

## Shark virgin birth produces multiple, viable offspring

Feldheim, K. A., Chapman, D. D., Sweet, D., Fitzpatrick, S., Prodohl, P., Shivji, M. S., & Snowden, B. (2010). Shark virgin birth produces multiple, viable offspring. Journal of Heredity, 101(3)(3), 374-377. DOI: 10.1093/jhered/esp129

Published in: Journal of Heredity

Queen's University Belfast - Research Portal:

Link to publication record in Queen's University Belfast Research Portal

General rights

Copyright for the publications made accessible via the Queen's University Belfast Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy
The Research Portal is Queen's institutional repository that provides access to Queen's research output. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact openaccess@qub.ac.uk.

© The American Genetic Association. 2010. All rights reserved. For permissions, please email: journals.permissions@oxfordjournals.org.

# Shark Virgin Birth Produces Multiple, Viable Offspring

Kevin A. Feldheim, Demian D. Chapman, Doug Sweet, Seán Fitzpatrick, Paulo A. Prodöhl, Mahmood S. Shivji, and Bob Snowden

From the Pritzker Laboratory for Molecular Systematics and Evolution, The Field Museum, 1400 South Lake Shore Drive, Chicago, IL 60605 (Feldheim); the Institute for Ocean Conservation Science, School of Marine and Atmospheric Science, Stony Brook University, Stony Brook, NY 11794-5000 (Chapman); the London State Fish Hatchery, Division of Wildlife, Ohio Department of Natural Resources, 2470 Robert's Mill Road SW, London, OH 43140 (Sweet); the School of Biological Sciences, Queen's University Belfast, Belfast, BT9 7BL, Northern Ireland, UK (Fitzpatrick and Prodöhl); The Guy Harvey Research Institute, Oceanographic Center, Nova Southeastern University, Dania Beach, FL 33004 (Shivji); and the Pittsburgh Zoo & PPG Aquarium, One Wild Place, Pittsburgh, PA 15206 (Snowden).

Address correspondence to Kevin A. Feldheim at the address above, e-mail: kfeldheim@fieldmuseum.org.

Facultative automictic parthenogenesis has only recently been confirmed in the most ancient jawed vertebrates, the chondrichthyan fishes (sharks, batoids, and chimeras). To date, however, in both documented cases, the females have only produced a single parthenogen offspring, and none of these have lived for more than 3 days. We present a genetically verified case of automictic parthenogenesis by a white-spotted bamboo shark (*Chiloscyllium plagiosum*), in which at least 2 parthenogens were produced and survived for 5 years or more. These findings demonstrate that some female sharks are capable of producing, multiple, viable offspring through parthenogenesis.

**Key words:** AFLPs, microsatellites, parthenogenesis, whitespotted bamboo shark

There is growing interest in the evolutionary significance and conservation implications of facultative parthenogenesis in vertebrates, as well as the potential biomedical applications that could result from understanding the underlying mechanisms involved (Lampert 2008). Though rare, cases of spontaneous parthenogenesis have now been documented in all major vertebrate lineages other than mammals (e.g., sharks [Chapman et al. 2007], teleost fish [Lampert et al. 2007], reptiles [Schuett et al. 1998; Groot et al. 2003; Watts et al. 2006], and birds [Olsen 1975]). Most of these appear to be the result of fusion of postmeiotic products (i.e., the ovum and a polar body: "automictic parthenogenesis"), which produces diploid offspring that are either homozygous for one or the other maternal alleles at many loci (through "terminal fusion") or that are genetically identical or nearly identical to the mother (through "central fusion," Lampert 2008). Automictic parthenogenesis was only recently genetically verified in chondrichthyan fish (sharks,

batoids, and chimeras; Chapman et al. 2007, 2008). In the first case, a newly mature female bonnethead shark (Sphyrna tiburo) that had been wild captured as a young-of-the-year and thereafter isolated from conspecific males in captivity for more than 3 years produced a single, live female offspring. This offspring was homozygous for a maternal allele at 4 polymorphic microsatellite loci (Chapman et al. 2007) and only survived for 3 days before being killed by another fish in the aquarium. In the second case, a welldeveloped female embryo was discovered during the necropsy of female blacktip shark (Carcharbinus limbatus) that had also been wild captured as a juvenile and subsequently isolated from males for 9 years. The embryo was homozygous for a maternal allele at 6 polymorphic microsatellite loci (Chapman et al. 2008). In summary: 1) verified cases of parthenogenesis in chondrichthyan fish are restricted to sharks of the Order Carcharhiniformes that exhibit placental viviparity, 2) in both cases, the offspring exhibited complete homozygosity at the microsatellite loci that were typed, 3) there is no evidence that females can produce multiple offspring from parthenogenesis, and 4) the verified cases provide no information on long-term viability of sharks produced in this manner. These issues make it difficult to assess the biological significance of parthenogenesis in sharks.

In 1996, the Belle Isle Aquarium of the Detroit Zoological Institute acquired one female brown-banded bamboo shark (*Chiloscyllium punctatum*; ISIS accession no. 5458) and one female white-spotted bamboo shark (*Chiloscyllium plagiosum*; ISIS accession no. 5455) as donations from 2 private aquarium hobbyists. Each bamboo shark was purchased by the hobbyist as an egg case and was hatched and reared with no contact with other bamboo sharks. Both animals were sexually immature when first acquired by the

Table 1. Genotypes of a bamboo shark mother (ISIS accession no. 5455) and 2 of her pups at 10 microsatellite loci

Shark	Cpl (II <sup>a</sup> )	Cp3 (6 <sup>a</sup> )	Cp4 (5 <sup>a</sup> )	Cp5 (6 <sup>a</sup> )	C <sub>P</sub> 7 (4 <sup>a</sup> )	Cp8 (6 <sup>a</sup> )	Cp9 (12 <sup>a</sup> )	Cpl80 (4 <sup>b</sup> )	Cpl150 (4 <sup>b</sup> )	Cpl471 (3 <sup>b</sup> )
Mother	315/315	228/228	377/377	418/422	176/176	283/283	200/208	265/293	202/202	224/226
Pup1	315/315	228/228	377/377	422/422	176/176	283/283	208/208	265/265	202/202	226/226
Pup2	315/315	228/228	377/377	422/422	176/176	283/283	208/208	293/293	202/202	226/226

<sup>&</sup>lt;sup>a</sup> Number of alleles observed in a sample of 33–35 individuals taken from the wild near Zhanjiang, China (Ding et al. 2009).

Belle Isle Aquarium. Because bamboo sharks are plentiful in zoological park facilities, there was no intent on breeding these individuals, and no males were ever added to cohabitate with these 2 females. In fact, there were no males of either species or any closely related species housed anywhere in the facility. The brown-banded bamboo shark never oviposited eggs. The white-spotted bamboo shark, on the other hand, oviposited egg cases on numerous occasions over a 6-year period. None of these egg cases were saved, as they were presumed to be infertile, until in early 2002 the Curator of Fishes (D.S.) heard about the virgin birth of a bonnethead shark, S. tiburo, at the Henry Doorly Zoo in Omaha, Nebraska (later genetically confirmed by Chapman et al. 2007). At this time, D.S. initiated an experiment to demonstrate that this female was reproducing in the absence of males (i.e., by parthenogenesis). Unfertilized eggs cases laid by this female were collected and incubated to see if they would develop into viable embryos. In 2005, genetic testing was initiated to 1) verify an absence of paternal genetic material in the offspring and 2) test the hypothesis that the offspring exhibit elevated homozygosity relative to their mother, as has been observed for other sharks (e.g., Chapman et al. 2007, 2008).

Seven eggs were deposited by the C. plagiosum between March and August of 2002. All 7 were saved and incubated for 15-16 weeks. Four of them developed embryos that went to full term (15 weeks) before hatching. All embryos were females and were kept in a separate tank from the mother. Three of the four individuals were vigorous within the egg case and hatched without assistance; the fourth appeared to be weakening as it surpassed its expected hatch date, and therefore, it's emergence was assisted by excising the shark from the egg case. This last individual (ISIS accession no. 10901) never initiated feeding and only lived for 14 days. The other 3 individuals (ISIS accession nos 10846, 10892, and 10895) were fully viable and grew well during their first year of life. One of these individuals (10846) perished after jumping from its tank at 15 months of age. Unfortunately, the 2 offspring that did not survive were not tissue sampled before being discarded. In contrast, the other 2 remaining offspring continued growing and were subadult size by May of 2005 at that time they were tissue sampled (via small clips of fin). The 2 surviving offspring and the mother were eventually transferred from the closing Belle Isle Aquarium to Denver's Landry's Downtown Aquarium and held in a tank with a large group of conspecific individuals, which precluded further tracking of these sharks.

We used microsatellites and amplified fragment length polymorphism (AFLP) to survey the genome of the suspected parthenogens to identify distinct nonmaternal alleles and potentially refute parthenogenesis. Seven variable microsatellite loci (Cp1, Cp3, Cp4, Cp5, Cp7, Cp8, and Cp9) described elsewhere (4-12 alleles/locus, Ding et al. 2009) and 3 new loci (Cpl80, Cpl150, and Cpl471) developed using an enrichment protocol (Glenn and Schable 2005) were used to genotype the mother and her 2 subadult offspring. To assess variability of the new loci, 9 captive animals (3 from the Pittsburgh Zoo and 6 from the University of South Florida) were also genotyped. All 3 loci amplified alleles in all 9 individuals and exhibited 3 (Cpl471) or 4 (Cpl80 and Cpl150) alleles. Microsatellite genotypes of all individuals, including the mother and her offspring, are shown in Table 1. Multilocus AFLP was also used to characterize the genetic diversity of the offspring. AFLP screening was carried on a LI-COR dual-laser system after the protocol of Zenger et al. (2006) using the selective primers in 14 different combinations (details available on request). Resulting fragments were scored and analyzed (band sharing) using the GeneProfiler software (Scanalytics Inc., Fairfax, VA).

The microsatellite and AFLP genotypes of the C. plagiosum offspring are consistent with their suspected parthenogenetic origin and are characteristic of terminal as opposed to central fusion automixis (Lampert 2008). Each proved to be homozygous at every microsatellite locus for one or the other of the possible maternal alleles (Table 1). Like the microsatellite analysis, all AFLP fragments observed in the suspected parthenogen offspring were also present in the mother (i.e., there were no unique paternal fragments in the offspring). Furthermore, both offspring displayed a reduced number of fragments in comparison to the mother (1.36% and 2.31% fewer fragments than the mother), and they differed in 1.74% of their fragments. The combined microsatellite and AFLP data set failed to reveal unique paternal genetic variation. Combining these genetic data with the well-documented captive history of the mother indicating lifetime isolation from males, the evidence that these births were due to parthenogenesis is thus unequivocal. The elevated homozygosity of the 2 offspring relative to the mother reflects previous cases of parthenogenesis in sharks and implies that terminal fusion automixis is the probable parthenogenetic mechanism (Chapman et al. 2007, 2008).

Notably, the 9 aquarium bred *C. plagiosum* we tested, including the mother, appeared to be from inbred lines.

<sup>&</sup>lt;sup>b</sup> Number of alleles observed in a sample of 9 aquarium-bred individuals in the United States (for loci Cpl80, Cpl150, and Cpl471).

Each individual exhibited a high proportion (~50%) of homozygous microsatellite loci, and several loci known to be polymorphic in a wild population (Ding et al. 2009) were monomorphic in the 9 captive animals we tested (e.g., mother in Table 1). Multilocus AFLP analysis supports the inbred status of the aquarium-bred individuals: they shared an average of 91.5% of AFLP fragments, despite being from different institutions. This is higher than what has been observed for other shark species. For instance, Chapman et al. (2007) showed that unrelated wild-captured bonnethead sharks (S. tiburo) share <69% of their AFLP fragments generated using similar methodology. Even given the background levels of homozygosity in the captive animals, the parthenogens were the only individuals that exhibited complete homozygosity at the 10 microsatellite loci, and there were a sufficient number of polymorphic AFLP markers (148) to have shown paternal alleles if they were present.

Though genetically untested, it is highly probable that the unsampled third and fourth of the hatched offspring were also parthenogens, which means that up to 4 of 7 eggs in this clutch (57%) developed asexually in the absence of sperm. This is the first genetically confirmed report of multiple offspring being produced by parthenogenesis by any female chondrichthyan. Moreover, the discovery of automictic parthenogenesis in C. plagiosum extends the occurrence of this form of reproduction from the Order Carcharhiniformes to the nonsister Order Orectolobiformes. Thus, automictic parthenogenesis is now known to occur in taxonomically diverse shark species with reproductive modes ranging from oviparity to placental viviparity (Chapman et al. 2007, 2008). It is therefore reasonable to speculate that more shark species are capable of asexual reproduction on occasion. Although sperm storage has been demonstrated in many species of shark (Pratt 1993), our findings suggest that inferences of long-term sperm storage in captive female sharks producing offspring, despite isolation from males (e.g., Castro et al. 1988), should be considered suspect unless genetic testing can exclude parthenogenesis.

Very little is known about the viability of automictic parthenogens in sharks (Chapman et al. 2007, 2008). Although the elevated homozygosity of automictic parthenogens could reduce their fitness by increasing expression of recessive genetic disorders and decreasing genomic diversity, long-term viability of automictic parthenogens has been reported in teleost fish, reptiles, and birds (Olsen 1975; Schuett et al. 1998; Watts et al. 2006; Lampert et al. 2007). We document that sharks also can produce viable offspring in this manner. Unfortunately, the closure of the Belle Isle aquarium and the disbursement of the sharks preclude an assessment of whether or not these parthenogens are also fertile.

Although the present case is the only genetically documented instance of parthenogenesis in *C. plagiosum*, we are aware of other cases of possible parthenogenesis by captive *C. plagiosum* females (e.g., Voss et al. 2001, http://www.azcentral.com/news/articles/1113phx-shark1114.html). This

species is the only elasmobranch where there is currently more than one report of possible parthenogenesis by a female in captivity; as such it may prove to be a useful model elasmobranch to study the mechanism of facultative parthenogenesis in fish.

The ability of captive C. plagiosum to produce multiple viable offspring through parthenogenesis may be related to the high levels of homozygosity and presumed inbred lines we observed in these aquarium animals. Hedrick (2007) noted that the likelihood of producing viable embryos by automictic parthenogenesis is greatly reduced when lethal recessive alleles are present in the genome. We speculate that inbred individuals, with elevated homozygosity relative to wild conspecifics, are already "purged" of highly deleterious and lethal recessive alleles to a certain extent, which could enhance their ability to produce viable offspring through automictic parthenogenesis. Automictic parthenogenesis might then be expected to result in viable offspring more often in inbred captive populations and, perhaps, in genetically depauperate wild populations of endangered species than in outbred lines.

We show that *C. plagiosum* is capable of automictic parthenogenesis and provide the first evidence in sharks that multiple offspring can be produced in this manner and that they can live for long periods (>5 years). We suggest that other reported cases of female *C. plagiosum* producing viable offspring in the absence of males are also likely to be the result of parthenogenesis. The demonstration of parthenogenesis in a third lineage of sharks raises the prospect that this reproductive capability may not be uncommon in these ancient fishes.

#### **Funding**

Institute for Ocean Conservation Science at Stony Brook. Microsatellite development was funded by the Grainger Foundation and was carried out in the Pritzker Laboratory for Molecular Systematics and Evolution operated with support from the Pritzker Foundation. AFLP analysis was carried out in the Fish Genetics and Molecular Ecology Laboratory at Queen's University Belfast with support of the Beaufort Fish Population Genetics Award carried out under the *Sea Change* Strategy and the Strategy for Science Technology and Innovation, funded under the Marine Research Sub-Programme of the National Development Plan 2007–2013.

#### **Acknowledgments**

We would like to thank P. Motta and A. Henningsen for providing tissues.

### References

Castro JI, Bubucis PM, Overstrom NA. 1988. The reproductive biology of the chain dogfish, *Scyliorbinus retifer*. Copeia. 1988:740–746.

Chapman DD, Firchau B, Shivji MS. 2008. Parthenogenesis in a large-bodied requiem shark, the blacktip *Carcharbinus limbatus*. J Fish Biol. 73:1473–1477.

Chapman DD, Shivji MS, Louis E, Sommer J, Prodöhl PA. 2007. Virgin birth in a hammerhead shark. Biol Lett. 3:425–427.

Ding S, Pan Y, Zeng H, Wang J. 2009. Characterization of 12 polymorphic microsatellite loci for the whitespotted bamboo shark (*Chiloscyllium plagiosum* Bennett). Mol Ecol Resour. 9:1398–1400.

Glenn TC, Schable NA. 2005. Isolating microsatellite DNA loci. Methods Enzymol. 395:202–222.

Groot TVM, Bruins E, Breeuwer JAJ. 2003. Molecular genetic evidence for parthenogenesis in the Burmese python, *Python molurus bivittatus*. Heredity. 90:130–135.

Hedrick PW. 2007. Virgin birth, genetic variation and inbreeding. Biol Lett. 3:715-716.

Lampert KP. 2008. Facultative parthenogenesis in vertebrates: reproductive error or chance? Sex Dev. 2:290–301.

Lampert KP, Lamatsch DK, Fischer P, Epplen JT, Nanda I, Schmid M, Schartl M. 2007. Automictic reproduction in interspecific hybrids of poeciliid fish. Curr Biol. 17:1948–1953.

Olsen MW. 1975. Avian parthenogenesis. United States of Agriculture Publication No: ARS-NE-65. Beltsville (MD): USDA.

Pratt HLJ. 1993. The storage of spermatozoa in the oviducal glands of western North Atlantic sharks. Environ Biol Fish. 38:139–149.

Schuett GW, Fernandez PJ, Chiszar D, Smith HM. 1998. Fatherless sons: a new type of parthenogenesis in snakes. Fauna. 1:19–25.

Voss J, Berti L, Michel C. 2001. Chiloscyllium plagiosum (Anon, 1830) born in captivity: hypothesis for gynogenesis. Bull Inst Oceanogr Monaco. 20:351–353.

Watts PC, Buley KR, Sanderson S, Boardman W, Ciofi C, Gibson R. 2006. Parthenogenesis in Komodo dragons. Nature. 444:1021–1022.

Zenger KR, Stow AJ, Peddemors V, Briscoe DA, Harcourt R. 2006. Widespread utility of highly informative AFLP molecular markers across divergent shark species. J Hered. 5:1–5.

Received September 24, 2009; Revised November 20, 2009; Accepted December 9, 2009

Corresponding Editor: Stephen Karl