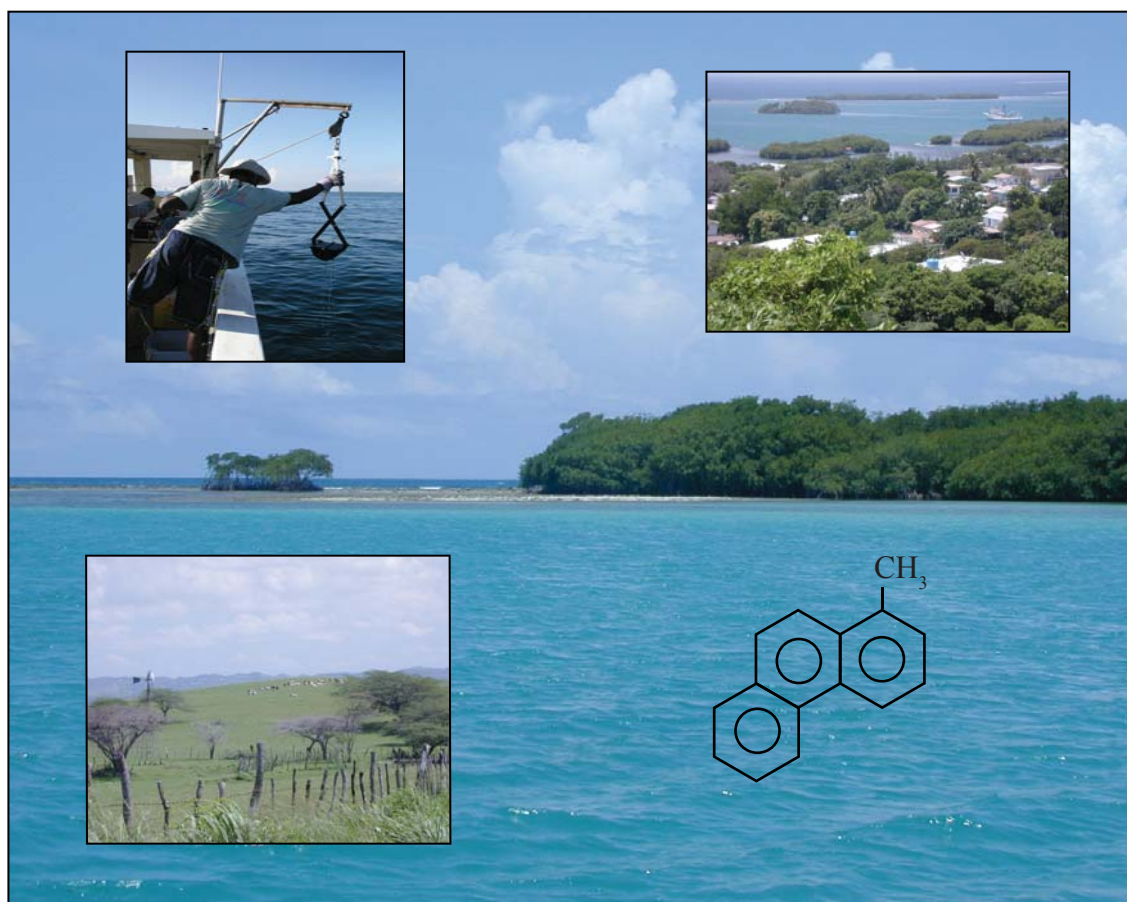


An Assessment of Chemical Contaminants in the Marine Sediments of Southwest Puerto Rico



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An Assessment of Chemical Contaminants in the Marine Sediments of Southwest Puerto Rico

Anthony S. Pait¹, David R. Whitall¹, Christopher F.G. Jeffrey¹,
Chris Caldwell¹, Andrew L. Mason¹, John D. Christensen¹, Mark E. Monaco¹, and Juan
Ramirez²

¹Center for Coastal Monitoring and Assessment
NOAA/NOS/NCCOS, 1305 East/West Highway, Silver Spring, MD 20910

²TDI-Brooks International, 1902 Pinon, College Station, TX 77845

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Carlos M. Gutierrez
Secretary

National Oceanic and
Atmospheric Administration

Conrad C. Lautenbacher, Jr.
Vice Admiral (Ret.), Administrator

National Ocean Service

John H. Dunnigan
Assistant Administrator

Table of Contents

List of Tables.....	iv
List of Figures.....	iv
I. Introduction.....	1
II. Assessment Framework.....	2
III. Materials and Methods.....	4
Study Area.....	4
Sampling Design.....	6
Compounds Analyzed.....	10
Statistical and Spatial Analyses.....	12
IV. Effects of Contaminants on Corals.....	13
Petroleum Hydrocarbons.....	13
Pesticides.....	21
Trace Elements.....	21
Nutrients.....	22
V. Results and Discussion.....	22
Field Data.....	22
Total Organic Carbon and Grain Size.....	22
Polycyclic Aromatic Hydrocarbons.....	22
Polychlorinated Biphenyls.....	33
Organochlorine Pesticides.....	38
Polybrominated Diphenyl Ethers.....	42
Butyltins.....	44
Trace Elements.....	46
Arsenic.....	46
Chromium.....	48
Nickel.....	51
Copper.....	53
Other Contaminants.....	57
<i>Clostridium perfringens</i>	57
Nutrients.....	58
Irgarol 1051.....	60
Contaminants and Coral Species Richness.....	62
Contaminant Modeling.....	62
Total PAHs and Coral Species Richness.....	65

VI. Summary and Conclusions.....	66
Project and Framework	66
Chemical Contaminant Concentrations.....	67
Contaminants and Coral Species Richness	67
Next Steps	68
VII. Acknowledgments	68
VIII. Literature Cited.....	70
IX. Appendices.....	78
A. Sample sites from May 2005 mission	78
B. Field data from May 2005 mission.....	79
C. Total organic carbon and grain size distribution in sediment samples from Southwest Puerto Rico, May 2005 mission.....	81
D. Total organic carbon and grain size distribution in sediment samples from Southwest Puerto Rico, August 2005 mission	82
E. Polycyclic aromatic hydrocarbons (PAHs) in sediment samples from Southwest Puerto Rico (ng/ dry g)	83
F. Polychlorinated biphenyls (PCBs) in sediment samples from Southwest Puerto Rico (ng/dry g).....	87
G. Organochlorine pesticides in sediment samples from Southwest Puerto Rico (ng/dry g)	94
H. Polybrominated diphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g).....	103
I. Butyltins in sediment samples from Southwest Puerto Rico (ng Sn/dry g).....	110
J. Trace and major elements in sediment samples from Southwest Puerto Rico (ug/dry g)	111
K. <i>Clostridium perfringens</i> in sediment samples from Southwest Puerto Rico.....	115
L. Total nitrogen and phosphorus detected in water samples from southwest Puerto Rico	116

List of Tables

1. Chemical contaminants analyzed in the sediments from southwest Puerto Rico	11
2. Effects of chemical contaminants on corals.....	14
3. Field data from the August 2005 mission	23
4. Concentration of other selected organochlorine pesticides found in the sediments in southwest Puerto Rico.....	40

List of Figures

1. Assessment framework developed for contaminants and coral condition.....	3
2. Study area in southwest Puerto Rico.....	5
3. Stratified random sampling design used in May 2005 reconnaissance in southwest Puerto Rico.....	7
4. Modeled total organic carbon levels in the sediments	8
5. Sites targeted in southwest Puerto Rico during the August 2005 mission.....	9
6. Map of total PAHs detected in sediments in southwest Puerto Rico	24
7. Total PAHs by site detected in sediments in southwest Puerto Rico	24
8. Concentration of selected PAHs detected in sediments in southwest Puerto Rico	25
9. Regression fit of percent silt and total PAH in the sediment samples from Puerto Rico.....	26
10. Relationship of total organic carbon (TOC) and total PAH concentration in the sediment samples from Puerto Rico.....	26
11. Comparison of total PAH to silt/clay normalized PAH concentrations	27
12. Low molecular weight vs high molecular weight PAHs in the sediments from southwest Puerto Rico.....	28
13. Comparison of selected high molecular weight (HMW) PAHs in sediments sampled from southwest Puerto Rico.....	29
14. Comparison of signatures for selected PAHs in outboard engines with PAHs in sediments in southwest Puerto Rico.....	30
15. Results of ANOVA and pair-wise comparison on transformed total PAH data and habitat type.....	31
16. Map of total PCBs detected in the sediments in southwest Puerto Rico	34
17. Total PCBs detected by site in the sediments in southwest Puerto Rico	34
18. Bivariate fit of total PCB (log10 transformed) and percent silt in the sediments from southwest Puerto Rico.....	35
19. Bivariate fit of total PCB and total organic carbon (TOC) of the sediment.....	36
20. Comparison of total PCB to silt/clay normalized PCB concentrations	36
21. Composition of various Aroclors expressed as a percent of PCB class present in each of the commercial mixtures	37
22. PCB detections in Guanica Bay by level of chlorination.....	37
23. Map of total DDT detected by site in the sediments in southwest Puerto Rico.....	39
24. Total DDT detected by site in the sediments in southwest Puerto Rico	39
25. Relationship of total DDT to (a) %silt and (b) total organic carbon (TOC).....	40

26. Map of endosulfan detected in the sediments in southwest Puerto Rico	41
27. Map of total PBDEs detected by site in the sediments in southwest Puerto Rico	43
28. Total PBDEs detected by site in the sediments in southwest Puerto Rico.....	43
29. Detections of sediment PBDEs in southwest Puerto Rico by class.....	44
30. Butyltins detected in the sediments in southwest Puerto Rico.....	45
31. Map of arsenic detected in the sediments in southwest Puerto Rico	47
32. Arsenic detected by site in the sediments in southwest Puerto Rico	47
33. Map of chromium detected in the sediments in southwest Puerto Rico	49
34. Chromium detected by site in the sediments in southwest Puerto Rico	49
35. Bivariate fit of chromium and %silt (a) and %clay (b) in the sediments.....	50
36. Map of nickel detected in the sediments in southwest Puerto Rico.....	52
37. Nickel detected by site in the sediments in southwest Puerto Rico.....	52
38. Bivariate fit of nickel and %silt (a) and %clay (b) in the sediments.....	53
39. Map of copper detected in the sediments in southwest Puerto Rico.....	54
40. Copper detected by site in the sediments in southwest Puerto Rico.....	54
41. Bivariate fit of copper and %silt (a) and %clay (b) in the sediments.....	55
42. Normalizations of chromium, nickel and copper to aluminum	56
43. Map of <i>Clostridium perfringens</i> detected in the sediments in southwest Puerto Rico	58
44. Total nitrogen in water samples from southwest Puerto Rico	59
45. Total phosphorus in water samples from southwest Puerto Rico	59
46. Detection of the antifoulant compound Irgarol 1051 in water samples from Puerto Rico in May 2005 (circles) and August 2005 (squares).....	61
47. Modeled vs observed concentrations for total PAHs in southwest Puerto Rico	63
48. Indicator Kriging and measured concentrations of total PAH in the sediments in southwest Puerto Rico.....	63
49. Indicator Kriging and measured concentrations of a) total PCBs, b) total DDTs, and c) copper.....	64
50. Kriging of total PAHs and coral species richness	65

An Assessment of Chemical Contaminants in the Marine Sediments of Southwest Puerto Rico

This report summarizes the results of a characterization of chemical contaminants in the sediments in southwest Puerto Rico. The report is part of a project to integrate various analytical specialties to assess linkages between chemical contaminants and the condition of coral reefs. In this phase of the project, over 120 chemical contaminants were analyzed in sediments collected, including a number of organic (e.g., hydrocarbons), inorganic (e.g., metals), and biological (bacterial) compounds/analytes. The report also provides a preliminary analysis of the association between sediment contaminants and coral species richness.

Overall, the levels of chemical contaminants in the study area between Guanica Bay and the town of La Parguera were fairly low. At most of the sites sampled, particularly adjacent to the town of La Parguera, concentrations of organic and inorganic contaminants were below the median values from NOAA's National Status and Trends Program, which monitors the Nation's coastal and estuarine waters for chemical contaminants. Elevated levels of a number of contaminant classes were seen at the two sites sampled within Guanica Bay.

An initial analysis of modeled PAH (hydrocarbon) data and coral species richness (reef building species) indicated a strong negative correlation between the presence of PAHs in the sediments and coral species richness. Additional work is needed to assess possible reasons for this observed pattern.

Introduction

Coral reefs are among the most productive and diverse ecosystems in the world (Bryant *et al.*, 1998). They occur in tropical and semitropical oceans and are home to thousands of species of fish, corals, crustaceans and molluscs. Coral reefs provide a variety of goods and services ranging from commercial and subsistence fisheries, tourism and recreation, sources of new medicines, to natural protection against storms for communities and ports. The global value of coral reefs has been estimated at \$375 billion/year (Costanza *et al.*, 1997).



San Cristobal emergent reef in southwest Puerto Rico.

Worldwide, however, coral reefs are experiencing significant degradation, much of it thought to be related to human activities. Pollution, disease, sedimentation, overfishing, global climate change, invasive species, ship groundings (Waddell *et al.*, 2005), and possibly ocean

acidification (Kleypas *et al.*, 2006), are some of the threats currently faced by coral reefs worldwide.

In their report entitled "America's Living Oceans", the Pew Oceans Commission cited both point and nonpoint sources of pollution as major threats to the oceans (Pew Oceans Commission, 2003). Waddell *et al.* (2005) have described some of the threats posed by pollution to coral reefs of the U.S. and Freely Associated States. Bryant *et al.* (1998) estimated that 58 percent of the world's reefs are at risk from human disturbance. The World Resources Institute (Burke and Maidens, 2004) noted that nearly two-thirds of the coral reefs in the Caribbean are threatened by

human activities. Gardner *et al.* (2003) estimated that coral cover in the Caribbean has been reduced by 80 percent.

In 2002, the U.S. Coral Reef Task Force (USCRTF) adopted the “Puerto Rico Resolution”, which called for the development of Local Action Strategies (LAS) to identify and implement priority actions needed to reduce key threats to valuable coral reef resources (FDEP, 2004). Pollution was identified as a focus area for priority action in the LAS for Puerto Rico, the Virgin Islands and Southeast Florida. Although pollution is frequently cited as impacting coral reef health, the concentration of chemical contaminants present in coral reefs is not well characterized, and typically even less is known regarding linkages between contaminants and coral condition.

Downs *et al.* (2005) concluded that coral decline in a section of the northern Florida Keys is likely related to chemical contaminant exposure, and noted that an analysis of contaminants present would greatly increase the power of determining the impact of this stressor. Developing an understanding of how and to what extent contaminants affect the health of corals and coral reefs would help focus management efforts and optimize the use of finite resources (funds and personnel) to conserve and restore coral reef habitats.

Project Goals. To address these needs, NOAA’s National Centers for Coastal Ocean Science (NCCOS), in partnership with the University of Puerto Rico - Mayaguez, conducted a project to assess chemical contamination in the coral reefs in southwest Puerto Rico, and to develop a better understanding of the relationships between chemical contaminants and coral condition. Objectives were to:

- 1) characterize both organic (*e.g.*, petroleum hydrocarbons) and inorganic (*e.g.*, metals) contaminants in the study area;
- 2) map and determine coral distribution/species richness (biogeography);
- 3) characterize the presence of bacterial pathogens that have been associated with coral disease; and
- 4) develop and implement an assessment

framework that can be applied throughout the Caribbean and elsewhere.

This project directly responds to the USCRTF’s call for an improved understanding of the effects of chemical contaminants on coral reefs. Because there is an overall lack of information available on contaminant effects, and even contaminant concentrations in coral reefs, management efforts may be missing an important and, in some locations, a critical information need for the sustained management of these valuable resources. Knowing how, where, and to what extent chemical contaminants are impacting coral reefs would provide managers and policy makers with the knowledge needed to make better informed decisions regarding both land-use and coastal resource management.

Assessment Framework

The assessment framework developed as part of this project integrates a number of specialties, including environmental chemistry, biogeography, microbiology and geographic information system (GIS) analysis (both terrestrial and marine), to better understand how anthropogenic inputs may affect coral ecosystems. The integration and concurrent analysis of these data types maximizes the likelihood of quantifying linkages between stressors (*e.g.*, chemical contaminants) and coral health. Figure 1 presents a simple schematic of the assessment framework, and shows the types of information brought together as part of the process. The information developed as part of the assessment framework can be used to help coral reef managers determine the degree to which chemical contaminants are impacting corals, so that appropriate management strategies can be put in place.

Chemical Contaminants. In the framework, sediments and coral tissues are analyzed for a suite of chemical contaminants (*e.g.*, petroleum hydrocarbons, metals and pesticides). In the next section, the compounds analyzed in sediments as part of this work will be discussed. An assessment of contaminants in the sediment and coral tissues is an important capability, and provides the means for correlating contaminants with other parameters measured in the framework.

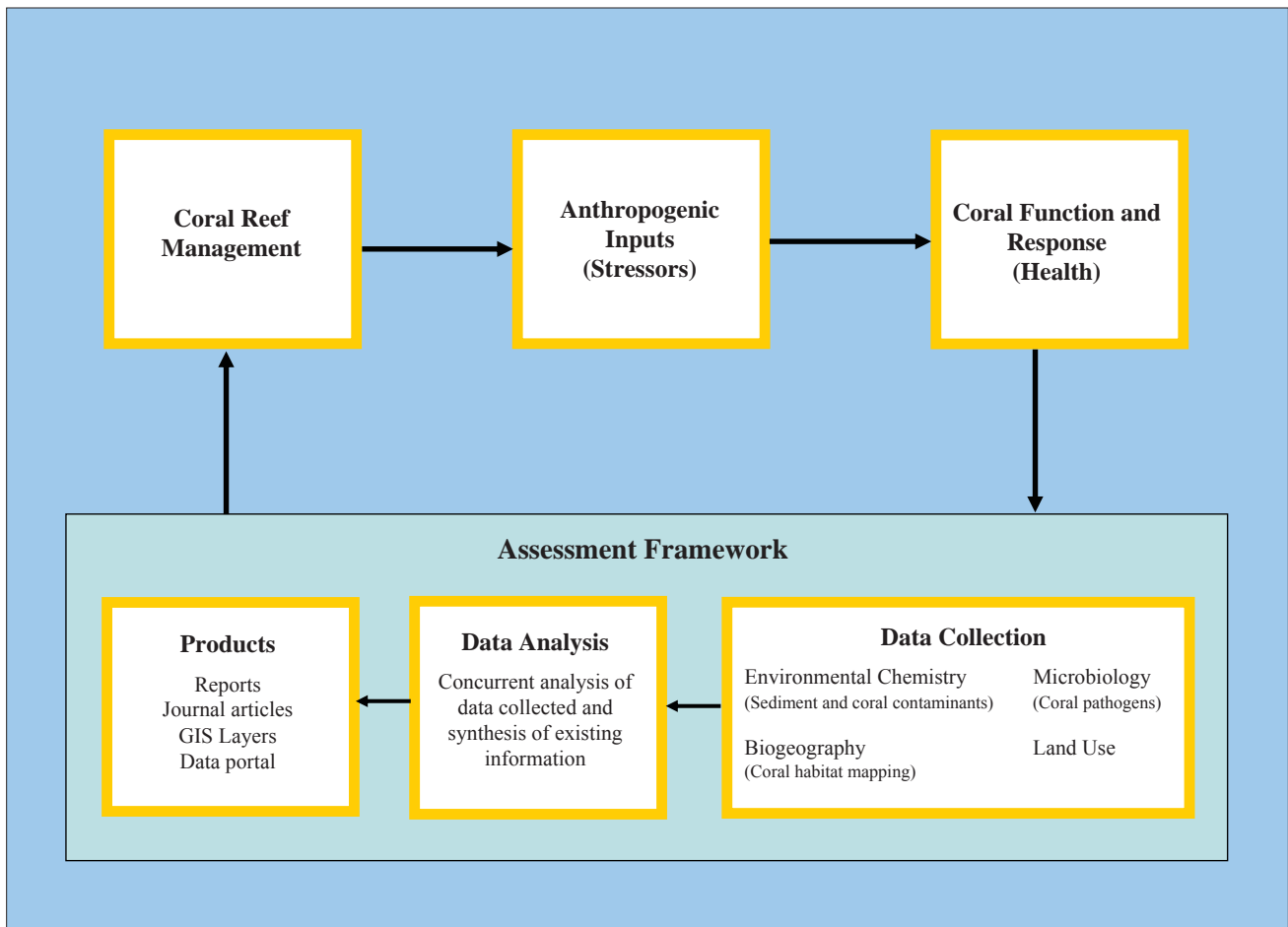


Figure 1. Assessment framework developed for contaminants and coral condition.

Coral Cover. Since 1999, NOAA’s Biogeography Branch has completed extensive mapping and monitoring of the reefs in southwest Puerto Rico and elsewhere in the U.S. Caribbean. These data can be used to help determine relationships between the presence of contaminants and the distribution and abundance of coral species.

Coral Pathogens. The presence of coral pathogens is also determined as part of the framework. In addition to providing a measure of their presence, exploring the relationship between contaminants and coral pathogens may provide a means for determining if chemical contaminants modulate pathogen susceptibility and ultimately the onset of coral disease.

Land Use. The types of contaminants found in the aquatic environment are a function, in part, of the

activities that occur on land. Nonpoint source runoff from agricultural areas can introduce pesticides, nutrients and sediments onto the reefs. Nonpoint source runoff from urban or suburban areas can also introduce a number of contaminant types, ranging from petroleum hydrocarbons to coliform bacteria. Point sources such as wastewater treatment plants or industrial operations also discharge a variety of contaminant types.

Project Partners. The Coastal Oceanographic Assessment, Status and Trends (COAST) Branch, and the Biogeography Branch of NOAA’s Center for Coastal Monitoring and Assessment (CCMA) are working to characterize chemical contaminants within sediments and coral tissues, and to document coral cover/species richness, respectively. COAST operates NOAA’s National Status and Trends

(NS&T) Program, which has monitored the Nation's coastal waters for contaminants and bioeffects for over 20 years. The NS&T Program contains the Mussel Watch Project which monitors approximately 280 coastal and estuarine sites around the Nation for chemical contaminants in both bivalve tissues and sediments. NOAA's Biogeography Branch has worked to characterize and map coastal habitats for over 20 years, and since 1999 has worked specifically in tropical ecosystems. The concurrent collection and analysis of data on chemical contaminants and coral community structure creates a powerful tool to assess contaminant effects on habitat and species distributions.

The University of Puerto Rico-Mayaguez provided expertise in local coral reef ecology, and a base of operations for field work. Partners from NOAA's Center for Coastal Environmental Health and Biomolecular Research and Hollings Marine Laboratory in Charleston, South Carolina provided laboratory space, and analyzed coral tissues for pathogens associated with coral disease. TDI-Brooks International analyzed the sediment samples for contaminants.

Materials and Methods

Study Area

The study area is approximately 23 km long, extending from Guanica Bay in southwest Puerto Rico, west past the town of La Parguera to the Margarita reef complex (Figure 2). The study area was selected based on established partnerships, data availability, and designation of the area as a NOAA funded Coral Reef Ecosystem Study (CRES) site. CRES, funded by NCCOS' Center for Sponsored Coastal Ocean Research (CSCOR), is a 5-year program focused on understanding the causes and effects of reef degradation, and to provide managers

with the information and tools needed to help reverse this degradation. CRES-Parguera is one of two CRES study sites in Puerto Rico. The Puerto Rico Planning Board has classified the terrestrial and marine environments of La Parguera as a Zone of Special Planning. The area has also been designated as a Natural Reserve by the Puerto Rico Department of Natural and Environmental Resources (Garcia

et al., 1998). NOAA's Biogeography Branch has completed extensive coral reef habitat mapping in this and other areas of Puerto Rico, and the University of Puerto Rico-Mayaguez has also done extensive research on the reefs around La Parguera.

Terrestrial Environment.

The southwest coast of Puerto Rico is generally a dry and warm region, and has been classified as a subtropical dry

forest life zone (Garcia *et al.*, 1998). The town of La Parguera is located in the municipality of Lajas, and is home to approximately 26,000 people (U.S. Census Bureau, 2006). Approximately 100,000 visitors come to La Parguera during the year for recreation, and over the last 10 years a number of guest houses, private vacation homes and resorts have been developed (Garcia *et al.*, 1998). A wastewater treatment plant is situated between the town of La Parguera and Isla Guayacan (Figure 2). The facility discharges approximately 228,000 liters of secondary treated effluent per day into a series of percolation ponds (Garcia *et al.*, 1998).

The area around La Parguera has relatively low rainfall and subsequently low runoff. A series of coastal hills known as the Sierra Bermeja separates the narrow coastal plain from the Lajas Valley. The valley drains primarily into Guanica Bay via a man-made canal system. Historically, the Lajas Valley was home to extensive sugarcane production, but now has significant pastureland along with the production of selected vegetable and melon crops (USDA, 2004).



View of La Parguera and Isla Maguayes.



Figure 2. Study area in southwest Puerto Rico. Yellow text denotes a number of the emergent reefs in the study area.

The municipality of Lajas has approximately 42 km² of pastureland (primarily for beef cattle), higher than any other in Puerto Rico, and Lajas ranks second in terms of percent land area devoted to pasture.

Marine Environment. The shelf along the southern coast of the island extends approximately 10 km from the shore, and then drops off from about 20 meters. In some areas this drop off is fairly abrupt (Garcia-Sais *et al.*, 2004), plunging to depths of 350 meters or more. The shelf, composed mainly of carbonates, was formed during the Cretaceous Period and then flooded around 5,000 - 9,000 years ago as a result of sea level rise (Garcia *et al.*, 1998). The shelf contains both emergent and submerged reefs along with extensive seagrass beds, coastal fringing mangroves and a series of mangrove islands. A number of the emergent reefs also shelter the inner parts of the shelf.

The reefs around La Parguera display a variety of reef forms from the typical spur and groove formations to extensive patch reefs, fringing reefs, and emergent island reefs or keys (Figure 2). In the La Parguera area, coral growth has been primarily responsible for the formation of these reefs (Garcia-Sais *et al.*, 2004). Many of the emergent reefs also contain stands of mangroves.

Typical corals include boulder coral (*Montastrea annularis*), brain corals (*Diploria spp.*), mustard hill coral (*Porites astreoides*), branching corals such as *Porites porites*, fire corals (*Millepora spp.*) and elkhorn and staghorn corals (*Acropora palmata* and *A. cervicornus*, respectively). In 2006, both acroporid species were listed as threatened under the Endangered Species Act. There are also extensive areas of soft sediment around the patch reefs (Morelock *et al.*, 1994).

Sampling Design

A reconnaissance mission (May 2005) was used

to characterize sediments in the study area, and qualitatively assess land use in the watershed. The primary sampling mission for the project took place in August 2005. During the primary mission, samples of sediment, water, and coral tissue were collected for the analysis of chemical contaminants.

The major goal for this phase of the project was to characterize sediment contaminants in the study area. To do this, a stratified random sampling design was used to select points where sediments would be collected for chemical contaminant analysis.

The location of sediments that could be sampled in the study area was made possible by the extensive benthic habitat mapping completed by CCMA's Biogeography Branch (Kendall *et al.*, 2001).

Using GIS (ArcGIS[®]), the hardbottom (coral) data layer was removed from the NOAA Biogeography benthic habitat map, leaving the soft bottom sediment data layer. The area to be sampled was then stratified by benthic

zone which included mangrove, lagoon, backreef and bankshelf areas (Figure 3). The study area was also stratified by longshore position. The sampling points were randomly selected using ArcGIS[®]. The number of samples per stratum was area weighted, however each habitat/longshore stratum contained at least three randomized sampling points.

Sediments collected during the reconnaissance mission in May 2005 were analyzed for total organic carbon (TOC) and grain size. These two parameters are important in determining where contaminants are more likely to accumulate if they are being introduced into a system, and were used to guide the selection of sampling sites for the August 2005 mission. Contaminants, particularly organics, tend to accumulate in sediments that have a higher TOC content. Grain size is also an important factor, as smaller grain size sediments have more surface area



Pastureland in Lajas.

available for the adsorption of contaminants and are found in depositional habitats.

The sediment TOC data were analyzed for spatial structure by quantitative/estimation autocorrelation using Moran's "I" and Geary's "C" tests. A positive test for spatial autocorrelation was found for TOC. Subsequently, indicator Kriging was used to interpolate the sediment TOC data to create a generalized map of high and low TOC (Figure 4). The contours in pink represent those areas modeled as having a high probability of elevated organic carbon, while areas in blue represent sediments with lower TOC. The map was then used to optimize the sampling for the August mission.



Sampling for sediment with the Van Veen grab.

strata, using standard NS&T Program protocols (Lauenstein and Cantillo, 1998). The sample sites are shown in Figure 5. The red half-circles indicate the higher organic carbon sites; blue half circles are the sites identified by Kriging as having lower organic carbon.

Samples were collected aboard the vessel *Aquanauta* using a GPS programmed with the station coordinates. A modified Van Veen grab (see inset) was deployed to collect the sediment samples using a pulley and davit, and retrieved by hand. Rocks or bits of seagrass were removed. If a particular grab did not result in 200-300 g of sediment, a second grab was made and composited with material

from the first grab. If enough sediment had not been collected after three deployments of the grab, the site was abandoned and the boat moved on to the next location.

Sampling Protocols. Sediments for contaminant analysis were collected in August 2005 from 43 sites, randomly selected within the high and low TOC

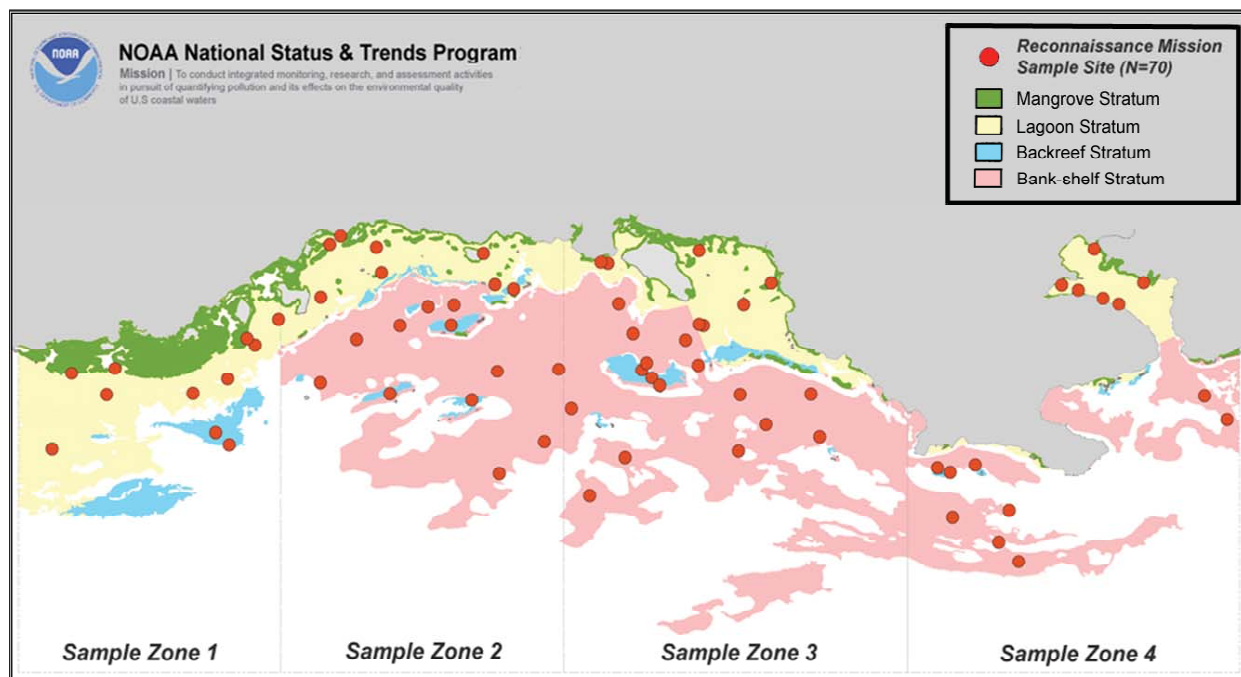


Figure 3. Stratified random sampling design used in May 2005 reconnaissance in southwest Puerto Rico.

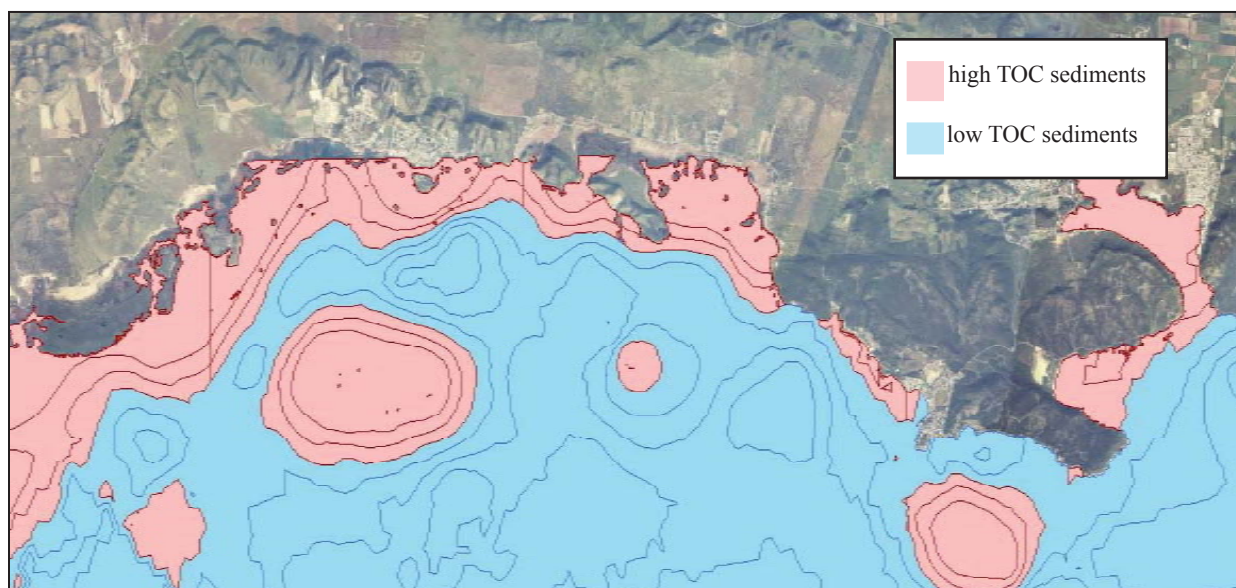


Figure 4. Modeled total organic carbon levels in the sediments.

Because the sediments collected were to be analyzed for chemical contaminants, protocols were used to avoid contamination of samples by equipment and cross contamination between samples. All equipment was rinsed with acetone and then distilled water just prior to use, to reduce the possibility of contaminating the sediment sample. Personnel handling the samples also wore disposable nitrile gloves. The top 3 cm of sediment were collected from the sediment grab using a Kynar-coated sediment scoop.

Sediments were placed into two certified clean (I-Chem[®]) 250 ml labeled jars, one for organic chemical analysis the other for metal analysis, capped and then placed on ice in a cooler. Sediments for grain size analysis were placed in a WhirlPack[®] bag, sealed and placed on ice in a cooler. At the end of each day, sediment samples for contaminant analysis were placed in a freezer; the WhirlPack[®] bags for the grain size analysis were placed in a refrigerator rather than frozen, to avoid altering the grain size structure of the sediment.

A series of water quality parameters (dissolved oxygen, temperature, salinity, and conductivity) were measured at each site using a YSI[®] salinity/conductivity/temperature meter. The instrument probe

was submerged to a depth of approximately one meter. Secchi depth was also measured at each site. Water samples were analyzed from 47 sites (Figure 5) for total nitrogen and phosphorus, and were collected by submerging a labeled 125 ml HDPE (high density polyethylene) bottle beneath the water surface, capping it and then placing it on ice. Nitrile disposable gloves were worn to prevent contamination. Bottles were rinsed three times with sample water prior to sample collection.

Water samples were also collected from seven sites for the analysis of Irgarol 1051, an antifoulant compound currently used in paint formulations for boat hulls. Water samples were collected in precleaned 500 ml amber glass bottles at a depth of 30-40 cm by hand, the bottle was then capped and placed on ice.

Finally, coral tissues (*Porites astreoides*) were collected at selected sites. The green half-circles in Figure 5 identify those sites where *P. astreoides* and soft sediments were co-located. The protocols used and results of the coral tissue analyses will be included in a later publication.

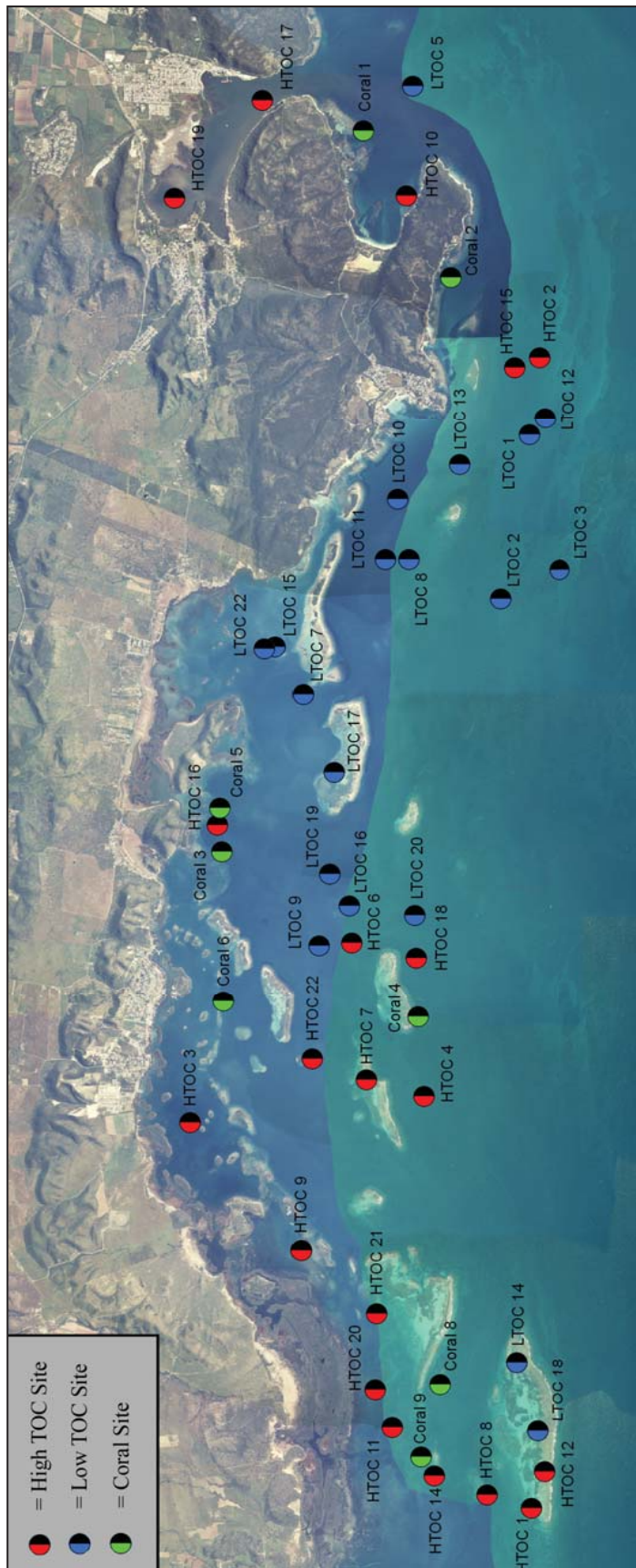


Figure 5. Sites targeted in southwest Puerto Rico during the August 2005 mission. TOC, total organic carbon.

Compounds Analyzed

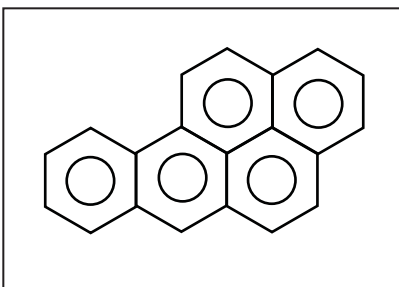
NOAA's NS&T Program monitors the Nation's estuarine and coastal waters for contaminants in bivalve mollusk tissues and sediments. The Program regularly quantifies over 120 organic and inorganic contaminants. The list of chemical contaminants analyzed in the sediments for this project is shown in Table 1. The compounds include 59 polycyclic aromatic hydrocarbons (PAHs), 30 organochlorine pesticides, 38 polychlorinated biphenyls (PCBs), four butyltins, 38 polybrominated diphenyl ether (PBDE) flame retardants, and 16 major and trace elements. All samples were analyzed using NS&T analytical protocols. Detailed descriptions of the protocols used for the analysis of organic contaminants can be found in Kimbrough *et al.* (2006); for inorganic analyses, Kimbrough and Lauenstein (2006). Each of the contaminant classes are discussed below.

PAHs. Polycyclic aromatic hydrocarbons are associated with the use and combustion of fossil fuels (*e.g.*, oil and gas) and other organic materials (*e.g.*, wood). Natural sources of PAHs include forest fires and volcanoes. The PAHs analyzed are two to six ring aromatic compounds (see inset). A number of PAHs can bioaccumulate in aquatic and terrestrial organisms, are toxic and some, including benzo[a]pyrene, benz[a]anthracene, chrysene, benzo[b]fluoranthene, benzo[k]fluoranthene, dibenzo[a,h]anthracene, and indeno[1,2,3-c,d]pyrene, are likely carcinogens (USDHHS, 1995). PAHs were analyzed using gas chromatography/mass spectrometry in the selected ion monitoring mode.

PCBs. Polychlorinated biphenyls are a class of synthetic compounds that have been used in numerous applications ranging from electrical transformers and capacitors, to hydraulic and heat transfer fluids, to pesticides and paints. Although no longer manufactured in the U.S., PCBs are now contaminants in the environment and in some cases are still in use in old equipment. PCBs have a biphenyl ring structure (two benzene rings with

a carbon to carbon bond) and a varying number of chlorine atoms. There are 209 PCB congeners possible. PCBs can bioaccumulate, and degradation in the environment proceeds slowly. Exposure to PCBs in fish has been linked to reduced growth, reproductive impairment and vertebral abnormalities (USEPA, 1997). PCBs were analyzed using gas chromatography/electron capture detection.

Butyltins. This class of compounds has a range of uses from biocides to catalysts to glass coatings. Tributyltin (TBT) in particular, used in antifoulant paint formulations on boat hulls, has been associated with endocrine disruption, specifically an imposex condition in marine gastropod molluscs. Beginning in 1989 in the U.S., the use of TBT as an antifouling agent was banned on vessels smaller than 25 meters in length (Gibbs and Bryan, 1996). Butyltins were analyzed using gas chromatography/flame photometric detection.



Benzo[a]pyrene, one of the PAHs analyzed in the sediments from Puerto Rico.

PBDEs. Polybrominated diphenyl ethers are currently a widely used class of flame retardants whose concentrations are rising in the environment as well as in human tissues. PBDEs are analogous to PCBs in structure, having 209 congeners, but containing bromine atoms (rather than chlorines) bonded to the aromatic rings. They are used in many products, most notably in

high impact polystyrene (plastics), foams, carpets and upholstery, textiles, office furniture, electronic equipment, and electronic circuits (McDonald, 2002). Worldwide production of PBDEs is estimated at about 67,000 metric tons per year, with the deca-BDE congener constituting the largest fraction. The penta-BDE is still used in large quantities in the United States but its use is now severely curtailed in Europe (Raloff, 2003).

Most PBDEs released into the environment tend to persist in sediment and soils. The fully brominated deca-BDE is not as readily bioaccumulated as the other PBDE congeners, however, it can degrade to penta-BDE through metabolic or environmental processes. The penta-BDE is rapidly and almost

Table 1. Chemical contaminants analyzed in the sediments from southwest Puerto Rico.

PAHs - Low MW Weight	PAHs - High MW Weight	Organochlorine Pesticides	PCBs	Major and Trace Elements	PBDEs
Naphthalene	Fluoranthene	Aldrin	PCB8/5	Aluminum (Al)	BDE 1 [2-MonoBDE]
1-Methylnaphthalene	Pyrene	Dieldrin	PCB18	Antimony (Sb)	BDE 2 [3-MonoBDE]
2-Methylnaphthalene	C1-Fluoranthenes/Pyrenes	Endrin	PCB28	Arsenic (As)	BDE 3 [4-MonoBDE]
2,6-Dimethylnaphthalene	C2-Fluoranthenes/Pyrenes	Heptachlor	PCB29	Cadmium (Cd)	BDE 10 [2,6-DiBDE]
1,6,7-Trimethylnaphthalene	C3-Fluoranthenes/Pyrenes	Heptachlor-Epoxide	PCB31	Chromium (Cr)	BDE 7 [2,4-DiBDE]
C1-Naphthalenes	Naphthobenzothiophene	Oxychlorane	PCB44	Copper (Cu)	BDE 8 [2,4'-DiBDE]
C2-Naphthalenes	C1-Naphthobenzothiophenes	Alpha-Chlordane	PCB45	Iron (Fe)	BDE 11 [3,3'-DiBDE]
C3-Naphthalenes	C2-Naphthobenzothiophenes	Gamma-Chlordane	PCB49	Lead (Pb)	BDE 12 [3,4-DiBDE]
C4-Naphthalenes	C3-Naphthobenzothiophenes	Trans-Nonachlor	PCB52	Manganese (Mn)	BDE 13 [3,4'-DiBDE]
Benzothiophene	Benz[a]anthracene	Cis-Nonachlor	PCB56/60	Mercury (Hg)	BDE 15 [4,4'-DiBDE]
C1-Benzothiophenes	Chrysene	Alpha-HCH	PCB66	Nickel (Ni)	BDE 30 [2,4,6-TriBDE]
C2-Benzothiophenes	C1-Chrysenes	Beta-HCH	PCB70	Selenium (Se)	BDE 32 [2,4',6'-TriBDE]
C3-Benzothiophenes	C2-Chrysenes	Delta-HCH	PCB74/61	Silicon (Si)	BDE 17 [2,2',4'-TriBDE]
Biphenyl	C3-Chrysenes	Gamma-HCH	PCB87/115	Silver (Ag)	BDE 25 [2,3',4'-TriBDE]
Acenaphthylene	C4-Chrysenes	2,4'-DDD	PCB95	Tin (Sn)	BDE 33 [2,3',4'-TriBDE]
Acenaphthene	Benzofluoranthene	4,4'-DDD	PCB99	Zinc (Zn)	BDE 28 [2,4,4'-TriBDE]
Dibenzofuran	Benzokjfluoranthene	2,4'-DDE	PCB101/90		BDE 35 [3,3',4'-TriBDE]
Fluorene	Benzofluorene	4,4'-DDE	PCB105		BDE 37 [3,4',4'-TriBDE]
C1-Fluorenes	Benzofluorene	2,4'-DDT	PCB110/77		BDE 75 [2,4,4',6'-TetraBDE]
C2-Fluorenes	Perylene	4,4'-DDT	PCB118		BDE 49/71 [2,2',4,5'-TetraBDE/ 2,3',4',6'-TetraBDE]
C3-Fluorenes	Indeno[1,2,3-c,d]pyrene	1,2,3,4-Tetrachlorobenzene	PCB128		BDE 47 [2,2',4,4'-TetraBDE]
Carbazole	Dibenzo[a,h]anthracene	1,2,4,5-Tetrachlorobenzene	PCB138/160		BDE 66 [2,2',3,4,4'-TetraBDE]
Anthracene	C1-Dibenzo[a,h]anthracenes	Hexachlorobenzene	PCB146		BDE 77 [3,3',4,4'-TetraBDE]
Phenanthrene	C2-Dibenzo[a,h]anthracenes	Pentachloroisole	PCB149/123		BDE 100 [2,2',4,4',6-PentaBDE]
1-Methylphenanthrene	C3-Dibenzo[a,h]anthracenes	Endosulfan II	PCB151		BDE 119 [2,2',4,4',5-PentaBDE]
C1-Phenanthrene/Anthracenes	Benzo[g,h,i]perylene	Endosulfan I	PCB153/132		BDE 99 [2,2',4,4',5-PentaBDE]
C2-Phenanthrene/Anthracenes	Butyltins	Endosulfan Sulfate	PCB156/171/202		BDE 116 [2,3,4,5,6-PentaBDE]
C3-Phenanthrene/Anthracenes	Monobutyltin	Mirex	PCB158		BDE 118 [2,3',4,4',5-PentaBDE]
C4-Phenanthrene/Anthracenes	Dibutyltin	Chlorpyrifos	PCB170/190		BDE 85 [2,2',3,4,4'-PentaBDE]
Dibenzothiophene	Tributyltin		PCB174		BDE 126 [3,3',4,4',5-PentaBDE]
C1-Dibenzothiophenes	Tetrabutyltin		PCB180		BDE 155 [2,2',4,4',6,6-HexaBDE]
C2-Dibenzothiophenes			PCB183		BDE 154 [2,2',4,4',5,6-HexaBDE]
C3-Dibenzothiophenes			PCB187		BDE 153 [2,2',4,4',5,5'-HexaBDE]
			PCB194		BDE 138 [2,2',3,4,4',5'-HexaBDE]
			PCB195/208		BDE 166 [2,3,4,4',5,6-HexaBDE]
			PCB201/157/173		BDE 183 [2,2',3,4,4',5',6-HeptaBDE]
			PCB206		BDE 181 [2,2',3,4,4',5,6-HeptaBDE]
			PCB209		BDE 190 [2,3,3',4,4',5,6-HeptaBDE]

Abbreviations: MW, molecular weight; PAH, polycyclic aromatic hydrocarbons; HCH, hexachlorocyclohexane; DDT, DDD, dichlorodiphenyldichloroethane; DDE, dichlorodiphenyldichloroethylene; PCB, polychlorinated biphenyl; BDE, brominated diphenyl ether

completely absorbed, not eliminated from the body quickly, and tends to bioaccumulate. Recently, there has been concern regarding PBDEs because of rising concentrations in the environment and in human tissues. PBDEs have been shown to affect thyroid hormone function, and have been associated with impaired liver function and neural development (McDonald, 2002). PBDEs were analyzed using gas chromatography/mass spectrometry in the selected ion monitoring mode.

Major and Trace Elements. A total of 16 major and trace elements were measured in the sediments for this project (Table 1). All occur naturally to some extent in the environment. Aluminum, iron, and silicon are major elements in the Earth's crust. As their name implies, trace elements occur at lower concentrations in the crustal material, however mining and manufacturing processes along with the use and disposal of products containing trace elements result in elevated concentrations in the environment. A number of trace elements are toxic at low concentrations. Cadmium, used in metal plating and solders, has been shown to impair development and reproduction in several invertebrate species, and ability to osmoregulate in herring larvae (USDHHS, 1999; Eisler, 1985). Mercury is volatile and can enter the atmosphere through processes including mining, manufacturing, combustion of coal and volcanic eruptions. Effects of mercury on copepods include reduced growth and reproductive rates (Eisler, 1987). Chromium has been shown to reduce survival and fecundity in the cladoceran *Daphnia magna*, and reduced growth in fingerling chinook salmon (*Oncorhynchus tshawytscha*) (Eisler, 1986). Copper has a number of uses such as in antifouling paints, wood preservatives, heat exchangers in power plants, electrical wires, coinage, and in agricultural fungicides. Although an essential element, elevated levels of copper can impact aquatic organisms, including reproduction and development in mysid shrimp (Eisler, 1998). Major and trace elements were analyzed using atomic spectroscopy techniques.

Clostridium perfringens. Although not a chemical contaminant, the bacterium *C. perfringens* has been used as an indicator of fecal pollution and was analyzed in the sediment samples from southwest

Puerto Rico. This bacterium occurs in the intestines of humans and in some domestic and feral animals, and ingestion can cause food poisoning.

Nitrogen and Phosphorus. Nitrogen and phosphorus are nutrients essential for primary production in plants. Excessive loading of nutrients, from sources such as land application of fertilizers and wastewater treatment plant effluent, can lead to eutrophication characterized by an overabundance of algae and reduced concentrations of dissolved oxygen in the water column. Water samples were analyzed for total nitrogen and total phosphorus using an autoanalyzer following persulfate oxidation.

Irgarol 1051. This triazine herbicide is currently used in copper-based antifoulant paints on boats. While Irgarol 1051 has been detected in both salt and freshwater environments (Berard *et al.*, 2003), impacts that might occur in corals and seagrass communities are currently under investigation. Water samples were analyzed for Irgarol 1051 using high performance liquid chromatography (HPLC)/mass spectrometry.

Statistical and Spatial Analyses

All contaminant data were analyzed using JMP[®] statistical software. A Shapiro-Wilk test was first run on individual parameters to see if the data were normally distributed. When data were normally distributed, regression analyses and Pearson Product moment correlations were run to explore relationships between parameters. If the data were not normally distributed and transformations were not effective, Spearman's nonparametric multivariate correlation was used.

Differences in contaminant classes among sample strata (habitat type) were also investigated. ANOVAs were run when tests (O'Brien's test) revealed adequate homogeneity of variance on either the raw or transformed data. Pair-wise comparison tests (Student's T or Tukey's HSD) were then run to assess differences among the means. A Wilcoxon nonparametric test was run on data that did not meet the required homogeneity of variance condition.

Data also were analyzed using the Spatial Analyst extension in ArcGIS®. The data (e.g., PAHs in sediments) were first tested for spatial autocorrelation. This correlation is used to measure the similarity of objects within an area, the degree to which a spatial phenomenon is correlated with itself, the level of interdependence between the variables, and the nature and strength of that interdependence (Childs, 2004). The significance of the spatial autocorrelation was tested using Moran's I and Geary's C test. Indicator Kriging was then used to model the concentrations of spatially autocorrelated contaminant data. In addition, correlations (e.g., Spearman's Rho) between contaminant class and coral community structure were evaluated to determine the degree to which coral richness and contamination correspond in the region.

Effects of Contaminants on Corals

A number of authors have found that pollution can significantly impact coral reefs. Pandolfi *et al.* (2003) noted that pollution and overfishing have resulted in massive declines in abundance, diversity and habitat structure in coral reefs and associated tropical nearshore ecosystems. Edinger *et al.* (1998) found that reefs exposed to land-based pollution in Indonesia showed a 30-50 percent reduction in coral diversity at a depth of 3 meters, and a 40-60 percent reduction in coral diversity at a depth of 10 meters.

This section of the report provides an overview of some of the documented effects of chemical contaminants on corals. More detailed discussions on the effects of individual pollutants or pollutant classes are included following each contaminant section in the Results and Discussion.

Understanding the effects that chemical contaminants can have on corals is important as it provides a basis for describing mechanistic linkages that may exist

between contaminants and coral condition, from the individual coral to the community. Determining the effects of individual contaminants on adult corals is a relatively new field of interest. One likely reason for this is the difficulty in maintaining and culturing corals in the laboratory.

The majority of the literature found on the effects of chemical contaminants on corals involves aqueous exposures (e.g., crude oil, trace elements and pesticides) to coral larvae or the direct application of contaminants (e.g., crude oil) to corals. Although most of the chemical contaminant data presented in this report are for sediments, the presence of

chemical contaminants in the sediments provides an indication of what the corals have been exposed to over time, as sediments act to accumulate many contaminant types, particularly those that are less water soluble. Most of these less water soluble or hydrophobic compounds were in the water column at some point, either in the dissolved phase or bound to particulates before partitioning out onto the sediments or into aquatic organisms.



An example of boulder star coral, Montastrea annularis.

Table 2 summarizes some of the documented effects of individual contaminants and crude oil on corals. From this table, it can be seen that exposure to a number of contaminants can affect coral fertilization, settlement and metamorphosis, primary production and formation of the coral skeleton. Much of the research has shown that contaminant concentrations as low as $\mu\text{g/L}$ (part per billion, or ppb) can have significant and in some cases serious effects on corals. Only one article involving the effects of contaminated sediments on corals was found.

Petroleum Hydrocarbons

Few studies have described the effects of individual PAHs, such as anthracene, phenanthrene, pyrene and benzo[a]pyrene (Peachey and Crosby, 1996; Knap *et al.*, 1982; Gassman and Kennedy, 1992;

Table 2. Effects of chemical contaminants on corals.

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Fungia scutaria</i> (mushroom coral)	Pyrene	PAH	Larval motility	32 ug/L	No toxicity to larvae exposed to pyrene (1-48 ug/L) in the dark; immobilization of larvae observed when exposed to pyrene in the presence of artificial sunlight (phototoxicity)	2 hours	Peachey and Crosby (1996)
<i>Montastrea annularis</i> (lobed star coral)	Benzo[a]pyrene	PAH	Uptake and depuration	5 ug/L	Rapid uptake of radiolabeled benzo[a]pyrene in both coral tissue and zooxanthellae; low rate of depuration and metabolism	25 hours	Kennedy <i>et al.</i> (1992)
<i>Favia fragum</i> (golfball coral)	Benzo[a]pyrene	PAH	Uptake and depuration	5 ug/L	Rapid uptake of radiolabeled benzo[a]pyrene in both the coral tissue and zooxanthellae; low rate of depuration and metabolism	25 hours	Kennedy <i>et al.</i> (1992)
<i>Diploria strigosa</i> (symmetrical brain coral)	Phenanthrene	PAH	Uptake and depuration	33 ug/L	Rapid uptake of radiolabeled phenanthrene in the coral; rapid depuration during first two days, followed by much slower rate between day two and day 10	24 hours	Knap <i>et al.</i> (1982)
<i>Diploria strigosa</i> (symmetrical brain coral)	Phenanthrene	PAH	Uptake and depuration	3 ug/L	Rapid uptake of radiolabeled phenanthrene in the coral; rapid depuration during first ten days, followed by much slower release; radioactivity still detectable after one year	24 hours	Solbakken <i>et al.</i> (1984)
<i>Montastrea annularis</i> (lobed star coral)	Phenanthrene	PAH	Uptake and depuration	3 ug/L	Rapid uptake of radiolabeled phenanthrene in the coral; roughly 10 percent of radioactivity still detectable after one month of depuration	24 hours	Solbakken <i>et al.</i> (1984)
<i>Diploria strigosa</i> (symmetrical brain coral)	Naphthalene	PAH	Uptake and depuration	16 ug/L	Rapid uptake of radiolabeled naphthalene in the coral; rapid depuration compared to phenanthrene; after 21 days less than one percent of original radioactivity present	24 hours	Solbakken <i>et al.</i> (1984)
<i>Madracis decactis</i> (green cactus coral)	Naphthalene	PAH	Uptake and depuration	16 ug/L	Rapid uptake of radiolabeled naphthalene in the coral; rapid depuration compared to phenanthrene; after 21 days less than two percent of original radioactivity present	24 hours	Solbakken <i>et al.</i> (1984)
<i>Oculina diffusa</i> (ivory bush coral)	Naphthalene	PAH	Uptake and depuration	42 ug/L	Rapid uptake of radiolabeled naphthalene in the coral; half life of the accumulated naphthalene was 24 hours; clearance of accumulated naphthalene estimated at 14 days	7 hours	Neff and Anderson (1981)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Diploria strigosa</i> (symmetrical brain coral)	2,4,5,2',4',5'-hexachlorobiphenyl	PCB	Uptake and depuration	15 ug/L	Rapid uptake of radiolabeled PCB in the coral; depuration very slow, after one year nearly a third of the original radioactivity was present	24 hours	Solbakken <i>et al.</i> (1984)
<i>Madracis decactis</i> (green cactus coral)	2,4,5,2',4',5'-hexachlorobiphenyl	PCB	Uptake and depuration	15 ug/L	Rapid uptake of radiolabeled PCB in the coral; depuration very slow, after one year nearly a third of the original radioactivity was present	24 hours	Solbakken <i>et al.</i> (1984)
<i>Acropora millepora</i> (no common name)	Water accommodated fraction	Crude oil	Fertilization success	165 ug/L	No effect of water accommodated fraction (WAF) of crude oil up to 165 ug/L	4 hours	Negri and Heyward (2000)
<i>Acropora millepora</i> (no common name)	Water accommodated fraction	Crude oil	Larval settlement	82 ug/L	Larval settlement significantly affected at 82 ug/L; no settlement at 165 ug/L	4 hours	Negri and Heyward (2000)
<i>Sylophora pistillata</i> (club foot coral)	Water soluble fraction	Crude oil	Larval settlement	0.1%	No settlement of larvae at a water soluble fraction concentration of the crude oil of 0.1%	12 hours	Epstein <i>et al.</i> (2000)
<i>Diploria strigosa</i> (symmetrical brain coral)	Water accommodated fraction	Crude oil	Coral growth	10,000 ug/L	Corals exposed to light crude oil for 24 hours and then moved back onto the reefs; no effect on growth of corals over the next year	24 hours	Dodge <i>et al.</i> (1984)
<i>Acropora formosa</i> (stony coral)	Water accommodated fraction	Crude oil	Coral survival	5,000 ug/L	Corals exposed to marine fuel oil for 12 and 24 hours expelled zooxanthellae and then died	12 hours	Harrison <i>et al.</i> (1990)
<i>Manicina areolata</i> (rose coral)	Water accommodated fraction	No. 2 fuel oil	Histologic effects	70 ug/L	Degraded reproductive tissues, degradation or loss of zooxanthellae and atrophy of mucous secretory cells in corals	12 weeks	Peters <i>et al.</i> (1981)
<i>Acropora surculosa</i> (no common name)	Copper	Trace element	Fertilization success	10 ug/L	Significantly reduced fertilization success at or above this concentration; at 200 ug/L less than 10 percent fertilization success	5 hours	Victor and Richmond (2005)
<i>Lobophytum compactum</i> (soft coral)	Copper	Trace element	Fertilization success	117 ug/L	Significantly reduced fertilization success at or above this concentration; EC ₅₀ (calculated concentration that decreased fertilization by 50%) was 261 ug/L	5 hours	Reichel-Brushett and Michalek-Wagner (2005)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Goniastrea aspera</i> (brain coral)	Copper	Trace element	Fertilization success	20 ug/L	Significantly reduced fertilization at or above this concentration; at 75 ppb less than 1% fertilization success; EC ₅₀ of 19 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Goniastrea retiformis</i> (no common name)	Copper	Trace element	Fertilization success	20 ug/L	Significantly reduced fertilization at or above this concentration; EC ₅₀ of 25 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora tenuis</i> (finger coral)	Copper	Trace element	Fertilization success	42 ug/L	Significantly reduced fertilization rate at or above this concentration; at 67 ug/L, less than 5% fertilization success occurred; EC ₅₀ of 40 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora longicyathus</i> (no common name)	Copper	Trace element	Fertilization success	24 ug/L	Significantly reduced fertilization rate at or above this concentration; at 61 ug/L, less than 3% fertilization success occurred	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora surculosa</i> (no common name)	Copper	Trace element	Development	12 ug/L	Embryo development significantly reduced at or above this concentration; at concentrations greater than 58 ug/L, no embryo development	12 hours	Victor and Richmond (2005)
<i>Acropora millepora</i> (no common name)	Copper	Trace element	Fertilization success	17 ug/L	Fertilization success EC ₅₀	4 hours	Negri and Heyward (2001)
<i>Acropora millepora</i> (no common name)	Copper	Trace element	Larval settlement	110 ug/L	Larval settlement EC ₅₀ in the presence of crustos algae	24 hours	Negri and Heyward (2001)
<i>Goniastrea aspera</i> (brain coral)	Copper	Trace element	Larval motility	21 ug/L	Larval motility EC ₅₀ ; a no observable effect concentration (NOEC) of 20 ug/L was determined	12 hours	Reichelt-Brushett and Harrison (2004)
<i>Montipora verrucosa</i> (rice coral)	Copper	Trace element	Zooxanthellae growth	40 ug/L	Significantly decreased growth rate in <i>Symbiodinium microadriaticum</i> , zooxanthellae cultured from the coral <i>Montipora verrucosa</i>	28 days	Goh and Chou (1997)
<i>Acropora tenuis</i> (finger coral)	Copper	Trace element	Larval settlement	42 ug/L	Larval settlement EC ₅₀ of 35 ug/L; at 200 ug/L, all larvae died	48 hours	Reichelt-Brushett and Harrison (2000)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Porites cylindrica</i> (yellow finger coral)	Copper	Trace element	Primary production	11 ug/L	No effect on primary production when coral exposed only to copper; reduced primary production when corals exposed to increased temperature and copper	24 hours	Nystrom <i>et al.</i> (2001)
<i>Acropora microphthalma</i> (no common name)	Tributyltin, Copper and Zinc	Antifoulant ingredients and trace elements	Larval settlement	8 ug/g TBT, 72 ug/g Cu, 92 ug/g Zn in diluted sediments	No larval settlement on preconditioned terracotta tiles; exposure of larvae to 5% contaminated field sediments containing tributyltin (TBT), copper and zinc; most larvae died before metamorphosis at this concentration	48 hours	Negri <i>et al.</i> (2002)
<i>Goniastrea aspera</i> (brain coral)	Nickel	Trace element	Fertilization success	100 ug/L	Significantly reduced fertilization rate at or above this concentration.	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Pocillopora damicornis</i> (lace coral)	Nickel	Trace element	Larval settlement	1,000 ug/L	Significantly reduced larval settlement at 1 ppm nickel after 9 days of recovery; effects seen for all durations of exposure (12-96 hours)	12 hours	Goh (1991)
<i>Goniastrea aspera</i> (brain coral)	Zinc	Trace element	Fertilization success	500 ug/L	No significant difference in fertilization success compared to controls for any concentration tested, up to 500 ug/L.	5 hours	Reichelt-Brushett and Harrison (1999)
<i>Acropora tenuis</i> (finger coral)	Zinc	Trace element	Fertilization success	10 ug/L	Significantly reduced fertilization at or above this concentration; NOEC <10 ug/L; at 5,000 ug/L, no fertilization occurred at all	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Montipora verrucosa</i> (rice coral)	Zinc	Trace element	Zooxanthellae growth	509 ug/L	No effect on growth rate of <i>Symbiodinium microadriaticum</i> , zooxanthellae cultured from the coral <i>Montipora verrucosa</i>	28 days	Goh and Chou (1997)
<i>Goniastrea aspera</i> (brain coral)	Cadmium	Trace element	Fertilization success	200 ug/L	No significant difference in fertilization success compared to controls for any concentration tested, up to 200 ug/L	5 hours	Reichelt-Brushett and Harrison (1999)
<i>Oxyypora lacera</i> (scroll coral)	Cadmium	Trace element	Fertilization success	1,000 ug/L	No significant difference in fertilization success compared to controls for any concentration tested, up to 1,000 ug/L	5 hours	Reichelt-Brushett and Harrison (1999)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Acropora tenuis</i> (finger coral)	Cadmium	Trace element	Fertilization success	5,000 ug/L	Significantly reduced fertilization rate at or above this concentration; NOEC of 2,000 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora longicyathus</i> (no common name)	Lead	Trace element	Fertilization success	855 ug/L	Significantly reduced fertilization at or above this concentration; EC ₅₀ of 1,453 ug/L; NOEC of 451 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora tenuis</i> (finger coral)	Lead	Trace element	Fertilization success	1,982 ug/L	Significantly reduced fertilization at or above this concentration; EC ₅₀ of 1,801 ug/L; NOEC of 7901 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Goniastrea aspera</i> (brain coral)	Lead	Trace element	Fertilization success	6,409 ug/L	Significantly reduced fertilization at or above this concentration; NOEC of 5,455 ug/L	5 hours	Reichelt-Brushett and Harrison (2005)
<i>Acropora millepora</i> (no common name)	Tributyltin (TBT)	Antifoulant ingredient	Fertilization success	200 ug/L	Fertilization success EC ₅₀	4 hours	Negri and Heyward (2001)
<i>Acropora millepora</i> (no common name)	Tributyltin (TBT)	Antifoulant ingredient	Larval settlement	2 ug/L	Larval settlement EC ₅₀ in the presence of crustos algae	24 hours	Negri and Heyward (2001)
<i>Madracis mirabilis</i> (yellow pencil coral)	Irgarol 1051	Antifoulant ingredient	Photosynthesis in intact coral	0.1 ug/L	Reduction in net photosynthesis (O ₂ evolution); at TBT concentrations exceeding 1 ug/L, little or no photosynthesis	8 hours	Owen <i>et al.</i> (2002)
<i>Seriatopora hystrix</i> (bird's nest coral)	Irgarol 1051	Antifoulant ingredient	Chlorophyll fluorescence	0.7 ug/L	Concentration resulting in 50% decrease (EC ₅₀) in chlorophyll fluorescence (maximum effect quantum yield)	10 hours	Jones and Kerswell (2003)
<i>Madracis mirabilis</i> (yellow pencil coral)	Irgarol 1051	Antifoulant ingredient	Photosynthesis in zooxanthellae	0.063 ug/L	Inhibition of photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	8 hours	Owen <i>et al.</i> (2002)
<i>Diploria strigosa</i> (symmetrical brain coral)	Irgarol 1051	Antifoulant ingredient	Photosynthesis in zooxanthellae	2 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Favia fragum</i> (golfball coral)	Irgarol 1051	Antifoulant ingredient	Photosynthesis in zooxanthellae	2 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)
<i>Madracis mirabilis</i> (yellow pencil coral)	Diuron	Antifoulant ingredient	Photosynthesis in intact coral	2 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2002)
<i>Diploria strigosa</i> (symmetrical brain coral)	Diuron	Antifoulant ingredient	Photosynthesis in zooxanthellae	2 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)
<i>Favia fragum</i> (golfball coral)	Diuron	Antifoulant ingredient	Photosynthesis in zooxanthellae	2 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)
<i>Acropora formosa</i> (stony coral)	Diuron	Herbicide	Chlorophyll fluorescence	2.3 ug/L	Concentration resulting in 50% decrease (EC ₅₀) in chlorophyll fluorescence (maximum effect quantum yield)	10 hours	Jones and Kerswell (2003)
<i>Acropora formosa</i> (stony coral)	Atrazine	Antifoulant ingredient	Chlorophyll fluorescence	45 ug/L	Concentration resulting in 50% decrease (EC ₅₀) in chlorophyll fluorescence (maximum effect quantum yield)	10 hours	Jones and Kerswell (2003)
<i>Madracis mirabilis</i> (yellow pencil coral)	Atrazine	Antifoulant ingredient	Photosynthesis in intact coral	100 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2002)
<i>Diploria strigosa</i> (symmetrical brain coral)	Atrazine	Antifoulant ingredient	Photosynthesis in zooxanthellae	100 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)
<i>Favia fragum</i> (golfball coral)	Atrazine	Antifoulant ingredient	Photosynthesis in zooxanthellae	100 ug/L	Reduction in net photosynthesis (¹⁴ C incorporation) in isolated zooxanthellae after 6 hours	6 hours	Owen <i>et al.</i> (2003)
<i>Acropora formosa</i> (stony coral)	Simazine	Antifoulant ingredient	Chlorophyll fluorescence	150 ug/L	Concentration resulting in 50% decrease (EC ₅₀) in chlorophyll fluorescence (maximum effect quantum yield)	10 hours	Jones and Kerswell (2003)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

Table 2. Effects of chemical contaminants on corals (cont.).

Coral/Species	Contaminant	Contaminant Class	Response	Concentration	Effect	Duration of Exposure	Reference
<i>Pocillopora damicornis</i> (lace coral)	Chlorpyrifos	Insecticide	Movement	100 ug/L	Reduced swimming motion in all larvae; at a concentration of 1,000 ug/L killed 50 to 100 percent of larvae within 24 hours	24 hours	Acevedo (1991)
<i>Acropora longicyathus</i> (no common name)	Ammonium (NH ₄)	Nutrient	Fertilization success	18 ug/L	Mean fertilization success reduced in gametes	5 hours	Harrison and Ward (2001)
<i>Acropora longicyathus</i> (no common name)	Phosphate (PO ₄)	Nutrient	Fertilization success	95 ug/L	Mean fertilization success reduced in gametes	5 hours	Harrison and Ward (2001)
<i>Porites porites</i> (thick finger coral)	Nitrate (NO ₃)	Nutrient	Photosynthesis and calcification	310 ug/L	Increased photosynthesis (↑ zooxanthellae), but decreased calcification	40 days	Marubini and Davis (1996)
<i>Montastrea annularis</i> (boulder coral)	Nitrate (NO ₃)	Nutrient	Photosynthesis and calcification	310 ug/L	Increased photosynthesis (↑ zooxanthellae), but decreased calcification	30 days	Marubini and Davis (1996)
<i>Acropora digitifera</i> (staghorn coral)	Suspended sediment	Sediment	Fertilization success	50 mg/L	Reduced fertilization, post embryonic development not affected	18 hours	Gilmour (1999)

Abbreviations: ug/L, micrograms/Liter or parts per billion (ppb); mg/L, milligrams/Liter

and Kennedy *et al.*, 1992) on corals. The available literature for petroleum hydrocarbon effects tend to focus on crude and refined oils rather than individual PAHs. Crude oil is a complex mixture containing aliphatic (saturated), aromatic (including PAHs) and other compound classes. A substantial amount of the research to assess the impacts of crude oil on corals has been from field studies, primarily as a result of oil spills or chronic discharges to reefs. Although this report focuses on the presence and effects of individual compounds, some of the studies of crude oil were included to supplement the available PAH data.

PAHs readily accumulate in the living tissues of corals and in the zooxanthellae, the symbiotic photosynthetic dinoflagellate algae found within coral tissues. The bioaccumulation appears to be related to the lipid content of both (Kennedy *et al.*, 1992). While the simple accumulation of PAHs by corals is not an impact by itself, the accumulation of a chemical contaminant in an organism increases the likelihood of adverse effects. Solbakken *et al.* (1984) showed that both phenanthrene and naphthalene are accumulated by the brain coral *Diploria strigosa* and green cactus coral *Madracis decatis*, and that the lower molecular weight naphthalene was eliminated at a higher rate than phenanthrene (Solbakken *et al.*, 1984). Peachey and Crosby (1996) have shown that the toxicity (expressed as a reduction in coral larval motility) of PAHs to corals was enhanced by exposure to light (phototoxicity). PAHs have also been shown to induce microsomal degradative enzymes (Cytochrome P450 class) in corals. In higher organisms, P450 enzymes can metabolize contaminants such as PAHs, often making them more water soluble, and enhancing their rate of excretion. Gassman and Kennedy (1992) were among the first researchers to show that at least some corals possess Cytochrome P450 enzymes.

Crude and refined oils can also impact coral health. Negri and Heyward (2000) found that concentrations of crude oil in water as low as 82 µg/L reduced coral larval settlement (Table 2). Peters *et al.* (1981) found that exposure to No. 2 fuel oil at concentrations as low as 70 µg/L in the rose coral (*Manicina areolata*) over a 12 week period resulted

in substantial histologic changes in both the somatic and reproductive tissues, along with the loss of the zooxanthellae.

Pesticides

A few studies have focused on the effects of pesticides on corals, but no studies were found on the effects of organochlorine pesticides such as DDT or chlordane. A number of studies have been conducted with Irgarol 1051 (Jones and Kerswell, 2003; Owen *et al.* 2002 and 2003). Data were also located for diuron, a substituted urea herbicide which is sometimes used in antifoulant paints. Both Irgarol 1051 and diuron have been shown to significantly affect corals, particularly the zooxanthellae. Concentrations of Irgarol 1051 as low as 0.063 µg/L (0.063 ppb or 63 ppt) were found to significantly inhibit photosynthesis in zooxanthellae (Owen *et al.*, 2002). Diuron was found to impact photosynthesis at a concentration of 2 µg/L (Owen *et al.*, 2003). Other herbicides, more commonly used in agriculture, such as atrazine and simazine were less effective in inhibiting photosynthesis (Table 2). At a concentration of 100 µg/L, the organophosphate insecticide chlorpyrifos significantly inhibited swimming motion in the larvae of the lace coral *Pocillopora damicornis* (Acevedo, 1991).

Trace Elements

A number of studies have assessed the effects of trace elements on corals, primarily on fertilization and settlement success. Many of these studies involved copper (*e.g.*, Reichelt-Brushett and Harrison, 1999; Reichelt-Brushett and Harrison, 2000; Negri and Heyward, 2001; Nystrom *et al.*, 2001; and Victor and Richmond, 2005). Copper is of environmental significance due in part to its use in boat hull antifoulant paints, and in agriculture and industry. Copper has been shown to impact coral fertilization success, settlement and metamorphosis in the µg/L range (Table 2). A number of studies have been conducted for other trace elements including nickel (Goh, 1991; and Reichelt-Brushett and Harrison, 2005), zinc (Goh and Chou, 1997; Reichelt-Brushett and Harrison, 1999), lead (Reichelt-Brushett and Harrison, 2004), and mercury (Bastidas and Garcia, 2004). Nickel and zinc can impact coral fertilization and settlement, but typically at higher concentrations.

Nutrients

In addition to promoting the growth of macroalgae which can outcompete and overgrow corals, nitrogen and phosphorus can impact corals directly. Harrison and Ward (2001) showed that phosphate at a concentration of 95 µg/L reduced fertilization success in the coral *Acropora longicyathus*. Marubini and Davis (1996) showed that nitrate at a concentration of 310 µg/L significantly reduced photosynthesis in two species, and perhaps even more of a concern, reduced the calcification rate in the coral skeleton.

Results and Discussion

Field Data

Data collected in the field during the August 2005 mission are presented in Table 3. A total of 50 sites were visited. At seven sites, the field team was unsuccessful in collecting sediments, resulting in sediment samples being collected at 43 sites. The average depth at the sites sampled was 10.3 m. The shallowest site was a backreef site (LTOC 14), with a depth of 1.2 m; the deepest site sampled was 21 m (HTOC 22). The average salinity encountered was 34.2 ppt, average temperature was 30.2 °C. Nonparametric analysis (Spearman's Rho test) indicated that salinity did not vary significantly ($P > 0.2824$) by habitat (lagoon, backreef, bankshelf and coral sites). Analysis of temperature data indicated differences among habitat types ($P < 0.0001$) with near shore areas in general being warmer than offshore areas.

Secchi depth also varied between sites. At a number of sites, including all of the backreef locations, the Secchi disk was visible to the bottom (Table 3). An ANOVA run on those sites where the Secchi depth was less than the bottom depth indicated significant differences ($P < 0.0001$) between habitat types (*e.g.*, lagoon, coral, and bankshelf). Results of a parametric Tukey-Kramer HSD test indicated not surprisingly, that visibility was better at the offshore (bankshelf) sites.

Total Organic Carbon and Grain Size

The average TOC in the sediments collected was 1.17%, and ranged from a low of 0.13% to a high of 4.99% (Appendix D). As noted earlier, a positive correlation between organic carbon in the sediments

and levels of contaminants is often found in coastal marine and estuarine sediments (Shine and Wallace, 2000). In addition, sediments with higher organic carbon levels (>3% organic carbon) have been associated with degraded benthic communities (USEPA, 2002).

Polycyclic Aromatic Hydrocarbons

Total PAHs as used in this report refers to the sum of the 59 compounds and compound classes (*e.g.*, anthracene, C1-naphthalenes) analyzed in the sediment samples. Appendix E contains detailed results of this analysis. A map showing total PAHs in the sediments is shown in Figure 6. Higher concentrations of PAHs were found in the sediments adjacent to the town of La Parguera and also in Guanica Bay. This spatial pattern, *i.e.*, higher concentrations of contaminants closer to shore and in Guanica Bay was seen for a number of chemical contaminant classes. A plot of total PAH concentrations is shown in Figure 7. The sampling sites in this figure are organized in a west to east direction, as are subsequent figures, to be comparable to the map. The pattern of elevated PAH concentrations around La Parguera and Guanica Bay is apparent.

Comparison with NOAA's NS&T Data.

Concentrations of total PAHs in the sediments at the majority of sites sampled were below 300 ng/g (ppb). Figure 7 also contains information related to the NS&T national median and 85th percentile. Because of the long-term, national-level contaminant monitoring carried out by the NS&T Program, data from southwest Puerto Rico can be compared with data from the rest of the Nation's coastal waters to see how the sites compare. Overall, the concentrations of total PAHs in the study area were low. Only two sites, HTOC 17 and HTOC 19, both in Guanica Bay, were above the national NS&T median of 380 ng/g for total PAHs. None of the sites sampled in southwest Puerto Rico approached the national NS&T 85th percentile of 2,688 ng/g.

EPA National Coastal Assessment Data. The U.S. Environmental Protection Agency's (EPA) National Coastal Assessment (NCA) Program provides information on the condition of the Nation's coastal resources (USEPA, 2004). In 2000, sediments were

Table 3. Field data from the August 2005 mission.

Site	Collection Date	Habitat	Latitude (DD)	Longitude (DD)	Water Depth (m)	Dissolved Oxygen (mg/L)	Temperature (°C)	Salinity (ppt)	Conductivity (mS)	Secchi Depth (m)
Coral 1	8/27/05	Coral	17.94219	-66.91481	9.1	4.63	30.0	33.9	56.8	3.7
Coral 2	8/27/05	Coral	17.93012	-66.93719	7.0	4.61	29.6	33.8	56.1	6.7
Coral 3	8/27/05	Coral	17.96335	-67.02396	10.7	5.32	30.1	34.4	57.5	5.8
Coral 4	8/28/05	Coral	17.93594	-67.04942	18.9	5.04	29.6	34.3	57.0	7.9
Coral 5	8/27/05	Coral	17.96351	-67.01735	9.1	5.31	30.1	34.5	52.5	bottom
Coral 6	8/27/05	Coral	17.96334	-67.04679	13.7	4.81	29.6	34.5	57.3	7.3
Coral 8	8/28/05	Coral	17.93339	-67.10513	4.3	4.89	30.1	34.5	57.7	bottom
Coral 9	8/28/05	Coral	17.9362	-67.11609	4.6	4.62	30.1	34.6	57.9	3.7
LTOC 1	8/30/05	Bank	17.91935	-66.96133	14.3	5.26	30.0	34.2	57.2	14.0
LTOC 2	8/30/05	Bank	17.92371	-66.98617	15.5	4.95	30.4	34.0	57.2	bottom
LTOC 3	8/30/05	Bank	17.91522	-66.98183	16.2	4.91	30.1	34.1	57.2	bottom
LTOC 7	8/30/05	Bank	17.95165	-67.00027	13.7	5.24	30.1	34.4	57.6	6.7
LTOC 8	8/30/05	Bank	17.93658	-66.98	14.9	5.02	30.2	34.3	57.5	11.9
LTOC 9	8/29/05	Bank	17.94982	-67.03835	18.3	4.86	29.8	34.4	57.3	7.6
LTOC 10	8/30/05	Bank	17.93802	-66.9709	11.6	5.10	30.1	34.4	57.5	9.1
LTOC 11	8/30/05	Bank	17.9399	-66.98	14.6	5.18	30.1	34.4	57.6	11.0
LTOC 12	8/30/05	Bank	17.91706	-66.95874	13.7	5.17	30.0	34.3	57.2	bottom
LTOC 13	8/30/05	Bank	17.92927	-66.9657	11.9	5.05	30.1	34.4	57.3	bottom
LTOC 14	8/29/05	Backreef	17.92256	-67.10196	1.2	3.86	30.7	34.4	57.5	bottom
LTOC 15	8/30/05	Lagoon	17.95547	-66.99307	11.6	5.04	30.2	34.5	57.8	6.1
LTOC 16	8/29/05	Bank	17.94553	-67.03236	18.6	4.95	29.7	34.5	57.3	10.7
LTOC 17	8/30/05	Backreef	17.94743	-67.01212	1.8	5.49	30.8	34.4	58.3	bottom
LTOC 18	8/29/05	Backreef	17.91975	-67.11229	1.8	4.73	31.5	34.4	58.5	bottom
LTOC 19	8/29/05	Bank	17.9482	-67.02749	15.8	4.90	29.8	34.4	57.3	9.8
LTOC 22	8/30/05	Lagoon	17.95699	-66.99334	10.7	5.12	30.2	34.4	57.7	6.7
HTOC 1	8/29/05	Lagoon	17.92071	-67.12399	4.0	6.18	30.7	34.4	58.2	bottom
HTOC 2	8/27/05	Bank	17.9177	-66.94946	13.7	5.15	30.0	34.1	56.8	7.6
HTOC 3	8/28/05	Lagoon	17.96823	-67.06515	7.6	5.43	31.0	34.2	58.3	4.9
HTOC 4	8/29/05	Bank	17.93527	-67.06151	19.5	4.85	30.0	34.4	57.4	11.9
HTOC 6	8/29/05	Bank	17.94524	-67.03802	18.3	4.85	29.9	34.5	57.4	8.2
HTOC 7	8/29/05	Backreef	17.94338	-67.05894	1.5	4.70	29.5	34.5	57.0	bottom
HTOC 8	8/29/05	Lagoon	17.92699	-67.12191	10.4	5.22	30.3	34.5	57.9	bottom
HTOC 9	8/28/05	Lagoon	17.95278	-67.08468	3.7	6.16	30.8	34.9	59.0	bottom
HTOC 10	8/27/05	Bank	17.93624	-66.92476	4.3	5.03	30.0	33.9	56.7	bottom
HTOC 12	8/29/05	Backreef	17.91879	-67.1185	1.9	4.80	30.4	34.4	57.7	bottom
HTOC 14	8/28/05	Backreef	17.93436	-67.11897	2.7	5.13	30.3	34.5	57.9	bottom
HTOC 15	8/27/05	Bank	17.92125	-66.95095	15.2	5.02	29.8	34.1	56.8	7.6
HTOC 16	8/30/05	Bank	17.96392	-67.01995	18.0	4.83	30.1	33.6	56.5	9.8
HTOC 17	8/27/05	Lagoon	17.95639	-66.91005	6.4	5.28	30.8	33.7	57.2	2.4
HTOC 19	8/27/05	Lagoon	17.96895	-66.92469	4.0	5.25	31.0	30.0	53.6	1.2
HTOC 20	8/28/05	Lagoon	17.9426	-67.10583	3.0	5.28	30.4	34.7	58.3	bottom
HTOC 21	8/28/05	Lagoon	17.94228	-67.09434	5.5	5.41	30.3	34.6	58.1	4.6
HTOC 22	8/29/05	Bank	17.95096	-67.05567	21.0	4.93	29.9	34.5	57.5	8.2

Abbreviations: DD, decimal degrees; m, meters; mg/L, milligrams/liter; ppt, parts per thousand; mS, milli-Siemans.

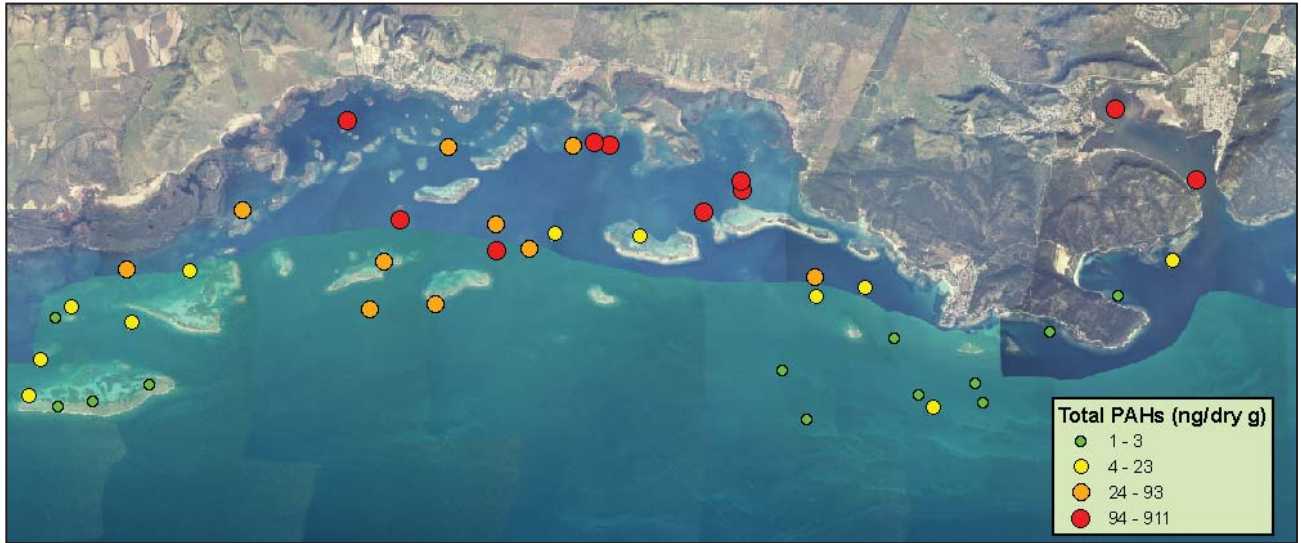


Figure 6. Map of total PAHs detected in sediments in southwest Puerto Rico. Data classified into quantiles.

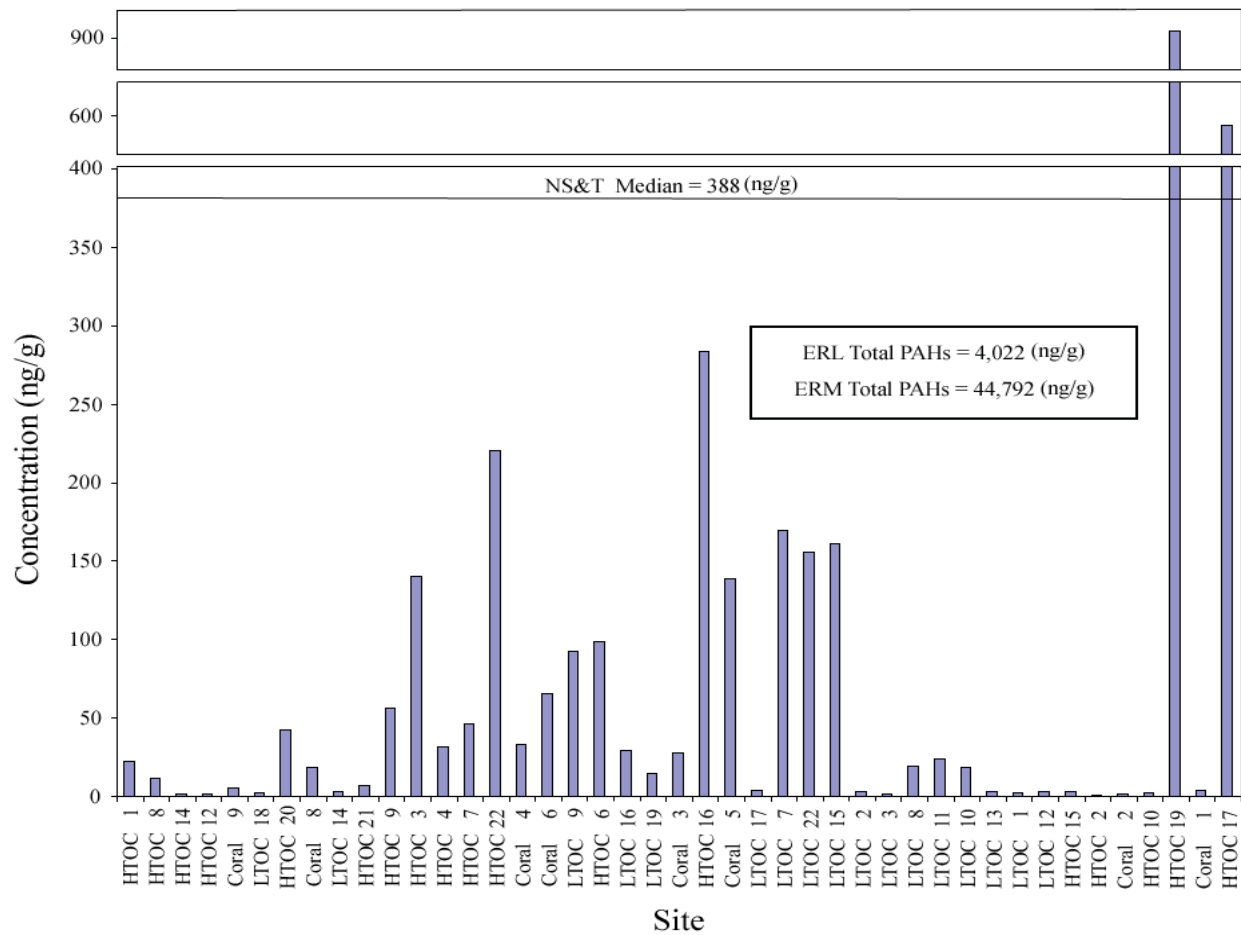


Figure 7. Total PAHs detected by site in sediments in southwest Puerto Rico.

sampled by the NCA in Bahia Montalva (Figure 2). One of the parameters measured by the NCA Program is total PAHs. The NCA list of total PAHs includes 22 individual compounds. These same 22 compounds can also be subsetted from the 59 compounds analyzed by NOAA and compared with the NCA results. The concentration of total PAHs detected at the NCA Bahia Montalva site was 25.2 ng/g (USEPA, 2004). Two sites (LTOC 15 and LTOC 22) in the present study were adjacent to the Bahia Montalva site. The sum of these same 22 PAHs measured by NS&T at each site was approximately 100 ng/g, higher than the adjacent NCA site.

In March 1973, the tanker *Zoe Colocotronis* ran aground on a reef near Cabo Rojo in Bahia Sucia, just west of the study area. Following the grounding, 37,000 barrels of Venezuelan crude oil were intentionally released to refloat the vessel (Corredor *et al.*, 1990). To assess if surface sediments in the western part of the study area showed evidence of that spill as would be indicated by increased PAHs, an ANOVA was run on total PAH concentration by longshore position. Results indicated no significant difference ($P > 0.0612$, $r^2 = 0.1701$) in PAH concentration among longshore positions.

The NS&T Program has developed effects-based, numerical guidelines to estimate the toxicological relevance of certain sediment contaminants (Long *et al.*, 1998). These guidelines, the Effects Range-Low (ERL) and the Effects Range-Median (ERM) define sediment contaminant concentration ranges that are rarely (<ERL), occasionally (ERL to ERM) or frequently (>ERM) associated with toxic effects in aquatic biota (typically amphipods) (NOAA, 1998). The ERL and ERM values for total PAHs are also in Figure 7. As can be seen, total PAHs in the sediments were well below not only the ERM, but also the ERL. Figure 8 shows the concentrations of a number of individual PAHs which have ERL and ERM values. Most of the exceedences of the NS&T national median occurred at the two sites in Guanica Bay, although there were a number of sites (*i.e.*, Coral 5, LTOC 7, LTOC 22, and HTOC 16) where the NS&T national median for naphthalene was exceeded, mostly in nearshore sites or sites near emergent reefs. None of the sites approached the NS&T 85th percentile,

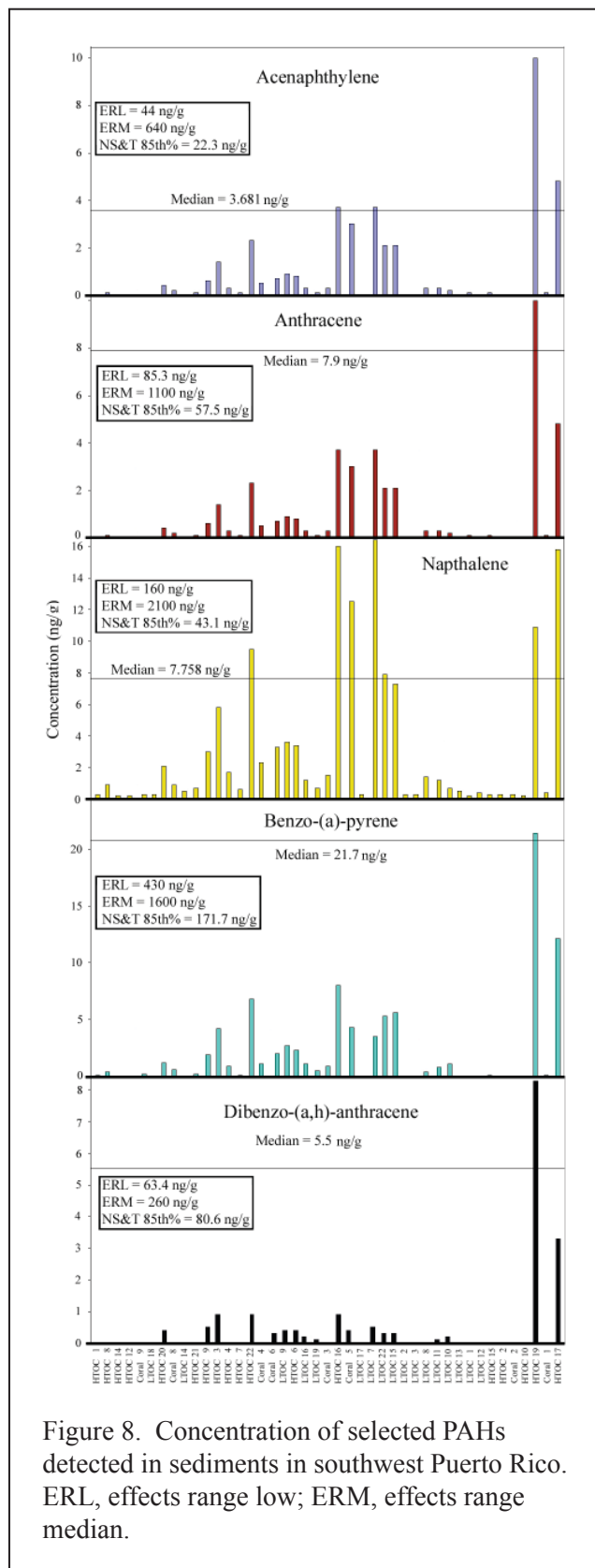
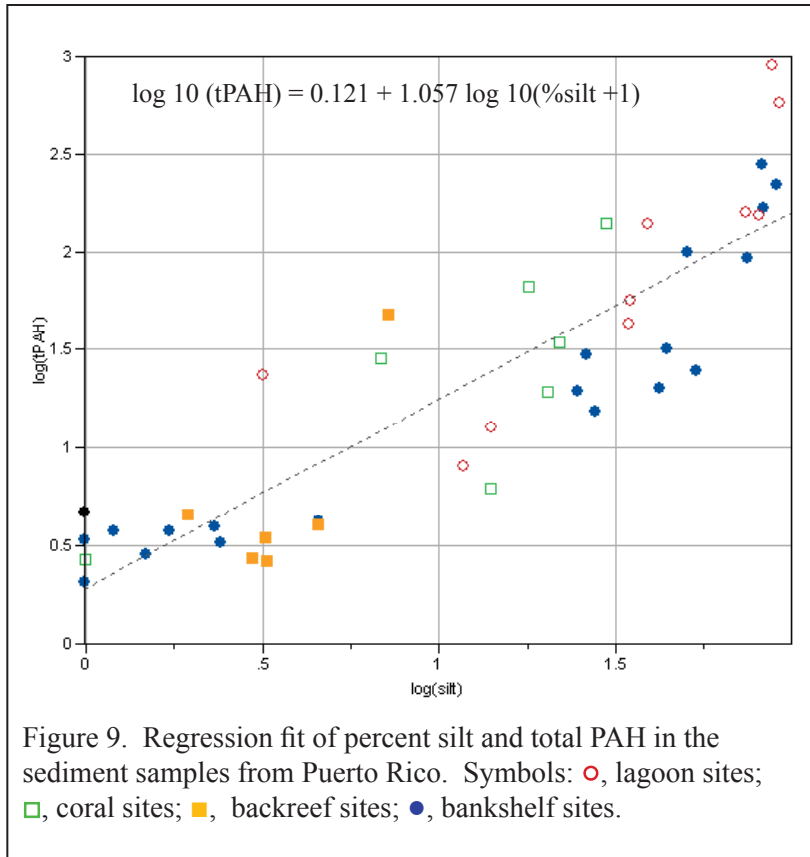


Figure 8. Concentration of selected PAHs detected in sediments in southwest Puerto Rico. ERL, effects range low; ERM, effects range median.



the ERL or ERM. Elevated levels of naphthalene (above the NS&T national median), a low molecular weight PAH, could be indicative of more recent PAH discharges, perhaps from boats.

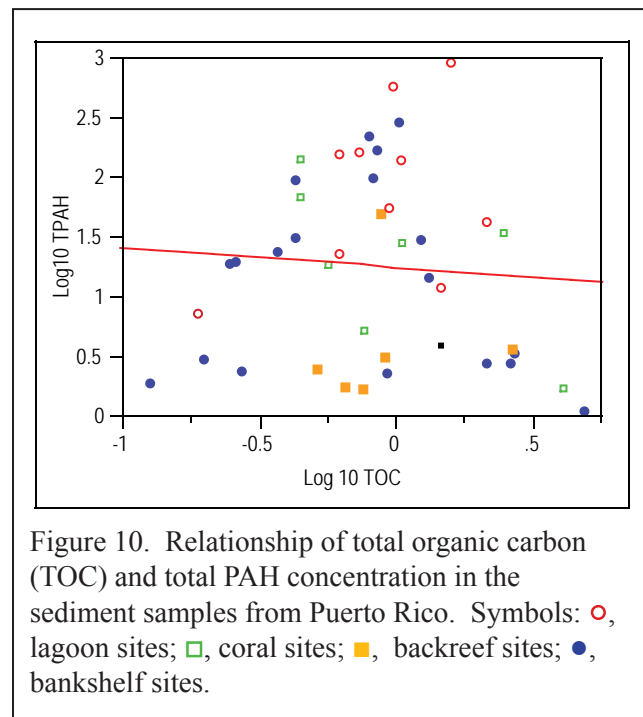
PAHs and Grain Size. As noted, the adsorption of organic contaminants onto sediments is strongly influenced by grain size (Hassett *et al.*, 1980). The smaller grain sizes of the silts and clays have proportionally higher surface areas available for the adsorption of contaminants. To assess this relationship for the samples collected in southwest Puerto Rico, a regression was run between grain size and the concentration of total PAHs found in the sediment samples. The data for grain size and PAH concentration were log₁₀ transformed. The results are shown in Figure 9. There was a highly significant ($P < 0.0001$) relationship between the silt fraction of the sediment and the concentration of total PAHs ($r^2 = 0.7944$). There was also a significant but weaker relationship ($P < 0.0001$, $r^2 = 0.3977$) between the concentration of PAHs and the clay fraction of the sediments. A number of the nearshore sites around La Parguera and also in Guanica Bay had higher silt

and clay fractions. Nonparametric tests indicated a negative correlation ($P < 0.0001$, Spearman Rho -0.8162) between the sand fraction of the sediments and total PAHs, and a significant negative correlation between the gravel fraction of the sediments and total PAH ($P > 0.0343$, Spearman Rho -0.3236).

PAHs and Total Organic Carbon.

Typically, a positive relationship exists between sediment total organic carbon (TOC) and chemical contaminants in freshwater, and in estuarine and coastal waters (Shine and Wallace, 2000; Hassett *et al.*, 1980). Because of this, organic contaminant concentrations are often normalized to the organic carbon content of sediments. A regression (Figure 10) of TOC against the concentration of PAHs (both log₁₀ transformed) in the sediments, however, was not significant ($P > 0.6531$, $r^2 = 0.005$).

Total organic carbon, particularly in estuarine systems, is usually associated with the silt fraction of



sediments. For example, a nonparametric analysis of nationwide data from the NS&T Program (excluding data from Hawaii, and other data from Puerto Rico) indicated a strong relationship between TOC and % silt ($P < 0.0001$, Spearman Rho = 0.5766). Using the Puerto Rico data from this study, however, regressions between TOC and silt ($P > 0.2474$), and TOC and clay ($P > 0.3948$) showed no positive correlation with either of these fractions. It is unclear why a significant relationship between TOC and grain size did not exist for the sediments in southwest Puerto Rico. Perhaps the organic carbon in these sediments is cycled faster or differently than in estuarine and/or temperate areas. Additional work is needed to assess TOC in the sediments in southwest Puerto Rico and possibly other tropical areas.

Total PAH concentrations in the sediments from southwest Puerto Rico were highly correlated

with sediment particle size, and not sediment organic carbon. The lack of association between contaminants and sediment TOC could also have implications in terms of the bioavailability of contaminants to coral reef organisms. Contaminants adsorbed onto sediments with lower organic carbon content could desorb from the sediments more readily (not held as tightly) and as a result, be more bioavailable to aquatic organisms.

Normalization of PAHs to Fines. An exercise was carried out to normalize the concentration of the PAHs detected in whole sediments to the percent fines in the sediment (sum of the silt and clay fractions). A plot of that normalization along with the actual concentrations of total PAHs in the sediments can be seen in Figure 11. Normalizations of this type are sometimes used to help identify sources of contaminants (Birch, 2003; Burgess *et al.*, 2001).

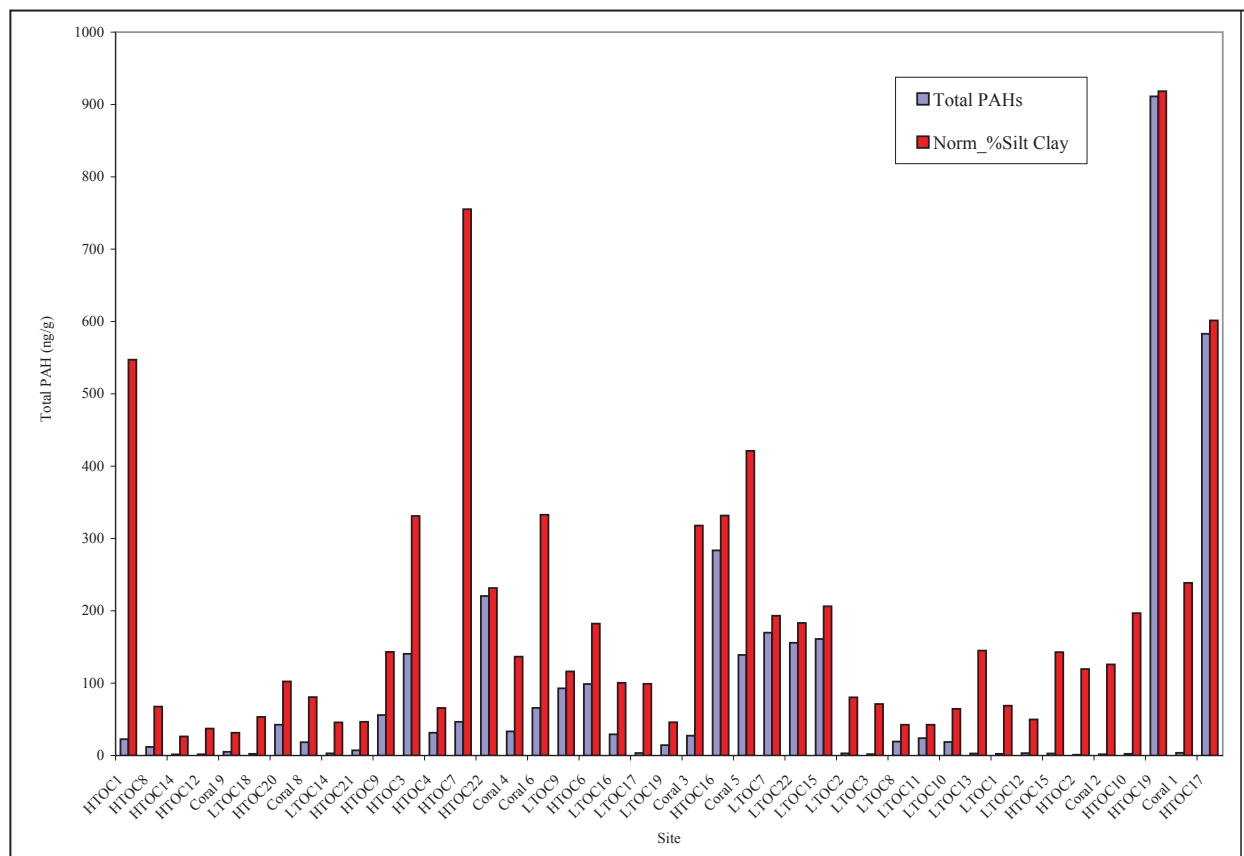


Figure 11. Comparison of total PAH to silt/clay normalized PAH concentrations. Normalized concentrations of PAHs were calculated as total PAH / silt clay fraction of the sediment.

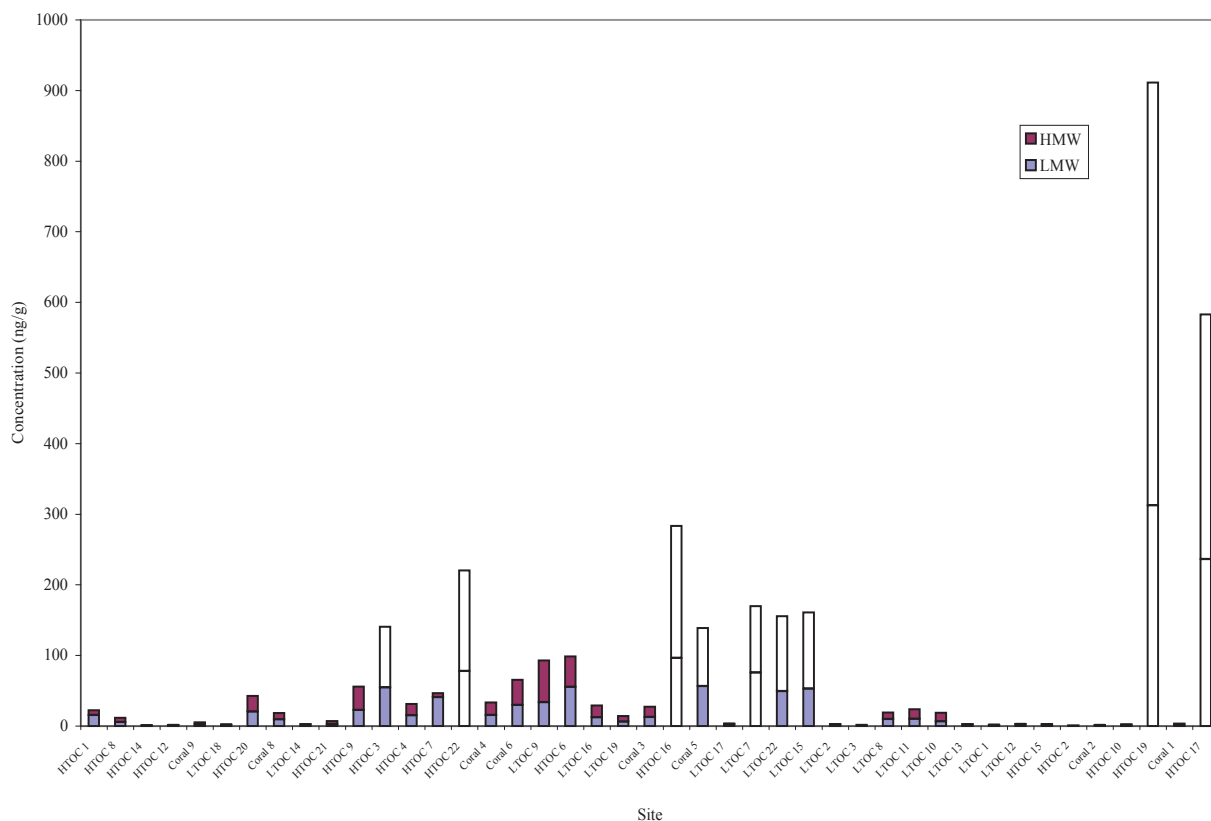


Figure 12. Low molecular weight vs high molecular weight PAHs in the sediments from southwest Puerto Rico.

From Figure 11, it can be seen that the patterns of silt/clay normalized concentrations are similar to those of total PAHs in the study area, that is elevated silt/clay normalized PAH levels occurred adjacent to La Parguera and in Guanica Bay, which would indicate that these areas are likely sources of at least some of the PAHs. At a couple of the sites (HTOC 1 and HTOC 7), elevated normalized levels may have been due to low silt/clay sediment fractions rather than high PAH levels (Appendix E).

Low and High Molecular Weight PAHs. The distribution of low (≤ 3 rings) and high (≥ 4 rings) molecular weight PAHs is shown in Figure 12. The PAHs in raw or refined petroleum products tend to be comprised of the lower molecular weight compounds. The major source of the higher molecular weight (≥ 4 rings) PAHs is the combustion of petroleum products and other organic matter (Neff *et al.*, 2005). From Figure 12, it can be seen that in

general, there was a mix of LMW and HMW PAHs (Prob > ChiSq = 0.4472) in the sediment samples, possibly indicating a mixture of both petrogenic (*e.g.*, petroleum product from fuel spills or other discharges), and pyrogenic sources.

The presence of alkylated (*e.g.*, containing methyl groups) PAHs has also been used to assess petrogenic versus pyrogenic sources (Colombo *et al.*, 1989). Alkylated PAHs are more abundant in petroleum products than in combusted materials. In Appendix E, alkylated PAHs are designated with the notations C1-C4, which indicates the number and type of alkyl group (*i.e.*, methyl, ethyl, propyl, and/or butyl) on the molecule. In the sediment samples from southwest Puerto Rico, the ratio of nonalkylated (sometimes termed the parent compound) to alkylated PAHs was 2.72, indicating a higher average concentration of nonalkylated compounds, those PAHs more commonly associated with pyrogenic

sources. This could also be due in part, however, to the limited number of alkylated PAHs analyzed in the samples.

PAH Fingerprinting. The pattern of individual PAHs present in environmental samples can be used to help discern their origins, and is termed “fingerprinting” (Zeng and Vista, 1997). Comparing individual PAHs from sources such as automobile exhaust and road dust, coal dust, creosote and even outboard engines, with the PAHs in sediments can assist in identifying major sources.

Kimbrough and Dickhut (2006), investigated PAH inputs to urban wetlands in relation to adjacent land use in the Elizabeth River, a sub-estuary of the Chesapeake Bay in Virginia. They analyzed a series of wetland sediments and compared the PAH profile to sources ranging from automobile exhaust to creosote to coal. Specifically, they looked at the patterns of five high molecular weight PAHs, including fluoranthene, pyrene, benzo[a]anthracene, benzo[a]pyrene, and benzo[ghi]perylene. Figure 13 contains a comparison of the selected high molecular weight (HMW) PAHs in the sediment samples from southwest Puerto Rico, expressed as a percent, to emissions from automobiles. The source of data on emissions from gasoline automobile engines was from Kimbrough and Dickhut (2006), Alsberg *et al.* (1985), Rogge *et al.* (1993), Singh *et al.* (1993), and Khalili *et al.* (1995). From Figure 13, it can be seen that the pattern of these five HMW PAHs from the sediments is similar to the automobile engine emission signature, indicating that automobile emissions could be an important source of PAHs in the sediments.

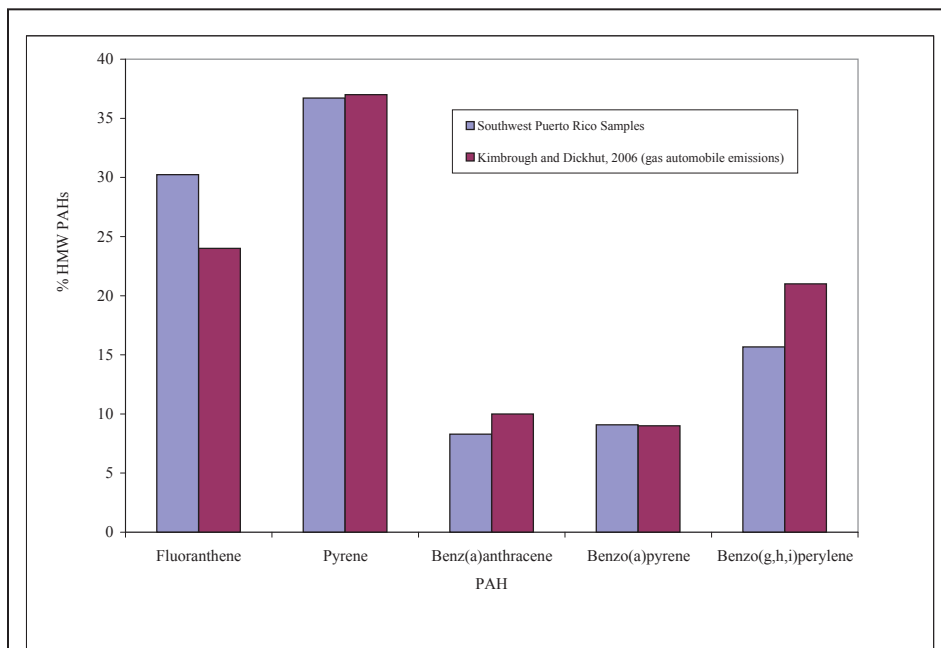


Figure 13. Comparison of selected high molecular weight (HMW) PAHs in sediment samples from southwest Puerto Rico. Data on gasoline automobile emissions from Kimbrough and Dickhut (2006), Alsberg *et al.* (1985), Rogge *et al.* (1993), Singh *et al.* (1993), and Khalili *et al.* (1995). %HMW PAHs = 100* HMW PAH concentration/ Σ fluoranthene, pyrene, benzo[a]anthracene, benzo[a]pyrene, benzo[ghi]perylene.

Neff *et al.* (2005) also investigated the signature of PAHs from a variety of sources. They looked at phenanthrene:anthracene, and fluoranthene:pyrene ratios from different sources. For auto exhaust soot, a ratio of 1.79 was reported for phenanthrene:anthracene, and a ratio of 0.90 for fluoranthene:pyrene. In southwest Puerto Rico, the ratio of phenanthrene:anthracene was 2.90, and the ratio of fluoranthene:pyrene was 0.82, somewhat similar to what Neff *et al.* (2005) found in automobile exhaust.

Miller *et al.* (2003) reported on the relationship between boating activity and contamination from PAHs on Lake Tahoe in Nevada and California, for a suite of 11 PAHs discharged in the exhaust from various types of outboard engines (four cycle, 2-cycle fuel injected, and 2 cycle carbureted). The study area in southwest Puerto Rico has a significant amount of recreational boating activity, particularly on weekends, along with some commercial fishing

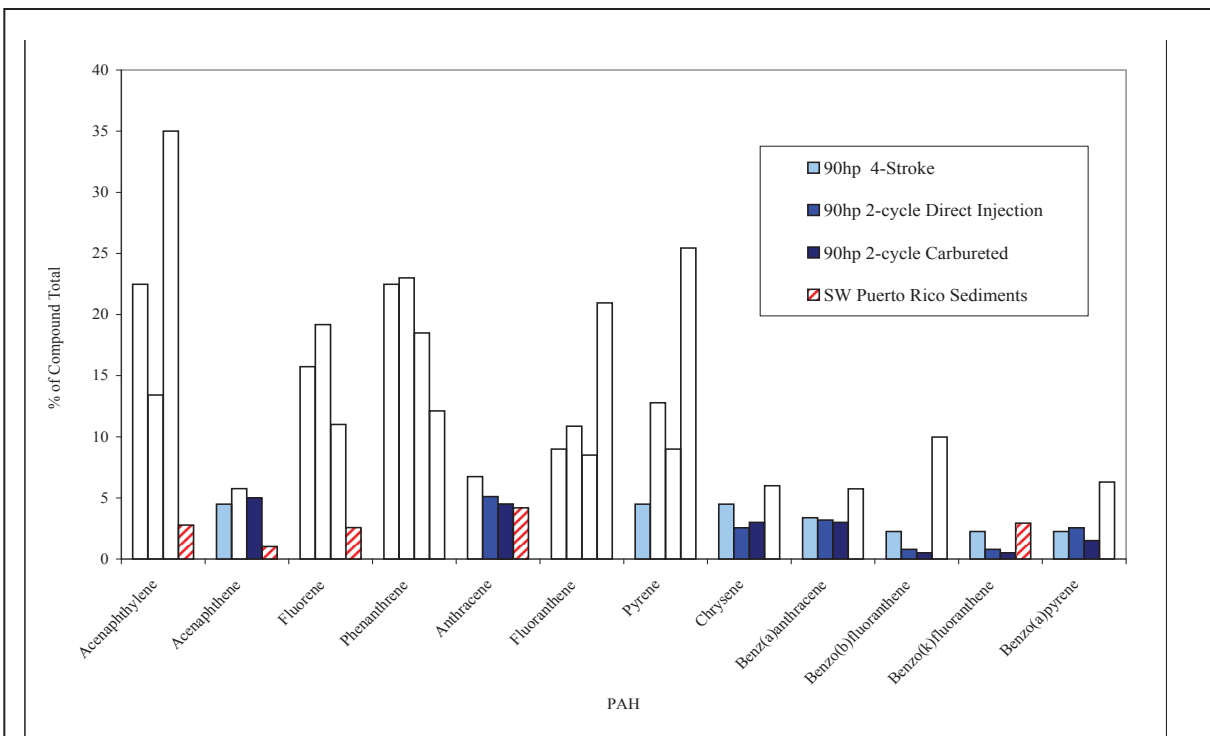


Figure 14. Comparison of signatures for selected PAHs in outboard engines with PAHs in sediments from southwest Puerto Rico. Outboard engine data reported in Miller *et al.* (2003).

boats. Comparing the PAH signal from outboard engines with the pattern of PAHs detected in the sediments in southwest Puerto Rico provides another opportunity to assess possible sources. Miller *et al.* (2003) reported that the outboard engine exhaust (blue bars) was enriched in acenaphthylene, fluorene and phenanthrene for all outboard engine types tested (Figure 14).

The sediments collected in southwest Puerto Rico (Figure 14, red striped bars), however, had higher relative contributions (percent) of fluoranthene and pyrene. The pattern of PAHs in the sediments appears more similar to emissions from automobile engines than outboard engines, although phenanthrene is elevated in both automobile and outboard engine discharges. Some of the boats around La Parguera have inboard engines, a number of which may be converted automobile engines which could have the same or similar exhaust discharge characteristics of automobile engines.

PAHs and Habitat. Habitats in the study area vary from nearshore mangrove and lagoon sites to

shallow areas behind emergent reefs (backreef), to the deeper bank reefs. To assess whether total PAHs in the sediments vary by habitat, an ANOVA was run followed by pairwise comparisons (Tukey-Kramer HSD). The results along with the 95% confidence intervals are shown in Figure 15. There was a significant difference ($P > 0.0077$, $r^2 = 0.2605$) in the total PAH concentration among habitats. The pairwise comparison indicated that the concentration of total PAHs in the backreef areas were significantly lower than the concentrations of PAHs in lagoons. In general, the backreef areas tended to have lower percentages of silt and clay.

Effects of PAHs and other Petroleum Hydrocarbons on Corals. This section of the report provides more detailed information on the documented effects of PAHs on corals. Most of the research on the effects of hydrocarbons on corals involves the accumulation and effects of crude and refined petroleum, with only very limited data available on individual PAHs. Only five papers were located that assessed individual PAHs in corals, four of those concerned with uptake

and depuration of PAHs. The remaining papers examined the effects of oil on corals. Because this project focused on individual PAHs, only a few of the studies investigating effects of oil on corals are presented here.

The research located on the effects of PAHs and other petroleum hydrocarbons on corals focused on aqueous exposures, i.e., the compound or mixture added to water. It is important to note that the concentrations of PAHs found to impact corals as a result of aqueous ($\mu\text{g/L}$) exposures cannot be directly compared with the concentrations of PAHs that have been found in the sediments (ng/g) in southwest Puerto Rico. PAHs and many other organic contaminants strongly bind to sediments, so that the overlying aqueous concentrations are typically orders of magnitude lower than what is seen in the sediments. PAHs and many other chemical contaminants, however, are continuously introduced into the water column as a result of processes such as stormwater runoff, atmospheric deposition, marine activities (such as boating and shipping), spills, resuspension as a result of storms, etc. Once in the water column, PAHs tend to partition out onto sediments and into biota. The sediments do provide an indicator or record of what has been introduced into the marine environment.

Effects of PAHs. Peachey and Crosby (1996) carried out a study to look at the phototoxicity of PAHs. Phototoxicity refers to the toxic effects of chemicals caused or enhanced by light (Peachey

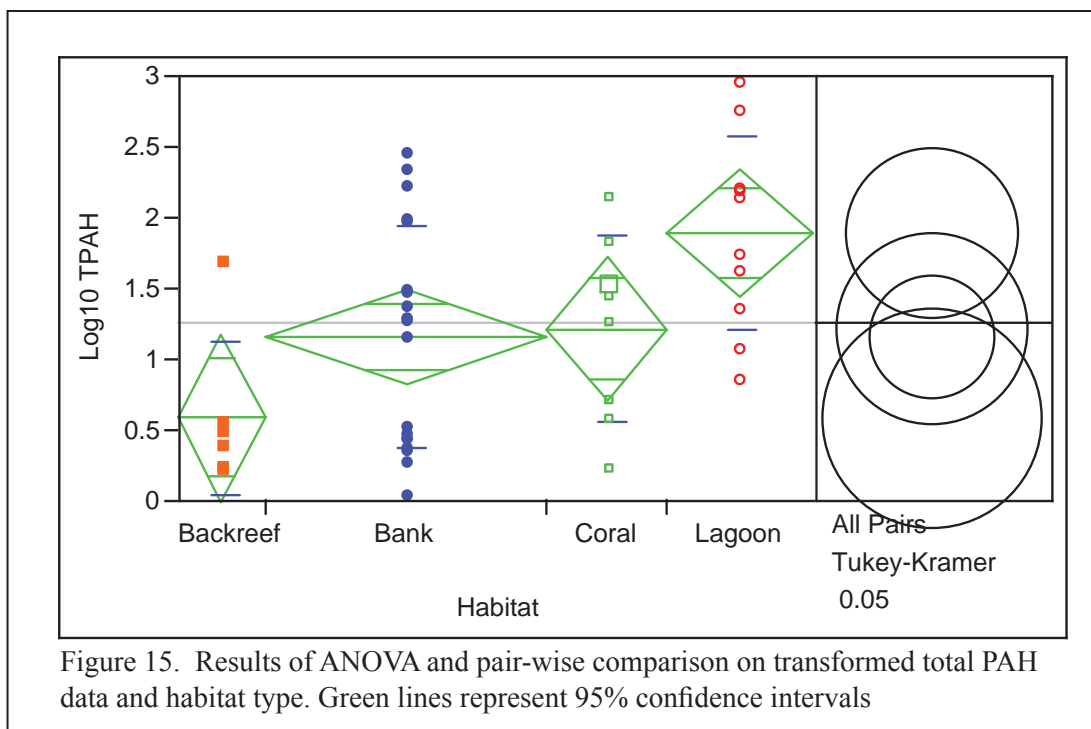


Figure 15. Results of ANOVA and pair-wise comparison on transformed total PAH data and habitat type. Green lines represent 95% confidence intervals

and Crosby, 1996). In the laboratory, larvae of the coral *Fungia scutaria* (mushroom coral) were first exposed to nominal concentrations of pyrene ranging from 1-48 $\mu\text{g/L}$ for a period of 2 hours. Larvae were then exposed to artificial sunlight and evaluated for changes in their mobility over a period of eight hours. A nominal pyrene concentration of 32 $\mu\text{g/L}$ resulted in a significant number (40%) of immobilized larvae. Changes in the mobility of the coral larvae, or their ability to find suitable substrate could impact recruitment, important in maintaining and building the reef structure. In addition, the phototoxicity of PAHs could be an issue especially in tropical areas, which receive intense sunlight during much of the year.

Kennedy *et al.* (1992) investigated the uptake and metabolism of radiolabeled (^{14}C) benzo[a]pyrene in two species of corals, *Montastrea annularis* and *Favia fragum*. Whole corals were exposed to a nominal concentration of 5 $\mu\text{g/L}$ benzo[a]pyrene for a period of 25 hours. Corals were then moved to uncontaminated flowing seawater, and depuration was followed over a period of 144 hours. Uptake of benzo[a]pyrene was rapid in both species, a little higher in *M. annularis*, which Kennedy *et al.* (1992)

noted could be related to a higher lipid content in *M. annularis*. Approximately 38 and 65 percent of the accumulated radioactivity remained in *F. fragum* and *M. annularis* respectively, 144 hours after moving them from the test solution.

Kennedy *et al.* (1992) noted that the rate of metabolism of benzo[a]pyrene in both species was low. After 144 hours, between 79 and 90 percent of the organic soluble radioactivity was in the form of unmetabolized benzo[a]pyrene. They noted that for both of these species, the low capacity to metabolize and eliminate xenobiotics compared to other marine invertebrates indicated that corals could be susceptible to environmental pollutants.

Knap *et al.* (1982) exposed the brain coral *Diploria strigosa* to ¹⁴C labeled phenanthrene at a measured concentration of 33 µg/L for a period of 24 hours. Corals were then removed to a flow-through aquarium with filtered water to observe depuration. Knap *et al.* (1982) found that the phenanthrene was readily accumulated by *D. strigosa*. When the corals were removed to the aquarium with flow-through seawater, there was a period of rapid decrease (~70 percent) in phenanthrene (and metabolites) concentration after two days as measured by the radioactivity remaining in the corals, followed by a much slower depuration rate between two and 10 days. Knap *et al.* (1982) concluded that the uptake of phenanthrene was similar to that of other invertebrates, and that the depuration rate was similar to the mussel *Modiolus modiolus*.

Solbakken *et al.* (1984) investigated the uptake of radiolabeled naphthalene and phenanthrene in *D. strigosa* and in a number of other corals. At a concentration of 3 µg/L, phenanthrene was rapidly accumulated by *F. fragum* and *M. annularis*. *D. strigosa* also accumulated the phenanthrene, but at a lower rate than the other two species. Solbakken *et al.* (1984) attributed this to taxonomic differences. While the uptake of phenanthrene was slower in *D. strigosa*, the depuration was also slower. Radioactivity, in the form of parent compound and any metabolites, was still detectable in the corals one year after the exposure. The uptake of another compound, naphthalene, was also rapid, and

depuration of this lower molecular weight PAH was faster than for phenanthrene in both *D. strigosa* and in the green cactus coral *Madracis decactis*. After two days, 92 percent of the naphthalene and any metabolites had been depurated (Solbakken *et al.*, 1984).

Neff and Anderson (1981) exposed the coral *Oculina diffusa* to radiolabeled naphthalene. Naphthalene was added to a 10 percent solution of crude oil, producing a naphthalene exposure concentration of approximately 42 µg/L. Similar to what Solbakken *et al.* (1984) found, the bioconcentration of naphthalene in *O. diffusa* was rapid during the seven hour exposure period. The corals were then moved to clean flowing seawater in order to assess depuration. As with *D. strigosa*, depuration of naphthalene proceeded rapidly, and Neff and Anderson (1981) calculated a naphthalene elimination half-life of 24 hours, and estimated that the naphthalene would be cleared from the tissues after 14 days.

Effects of Crude and Refined Oil. Negri and Heyward (2000) investigated the effects of the water accommodated fraction (WAF) of heavy crude oil on the scleractinian coral *Acropora millepora*. The WAF is the amount of oil that remains in the aqueous phase after mixing and then phase separation. In the WAF, oil can be truly dissolved or in particulate (*e.g.*, emulsions or oil droplets) forms. In these experiments, the oil and water (1% solution) were stirred vigorously for two hours, and then allowed to separate for an additional two hours. Coral gametes collected from reefs during spawning events were used in the experiments. Concentrations of total hydrocarbon (THC) up to 165 µg/L failed to significantly inhibit fertilization. However, larval settlement and metamorphosis was significantly affected by a THC concentration of 82 µg/L, and *A. millepora* larval settlement was completely inhibited at a concentration of 165 µg/L.

Epstein *et al.* (2000) investigated the effects of the water soluble fraction (WSF) of Egyptian crude oil on the settlement of coral larval *Stylophora pistillata*. The WSF is that amount of oil truly dissolved in an aqueous solution. A 0.1% WSF test solution was used. No mortality was observed during these

experiments, and there were no observed alterations in swimming behavior. However, the 0.1% WSF prevented any larval settlement after 12 hours. (Epstein *et al.*, 2000). Dodge *et al.* (1984) exposed the brain coral *D. strigosa* in a series of field and laboratory experiments to light crude oil at measured concentrations ranging from 1,000 - 50,000 µg/L. Corals were exposed for up to 24 hours, and the corals were then moved back to or left on the reefs for a year. Dodge *et al.* (1984) found no effects of any concentration on the growth on *D. strigosa*.

Downs *et al.* (2006) investigated cellular physiological condition in the hard coral *Porites lobata* in an area where an oil spill occurred in Yap, Federated States of Micronesia. Using a series of molecular biomarkers (e.g., heat shock proteins, ferrochelatase, catalase, and glutathione-S-transferase), Downs *et al.* (2006) showed that physiological processes had been affected in the corals. A followup study that exposed healthy *P. damicornis* corals to the fuel oil (Rougee *et al.*, 2006) in the laboratory produced results that were nearly identical to those observed in the field.

Several studies have documented the association between the presence of oil and impacts in coral communities. Bak (1987) documented a deterioration in the reef structure adjacent to, and downstream of an abandoned oil refinery. Some coral species such as *M. annularis* and *A. palmata* were completely absent in the transects just upstream of the refinery and 5 km downstream of the site, along the coast. Interestingly, however, some corals such as *D. strigosa* were relatively abundant in the area adjacent to the abandoned refinery. Loya (1975) also documented an association between large oil refineries and low coral larval recruitment.

From these studies, it can be seen that PAHs along with crude and refined petroleum products can significantly affect corals. The effects of individual PAHs and crude or refined petroleum products in the environment are certain to be variable and dependent on factors including species involved, contaminants present, how the contaminants are introduced (e.g., spill or chronic discharges), and environmental conditions such as depth, currents and time of year.

In addition, impacts may not become apparent immediately if the contaminants have more subtle effects, for example on metabolism or reproduction, the results of which may take longer to be manifested. Additional work is needed to better understand linkages that may exist between contaminants such as PAHs and effects at the community and species levels.

Other Effects of PAHs. PAHs can also accumulate in the tissues of fish, aquatic plants, and marine invertebrates. Fish, however, have the ability to metabolize PAHs, and as such their tissues often do not show high levels of PAHs in known areas of contamination, while many marine invertebrates and mollusks are less able to efficiently metabolize PAHs (Neff, 1985). The effects of PAHs on aquatic organisms are fairly well documented. For example, Ott *et al.* (1978) found that life cycle exposure of the marine copepod *Eurytemora affinis* to naphthalene at 10 µg/L resulted in significantly reduced life spans, brood sizes and total number of nauplii produced. Hose *et al.* (1982) found that a concentration of 0.1 µg benzo[a]pyrene for five days resulted in reduced and delayed hatch in the eggs of the sand sole *Psettichthys melanostictus*.

Polychlorinated Biphenyls

Total PCBs detected in the sediments are shown in Figures 16 and 17. Appendix F contains results of the analysis for individual PCBs. Total PCBs as included in this report represents the sum of 18 congeners multiplied by 2.19 to estimate the total amount of all (209 congeners) PCBs present in a sample (Lauenstein and Cantillo, 1993).

Comparison with NS&T Data. As with the PAHs, there was a pattern of elevated total PCB levels adjacent to La Parguera and especially in Guanica Bay (Figure 16 and 17). When compared with the NS&T sediment data, a number of the sites around La Parguera had total PCB levels above the NS&T national median of 8 ng/g, including HTOC 3, Coral 6, and Coral 3. Several sites around La Parguera, including LTOC 7, 15, 22, and HTOC 16 and 22 were also above the ERL for total PCBs of 22.7 ng/g. None of the sites adjacent to La Parguera were above the NS&T 85th percentile of 80 ng/g, or the

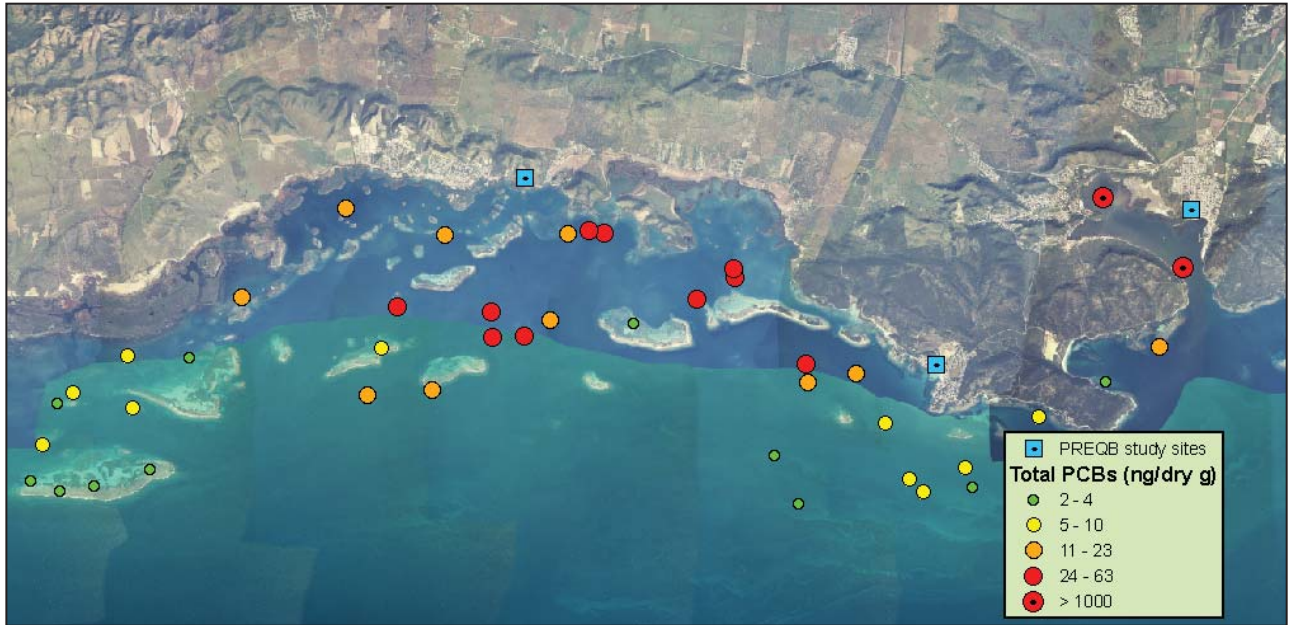


Figure 16. Map of total PCBs detected in the sediments in southwest Puerto Rico. Data classified into quantiles. (PREQB = Puerto Rico Environmental Quality Board)

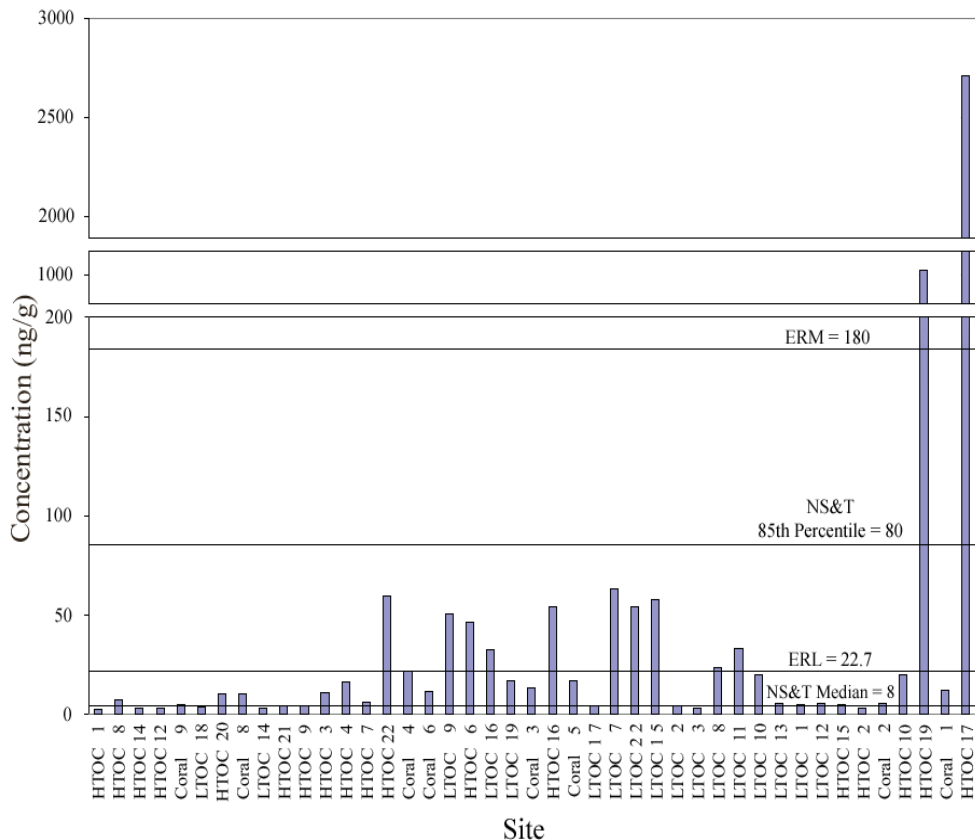


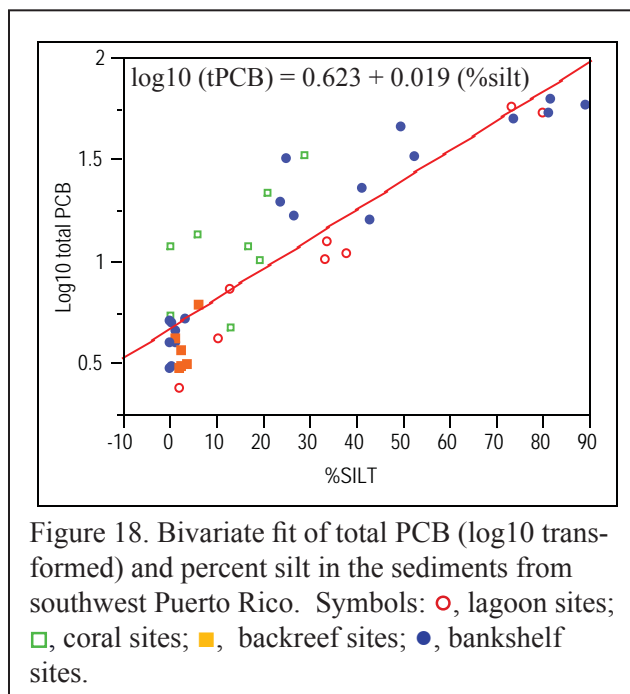
Figure 17. Total PCBs detected by site in the sediments in southwest Puerto Rico. The ERL or Effects Range-Low is the concentration of a contaminant below which toxicity (10th percentile) is rarely observed. The ERM or the Effects Range-Median is the concentration above which toxicity in test organisms is more frequently (50th percentile) observed.

ERM of 180 ng/g. In Guanica Bay (HTOC 17 and 19), however, the concentration of total PCBs at the sites sampled was not only above the NS&T 85th percentile, it was also substantially above the ERM value of 180 ng/g, indicating that PCBs are more likely to be impacting aquatic biota at these sites. Total PCBs at HTOC 17 in Guanica Bay (estimated at 2,700 ng/g) were over two orders of magnitude higher than the average found at the other sites in the study area. The other site sampled in Guanica Bay, HTOC 19, had an estimated total PCB sediment concentration of over 1,000 ng/g.

PCBs were manufactured in the U.S. between 1929 and 1977, and used in electrical applications including transformers and capacitors as insulators and coolants, and also in hydraulic and heat transfer systems, lubricants, paints, plasticizers, adhesives, flame retardants, brake linings, and asphalt. Approximately 60 percent of PCBs manufactured in the U.S. were used in electrical applications (USEPA, 1997). In the United States, all PCBs produced in North America were made by a single manufacturer, and the commercial products were referred to as Aroclors. Aroclors are mixtures of PCB congeners. For example, Aroclor 1242 refers to a commercial mixture of PCBs that is 42 percent chlorine by weight.

The presence of highly elevated levels of PCBs at the sites sampled in Guanica Bay could be the result of a spill, industrial discharges, dumping of PCB contaminated equipment (*e.g.*, transformers), discharges from ships, or runoff (*e.g.*, from rainfall or cleaning activities) from industrial sites, to mention a few possibilities. There have been a number of industrial operations within Guanica Bay, ranging from sugar refining to fertilizer production to textile manufacture to electronics. It would be useful, at some point, to better characterize the spatial distribution of PCBs within Guanica Bay, including PCBs in sediment cores, along with possible biological effects.

Comparison with Other Data Sources. Data from the Puerto Rico Environmental Quality Board (PREQB) was also accessed for this report using EPA's STORET database (USEPA, 2005).



The PREQB periodically monitors a variety of parameters, including a number of contaminants in water samples. Five PREQB sites are located inland or along the shore in the study area. Three of those sites are shown in Figure 16. PCBs were not detected in any of the water samples analyzed by PREQB. As part of EPA's NCA Program, total PCBs in the sediments were measured. The sum of the 10 individual congeners measured by the NCA was 3.10 ng/g. The sums of these same congeners measured at the two adjacent sites in Bahia Montalva (LTOC 15 and LTOC 22) by the NS&T Program were 8.04 and 7.09 ng/g, respectively, similar to what was measured by NCA.

PCBs and Grain Size. The relationship between PCBs, grain size and TOC was also examined. Figure 18 contains a plot of total PCB (log 10 transformed) and percent silt in the sediments. A nonparametric analysis of the data revealed a significant positive correlation between the silt fraction of the sediment, and total PCBs ($P < 0.0001$, Spearman Rho = 0.8578). An analysis of total PCBs and TOC (Figure 19), however, revealed no significant relationship ($P > 0.9242$, Spearman Rho = 0.0150), similar to what was observed for PAHs. For both PAHs and PCBs, sediment chemical

contaminant concentration appeared to be influenced by sediment particle size and not sediment TOC.

Normalization of PCBs to Fines. Figure 20 contains the results of the normalization of total PCBs to the silt/clay fraction (fines) of the sediment. The sites in Guanica Bay, particularly HTOC 17 dominate the graph, indicating that PCB concentrations are high in the Bay, even when normalized for the fine fraction of the sediment. It is also interesting to note that a number of the sites adjacent to Guanica Bay where sediments were collected (e.g., Coral 1, HTOC 10, Coral 2, HTOC 2 and HTOC 15) also have higher normalized PCB concentrations, possibly indicating some degree of transport of PCBs out of Guanica Bay. The predominant longshore flow in the study area is from east to west (CFMC, 1998).

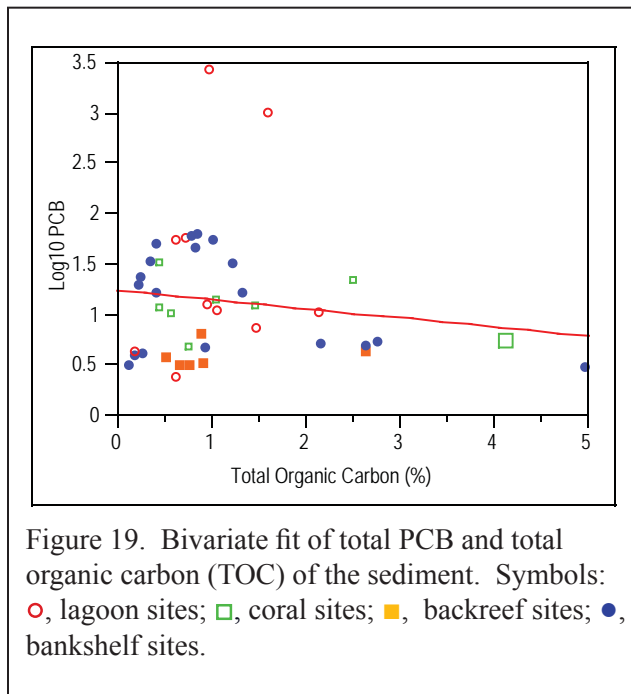


Figure 19. Bivariate fit of total PCB and total organic carbon (TOC) of the sediment. Symbols: ○, lagoon sites; □, coral sites; ■, backreef sites; ●, bankshelf sites.

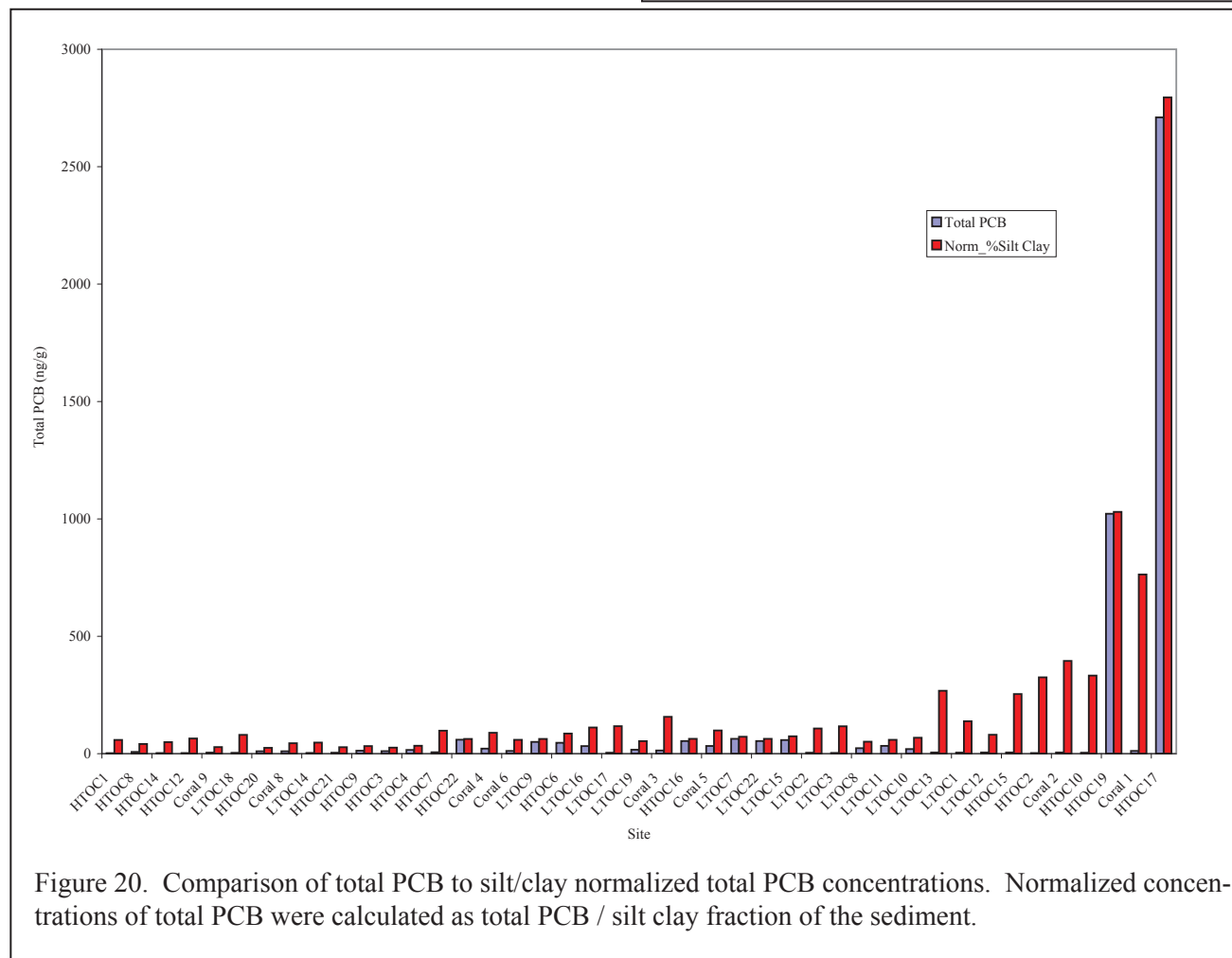


Figure 20. Comparison of total PCB to silt/clay normalized total PCB concentrations. Normalized concentrations of total PCB were calculated as total PCB / silt clay fraction of the sediment.

Comparison of PCBs in the Sediments with Aroclors.

A type of fingerprinting is also possible with PCBs. The commercial Aroclor mixtures manufactured in the U.S. contained PCBs ranging in chlorination from two (DiPCBs) to the fully chlorinated PCB containing 10 chlorine atoms, referred to as DecaPCB. By comparing the “signature” of the various Aroclor formulations to the PCBs found in the sediments in southwest Puerto Rico, a better understanding can be developed of the Aroclors that may have been used.

Figure 21 shows the relative proportions of the different classes (e.g., TetraPCBs, PentaPCBs, etc.) of PCBs in Aroclor 1242, 1248, 1254 and 1260. These formulations consist of PCBs containing 2-7 chlorines. In Figure 22, the relative proportions of these same PCB classes found in the sediment samples found at the two sites in Guanica Bay are presented. By comparing the two graphs, it can be seen that the pattern of PCBs in the two sediment samples from Guanica Bay shares some similarity with Aroclors 1248 and 1254. It is likely, however, that some degradation of PCBs in the sediments in southwest Puerto Rico has occurred making this comparison more difficult.

Effects of PCBs on Corals.
Only one laboratory study on the effects of PCBs on

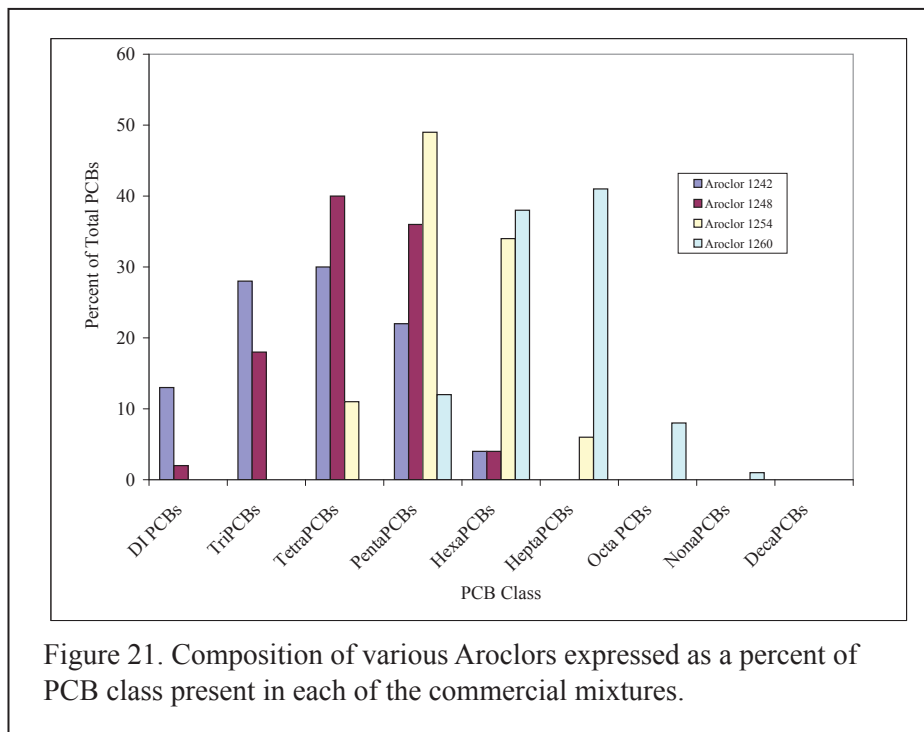


Figure 21. Composition of various Aroclors expressed as a percent of PCB class present in each of the commercial mixtures.

corals was located. Solbakken *et al.* (1984) assessed the bioconcentration of radiolabeled HexaPCB (2,4,5,2',4',5'-hexachlorobiphenyl) in *D. strigosa* and *M. decatis*. The PCB was accumulated in both species, but at lower levels than what was found for

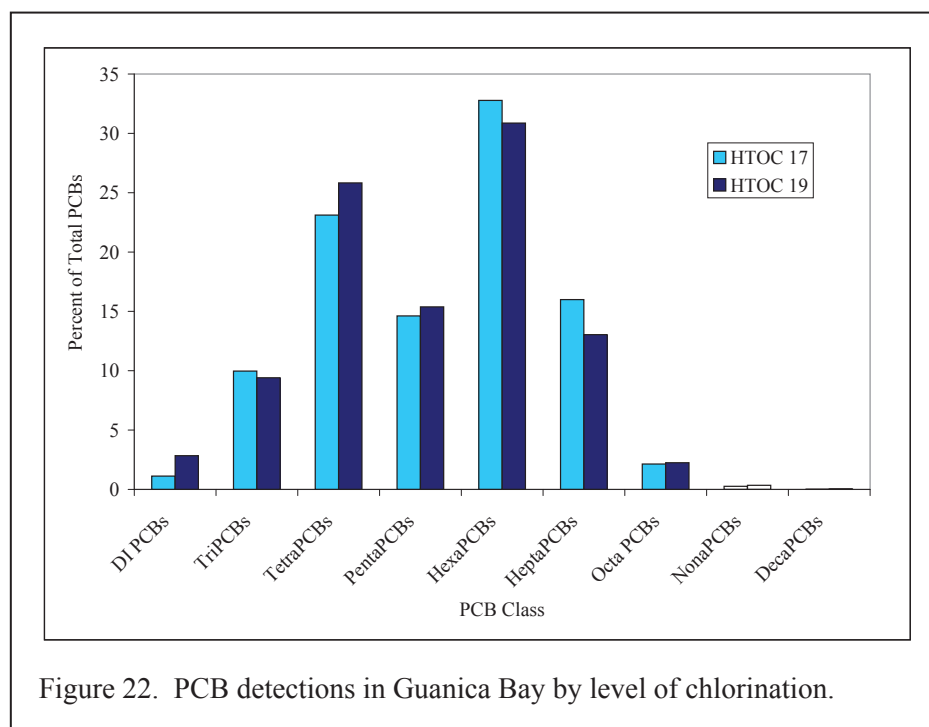


Figure 22. PCB detections in Guanica Bay by level of chlorination.

phenanthrene and naphthalene by Solbakken *et al.* (1984). Depuration of this PCB also proceeded at a slower rate than for either phenanthrene or naphthalene. After 275 days, nearly 33 percent of the original radioactivity from the PCB remained in the coral. Additional information on PCBs, particularly effects on corals would be useful as elevated concentrations of a number of the PCB congeners were found in the sediments in the study area. The slower depuration of this PCB congener, at least in the two species investigated by Solbakken *et al.* (1984) would indicate that once accumulated, PCBs would remain in the tissues, increasing the likelihood of adverse effects.

Other Effects of PCBs. PCBs readily accumulate in the tissues of other organisms including many marine animals such as filter feeders, fish, and marine mammals. PCBs can take years to degrade in sediments. Biomagnification of PCBs occurs with higher levels found in the tissues of animals at the top of the food chain.

In general, PCBs have been shown to adversely affect reproduction, growth, metabolism and survival. Effects can include hepatotoxicity, immunotoxicity, neurotoxicity, low birth weight, and teratogenicity (Eisler and Belise, 1996).

Organochlorine Pesticides

Organochlorine pesticides like DDT and chlordane were widely used in the past to control a variety of insect pests, both in agriculture and in and around structures, including houses (*e.g.*, mosquito, ant, and termite control). Most organochlorine pesticides have been banned due to their long environmental half lives, ability to bioaccumulate, and toxic effects on nontarget organisms. DDT (dichloro-diphenyltrichloroethane), was banned by the EPA for most uses in the U.S. in 1972. DDT is still effectively used in many developing countries to help control malaria. Most uses of chlordane were canceled in 1978, and all uses were canceled by 1988. Endosulfan is another chlorinated insecticide that is still in use, primarily in agriculture.

DDT. Figure 23 and 24 contain the results of the analyses of total DDTs in sediments. The results for

the individual organochlorine pesticides are included in Appendix G. Total DDTs, as included in Figure 23 and 24 and in Appendix G, is the sum of the parent isomers (4,4'-DDT and 2,4'-DDT) and the DDE and DDD degradation products.

Comparison of DDT with NS&T Data. As with the PAHs and the PCBs, there were elevated levels of DDT in the sediments near the town of La Parguera, and even higher concentrations at the two sites in Guanica Bay. With the exception of the two sites in Guanica Bay, all concentrations of total DDTs were below the ERL value of 1.58 ng/g, and below the NS&T national median and 85th percentile. A Wilcoxon nonparametric test revealed no differences in total DDTs concentration by habitat type.

In Guanica Bay, total DDTs exceeded the NS&T 85th percentile of 18 ng/g, and in HTOC 17, total DDTs concentration of 46.93 ng/g slightly exceeded the ERM value (46.1 ng/g), indicating a greater likelihood of biological effects on biota inhabiting this area. However, elevated levels of total DDTs did not appear to be present at the sampling sites just outside of Guanica Bay.

Comparison with Other Data Sources. The PREQB measured DDT and its metabolites in water samples at two sites south of Guanica Bay. Both DDT and its metabolites were below the level of quantification in the water samples. EPA's NCA Program also quantified total DDTs in the sediments in Bahia Montalva in 2000. Total DDTs in the sediment was 0.072 ng/g. Total DDTs measured in the sediments at LTOC 15 and LTOC 22 were 1.45 and 1.28 ng/g, respectively, higher than that found by the NCA Program.

DDT and % Silt. A regression of total DDTs (log₁₀ normalized) and % silt is shown in Figure 25a. A nonparametric analysis of the data indicated a significant positive correlation between total DDTs and the silt fraction of the sediment ($P < 0.0001$, Spearman Rho = 0.8381). A similar analysis to look at the association between total DDTs and sediment TOC (Figure 25b), however, revealed no significant association ($P > 0.7755$, Spearman Rho = 0.0448).

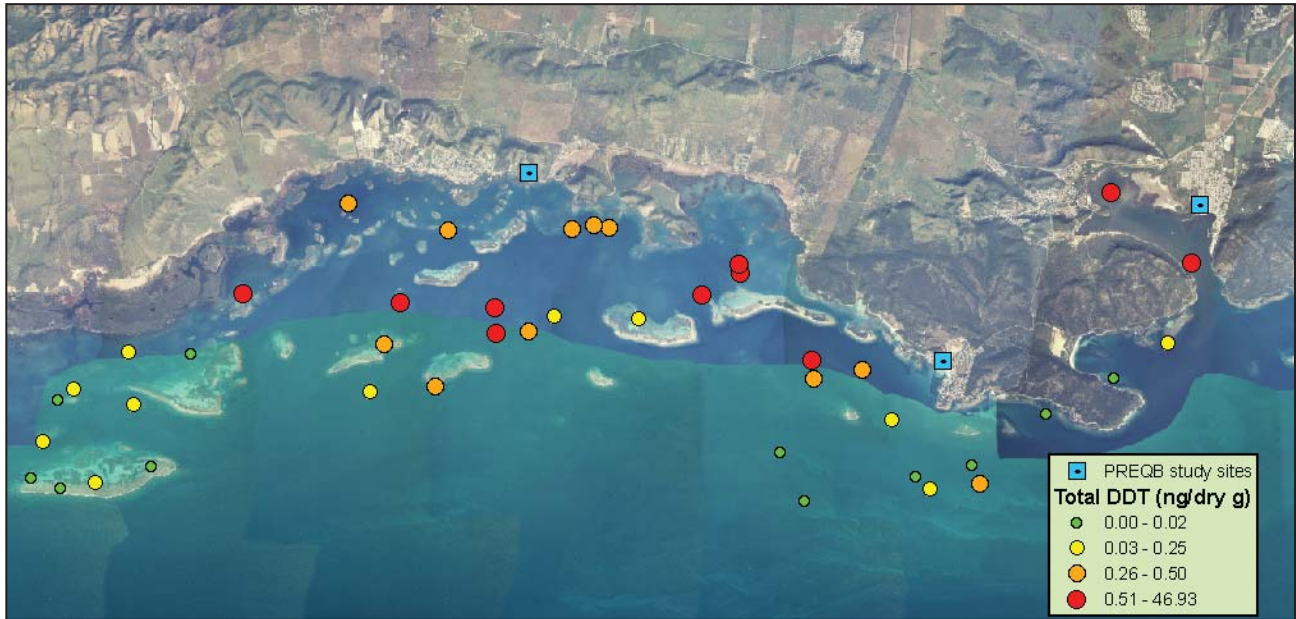


Figure 23. Map of total DDT detected by site in the sediments in southwest Puerto Rico. Data classified into quartiles.

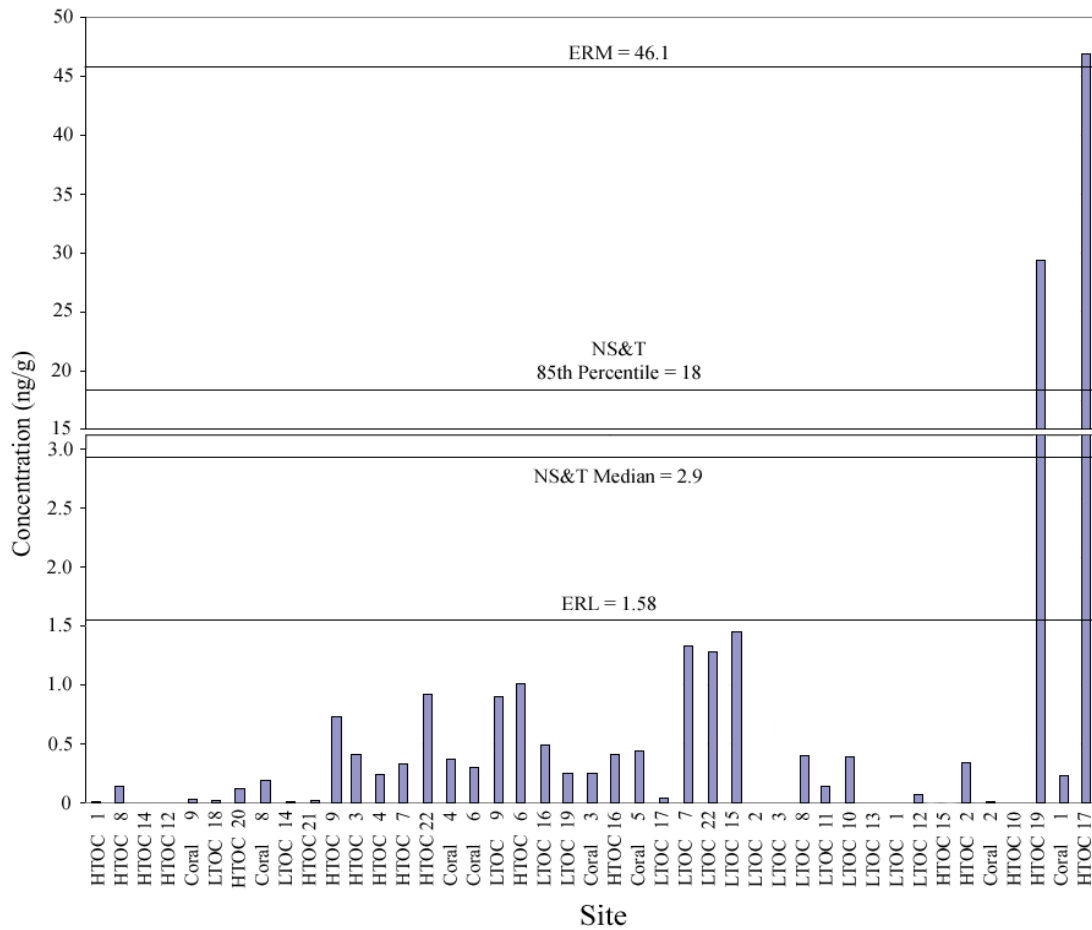


Figure 24. Total DDT detected by site in the sediments in southwest Puerto Rico.

The elevated levels of DDT and its metabolites (Appendix G) in Guanica Bay could be related to past agricultural activity. Much of the Lajas Valley drains to Guanica Bay through a series of manmade canals. The area has had extensive agricultural activity over the years, including sugarcane and pineapple production. DDT may have been applied to both of these crops to control insects on the foliage and in the soil.

Effects of DDT on Corals. No information was found on the effects of DDT in corals.

Other Effects of DDT. DDT readily accumulates in the tissues of marine organisms, especially in the adipose tissues of fish, shellfish, whales, and seals. DDT and the metabolites DDD and DDE are also highly persistent in the environment. The acute toxicity of DDT is due to its neurotoxic action, affecting the central nervous system by disrupting neural ion regulation. The chronic effects of DDT, however, have proved to be more of a problem. DDT was found to adversely affect avian reproduction, particularly in predatory birds as a result of eggshell thinning. DDT and its metabolites have also been associated with infertility and embryo- and fetotoxicity. These effects may be related to the estrogenic activity of *o,p'*-DDT and *o,p'*-DDE (Bustos *et al.*, 1996).

Other Organochlorine Pesticides. A number of other organochlorine pesticides included in this study

Table 4. Concentrations of other selected organochlorine pesticides found in the sediments in southwest Puerto Rico.

Compound(s)	Sites	Concentration (ng/g)	NS&T Median (ng/g)	NS&T 85th Percentile (ng/g)
Total dieldrin	HTOC 17	0.41	0.30	1.90
	HTOC 19	0.42		
	HTOC 22	0.14		
Endrin	Coral 4	0.06	0.00	0.06
	Coral 6	0.16		
	HTOC 6	0.27		
	HTOC 8	0.06		
	HTOC 9	0.13		
Total chlordane	HTOC 17	2.54	0.51	3.10
	HTOC 19	1.08		
Lindane (γ -HCH)	Coral 8	0.69	0.00	0.82

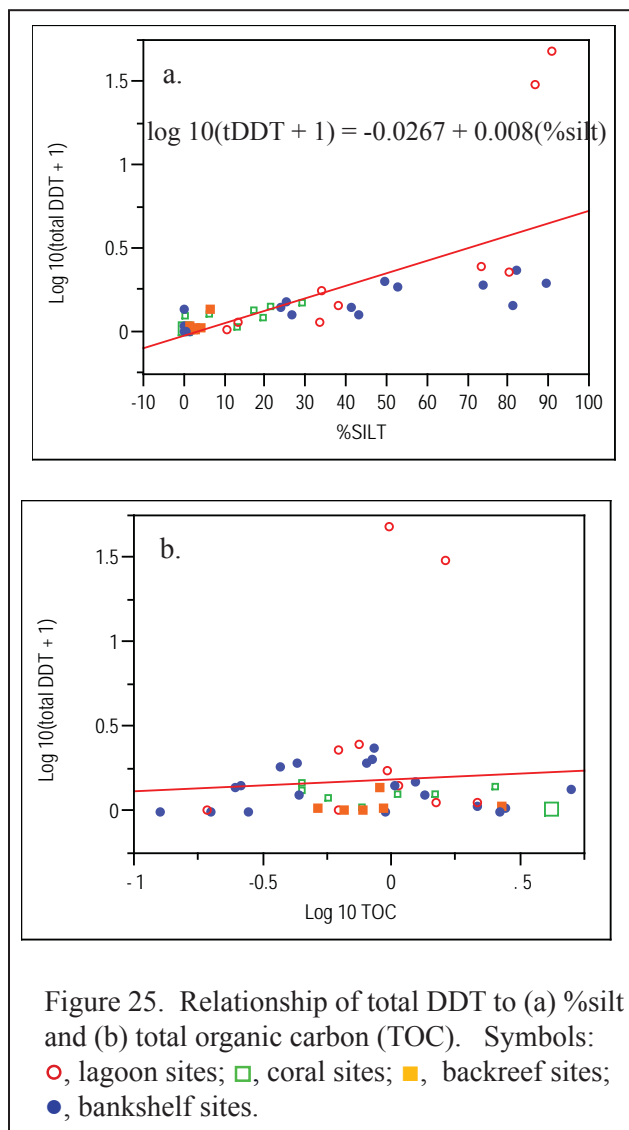


Figure 25. Relationship of total DDT to (a) %silt and (b) total organic carbon (TOC). Symbols: \circ , lagoon sites; \square , coral sites; \blacksquare , backreef sites; \bullet , bankshelf sites.

were also detected in the sediments in southwest Puerto Rico (Table 4, Appendix G), including total dieldrin (sum of dieldrin and aldrin), endrin, and total chlordane (alpha-chlordane, gamma-chlordane, heptachlor, heptachlorepoide, oxychlordane, *trans*-nonachlor, and *cis*-nonachlor). None of these compounds or

compound classes currently have ERL or ERM values. Most concentrations were below the NS&T national median (Table 4). A few sites, particularly the two in Guanica Bay (HTOC 17 and HTOC 19) had sediment concentrations above the median for total dieldrin, endrin, and total chlordane. Three sites, (Coral 6, HTOC 6 and HTOC 9) had endrin concentrations above the NS&T 85th percentile.

Concentrations of these same pesticides in almost all of the water samples collected by the PREQB were below the detection limit or not detected. Dieldrin was detected once by the PREQB at a site upstream of Guanica Bay at a concentration of 0.6 µg/L.

The chlorinated insecticide endosulfan is currently used in agriculture on a variety of vegetable and fruit crops. Endosulfan is a restricted use pesticide, meaning that it can only be applied by a certified applicator. In the past, endosulfan was classified as a general use pesticide, and could be used by homeowners to control insect pests. A map of endosulfan detections is shown in Figure 26. It is interesting to note that while there were detections of endosulfan adjacent to La Parguera, there were no endosulfan detections in Guanica Bay. Although there were only two sites sampled in Guanica Bay, the results may indicate that endosulfan was not a pesticide commonly applied agriculturally in the Lajas Valley.

The NCA Program measured a sediment concentration of 0.43 ng/g total endosulfan at Bahia Montalva. The sediment concentration of total endosulfan in the sediment sample at LTOC

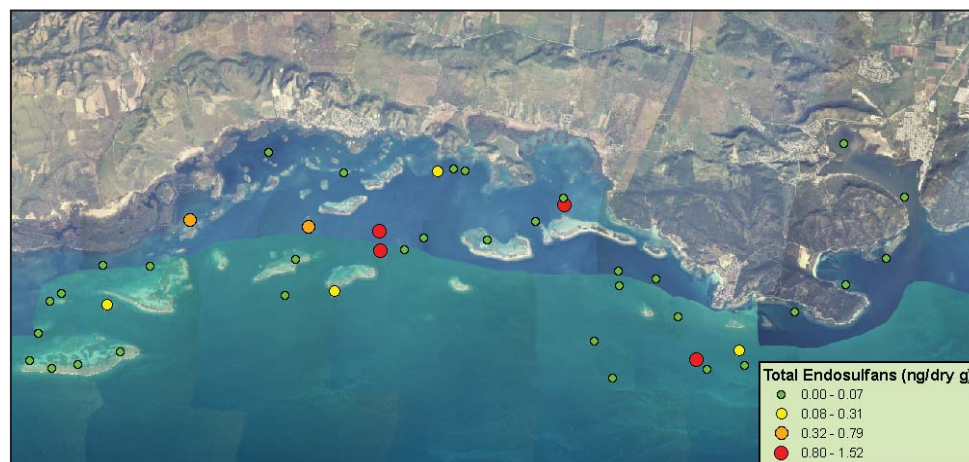


Figure 26. Map of endosulfan detected in the sediments in southwest Puerto Rico. Data classified into quartiles.

15 was 1.32 ng/g, and at LTOC 22 endosulfan was not detected. Another currently used pesticide, the chlorinated organophosphate insecticide chlorpyrifos was detected in the sediments at one site in Guanica Bay (HTOC 19), at a concentration of 0.60 ng/g.

Effects of Other Chlorinated Pesticides on Corals.

The only data found on the effects of other chlorinated pesticides analyzed in this project on corals was for chlorpyrifos. Acevedo (1991) exposed larvae of the coral *P. damicornis* to concentrations of chlorpyrifos ranging from 10 to 10,000 µg/L for up to 96 hours. Chlorpyrifos concentrations of 10 µg/L had no noticeable adverse effects on the coral larvae. A chlorpyrifos concentration of 100 µg/L slowed the swimming of larvae. A concentration of 1,000 µg/L killed 50 to 100 percent of the larvae within 24 hours. At 10,000 µg/L, all larvae were killed within 12 hours. Acevedo (1991) also tested two other compounds, the insecticide carbaryl, and 1-naphtho, a degradation product of carbaryl. Both compounds were found to have no noticeable effects on the larvae of *P. damicornis* after 24 hours at concentrations up to 10,000 µg/L (Acevedo, 1991).

Other Effects of Chlorinated Pesticides. Most chlorinated pesticides have the ability to bioaccumulate in aquatic and terrestrial animals. Accumulation typically occurs in the adipose tissues. Like DDT, many of the other organochlorine pesticides affect the central nervous system. In

fish, the signs of chlordane poisoning include hyperexcitability, increased respiration rate, erratic swimming, and loss of equilibrium (Eisler, 1990). Chlordane has been shown to adversely affect sensitive vertebrate and invertebrate species at fairly low concentrations. Reduced survival has been measured in shrimp and crabs at water concentrations of 0.2 to 2.0 µg/L, and in shrimp and fish, immobilization, impaired reproduction, and histopathologies resulted from exposures ranging between 0.8 and 3.0 µg/L (Eisler, 1990).

Polybrominated Diphenyl Ethers

The concentrations of total PBDEs (sum of 38 congeners) found in the sediments in southwest Puerto Rico ranged from 0 to 7 ng/g, and are shown in Figure 27 and 28, and in Appendix H. Polybrominated diphenyl ethers or PBDEs belong to a group of flame retardant chemicals used in products ranging from foam cushions in furniture to the plastics in televisions and computers. The use of PBDEs results in a slower ignition time and rate of fire growth, providing additional time to escape or extinguish the fire. PBDEs have recently come under scrutiny, however, because of increasing concentrations of these compounds in the environment and evidence of immunological, endocrine and hepatic effects (USEPA, 2006).

PBDE Concentrations. The concentrations of total PBDEs found in the sediment samples were low, the average concentration was 0.54 ng/g. Total PBDEs were fairly evenly distributed across the study area (Figure 27), with a spike in Guanica Bay at HTOC 17 (7.2 ng/g). It is interesting to note that around La Parguera, slightly higher concentrations of total PBDEs were found offshore, in the areas around the emergent reefs as opposed to higher concentrations nearshore, the pattern found for a number of other contaminant classes.

The analysis of PBDEs within the NS&T Program is a new activity. As a result, NS&T national median and 85th percentile values have not been established for PBDEs in sediments. In addition, ERL and ERM values have not been developed for either total PBDEs or for any of the congeners.

In the U.S. there is one manufacturer of PBDEs. The commercial PBDE products, analogous to the Aroclors, contain mixtures of various congeners. Theoretically, 209 congeners could be included, but a much smaller number are actually found in the PBDE-containing products sold in the U.S. Three formulations of commercial PBDE products are produced in the U.S., and are referred to as pentabromodiphenyl ether (pentaBDE), octabromodiphenyl ether (octaBDE), and decabromodiphenyl ether (decaBDE). Each commercial product contains a mixture of the various congeners. For example, the pentaBDE is composed of tetraBDEs (24-38%), pentaBDEs (50-62%), hexaBDEs (4-12%) and triBDEs (0-1%) (USEPA, 2006).

Figure 29 presents the results of the analysis by class of PBDE (*i.e.*, mono to hepta PBDEs). It can be seen that the diBDEs contributed significantly to the PBDEs detected, followed by the tetra- and pentaBDEs. The highest diBDE detected in a sample was BDE 11 (3,3'-DiBDE), at a concentration of 7 ppb at HTOC 17 in Guanica Bay (Appendix H). A number of other sites also had detections of this congener, but at lower concentrations. Most of the remaining PBDEs found in the sediment samples were the tetra and penta congeners. The penta- and octaBDE commercial products are made up primarily of the tetra, penta, and hepta congeners. The diBDE congener is not a component of the mixtures produced and used in the U.S. The presence of this congener in the samples could be the result of the breakdown of some of the higher brominated PBDEs, or perhaps diBDE is used in the area.

No other data sources were located for concentrations of PBDEs in the study area. In the U.S., the majority (74%) of the commercial PBDE product used is actually the decaBDE mixture (USEPA, 2006). The decaBDE formulation contains deca congeners (10 bromines) almost exclusively. Unfortunately, the sediment analysis for this project did not include the decaBDEs.

Effects of PBDEs on Corals. No data on the effects of PBDEs on corals was found in the literature

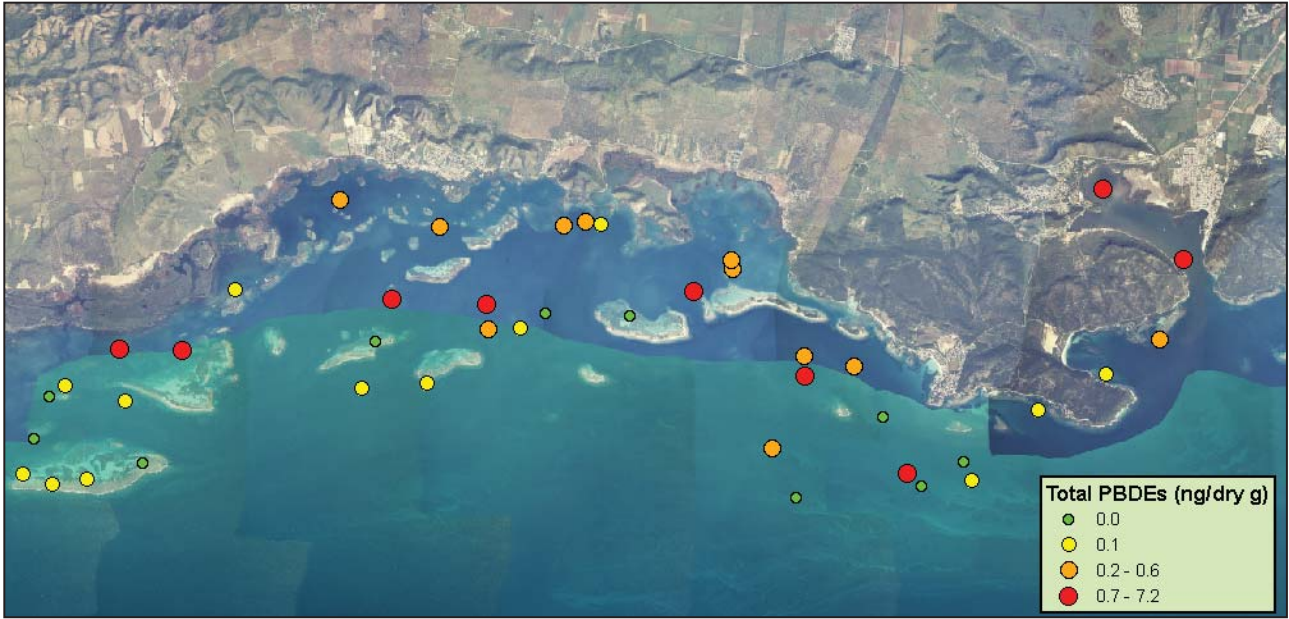


Figure 27. Map of total PBDEs detected by site in the sediments in southwest Puerto Rico. Data classified into quartiles.

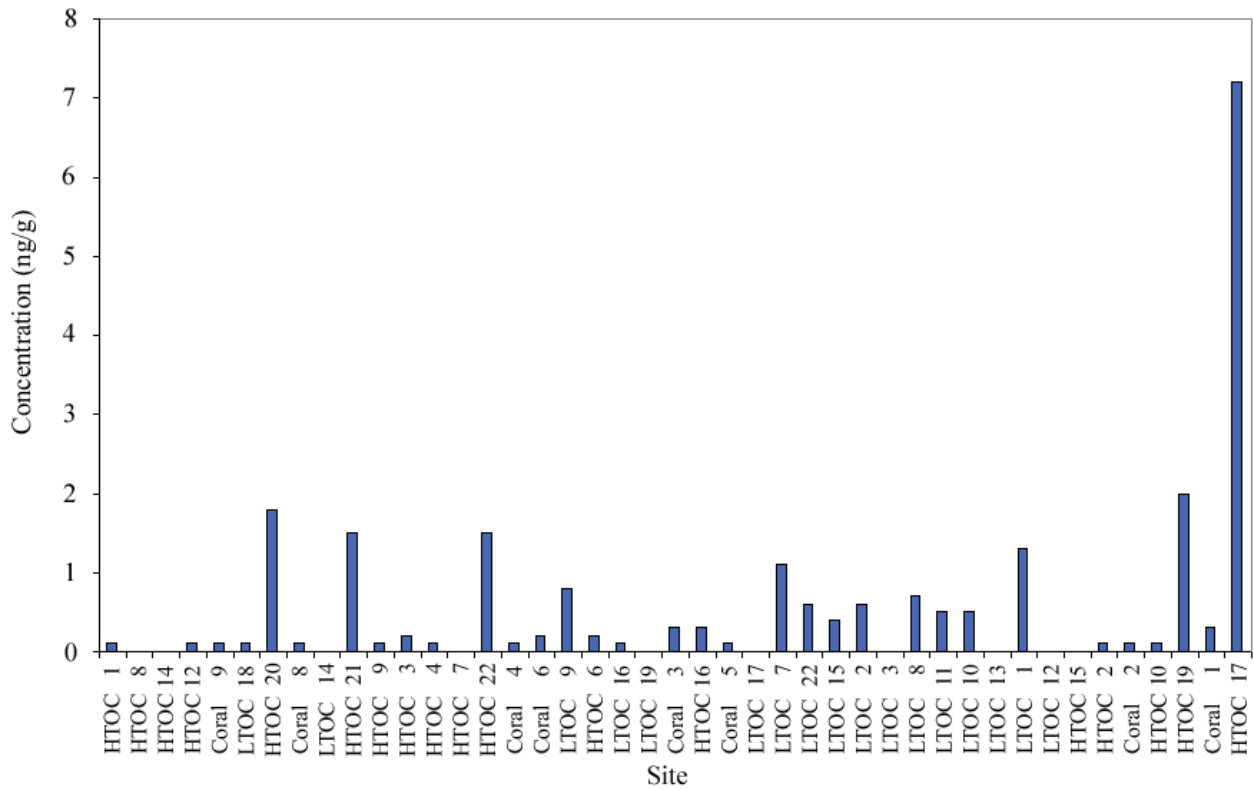


Figure 28. Total PBDEs detected by site in the sediments in southwest Puerto Rico.

search. PBDEs have been found in aquatic and terrestrial organisms, and as a result could also be found in coral tissues.

Other Effects of PBDEs.

There is a significant and growing body of work on the effects of PBDEs, primarily for vertebrates. The increasing concentrations of PBDEs in human breast milk, from nearly zero in 1985 to over 200 ng/g lipid weight recently, is seen as an alarming trend and a cause for more effective control of their manufacture and use (McDonald, 2002).

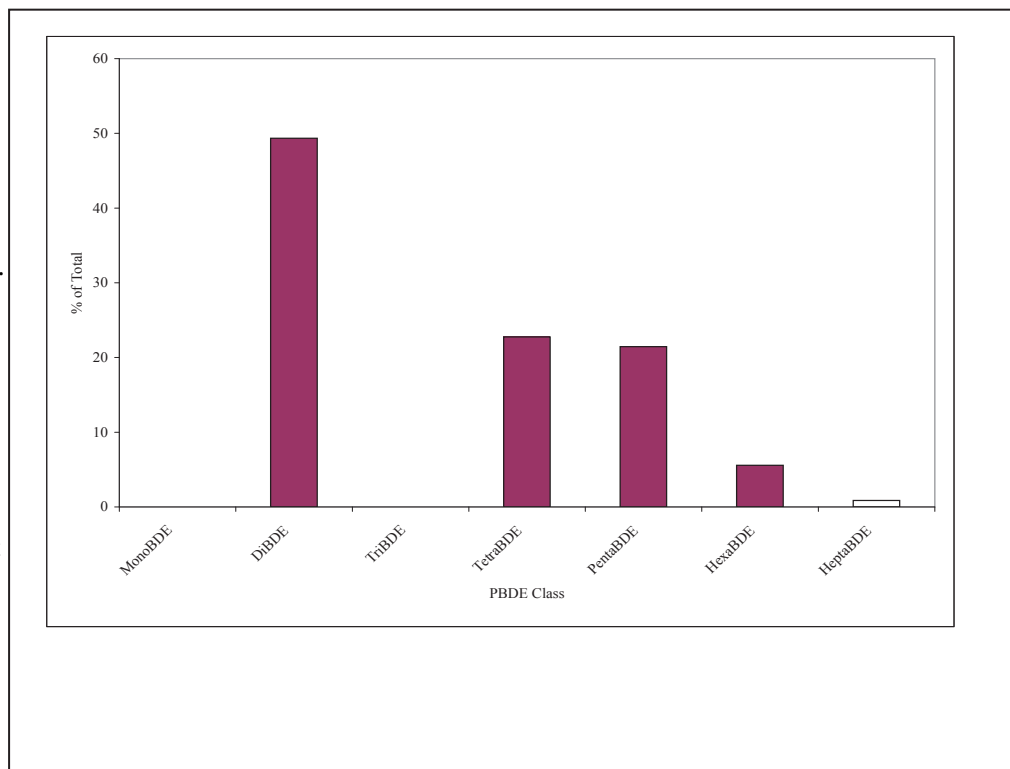


Figure 29. Detections of sediment PBDEs in southwest Puerto Rico by class.

Much of the work to date on the effects of PBDEs has focused on the penta congeners. Fowles *et al.* (1994) found that pentaBDE mixtures resulted in suppressed antibody production in mice, and Thuvander and Darnerud (1999) found decreased immunoglobulin production, also in mice. Developmental effects have also been found in mice exposed in the laboratory to PBDEs (Branchi *et al.*, 2002).

It is also now fairly well established that due to their structural similarity to the hormone thyroxine, PBDEs have the ability to mimic thyroid hormones, and PBDEs have been associated with a number of health consequences including goiters, benign and neoplastic thyroid diseases, and impaired neural development. Zhou *et al.* (2001) found effects on the thyroid system, mainly reduced serum levels of thyroxine in rats, following exposure to pentaBDE. Meerts *et al.* (2001) found evidence of estrogenic effects for 11 PBDE congeners and 2 hydroxylated metabolites. Stoker *et al.* (2004) found evidence of anti-androgenic activity in rats.

Butyltins

The results of the analyses of butyltins are shown in Figure 30 and in Appendix I. Mono-, di-, tri-, and tetrabutyltins were analyzed in the sediments. In the 1950s, tributyltin or TBT was first shown to have biocidal properties (Bennett, 1996). Beginning in the late 1960s, TBT was incorporated into a very effective antifoulant paint system, quickly becoming one of the most effective paints ever used on boat hulls (Birchenough *et al.*, 2002). TBT was incorporated into a polymer paint system that released the biocide at a constant and minimal rate, to control fouling organisms such as barnacles, mussels, weeds, and algae (Bennett, 1996).

In the aquatic environment, TBT is degraded by microorganisms and sunlight (Bennett, 1996). The transformation involves sequential debutylation resulting in dibutyltin, monobutyltin, and finally inorganic tin (Batley, 1996). Experiments have shown that the half-life of TBT, the amount of time needed to convert half of the TBT to dibutyltin

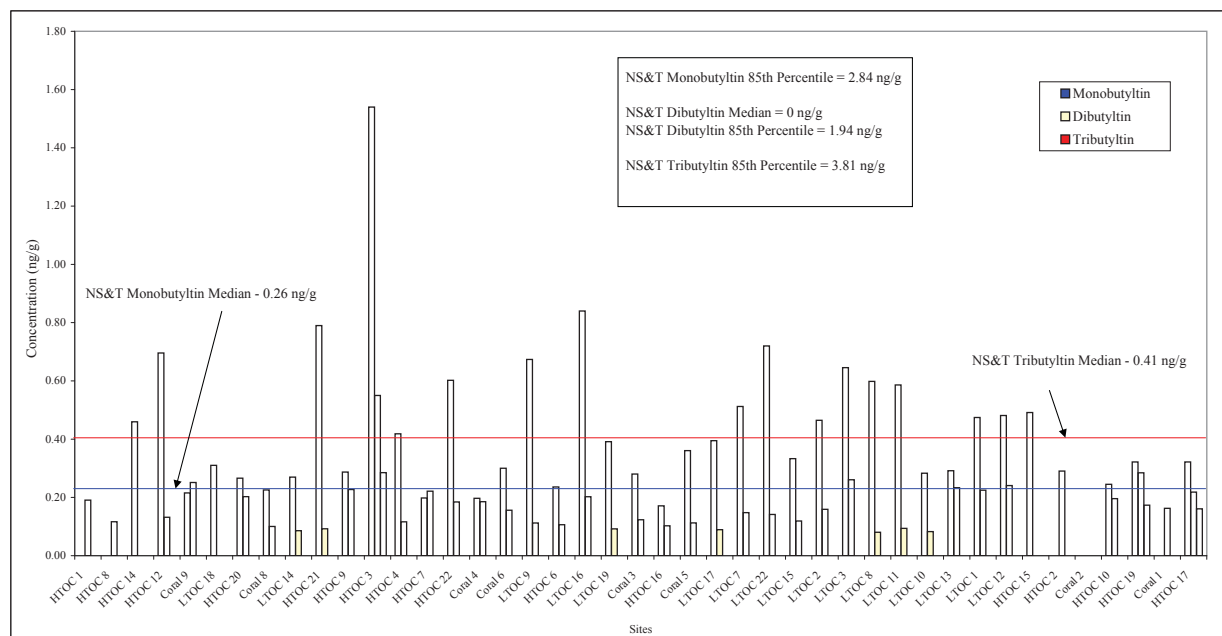


Figure 30. Butyltins detected in the sediments in southwest Puerto Rico.

in natural water samples, is on the order of days; degradation to monobutyltin takes approximately a month (Batley, 1996). Experiments with aerobic sediments have shown that the half-life of TBT is similar to that measured in solution. In deeper, anoxic sediments, however, the half life of TBT appears to be considerably longer, on the order of 2-4 years (Batley, 1996).

There were few detections of TBT in the sediments sampled in southwest Puerto Rico (Figure 30). Detectable TBT was only found at three sites, two of which were in Guanica Bay. At all three sites, TBT concentrations were below the NS&T national median. In the sediment samples, monobutyltin was the dominant butyltin found, and is likely the result of TBT debutylization. No samples in the study area had concentrations of dibutyltin or monobutyltin above the NS&T 85th percentile. Several sites, however, had sediment concentrations above the NS&T national median for monobutyltin (0.26 ng/g). Tetrabutyltin, an impurity in the manufacture of tributyltin oxide, was not detected in any of the sediments. No other data sources for TBT in the study area were found.

Effects of TBT on Corals. Negri and Heyward (2001) investigated the effects of tributyltin on fertilization and larval settling of *Acropora millepora*. Gametes were collected from reefs in Western Australia, and brought back to the laboratory. The effective concentration of TBT which caused 50 percent inhibition (EC_{50}) of fertilization after four hours was 200 $\mu\text{g/L}$. A second set of experiments was carried out to assess the effects on settlement and metamorphosis of larval *A. millepora*. The calculated concentration needed to inhibit 50 percent larval metamorphosis was low, only 2 $\mu\text{g/L}$.

Negri *et al.* (2002) investigated the effects of contaminants in sediments on the coral *Acropora microphthalma*. This was the only work located that directly assessed the effects of contaminants in sediments on corals. Sediments were collected from an area where a cargo ship had grounded on the Great Barrier Reef in Australia. The grounding of the ship and subsequent wave action resulted in TBT sediment concentrations of approximately 160 $\mu\text{g/g}$, copper concentrations of 1,180 $\mu\text{g/g}$, and zinc concentrations of 1,570 $\mu\text{g/g}$. Negri *et al.* (2002) diluted the sediments taken at the site of the grounding with sediments from a control site. Sediments diluted

to 5 percent of the original concentration prevented successful settlement of coral larvae in the laboratory on preconditioned terracotta tiles. The tiles were preconditioned by placing the tiles on control reefs for three months.

Other Effects of TBT. Beginning in the 1970s, a number of effects related to TBT exposure became apparent in the environment, particularly in gastropods and bivalves. In 1971, the term “imposex” was first used to describe the occurrence of male characteristics in the intertidal snail *Ilyanassa obsoleta* (Gibbs and Bryan, 1996). In the 1980s, TBT was implicated in the near collapse of oyster farming in western France and southern England, and in the dogwhelk *Nucella lapillus* in areas of high shipping and boating activity in southwest England. In North America, sterilization of the gastropods *Nucella emarginata*, *Nucella lamellosa* (northwest coast) and *I. obsoleta* (northeast coast) was found. TBT has also been shown to cause shell deformation in the Pacific oyster *Crassostrea gigas* in the U.S., along with abnormal development of larvae (Peachey, 2003).



An image of symmetrical brain coral, *Diploria strigosa*.

France enacted a ban on TBT in 1982. In the U.S., the Organotin Antifouling Paint Control Act of 1988 called for a ban on the use of tributyltin-based paints on vessels 25 meters in length or less (except for aluminum vessels). A number of other countries, primarily in Europe and Australia, adopted regulations prohibiting the use of TBT shortly thereafter.

The physiological and biochemical mechanisms of TBT are poorly understood, but in gastropods the mode of action appears to be related to effects on the endocrine system. Observations with *N. lapillus* have suggested that TBT elevates levels of testosterone, which may result from an inhibition of the Cytochrome P-450 dependant enzyme aromatase, responsible for the conversion of testosterone to estradiol (Gibbs and Bryan, 1996).

Trace Elements

Sixteen major and trace elements were analyzed in the sediment samples collected. The results of the analyses are contained in Appendix J. An assessment of arsenic, chromium, nickel and copper follows.

Arsenic. The results of the sediment analysis are shown in Figures 31 and 32. Arsenic concentrations were low. Only one site, Coral 1 (Figure 32), just outside of Guanica Bay, had a concentration above the NS&T national median of 6.8 $\mu\text{g/g}$. The concentration of arsenic in sediments at Coral 1 (10.3 $\mu\text{g/g}$) was also somewhat higher than the ERL value of 8.2, but appreciably below the ERM value. A nonparametric analysis indicated no significant relationship between arsenic concentration and

habitat, and unlike a number of other chemical contaminants, there was no relationship between arsenic and grain size.

No data were available from the PREQB for arsenic. In 2000, EPA's NCA Program sampled sediments in Bahia Montalva. The concentration of arsenic at this site was 5 $\mu\text{g/g}$, similar to the 7.7 $\mu\text{g/g}$ and 5.84 $\mu\text{g/g}$ found at LTOC 22 and LTOC 15, respectively.

Figure 31 shows that the spatial pattern for arsenic in the sediments is somewhat different from that of the organic compounds and as will be seen, for a number of the other trace elements. In this study, elevated levels of chemical contaminants were typically found nearshore, adjacent to the town of La Parguera, and in Guanica Bay. This pattern suggests, at least for the organic compounds, that the contamination is associated with anthropogenic activities (*e.g.*, land-based or nearshore marine). However, the elevated levels of arsenic (Figure 31) were outside of Guanica Bay and to the east of La Parguera.

Because trace elements like arsenic occur naturally in the Earth's crust, their presence in the environment



Figure 31. Map of arsenic detected in the sediments in southwest Puerto Rico. Data classified into quartiles.

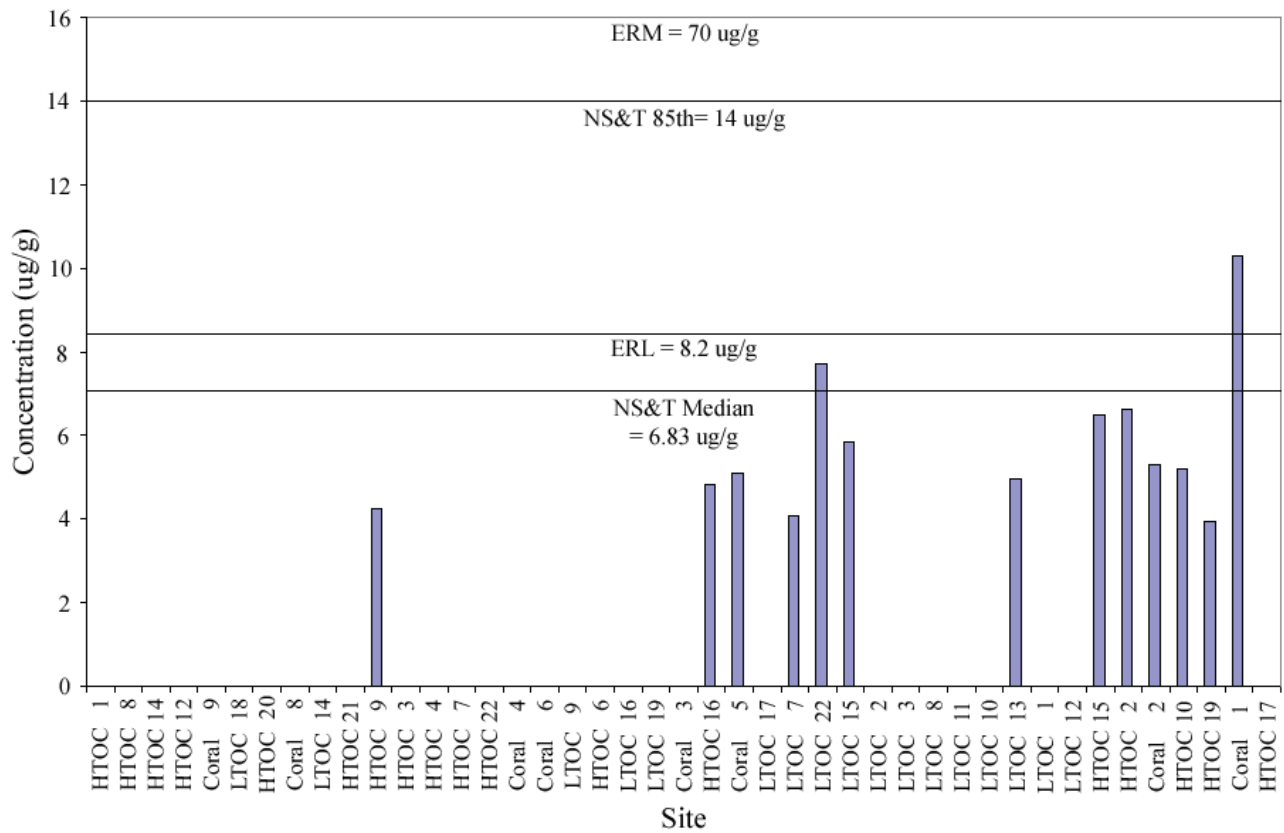


Figure 32. Arsenic detected by site in the sediments in southwest Puerto Rico.

can be the result of both natural and anthropogenic inputs. Valette-Silver *et al.* (1999) hypothesized that arsenic contained in phosphatic rock was a significant source contributing to elevated arsenic concentrations in both sediments and bivalve tissues in the southeast U.S. The results presented here for southwest Puerto Rico do not appear to indicate a large anthropogenic source for arsenic, either around the town of La Parguera or in Guanica Bay. The presence of slightly higher arsenic concentrations outside of these populated areas could be the result, in part, of natural processes. Additional work would be needed to assess this possibility.

Effects of Arsenic on Corals. No data were located on the effects of arsenic on corals.

Other Effects of Arsenic. Arsenic is a widely distributed element occurring most commonly in the environment as inorganic and organic combinations in soil, sediments, water and air. There has been a significant amount of work to assess the effects of arsenic on other aquatic organisms.

Production of offspring in the opossum shrimp (*Americamysis bahia*) was affected by a total arsenic concentration of 1,270 $\mu\text{g/L}$ (Lussier *et al.*, 1985). In rainbow trout (*Salmo gairdneri*), fish fed diets containing 120 mg As/kg avoided food, grew slower than the controls, and did not metabolize ingested food as efficiently (Eisler, 1988). Arsenic does have the potential for accumulation in the tissues of fish and shellfish, but usually in the less harmful arsenobetaine form. Arsenobetaine, containing three methyl groups bonded to the arsenic atom, results from the reduction and methylation of arsenic.

Chromium. Concentrations of chromium detected in the sediments are shown in Figures 33 and

34. The spatial pattern found for chromium in the sediments shows elevated levels adjacent to the town of La Parguera and at the two sites in Guanica Bay. This pattern is similar to a number of the organic compound classes analyzed in this project.

With the exception of Guanica Bay, chromium levels in the sediments were below the NS&T national median of 55 $\mu\text{g/g}$, and also below the ERL value of 81 $\mu\text{g/g}$. The PREQB detected a concentration of 11 $\mu\text{g/L}$ in a water sample north of Guanica Bay. EPA's NCA Program detected a concentration

of 23 $\mu\text{g/g}$ in the sediments in Bahia Montalva, similar to the 31 $\mu\text{g/g}$ and 30.4 $\mu\text{g/g}$ concentrations detected at LTOC 15 and LTOC 22 (Appendix J), respectively. In Guanica Bay, however, chromium levels were not only above the ERL value, but also above the ERM value of 370 $\mu\text{g/g}$, indicating that effects are more likely as a result of chromium contamination at these sites.



*Staghorn coral (*Acropora cervicornis*), recently listed as threatened under the Endangered Species Act. Image courtesy of NOAA CCMA Biogeography Branch.*

A nonparametric analysis (Wilcoxon Test) indicated a significant association between habitat and chromium concentrations ($P < 0.0005$, ChiSquare = 17.8641). Similar to a number of organic chemical contaminants, trace elements are often associated with the silt and clay fractions of the sediments. Figure 35 shows the relationship between chromium levels and the silt and clay fractions of the sediment. A nonparametric analysis of the data indicated that both the silt ($P < 0.0001$, Spearman Rho = 0.7259) and the clay ($P < 0.0001$, Spearman Rho = 0.6354) fractions were positively correlated with chromium in the sediments. Rieuwerts *et al.* (1998) noted that metal accumulation in these size fractions is

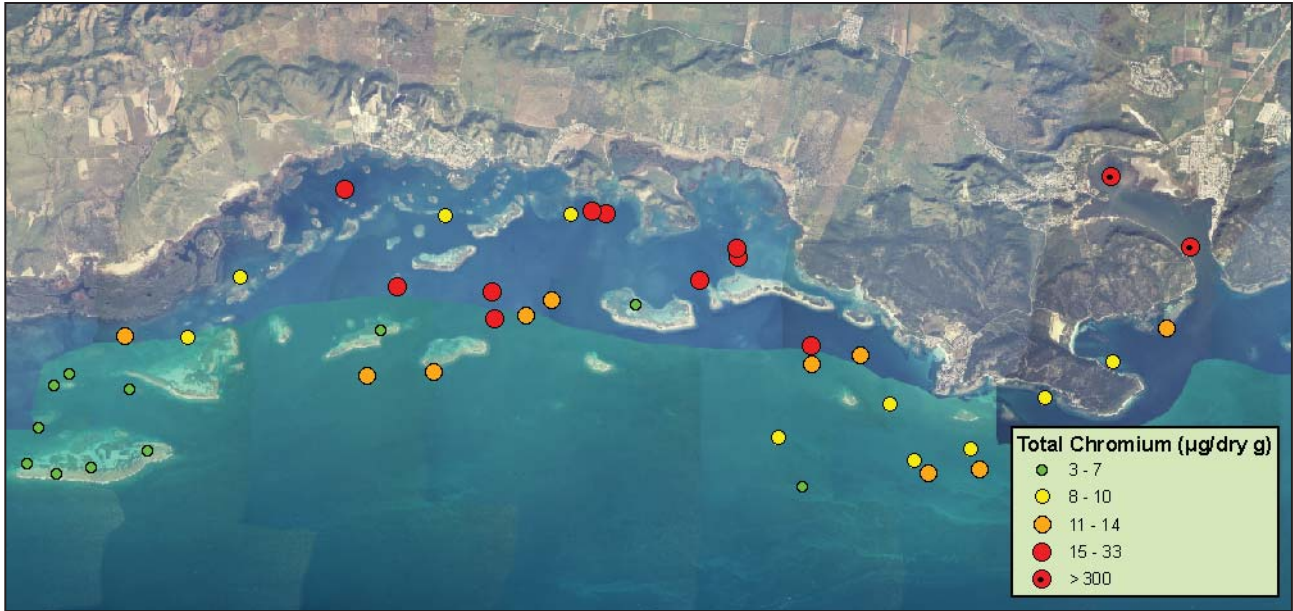


Figure 33. Map of chromium detected in the sediments in southwest Puerto Rico. Data classified into quantiles.

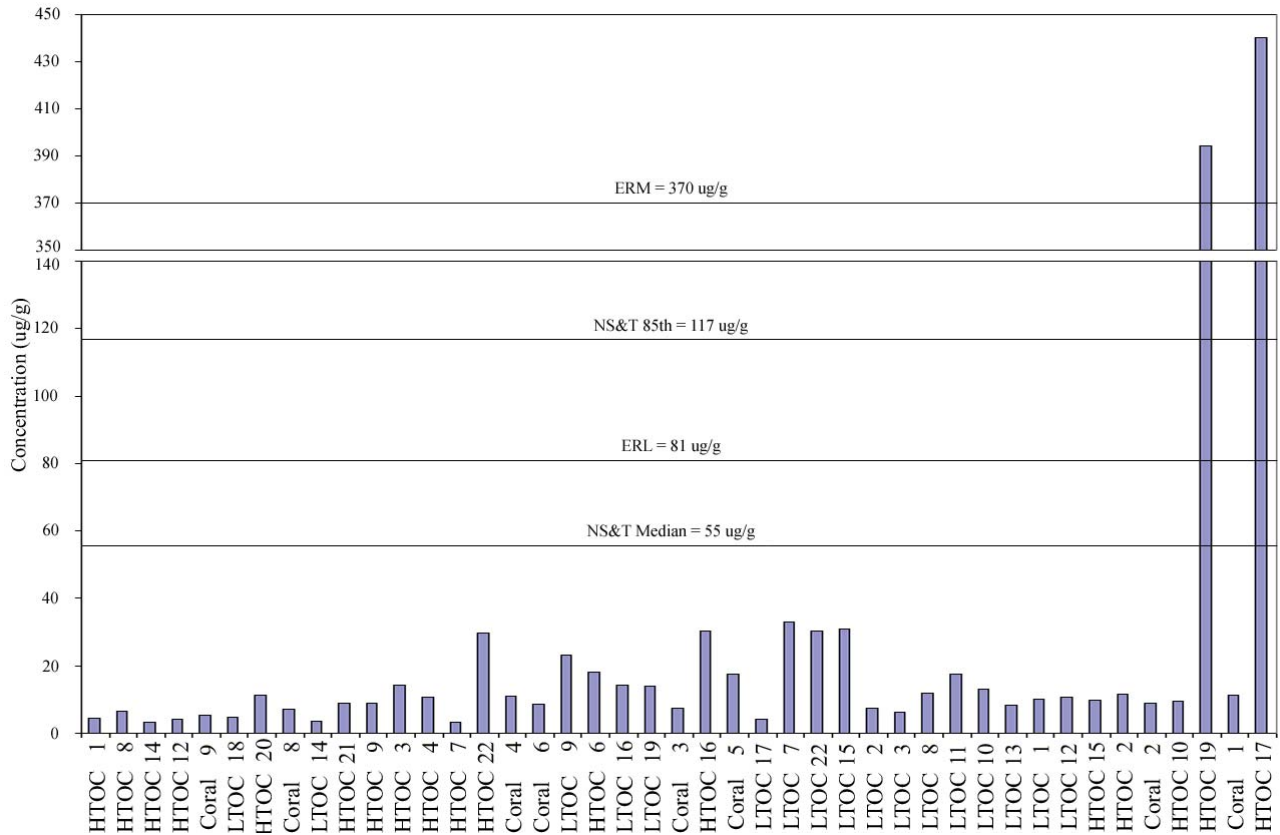


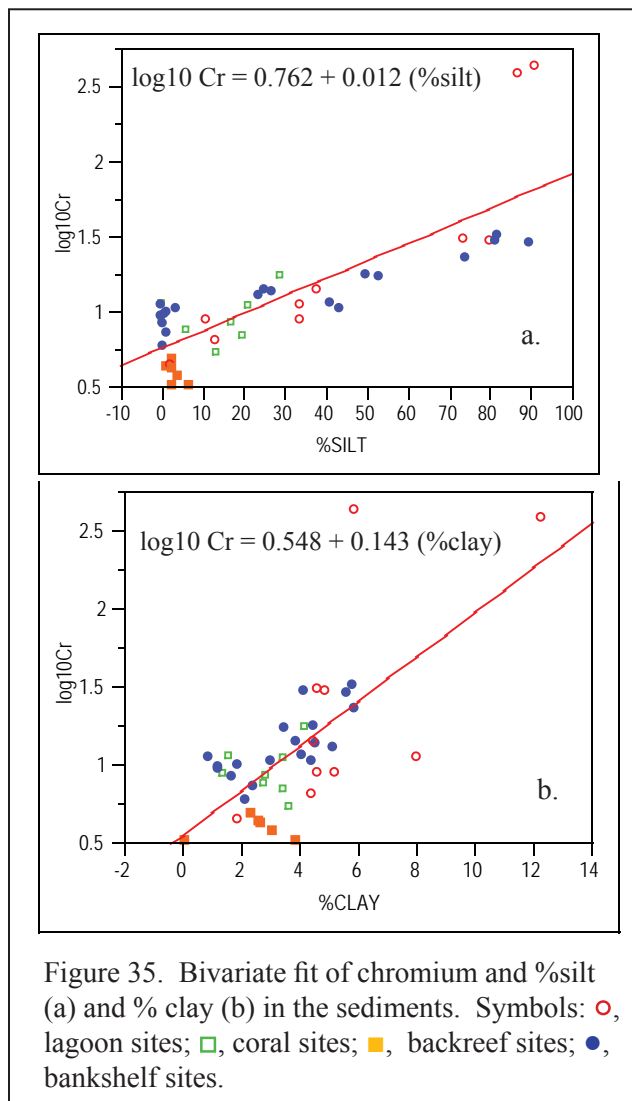
Figure 34. Chromium detected by site in the sediments in southwest Puerto Rico.

a function of the high relative surface areas for adsorption. In addition, metal ions are absorbed through both ion exchange and specific adsorption processes (Rieuwerts *et al.*, 1998).

Chromium, like other metals is a component of the Earth's crust. Its presence in the marine environment can be the result of natural crustal weathering in addition to the anthropogenic inputs. Much of the Lajas Valley, which has had significant agricultural activity over the years, drains into Guanica Bay and is likely a source of metals to the Bay.

Chromium also has a number of uses in industry, such as in stainless steel, chrome plating, and as a catalyst. In Guanica Bay, there have been various industrial activities over the years, ranging from the production of sugar to fertilizer mixing to textile manufacture. Inorganic fertilizers can contain certain trace elements, including cadmium, arsenic, copper, chromium and lead (Mortvedt, 1996). Elevated levels of cadmium in phosphatic rock, used in the manufacture of inorganic fertilizers, have been cited as a concern due to potential effects on human health (Smil, 1999; Mortvedt, 1996). At the two sites sampled in Guanica Bay, however, cadmium levels were low (Appendix J) in comparison to the amount of chromium in the sediments. Chromium is frequently used along with nickel and sometimes manganese in the production of stainless steel. Chromium can constitute up to 18 percent of the chemical composition of stainless steel (Wallinder *et al.*, 2004). Although only two sediment samples were analyzed from Guanica Bay, the levels of chromium, and also nickel (next section) at both of these sites were over an order of magnitude higher than at any of the other sites sampled in this study. Manganese, which is also a fairly common component in stainless steel formulations, was elevated at the two sites in Guanica Bay as well (Appendix J).

Stainless steel was first manufactured and used in the early part of the 20th century. Chromium and nickel have both been shown to leach out of stainless steel as a result of industrial use or environmental (*e.g.*, weathering) processes (Wallinder *et al.*, 2004). One of the common industrial uses of stainless steel is in the machinery used in the refining of sugar. The



elevated levels of chromium, nickel, manganese, and also iron in the sediments sampled in Guanica Bay (Appendix J), could be related to the use of stainless steel in the production of sugar, or other industrial activities that have occurred in the area around Guanica Bay. The production of sugar in the Guanica Bay area ceased in the early 1950s.

Effects of Chromium on Corals. No information was located on the effects of chromium on coral. This type of information would be useful for this study, and in other coral reef areas where chromium may be present at elevated levels.

Other Effects of Chromium. Chromium has a low bioaccumulation potential in the tissues of fish

and shellfish, but in the laboratory has been found to be mutagenic, carcinogenic and teratogenic (Eisler, 1986). Although chromium can exist in oxidation states from -2 to +6, it is most often found in the environment in either the trivalent (Cr^{+3}) or hexavalent (Cr^{+6}) oxidation states (Eisler, 1986). Hexavalent chromium appears to have the most biological activity. Hexavalent chromium at a concentration of 10 $\mu\text{g/L}$ over 32 days was associated with reduced survival and fecundity of the cladoceran *D. magna*. A number of species of fish have also shown sensitivity to chromium, particularly hexavalent chromium. Concentrations ranging from 16 to 21 $\mu\text{g/L}$ resulted in reduced growth in fingerlings of rainbow trout and chinook salmon during a 16 week exposure (USEPA, 1980a). As with the PAHs, the sediment concentrations of chromium found in the sediments in southwest Puerto Rico cannot be compared directly with the aqueous concentrations found to elicit effects in test organisms.

Nickel. The concentrations of nickel detected in the sediments are shown in Figures 36 and 37. Other than Guanica Bay, there were few detections above the NS&T national median. Four sites outside of Guanica Bay (LTOC 15, LTOC 22, HTOC 16, and HTOC 22) were very close but slightly below the ERL value of 21 $\mu\text{g/g}$. LTOC 7 was slightly above the ERL value. In Guanica Bay, however, both sites were substantially above the ERM value of 52 $\mu\text{g/g}$, indicating that impacts on biota were likely. EPA's NCA Program detected a concentration of 33 $\mu\text{g/g}$ nickel in a sediment sample from Bahia Montalva, similar to the 20.9 and 20.3 $\mu\text{g/g}$ values detected at LTOC 15 and LTOC 22 sites, respectively.

The regression of nickel with the silt and the clay fractions of the sediments is shown in Figure 38. Because the concentrations of nickel were not normally distributed, nonparametric analyses were run to assess the relationship between nickel and sediment particle size. As with chromium, the concentration of nickel was associated with both the silt ($P < 0.0001$, Spearman Rho = 0.8393) and clay ($P < 0.0001$, Spearman Rho = 0.7326) fractions of the sediments. These associations remained highly

significant even when the data from the two sites in Guanica Bay were excluded.

As noted, the association of metals with these sediment fractions are due to high surface areas, as well as ionic forces for the clay fraction. The silt and particularly the clay fractions have higher total surface areas providing sites for the adsorption of contaminants. The clays also have inherently negative charges within their layered structures allowing for cation exchange with various metals including nickel. For clays, both the surface area and ionic forces are important factors for the association with metals. There was also a significant negative correlation between the sand fraction of the sediment and nickel, while no correlation existed between percent gravel and nickel.

Nickel is also a metal found in crustal materials, and natural erosional processes can contribute to its presence in the marine environment. In addition, nickel has many applications in both industrial and consumer products. Approximately 65% of the nickel in the U.S. is used to make stainless steel. Other uses include its incorporation into a series of alloys, in rechargeable batteries, catalysts and other chemicals, coins, plating, and in foundry products. Nickel and chromium were elevated along with iron at the two sites sampled in Guanica Bay (Appendix J). As noted, all three metals are components of stainless steel, and the higher concentrations in Guanica Bay could be indicative of the use, particularly the industrial use, of stainless steel.

Effects of Nickel on Corals. Two articles were found on the effects of nickel in corals. Reichelt-Brushett and Harrison (2005) exposed the gametes of *Goniastrea aspera* to nickel concentrations ranging from 5 to 2,000 $\mu\text{g/L}$. Concentrations of nickel at or above 100 $\mu\text{g/L}$ resulted in decreased fertilization success. In another study, Goh (1991) exposed larvae of the coral *P. damicornis* to nickel. The larvae were exposed to concentrations ranging from 1 to 25 mg/L (ppm) for durations of 12 to 96 hours. After the exposure, the larvae were placed in natural filtered seawater, and settlement success was followed over a period of days. Goh (1991) found significantly lower settlement success of the larvae nine days after the

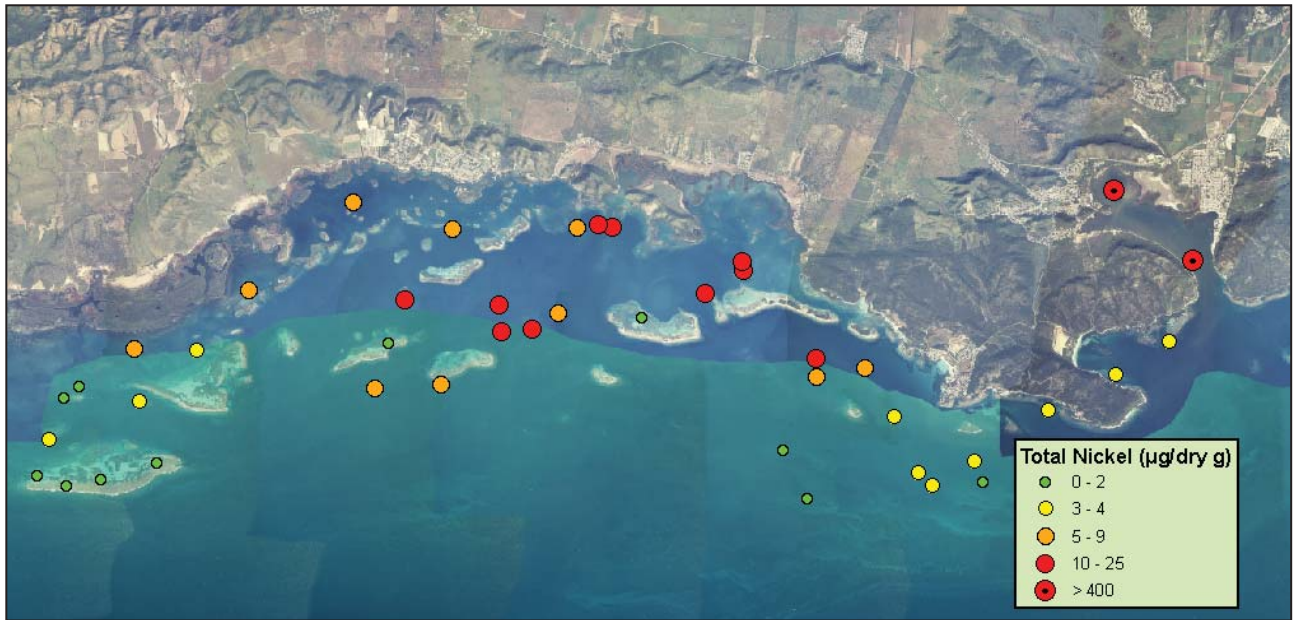


Figure 36. Map of nickel detected in the sediments in southwest Puerto Rico. Data classified into quantiles.

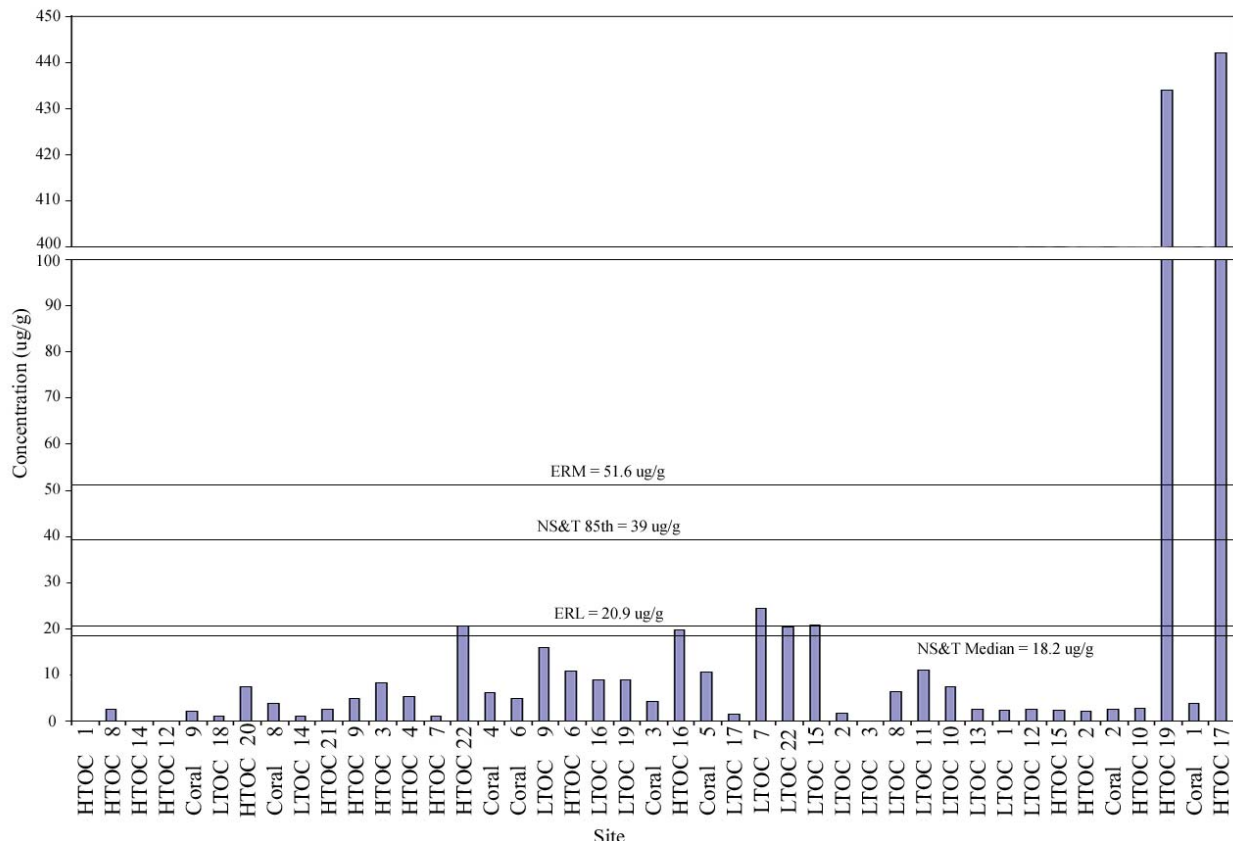


Figure 37. Nickel detected by site in the sediments in southwest Puerto Rico.

recovery period in all concentrations, regardless of the duration of the treatment.

Other Effects of Nickel. Nickel in the environment can come from a variety of sources including the use and disposal of the products mentioned above, the combustion of fuels in power plants, and waste material in incinerators. Nickel has a low potential for bioaccumulation in the tissues of fish and most mollusks. In the cladoceran *D. magna*, nickel concentrations as low as 30 µg/L have been reported to impair reproduction after 21 days, and at a concentration of 100 µg/L, growth was inhibited in this species after 9 days (USEPA, 1980b). In the zebrafish *Brachydanio rerio*, nickel concentrations greater than 40 µg/L delayed hatching time (Dave and Xiu, 1991).

Copper. The results of the analysis of copper are shown in Figures 39 and 40, and in Appendix J. The highest copper concentrations were found in the sediments at the two sites in Guanica Bay. There were some elevated levels of copper around La Parguera, but all were below the NS&T national median of 15 µg/g. In Guanica Bay, copper in the sediment samples from HTOC 17 was above the ERL of 34 µg/g. At HTOC 19, copper (80.6 µg/g) was above the NS&T 85th percentile, but below the ERM value of 270 µg/g. No data were located from the PREQB for copper, however, EPA's NCA Program detected copper at a concentration of 9 µg/g in the sediments at Bahia Montalva, similar to the 7.83 and 7.4 µg/g copper detected at LTOC 15 and LTOC 22, respectively.

In Figure 41, copper is plotted against the silt and clay fractions of the sediment. Nonparametric analyses indicated a strong positive relationship between copper and the silt ($P < 0.0001$, Spearman $Rho = 0.8835$) and clay ($P < 0.0001$, Spearman $Rho = 0.7879$) fractions of the sediment, similar to chromium and nickel. Copper has many applications, including its use as an antifoulant on boat hulls and in many industrial and consumer applications, including copper wire, electronic circuits, copper plumbing, cookware, coins and as industrial catalysts. Copper is also commonly used in agriculture as a fungicide on citrus crops. The higher levels of copper in the

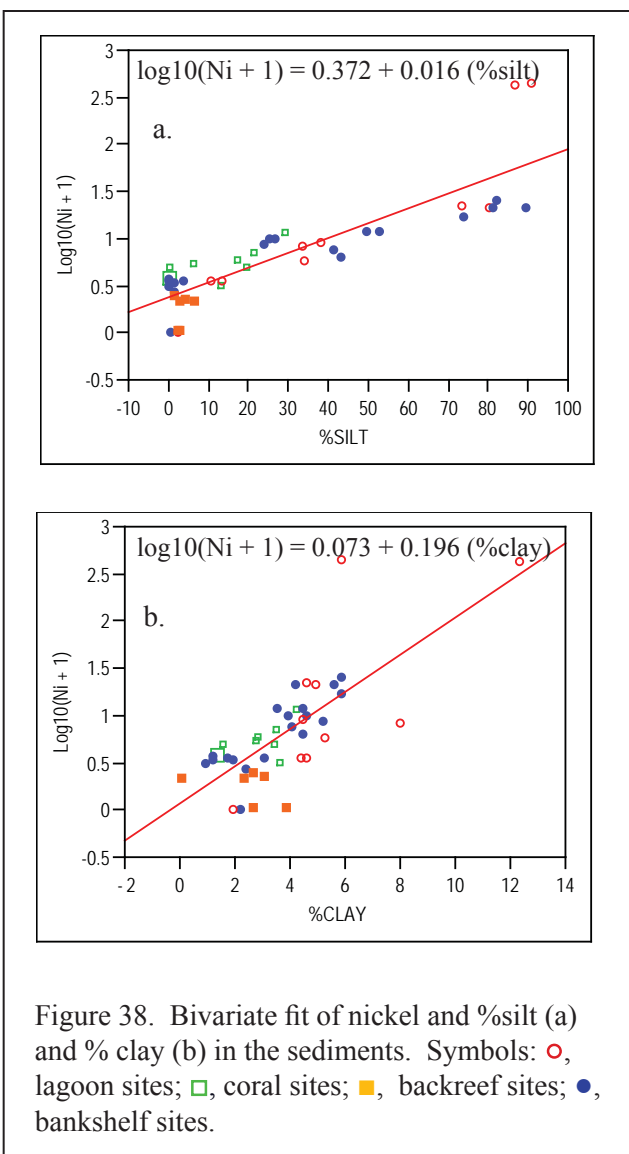


Figure 38. Bivariate fit of nickel and %silt (a) and % clay (b) in the sediments. Symbols: ○, lagoon sites; □, coral sites; ■, backreef sites; ●, bankshelf sites.

sediments in Guanica Bay may be the result of a combination of uses in industry, agriculture and as an antifoulant on boat hulls.

Normalization of Metals. As mentioned, the natural occurrence of metals in the Earth's crust and their appearance in the marine environment as a result of crustal weathering makes it more challenging to determine when elevated concentrations are due to anthropogenic activities versus normal weathering of rock (Weisberg *et al.*, 2000). One approach that has been taken to separate natural from anthropogenic contributions is to normalize the concentrations of metals such as chromium, nickel and copper to either aluminum or iron (Windom *et al.*, 1989; Daskalakis

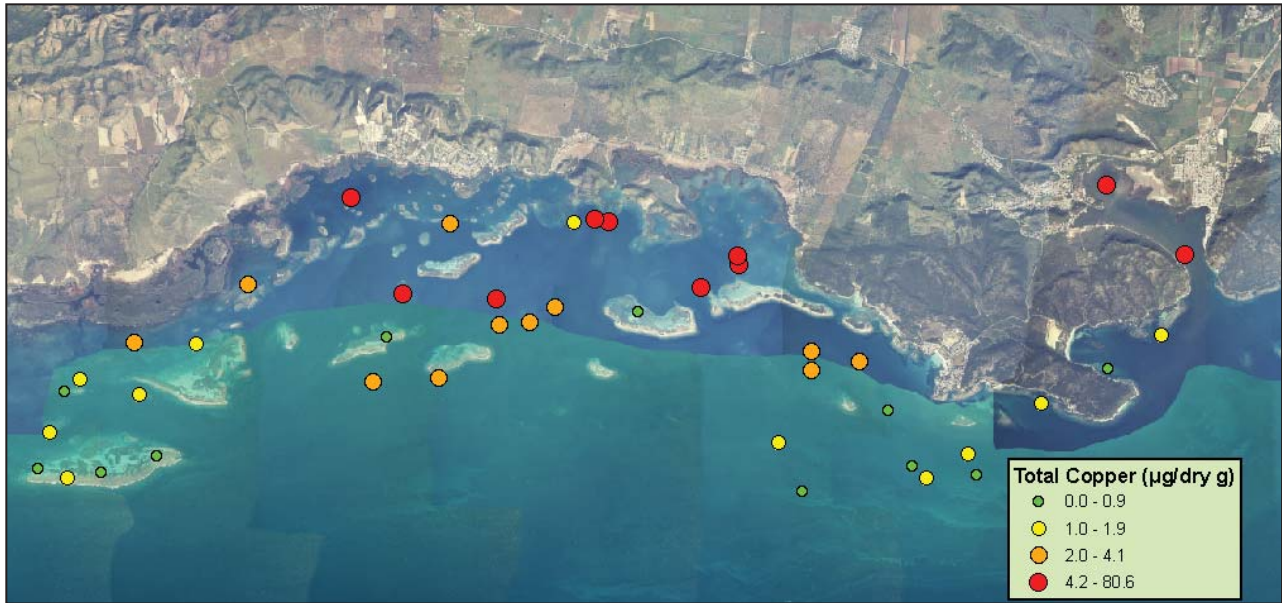


Figure 39. Map of copper detected in the sediments in southwest Puerto Rico. Data classified into quartiles.

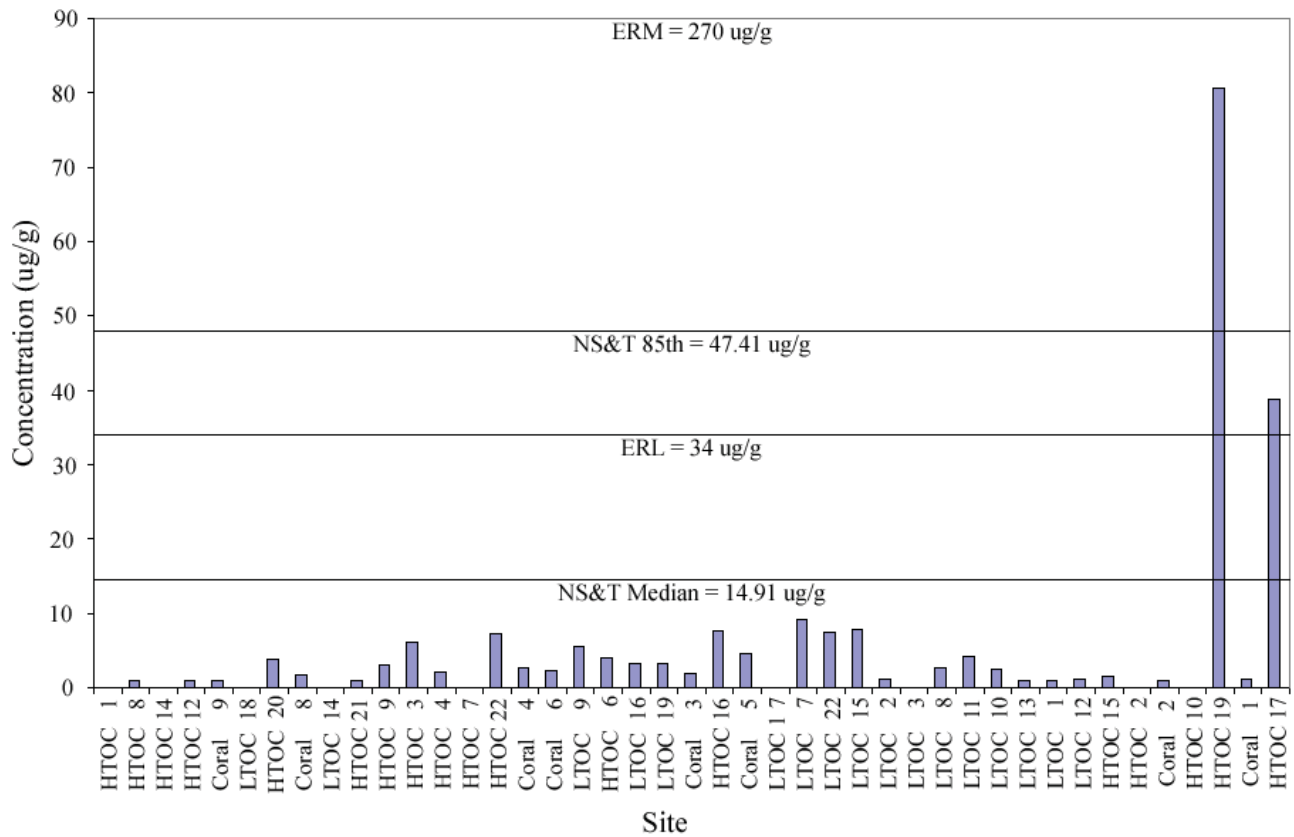


Figure 40. Copper detected by site in the sediments in southwest Puerto Rico.

and O'Connor, 1995). Aluminum and iron are treated as conservative tracers of natural metal-bearing sediments (Weisberg *et al.*, 2000).

For the sediments from southwest Puerto Rico, chromium, nickel, and copper were normalized to aluminum. Iron was not used as Daskalakis and O'Connor (1995) found that aluminum worked better on the East Coast of the U.S.

Regression analyses were first run to verify that there was a significant correlation between each of the metals and aluminum. The results of the normalizations can be seen in Figure 42. From Figure 42 it can be seen that elevated chromium/aluminum and nickel/aluminum ratios occurred primarily in the eastern portion of study area, adjacent and inside Guanica Bay, and for chromium, to some extent in the western portion of the study area as well. In the western portion of the study area, the actual concentration of chromium at the sites with elevated ratios was less than 10 µg/g.

Schropp and Windom (1988), Schropp *et al.* (1990), and Weisberg *et al.* (2000) have looked at the relationship between aluminum and other metals in sediments using simple linear regressions of log10 transformed data, followed by calculation of the 95% confidence limits. Concentrations of metals that exceed the 95% confidence limits are seen as evidence of anthropogenic enrichment. Using NS&T sediment data for the East Coast, linear regressions were run between aluminum and chromium, nickel and copper. The results indicated that HTOC 17 and HTOC 19 in Guanica Bay were both above the 95% confidence limits for chromium and nickel, providing evidence that the concentrations of these two metals in the Bay showed evidence of anthropogenic enrichment in addition to the input of sediments from the Lajas Valley. The same was true when the concentrations of chromium and nickel at the two sites in Guanica Bay were compared with the regressions found in Weisberg *et al.* (2000) for sediments from EPA's Environmental Monitoring and Assessment Program (EMAP) Virginian (Long Island to Chesapeake Bay) and Louisianian (Tampa Bay to Texas) Provinces. The only other metal concentrations that were slightly above the 95%

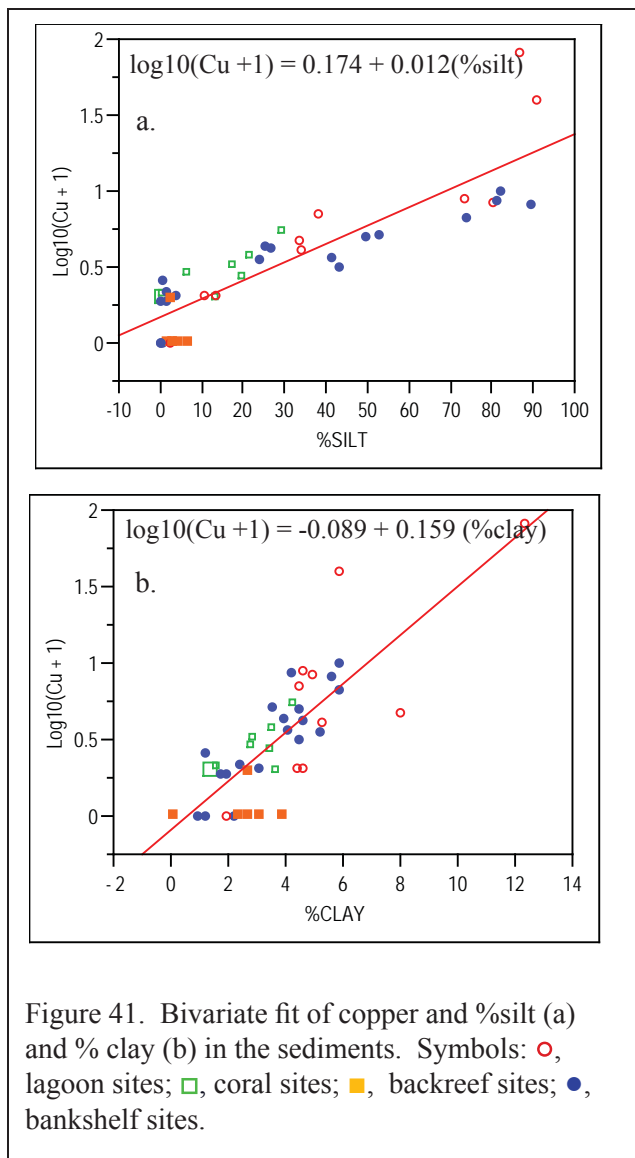


Figure 41. Bivariate fit of copper and %silt (a) and % clay (b) in the sediments. Symbols: ○, lagoon sites; □, coral sites; ■, backreef sites; ●, bankshelf sites.

confidence limits in the study area using NS&T data was nickel at four sites around La Parguera (HTOC 16, LTOC 7, LTOC 22, and LTOC 15).

Effects of Copper on Corals. Copper appears to be the trace element most commonly used in toxicity tests in corals. A number of studies have investigated the effects of this trace element on coral fertilization and development. Victor and Richmond (2005) found that a copper concentration of 10 µg/L and greater significantly affected fertilization in the reef building coral *Acropora surculosa* in the laboratory after five hours. In a second experiment, Victor and Richmond (2005) found that embryo development was affected when gametes were

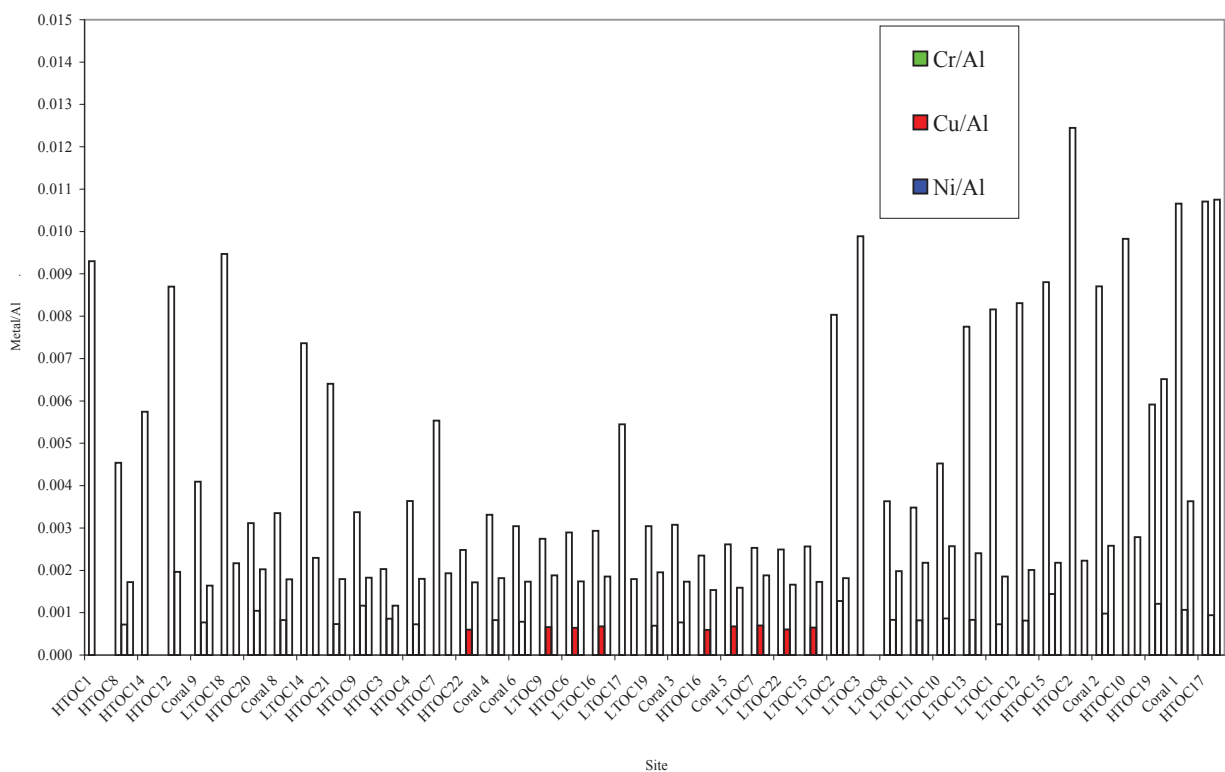


Figure 42. Normalizations of chromium, nickel and copper to aluminum. Metals were normalized by dividing the concentration of each metal by the concentration of aluminum in the sediment at the site.

exposed to copper at concentrations of 12 $\mu\text{g/L}$ and higher. At concentrations of 58 $\mu\text{g/L}$ and greater, no embryo development was observed. As noted earlier, the concentrations of contaminants found in the sediments in southwest Puerto Rico cannot be compared with aqueous concentrations which have been found to affect corals. The corresponding concentration of copper and many other contaminants in the water column is different, typically much less than the sediment concentration. This is due to the contaminant preferentially binding to the sediment, with the corresponding water concentration being much lower.

Reichelt-Brushett and Harrison (2005) exposed the gametes from a number of coral species to copper. Eggs and sperm were first dosed separately with the appropriate copper concentrations and then the gametes were mixed and allowed to incubate in sealed vials on the reef for five hours. A copper concentration of 20 $\mu\text{g/L}$ and higher

significantly reduced the fertilization success in *G. aspera*. At copper concentrations at or above 75 $\mu\text{g/L}$, fertilization success was one percent or less. Reichelt-Brushett and Harrison (2005) also found that copper at or above 20 $\mu\text{g/L}$ significantly decreased fertilization success in the coral *Goniastrea retiformis*, the same as for *G. aspera*. Fertilization success was also significantly reduced in *Acropora longicyathus* at 24 $\mu\text{g/L}$, similar to *G. retiformis*, however, in the finger coral *Acropora tenuis*, a significant effect on fertilization was only seen at concentrations of 42 $\mu\text{g/L}$ and greater. At 67 $\mu\text{g/L}$ and higher, fertilization success was 5 percent or less.

Reichelt-Brushett and Michalek-Wagner (2005) investigated the effects of copper on the soft coral *Lobophytum compactum*. A significant difference in fertilization success was found at copper concentrations of 117 $\mu\text{g/L}$ and above, higher than that found for a number of other coral species. The EC_{50} value (that which decreased fertilization

by 50%) was 261 µg/L. Reichelt-Brushett and Michalek-Wagner (2005) speculated the higher copper concentrations needed to affect fertilization in this species could be a function of its longer gametogenic cycle (nearly two years), leading the authors to suggest that *L. compactum* may have developed some type of biochemical copper detoxification mechanism.

Negri and Heyward (2001) calculated an EC₅₀ of 17.4 µg/L copper for fertilization in *A. millepora* (4 hour exposure). They also found there was a 50 percent inhibition of larval metamorphosis in the presence of coralline algae at a copper concentration of 110 µg/L.

Reichelt-Brushett and Harrison (2004) exposed 4-6 day old larvae of *G. aspera* to varying concentrations of copper for 12 hours and calculated an EC₅₀ for larval motility of 21 µg/L. They also noted that the EC₅₀ was much lower than the 24 hour LC₅₀ (137 µg/L) or lethal concentration.

Reichelt-Brushett and Harrison (2000) investigated the settlement of larval *A. tenuis* on biologically preconditioned terracotta ceramic tiles. A copper concentration of 42 µg/L was found to significantly reduce settlement success of the larvae after 48 hours. The 48 hour EC₅₀ was 35 µg/L, and the calculated no observed effect concentration (NOEC) was 20 µg/L.

Goh and Chou (1997) investigated the effects of copper on the zooxanthellae *Symbiodinium microadriaticum*, isolated from the coral *Montipora verrucosa*. A concentration of 40 µg/L copper resulted in growth inhibition in the symbiotic dinoflagellate. Goh and Chou (1997) also conducted a series of experiments where the zooxanthellae were exposed to both copper and zinc, and noted there was an apparent synergistic effect when the metals were combined.

Nystrom *et al.* (2001) investigated the effects of copper on *Porites cylindrica*. There was no change in primary production, (oxygen production/mg chlorophyll *a*) when adult corals were exposed to 11 µg/L copper for 24 hours. However, in corals that were preexposed to elevated temperatures (4°C higher than ambient) for five days and then exposed

to the copper, there was a significantly decreased primary production rate. These studies suggest that copper has the ability to affect coral larvae at low concentrations. This could be of concern in some locations, particularly in areas of higher boat traffic, or in areas where copper is used in agriculture.

Other Effects of Copper. Bioaccumulation of copper is generally low in aquatic animals, with the exception of oysters and squid. The oxidation states of copper includes Cu⁰, Cu⁺¹, Cu⁺², and Cu⁺³, with the cupric ion (Cu⁺²) appearing to be the most toxic. A fair amount of research has been conducted to look at the toxicity of copper to a variety of other aquatic organisms. Exposure of the amphipod *Allorchestes compressa* for 4 weeks at a concentration of 10 µg/L caused adverse effects on growth and survival (Ahsanullah and Williams, 1991). Wong *et al.* (1993) found that feeding was inhibited by more than 50 percent in the penaeid shrimp *Metapenaeus ensis*, at a total copper concentration of 250 µg/L after two hours. In rainbow trout (*Oncorhynchus mykiss*), Saucier *et al.* (1991) found that embryo survival was reduced by 30 percent as a result of exposure to a copper concentration of 22 µg/L for 41 weeks.

Other Contaminants

As part of this project, a number of other contaminants, both biological and chemical, were measured. The bacteria *Clostridium perfringens*, often used as an indicator of sewage was analyzed in the sediments collected. Total nitrogen and total phosphorus were also measured in water samples. Finally, water samples were collected from selected sites and analyzed for Irgarol 1051, a herbicide currently used in copper-based antifoulant paints for boat hulls. Water samples were collected in May and then again in August 2005. The results of each of these analyses are discussed below.

Clostridium perfringens. This anaerobic, gram-positive staining rod-shaped bacteria frequently occurs in the intestines of humans as well as in domestic and wild animals. The results of the analysis of sediments for *C. perfringens* are shown in Figure 43 and in Appendix K.

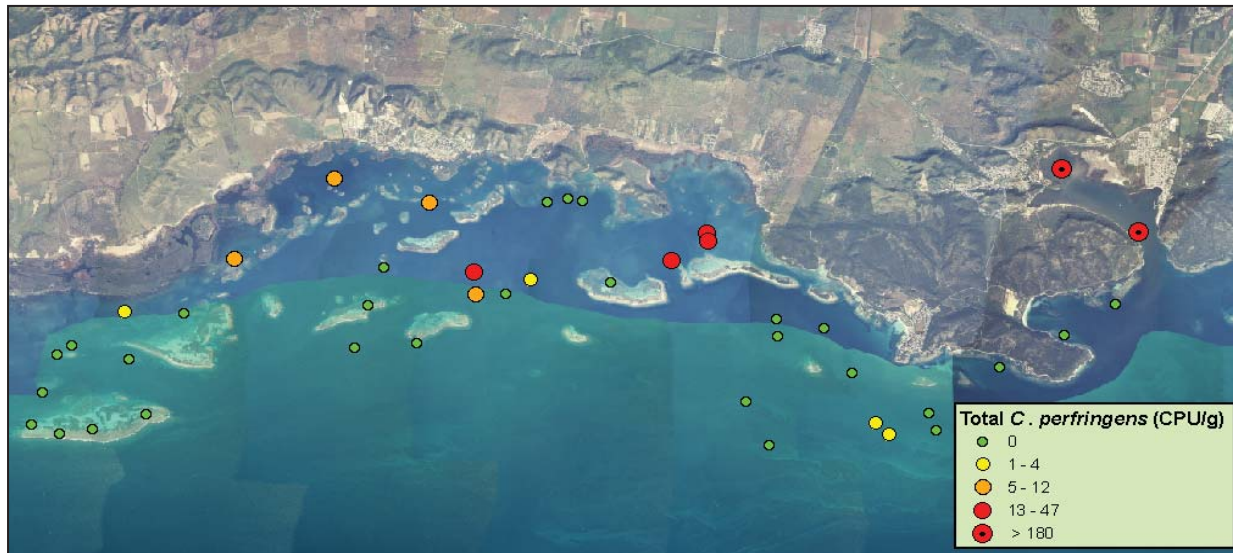


Figure 43. Map of *Clostridium perfringens* detected in the sediments in southwest Puerto Rico. Data classified into quantiles.

To assess the presence of viable *C. perfringens*, sediment extracts are plated on growth medium and the number of colonies that develop are counted. The highest concentrations of *C. perfringens* were found, similar to a number of the other contaminant classes, in Guanica Bay. The sediment sample from HTOC 17 had over 180 colony forming units (CFU) per gram of sediment. Sediment collected from HTOC 19 contained over 1,700 CFUs per gram of sample. Sources of the elevated levels of *C. perfringens* would include wastewater treatment plants or septic systems that discharge to Guanica Bay, along with inputs from the pastures in the Lajas Valley.

No health guidelines were located for concentrations of *C. perfringens* in sediments. *C. perfringens* is a common cause of foodborne illnesses. A more severe form of the disease is often fatal and results from ingesting large numbers of the active bacteria. *C. perfringens* also has the capability of forming spores which can persist in soils and sediments in areas subject to human and animal fecal pollution.

Nutrients. Water samples were collected and analyzed for both total nitrogen and total phosphorus from 47 sites (Figure 44 and 45). The samples were taken at the sites where sediment samples were taken, plus at an additional four sites where sediments were not available.

Total nitrogen and total phosphorus are the sums of the organic and inorganic forms for each nutrient. There were a number of areas, for example, around La Parguera (HTOC 16 and Coral 5), and especially in Guanica Bay that had elevated levels of nitrogen and phosphorus. A Wilcoxon Test run on the nutrient concentrations indicated no significant relationship between habitat and nitrogen, but that habitat type was correlated with phosphorus concentration. The NS&T Program does not regularly analyze for nutrients, so no median or 85th percentile values are available.

No criteria for total nitrogen or total phosphorus were located for coral reefs. These types of criteria for the region would be useful as the nitrogen and phosphorus levels detected in the study area in southwest Puerto Rico could be compared with values deemed protective of reef habitats.

Effects of Nutrients on Corals. A number of studies have found that elevated nitrogen and phosphorus levels can affect corals, both directly and indirectly. In the laboratory, Harrison and Ward (2001) found that mean coral fertilization rates were significantly reduced in the branching coral *Acropora longicyathus*, when gametes were exposed to ammonium (NH_4^+) concentrations equal or greater

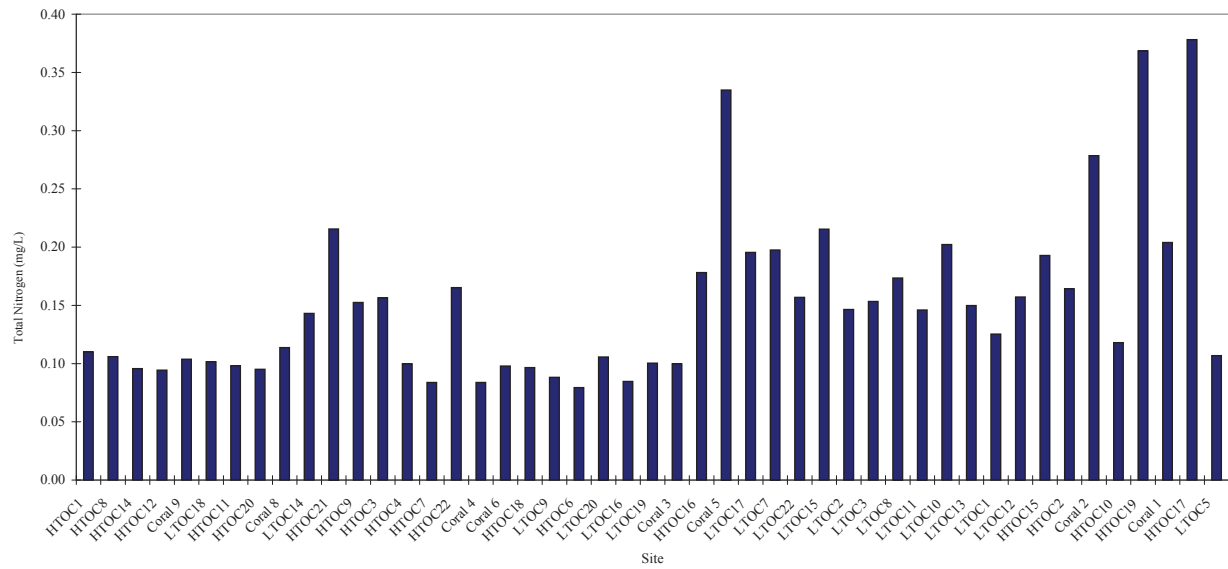


Figure 44. Total nitrogen in water samples from southwest Puerto Rico.

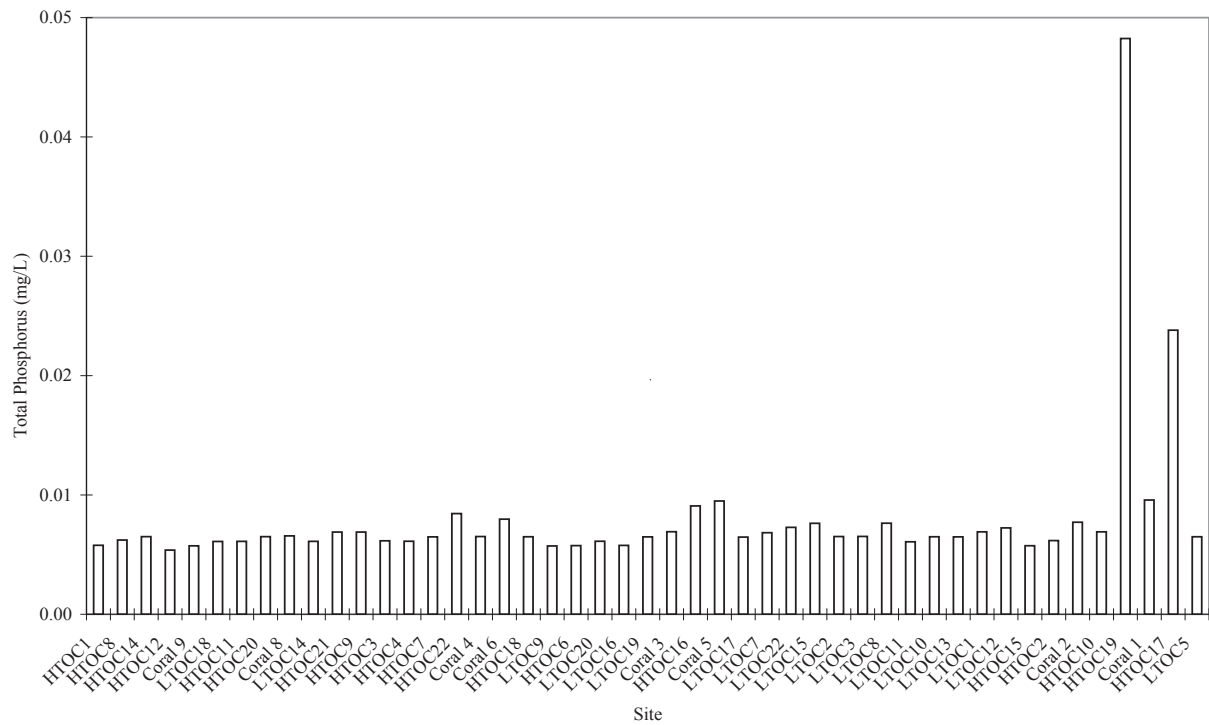


Figure 45. Total phosphorus in water samples from southwest Puerto Rico.

than 18 µg/L. Similarly, mean fertilization was also reduced in gametes from *A. longicyathus* exposed to phosphate (PO₄⁻³) concentrations equal to or greater than 95 µg/L. In addition, the mean percentage of embryos that developed normally was significantly reduced.

Marubini and Davies (1996) investigated the effects of nitrate (NO₃⁻) on two species of coral, *Porites porites* and the massive coral *M. annularis*. At nitrate concentrations of 310 µg/L or greater, the population density of zooxanthellae within the corals increased significantly. The amount of chlorophyll *a* and algal protein increased as well. The most dramatic change found in this study, however, was that the rate of calcification or skeletogenesis of the coral decreased by approximately 50 percent in both species when exposed to the nitrate enrichment. Lower calcification rates are of concern as they could result in the corals becoming more fragile and thus more susceptible to damage.

In the field, Hunte and Wittenberg (1992) found that the number of coral recruits decreased with increasing eutrophication along the fringing reefs on the west coast of Barbados. In addition, coral settlement rates on artificial plates were lower on reefs in the more eutrophic areas, and non-coraline settlement was heavier on the plates located in eutrophic areas. Also in the field, Abelson *et al.* (2005) found the lowest coral recruitment on ceramic tiles closest to a major eutrophication source in the Red Sea. Indirectly, increased nutrient levels can also result in macroalgal species outcompeting corals and forming dense mats that overgrow coral areas (Fabricius, 2005). As development pressures increase in areas adjacent to coral reefs, the risk of eutrophication and subsequent algal growth overwhelming corals becomes a greater concern and more likely phenomenon.

Other Effects of Nutrients. Nitrogen and phosphorus are essential elements for both plants and animals. Nitrogen, for example, is a component of proteins, RNA and DNA. Phosphorus is found in nucleic acids and also in certain types of lipids, essential for example, in the structure of cell membranes. In marine systems nitrogen is generally the limiting factor for primary production, however, under

certain conditions phosphorus can become limiting. Although essential for plant and animal life, elevated levels of these nutrients can lead to eutrophication, characterized by an overabundance of algae and depressed dissolved oxygen levels in a river, bay or coral reef. Eutrophication is an overproduction of organic matter, particularly algae, and is commonly caused by an increase in the amount of nutrients coming into a waterbody (Bricker *et al.*, 1999). The additional algae shade out the light needed by native plants. As the algae produced dies, additional oxygen is needed for its decay, leading to low levels of oxygen in the water. These low oxygen or hypoxic conditions make it difficult for other aquatic organisms to survive.

Irgarol 1051. As part of the collaborative effort for this project, water samples from selected sites around La Parguera were analyzed by NOAA's Center for Coastal Environmental Health and Biomolecular Research in Charleston, South Carolina for Irgarol 1051. Since the ban on the use of tributyltin as an antifoulant on smaller vessels in the late 1980s, a number of "TBT-free" formulations containing copper combined with organic boosters have been used (Konstantinou and Albanis, 2004). One of the more frequently used compounds is Irgarol 1051 (2-methylthio-4-*tert*-butylamino-6-cyclopropylamino-*s*-triazine), similar in structure to atrazine, another triazine herbicide which is used in agriculture. Irgarol 1051 is highly phototoxic, inhibiting the electron transport chain in Photosystem II within the chloroplasts of algal cells (Carbery *et al.*, 2006).

Water samples were analyzed from the May and August 2005 missions. The results of the analyses are shown in Figure 46. The highest concentration of Irgarol 1051 was found adjacent to a marina (15.4 ng/L) in La Parguera in August 2005, followed by a concentration of 6.3 ng/L adjacent to a dry dock. Water samples from several sites where corals were sampled, or from reference sites were below the method detection limit or MDL (1 ng/L) for Irgarol 1051 (Figure 46).

Effects of Irgarol 1051 on Corals. Several papers were located on the effects of Irgarol 1051 on corals, specifically related to primary production. Owen



Figure 46. Detection of the antifoulant compound Irgarol 1051 in water samples from Puerto Rico in May 2005 (circles) and August 2005 (squares). Data classified into quantiles (25%).

et al. (2002) investigated the effects of Irgarol 1051 on the yellow pencil coral *Madracis mirabilis*. Experiments were carried out on both the isolated zooxanthellae and the intact coral. For the isolated zooxanthellae, changes in photosynthesis were measured by assessing the uptake of ^{14}C bicarbonate from the exposure medium, using a suite of irradiances over an 8 hour period. Concentrations of Irgarol 1051 as low as $0.063 \mu\text{g/L}$ resulted in complete inhibition of ^{14}C uptake by zooxanthellae after 6 hours of exposure. Incorporation of ^{14}C provides an estimate of photosynthetic primary production. Whole coral respiration over an 8 hour period was also investigated. Irgarol 1051 concentrations as low as $0.1 \mu\text{g/L}$ resulted in reduced photosynthesis in *M. mirabilis* as measured by the generation of oxygen after four hours. At $1 \mu\text{g/L}$, little or no photosynthesis occurred.

An investigation by Owen *et al.* (2003) examined the effects of Irgarol 1051 on a number of other coral species. Incorporation of ^{14}C was used to measure changes in net photosynthesis. Concentrations of $2 \mu\text{g/L}$ Irgarol 1051 were found to significantly inhibit photosynthesis in the symmetrical brain coral *D. strigosa* and in the golfball coral *F. fragum*. A

concentration of $2 \mu\text{g/L}$ diuron in these species produced similar results, while a concentration of at least $100 \mu\text{g/L}$ of the herbicides atrazine or simazine was needed to decrease photosynthesis in these species (Owen *et al.*, 2003).

Jones and Kerswell (2003) examined the effect of Irgarol 1051 and other herbicides on two species of corals from Australia. Intact *Seriatopora hystrix* or bird's nest coral were exposed in the laboratory to five test concentrations of Irgarol 1051 along with a control. Changes in chlorophyll fluorescence (maximum effective yield) in coral fragments containing zooxanthellae were determined to assess the effects of the herbicide. In *S. hystrix*, an EC_{50} (concentration needed to reduce maximum fluorescence by 50 percent) of $0.7 \mu\text{g/L}$ was calculated for a 10 hour exposure. The effect of Irgarol 1051 on chlorophyll fluorescence was also tested in the stony coral *A. formosa*. Results indicated an LC_{50} for this species of $1.3 \mu\text{g/L}$. After the exposures, Jones and Kerswell (2003) moved the coral fragments of *S. hystrix* to fresh running seawater. The amount of time needed to recover the preexposure fluorescence levels was measured. While the level of fluorescence returned to normal in a matter of minutes or hours in coral exposed to

herbicides such as diuron, atrazine, and simazine, *S. hystrix* exposed to Irgarol 1051 recovered more slowly, on the order of hours to days, which would have a greater impact on the corals exposed to Irgarol 1051.

Other Effects of Irgarol 1051. Much of the work done to assess other effects of Irgarol 1051 has focused on algae, with some work done on invertebrates. Okamura *et al.* (2000) calculated an EC₅₀ of 2 µg/L for the growth of germlings from the marine brown macroalga *Eisenia bicyclis* after 7 days. Okamura *et al.* (2000) also investigated the effects of Irgarol 1051 on growth in the freshwater duckweed *Lemna gibba*, and calculated a 7-day EC₅₀ of 11 µg/L. These values are similar to those found for coral zooxanthellae.

There have also been investigations into invertebrate toxicity. Bellas (2006) investigated the toxicity of five antifouling biocides, including Irgarol 1051, to larvae of the mussel *Mytilus edulis*, the sea-urchin *Paracentrotus lividus* and the tunicate *Ciona intestinalis*. Toxicity was measured in terms of embryonic development, larval growth and larval settlement. All tests ran for 48 hours. Irgarol 1051 was the least toxic compound tested. The EC₅₀ for embryonic development in *M. edulis* was 1,540 µg/L, in *P. lividus*, 4,021 µg/L, and *C. intestinalis*, the EC₅₀ was 2,115 µg/L. In *P. lividus*, the larval growth EC₅₀ was 6,032 µg/L. An EC₅₀ for larval settlement in *C. intestinalis* could not be calculated, as Irgarol 1051 showed no toxicity at concentrations approaching 6,500 µg/L. Irgarol 1051 is an effective inhibitor of the photosynthetic electron transport chain in chloroplasts (Konstantinou and Alabanis, 2004), and so it is perhaps not unexpected that some nonphotosynthetic organisms would be less sensitive to this compound.

Correlations Between Sediment Contaminants. A nonparametric Spearman's Rho test was run to look for correlations between some of the sediment contaminant classes described in this report. The results indicated that the sediment organic contaminant classes (total PAHs, total PCBs, and total DDTs) were highly correlated ($P < 0.0001$) with

each other. The trace elements chromium, nickel, copper, zinc, and lead were likewise highly correlated with each other. In addition, the organic contaminant classes were also highly correlated ($P < 0.0001$) with the trace elements listed above. The only contaminant in this exercise that did not correlate well with the other contaminants was arsenic. As mentioned previously, the presence of arsenic in the study area did not have the same geographic distribution as the other contaminants, and the low concentrations and distribution of this trace element in the study area was suggestive of natural sources rather than anthropogenic activities.

Contaminants and Coral Species Richness

One of the goals of this project and the assessment framework is to examine linkages between chemical contaminants and coral ecosystem condition. Because of the detailed mapping of coral reefs that has been completed in this part of Puerto Rico, it is possible to begin assessing relationships between the spatial distribution of corals in the study area and chemical contaminants. For this report, an analysis was performed to quantify correlations between concentrations of total PAHs in sediments and coral species richness.

Contaminant Modeling

An analysis of spatial structure for total PAH using Geary's C test indicated a significant spatial autocorrelation ($P < 0.0001$). Subsequently, a geospatial model was developed using indicator Kriging. Figure 47 shows the relationship between the modeled and observed total PAH concentrations ($P < 0.0001$, $r^2 = 0.78$). Figure 48 shows the modeled total PAH and includes the relative PAH concentrations (dots) detected in the sediments. The map (Figure 48) shows higher modeled concentrations of total PAHs in the sediments (red shading) in the nearshore areas around the town of La Parguera. The model also shows higher concentrations east of La Parguera and in Guanica Bay. Moving offshore, the modeled concentrations decrease.

Figure 49 contains results of some additional modeling for total PCBs, total DDTs, and copper. It

can be seen that the concentrations of contaminants in the sediments result in somewhat different geospatial models, however all show an area of higher modeled contaminant concentrations in and around La Parguera and to the east. Although contaminant concentrations detected in Guanica Bay were higher for most contaminant classes, because there were only two sediment samples taken there, the model was not able to capture the magnitude of the higher concentrations.

The patterns of modeled PAHs in Figure 48 could be the result of a number of processes. Garcia *et al.* (1998) identified improvements to transportation and infrastructure as transforming the village of La Parguera from a “mostly undeveloped and quiet fishing village” to a center of tourism. That transition has been reflected in the increased number of resorts, guest houses and vacation homes, and proliferation of the transient population over the last 10 years from approximately 35,000 to over 100,000 per year. (Garcia *et al.*, 1998). Atmospheric deposition and surface runoff related to motor vehicle traffic are likely means of introducing PAHs into the surrounding waters. These fluxes would be highest particularly on weekends, when automobile traffic is at its peak. The use of outboard and inboard engines on boats in the area also likely contributes to the PAH load in the study area. The area around La Parguera has a substantial number of boats, both

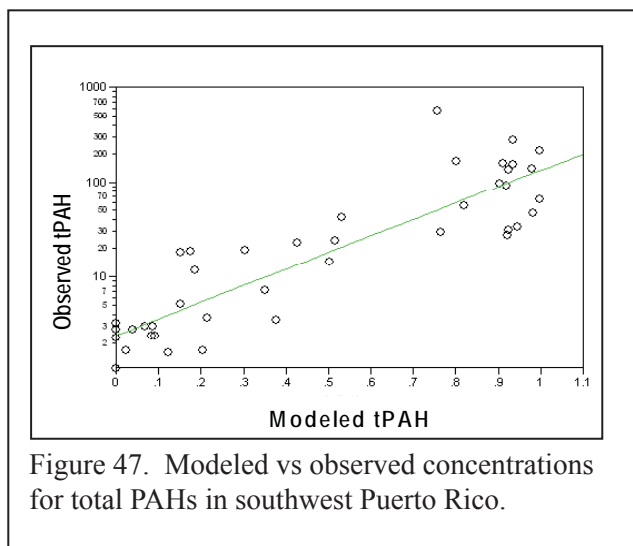


Figure 47. Modeled vs observed concentrations for total PAHs in southwest Puerto Rico.

for recreational and commercial fishing. Between 1996 and 2003, the total number of boats registered in Puerto Rico increased by 28% to over 60,000 (Garcia-Sais *et al.*, 2004).

A number of emergent reefs in the area (*e.g.*, Enrique, Media Luna, and Corral) have shallow areas on the leeward side, that serve as gathering areas for recreational boaters, particularly on weekends and holidays. It has been reported that over 200 boats can be anchored on a small reef in La Parguera (Garcia-Sais *et al.*, 2004). The presence of boats, either moored or idling on the backsides of the emergent

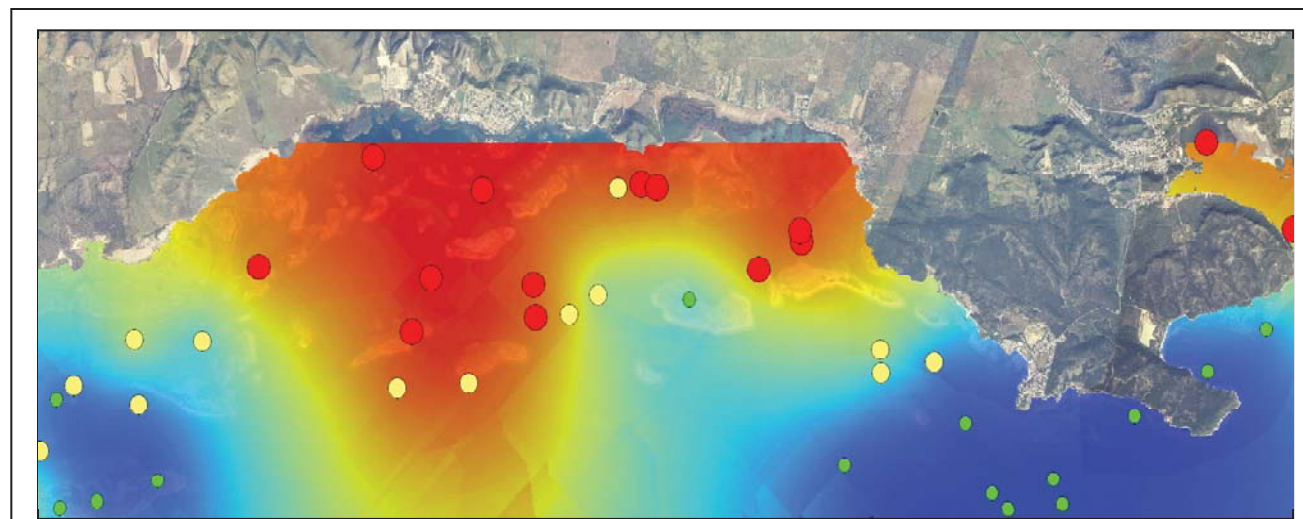


Figure 48. Indicator Kriging and measured concentrations of total PAH in the sediments in southwest Puerto Rico. Red areas indicate areas of higher modeled total PAH concentrations. Colored dots represent sediment contaminant quantiles (●, low; ●, medium; ●, high).

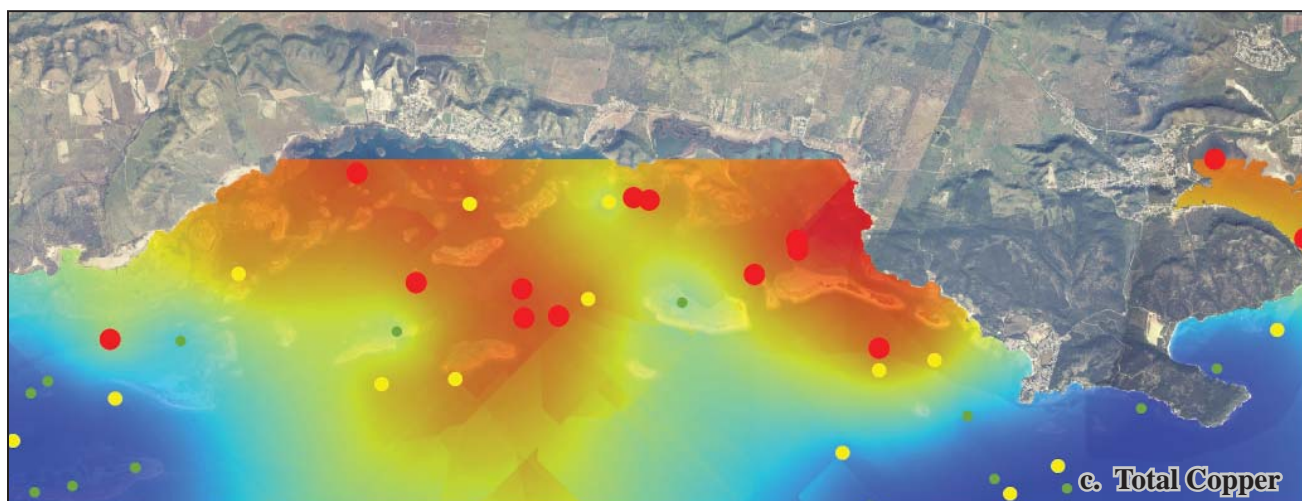
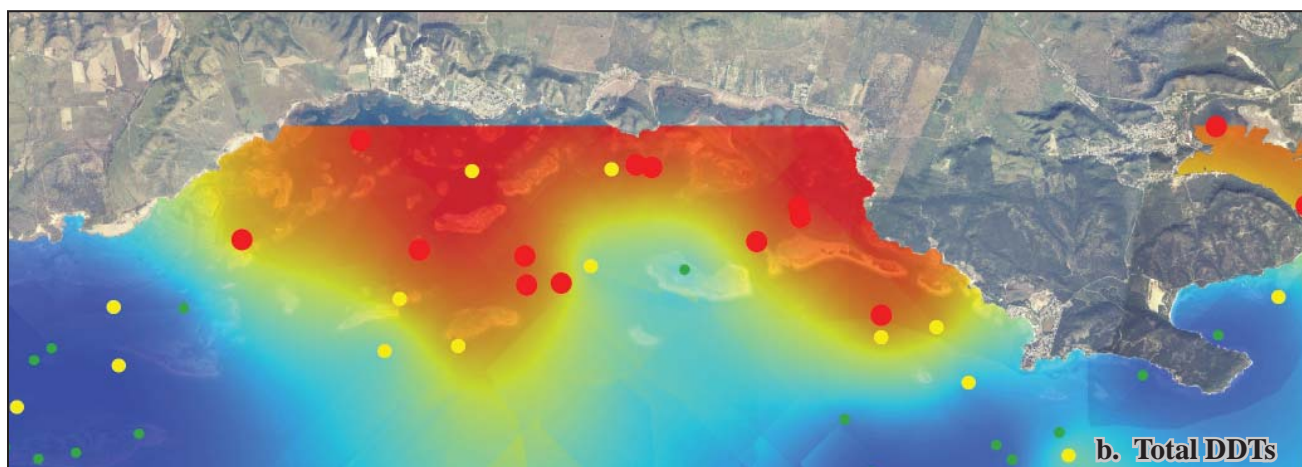
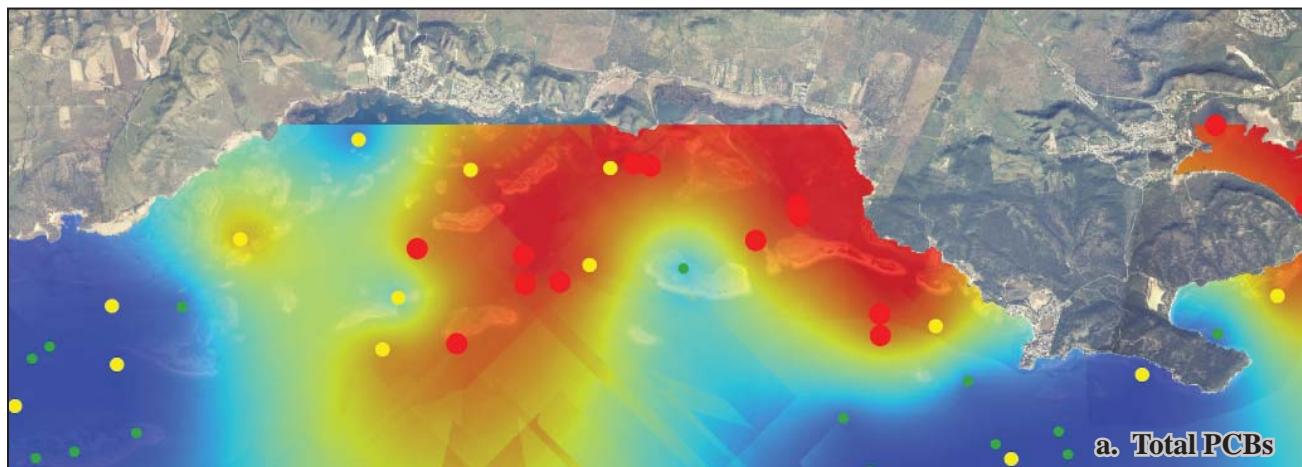


Figure 49. Indicator Kriging and measured concentrations of a) total PCBs, b) total DDTs, and c) copper. Colored dots represent sediment contaminant quantiles (●, low; ●, medium; ●, high).

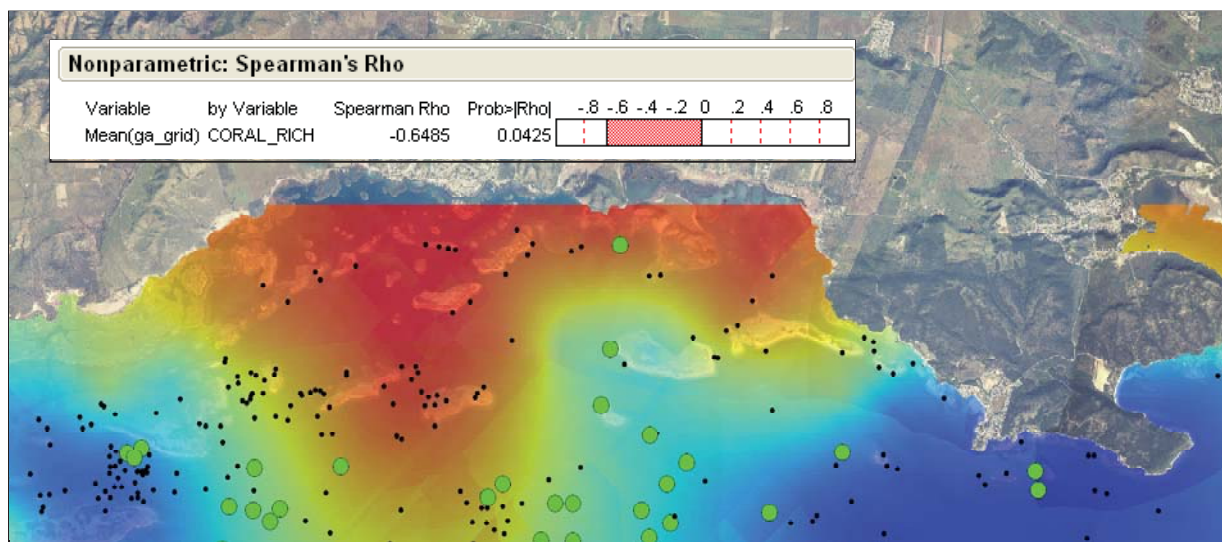


Figure 50. Kriging of total PAHs and coral species richness. Dots indicate survey points for NOAA's Biogeography Branch. Green dots indicate those locations where coral species richness was in the top 25th percent for brain, branching, pillar, encrusting, mound and boulder corals. Black dots are the remaining Biogeography Branch survey locations.

reefs could also be sources of the PAHs found in the sediments. However, the concentrations of total PAHs at the backreef sites did not appear to be elevated relative to the other habitat types.

Total PAHs and Coral Species Richness

Chemical contaminants, ranging from PAHs to metals to pesticides, can be toxic to corals and other organisms living on the reef (Negri and Heyward, 2001; Owen *et al.*, 2003; Peachey and Crosby, 1996). Chemical contaminants can impact a number of vital life processes in corals including fertilization, settlement, metamorphosis, development and growth. These effects can be manifested from the molecular to the community levels. At the community level, toxic contaminants could impact the ability of various coral species to survive in an area. Developing correlations between the presence of chemical contaminants and coral species richness (number of species in an area), can help establish linkages between contaminants and coral distribution.

In Figure 50, coral reef survey data is overlaid on the total PAH model. Each point represents a survey point by NOAA's Biogeography Branch to

characterize coral cover in the study area. Green dots represent those areas that were most speciose (top 25th percent), while the remaining dots indicate survey sites where the coral species richness was below this 75th percentile. In order to assess whether a correlation existed between the modeled concentrations of total PAHs and species richness, a nonparametric Spearman's Rho test was carried out.

There was a strong negative correlation ($P > 0.0425$; Spearman's Rho = -0.6485) between the modeled PAH concentrations and coral species richness indicating that as PAH concentrations increase, coral species richness decreases. The corals used in the analysis included the major reef building species such as boulder corals (*M. annularis*), brain corals (*Diploria spp.*), and branching corals (*e.g.*, *Porites porites*, *A. palmata* and *A. cervicornus*).

The results presented in Figure 50, however, do not represent a causal relationship, but rather an indication of a negative association between PAHs and coral species richness for this area. Many factors could be responsible for the observed pattern, ranging from physical processes (currents, freshwater runoff),

damage to corals caused by boat traffic coming in and out of the La Parguera area, to sedimentation resulting from natural or anthropogenic activities and, the presence of chemical contaminants such as PAHs. The data could also simply be indicative of areas of finer grain sediments which tend to accumulate contaminants and which are also less suitable for coral settlement and development. Future work to assess contaminants in corals will provide a better understanding of the factors influencing the patterns observed.

The results presented here do, however, provide an opportunity to develop testable hypotheses to better assess how contaminants may be impacting corals. Additional results, including the coral contaminant data will be used to better understand associations between contaminants and corals from the molecular to the community levels.

Summary and Conclusions

The degradation of coral reef ecosystems worldwide has led to intensive efforts to understand and mitigate the stressors responsible for the declines of these ecosystems. The role of pollution is often cited as a major factor, but the degree to which pollution, and more specifically, chemical contaminants, impact coral reefs is in most cases unknown. Because of this, coral reef managers may be missing an important, and in some locations, a critical piece of information needed to effectively manage the reefs. The impacts of chemical contaminants on corals at the community (species diversity and distributions) and individual (*e.g.*, metabolism and reproduction) levels are both needed for a complete assessment of contaminant impacts.

Project and Framework

NOAA and its partners initiated this project to characterize chemical contaminants in southwest Puerto Rico and to provide a mechanism for assessing

linkages between chemical contaminants and coral health (function and coral species distribution/richness). This project has been developed in direct response to the need to assess the effects of pollution on coral reefs as called for by the U.S. Coral Reef Task Force, the “State of the Reefs” report (Waddell *et al.*, 2005), the Pew Oceans Commission (Pew Oceans Commission, 2003), and the World Resources Institute (Burke and Maidens, 2004), among others.



Elkhorn coral (Acropora palmata), recently listed as threatened under the U.S. Endangered Species Act (ESA).

The assessment framework established as part of this project brings together various specialties ranging from environmental chemistry to biogeography to molecular and microbiology. By bringing these specialties and the resulting data types together, it is more likely that linkages between contaminants and coral condition can be established.

This report is the first product of this collaborative effort, and contains the results of the analysis of chemical contaminants in the sediments in southwest Puerto Rico. The study area, from Guanica Bay west past the town of La Parguera was chosen because of established partnerships, data availability, and designation of the area as a Coral Reef Ecosystem Study (CRES) site, the 5-year program funded by NOAA’s Center for Sponsored Coastal Ocean Research.

A stratified random sampling design was developed for the study area to characterize the distribution and concentrations of chemical contaminants in the study area. Sediment samples for the analysis of both organic contaminants (*e.g.*, polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs) and pesticides), and inorganic contaminants (trace elements such as chromium, nickel and copper) were collected during a mission in August 2005. The data were analyzed using a series of statistical and geospatial tools.

Chemical Contaminant Concentrations

Overall, the levels of chemical contaminants in the sediments in the study area were fairly low. At most locations, sediment contaminant concentrations were less than the median values in NOAA's National Status and Trends (NS&T) Program, which monitors chemical contamination in coastal waters of the U.S. A pattern seen in the study area for a number of the contaminant classes, included higher concentrations of contaminants in the embayments and behind the emergent reefs, while offshore sites tended to have lower concentrations.

Polycyclic aromatic hydrocarbons or PAHs are associated with the use and combustion of petroleum hydrocarbons and other fuels (e.g., wood). Sediments nearer the shore and adjacent to La Parguera tended to have higher PAH concentrations. As was seen for a number of the contaminant classes, sediment concentrations of total PAHs were above NOAA's National Status and Trends (NS&T) Program median at both sites in Guanica Bay. Corals, however, are typically not found in Guanica Bay.

Polychlorinated biphenyls or PCBs, were used in the past in electrical transformers and capacitors, and are persistent in the environment. The results from the analysis of sediments indicated that PCBs were above the NS&T national median at a number of sites in nearshore areas, adjacent to the town of La Parguera. The NS&T Program has also developed a series of sediment contaminant concentrations associated with toxic effects. The ERL or Effects Range-Low is the concentration of a contaminant below which toxicity (10th percentile) is rarely observed. The ERM or the Effects Range-Median is the concentration above which toxicity in test organisms is more frequently (50th percentile) observed and therefore effects at or above this concentration are more likely. In Guanica Bay, the concentration of total PCBs at the two sites was not only above the ERL, it was also substantially above the ERM value, indicating that PCBs, at least at the two sites sampled in Guanica Bay, are more likely to be impacting the biota there.

Levels of the organochlorine pesticide DDT in the sediments were all below the NS&T national median in the sediments around La Parguera. Elevated levels

of DDT were detected in Guanica Bay. Sediments from both Guanica sites were above the ERL level for DDT, and at one site in Guanica Bay, total DDTs were above the ERM, indicating the DDT in the sediment may have an impact on biota there. The drainage from the Lajas Valley flows into Guanica Bay, and the Lajas Valley has had extensive agricultural activity over the years. The elevated levels of DDT found in the sediments at the two sites in Guanica Bay may be related to the past agricultural use of this insecticide.

Sediment concentrations of chromium, nickel and copper were below the NS&T national median in most of the sediment samples taken in the study area. At the two sites sampled in Guanica Bay, levels of both chromium and nickel were above the ERM value for these trace elements, indicating these two metals may be having adverse impacts on the aquatic biota.

Contaminants and Coral Species Richness

One of the major goals of the assessment framework developed is to understand linkages that may exist between chemical contaminants and coral ecosystem condition. To begin to address this goal, an exercise was carried out to assess correlations between PAH concentrations in the sediments and coral species richness. A geospatial model of total PAHs in the sediments was developed using the PAH data which was spatially autocorrelated. Existing mapped data on coral species richness for the study area was then overlaid on the modeled PAH concentrations. A nonparametric analysis of the modeled PAH data and coral species richness for the major reef building species indicated a strong negative correlation between modeled PAH concentrations and coral species richness. That is, higher total PAHs in the sediments were associated with lower coral species richness.

The cause for the negative correlation between sediment PAHs and coral species richness is unknown. As mentioned, there could be many other physical (e.g., currents, sedimentation), chemical (e.g., temperature, salinity) and biological (substrate preference) factors that could be responsible for the observed pattern. The results from this analysis,

however, will help direct additional data analysis and the development and testing of hypotheses.

Next Steps

Additional products will be developed as the data from this project are received and analyzed. In the short term, additional data analyses will be carried out to look at correlations between other contaminant classes and coral species diversity, including individual species distributions. If contaminants are in fact impacting corals in the study area in southwest Puerto Rico, it is likely that some contaminants will have a greater effect than others, and it is also likely that certain species of corals would have a greater sensitivity to one or more contaminants or contaminant classes.

As part of this project, an analysis of contaminants has been completed for the *Porites astreoides* coral tissues collected. Pathogens in the corals have also been quantified. The analysis of contaminants in the corals provides a better idea of not only what the corals have been exposed to, but what they have accumulated, making it easier to correlate the presence of contaminants in the corals with effects.

The additional data types to be included in this project will add to the power of the analysis and interpretation of the information. Combining the chemical contaminant data along with information on coral species distributions, creates a powerful tool that can be used to better assess effects at different levels of organization. As an example, chemical contaminants can have a toxicological impact on individual corals, affecting metabolism and reproduction. If these effects are severe enough over time, they would be manifested at the community level, in terms of coral species distribution and diversity. Generating data that can be indicative of effects at the individual and community levels provides the opportunity for understanding the broader effects that chemical contaminants may have on corals and coral reefs.

The inclusion in this project of pathogens associated with coral disease is another important data type. A number of bacterial and fungal pathogens have been associated with coral disease. It is possible that the

presence of chemical contaminants could cause corals to put more energy into metabolizing or responding to the chemical's toxic effects stressing the coral, and making it more susceptible to disease.

The results from this project will also be used to drive additional assessments. These might be in the form of applying the framework developed to other areas in the Caribbean or possibly the Pacific. Assessments could also be carried out to pose additional hypotheses in the current study area in southwest Puerto Rico.

At the present time, chemical contaminants are analyzed in sediments and coral tissues. In the future, additional organisms, both invertebrates and vertebrates, could be assessed in terms of chemical contaminants. This additional information, combined with biogeographic data would further increase the power of the approach.

Maintaining the multi-discipline nature of this study (*e.g.*, environmental chemistry, biogeography, molecular and microbiology) is critical to any additional work. Combining these specialties into a standardized ecosystem approach for assessing coral reefs would greatly enhance the characterization of coral reefs, and is an approach that will evolve over time, and can also be transferred to other institutions. The approach would greatly increase our ability to understand not only the overall concentration of contaminants in corals reefs, but the extent to which the stressors are impacting these valuable and fragile ecosystems.

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Literature Cited

- Abelson, A., R. Olinky, and S. Gaines. 2005. Coral recruitment to the reefs of Eilat, Red Sea: temporal and spatial variation, and possible effects of anthropogenic disturbances. *Marine Pollution Bulletin*. 50: 576-582.
- Acevedo, R. 1991. Preliminary observations on effects of pesticides carbaryl, naphthol, and chlorpyrifos on planulae of the hermatypic coral *Pocillopora damicornis*. *Pacific Science*. 45(3): 287-289.
- Ahsanullah, M., and A. R. Williams. 1991. Sublethal effects and bioaccumulation of cadmium, chromium, copper and zinc in the marine amphipod *Allorchestes compressa*. *Marine Biology* 108:59-65.
- Alsberg, T.E., U. Sternberg, R.N. Westerholm, M.E. Strandell, U. Rannug, A. Sundvall, L. Romert, V. Bernson, B. Pettersson, R. Tofgard, B. Franzen, M. Jansson, J.A. Gustafsson, K.E. Egeback, and G. Tejle. 1985. Chemical and biological characterization of organic material from gasoline exhaust particles. *Environmental Science and Technology*. 19: 43-50.
- Bak, R.P.M. 1987. Effects of chronic oil pollution on a Caribbean coral reef. *Marine Pollution Bulletin*. 18: 534-539.
- Bastidas, C., and E.M. Garcia. 2004. Sublethal effects of mercury and its distribution in the coral *Porites astreoides*. *Marine Ecology Progress Series*. 267:137-143.
- Batley, G. 1996. Distribution and fate of tributyltin. In: S. J. de Mora, editor, *Tributyltin: a case study of an environmental contaminant*. Cambridge University Press. Cambridge, England. 301pp.
- Bellas, J. 2006. Comparative toxicity of alternative antifouling biocides on embryos and larvae of marine invertebrates. *Science of the Total Environment*. 367: 573-585.
- Bennett, R.F. 1996. Industrial manufacture and applications of tributyltin compounds. In: S. J. de Mora, editor, *Tributyltin: a case study of an environmental contaminant*. Cambridge University Press. Cambridge, England. 301pp.
- Berard, A., U. Dorigo, I. Mercier, K. Becker-van Slooten, D. Grandjean, and C. Le Boulanger. 2003. Comparison of the ecotoxicological impact of the triazines Irgarol 1051 and atrazine on microalgal cultures and natural microalgal communities in Lake Geneva. *Chemosphere*. 53(8): 935-944.
- Birch, G.F. 2003. A test of normalization methods for marine sediment, including a new post-extraction normalization (PEN) technique. *Hydrobiologia*. 492: 5-13.
- Birchenough, A.C., N. Barnes, S.M. Evans, H. Hinz, I. Kronke, and C. Moss. 2002. A review and assessment of tributyltin contamination in the North Sea, based on surveys of butyltin tissue burdens and imposex/intersex in four species of neogastropods. *Marine Pollution Bulletin*. 44: 534-543.
- Branchi I., E. Alleva and L.G. Costa. 2002. Effects of perinatal exposure to a polybrominated diphenyl ether (PBDE 99) on mouse neurobehavioural development. *Neurotoxicology*. 23:375-384
- Bricker, S.B., C.G. Clement, D.E. Pirhalla, S.P. Orlando, and D.R.G. Farrow. 1999. *National Estuarine Eutrophication Assessment: Effects of Nutrient Enrichment in the Nation's Estuaries*. NOAA, National Ocean Service, Special Projects Office and the National Centers for Coastal Ocean Science. Silver Spring, MD 71pp.
- Bryant, D., L. Burke, J. McManus, and M. Spalding. 1998. *Reefs at risk: a map-based indicator of threats to the world's coral reefs*. World Resources Institute. Washington, D.C. 56pp.
- Burgess, R.M., S.A. Ryba, M.G. Cantwell, and J.L. Gundersen. 2001. Exploratory analysis of the effects of particulate characteristics on the variation in partitioning of nonpolar organic contaminants to marine sediments. *Water Research*. 35(18): 4390-4404.
- Burke, L., and J. Maidens. 2004. *Reefs at risk in the*

-
- Caribbean. World Resources Institute. Washington, D.C. 80pp.
- Bustos, S., J. Soto, and A.N. Tchernitchin. 1996. Estrogenic activity of p,p-DDT. *Environmental Toxicology and Water Quality*. 11(3): 265-271 .
- Carbery, K., R. Owen, T. Frickers, E. Otero and J. Readman. 2006. Contamination of Caribbean coastal waters by the antifouling herbicide Irgarol 1051. *Marine Pollution Bulletin*. 52(6): 635-644.
- Caribbean Fishery Management Council (CFMC). 1998. Essential fish habitat (EFH) generic amendment to the fisheries management plans (FMPs) of the U.S. Caribbean including a draft environmental Assessment: Volume 1. 169pp.
- Childs, C. 2004. Interpolating surfaces in ArcGIS Spatial Analyst. *ArcUser* July/September: 32-35.
- Colombo, J. C., E. Pelletier, C. Brochu and M. Khalil. 1989. Determination of hydrocarbon sources using n-alkane and polyaromatic hydrocarbon distribution indexes. Case study: Rio de La Plata Estuary, Argentina. *Environmental Science and Technology*. 23(7): 888-894.
- Corredor, J.E., J.M. Morell, and C.E. Del Castillo. 1990. Persistence of spilled crude oil in a tropical intertidal environment. *Marine Pollution Bulletin*. 21(8): 385-388.
- Costanza, R., R. d'Arge, R. de Groot, S. Farber, M. Grasso, B. Hannon, K. Limburg, S. Naeem, R. V. O'Neill, J. Paruelo, R. G. Raskin, P. Sutton, and M. van den Belt. 1997. The value of the world's ecosystem services and natural capital. *Nature*. 387: 253-260.
- Daskalakis, K.D., and T.P. O'Connor. 1995. Normalization and elemental sediment contamination in the coastal United States. *Environmental Science and Technology*. 29: 470-477.
- Dave, G., and R. Xiu. 1991. Toxicity of mercury, copper, nickel, lead, and cobalt to embryos and larvae of zebrafish, *Brachydanio rerio*. *Archives of Environmental Contamination and Toxicology* 21:126-134.
- Dodge, R.E., S.C. Wyers, H.R. Frith, A.H. Knap, S.R. Smith, and T.D. Sleeter. 1984. The effects of oil and oil dispersants on the skeletal growth of the hermatypic coral *Diploria strigosa*. *Coral Reefs*. 3:191-198.
- Downs, C.A., J.E. Fauth, C.E. Robinson, R. Curry, B. Lanzendorf, J.C. Halas, J. Halas, and C.M. Woodley. 2005. Cellular diagnostics and coral health: declining coral health in the Florida Keys. *Marine Pollution Bulletin*. 51: 558-569.
- Downs, C.A., R.H. Richmond, W.J. Mendiola, L. Rougee, and G.K. Ostrander. 2006. Cellular physiological effects of the MV *Kyowa Violet* fuel-oil spill on the hard coral, *Porites lobata*. *Environmental Toxicology and Chemistry*. 25(12): 3171-3180.
- Edinger, E.N., J. Jompa, G.V. Limmon, W. Widjatmoko, and M.J. Risk. 1998. Reef degradation and coral diversity in Indonesia: effects of land-based pollution, destructive fishing practices and changes over time. *Marine Pollution Bulletin*. 36(8): 617-630.
- Eisler, R. 1985. Cadmium hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service Biological Report 85. Contaminant Hazard Reviews Report No. 2. 46 pp.
- Eisler, R. 1986. Chromium hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service Biological Report 85. 60 pp.
- Eisler, R. 1987. Mercury hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service Biological Report 85(1.10).
- Eisler, R. 1988. Arsenic hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service. US Fish and Wildlife Service. Contaminant Hazard Reviews. Report No. 12. 65pp.
- Eisler, R. 1990. Chlordane hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service. Contaminant Hazard Reviews. Report No. 21. 63pp.
-

-
- Eisler, R. 1998. Copper hazards to fish, wildlife, and invertebrates: a synoptic review. U.S. Fish and Wildlife Service. Contaminant Hazard Reviews. Report No. 33. 120pp.
- Eisler, R., and A.A. Belisle. 1996. Planar PCB hazards to fish, wildlife, and invertebrates: a synoptic review. Contaminant hazard reviews. Report No. 31. U.S. National Biological Service. 96pp.
- Epstein, N., R.P.M. Bak, and B. Rinkevich. 2000. Toxicity of third generation dispersants and dispersed Egyptian crude oil on Red Sea coral larvae. *Marine Pollution Bulletin*. 40(6): 497-503.
- Fabricius, K.E. 2005. Effects of terrestrial runoff on the ecology of corals and coral reefs: review and synthesis. *Marine Pollution Bulletin*. 50: 125-146.
- Florida Department of Environmental Protection (FDEP). 2004. Southeast Florida coral reef initiative: a local action strategy. FDEP, Office of Coastal and Aquatic Managed Areas, Coral Reef Conservation Program. Miami, FL. 28pp.
- Fowles J.R., A. Fairbrother, L. Baecher-Steppan, and N.I. Kerkvliet. 1994. Immunologic and endocrine effects of the flame-retardant pentabromodiphenyl ether (DE-71) in C57BL/6J mice. *Toxicology* 86:49-61.
- Garcia, J.R., C. Schmitt, C. Heberer, and A. Winter. 1998. La Parguera, Puerto Rico, USA. In: CARI-COMP Caribbean coral reef, seagrass and mangrove sites. Coastal region and small island paper 3. UNESCO, Paris. 347 pp.
- Garcia-Sais, J.R., R. Castro, J. Sabater, and M. Carlo. 2004. Inventory and atlas of corals and coral reefs, with emphasis on deep-water coral reefs from the U.S. Caribbean EEZ. Caribbean Fisheries Management Council. San Juan, Puerto Rico. 36pp.
- Gardner, T.A., I.M. Cote, J.A. Gill, A. Grant, and A.R. Watkinson. 2003. Long-term region-wide declines in Caribbean corals. *Science*. 301: 958-960.
- Gassman, N.J., and C.J. Kennedy. 1992. Cytochrome P-450 content and xenobiotic metabolizing enzyme activities in the scleractinian coral, *Favia fragum*. *Bulletin of Marine Science*. 50(2): 320-330
- Gibbs, P.E., and G.W. Bryan. 1996. TBT-induced imposex in neogastropod snails: masculinization to mass extinction. In: S. J. de Mora, editor, Tributyltin: a case study of an environmental contaminant. Cambridge University Press. Cambridge, England. 301pp.
- Goh, B.P.L. 1991. Mortality and settlement success of *Pocillopora damicornis* planula larvae during recovery from low levels of nickel. *Pacific Science*. 45(3): 276-286.
- Goh, B.P.L., and L.M. Chou. 1997. Effects of the heavy metals copper and zinc on zooxanthellae cells in culture. *Environmental Monitoring and Assessment*. 44:11-19.
- Harrison, P.L., and S. Ward. 2001. Elevated levels of nitrogen and phosphorus reduce fertilisation success of gametes from scleractinian reef corals. *Marine Biology*. 139: 1057-1068.
- Hassett, J.J., J.C. Means, W.L. Banwart, and S.G. Wood. 1980. Sorption related properties of sediments and energy related pollutants. U.S. Environmental Protection Agency. EPA-600/3-8-041. 133pp.
- Hose, J.E., J.B. Hannah, D. DiJulio, M.L. Landolt, B.S. Miller, W.T. Iwaoka, and S.P. Felton. 1982. Effects of benzo[a]pyrene on early development of flatfish. *Archives of Environmental Contamination and Toxicology*. 11:167-171.
- Hunte, W. and M. Wittenberg. 1992. Effects of eutrophication and sedimentation on juvenile corals. I. settlement. *Marine Biology*. 114: 625-631.
- Jones, R.J., and A.P. Kerswell. 2003. Phototoxicity of photosystem II (PSII) herbicides to coral. *Marine Ecology Progress Series*. 261: 149-159.
- Khalili, N.R., P.A. Scaff and T.M. Holsen. 1995. PAH source fingerprints for coke ovens, diesel and gasoline engines, highway tunnels, and wood combustion emissions. *Atmospheric Environment*. 295: 33-542.
-

-
- Kendall, M.S., C.R. Krueger, K.R. Buja, J.D. Christensen, M. Finkbeiner, and M.E. Monaco. 2001. Methods used to map benthic habitats of Puerto Rico and the U.S. Virgin Islands. National Oceanic and Atmospheric Administration, National Ocean Service, Biogeography Team Technical Report. Silver Spring, MD USA. 45pp.
- Kennedy, C.J., N.J. Gassman, and P.J. Walsh. 1992. The fate of benzo[a]pyrene in the scleractinian corals *Favia fragum* and *Montastrea annularis*. *Marine Biology*. 113: 3131-318.
- Kimbrough, K.L., and R.M. Dickhut. 2006. Assessment of polycyclic aromatic hydrocarbon input to urban wetlands in relation to adjacent land use. *Marine Pollution Bulletin*. 52(11): 1355-1363.
- Kimbrough, K. L., and G. G. Lauenstein (Editors). 2006. Major and trace element analytical methods of the National Status and Trends Program: 2000-2006. Silver Spring, MD. NOAA Technical Memorandum NOS NCCOS 29, 19 pp.
- Kimbrough, K. L., G. G. Lauenstein and W. E. Johnson (Editors). 2006. Organic Contaminant Analytical Methods of the National Status and Trends Program: Update 2000-2006. NOAA Technical Memorandum NOS NCCOS 30. 137 pp.
- Kleypas, J.A., R.A. Feely, V.J. Fabry, C. Langdon, C.L. Sabine, and L.L. Robbins. 2006. Impacts of ocean acidification on coral reefs and other marine calcifiers: a guide to future research, a report of a workshop held 18-20 April 2005. St. Petersburg, FL. Sponsored by NSF, NOAA, and the USGS. 88pp.
- Knap, A.H., J.E. Solbakken, R.E. Dodge, T.D. Sleeter, S.J. Wyers, and K.H. Palmork. 1982. Accumulation and elimination of (9-¹⁴C)phenanthrene in the reef-building coral (*Diploria strigosa*). *Bulletin of Environmental Contamination and Toxicology*. 28: 281-284.
- Konstantinou, I.K., and T.A. Albanis. 2004. World-wide occurrence and effects of antifouling paint booster biocides in the aquatic environment: a review. *Environment International* 30:235-248.
- Lauenstein, G. G. and A. Y. Cantillo (eds.) 1993. Sampling and analytical methods of the National Status and Trends Programs, National Benthic Surveillance and Mussel Watch Projects, 1984-1992. NOAA Technical Memorandum NOS ORCA 71, Vol 1.
- Lauenstein, G.G., and A.Y. Cantillo. 1998. Sampling and analytical methods of the National Status and Trends Program Mussel Watch Project: 1993-1996 update. NOAA Technical Memorandum NOS ORCA 130. 233pp.
- Long, E.R., L. J. Field, and D.D. MacDonald. 1998. Predicting toxicity in marine sediments with numerical sediment quality guidelines. *Environmental Toxicology and Chemistry*. 17(4): 714-727.
- Loya, Y. 1975. Possible effects of water pollution on the community structure of Red Sea corals. *Marine Biology*. 29: 177-185.
- Lussier, S.M., J.H. Gentile, and J. Walker. 1985. Acute and chronic effects of heavy metals and cyanide on *Mysidopsis bahia* (Crustacea:Mysidacea). *Aquatic Toxicology* 7(1-2): 25-35.
- Marubini, F., and P.S. Davies. 1996. Nitrate increases zooxanthellae population density and reduces skeletogenesis in corals. *Marine Biology*. 127: 319-328.
- McDonald, T.A. 2002. A perspective on the potential health risks of PBDEs. *Chemosphere*. 46: 745-755.
- Meerts I.A., R.J. Letcher, S. Hoving, G. Marsh, A. Bergman, J.G. Lemmen, B. van der Burg, and A. Brouwer. 2001. In vitro estrogenicity of polybrominated diphenyl ethers, hydroxylated PBDEs, and polybrominated bisphenol A compounds. *Environmental Health Perspectives*. 109: 399-407.
- Miller, G.C., C. Hoonhout, E. Sufka, S. Carroll, V. Edirveerasingam, B. Allen, J. Reuter, J. Oris and M.S. Lico. 2003. Environmental Assessment of the Impacts of Polycyclic Aromatic Hydrocarbons (PAH) in Lake Tahoe and Donner Lake. A Final Report to the California State Water Resources Board. 63p.
- Morelock, J., E.A. Winget, and C. Goenaga. 1994. Geologic maps of the southwestern Puerto Rico Par-
-

-
- guera to Guanica insular shelf. U.S. Geologic Report 2387. 6pp.
- Mortvedt, J.J. 1996. Heavy metal contaminants in inorganic and organic fertilizers. *Fertilizer Research* 43: 55-61.
- National Oceanic and Atmospheric Administration (NOAA). 1998. NS&T Program: National Status and Trends Program for Marine Environmental Quality. NOAA/National Ocean Service. 32pp.
- Neff, J.M. 1985. Polycyclic aromatic hydrocarbons. In: G.M. Rand and S.R. Petrocelli. *Fundamentals of Aquatic Toxicology*. Hemisphere Publishing Corporation. New York. 666pp.
- Neff, J.M., and J.W. Anderson. 1981. *Response of Marine Animals to Petroleum Hydrocarbons*. John Wiley and Sons, New York. 177pp.
- Neff, J.M., S.A. Stout, and D.G. Gunster. 2005. Ecological risk assessment of polycyclic aromatic hydrocarbons in sediments: identifying sources and ecological hazard. *Integrated Environmental Assessment and Management*. 1(1): 22-33.
- Negri, A.P., and A.J. Heyward. 2000. Inhibition of fertilization and larval metamorphosis of the coral *Acropora millepora* (Ehrenberg, 1834) by petroleum products. *Marine Pollution Bulletin*. 41(7-12): 420-427.
- Negri, A.P., and A.J. Heyward. 2001. Inhibition of coral fertilisation and larval metamorphosis by tributyltin and copper. *Marine Environmental Research*. 51: 17-27.
- Negri, A.P., L.D. Smith, N.S. Webster, and A.J. Heyward. 2002. Understanding ship-grounding impacts on a coral reef: potential effects of anti-foulant paint contamination on coral recruitment. *Marine Pollution Bulletin*. 44: 111-117.
- Nystrom, M., I. Nordemar, and M. Tedengren. 2001. Simultaneous and sequential stress from increased temperature and copper on the metabolism of the hermatypic coral *Porites cylindrica*. *Marine Biology*. 138: 1225-1231.
- Okamura, H., I. Aoyama, T. Takami, T. Maruyama, Y. Suzuki, M. Matsumoto, I. Katsuyama, J. Hamada, T. Beppu, O. Tanaka, R.J. Maguire, D. Liu, Y.L. Lau, and G.J. Pacepavicius. 2000. Phytotoxicity of the new antifouling compound Irgarol 1051 and a major degradation product. *Marine Pollution Bulletin*. 40(9): 754-763.
- Ott, F.S., R.P. Harris, and S.C.M. O'Hara. 1978. Acute and sublethal toxicity of naphthalene and three methylated derivatives to the estuarine copepod *Eurytemora affinis*. *Marine Environmental Research*. 1:49-58.
- Owen, R., A. Knap, M. Toaspern, and K. Carbery. 2002. Inhibition of coral photosynthesis by the antifouling herbicide Irgarol 1051. *Marine Pollution Bulletin*. 44: 623-632.
- Owen, R., A. Knap, N. Ostrander, and K. Carbery. 2003. Comparative acute toxicity of herbicides to photosynthesis of coral zooxanthellae. *Bulletin of Environmental Contamination and Toxicology*. 70: 541-548.
- Pandolfi, J.M., R.H. Bradbury, E. Sala, T.P. Hughes, K.A. Bjorndal, R.G. Cooke, D. McArdle, L. McClenachan, M.J.H. Newman, G. Paredes, R.R. Warner, and J.B.C. Jackson. 2003. Global trajectories of the long-term decline of coral reef ecosystems. *Science*. 301: 955-958.
- Peachey, R.B.J. 2003. Tributyltin and polyaromatic hydrocarbon levels in Mobile Bay, Alabama: a review. *Marine Pollution Bulletin*. 46: 1365-1371.
- Peachey, R.L., and D.G. Crosby. 1996. Phototoxicity in tropical reef animals. *Marine Environmental Research*. 42(1-4): 359-362.
- Peters, E.C., P.A. Meyers, P.P. Yevich, and N.J. Blake. 1981. Bioaccumulation and histopathological effects of oil on a stony coral. *Marine Pollution Bulletin*. 12(10): 333-339.
- Pew Oceans Commission. 2003. *America's Living Oceans: Charting a Course for Sea Change*. A Report
-

-
- to the Nation. Pew Oceans Commission, Arlington, Virginia. 144pp.
- Raloff, J. 2003. New PCBs? Science News. 164: 266-268.
- Reichelt-Brushett, A.J., and P.L. Harrison. 1999. The effect of copper, zinc and cadmium on fertilization success of gametes from scleractinian reef corals. Marine Pollution Bulletin. 38(3): 182-187.
- Reichelt-Brushett, A.J., and P.L. Harrison. 2000. The effect of copper on the settlement success of larvae from the scleractinian coral *Acropora tenuis*. Marine Pollution Bulletin. 41(7-12): 385-391.
- Reichelt-Brushett, A.J., and P.L. Harrison. 2004. Development of a sublethal test to determine the effects of copper and lead on scleractinian coral larvae. Archives of Environmental Contamination and Toxicology. 47: 40-55.
- Reichelt-Brushett, A.J., and P.L. Harrison. 2005. The effect of selected trace elements on the fertilization success of several scleractinian coral species. Coral Reefs. 24: 524-534.
- Reichelt-Brushett, A.J., and K. Michalek-Wagner. 2005. Effects of copper on the fertilization success of the soft coral *Lobophytum compactum*. Aquatic Toxicology. 74: 280-284.
- Rieuwerts, J.S., I. Thornton, M.E. Farago, and M.R. Ashmore. 1998. Factors influencing metal bioavailability in soils: preliminary investigations for the development of a critical loads approach for metals. Chemical Speciation and Bioavailability 10(2):61-75.
- Rogge, W.F., L.M. Hildeman, M.A. Mazurek, G. Cass and B.R. Simonelt. 1993. Sources of fine organic aerosol: 3. Road dust, tire debris, and organometallic brake lining dust: roads as sources and sinks. Environmental Science and Technology 27: 1892-1904.
- Rougee, L., C.A. Downs, R.H. Richmond, and G.K. Ostrander. 2006. Alteration of normal cellular profiles in the scleractinian coral (*Pocillopora damicornis*) following laboratory exposure to fuel oil. Environmental Toxicology and Chemistry. 25(12): 3181-3187.
- Saucier, D., L. Astic, and P. Rioux. 1991. The effects of early chronic exposure to sublethal copper on the olfactory discrimination of rainbow trout, *Oncorhynchus mykiss*. Environmental Biology of Fishes 30:345-351.
- Schropp, S.J., and H.L. Windom. 1988. A guide to the interpretation of metal concentrations in estuarine sediments. Florida Department of Environmental Regulation. 44pp.
- Schropp, S.J., F.G. Lewis, H.L. Windom, J.D. Ryan, F.D. Calder, and L.C. Burney. 1990. Interpretation of metal concentrations in estuarine sediments of Florida using aluminum as a reference metal. Estuaries. 13(3): 227-235.
- Shine, J., and G. Wallace. 2000. Chemical aspects of organic carbon and ecosystem stress in benthic ecosystems. In: Ad Hoc Benthic Indicator Group - results of initial planning meeting. IOC Technical Series No 57. UNESCO. 65pp.
- Singh, A.K., M.F. Gin, F. Ni and E.R. Christensen. 1993. A source-receptor method for determining non-point source of PAHs to the Milwaukee Harbor estuary. Water Science and Technology 28: 91-102.
- Smil, V. 1999. Long-Range Perspectives on Inorganic Fertilizers in Global Agriculture. 1999 Travis P. Hignett Lecture. International Fertilizer Development Center. 36pp.
- Solbakken, J.E., A.H. Knap, T.D. Sleeter, C.E. Searle, and K.H. Palmork. 1984. Investigation into the fate of 14C-labeled xenobiotics (naphthalene, phenanthrene, 2,4,5,2',4',5'-hexachlorobiphenyl, octachlorostyrene) in Bermudian corals. Marine Ecology Progress Series. 16: 149-154.
- Stoker T.E., S.C. Laws., K.M. Crofton., J.M. Hedge, J.M. Ferrell, and R.L. Cooper. 2004. Assessment of DE-71, a commercial polybrominated diphenyl ether (PBDE) mixture, in the EDSP male and female pu-
-

-
- bertal protocols. *Toxicological Science*. 78:144-155.
- Thuvander A., and P.O. Darnerud. 1999. Effects of polybrominated diphenyl ether (PBDE) and polychlorinated biphenyl (PCB) on some immunological parameters after oral exposure in rats and mice. *Toxicological and Environmental Chemistry* 79:229-42.
- U.S. Department of Agriculture. 2004. 2002 Census of agriculture - Puerto Rico volume 1. geographic area series, part 52. Report AC-02-A-52. 219pp.
- U.S. Census Bureau. 2006. United States Census 2000. U.S. Department of Commerce. Available at: <http://www.census.gov/census2000/states/pr.html>.
- U.S. Department of Human Health and Services (USDHHS). 1995. Polycyclic Aromatic Hydrocarbons Toxicology Profile. Agency for Toxic Substance and Disease Registry (ASTDR). Atlanta, GA pp.487
- U.S. Department of Human Health and Services (USDHHS). 1999. Toxicological Profile for Cadmium. Agency for Toxic Substance and Disease Registry (ASTDR). Atlanta, GA pp.439.
- U.S. Environmental Protection Agency (USEPA) 1980a. Ambient water quality criteria for chromium. EPA Report 440/5-80-035.
- U.S. Environmental Protection Agency (USEPA). 1980b. Ambient water quality criteria for nickel. EPA Report 440/5-80-060.
- U.S. Environmental Protection Agency (USEPA). 1997. Environmental Protection Agency. Management of Polychlorinated Biphenyls in the United States. Available at: <http://www.chem.unep.ch/pops>. Office of Pollution Prevention and Toxics U.S. Environmental Protection Agency. 6pp.
- U.S. Environmental Protection Agency (USEPA). 2002. Mid-Atlantic integrated assessment (MAIA) estuaries 1997-1998: summary report. EPA/620/R-02/003. 115pp.
- U.S. Environmental Protection Agency (USEPA). 2004. National coastal condition report II. EPA-620/R-03/002. Office of Research and Development and Office of Water. 286pp.
- U.S. Environmental Protection Agency (USEPA). 2005. STOrage and RETrieval data base. Available at: www.epa.gov/storet.
- U.S. Environmental Protection Agency (USEPA). 2006. Polybrominated diphenyl ethers (PBDEs) Project Plan. Available at: <http://www.epa.gov/oppt/pbde/pubs/proj-plan32906a.pdf>. 39pp.
- Valette-Silver, N.J., G.F. Riedel, E.A. Crecelius, H. Windom, R.G. Smith, and S.S. Dolvin. 1999. Elevated arsenic concentrations in bivalves from the southeast coasts of the USA. *Marine Environmental Research* 48: 311-333.
- Victor, S., and R.R. Richmond. 2005. Effect of copper on fertilization success in the reef coral *Acropora surculosa*. *Marine Pollution Bulletin*. 50: 1433-1456.
- Waddell, J.E. (ed.) 2005. The state of coral reef ecosystems of the United States and Pacific Freely Associated States: 2005. NOAA Technical Memorandum NOS NCCOS 11. NOAA/NCCOS Center for Coastal Monitoring and Assessment's Biogeography Team. Silver Spring, MD. 522pp.
- Wallinder, I.O., S. Bertling, G. Herting, C. Leygraf, D. Heijerick, D. Berggren, and P. Koundakjian. 2004. Release of chromium, nickel and iron from stainless steel exposed under atmospheric conditions and the environmental interaction of these metals. A combined field and laboratory investigation. European Confederation of Iron and Steel Industries. 53pp.
- Weisberg., S.B. H.T. Wilson, D.G. Heimbuch, H.L. Windom, and J.K. Summers. 2000. Comparison of sediment metal: aluminum relationships between the eastern and Gulf coasts of the United States. *Environmental Monitoring and Assessment*. 61: 373-385.
- Windom, H.L., S.J. Schropp, F.D. Calder, J.D. Ryan, R.G. Smith, L.C. Burney, F.G. Lewis, and C.H. Rawlinson. 1989. Natural trace metal concentrations in estuarine and coastal marine sediments of the south-
-

eastern United States. *Environmental Science and Technology*. 3: 314-320.

Wong, C. K., K. H. Chu, K. W. Tang, T. W. Tam, and L. J. Wong. 1993. Effects of chromium, copper and nickel on survival and feeding behavior of *Metapenaeus ensis* larvae and postlarvae (Decapoda:Penaeidae). *Marine Environmental Research* 36:63-78.

Zeng, E.Y., and C.L. Vista. 1997. Organic pollutants in the coastal environment off San Diego, California. 1. Source identification and assessment by compositional indices of polycyclic aromatic hydrocarbons. *Environmental Toxicology and Chemistry*. 16(2):179-188.

Zhou T., D.G. Ross, M.J. DeVito, and K.M. Crofton 2001. Effects of short-term in vivo exposure to polybrominated diphenyl ethers on thyroid hormones and hepatic enzyme activities in weanling rats. *Toxicological Science*. 61:76-82.

Appendix A. Sediment sample sites from May 2005 mission.



Abbreviations: PRBR, backreef sites; PRBS, bankshelf sites; PRMG, mangrove sites; PRLG, lagoon sites.

Appendix B. Field data from May 2005 mission.

Site	Collection Date	Habitat	Latitude (DD)	Longitude (DD)	Water Depth (m)	Dissolved Oxygen (mg/L)	Temperature (°C)	Salinity (ppt)	Conductivity (mS)	Secchi Depth (m)
PRMG1	5/26/2005	Mangrove	N17.947490	W067.115060	0.6	4.44	31.3	34.2	58.5	bottom
PRMG2	5/26/2005	Mangrove	N17.946600	W067.123610	0.6	3.40	31.4	34.4	59.0	bottom
PRMG3	5/26/2005	Mangrove	N17.953350	W067.089000	0.3	5.34	31.9	34.1	59.0	bottom
PRMG7	5/25/2005	Mangrove	N17.971420	W067.072660	2.7	5.35	31.9	33.7	58.2	bottom
PRMG8	5/23/2005	Mangrove	N17.969690	W067.042400	1.5	3.89	31.0	32	55.5	bottom
PRMG9	5/25/2005	Mangrove	N17.973070	W067.070590	0.9	4.82	31.9	33.8	58.7	bottom
PRMG13	5/23/2005	Mangrove	N17.970200	W066.999830	1.2	5.29	31.1	33.3	57.0	bottom
PRMG14	5/26/2005	Mangrove	N17.964000	W066.985630	0.9	3.16	30.3	34.7	58.2	bottom
PRMG15	5/26/2005	Mangrove	N17.968040	W067.019200	0.6	3.95	30.4	34.6	52.7	bottom
PRMG19	5/24/2005	Mangrove	N17.963620	W066.928380	1.5	4.77	30.3	14.4	35.2	0.8
PRMG21	5/24/2005	Mangrove	N17.970560	W066.921920	1.5	4.12	30.2	24.2	41.9	0.9
PRMG24	5/24/2005	Mangrove	N17.964090	W066.912160	1.8	5.29	31.1	32.1	54.4	1.5
PRLG1	5/25/2005	Lagoon	N17.942687	W067.099647	4.9	5.12	30.2	34.1	57.2	bottom
PRLG3	5/26/2005	Lagoon	N17.952112	W067.087452	1.2	5.20	30.5	34.1	57.5	bottom
PRLG4	5/25/2005	Lagoon	N17.956989	W067.082784	1.8	5.81	32.2	33.6	58.5	bottom
PRLG6	5/25/2005	Lagoon	N17.931980	W067.127456	8.5	5.30	30.3	34.1	57.3	bottom
PRLG7	5/25/2005	Lagoon	N17.945519	W067.092832	6.7	5.06	30.4	34.1	57.3	bottom
PRLG8	5/25/2005	Lagoon	N17.942424	W067.116750	5.2	4.92	31.6	33.6	51.0	bottom
PRLG17	5/25/2005	Lagoon	N17.961227	W067.074486	5.8	6.01	31.9	33.5	58.0	bottom
PRLG18	5/25/2005	Lagoon	N17.970818	W067.063506	6.1	6.81	30.9	33.8	57.4	5.8
PRLG19	5/23/2005	Lagoon	N17.963663	W067.040107	5.5	5.40	30.2	33.6	56.4	bottom
PRLG20	5/25/2005	Lagoon	N17.965985	W067.062499	7.0	5.24	31.5	33.8	51.5	bottom
PRLG27	5/23/2005	Lagoon	N17.959831	W066.991050	7.9	5.54	30.0	33.3	55.9	4.9
PRLG28	5/23/2005	Lagoon	N17.956087	W066.999836	13.4	5.39	29.9	33.4	55.2	7.6
PRLG29	5/23/2005	Lagoon	N17.967738	W067.017806	3.0	5.83	30.9	33.6	56.9	3.0
PRLG30	5/25/2005	Lagoon	N17.955862	W066.998984	13.1	5.08	29.3	34.5	57.0	7.0
PRLG37	5/24/2005	Lagoon	N17.962632	W066.925094	3.0	4.24	29.5	32.3	55.0	1.2
PRLG38	5/24/2005	Lagoon	N17.959960	W066.917055	2.7	5.06	30.1	32.4	54.7	1.5
PRLG39	5/24/2005	Lagoon	N17.961028	W066.920218	3.0	4.37	29.9	32.9	55.1	0.9
PRBR1	5/25/2005	Back Reef	N17.923550	W067.114357	2.7	5.53	30.6	34.2	57.4	bottom
PRBR2	5/25/2005	Back Reef	N17.935098	W067.095237	1.2	5.37	30.5	34.3	57.9	bottom
PRBR3	5/25/2005	Back Reef	N17.932738	W067.092546	1.2	5.39	31.4	34.1	58.3	bottom
PRBR7	5/24/2005	Back Reef	N17.942606	W067.060851	1.8	6.85	30.7	34.2	58.2	bottom

Appendix B. Field data from May 2005 mission (cont.).

Site	Collection Date	Habitat	Latitude (DD)	Longitude (DD)	Water Depth (m)	Dissolved Oxygen (mg/L)	Temperature (°C)	Salinity (ppt)	Conductivity (mS)	Secchi Depth (m)
PRBR12	5/25/2005	Back Reef	N17.955912	W067.048727	1.5	5.21	30.4	34.4	52.3	bottom
PRBR13	5/24/2005	Back Reef	N17.945716	W067.009205	1.2	5.66	30.9	34.2	58.0	bottom
PRBR14	5/24/2005	Back Reef	N17.944234	W067.007594	0.9	6.13	31.0	34.5	58.6	bottom
PRBR15	5/24/2005	Back Reef	N17.947177	W067.011145	1.5	5.57	30.6	33.9	51.5	bottom
PRBR25	5/24/2005	Back Reef	N17.928947	W066.945380	2.1	5.68	30.1	33.9	56.8	bottom
PRBR26	5/24/2005	Back Reef	N17.927524	W066.950354	4.6	6.58	30.6	33.9	51.6	bottom
PRBR27	5/24/2005	Back Reef	N17.928358	W066.952837	4.0	6.62	30.4	34	58.0	bottom
PRBS1	5/23/2005	Bank Shelf	N17.953140	W067.067396	18.6	5.33	30.5	32.6	55.0	8.8
PRBS2	5/23/2005	Bank Shelf	N17.927291	W067.039238	17.4	5.49	29.4	33.9	56.3	14.6
PRBS3	5/23/2005	Bank Shelf	N17.944686	W067.074537	18.6	5.79	30.0	33.4	55.9	9.8
PRBS4	5/24/2005	Bank Shelf	N17.958692	W067.028657	12.2	5.51	30.1	34.1	57.2	8.2
PRBS5	5/25/2005	Bank Shelf	N17.933401	W067.030351	18.9	5.30	29.3	34.5	56.9	11.3
PRBS7	5/24/2005	Bank Shelf	N17.947373	W067.027537	16.8	5.50	29.6	34.6	57.3	7.6
PRBS8	5/24/2005	Bank Shelf	N17.946996	W067.039631	17.7	5.63	29.5	34.5	57.2	9.4
PRBS9	5/25/2005	Bank Shelf	N17.955879	W067.058907	15.2	5.21	29.9	34.2	57.0	9.4
PRBS16	5/25/2005	Bank Shelf	N17.959484	W067.053296	18.3	5.13	30.9	34.2	58.1	9.4
PRBS20	5/25/2005	Bank Shelf	N17.959799	W067.048179	16.8	5.20	30.3	34.3	57.6	8.5
PRBS24	5/24/2005	Bank Shelf	N17.941551	W067.031722	12.2	5.65	29.7	33.9	56.5	11.3
PRBS29	5/23/2005	Bank Shelf	N17.942431	W066.991808	15.8	5.18	29.7	33.4	55.8	7.6
PRBS30	5/23/2005	Bank Shelf	N17.930319	W067.014447	18.9	5.36	29.7	33.8	56.1	10.7
PRBS31	5/23/2005	Bank Shelf	N17.942497	W066.977832	13.7	5.19	29.6	33.7	51.5	8.8
PRBS32	5/25/2005	Bank Shelf	N17.939746	W067.025056	17.4	5.31	28.3	34.5	56.9	8.2
PRBS33	5/24/2005	Bank Shelf	N17.948607	W067.010185	12.2	5.39	30.0	34.5	57.4	7.9
PRBS34	5/25/2005	Bank Shelf	N17.923034	W067.021467	22.6	5.39	28.2	34.5	56.7	14.3
PRBS35	5/24/2005	Bank Shelf	N17.960042	W067.015735	15.8	5.86	30.2	34.2	57.3	7.6
PRBS36	5/24/2005	Bank Shelf	N17.952984	W067.002496	12.2	5.60	31.2	33.7	52.2	8.2
PRBS40	5/24/2005	Bank Shelf	N17.934368	W066.976107	12.2	5.44	29.6	34.5	57.2	11.0
PRBS41	5/25/2005	Bank Shelf	N17.948125	W067.000005	15.8	4.99	29.4	34.3	56.9	9.4
PRBS43	5/25/2005	Bank Shelf	N17.931560	W066.992106	16.5	5.11	28.1	33.9	55.8	6.1
PRBS60	5/25/2005	Bank Shelf	N17.936707	W066.986643	16.8	5.18	29.2	34.4	56.7	9.4
PRBS62	5/25/2005	Bank Shelf	N17.954316	W067.012856	17.7	5.15	30.2	34.2	57.3	7.9
PRBS63	5/24/2005	Bank Shelf	N17.918851	W066.949879	13.7	5.10	28.9	34	55.8	9.4

Appendix C. Total organic carbon and grain size distribution in sediment samples from southwest Puerto Rico, May 2005 mission.

Site	TOC %	Gravel (%)	Sand (%)	Silt (%)	Clay (%)	Site	TOC %	Gravel (%)	Sand (%)	Silt (%)	Clay (%)
PRMG-8	3.74	0.75	37.62	48.02	13.61	PRMG-3	2.89	32.82	34.42	2.58	30.18
PRMG-13	3.02	2.72	44.16	41.46	11.65	PRMG-7	2.54	4.75	54.98	28.30	11.96
PRMG-19	4.16	6.16	48.74	38.24	6.86	PRMG-9	2.70	1.87	27.91	54.96	15.25
PRMG-21	2.69	1.09	39.94	48.26	10.71	PRMG-14	3.31	3.64	43.86	41.33	11.16
PRMG-24	1.91	0.04	22.35	69.56	8.06	PRMG-15	1.61	12.88	71.73	7.87	7.51
PRBS-3	0.64	3.77	64.81	29.03	2.39	PRBS-35	0.57	0.07	7.55	88.63	3.75
PRBS-5	0.97	1.58	60.08	37.01	1.33	PRBS-36	0.87	0.22	5.77	89.40	4.61
PRBS-9	0.60	3.20	39.22	54.93	2.66	PRBS-40	0.50	17.34	70.50	10.25	1.91
PRBS-10	0.37	3.32	93.45	1.10	2.13	PRBS-63	2.90	n/a	n/a	n/a	n/a
PRBS-16	1.34	0.00	7.32	89.09	3.59	PRBS-65	0.50	n/a	n/a	n/a	n/a
PRBS-20	0.78	0.00	5.16	91.61	3.23	PRBS-66	2.29	n/a	n/a	n/a	n/a
PRBS-29	1.03	34.43	35.09	28.33	2.15	PRBS-67	0.40	2.29	59.82	35.73	2.16
PRBS-31	0.29	0.02	34.90	64.44	1.65	PRBS-69	0.29	0.77	58.99	37.72	2.51
PRBS-32	0.52	1.69	46.65	46.88	4.77	PRBS-73	1.39	14.48	82.08	1.27	2.18
PRBS-34	0.23	0.16	53.03	43.16	1.65	PRLG-19	0.66	4.76	65.03	26.95	3.26
PRBS-41	0.92	0.00	4.50	90.64	4.86	PRLG-27	0.96	3.09	48.59	44.25	4.08
PRBS-43	0.80	29.28	65.91	3.51	1.30	PRLG-28	0.81	0.05	4.57	90.75	4.62
PRBS-60	0.28	0.50	64.72	33.24	1.54	PRLG-37	2.14	0.00	12.29	74.84	12.87
PRBS-62	0.90	0.07	5.79	90.57	3.57	PRLG-38	2.01	39.31	40.95	16.66	3.08
PRBR-2	0.72	0.11	98.56	1.31	0.02	PRLG-39	1.86	49.87	35.03	12.91	2.19
PRBR-3	0.50	3.38	88.66	6.56	1.40	PRBR-7	3.41	4.20	90.06	4.04	1.70
PRBR-8	0.29	n/a	n/a	n/a	n/a	PRBR-9	3.08	6.46	87.49	4.63	1.42
PRBR-12	0.34	0.89	93.38	3.59	2.15	PRBR-13	1.56	0.08	93.48	6.22	0.22
PRLG-1	1.06	1.46	51.71	41.85	4.98	PRBR-14	1.89	3.19	91.00	4.61	1.20
PRLG-3	2.24	10.26	28.96	43.63	17.14	PRBR-15	0.36	0.18	82.97	15.68	1.17
PRLG-4	2.77	23.63	27.94	37.32	11.11	PRBR-25	0.54	0.09	97.42	1.43	1.07
PRLG-6	0.74	14.39	55.87	24.71	5.02	PRBR-26	1.57	26.58	69.46	2.66	1.30
PRLG-7	1.80	33.94	47.97	14.58	3.51	PRBR-27	0.55	11.36	85.18	2.42	1.04
PRLG-8	2.08	17.07	27.74	42.81	12.38	PRBS-1	0.63	2.46	16.93	77.49	3.12
PRLG-17	1.97	16.37	35.19	40.95	7.50	PRBS-2	0.23	0.21	78.17	20.33	1.29
PRLG-18	1.18	8.52	44.93	40.65	5.90	PRBS-4	NA	13.14	56.86	28.30	1.70
PRLG-20	1.25	5.86	50.96	36.37	6.82	PRBS-7	0.93	22.50	30.90	44.41	2.19
PRLG-29	2.48	0.12	24.84	63.17	11.87	PRBS-8	1.03	1.21	32.45	63.70	2.64
PRLG-30	0.80	0.02	4.79	89.23	5.96	PRBS-30	0.48	0.30	76.30	21.66	1.74
PRMG-1	3.47	7.91	67.71	16.79	7.58	PRBS-33	0.51	3.48	46.92	47.54	2.06
PRMG-2	6.45	15.75	38.57	22.47	23.21						

Appendix D. Total organic carbon and grain size distribution in sediment samples from southwest Puerto Rico, August 2005 mission.

Site	TOC %	TIC %	Gravel (%)	Sand (%)	Silt (%)	Clay (%)
Coral 1	1.47	10.06	1.48	97.13	0.00	1.55
Coral 2	4.13	7.36	0.67	98.23	0.00	1.35
Coral 3	1.05	10.35	9.37	82.02	5.86	2.76
Coral 4	2.51	8.78	16.74	58.88	20.94	3.44
Coral 5	0.45	10.46	1.06	65.93	28.84	4.17
Coral 6	0.45	10.82	7.63	72.63	16.91	2.83
Coral 8	0.57	10.89	12.73	64.49	19.38	3.40
Coral 9	0.76	10.76	6.65	76.83	12.92	3.60
LTOC1	0.94	10.84	28.96	67.70	1.41	1.93
LTOC2	0.20	11.62	0.70	95.57	1.33	2.40
LTOC3	0.13	11.76	0.03	97.30	0.49	2.18
LTOC7	0.86	9.66	0.05	12.04	82.04	5.86
LTOC8	0.26	10.87	0.15	54.45	41.33	4.07
LTOC9	0.43	10.35	0.53	19.67	73.93	5.87
LTOC10	0.25	11.07	0.95	70.07	23.79	5.19
LTOC11	0.37	10.74	0.14	43.52	52.83	3.51
LTOC12	2.78	8.99	7.21	86.17	3.58	3.04
LTOC13	2.17	9.59	13.40	84.67	0.21	1.72
LTOC14	0.91	10.73	3.12	90.31	3.53	3.03
LTOC15	0.75	9.63	0.12	21.76	73.51	4.61
LTOC16	1.25	9.70	34.07	36.76	25.25	3.92
LTOC17	2.64	8.90	0.08	96.39	0.94	2.59
LTOC18	0.51	11.26	2.20	93.29	2.22	2.29
LTOC19	1.35	9.82	18.26	50.40	26.77	4.57
LTOC22	0.63	9.70	0.13	14.81	80.13	4.93
HTOC1	0.63	11.04	0.07	95.79	2.20	1.93
HTOC2	4.99	6.73	11.43	87.93	0.00	0.92
HTOC3	1.06	10.43	6.72	50.82	38.00	4.47
HTOC4	0.44	10.97	2.01	50.23	43.32	4.45
HTOC6	0.84	10.01	4.26	41.55	49.72	4.47
HTOC7	0.88	10.85	2.98	91.49	6.17	0.00
HTOC8	1.49	10.23	0.70	81.71	13.18	4.41
HTOC9	0.96	10.86	6.18	54.75	33.80	5.26
HTOC10	0.28	11.35	0.44	98.76	0.00	1.22
HTOC12	0.65	11.37	2.45	92.98	1.96	2.60
HTOC14	0.76	11.03	5.16	88.76	2.26	3.82
HTOC15	2.65	9.18	20.15	77.89	0.74	1.22
HTOC16	1.04	9.40	0.04	14.49	81.30	4.18
HTOC17	0.99	4.46	0.41	2.64	91.06	5.89
HTOC19	1.61	1.22	0.00	0.78	86.92	12.30
HTOC20	2.16	10.14	11.84	46.54	33.61	8.01
HTOC21	0.19	11.49	1.24	83.33	10.80	4.63
HTOC22	0.80	9.86	0.06	4.76	89.57	5.61

Abbreviations: TOC, total organic carbon; TIC, total inorganic carbon

Appendix E. Polycyclic aromatic hydrocarbons (PAHs) in sediment samples from southwest Puerto Rico (ng/dry g).

Compound	Coral 1	Coral 2	Coral 3	Coral 4	Coral 5	Coral 6	Coral 8	Coral 9	LTOC1	LTOC2
Decalin	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Naphthalene	0.4	0.3	1.5	2.3	12.5	3.3	0.9	0.3	0.2	0.3
C1-Naphthalenes	0.2 J	0.1 J	0.3 J	0.5	1.3	0.6	0.3 J	0.2 J	0.1 J	0.1 J
C2-Naphthalenes	0.0 U	0.2 J	0.7	1.0	1.9	1.1	0.8	0.3 J	0.3 J	0.3 J
C3-Naphthalenes	0.0 U	0.0 U	0.0 U	0.0 U	1.0	0.8	0.0 U	0.0 U	0.0 U	0.0 U
C4-Naphthalenes	0.0 U	0.0 U	0.0 U	0.0 U	1.3	1.4	0.0 U	0.0 U	0.0 U	0.0 U
Benzothiophene	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Biphenyl	0.2	0.2	0.5	0.6	2.0	0.8	0.3	0.2	0.2	0.2
Acenaphthylene	0.1 J	0.0 U	0.3	0.5	3.0	0.7	0.2	0.0 U	0.1 J	0.0 U
Acenaphthene	0.1 J	0.0 U	0.3	0.2	1.0	0.3	0.0 U	0.0 U	0.1 J	0.1 J
Dibenzofuran	0.1 J	0.1 J	0.4	0.5	2.0	0.8	0.3	0.1 J	0.2 J	0.2 J
Fluorene	0.2	0.0 U	1.1	0.6	1.3	1.7	0.7	0.1 J	0.2	0.3
C1-Fluorenes	0.0 U	0.0 U	0.5	0.5	0.7	0.9	0.5	0.0 U	0.0 U	0.0 U
C2-Fluorenes	0.0 U	0.0 U	1.8	1.6	1.8	2.5	1.9	0.0 U	0.0 U	0.0 U
C3-Fluorenes	0.0	0.0	0.0	0.0	0.0	2.8	0.0	0.0	0.0	0.0
Carbazole	0.0 U	0.0 U	0.0 U	0.0 U	0.3 J	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Anthracene	0.0 U	0.0 U	0.4	0.6	2.9	1.0	0.2	0.1 J	0.0 U	0.0 U
Phenanthrene	0.3	0.2	2.5	3.0	14.2	5.2	1.5	0.5	0.3	0.5
C1-Phenanthrene/Anthracenes	0.3	0.1 J	0.8	0.8	3.1	1.6	0.7	0.3	0.2 J	0.3
C2-Phenanthrene/Anthracenes	0.4	0.4	1.1	1.3	2.3	1.8	0.9	0.7	0.0 U	0.0 U
C3-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	0.8	1.5	1.1	0.0 U	0.0 U	0.0 U	0.0 U
C4-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Dibenzothiophene	0.0 U	0.0 U	0.1 J	0.1 J	0.3	0.1 J	0.1 J	0.0 U	0.0 U	0.0 U
C1-Dibenzothiophenes	0.0 U	0.0 U	0.2 J	0.2 J	0.5	0.3 J	0.2 J	0.0 U	0.0 U	0.0 U
C2-Dibenzothiophenes	0.0 U	0.0 U	0.4	0.5	0.8	0.5	0.0 U	0.0 U	0.0 U	0.0 U
C3-Dibenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.8	0.5	0.0 U	0.0 U	0.0 U	0.0 U
Fluoranthene	0.3	0.1 J	2.5	3.4	19	6.7	1.6	0.4	0.2 J	0.2 J
Pyrene	0.2	0.0 U	3.1	4.1	24	7.9	1.7	0.4	0.2	0.2
C1-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.4	0.7	2.1	1.1	0.3 J	0.0 U	0.0 U	0.0 U
C2-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.6	0.4	1.4	1.9	0.0 U	0.0 U	0.0 U	0.0 U
C3-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.0 U	0.0 U	0.4	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Naphthobenzothiophene	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benz(a)anthracene	0.1 J	0.0 U	0.4	0.6	2.0	1.0	0.3	0.1 J	0.0 U	0.1 J
Chrysene	0.1 J	0.0 U	0.6	0.8	2.8	1.4	0.4	0.1 J	0.0 U	0.1 J
C1-Chrysenes	0.0 U	0.0 U	0.2 J	0.4	0.9	0.6	0.0 U	0.0 U	0.0 U	0.0 U
C2-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	1.0	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(b)fluoranthene	0.2 J	0.0 U	1.2	1.4	4.8	2.8	0.9	0.3	0.0 U	0.0 U
Benzo(k)fluoranthene	0.1 J	0.0 U	0.4	0.4	1.5	0.8	0.2 J	0.1 J	0.0 U	0.0 U
Benzo(e)pyrene	0.1 J	0.0 U	0.7	0.8	3.4	1.7	0.5	0.1 J	0.0 U	0.0 U
Benzo(a)pyrene	0.1 J	0.0 U	0.9	1.1	4.3	2.0	0.6	0.2 J	0.0 U	0.0 U
Perylene	0 U	0.0 U	0.3 J	0.1 J	1.2 J	0.5 J	0.1 J	0.0 U	0.0 U	0.0 U
Indeno(1,2,3-c,d)pyrene	0.1 J	0.0 U	1.1	1.2	4.3	2.3	0.8	0.2 J	0.0 U	0.0 U
Dibenzo(a,h)anthracene	0.0 U	0.0 U	0.0 U	0.0 U	0.4	0.3	0.0 U	0.0 U	0.0 U	0.0 U
C1-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(g,h,i)perylene	0.1 J	0.0 U	2.1	2.3	9.2	4.9	1.5	0.5	0.0 U	0.1 J
Total PAHs	3.7	1.7	27.4	33.3	139	65.7	18.4	5.2	2.3	3.0
Individual Isomers*										
2-Methylnaphthalene	0.2 J	0.1 J	0.3	0.5	1.4	0.7	0.3	0.2 J	0.1 J	0.1 J
1-Methylnaphthalene	0.1 J	0.1 J	0.2	0.3	0.7	0.3	0.2	0.1 J	0.1 J	0.1 J
2,6-Dimethylnaphthalene	0.0 U	0.1 J	0.7	1.0	2.2	1.0	0.9	0.1 J	0.2 J	0.1 J
1,6,7-Trimethylnaphthalene	0.0 U	0.0 U	0.0 U	0.0 U	0.1	0.1	0.0 U	0.0 U	0.0 U	0.0 U
1-Methylphenanthrene	0.1 J	0.0 U	0.2 J	0.2 J	0.5	0.4	0.2 J	0.1 J	0.1 J	0.1 J

Notes: J, below method detection level, MDL; U, not detected

*Individual isomers contained in alkylated (C1-C4 sums)

Appendix E. Polycyclic aromatic hydrocarbons (PAHs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC3	LTOC7	LTOC8	LTOC9	LTOC10	LTOC11	LTOC12	LTOC13	LTOC14	LTOC15	LTOC16
Decalin	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Naphthalene	0.3	16.4	1.4	3.6	0.7	1.2	0.4	0.5	0.5	7.3	1.2
C1-Naphthalenes	0.1 J	1.8	0.4	0.6	0.3 J	0.3 J	0.2 J	0.2 J	0.2 J	1.1	0.5
C2-Naphthalenes	0.3 J	2.9	0.8	1.5	0.4	0.5	0.3 J	0.3 J	0.3 J	1.7	0.7
C3-Naphthalenes	0.0 U	1.6	0.0 U	1.3	0.0 U	0.7	0.0 U	0.0 U	0.0 U	1.4	0.8
C4-Naphthalenes	0.0 U	2.2	0.0 U	2.0	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.7	0.0 U
Benzothiophene	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Benzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Biphenyl	0.2	2.7	0.4	0.9	0.3	0.4	0.2	0.3	0.3	1.5	0.4
Acenaphthylene	0.0 U	3.7	0.3	0.9	0.2	0.3	0.0 U	0.0 U	0.0 U	2.1	0.3
Acenaphthene	0.0 U	1.5	0.1 J	0.3	0.0 U	0.0 U	0.0 U	0.1 J	0.0 U	0.7	0.2
Dibenzofuran	0.2 J	2.1	0.4	0.7	0.0 U	0.2 J	0.1 J	0.2 J	0.0 U	1.2	0.3
Fluorene	0.1 J	1.4	0.9	1.7	0.3	0.3	0.1 J	0.1 J	0.2	1.2	0.6
C1-Fluorenes	0.0 U	1.3	0.7	1.1	0.4	0.4	0.2 J	0.3 J	0.0 U	1.0	0.7
C2-Fluorenes	0.0 U	1.8	0.0 U	1.9	0.0 U	1.0	0.7	0.0 U	0.0 U	2.4	1.7
C3-Fluorenes	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0	3.4	0.0
Carbazole	0.0 U	0.4	0.0 U	0.1 J	0.1 J	0.1 J	0.1 J	0.0 U	0.0 U	0.2 J	0.1 J
Anthracene	0.0 U	3.3	0.3	1.4	0.2	0.3	0.0 U	0.0 U	0.0 U	2.6	0.4
Phenanthrene	0.3	15.7	1.6	5.9	1.1	1.5	0.2	0.3	0.3	10.2	1.8
C1-Phenanthrene/Anthracenes	0.2 J	4.5	0.8	2.4	0.7	1.0	0.2 J	0.3	0.2 J	4.6	1.0
C2-Phenanthrene/Anthracenes	0.0 U	3.9	1.2	2.7	1.5	1.4	0.0 U	0.0 U	0.6	3.2	1.3
C3-Phenanthrene/Anthracenes	0.0 U	3.0	0.0 U	2.2	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	2.6	0.0 U
C4-Phenanthrene/Anthracenes	0.0 U	1.1	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Dibenzothiophene	0.0 U	0.4	0.1 J	0.2	0.1 J	0.1 J	0.0 U	0.0 U	0.0 U	0.3	0.0 U
C1-Dibenzothiophenes	0.0 U	0.9	0.2 J	0.5	0.2 J	0.2 J	0.0 U	0.0 U	0.0 U	0.6	0.2 J
C2-Dibenzothiophenes	0.0 U	1.5	0.4	1.0	0.2 J	0.4	0.0 U	0.0 U	0.0 U	1.1	0.4
C3-Dibenzothiophenes	0.0 U	1.6	0.0 U	1.0	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.1	0.0 U
Fluoranthene	0 J	21.2	2	11	1.4	1.9	0.1 J	0.1 J	0.1 J	20	2.6
Pyrene	0 J	27.0	2	12	1.3	2.3	0.2	0.1 J	0.1 J	26	2.9
C1-Fluoranthenes/Pyrenes	0.0 U	3.7	1	2.2	0.5	0.8	0.0 U	0.0 U	0.1 J	3.2	0.7
C2-Fluoranthenes/Pyrenes	0.0 U	2.6	0.6	1.7	0.6	0.6	0.0 U	0.0 U	0.0 U	2.2	0.0 U
C3-Fluoranthenes/Pyrenes	0.0 U	1.5	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.9	0.0 U
Naphthobenzothiophene	0.0 U	0.8	0.0 U	0.5	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.8	0.0 U
C1-Naphthobenzothiophenes	0.0 U	1.4	0.0 U	1.0	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.3	0.0 U
C2-Naphthobenzothiophenes	0.0 U	2.1	0.0 U	1.5	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.9	0.0 U
C3-Naphthobenzothiophenes	0.0 U	1.5	0.0 U	1.1	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benz(a)anthracene	0.0 U	2.9	0	2.1	0.6	0.5	0.0 U	0.0 U	0.0 U	2.6	0.5
Chrysene	0 U	2.9	0	2	0.6	0.7	0.1 J	0.0 U	0.0 U	4.0	0.7
C1-Chrysenes	0.0 U	1.6	0	1.4	0.4	0.6	0.0 U	0.0 U	0.0 U	1.8	0.5
C2-Chrysenes	0.0 U	1.8	0.0 U	1.6	0.0 U	0.7	0.0 U	0.0 U	0.0 U	1.9	0.8
C3-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(b)fluoranthene	0 U	4.5	1	4	1.4	1.2	0.0 U	0 U	0.0 U	6.8	1.4
Benzo(k)fluoranthene	0.0 U	1.1	0.2 J	1.0	0.5	0.4	0.0 U	0.0 U	0.0 U	2.2	0.4
Benzo(e)pyrene	0.0 U	2.9	0.4	2.4	0.8	0.7	0.0 U	0.0 U	0.0 U	4.8	0.8
Benzo(a)pyrene	0.0 U	3.5	0.4	2.7	1.1	0.8	0.0 U	0.0 U	0.0 U	5.6	1.1
Perylene	0.0 U	1.1 J	0.2 J	1 J	0 J	0.4 J	0.0 U	0.0 U	0.0 U	2.1	0.5 J
Indeno(1,2,3-c,d)pyrene	0.0 U	3.4	0.6	3.4	1.0	0.8	0.1 J	0.0 U	0.0 U	6.3	1.3
Dibenzo(a,h)anthracene	0.0 U	0.5	0.0 U	0.4	0.2	0.1 J	0.0 U	0.0 U	0.0 U	0.3	0.2
C1-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(g,h,i)perylene	0.0 U	6.1	0.9	5.6	1.2	1.2	0.1 J	0.0 U	0.1 J	13.5	2.3
Total PAHs	2	170	19	93	18.7	24.0	3.3	2.8	3.0	161	29.3
Individual Isomers*											
2-Methylnaphthalene	0.1 J	1.8	0.4	0.7	0.3	0.3	0.2 J	0.2 J	0.2 J	1.2	0.5
1-Methylnaphthalene	0.1 J	1.1	0.2	0.3	0.2	0.2	0.1 J	0.1 J	0.1 J	0.6	0.3
2,6-Dimethylnaphthalene	0.1 J	3.0	0.5	1.4	0.2 J	0.3	0.1 J	0.1 J	0.1 J	1.7	0.5
1,6,7-Trimethylnaphthalene	0.0 U	0.1	0.0 U	0.1	0.0 U	0.1	0.0 U	0.0 U	0.0 U	0.2	0.1
1-Methylphenanthrene	0.1 J	0.7	0.2 J	0.4	0.2 J	0.2 J	0.1 J	0.1 J	0.1 J	0.6	0.2 J

Notes: J, below method detection level, MDL; U, not detected

*Individual isomers contained in alkylated (C1-C4 sums)

Appendix E. Polycyclic aromatic hydrocarbons (PAHs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC17	LTOC18	LTOC19	LTOC22	HTOC1	HTOC2	HTOC3	HTOC4	HTOC6	HTOC7	HTOC8
Decalin	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Naphthalene	0.3	0.3	0.7	7.9	0.3	0.3	5.8	1.7	3.4	0.6	0.9
C1-Naphthalenes	0.2 J	0.1 J	0.3 J	1.2	0.4	0.1 J	1.1	0.5	0.9	0.5	0.3 J
C2-Naphthalenes	0.2 J	0.2 J	0.4	1.5	0.7	0.2 J	2.2	1.0	2.1	1.3	0.5
C3-Naphthalenes	0.0 U	0.0 U	0.0 U	1.2	1.5	0.0 U	1.8	0.6	3.2	5.0	0.0 U
C4-Naphthalenes	0.0 U	0.0 U	0.0 U	1.6	1.4	0.0 U	2.8	1.1	5.9	12.9	0.0 U
Benzo(b)thiophene	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Benzo(b)thiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Benzo(b)thiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Benzo(b)thiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Biphenyl	0.2	0.2	0.3	1.5	0.2	0.2	1.3	0.4	0.8	0.3	0.2
Acenaphthylene	0.0 U	0.0 U	0.1 J	2.1	0.0 U	0.0 U	1.4	0.3	0.8	0.1 J	0.1 J
Acenaphthene	0.1 J	0.1 J	0.2	0.6	0.0 U	0.0 U	0.4	0.1 J	0.3	0.1 J	0.1 J
Dibenzofuran	0.1 J	0.1 J	0.2 J	1.2	0.1 J	0.1 J	1.2	0.4	0.8	0.3	0.2 J
Fluorene	0.1 J	0.1 J	0.3	1.2	0.1 J	0.1 J	3.1	0.8	2.0	0.4	0.3
C1-Fluorenes	0.2 J	0.2 J	0.5	1.0	0.5	0.0 U	1.6	0.7	2.0	2.5	0.3 J
C2-Fluorenes	0.5	0.0 U	0.0 U	2.1	1.3	0.0 U	5.0	1.4	4.6	5.4	0.7
C3-Fluorenes	0.0	0.0	0.0	0.0	1.2	0.0	3.6	0.0	3.9	0.0	0.0
Carbazole	0.0 U	0.0 U	0.0 U	0.2 J	0.0 U	0.0 U	0.3 J	0.0 U	0.3 J	0.0 U	0.0 U
Anthracene	0.0 U	0.0 U	0.2	2.6	0.1 J	0.0 U	2.7	0.5	1.3	0.2	0.2
Phenanthrene	0.3	0.2	0.9	9.2	0.5	0.1 J	9.0	2.7	6.7	2.8	1.0
C1-Phenanthrene/Anthracenes	0.2 J	0.2 J	0.7	4.7	1.8	0.0 U	4.3	1.0	3.6	4.0	0.4
C2-Phenanthrene/Anthracenes	0.5	0.5	0.9	2.9	2.8	0.0 U	4.1	1.3	4.2	3.9	0.5
C3-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	3.2	1.6	0.0 U	0.0 U	0.0 U	4.0	0.0 U	0.0 U
C4-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	0.8	0.6	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Dibenzothiophene	0.0 U	0.0 U	0.1 J	0.2	0.0 U	0.0 U	0.3	0.1 J	0.5	0.6	0.0 U
C1-Dibenzothiophenes	0.0 U	0.0 U	0.2 J	0.6	0.2 J	0.0 U	0.6	0.3 J	1.5	0.0 U	0.0 U
C2-Dibenzothiophenes	0.0 U	0.0 U	0.3 J	1.0	0.3 J	0.0 U	1.1	0.5	1.6	0.0 U	0.0 U
C3-Dibenzothiophenes	0.0 U	0.0 U	0.0 U	1.1	0.0 U	0.0 U	1.2	0.0 U	1.3	0.0 U	0.0 U
Fluoranthene	0.2 J	0.1 J	1.2	20	0.2 J	0.0 U	15	2.8	7	0.8	1
Pyrene	0.2	0.1 J	1.6	29	0.2	0.0 U	16	3.3	9	0.7	1
C1-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.3 J	3.0	0.8	0.0 U	3.5	0.6	2.4	2.3	0 J
C2-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.0 U	1.7	1.1	0.0 U	2.9	0.5	1.5	0.8	0.2 J
C3-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.0 U	0.8	0.9	0.0 U	1.7	0.0 U	0.7	0.0 U	0.0 U
Naphthobenzothiophene	0.0 U	0.0 U	0.0 U	0.6	0.2 J	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	1.2	0.4 J	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	1.8	0.4 J	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benz(a)anthracene	0.0 U	0.0 U	0.2	2.6	0.2	0.0 U	2.9	0.5	1.2	0.1 J	0
Chrysene	0.0 U	0.0 U	0.3	3.6	0.3	0.0 U	3.6	0.6	2	0.2	0
C1-Chrysenes	0.0 U	0.0 U	0.3 J	1.7	0.8	0.0 U	2.1	0.4	1.0	0.0 U	0 U
C2-Chrysenes	0.0 U	0.0 U	0.4	2.2	0.8	0.0 U	2.1	0.8	1.4	0.0 U	0.0 U
C3-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(b)fluoranthene	0.0 U	0.0 U	0.7	6.4	0.2 J	0.0 U	8.1	1.3	4	0.1 J	1
Benzo(k)fluoranthene	0.0 U	0.0 U	0.2 J	1.9	0.0 U	0.0 U	2.2	0.1 J	0.9	0.0 U	0.2 J
Benzo(e)pyrene	0.0 U	0.0 U	0.4	4.4	0.2 J	0.0 U	4.5	0.7	2.0	0.1 J	0.3 J
Benzo(a)pyrene	0.0 U	0.0 U	0.5	5.3	0.1 J	0.0 U	4.2	0.9	2.3	0.1 J	0.4
Perylene	0.0 U	0.0 U	0.2 J	2.3	0.0 U	0.0 U	1.3 J	0.3 J	0.7 J	0.2 J	0.1 J
Indeno(1,2,3-c,d)pyrene	0.0 U	0.0 U	0.6	5.6	0.0 U	0.0 U	5.6	1.1	2.8	0.1 J	0.5
Dibenzo(a,h)anthracene	0.0 U	0.0 U	0.1 J	0.3	0.0 U	0.0 U	0.9	0.0 U	0.4	0.0 U	0.0 U
C1-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(g,h,i)perylene	0.2	0.0 U	1.1	11.9	0.2	0.0 U	9.5	2.1	5.0	0.2	0.9
Total PAHs	3.5	2.4	14.4	156	22.6	1.1	141	31.4	99	47	12
Individual Isomers*											
2-Methylnaphthalene	0.2 J	0.2 J	0.3	1.2	0.4	0.2 J	1.3	0.6	1.0	0.5	0.3
1-Methylnaphthalene	0.1 J	0.1 J	0.2	0.6	0.2	0.0 U	0.6	0.3	0.5	0.3	0.1 J
2,6-Dimethylnaphthalene	0.1 J	0.1 J	0.3	1.5	0.3	0.1 J	1.7	0.7	1.7	0.4	0.3
1,6,7-Trimethylnaphthalene	0.0 U	0.0 U	0.0 U	0.1	0.2	0.0 U	0.2	0.1	0.6	1.4	0.0 U
1-Methylphenanthrene	0.0 U	0.0 U	0.2 J	0.5	0.6	0.0 U	1.0	0.3	1.2	1.5	0.1 J

Notes: J, below method detection level, MDL; U, not detected

*Individual isomers contained in alkylated (C1-C4 sums)

Appendix E. Polycyclic aromatic hydrocarbons (PAHs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	HTOC9	HTOC10	HTOC12	HTOC14	HTOC15	HTOC16	HTOC17	HTOC19	HTOC20	HTOC21	HTOC22
Decalin	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C1-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C4-Decalins	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Naphthalene	3.0	0.2	0.2	0.2	0.3	16.0	15.8	10.9	2.1	0.7	9.5
C1-Naphthalenes	0.6	0.1 J	0.1 J	0.1 J	0.1 J	1.8	6.1	6.9	0.5	0.2 J	1.7
C2-Naphthalenes	0.9	0.4	0.0 U	0.0 U	0.2 J	3.1	10.9	12.9	1.8	0.3 J	3.0
C3-Naphthalenes	0.6	0.0 U	0.0 U	0.0 U	0.0 U	2.0	9.7	11.4	1.2	0.0 U	2.4
C4-Naphthalenes	1.2	0.0 U	0.0 U	0.0 U	0.0 U	2.8	8.0	9.0	2.2	0.0 U	3.1
Benzo[thiophene]	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.2	0.0 U	0.0 U	0.0 U	0.0 U
C1-Benzo[thiophenes]	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Benzo[thiophenes]	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Benzo[thiophenes]	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Biphenyl	0.6	0.2	0.2	0.2	0.3	2.9	3.5	2.7	0.6	0.3	2.2
Acenaphthylene	0.6	0.0 U	0.0 U	0.0 U	0.1 J	3.7	4.8	10.0	0.4	0.1 J	2.3
Acenaphthene	0.3	0.2	0.1 J	0.1 J	0.1 J	1.3	2.0	1.7	1.1	0.0 U	0.8
Dibenzofuran	0.6	0.1 J	0.1 J	0.2 J	0.2 J	2.6	3.4	3.3	0.6	0.2 J	1.9
Fluorene	1.2	0.0 U	0.3	0.0 U	0.0 U	2.8	3.4	3.7	2.3	0.0 U	1.4
C1-Fluorenes	0.8	0.0 U	0.0 U	0.0 U	0.0 U	1.7	5.5	6.6	0.0 U	0.0 U	2.4
C2-Fluorenes	2.2	0.0 U	0.0 U	0.0 U	0.0 U	3.4	10.8	12.2	0.0 U	0.0 U	3.5
C3-Fluorenes	0.0	0.0	0.0	0.0	0.0	5.8	12.0	18.1	0.0	0.0	5.7
Carbazole	0.2 J	0.0 U	0.0 U	0.0 U	0.0 U	0.5	1.4	3.1	0.2 J	0.0 U	0.3 J
Anthracene	1.2	0.0 U	0.0 U	0.0 U	0.0 U	5.5	9.0	17.9	0.7	0.2	3.8
Phenanthrene	4.7	0.3	0.3	0.3	0.2	19.7	13.6	16.5	2.5	0.7	13.9
C1-Phenanthrene/Anthracenes	1.9	0.2 J	0.2 J	0.0 U	0.2 J	7.0	18.4	31.2	2.0	0.3	5.9
C2-Phenanthrene/Anthracenes	1.7	0.5	0.0 U	0.0 U	0.0 U	4.6	26.5	42.2	1.8	0.0 U	4.5
C3-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	3.7	32.4	41.0	0.0 U	0.0 U	3.5
C4-Phenanthrene/Anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.9	10.9	14.8	0.0 U	0.0 U	1.1
Dibenzothiophene	0.2	0.0 U	0.0 U	0.0 U	0.0 U	0.5	1.9	2.4	0.1 J	0.0 U	0.4
C1-Dibenzothiophenes	0.4	0.0 U	0.0 U	0.0 U	0.0 U	1.0	5.1	6.8	0.4	0.0 U	0.9
C2-Dibenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.6	10.0	13.0	0.3 J	0.0 U	1.8
C3-Dibenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.7	11.3	14.4	0.0 U	0.0 U	1.9
Fluoranthene	6	0.1 J	0.1 J	0.2 J	0.1 J	40.9	29.6	46.6	4.1	1.0	27
Pyrene	7	0.1 J	0.1 J	0.2	0.2	54.6	38.9	45.3	4.8	1.2	35
C1-Fluoranthenes/Pyrenes	1.1	0.0 U	0.0 U	0.0 U	0.0 U	5.6	20.9	33.0	0.9	0.0 U	5.0
C2-Fluoranthenes/Pyrenes	1.2	0.0 U	0.0 U	0.0 U	0.0 U	3.5	17.7	34.6	0.7	0.0 U	3.2
C3-Fluoranthenes/Pyrenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.4	8.9	15.6	0.0 U	0.0 U	1.7
Naphthobenzothiophene	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.9	6.0	9.5	0.0 U	0.0 U	0.9
C1-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.6	8.4	13.9	0.0 U	0.0 U	1.9
C2-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	2.7	13.5	23.1	0.0 U	0.0 U	2.7
C3-Naphthobenzothiophenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.6	13.0	22.7	0.0 U	0.0 U	2.1
Benz(a)anthracene	0.9	0.0 U	0.0 U	0.0 U	0.1 J	8.4	15.9	30.1	0.7	0.1 J	3.8
Chrysene	1	0.0 U	0.0 U	0.0 U	0.1 J	4.4	17.5	28.0	1.0	0.1 J	5.2
C1-Chrysenes	0.6	0.0 U	0.0 U	0.0 U	0.0 U	2.8	18.9	32.5	0.0 U	0.0 U	2.9
C2-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	2.7	15.8	27.2	0.0 U	0.0 U	2.6
C3-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	10.1	17.0	0.0 U	0.0 U	0.0 U
C4-Chrysenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	3.9	7.2	0.0 U	0.0 U	0.0 U
Benzo(b)fluoranthene	3	0.0 U	0.0 U	0.0 U	0.1 J	9.8	25.8	40.7	1.7	0.4	9.2
Benzo(k)fluoranthene	0.7	0.0 U	0.0 U	0.0 U	0.0 U	1.9	6.8	15.3	0.5	0.1 J	1.8
Benzo(e)pyrene	1.5	0.0 U	0.0 U	0.0 U	0.1 J	6.5	14.4	23.0	0.8	0.2 J	5.7
Benzo(a)pyrene	1.9	0.0 U	0.0 U	0.0 U	0.1 J	8.0	12.1	21.4	1.2	0.2 J	6.8
Perylene	0 J	0.0 U	0.0 U	0.0 U	0.1 J	3.0	22.2	68.0	0.3 J	0.1 J	1.8
Indeno(1,2,3-c,d)pyrene	2.5	0.0 U	0.0 U	0.0 U	0.1 J	8.6	8.7	17.1	1.8	0.3	8.0
Dibenzo(a,h)anthracene	0.5	0.0 U	0.0 U	0.0 U	0.0 U	0.9	3.3	8.3	0.4	0.0 U	0.9
C1-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C2-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
C3-Dibenzo(a,h)anthracenes	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Benzo(g,h,i)perylene	4.9	0.0 U	0.0 U	0.1 J	0.1 J	17.2	14.2	18.4	2.9	0.5	14.0
Total PAHs	56	2.4	1.7	1.6	2.8	284	583	911	42.6	7.2	220
Individual Isomers*											
2-Methylnaphthalene	0.7	0.1 J	0.1 J	0.1 J	0.1 J	1.8	6.7	7.6	0.6	0.2 J	1.8
1-Methylnaphthalene	0.3	0.1 J	0.1 J	0.1 J	0.1 J	1.0	2.9	3.2	0.3	0.1 J	0.9
2,6-Dimethylnaphthalene	0.8	0.2 J	0.0 U	0.0 U	0.1 J	3.1	8.4	10.3	1.9	0.2 J	2.8
1,6,7-Trimethylnaphthalene	0.1	0.0 U	0.0 U	0.0 U	0.0 U	0.3	1.0	1.2	0.1	0.0 U	0.2
1-Methylphenanthrene	0.4	0.0 U	0.1 J	0.0 U	0.0 U	0.8	3.0	4.6	0.3	0.1 J	0.9

Notes: J, below method detection level, MDL; U, not detected
 *Individual isomers contained in alkylated (C1-C4 sums)

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g).

Compound	Coral 1	Coral 2	Coral 3	Coral 4	Coral 5	Coral 6
PCB8/5	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB18	0.00 U	0.00 U	0.00 U	0.31	0.00 U	0.00 U
PCB28	0.14	0.08 J	0.25	0.48	0.60	0.11
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.00 U	0.00 U	0.25	0.55	0.59	0.10
PCB44	0.00 U	0.00 U	0.00 U	0.02 J	0.04 J	0.00 U
PCB45	0.00 U	0.00 U	1.35	0.00 U	0.00 U	0.00 U
PCB49	1.27	0.00 U	0.94	1.67	2.62	0.83
PCB52	0.52	0.00 U	0.56	1.21	1.51	0.44
PCB56/60	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB66	0.07	0.34	0.30	0.27	0.39	0.05 J
PCB70	0.07 J	0.00 U	0.30	0.32	0.00 U	0.00 U
PCB74/61	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.01 J	0.00 U	0.00 U	0.04 J	0.00 U	0.02 J
PCB95	0.21	0.00 U	0.00 U	0.44	0.65	0.14
PCB99	0.26	0.00 U	0.30	0.60	1.00	0.26
PCB101/90	0.51	0.12	0.45	0.77	1.53	0.52
PCB105	0.02 J	0.01 J	0.04 J	0.05	0.05	0.05 J
PCB110/77	0.12	0.00 U	0.11	0.22	0.30	0.17
PCB118	0.17	0.02 J	0.21	0.29	0.59	0.20
PCB128	0.00 U	0.00 U	0.00 U	0.00 U	0.08 J	0.00 U
PCB138/160	0.48	0.26	0.61	1.02	1.56	0.57
PCB146	0.10 J	0.02 J	0.20	0.27	0.54	0.17
PCB149/123	0.39	0.08 J	0.49	1.04	1.03	0.44
PCB151	0.15	0.00 U	0.18	0.32	0.30	0.00 U
PCB153/132	1.30	0.31	1.49	2.55	4.46	1.57
PCB156/171/202	0.10 J	0.01 J	0.04 J	0.04 J	0.04 J	0.00 U
PCB158	0.08 J	0.00 U	0.10 J	0.14	0.20	0.05 J
PCB170/190	0.30	0.08	0.49	0.20	0.73	0.20
PCB174	0.19	0.04 J	0.10	0.29	0.34	0.02 J
PCB180	0.48	0.14	0.53	1.11	1.45	0.46
PCB183	0.10	0.02 J	0.12	0.24	0.41	0.11
PCB187	0.38	0.06 J	0.24	0.60	0.97	0.16
PCB194	0.13	0.02 J	0.09	0.17	0.17	0.00 U
PCB195/208	0.05	0.00 U	0.00 U	0.11	0.00 U	0.00 U
PCB201/157/173	0.00 U	0.00 U	0.00 U	0.02 J	0.03 J	0.00 U
PCB206	0.01 J	0.00 U	0.00 U	0.02 J	0.01 J	0.00 U
PCB209	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total PCB	11.83	5.33	13.51	21.73	32.68	11.69

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	Coral 8	Coral 9	LTOC 1	LTOC 2	LTOC 3	LTOC 7
PCB8/5	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	1.16
PCB18	0.00 U	0.03 J	0.00 U	0.00 U	0.00 U	0.33
PCB28	0.21	0.00 U	0.10	0.00 U	0.00 U	1.24
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.19	0.00 U	0.06	0.00 U	0.00 U	1.57
PCB44	0.00 U	0.03 J	0.00 U	0.00 U	0.00 U	0.11
PCB45	0.02 J	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB49	0.57	0.19	0.21	0.14	0.05 J	6.17
PCB52	0.39	0.16	0.18	0.11	0.03 J	3.77
PCB56/60	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB66	0.03 J	0.00 U	0.00 U	0.00 U	0.00 U	0.58
PCB70	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.34
PCB74/61	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.11
PCB95	0.17	0.00 U	0.00 U	0.00 U	0.00 U	0.88
PCB99	0.20	0.05	0.09	0.07	0.00 U	1.91
PCB101/90	0.42	0.14	0.11	0.11	0.10	2.77
PCB105	0.03 J	0.02 J	0.00 U	0.00 U	0.00 U	0.08
PCB110/77	0.10	0.00 U	0.02 J	0.02 J	0.00 U	0.68
PCB118	0.12	0.03 J	0.03 J	0.01 J	0.00 U	0.88
PCB128	0.00 U	0.06 J	0.00 U	0.00 U	0.00 U	0.13
PCB138/160	0.44	0.22	0.15	0.13	0.11 J	3.04
PCB146	0.11 J	0.04 J	0.02 J	0.00 U	0.00 U	0.78
PCB149/123	0.40	0.12 J	0.11 J	0.07 J	0.02 J	1.63
PCB151	0.00 U	0.00 U	0.04 J	0.00 U	0.00 U	1.02
PCB153/132	1.10	0.29	0.28	0.23	0.09	7.73
PCB156/171/202	0.04 J	0.00 U	0.01 J	0.00 U	0.00 U	0.29
PCB158	0.06 J	0.00 U	0.00 U	0.00 U	0.00 U	0.28
PCB170/190	0.13	0.06 J	0.07 J	0.03 J	0.02 J	1.02
PCB174	0.12	0.03 J	0.03 J	0.02 J	0.01 J	0.72
PCB180	0.48	0.12	0.14	0.12	0.05	3.13
PCB183	0.10	0.00 U	0.01 J	0.00 U	0.00 U	0.53
PCB187	0.28	0.00 U	0.05 J	0.07	0.02 J	1.69
PCB194	0.07	0.00 U	0.00 U	0.00 U	0.00 U	0.56
PCB195/208	0.00 U	0.00 U	0.02 J	0.00 U	0.00 U	0.26
PCB201/157/173	0.02 J	0.00 U	0.00 U	0.00 U	0.00 U	0.04 J
PCB206	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.10
PCB209	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.12
Total PCB	10.15	4.68	4.61	4.00	3.11	63.27

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 8	LTOC 9	LTOC 10	LTOC 11	LTOC 12	LTOC 13
PCB8/5	0.13	0.40	0.00 U	0.00 U	0.00 U	0.00 U
PCB18	0.13	0.21	0.10	0.00 U	0.00 U	0.00 U
PCB28	0.43	0.80	0.39	0.69	0.19	0.09 J
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.65	1.07	0.79	1.11	0.10	0.07
PCB44	0.04 J	0.08 J	0.03 J	0.05 J	0.00 U	0.00 U
PCB45	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.02 J
PCB49	2.00	4.12	1.69	2.97	0.27	0.22
PCB52	1.35	3.06	1.19	2.10	0.22	0.17
PCB56/60	0.00 U	0.00 U	0.04 J	0.00 U	0.00 U	0.00 U
PCB66	0.14	0.34	0.10	0.22	0.00 U	0.00 U
PCB70	0.07 J	0.20	0.00 U	0.05 J	0.00 U	0.09
PCB74/61	0.07	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.04 J	0.14	0.00 U	0.00 U	0.00 U	0.00 U
PCB95	0.25	0.69	0.30	0.53	0.00 U	0.00 U
PCB99	0.59	1.40	0.39	0.92	0.08	0.06
PCB101/90	1.10	2.13	0.68	1.57	0.22	0.14
PCB105	0.07	0.10	0.04 J	0.04 J	0.00 U	0.00 U
PCB110/77	0.23	0.56	0.18	0.32	0.00 U	0.02 J
PCB118	0.32	0.67	0.24	0.43	0.02 J	0.04 J
PCB128	0.00 U	0.10 J	0.00 U	0.00 U	0.00 U	0.00 U
PCB138/160	0.96	2.43	0.97	1.72	0.10 J	0.18
PCB146	0.20	0.59	0.22	0.37	0.03 J	0.03 J
PCB149/123	0.64	1.35	0.70	1.21	0.08 J	0.15
PCB151	0.20	0.71	0.29	0.55	0.02 J	0.04 J
PCB153/132	2.69	6.32	2.28	3.99	0.39	0.37
PCB156/171/202	0.11 J	0.09 J	0.10 J	0.21	0.00 U	0.02 J
PCB158	0.12	0.25	0.09 J	0.18	0.01 J	0.00 U
PCB170/190	0.48	1.09	0.56	0.58	0.00 U	0.09
PCB174	0.32	0.67	0.26	0.50	0.04 J	0.05
PCB180	1.15	2.51	0.86	1.83	0.19	0.19
PCB183	0.20	0.38	0.20	0.40	0.04 J	0.03 J
PCB187	0.64	1.50	0.56	1.01	0.10	0.10
PCB194	0.24	0.42	0.22	0.39	0.06	0.04
PCB195/208	0.10	0.21	0.07	0.15	0.07	0.03 J
PCB201/157/173	0.00 U	0.03 J	0.00 U	0.05 J	0.01 J	0.00 U
PCB206	0.00 U	0.08	0.00 U	0.00 U	0.00 U	0.00 U
PCB209	0.00 U	0.06 J	0.00 U	0.00 U	0.00 U	0.00 U
Total PCB	23.24	50.23	19.67	33.23	5.32	5.17

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 14	LTOC 15	LTOC 16	LTOC 17	LTOC 18	LTOC 19
PCB8/5	0.00 U	0.24	0.00 U	0.00 U	0.00 U	0.00 U
PCB18	0.00 U	0.25	0.18	0.00 U	0.00 U	0.00 U
PCB28	0.00 U	0.98	0.55	0.00 U	0.00 U	0.33
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.00 U	1.59	0.84	0.00 U	0.00 U	0.29
PCB44	0.00 U	0.11	0.03 J	0.02 J	0.00 U	0.00 U
PCB45	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB49	0.05 J	5.23	2.85	0.17	0.06 J	1.27
PCB52	0.04 J	3.21	1.86	0.09	0.04 J	0.85
PCB56/60	0.00 U	0.41	0.15	0.00 U	0.00 U	0.07 J
PCB66	0.04 J	0.38	0.15	0.00 U	0.02 J	0.00 U
PCB70	0.00 U	0.26	0.07 J	0.00 U	0.00 U	0.00 U
PCB74/61	0.00 U	0.30	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.00 U	0.13	0.07 J	0.00 U	0.00 U	0.03 J
PCB95	0.00 U	0.96	0.36	0.00 U	0.00 U	0.00 U
PCB99	0.00 U	1.78	0.93	0.04 J	0.02 J	0.47
PCB101/90	0.15	2.73	1.59	0.14	0.17	0.65
PCB105	0.00 U	0.11	0.04 J	0.00 U	0.00 U	0.03 J
PCB110/77	0.02 J	0.63	0.32	0.07	0.02 J	0.14
PCB118	0.00 U	0.91	0.41	0.02 J	0.01 J	0.21
PCB128	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB138/160	0.00 U	3.18	1.69	0.16	0.21	0.88
PCB146	0.00 U	0.84	0.36	0.02 J	0.00 U	0.19
PCB149/123	0.02 J	2.23	1.20	0.08 J	0.03 J	0.65
PCB151	0.02 J	1.00	0.50	0.03 J	0.01 J	0.23
PCB153/132	0.11	7.51	3.89	0.27	0.10	1.96
PCB156/171/202	0.00 U	0.27	0.10 J	0.01 J	0.00 U	0.03 J
PCB158	0.00 U	0.29	0.17	0.01 J	0.00 U	0.09 J
PCB170/190	0.00 U	1.19 I	0.77	0.04 J	0.02 J	0.31
PCB174	0.00 U	0.68	0.49	0.00 U	0.00 U	0.20
PCB180	0.05	2.91	1.70	0.12	0.05	0.90
PCB183	0.01 J	0.64	0.32	0.00 U	0.00 U	0.18
PCB187	0.02 J	1.64	0.94	0.04 J	0.02 J	0.48
PCB194	0.00 U	0.55	0.30	0.02 J	0.00 U	0.17
PCB195/208	0.00 U	0.28	0.15	0.00 U	0.00 U	0.08
PCB201/157/173	0.00 U	0.06 J	0.00 U	0.00 U	0.00 U	0.00 U
PCB206	0.00 U	0.09	0.00 U	0.00 U	0.00 U	0.00 U
PCB209	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total PCB	3.09	57.89	32.51	4.13	3.61	16.69

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 22	HTOC 1	HTOC 2	HTOC 3	HTOC 4	HTOC 6
PCB8/5	0.34	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB18	0.16	0.00 U	0.00 U	0.00 U	0.22	0.00 U
PCB28	0.82	0.00 U	0.01 J	0.26	0.31	0.93
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	1.30	0.00 U	0.00 U	0.15	0.50	1.39
PCB44	0.10	0.00 U	0.00 U	0.00 U	0.04 J	0.10 J
PCB45	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB49	4.61	0.00 U	0.05 J	0.62	1.26	3.84
PCB52	2.78	0.00 U	0.05 J	0.34	0.86	2.69
PCB56/60	0.41	0.00 U	0.00 U	0.00 U	0.00 U	0.71
PCB66	0.38	0.00 U	0.00 U	0.11	0.00 U	0.39
PCB70	0.29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB74/61	0.34	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.11	0.00 U	0.00 U	0.00 U	0.00 U	0.14
PCB95	0.89	0.00 U	0.00 U	0.21	0.00 U	0.72
PCB99	1.66	0.00 U	0.00 U	0.31	0.47	1.26
PCB101/90	2.08	0.00 U	0.00 U	0.33	0.75	2.05
PCB105	0.10	0.00 U	0.00 U	0.08	0.04 J	0.12
PCB110/77	0.62	0.00 U	0.19	0.00 U	0.19	0.59
PCB118	0.88	0.00 U	0.01 J	0.20	0.28	0.73
PCB128	0.09 J	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB138/160	3.05	0.07 J	0.14	0.73	0.73	2.08
PCB146	0.80	0.00 U	0.00 U	0.14	0.20	0.73
PCB149/123	2.02	0.00 U	0.03 J	0.38	0.62	1.63
PCB151	0.91	0.00 U	0.00 U	0.00 U	0.29	0.86
PCB153/132	7.11	0.00 U	0.08	1.21	1.96	5.84
PCB156/171/202	0.28	0.00 U	0.00 U	0.00 U	0.00 U	0.11 J
PCB158	0.25	0.00 U	0.00 U	0.00 U	0.09 J	0.31
PCB170/190	1.40	0.00 U	0.04 J	0.27	0.13	0.77
PCB174	0.64	0.00 U	0.00 U	0.00 U	0.22	0.73
PCB180	2.68	0.03 J	0.04 J	0.49	0.73	2.87
PCB183	0.64	0.00 U	0.00 U	0.00 U	0.16	0.58
PCB187	1.51	0.00 U	0.00 U	0.00 U	0.33	1.55
PCB194	0.56	0.00 U	0.00 U	0.00 U	0.14	0.44
PCB195/208	0.26	0.00 U	0.00 U	0.00 U	0.07	0.27
PCB201/157/173	0.04 J	0.00 U	0.00 U	0.00 U	0.00 U	0.09
PCB206	0.08	0.00 U	0.00 U	0.00 U	0.00 U	0.08
PCB209	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total PCB	53.84	2.41	2.99	11.01	16.11	46.53

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	HTOC 7	HTOC 8	HTOC 9	HTOC 10	HTOC 12	HTOC 14
PCB8/5	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB18	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB28	0.02 J	0.03 J	0.12	0.00 U	0.00 U	0.00 U
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.00 U	0.18	0.00 U	0.00 U	0.00 U	0.00 U
PCB44	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB45	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB49	0.23	0.52	0.21	0.12	0.07 J	0.07 J
PCB52	0.13	0.35	0.13	0.08	0.04 J	0.06 J
PCB56/60	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB66	0.07	0.02 J	0.06	0.00 U	0.00 U	0.00 U
PCB70	0.13	0.00 U	0.14	0.00 U	0.00 U	0.00 U
PCB74/61	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB87/115	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB95	0.09	0.16	0.08	0.00 U	0.00 U	0.00 U
PCB99	0.02 J	0.13	0.07	0.00 U	0.00 U	0.00 U
PCB101/90	0.19	0.28	0.12	0.00 U	0.00 U	0.00 U
PCB105	0.06	0.03 J	0.03 J	0.00 U	0.00 U	0.00 U
PCB110/77	0.03 J	0.08	0.00 U	0.03 J	0.00 U	0.00 U
PCB118	0.02 J	0.10	0.04 J	0.03 J	0.00 U	0.02 J
PCB128	0.07 J	0.00 U	0.85	0.00 U	0.00 U	0.00 U
PCB138/160	0.28	0.34	0.79	0.15	0.10 J	0.16
PCB146	0.00 U	0.09 J	0.00 U	0.00 U	0.00 U	0.00 U
PCB149/123	0.11 J	0.24	0.37	0.00 U	0.00 U	0.04 J
PCB151	0.00 U	0.11 J	0.00 U	0.06 J	0.00 U	0.00 U
PCB153/132	0.19	0.72	0.46	0.28	0.12	0.12
PCB156/171/202	0.00 U	0.00 U	0.20	0.00 U	0.00 U	0.00 U
PCB158	0.01 J	0.03 J	0.22	0.00 U	0.00 U	0.00 U
PCB170/190	0.47	0.07 J	1.25	0.09	0.00 U	0.00 U
PCB174	0.00 U	0.05	0.00 U	0.03 J	0.00 U	0.00 U
PCB180	0.19	0.20	0.58	0.12	0.05	0.00 U
PCB183	0.06	0.06	0.28	0.00 U	0.00 U	0.00 U
PCB187	0.07	0.17	0.23	0.11	0.04 J	0.00 U
PCB194	0.01 J	0.00 U	0.51	0.00 U	0.00 U	0.00 U
PCB195/208	0.00 U	0.00 U	1.05	0.00 U	0.00 U	0.00 U
PCB201/157/173	0.13	0.00 U	0.30	0.00 U	0.00 U	0.00 U
PCB206	0.00 U	0.00 U	0.16	0.00 U	0.00 U	0.00 U
PCB209	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total PCB	6.05	7.29	12.69	4.06	2.95	2.98

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix F. Polychlorinated biphenyls (PCBs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	HTOC 15	HTOC 16	HTOC 17	HTOC 19	HTOC 20	HTOC 21	HTOC 22
PCB8/5	0.00 U	0.00 U	21.86 D	23.90 D	0.00 U	0.00 U	0.00 U
PCB18	0.00 U	0.21	22.93 D	9.20	0.16	0.00 U	0.25
PCB28	0.00 U	0.84	55.86 D	24.05 D	0.00 U	0.00 U	0.82
PCB29	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
PCB31	0.00 U	1.16	115.04 D	39.18 D	2.91	0.00 U	1.20
PCB44	0.00 U	0.10 J	3.27	1.45	0.00 U	0.00 U	0.13
PCB45	0.00 U	0.00 U	0.34	0.00 U	0.00 U	0.00 U	0.00 U
PCB49	0.30	4.47	198.52 D	88.10 D	0.66	0.14	4.65
PCB52	0.24	2.49	202.91 D	89.43 D	0.45	0.09	3.48
PCB56/60	0.00 U	0.49	6.29	2.75	0.09	0.00 U	0.40
PCB66	0.00 U	0.29	21.39 D	9.10	0.00 U	0.00 U	0.35
PCB70	0.00 U	0.18	10.81	5.00	0.16	0.02 J	0.21
PCB74/61	0.00 U	0.23	5.98	2.94	0.00 U	0.00 U	0.18
PCB87/115	0.00 U	0.12	5.21	2.39	0.00 U	0.00 U	0.00 U
PCB95	0.00 U	1.13	30.07 D	13.89 D	0.00 U	0.00 U	0.75
PCB99	0.13	1.61	86.48 D	36.50 D	0.31	0.10	1.70
PCB101/90	0.00 U	2.47	100.50 D	43.47 D	0.37	0.26	2.60
PCB105	0.00 U	0.18	4.00	1.53	0.00 U	0.00 U	0.13
PCB110/77	0.04 J	0.60	26.77 D	7.14 D	0.10	0.03 J	0.69
PCB118	0.05 J	0.86	31.29 D	13.45 D	0.13	0.02 J	0.91
PCB128	0.00 U	0.06 J	5.58	2.48	0.00 U	0.00 U	0.10 J
PCB138/160	0.16	3.40	98.69 D	48.79 D	0.56	0.16	3.61
PCB146	0.03 J	0.77	26.42 D	11.92	0.00 U	0.03 J	0.78
PCB149/123	0.11 J	2.01	64.34 D	29.84 D	0.50	0.09 J	2.14
PCB151	0.00 U	0.86	39.38 D	15.89 D	0.11 J	0.03 J	0.92
PCB153/132	0.44	7.12	377.58 D	117.16 D	1.07	0.25	7.50
PCB156/171/202	0.02 J	0.24	15.08 D	7.25	0.00 U	0.00 U	0.23
PCB158	0.00 U	0.26	10.28	4.28	0.03 J	0.00 U	0.25
PCB170/190	0.08	1.02	61.72 D	17.79 D	0.35	0.00 U	1.44
PCB174	0.00 U	0.59	0.00 U	10.77 D	0.08	0.02 J	0.64
PCB180	0.20	2.78	138.80 D	39.17 D	0.43	0.10	3.16
PCB183	0.00 U	0.53	27.61 D	11.94	0.09	0.00 U	0.58
PCB187	0.10	1.51	82.92 D	20.71 D	0.25	0.05 J	1.67
PCB194	0.00 U	0.54	30.01 D	11.75 D	0.08	0.00 U	0.60
PCB195/208	0.00 U	0.42	8.65	4.07	0.00 U	0.00 U	0.32
PCB201/157/173	0.00 U	0.05 J	2.96	1.50	0.00 U	0.00 U	0.06 J
PCB206	0.00 U	0.06	5.10	2.67	0.00 U	0.00 U	0.10
PCB209	0.00 U	0.20	0.07 J	0.29	0.00 U	0.00 U	0.14
Total PCB	4.98	53.89	2709.90	1021.78	10.41	4.23	59.67

Qualifiers (Q): J=Below the MDL; U=Not detected; D=Sample diluted

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g).

Compound	Coral 1	Coral 2	Coral 3	Coral 4	Coral 5
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.07	0.00 U	0.00 U	0.00 U	0.00 U
Endrin	0.00 U	0.00 U	0.00 U	0.06 J	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordan	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-Chlordane	0.00 U	0.00 U	0.02 J	0.00 U	0.00 U
Trans-Nonachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.01 J	0.00 U	0.01 J	0.02 J	0.05
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.13	0.00 U	0.13	0.22	0.28
4,4'-DDD	0.00 U	0.00 U	0.00 U	0.00 U	0.01 J
2,4'-DDE	0.02 J	0.00 U	0.00 U	0.05 J	0.08
4,4'-DDE	0.06	0.00 U	0.04 J	0.00 U	0.02 J
2,4'-DDT	0.01 J	0.00 U	0.00 U	0.00 U	0.02 J
4,4'-DDT	0.01 J	0.01 J	0.08	0.10	0.03 J
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Pentachloroanisole	0.00 U	0.00 U	0.08	0.00 U	0.09
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.12 J	0.00 U	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan Sulfate	0.00 U	0.05 J	0.00 U	0.31	0.00 U
Mirex	0.00 U	0.00 U	0.12	0.03 J	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U	0.36	0.00 U
Total HCH	0.00	0.00	0.00	0.00	0.00
Total Chlordane	0.01	0.00	0.03	0.02	0.05
Total DDT	0.23	0.01	0.25	0.37	0.44

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	Coral 6	Coral 8	Coral 9	LTOC 1	LTOC 2
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.03 J	0.00 U
Endrin	0.16 J	0.02 J	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxyde	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlorane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.09	0.14	0.01 J	0.00 U	0.00 U
Gamma-Chlordane	0.03 J	0.03 J	0.09	0.01 J	0.01 J
Trans-Nonachlor	0.00 U	0.00 U	0.02 J	0.00 U	0.00 U
Cis-Nonachlor	0.01 J	0.01 J	0.00 U	0.00 U	0.00 U
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.00 U	0.00 U	0.19	0.07	0.06 J
Delta-HCH	0.00 U	0.00 U	0.00 U	0.01 J	0.02 J
Gamma-HCH	0.00 U	0.69	0.00 U	0.01 J	0.01 J
2,4'-DDD	0.15	0.09	0.00 U	0.00 U	0.00 U
4,4'-DDD	0.01 J	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDE	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDE	0.02 J	0.03 J	0.00 U	0.00 U	0.00 U
2,4'-DDT	0.01 J	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDT	0.10	0.07	0.03 J	0.00 U	0.00 U
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.01 J	0.01 J
Pentachloroanisole	0.25	0.14	0.04 J	0.01 J	0.02 J
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.07 J	0.00 U	0.00 U	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	1.44	0.00 U
Endosulfan Sulfate	0.00 U	0.10	0.00 U	0.00 U	0.00 U
Mirex	0.00 U	0.03 J	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.15	0.00 U	0.06 J	0.00 U	0.00 U
Total HCH	0.00	0.69	0.19	0.10	0.09
Total Chlordane	0.14	0.19	0.13	0.01	0.01
Total DDT	0.30	0.19	0.03	0.00	0.00

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	LTOC 3	LTOC 7	LTOC 8	LTOC 9	LTOC 10
Aldrin	0.00 U	0.00 U	0.06	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.05 J	0.00 U
Endrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordan	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-Chlordane	0.01 J	0.00 U	0.00 U	0.00 U	0.00 U
Trans-Nonachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.00 U	0.08	0.03 J	0.06	0.02 J
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.00 U	0.06 J	0.00 U	0.12	0.00 U
Delta-HCH	0.00 U	0.00 U	0.02 J	0.02 J	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.00 U	0.56	0.18	0.41	0.16
4,4'-DDD	0.00 U	0.04 J	0.00 U	0.00 U	0.00 U
2,4'-DDE	0.00 U	0.15	0.07	0.26	0.13
4,4'-DDE	0.00 U	0.41	0.15	0.00 U	0.06
2,4'-DDT	0.00 U	0.18	0.00 U	0.24	0.04
4,4'-DDT	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Pentachloroanisole	0.00 U	0.04 J	0.04 J	0.08	0.01 J
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.00 U	0.53	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	0.23	0.00 U
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U	0.76	0.00 U
Mirex	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total HCH	0.00	0.06	0.02	0.14	0.00
Total Chlordane	0.01	0.08	0.03	0.06	0.02
Total DDT	0.00	1.33	0.40	0.91	0.39

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	LTOC 11	LTOC 12	LTOC 13	LTOC 14	LTOC 15
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordan	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Trans-Nonachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.03 J	0.00 U	0.00 U	0.00 U	0.07
Alpha-HCH	0.00 U	0.00 U	0.05 J	0.00 U	0.00 U
Beta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.00 U	0.00 U	0.00 U	0.02 J	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.31	0.04 J	0.03 J	0.01 J	0.51
4,4'-DDD	0.01 J	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDE	0.23	0.00 U	0.02 J	0.00 U	0.30
4,4'-DDE	0.20	0.00 U	0.02 J	0.00 U	0.37
2,4'-DDT	0.08	0.00 U	0.00 U	0.00 U	0.22
4,4'-DDT	0.00 U	0.00 U	0.00 U	0.00 U	0.05 J
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Pentachloroanisole	0.02 J	0.01 J	0.00 U	0.01 J	0.11
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U	0.00 U	1.32
Mirex	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total HCH	0.00	0.00	0.05	0.02	0.00
Total Chlordane	0.03	0.00	0.00	0.00	0.07
Total DDT	0.83	0.04	0.07	0.01	1.45

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	LTOC 16	LTOC 17	LTOC 18	LTOC 19	LTOC 22
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-Chlordane	0.04 J	0.00 U	0.00 U	0.00 U	0.00 U
Trans-Nonachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.03 J	0.00 U	0.00 U	0.00 U	0.07
Alpha-HCH	0.04 J	0.00 U	0.00 U	0.04 J	0.00 U
Beta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.00 U	0.00 U	0.01 J	0.00 U	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.29	0.04 J	0.02 J	0.17	0.51
4,4'-DDD	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDE	0.00 U	0.00 U	0.00 U	0.00 U	0.21
4,4'-DDE	0.13	0.00 U	0.00 U	0.08	0.36
2,4'-DDT	0.07	0.00 U	0.00 U	0.00 U	0.21
4,4'-DDT	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.08	0.00 U
Pentachloroanisole	0.05 J	0.00 U	0.01 J	0.00 U	0.05 J
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Mirex	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Total HCH	0.04	0.00	0.01	0.04	0.00
Total Chlordane	0.07	0.00	0.00	0.00	0.07
Total DDT	0.50	0.04	0.02	0.25	1.28

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	HTOC 1	HTOC 2	HTOC 3	HTOC 4	HTOC 6
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endrin	0.00 U	0.00 U	0.00 U	0.00 U	0.27
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordan	0.00 U	0.00 U	0.00 U	0.00 U	0.05 J
Alpha-Chlordane	0.00 U	0.00 U	0.26	0.00 U	0.16
Gamma-Chlordane	0.00 U	0.00 U	0.09	0.00 U	0.00 U
Trans-Nonachlor	0.00 U	0.00 U	0.04 J	0.00 U	0.00 U
Cis-Nonachlor	0.00 U	0.00 U	0.00 U	0.01 J	0.05
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.00 U	0.01 J	0.07	0.18	0.68
4,4'-DDD	0.00 U	0.00 U	0.08	0.00 U	0.00 U
2,4'-DDE	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDE	0.00 U	0.00 U	0.05	0.00 U	0.00 U
2,4'-DDT	0.00 U	0.32	0.00 U	0.00 U	0.11
4,4'-DDT	0.01 J	0.01 J	0.21	0.06	0.22
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Pentachloroanisole	0.00 U	0.00 U	0.22	0.20	0.38
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.05 J	0.00 U	0.00 U	0.59
Endosulfan I	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U	0.00 U	0.47
Mirex	0.00 U	0.00 U	0.00 U	0.00 U	0.04 J
Chlorpyrifos	0.00 U	0.00 U	0.45	0.00 U	0.54
Total HCH	0.00	0.00	0.00	0.00	0.00
Total Chlordane	0.00	0.00	0.39	0.01	0.27
Total DDT	0.01	0.34	0.41	0.24	1.01

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	HTOC 7	HTOC 8	HTOC 9	HTOC 10	HTOC 12
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.05 J
Endrin	0.00 U	0.06 J	0.13 J	0.00 U	0.00 U
Heptachlor	0.00 U	0.02 J	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlorane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.09	0.07	0.08	0.00 U	0.00 U
Gamma-Chlordane	0.07	0.09	0.00 U	0.01 J	0.04 J
Trans-Nonachlor	0.03 J	0.03 J	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.00 U	0.01 J	0.02 J	0.00 U	0.00 U
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.00 U	0.00 U	0.00 U	0.03 J	0.02 J
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.18	0.08	0.03 J	0.00 U	0.00 U
4,4'-DDD	0.03 J	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDE	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDE	0.00 U	0.03 J	0.00 U	0.00 U	0.00 U
2,4'-DDT	0.03 J	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDT	0.08	0.02 J	0.70	0.00 U	0.00 U
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Pentachloroanisole	0.09	0.06 J	0.15	0.01 J	0.01 J
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan I	0.00 U	0.00 U	0.00 U	0.06 J	0.04 J
Endosulfan Sulfate	0.00 U	0.00 U	0.47	0.00 U	0.00 U
Mirex	0.06 J	0.00 U	0.92	0.00 U	0.00 U
Chlorpyrifos	0.11 J	0.00 U	0.29	0.00 U	0.00 U
Total HCH	0.00	0.00	0.00	0.03	0.02
Total Chlordane	0.19	0.23	0.10	0.01	0.04
Total DDT	0.33	0.14	0.73	0.00	0.00

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	HTOC 14	HTOC 15	HTOC 16	HTOC 17	HTOC 19
Aldrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Dieldrin	0.00 U	0.00 U	0.00 U	0.41	0.42
Endrin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Oxychlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-Chlordane	0.01 J	0.01 J	0.03 J	0.03 J	0.02 J
Trans-Nonachlor	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Cis-Nonachlor	0.00 U	0.00 U	0.08	2.51	1.05
Alpha-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Beta-HCH	0.04 J	0.00 U	0.00 U	0.00 U	0.00 U
Delta-HCH	0.04 J	0.00 U	0.00 U	0.00 U	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.00 U	0.00 U	0.00 U	28.85 D	13.46 D
4,4'-DDD	0.00 U	0.00 U	0.02 J	0.97	1.08
2,4'-DDE	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
4,4'-DDE	0.00 U	0.00 U	0.39	10.26	9.48
2,4'-DDT	0.00 U	0.00 U	0.00 U	5.29	3.16
4,4'-DDT	0.00 U	0.00 U	0.00 U	1.56	2.17
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.01 J	0.00 U	0.00 U
Pentachloroanisole	0.00 U	0.00 U	0.15	0.00 U	0.00 U
Pentachlorobenzene	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Endosulfan I	0.04 J	0.17	0.00 U	0.00 U	0.00 U
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Mirex	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U	0.00 U	0.60
Total HCH	0.08	0.00	0.00	0.00	0.00
Total Chlordane	0.01	0.01	0.11	2.54	1.08
Total DDT	0.00	0.00	0.41	46.93	29.33

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix G. Organochlorine pesticides in sediment samples from southwest Puerto Rico (ng/dry g)
(continued).

Compound	HTOC 20	HTOC 21	HTOC 22
Aldrin	0.00 U	0.00 U	0.00 U
Dieldrin	0.07	0.00 U	0.14
Endrin	0.00 U	0.00 U	0.00 U
Heptachlor	0.00 U	0.00 U	0.00 U
Heptachlor-Epoxide	0.00 U	0.00 U	0.00 U
Oxychlordane	0.00 U	0.00 U	0.00 U
Alpha-Chlordane	0.00 U	0.12	0.00 U
Gamma-Chlordane	0.00 U	0.06	0.04 J
Trans-Nonachlor	0.00 U	0.02 J	0.00 U
Cis-Nonachlor	0.02 J	0.00 U	0.07
Alpha-HCH	0.00 U	0.00 U	0.00 U
Beta-HCH	0.10	0.06 J	0.11
Delta-HCH	0.00 U	0.02 J	0.00 U
Gamma-HCH	0.00 U	0.00 U	0.00 U
2,4'-DDD	0.07	0.02 J	0.50
4,4'-DDD	0.00 U	0.00 U	0.04 J
2,4'-DDE	0.00 U	0.00 U	0.00 U
4,4'-DDE	0.00 U	0.00 U	0.39
2,4'-DDT	0.05	0.00 U	0.00 U
4,4'-DDT	0.00 U	0.00 U	0.00 U
1,2,3,4-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U
1,2,4,5-Tetrachlorobenzene	0.00 U	0.00 U	0.00 U
Hexachlorobenzene	0.00 U	0.00 U	0.01 J
Pentachloroanisole	0.32	0.00 U	0.16
Pentachlorobenzene	0.00 U	0.00 U	0.00 U
Endosulfan II	0.00 U	0.00 U	0.66
Endosulfan I	0.07 J	0.00 U	0.13
Endosulfan Sulfate	0.00 U	0.00 U	0.00 U
Mirex	0.00 U	0.00 U	0.00 U
Chlorpyrifos	0.00 U	0.00 U	0.00 U
Total HCH	0.10	0.08	0.11
Total Chlordane	0.02	0.20	0.11
Total DDT	0.12	0.02	0.92

Qualifiers (Q): J=Below the MDL, U=Not detected

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g).

Compound	Coral 1		Coral 2		Coral 3		Coral 4		Coral 5		Coral 6	
BDE 1	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 2	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 3	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 10	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 7	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 8	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 11	0.1	J	0.0	U	0.0	U	0.0	U	0.1	J	0.0	U
BDE 12	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 13	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 15	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 30	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 32	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 17	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 25	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 33	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 28	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 35	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 37	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 75	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 49/71	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 47	0.2	J	0.1	J	0.1	J	0.1	J	0.0	U	0.2	J
BDE 66	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 77	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 100	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 119	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 99	0.0	U	0.0	U	0.2	J	0.0	U	0.0	U	0.0	U
BDE 116	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 118	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 85	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 126	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 155	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 154	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 153	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 138	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 166	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 183	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 181	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 190	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
Total PBDE	0.3		0.1		0.3		0.1		0.1		0.2	

Qualifiers (Q): J=Below the MDL; U=Not detected; I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	Coral 8		Coral 9		LTOC 1		LTOC 2		LTOC 3		LTOC 7	
BDE 1	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 2	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 3	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 10	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 7	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 8	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 11	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.2	J
BDE 12	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 13	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 15	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 30	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 32	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 17	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 25	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 33	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 28	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 35	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 37	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 75	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 49/71	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 47	0.1	J	0.1	J	0.0	U	0.0	U	0.0	U	0.1	J
BDE 66	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 77	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 100	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 119	0.0	U	0.0	U	0.0	U	0.6	JI	0.0	U	0.8	I
BDE 99	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 116	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 118	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 85	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 126	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 155	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 154	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 153	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 138	0.0	U	0.0	U	1.3		0.0	U	0.0	U	0.0	U
BDE 166	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 183	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 181	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 190	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
Total PBDE	0.1		0.1		1.3		0.6		0.0		1.1	

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 8		LTOC 9		LTOC 10		LTOC 11		LTOC 12		LTOC 13	
BDE 1	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 2	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 3	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 10	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 7	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 8	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 11	0.0	U	0.1	J	0.0	U	0.0	U	0.0	U	0.0	U
BDE 12	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 13	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 15	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 30	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 32	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 17	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 25	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 33	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 28	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 35	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 37	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 75	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 49/71	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 47	0.1	J	0.1	J	0.1	J	0.1	J	0.0	U	0.0	U
BDE 66	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 77	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 100	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 119	0.6	JI	0.6	JI	0.4	JI	0.4	JI	0.0	U	0.0	U
BDE 99	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 116	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 118	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 85	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 126	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 155	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 154	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 153	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 138	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 166	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 183	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 181	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 190	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
Total PBDE	0.7		0.8		0.5		0.5		0.0		0.0	

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 14	LTOC 15	LTOC 16	LTOC 17	LTOC 18	LTOC 19
BDE 1	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 2	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 3	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 10	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 7	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 8	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 11	0.0 U	0.1 J	0.0 U	0.0 U	0.0 U	0.0 U
BDE 12	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 13	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 15	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 30	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 32	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 17	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 25	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 33	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 28	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 35	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 37	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 75	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 49/71	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 47	0.0 U	0.3 J	0.1 J	0.0 U	0.1 J	0.0 U
BDE 66	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 77	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 100	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 119	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 99	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 116	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 118	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 85	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 126	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 155	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 154	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 153	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 138	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 166	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 183	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 181	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 190	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Total PBDE	0.0	0.4	0.1	0.0	0.1	0.0

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	LTOC 22	HTOC 1	HTOC 2	HTOC 3	HTOC 4	HTOC 6
BDE 1	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 2	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 3	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 10	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 7	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 8	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 11	0.2 J	0.0 U	0.0 U	0.0 U	0.0 U	0.1 J
BDE 12	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 13	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 15	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 30	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 32	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 17	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 25	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 33	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 28	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 35	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 37	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 75	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 49/71	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 47	0.1 J	0.1 J	0.1 J	0.2 J	0.1 J	0.1 J
BDE 66	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 77	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 100	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 119	0.3 J	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 99	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 116	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 118	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 85	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 126	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 155	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 154	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 153	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 138	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 166	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 183	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 181	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 190	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
Total PBDE	0.6	0.1	0.1	0.2	0.1	0.2

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	HTOC 7		HTOC 8		HTOC 9		HTOC 10		HTOC 12		HTOC 14	
BDE 1	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 2	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 3	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 10	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 7	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 8	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 11	0.0	U	0.0	U	0.0	U	0.0	U	0.1	J	0.0	U
BDE 12	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 13	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 15	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 30	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 32	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 17	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 25	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 33	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 28	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 35	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 37	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 75	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 49/71	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 47	0.0	U	0.0	U	0.1	J	0.1	J	0.0	U	0.0	U
BDE 66	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 77	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 100	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 119	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 99	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 116	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 118	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 85	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 126	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 155	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 154	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 153	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 138	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 166	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 183	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 181	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
BDE 190	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U	0.0	U
Total PBDE	0.0		0.0		0.1		0.1		0.1		0.0	

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix H. Polybrominated biphenyl ethers (PBDEs) in sediment samples from southwest Puerto Rico (ng/dry g) (continued).

Compound	HTOC 15	HTOC 16	HTOC 17	HTOC 19	HTOC 20	HTOC 21	HTOC 22
BDE 1	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 2	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 3	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 10	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 7	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 8	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 11	0.0 U	0.2 J	7.0	1.8	0.0 U	1.4	0.1 J
BDE 12	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 13	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 15	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 30	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 32	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 17	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 25	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 33	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 28	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 35	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 37	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 75	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 49/71	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 47	0.0 U	0.1 J	0.2 J	0.2 J	1.8 J	0.1 J	0.1 J
BDE 66	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 77	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 100	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 119	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	1.1 I
BDE 99	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 116	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 118	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 85	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 126	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 155	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 154	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 153	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 138	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 166	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 183	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U
BDE 181	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.0 U	0.2 J
BDE 190	0.0 U	0.0 U	0.0 U	0.0 U	0.5 U	0.0 U	0.0 U
Total PBDE	0.0	0.3	7.2	2.0	1.8	1.5	1.5

Qualifiers (Q): J=Below the MDL, U=Not detected, I=Interference

Appendix I. Butyltins in sediment samples from southwest Puerto Rico (ng Sn/dry g).

Site	Coral 1	Coral 2	Coral 3	Coral 4	Coral 5	Coral 6	Coral 8
Monobutyltin	0.00 U	0.00 U	0.28 J	0.20 J	0.36 J	0.30 J	0.23 J
Dibutyltin	0.16 J	0.00 U	0.12 J	0.19 J	0.11 J	0.16 J	0.10 J
Tributyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	Coral 9	LTOC1	LTOC2	LTOC3	LTOC7	LTOC8	LTOC9
Monobutyltin	0.22 J	0.47 J	0.46 J	0.65 J	0.51 J	0.60 J	0.67 J
Dibutyltin	0.25 J	0.22 J	0.16 J	0.26 J	0.15 J	0.08 J	0.11 J
Tributyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	LTOC10	LTOC11	LTOC12	LTOC13	LTOC14	LTOC15	LTOC16
Monobutyltin	0.28 J	0.59 J	0.48 J	0.29 J	0.27 J	0.33 J	0.84
Dibutyltin	0.08 J	0.09 J	0.24 J	0.23 J	0.09 J	0.12 J	0.20 J
Tributyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	LTOC17	LTOC18	LTOC19	LTOC22	HTOC1	HTOC2	HTOC3
Monobutyltin	0.39 J	0.31 J	0.39 J	0.72	0.00 U	0.00 U	1.54
Dibutyltin	0.09 J	0.00 U	0.09 J	0.14 J	0.19 J	0.29 J	0.55
Tributyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.29 J
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	HTOC4	HTOC6	HTOC7	HTOC8	HTOC9	HTOC10	HTOC12
Monobutyltin	0.42 J	0.24 J	0.20 J	0.00 U	0.29 J	0.24 J	0.70 J
Dibutyltin	0.12 J	0.11 J	0.22 J	0.12 J	0.23 J	0.20 J	0.13 J
Tributyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	HTOC14	HTOC15	HTOC16	HTOC17	HTOC19	HTOC20	HTOC21
Monobutyltin	0.46 J	0.49 J	0.17 J	0.32 J	0.32 J	0.27 J	0.79
Dibutyltin	0.00 U	0.00 U	0.10 J	0.22 J	0.28 J	0.20 J	0.09 J
Tributyltin	0.00 U	0.00 U	0.00 U	0.16 J	0.17 J	0.00 U	0.00 U
Tetrabutyltin	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U	0.00 U

Site	HTOC22
Monobutyltin	0.60 J
Dibutyltin	0.18 J
Tributyltin	0.00 U
Tetrabutyltin	0.00 U

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix J. Trace and major elements in sediment samples from southwest Puerto Rico ($\mu\text{g}/\text{dry g}$).

Compound	Coral 1	Coral 2	Coral 3	Coral 4	Coral 5	Coral 6
Ag	0 U	0 U	0 U	0 U	0 U	0 U
Al	1,060	1,020	2,460	3,350	6,660	2,840
As	10.3	5.3	0 U	0 U	5.11	0 U
Cd	0 U	0 U	0 U	0 U	0 U	0 U
Cr	11.3	8.88	7.57	11.1	17.4	8.64
Cu	1.13	1	1.89	2.77	4.51	2.24
Fe	2,700	1,440	1,950	2,870	5,580	2,050
Hg	0 U	0 U	0 U	0 U	0 U	0 U
Mn	191	83.2	63.5	86.5	111	56
Ni	3.85	2.63	4.26	6.08	10.6	4.92
Pb	1.03	0.852	0.817	1.25	1.68	1.01
Sb	0.196	0.111	0.055	0.101	0.085	0.0597
Se	0.144	0.09186	0.16	0.147	0.254	0.147
Si	4,500	3,670	8,350	11,500	20,900	10,200
Sn	0 U	0 U	0 U	0 U	0 U	0 U
Zn	1.67	1.55	3.34	4.48	8.63	4.18

Compound	Coral 8	Coral 9	LTOC1	LTOC2	LTOC3	LTOC7
Ag	0 U	0 U	0 U	0 U	0 U	0 U
Al	2,100	1,310	1,250	931	617	13,000
As	0 U	0 U	0 U	0 U	0 U	4.06
Cd	0 U	0 U	0 U	0 U	0 U	0 U
Cr	7.04	5.36	10.2	7.48	6.1	32.9
Cu	1.73	1.01	0.906	1.19	0 U	9.08
Fe	1,420	900	1,650	924	559	8,620
Hg	0 U	0 U	0 U	0 U	0 U	0 U
Mn	65.7	29	58.5	48.1	30	197
Ni	3.75	2.15	2.32	1.69	0 U	24.5
Pb	0.692	2.48	0.821	0.482	0.35	3.19
Sb	0.0513	0.055	0.133	0.0773	0.0434	0.0867
Se	0.16	0.158	0.09315	0.1	0.09964	0.28
Si	7,440	8,440	3,480	2,710	1,830	38,300
Sn	0 U	0 U	0 U	0 U	0 U	0.351
Zn	2.71	1.88	1.68	1.13	0 U	16.2

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix J. Trace and major elements in sediment samples from southwest Puerto Rico ($\mu\text{g}/\text{dry g}$) (continued).

Compound	LTOC8	LTOC9	LTOC10	LTOC11	LTOC12	LTOC13
Ag	0 U	0 U	0 U	0 U	0 U	0 U
Al	3,250	8,450	2,920	5,050	1,300	1,090
As	0 U	0 U	0 U	0 U	0 U	4.95
Cd	0 U	0 U	0 U	0 U	0 U	0 U
Cr	11.8	23.2	13.2	17.6	10.8	8.45
Cu	2.7	5.59	2.52	4.13	1.06	0.905
Fe	2,690	6,180	2,720	4,010	1,410	1,970
Hg	0 U	0 U	0 U	0 U	0 U	0 U
Mn	100	166	108	129	59.3	82.6
Ni	6.44	15.9	7.51	11	2.61	2.62
Pb	1.15	2.51	0.973	1.52	0.77	1.01
Sb	0.0507	0.0798	0.0534	0.0538	0.0891	0.171
Se	0.135	0.231	0.177	0.145	0.118	0.117
Si	10,900	27,100	9,980	16,800	3,820	3,020
Sn	0 U	0.302	0 U	0 U	0 U	0 U
Zn	4.32	11	4.06	6.66	1.41	2.04

Compound	LTOC14	LTOC15	LTOC16	LTOC17	LTOC18	LTOC19	LTOC22
Ag	0 U	0 U	0 U	0 U	0 U	0 U	0 U
Al	497	12,100	4,840	780	508	4,570	12,200
As	0 U	5.84	0 U	0 U	0 U	0 U	7.7
Cd	0 U	0 U	0 U	0 U	0 U	0 U	0 U
Cr	3.66	31	14.2	4.25	4.81	13.9	30.4
Cu	0 U	7.83	3.28	0 U	0 U	3.16	7.4
Fe	428	8,850	5,510	662	456	5,480	8,820
Hg	0 U	0 U	0 U	0 U	0 U	0 U	0 U
Mn	20.1	174	108	16.8	18.8	104	173
Ni	1.14	20.9	8.97	1.4	1.1	8.92	20.3
Pb	0.206	2.77	1.59	0.187	0.232	1.48	2.78
Sb	0 U	0.1	0.0866	0 U	0.0425	0.0918	0.11
Se	0.05744	0.315	0.146	0 U	0.08689	0.161	0.322
Si	1,680	35,900	16,000	2,250	2,180	15,000	36,600
Sn	0 U	0.429	0 U	0 U	0 U	0.231	0.419
Zn	0.42	15	7.7	1.21	0 U	7.54	14.9

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix J. Trace and major elements in sediment samples from southwest Puerto Rico ($\mu\text{g}/\text{dry g}$) (continued).

Compound	HTOC1	HTOC2	HTOC3	HTOC4	HTOC6	HTOC7
Ag	0 U	0 U	0 U	0 U	0 U	0 U
Al	485	924	7,050	2,970	6,220	580
As	0 U	6.61	0 U	0 U	0 U	0 U
Cd	0 U	0 U	0 U	0 U	0 U	0 U
Cr	4.51	11.5	14.3	10.8	18	3.21
Cu	0 U	0 U	6.04	2.15	4.01	0 U
Fe	506	1,390	4,280	2,460	5,990	477
Hg	0 U	0 U	0 U	0 U	0 U	0 U
Mn	22.8	66.1	61	85.4	125	26.9
Ni	0 U	2.06	8.21	5.35	10.8	1.12
Pb	1.54	0.749	3.49	1.04	1.9	0.247
Sb	0 U	0.145	0.127	0.0472	0.0806	0 U
Se	0.07967	0.153	0.301	0.13	0.169	0.07136
Si	1,950	2,580	21,700	10,500	19,700	1,990
Sn	0 U	0 U	0.34	0 U	0 U	0 U
Zn	0.577	0.898	11.9	3.9	9.18	1.01

Compound	HTOC8	HTOC9	HTOC10	HTOC12	HTOC14	HTOC15
Ag	0 U	0 U	0 U	0 U	0 U	0 U
Al	1,440	2,670	973	476	564	1,110
As	0 U	4.23	5.19	0 U	0 U	6.5
Cd	0 U	0 U	0 U	0.223	0 U	0 U
Cr	6.54	9	9.56	4.14	3.24	9.77
Cu	1.04	3.11	0 U	0.934	0 U	1.6
Fe	1,030	1,890	1,110	399	444	1,680
Hg	0 U	0 U	0 U	0 U	0 U	0 U
Mn	32.9	24.4	58.8	20.3	17.8	66.7
Ni	2.48	4.88	2.71	0 U	0 U	2.42
Pb	0.441	2.23	0.524	4	0.345	0.95
Sb	0.0608	0.0838	0.0915	0.196	0.0472	0.176
Se	0.124	0.267	0.094	0.079	0.104	0.135
Si	4,810	11,700	3,630	1,650	2,080	2,820
Sn	0 U	0 U	0 U	2.93	0 U	0 U
Zn	1.91	4.03	1.32	0.437	1.03	1.19

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix J. Trace and major elements in sediment samples from southwest Puerto Rico ($\mu\text{g}/\text{dry g}$) (continued).

Compound	HTOC16	HTOC17	HTOC19	HTOC20	HTOC21	HTOC22
Ag	0 U	0 U	0.0806	0 U	0 U	0 U
Al	12900	41100	66600	3630	1410	12000
As	4.83	0 U	3.94	0 U	0 U	0 U
Cd	0 U	0.0578	0.11	0 U	0 U	0 U
Cr	30.3	440	394	11.3	9.03	29.8
Cu	7.67	38.7	80.6	3.79	1.03	7.23
Fe	9910	42400	58900	2460	1030	7830
Hg	0 U	0.0954	0.0879	0 U	0 U	0 U
Mn	199	774	567	39	34	204
Ni	19.8	442	434	7.35	2.53	20.6
Pb	2.81	12.9	13.7	1.01	0.436	3.06
Sb	0.0989	0.243	0.325	0.0908	0.0453	0.0827
Se	0.279	0.252	0.432	0.356	0.123	0.279
Si	41400	143000	187000	12700	4310	36000
Sn	0.329	1.22	2.17	0 U	0 U	0.316
Zn	15.1	55.2	90.7	5.4	1.94	14

Qualifiers (Q): J=Below the MDL; U=Not detected

Appendix K. *Clostridium perfringens* in sediment samples from southwest Puerto Rico.

Site	Sample composition	% water	% sediment	<i>Cperf</i> dry (CFU/g)
Coral 1	lg. particles/sandy	25.49	74.51	0
Coral 2	fine sand	26.67	73.33	0
Coral 3	crushed shells	36.49	63.51	0
Coral 4	sand, mostly crushed shells	29.41	70.59	0
Coral 5	fine sand	33.93	66.07	0
Coral 6	crushed shells, sand	30.77	69.23	5.21
Coral 8	crushed shells, sand	37.50	62.50	0
Coral 9	crushed shells, sand	40.00	60.00	0
LTOC1	lg. particles/sand/shells	22.95	77.05	2.30
LTOC2	sandy	26.42	73.58	0
LTOC3	sandy	29.63	70.37	0
LTOC7	mud	40.68	59.32	13.49
LTOC8	mud	35.21	64.79	0
LTOC9	mud	40.63	59.38	25.26
LTOC10	sandy	36.05	63.95	0
LTOC11	dry sand	32.08	67.92	0
LTOC12	lg. particles/dry sand	27.54	72.46	2.51
LTOC13	lg. particles/dry sand	28.41	71.59	0
LTOC13	lg. particles/dry sand	28.57	71.43	0
LTOC14	crushed shells	30.43	69.57	0
LTOC15	watery mud	43.59	56.41	46.65
LTOC16	crushed shells/mud	34.43	65.57	0
LTOC17	sand	25.86	74.14	0
LTOC18	crushed shells	29.17	70.83	0
LTOC19	crushed shells	34.38	65.63	2.72
LTOC22	mud	43.28	56.72	18.08
HTOC1	fine sand	29.85	70.15	0
HTOC2	wet sand, shells	19.74	80.26	0
HTOC3	wet sand, shells	44.64	55.36	9.68
HTOC4	sandy	30.00	70.00	0
HTOC4	sandy	29.82	70.18	0
HTOC6	watery sand	35.71	64.29	12.32
HTOC7	crushed shells, sand	30.00	70.00	0
HTOC8	crushed shells, sand	27.78	72.22	0
HTOC9	wet sand, crushed shells	49.09	50.91	7.21
HTOC10	crushed shells, sand	25.61	74.39	0
HTOC12	crushed shells	28.75	71.25	0
HTOC14	crushed shells	33.78	66.22	0
HTOC15	lg. particles/sand/shells	22.41	77.59	0
HTOC16	watery mud	41.18	58.82	0
HTOC17	watery mud	42.86	57.14	184.21
HTOC19	watery sand	63.41	36.59	1760.69
HTOC20	watery sand	52.83	47.17	3.82
HTOC20	watery sand	56.90	43.10	0
HTOC21	fine sand	32.00	68.00	0
HTOC22	mud	42.31	57.69	9.37

CFU = [(Mean *Clostridium* spores x 40)/volume assayed]/assay weight

Appendix L. Total nitrogen and phosphorus detected in water samples from southwest Puerto Rico (mg/L).

Site	Total Nitrogen	Total Phosphorus
Coral 1	0.204	0.0096
Coral 2	0.278	0.0077
Coral 3	0.100	0.0069
Coral 4	0.084	0.0065
Coral 5	0.335	0.0095
Coral 6	0.098	0.0080
Coral 8	0.114	0.0066
Coral 9	0.104	0.0057
LTOC1	0.125	0.0069
LTOC2	0.146	0.0065
LTOC3	0.153	0.0065
LTOC5	0.107	0.0065
LTOC7	0.197	0.0068
LTOC8	0.173	0.0076
LTOC9	0.088	0.0057
LTOC10	0.202	0.0065
LTOC11	0.146	0.0061
LTOC12	0.157	0.0072
LTOC13	0.150	0.0065
LTOC14	0.143	0.0061
LTOC15	0.215	0.0076
LTOC16	0.085	0.0058
LTOC17	0.196	0.0065
LTOC18	0.102	0.0061
LTOC19	0.100	0.0065
LTOC20	0.106	0.0061
LTOC22	0.157	0.0073
HTOC1	0.110	0.0058
HTOC2	0.164	0.0062
HTOC3	0.156	0.0062
HTOC4	0.100	0.0061
HTOC6	0.079	0.0058
HTOC7	0.084	0.0065
HTOC8	0.106	0.0062
HTOC9	0.152	0.0069
HTOC10	0.118	0.0069
HTOC11	0.098	0.0061
HTOC12	0.094	0.0054
HTOC14	0.096	0.0065
HTOC15	0.193	0.0057
HTOC16	0.178	0.0091
HTOC17	0.378	0.0238
HTOC18	0.097	0.0065
HTOC19	0.368	0.0483
HTOC20	0.095	0.0065
HTOC21	0.216	0.0069
HTOC22	0.165	0.0084

United States Department of Commerce

Carlos M. Gutierrez
Secretary

National Oceanic and Atmospheric Administration

Vice Admiral Conrad C. Lautenbacher, Jr. USN (Ret.)
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National Ocean Service

John H. Dunnigan
Assistant Administrator for Ocean Service and Coastal Zone
Management

